

<b>REPORT DOCUMENTATION PAGE</b>				Form Approved OMB NO. 0704-0188	
<p>The public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington VA, 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number.</p> <p>PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.</p>					
1. REPORT DATE (DD-MM-YYYY) 05-06-2021		2. REPORT TYPE Final Report		3. DATES COVERED (From - To) 1-Jul-2016 - 31-Jan-2021	
4. TITLE AND SUBTITLE Final Report: Chemical Sciences: Transient Nanopatterns by Biocatalytic Self-Assembly				5a. CONTRACT NUMBER W911NF-16-1-0113	
				5b. GRANT NUMBER	
				5c. PROGRAM ELEMENT NUMBER 611102	
				5d. PROJECT NUMBER	
6. AUTHORS				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAMES AND ADDRESSES Research Foundation CUNY - Advanced Sc 85 Saint Nicolas Terrace  New York, NY 10031 -1246				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS (ES) U.S. Army Research Office P.O. Box 12211 Research Triangle Park, NC 27709-2211				10. SPONSOR/MONITOR'S ACRONYM(S) ARO	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S) 69180-CH.21	
12. DISTRIBUTION AVAILABILITY STATEMENT Approved for public release; distribution is unlimited.					
13. SUPPLEMENTARY NOTES The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision, unless so designated by other documentation.					
14. ABSTRACT					
15. SUBJECT TERMS					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT	15. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON
a. REPORT UU	b. ABSTRACT UU	c. THIS PAGE UU			Rein Ulijn
			UU		19b. TELEPHONE NUMBER 212-413-3380

# RPPR Final Report

as of 15-Oct-2021

Agency Code: 21XD

Proposal Number: 69180CH

Agreement Number: W911NF-16-1-0113

## INVESTIGATOR(S):

**Name:** PhD Elisa Riedo  
**Email:** elisa.riedo@asrc.cuny.edu  
**Phone Number:** 2124133384  
**Principal:** N

**Name:** Rein Ulijn  
**Email:** Rein.Ulijn@asrc.cuny.edu  
**Phone Number:** 2124133380  
**Principal:** Y

Organization: **Research Foundation CUNY - Advanced Science Research Center**

Address: 85 Saint Nicolas Terrace, New York, NY 100311246

Country: USA

DUNS Number: 831361857

EIN:

**Report Date:** 30-Apr-2021

Date Received: 05-Jun-2021

**Final Report** for Period Beginning 01-Jul-2016 and Ending 31-Jan-2021

**Title:** Chemical Sciences: Transient Nanopatterns by Biocatalytic Self-Assembly

**Begin Performance Period:** 01-Jul-2016

**End Performance Period:** 31-Jan-2021

**Report Term:** 0-Other

Submitted By: Rein Ulijn

Email: Rein.Ulijn@asrc.cuny.edu

Phone: (212) 413-3380

**Distribution Statement:** 1-Approved for public release; distribution is unlimited.

**STEM Degrees:** 3

**STEM Participants:** 3

**Major Goals:** It was the overall aim of this research program to develop transient nanoscale patterns that change their chemical information content and functionality over time with nanoscale resolution. To achieve this, we combined pioneering work from Riedo's Lab (thermochemical nanolithography–TCNL), and Ulijn's Lab (biocatalytic self-assembly).

A major goal was to provide fundamental understanding and take concrete steps towards laboratory-based mimicry of fabrication approaches used by living systems. We achieved this by exploiting similar mechanisms used by living systems (catalytic amplification, self-assembly, structural confinement) but harness them as new tools for nanoscale fabrication, in non-biological contexts.

**Accomplishments:** The objective of this collaborative grant between the labs of Rein Ulijn (CUNY) and Elisa Riedo (NYU) was to combine revolutionary nanoscale patterning with catalytic self-assembly to obtain spatio-temporal control of nanostructure formation and breakdown and consequent dynamic control of functionality. By using thermochemical scanning probe lithography (tSPL) we were able to precisely tune the film 3D topographical landscape simultaneously with surface chemistry. This provided control over positioning and concentration of surface-immobilized enzymes [1,2]. Furthermore, by making use of competing enzyme-catalyzed assembly and dis-assembly reactions, we could demonstrate the transient formation of nanoscale fibers that have programmable chemical content and (electronic) functionality, as well as time programmable assembly - disassembly behavior. We studied fundamental aspects of biocatalytic assembly at solid-liquid interfaces by using both synthetic and biological surfaces and particles. Finally, we developed a versatile and modular chemical method to incorporate fluorescent dyes into peptide nanostructures, which allowed us to visualize in real-time the formation and breakdown of nanostructures with super-resolution (STED) fluorescent microscopy. More recently, we demonstrated the possibility to apply tSPL for the production of high performing 2D materials-based field effect transistors (FETs) fabricated on flexible materials without the need of masks, or markers, and in ambient conditions. Also, we have developed innovative methodologies for the time- and cost-effective scaling up of tSPL in combination with a biocompatible polymer, and we have replicated the exact topography of the bone tissue microenvironment with unprecedented sub-15 nm resolution over millimeter scale areas creating new possibilities for biomedical research and applications.

## RPPR Final Report as of 15-Oct-2021

The project resulted in 15 publications, one patent and training opportunities for several postdocs, graduate and undergraduate researchers as detailed in the uploaded report.

A pdf is attached with more details.

**Training Opportunities:** Training opportunities provided to students and postdocs include:

- experimental training in nano fabrication, surface analysis, imaging provided at CUNY ASRC and NYU. Training is provided by core facility directors and students eventually run these instruments and acquire data independently.
- mentoring opportunities for all postdocs and graduate students through Sumer research programs at NYU and CUNY
- all students and postdocs regularly participated in national and international conferences and workshops through presentation of posters and contributed talks. Examples in crude MRS, ACS, APS.

**Results Dissemination:** Dissemination of research results through regular interaction with groups of high school children to CUNY ASRC as part of our Illumination Space science exhibits. Postdocs and Grad students are involved in hosting students at these events.

Ulijn participated in a summer program for high school students through the World Science Festival in NYC and created an online course for gifted high school students across the world.

Given the broad impact of the project on different research area, students and postdocs worked on diverse research topics and areas and even interdisciplinary fields. Students and postdocs were encouraged to work in diverse groups and to build a research group consisting of people from different backgrounds. Relating with people from different areas and disciplines often brought up new perspectives and new ideas.

Part of the dissemination activity of the project was devoted to the Diversity, Equity and Inclusion (DEI) initiative organized by Riedo (Chair of the Committee) in the Chemical and Biochemical department at Tandon. This activity is aimed at giving voice to racial and gender diversity problematics and addresses issues such as sexual harassment and work-life balance issues within STEM career path development, and how they will prepare themselves for key career-based transitions.

**Honors and Awards:** Ulijn was awarded a Vannevar Bush Faculty Fellowship from ONR (2021).

Ulijn was elected co-Chair of the editorial board of ChemSystemsChem (Wiley) (2020).

Ulijn was elected to World Science University Faculty for the World Science Festival's global science scholar program (2020).

Ulijn awarded Batsheva de Rothschild Fellowship, personal travel fellowship to support lecture tour and University visits in Israel (Tel Aviv, Weizmann Institute, Ben Gurion, Hebrew University) (2020).

AFOSR DURIP ( Rein Ulijn Co-PI) Dual Source Single Crystal X-ray Diffractometer for the Characterization of Small Molecules and Materials, \$399,000 (PI Share-\$0), 7/15/20-7/14/21

Ulijn (in his role chair) received sponsorship from ARO for the Gordon Research Conference on Systems Chemistry (June 2020). The conference was postponed to 2022 due to COVID.

Riedo was invited speaker at the Faraday Discussions on "Nanolithography of Biointerfaces" (ARO UK office program managers among the participants, Hollie Pietsch CCDC Atlantic).

Riedo became Editorial Board Member of Nature 2D Materials and Applications

Riedo Invited Speaker at the 2021 TMS Meeting

Riedo Invited Speaker at the 2020 Fall MRS Meeting

Riedo Invited Speaker at the Workshop on 2D and Quantum Materials at the 2020 CFN/NSLS-II Brookhaven Meeting

Riedo's work on tSPL selected to appear in the National Nanotechnology Initiative (NNI) Supplement to the President's 2020 Budget (which also serves as the annual report to Congress for the NNI).

ARO Materials Science (Elisa Riedo, Single PI) Novel properties of pressure activated 2D materials, \$569,887, 08/01/20-07/31/23

Post-Doc Zanut received the NYU Provost Fellowship.

Ph.D. Xiangyu Liu won a Conference Award for the World Congress on Micro and Nano Manufacturing.

**Protocol Activity Status:**

## RPPR Final Report as of 15-Oct-2021

**Technology Transfer:** Ulijn: Virtual seminar at Soldier Center, US Army Future Command, Natick MA (May 2020).  
Riedo: Provisional Patent Application: "Biomimetic Tissue And Method Of Use Thereof", NYU and NYSCF (2021)  
Riedo: new collaboration with Thomas J. Kiel, Armaments Center, CCDC U.S. Army Futures Command Picatinny Arsenal  
Riedo: ongoing partnership with SwissLitho and IBM for commercialized tSPL.  
Visit by Wendy Mills and Dawanne Poree to CUNY ASRC (October 2018).  
Visit by Rajesh Naik (AFoSR) to CUNY ASRC as part of 5-year review (February 2019).  
Visit by Thomas J. Kiel, U.S. Army Futures Command Picatinny Arsenal to NYU Tandon Engineering (May 2019)  
Riedo participated in a virtual meeting with Dr. Daniel Cole, Dr. Dan Magagnosc, and Dr. Matt Guziewski from ARL to discuss possible interactions and collaborations.

### PARTICIPANTS:

**Participant Type:** PD/PI

**Participant:** Rein Ulijn

**Person Months Worked:** 2.00

Project Contribution:

National Academy Member: N

**Funding Support:**

**Participant Type:** Co PD/PI

**Participant:** Elisa Riedo

**Person Months Worked:** 2.00

Project Contribution:

National Academy Member: N

**Funding Support:**

**Participant Type:** Postdoctoral (scholar, fellow or other postdoctoral position)

**Participant:** Mohit Kumar

**Person Months Worked:** 15.00

Project Contribution:

National Academy Member: N

**Funding Support:**

**Participant Type:** Postdoctoral (scholar, fellow or other postdoctoral position)

**Participant:** Ankit Jain

**Person Months Worked:** 2.00

Project Contribution:

National Academy Member: N

**Funding Support:**

**Participant Type:** Postdoctoral (scholar, fellow or other postdoctoral position)

**Participant:** Alessandra Zanut

**Person Months Worked:** 3.00

Project Contribution:

National Academy Member: N

**Funding Support:**

**Participant Type:** Postdoctoral (scholar, fellow or other postdoctoral position)

**Participant:** Xiaouri Zheng

**Person Months Worked:** 3.00

Project Contribution:

**Funding Support:**

**RPPR Final Report**  
as of 15-Oct-2021

National Academy Member: N

**Participant Type:** Postdoctoral (scholar, fellow or other postdoctoral position)

**Participant:** Annalisa Calo

**Person Months Worked:** 8.00

**Funding Support:**

Project Contribution:

National Academy Member: N

**Participant Type:** Postdoctoral (scholar, fellow or other postdoctoral position)

**Participant:** Edoardo Albisetti

**Person Months Worked:** 3.00

**Funding Support:**

Project Contribution:

National Academy Member: N

**Participant Type:** Graduate Student (research assistant)

**Participant:** Naxhije Berisha

**Person Months Worked:** 2.00

**Funding Support:**

Project Contribution:

National Academy Member: N

**Participant Type:** Graduate Student (research assistant)

**Participant:** Maria-Paola Conte

**Person Months Worked:** 6.00

**Funding Support:**

Project Contribution:

National Academy Member: N

**Participant Type:** Graduate Student (research assistant)

**Participant:** Xiangyu Liu

**Person Months Worked:** 15.00

**Funding Support:**

Project Contribution:

National Academy Member: N

**Participant Type:** Graduate Student (research assistant)

**Participant:** Nadeesha Wijerathne

**Person Months Worked:** 3.00

**Funding Support:**

Project Contribution:

National Academy Member: N

**Participant Type:** Graduate Student (research assistant)

**Participant:** Liyuan Xie

**Person Months Worked:** 3.00

**Funding Support:**

Project Contribution:

National Academy Member: N

**RPPR Final Report**  
as of 15-Oct-2021

**ARTICLES:**

**Publication Type:** Journal Article      Peer Reviewed: Y      **Publication Status:** 1-Published

**Journal:** Nanotechnology

Publication Identifier Type: DOI

Publication Identifier: 10.1088/0957-4484/27/31/315302

Volume: 27

Issue: 31

First Page #: 315302

Date Submitted: 8/30/18 12:00AM

Date Published: 8/1/16 4:00PM

Publication Location:

**Article Title:** Thermochemical scanning probe lithography of protein gradients at the nanoscale

**Authors:** E Albisetti, K M Carroll, X Lu, J E Curtis, D Petti, R Bertacco, E Riedo

**Keywords:** scanning probe lithography, biofunctionalization, nanopatterning, protein gradient, extracellular matrix, surface functionalization

**Abstract:** Patterning nanoscale protein gradients is crucial for studying a variety of cellular processes in vitro. Despite the recent development in nano-fabrication technology, combining nanometric resolution and the control of protein concentrations is still an open challenge. Here, we demonstrate the use of thermochemical scanning probe lithography (tc-SPL) for defining micro- and nano-sized patterns with precisely controlled protein concentration. First, tc-SPL is performed by scanning a heatable atomic force microscopy tip on a polymeric substrate, locally exposing reactive amino groups on the surface, then the substrate is functionalized with streptavidin and laminin proteins. We show, by fluorescence microscopy on the patterned gradients, that it is possible to precisely tune the concentration of the immobilized proteins by varying the patterning parameters during tc-SPL. This paves the way to the use of tc-SPL for defining protein gradients at the nanoscale, to be used as chemical cues e.g. for stem cell differentiation.

**Distribution Statement:** 3-Distribution authorized to U.S. Government Agencies and their contractors

Acknowledged Federal Support: Y

**Publication Type:** Journal Article      Peer Reviewed: Y      **Publication Status:** 1-Published

**Journal:** ACS Applied Materials & Interfaces

Publication Identifier Type: DOI

Publication Identifier: 10.1021/acsami.6b13162

Volume: 9

Issue: 4

First Page #: 3266

Date Submitted: 8/21/18 12:00AM

Date Published: 1/1/17 8:00PM

Publication Location:

**Article Title:** Biocatalytic Self-Assembly Using Reversible and Irreversible Enzyme Immobilization

**Authors:** M. P. Conte, K. H. A. Lau, R. V. Ulijn

**Keywords:** bioinspired materials; biointerfaces; hydrogel; polydopamine; polyphenol; protein; supramolecular chemistry; surface functionalization

**Abstract:** Biocatalytic control of molecular self-assembly provides an effective approach for developing smart biomaterials, allowing versatile enzyme-mediated tuning of material structure and properties as well as enabling biomedical applications. We functionalized surfaces with bioinspired polydopamine and polyphenol coatings to study the effects of enzyme surface localization and surface release on the self-assembly process. We show how these coatings could be conveniently used to release enzymes for bulk gelation as well as to irreversibly immobilize enzymes for localizing the self-assembly to the surface. The results provide insights to the mode of action of biocatalytic self-assembly relevant to nanofabrication and enzyme-responsive materials.

**Distribution Statement:** 3-Distribution authorized to U.S. Government Agencies and their contractors

Acknowledged Federal Support: Y

## RPPR Final Report as of 15-Oct-2021

**Publication Type:** Journal Article      Peer Reviewed: Y      **Publication Status:** 1-Published

**Journal:** Nanoscale

Publication Identifier Type: DOI

Publication Identifier: 10.1039/C7NR04624C

Volume:      Issue:

First Page #:

Date Submitted: 8/30/18 12:00AM

Date Published:

Publication Location:

**Article Title:** Pathway-dependent gold nanoparticle formation by biocatalytic self-assembly

**Authors:** Jugal Kishore Sahoo, Sangita Roy, Nadeem Javid, Krystyna Duncan, Lynsey Aitken, Rein V. Ulijn

**Keywords:** biocatalytic, self-assembly, nanoparticles, template, dynamic

**Abstract:** We report on the use of non-equilibrium biocatalytic self-assembly and gelation to guide the reductive synthesis of gold nanoparticles. We show that biocatalytic rates simultaneously dictate supramolecular order and presentation of reductive phenols which in turn results in size control of nanoparticles that are formed.

**Distribution Statement:** 3-Distribution authorized to U.S. Government Agencies and their contractors

Acknowledged Federal Support: Y

**Publication Type:** Journal Article      Peer Reviewed: Y      **Publication Status:** 1-Published

**Journal:** Nature Chemistry

Publication Identifier Type: DOI

Publication Identifier: 10.1038/s41557-018-0047-2

Volume: 10

Issue: 7

First Page #: 696

Date Submitted: 8/21/18 12:00AM

Date Published: 4/1/18 4:00AM

Publication Location:

**Article Title:** Amino-acid-encoded biocatalytic self-assembly enables the formation of transient conducting nanostructures

**Authors:** Mohit Kumar, Nicole L. Ing, Vishal Narang, Nadeesha K. Wijerathne, Allon I. Hochbaum, Rein V. Ulijn

**Keywords:** peptide, supramolecular materials, biocatalytic self-assembly

**Abstract:** Aqueous compatible supramolecular materials hold promise for applications in environmental remediation, energy harvesting and biomedicine. One remaining challenge is to actively select a target structure from a multitude of possible options, in response to chemical signals, while maintaining constant, physiological conditions. Here, we demonstrate the use of amino acids to actively decorate a self-assembling core molecule in situ, thereby controlling its amphiphilicity and consequent mode of assembly. The core molecule is the organic semiconductor naphthalene diimide, functionalized with D- and L- tyrosine methyl esters as competing reactive sites. In the presence of  $\beta$ -chymotrypsin and a selected encoding amino acid, kinetic competition between ester hydrolysis and amidation results in covalent or non-covalent amino acid incorporation, and variable supramolecular self-assembly pathways. Taking advantage of the semiconducting nature of the naphthalene diimide core, electronic wires could

**Distribution Statement:** 3-Distribution authorized to U.S. Government Agencies and their contractors

Acknowledged Federal Support: Y

## RPPR Final Report as of 15-Oct-2021

**Publication Type:** Journal Article      Peer Reviewed: Y      **Publication Status:** 1-Published

**Journal:** Communication Physics of Nature Research

Publication Identifier Type:      Publication Identifier:

Volume:      Issue:      First Page #:

Date Submitted: 6/5/21 12:00AM      Date Published:

Publication Location:

**Article Title:** Nanoscale spin-wave circuits based on engineered reconfigurable spin-textures

**Authors:** Edoardo Albisetti, Daniela Petti, Giacomo Sala, Raffaele Silvani, Silvia Tacchi, Simone Finizio, Sebastia

**Keywords:** reconfigurable spin-textures, spin-wave, Nanoscale

**Abstract:** The development of a scalable and cost-effective nanofabrication method is of key importance for future advances in nanoelectronics. Thermalscanning probe lithography (t-SPL) is a growing nanopatterning method with potential for parallelization, offering unique capabilities that make it an attractive candidate for industrial nanomanufacturing. Here, we demonstrate the possibility to apply t-SPL for the fabrication of graphene devices. In particular, we use t-SPL to produce high performing graphene-based field effect transistors (FETs). The here described t-SPL process includes the fabrication of high-quality metal contacts, as well as patterning and etching of graphene to define the active region of the device. The electrical measurements on the t-SPL fabricated FETs indicate a symmetric conductance at the Dirac point and a low specific contact resistance without the use of any contact engineering strategy.

**Distribution Statement:** 1-Approved for public release; distribution is unlimited.

Acknowledged Federal Support: Y

**Publication Type:** Journal Article      Peer Reviewed: Y      **Publication Status:** 1-Published

**Journal:** ACS Applied Materials & Interfaces

Publication Identifier Type: DOI      Publication Identifier: 10.1021/acsami.7b15456

Volume: 10      Issue: 3      First Page #: 3069

Date Submitted: 11/14/18 12:00AM      Date Published: 1/1/18 10:00AM

Publication Location:

**Article Title:** Biocatalytic Self-Assembly on Magnetic Nanoparticles

**Authors:** Maria P. Conte, Jugal Kishore Sahoo, Yousef M. Abul-Haija, K. H. Aaron Lau, Rein V. Ulijn

**Keywords:** biocatalysts, molecular self-assembly, magnetic nanoparticles

**Abstract:** Combining (bio)catalysis and molecular self-assembly provides an effective approach for the production and processing of self-assembled materials by exploiting catalysis to direct the assembly kinetics and hence controlling the formation of ordered nanostructures. Applications of (bio)catalytic self-assembly in biologically interfacing systems and in nanofabrication have recently been reported. Inspired by self-assembly in biological cells, efforts to confine catalysts on flat or patterned surfaces to exert spatial control over molecular gelator generation and nanostructure self-assembly have also emerged. Building on our previous work in the area, we demonstrate in this report the use of enzymes immobilized onto magnetic nanoparticles (NPs) to spatially localize the initiation of peptide self-assembly into nanofibers around NPs. The concept is generalized for both an equilibrium biocatalytic system that forms stable hydrogels and a nonequilibrium system that normally has a preset life

**Distribution Statement:** 3-Distribution authorized to U.S. Government Agencies and their contractors

Acknowledged Federal Support: Y



## RPPR Final Report as of 15-Oct-2021

**Publication Type:** Journal Article      Peer Reviewed: Y      **Publication Status:** 1-Published

**Journal:** Angewandte Chemie International Edition

Publication Identifier Type: DOI

Publication Identifier: 10.1002/anie.201708036

Volume: 56

Issue: 46

First Page #: 14511

Date Submitted: 11/14/18 12:00AM

Date Published: 11/1/17 4:00AM

Publication Location:

**Article Title:** Switchable Hydrolase Based on Reversible Formation of Supramolecular Catalytic Site Using a Self-Assembling Peptide

**Authors:** Chunqiu Zhang, Ramim Shafi, Ayala Lampel, Douglas MacPherson, Charalampos G. Pappas, Vishal N:

**Keywords:** peptide catalysis stimuli-responsive

**Abstract:** The reversible regulation of catalytic activity is a feature found in natural enzymes which is not commonly observed in artificial catalytic systems. Here, we fabricate an artificial hydrolase with pH-switchable activity, achieved by introducing a catalytic histidine residue at the terminus of a pH-responsive peptide. The peptide exhibits a conformational transition from random coil to  $\beta$ -sheet by changing the pH from acidic to alkaline. The  $\beta$ -sheet self-assembles to form long fibrils with the hydrophobic edge and histidine residues extending in an ordered array as the catalytic microenvironment, which shows significant esterase activity. Catalytic activity can be reversibly switched by pH-induced assembly/disassembly of the fibrils into random coils. At higher concentrations, the peptide forms a hydrogel which is also catalytically active and maintains its reversible (de-)activation.

**Distribution Statement:** 3-Distribution authorized to U.S. Government Agencies and their contractors

Acknowledged Federal Support: Y

**Publication Type:** Journal Article

Peer Reviewed: Y

**Publication Status:** 1-Published

**Journal:** Langmuir

Publication Identifier Type: DOI

Publication Identifier: 10.1021/acs.langmuir.6b03471

Volume: 32

Issue: 51

First Page #: 13600

Date Submitted: 11/14/18 12:00AM

Date Published: 12/1/16 10:00AM

Publication Location:

**Article Title:** Understanding How Charged Nanoparticles Electrostatically Assemble and Distribute in 1-D

**Authors:** Keith M. Carroll, Heiko Wolf, Armin Knoll, Jennifer E. Curtis, Yadong Zhang, Seth R. Marder, Elisa Riedl

**Keywords:** Nanoparticles

**Abstract:** The effects of increasing the driving forces for a 1-D assembly of nanoparticles onto a surface are investigated with experimental results and models. Modifications, which take into account not only the particle-particle interactions but also particle-surface interactions, to previously established extended random sequential adsorption simulations are tested and verified. Both data and model are compared against the heterogeneous random sequential adsorption simulations, and finally, a connection between the two models is suggested. The experiments and models show that increasing the particle-surface interaction leads to narrower particle distribution; this narrowing is attributed to the surface interactions compensating against the particle-particle interactions. The long-term advantage of this work is that the assembly of nanoparticles in solution is now understood as controlled not only by particle-particle interactions but also by particle-surface interactions. Both particle-partic

**Distribution Statement:** 3-Distribution authorized to U.S. Government Agencies and their contractors

Acknowledged Federal Support: Y

## RPPR Final Report as of 15-Oct-2021

**Publication Type:** Journal Article      Peer Reviewed: Y      **Publication Status:** 1-Published

**Journal:** Applied Physics Letters

Publication Identifier Type: DOI

Publication Identifier: 10.1063/1.5047222

Volume: 113      Issue: 16

First Page #: 162401

Date Submitted: 8/30/19 12:00AM

Date Published: 10/1/18 4:00AM

Publication Location:

**Article Title:** Stabilization and control of topological magnetic solitons via magnetic exchange bias systems      nanopatterning of

**Authors:** Edoardo Albisetti, Annalisa Calò, Martin Spieser, Armin W. Knoll, Elisa Riedo, Daniela Petti

**Keywords:** topological spin textures, nano devices

**Abstract:** Stabilizing and manipulating topological magnetic quasiparticles in thin films is of great interest for potential applications in data storage and information processing. Here, we present a strategy for stabilizing magnetic vortices and Bloch lines with controlled position, vorticity, and chirality in a continuous exchange bias system. By tailoring vectorially the unidirectional anisotropy of the system at the nanoscale, via thermally assisted magnetic scanning probe lithography, we show experimentally and via micromagnetic simulations the non-volatile creation of vortex-antivortex pairs. In addition, we demonstrate the deterministic stabilization of cross and circular Bloch lines within patterned Néel magnetic domain walls. This work enables the implementation of complex functionalities based on the control of tailored topological spin-textures in spintronic and magnonic nanodevices.

**Distribution Statement:** 2-Distribution Limited to U.S. Government agencies only; report contains proprietary info  
**Acknowledged Federal Support:** Y

**Publication Type:** Journal Article      Peer Reviewed: Y      **Publication Status:** 1-Published

**Journal:** Nature Electronics

Publication Identifier Type: DOI

Publication Identifier: <https://doi.org/10.1038/s41928-018-0191-0>

Volume: 2      Issue:

First Page #: 17

Date Submitted: 8/30/19 12:00AM

Date Published: 1/1/19 5:00AM

Publication Location:

**Article Title:** Patterning metal contacts on monolayer MoS<sub>2</sub> with vanishing Schottky barriers using thermal nano lithography

**Authors:** X. Zheng, A. Calò, E. Albisetti, X. Liu, A.S.M. Alharbi, G. Arefe, X.i Liu, M. Spieser, W.J. Yoo, T. Taniguchi

**Keywords:** 2D materials, nano lithography

**Abstract:** Two-dimensional semiconductors, such as molybdenum disulfide (MoS<sub>2</sub>), exhibit a variety of properties that could be useful in the development of novel electronic devices. However, nanopatterning metal electrodes on such atomic layers, which is typically achieved using electron beam lithography, is currently problematic, leading to non-ohmic contacts and high Schottky barriers. Here, we show that thermal scanning probe lithography can be used to pattern metal electrodes with high reproducibility, sub-10-nm resolution, and high throughput (105 nm<sup>2</sup> h<sup>-1</sup> per single probe). The approach, which offers simultaneous in situ imaging and patterning, does not require a vacuum, high energy, or charged beams, in contrast to electron beam lithography. Using this technique, we pattern metal electrodes in direct contact with monolayer MoS<sub>2</sub> for top-gate and back-gate field-effect transistors.

**Distribution Statement:** 2-Distribution Limited to U.S. Government agencies only; report contains proprietary info  
**Acknowledged Federal Support:** Y

## RPPR Final Report

as of 15-Oct-2021

**Publication Type:** Journal Article

Peer Reviewed: Y

**Publication Status:** 1-Published

**Journal:** Advanced Functional Materials

Publication Identifier Type: DOI

Publication Identifier: 10.1002/adfm.202008662

Volume: 31

Issue: 19

First Page #: 2008662

Date Submitted: 6/5/21 12:00AM

Date Published: 2/1/21 5:00AM

Publication Location:

**Article Title:** Cost and Time Effective Lithography of Reusable Millimeter Size Bone Tissue Replicas With Sub? 15 nm Feature Size on A Biocompatible Polymer

**Authors:** Xiangyu Liu, Alessandra Zanut, Martina Sladkova?Faure, Liyuan Xie, Marcus Weck, Xiaorui Zheng, Elis

**Keywords:** bone tissue regeneration scanning probe lithography

**Abstract:** The development of a scalable and cost-effective nanofabrication method is of key importance for future advances in nanoelectronics. Thermal scanning probe lithography (t-SPL) is a growing nanopatterning method with potential for parallelization, offering unique capabilities that make it an attractive candidate for industrial nanomanufacturing. Here, we demonstrate the possibility to apply t-SPL for the fabrication of graphene devices. In particular, we use t-SPL to produce high performing graphene-based field effect transistors (FETs). The here described t-SPL process includes the fabrication of high-quality metal contacts, as well as patterning and etching of graphene to define the active region of the device. The electrical measurements on the t-SPL fabricated FETs indicate a symmetric conductance at the Dirac point and a low specific contact resistance without the use of any contact engineering strategy.

**Distribution Statement:** 1-Approved for public release; distribution is unlimited.

Acknowledged Federal Support: Y

**Publication Type:** Journal Article

Peer Reviewed: Y

**Publication Status:** 1-Published

**Journal:** APL Materials

Publication Identifier Type: DOI

Publication Identifier: 10.1063/5.0026159

Volume: 9

Issue: 1

First Page #: 011107

Date Submitted: 6/5/21 12:00AM

Date Published: 1/1/21 5:00AM

Publication Location:

**Article Title:** Nanofabrication of graphene field-effect transistors by thermal scanning probe lithography

**Authors:** Xiangyu Liu, Zhujun Huang, Xiaorui Zheng, Davood Shahrjerdi, Elisa Riedo

**Keywords:** nanofabrication graphene field-effect transistor scanning probe lithography

**Abstract:** The development of a scalable and cost-effective nanofabrication method is of key importance for future advances in nanoelectronics. Thermal scanning probe lithography (t-SPL) is a growing nanopatterning method with potential for parallelization, offering unique capabilities that make it an attractive candidate for industrial nanomanufacturing. Here, we demonstrate the possibility to apply t-SPL for the fabrication of graphene devices. In particular, we use t-SPL to produce high performing graphene-based field effect transistors (FETs). The here described t-SPL process includes the fabrication of high-quality metal contacts, as well as patterning and etching of graphene to define the active region of the device. The electrical measurements on the t-SPL fabricated FETs indicate a symmetric conductance at the Dirac point and a low specific contact resistance without the use of any contact engineering strategy.

**Distribution Statement:** 1-Approved for public release; distribution is unlimited.

Acknowledged Federal Support: Y

## RPPR Final Report as of 15-Oct-2021

**Publication Type:** Journal Article

Peer Reviewed: Y

**Publication Status:** 1-Published

**Journal:** ACS Nano

Publication Identifier Type: DOI

Publication Identifier: 10.1021/acsnano.0c05029

Volume: 14

Issue: 11

First Page #: 15056

Date Submitted: 6/5/21 12:00AM

Date Published: 11/1/20 4:00AM

Publication Location:

**Article Title:** In Situ, Noncovalent Labeling and Stimulated Emission Depletion-Based Super-Resolution Imaging of Supramolecular Peptide Nanostructures

**Authors:** Mohit Kumar, Jiye Son, Richard H. Huang, Deborah Sementa, Magdelene Lee, Stephen O'Brien, Rein V.

**Keywords:** Nanostructures peptides STED superresolution microscopy

**Abstract:** The development of a scalable and cost-effective nanofabrication method is of key importance for future advances in nanoelectronics. Thermalscanning probe lithography (t-SPL) is a growing nanopatterning method with potential for parallelization, offering unique capabilities that make it an attractive candidate for industrial nanomanufacturing. Here, we demonstrate the possibility to apply t-SPL for the fabrication of graphene devices. In particular, we use t-SPL to produce high performing graphene-based field effect transistors (FETs). The here described t-SPL process includes the fabrication of high-quality metal contacts, as well as patterning and etching of graphene to define the active region of the device. The electrical measurements on the t-SPL fabricated FETs indicate a symmetric conductance at the Dirac point and a low specific contact resistance without the use of any contact engineering strategy.

**Distribution Statement:** 1-Approved for public release; distribution is unlimited.

Acknowledged Federal Support: Y

**Publication Type:** Journal Article

Peer Reviewed: Y

**Publication Status:** 1-Published

**Journal:** Chemical Science

Publication Identifier Type: DOI

Publication Identifier: 10.1039/D0SC00954G

Volume: 11

Issue: 14

First Page #: 3737

Date Submitted: 6/5/21 12:00AM

Date Published:

Publication Location:

**Article Title:** Inhibiting cancer metabolism by aromatic carbohydrate amphiphiles that act as antagonists of the glucose transporter GLUT1

**Authors:** Alexandra Brito, Patrícia M. R. Pereira, Diana Soares da Costa, Rui L. Reis, Rein V. Ulijn, Jason S. Lew

**Keywords:** aromatic peptide Amphiphiles antagonist self-assembly cancer

**Abstract:** The development of a scalable and cost-effective nanofabrication method is of key importance for future advances in nanoelectronics. Thermalscanning probe lithography (t-SPL) is a growing nanopatterning method with potential for parallelization, offering unique capabilities that make it an attractive candidate for industrial nanomanufacturing. Here, we demonstrate the possibility to apply t-SPL for the fabrication of graphene devices. In particular, we use t-SPL to produce high performing graphene-based field effect transistors (FETs). The here described t-SPL process includes the fabrication of high-quality metal contacts, as well as patterning and etching of graphene to define the active region of the device. The electrical measurements on the t-SPL fabricated FETs indicate a symmetric conductance at the Dirac point and a low specific contact resistance without the use of any contact engineering strategy.

**Distribution Statement:** 1-Approved for public release; distribution is unlimited.

Acknowledged Federal Support: Y

### WEBSITES:

**URL:** ulijnlab.com

Date Received: 21-Aug-2018

**Title:** ulijn group website

**Description:** research group website

# **RPPR Final Report**

as of 15-Oct-2021

## **Partners**

,

Dr. Giuseppe De Peppo, New York Stem Cell foundation (New York)Dr. Ricardo Pires, University of Minho (Portuga

I certify that the information in the report is complete and accurate:

Signature: rein Ulijn

Signature Date: 6/5/21 12:31PM

## Major accomplishments

### Summary

The objective of this collaborative grant between the labs of Rein Ulijn (CUNY) and Elisa Riedo (NYU) was to combine revolutionary nanoscale patterning with catalytic self-assembly to obtain spatio-temporal control of nanostructure formation and breakdown and consequent dynamic control of functionality. By using thermochemical scanning probe lithography (tSPL) we were able to precisely tune the film 3D topographical landscape simultaneously with surface chemistry. This provided control over positioning and concentration of surface-immobilized enzymes [1,2]. Furthermore, by making use of competing enzyme-catalyzed assembly and dis-assembly reactions, we could demonstrate the transient formation of nanoscale fibers that have programmable chemical content and (electronic) functionality, as well as time programmable assembly - disassembly behavior [3-8]. We studied fundamental aspects of biocatalytic assembly at solid-liquid interfaces by using both synthetic and biological surfaces and particles [9-11]. Finally, we developed a versatile and modular chemical method to incorporate fluorescent dyes into peptide nanostructures, which allowed us to visualize in real-time the formation and breakdown of nanostructures with super-resolution (STED) fluorescent microscopy [12]. More recently, we demonstrated the possibility to apply tSPL for the production of high performing 2D materials-based field effect transistors (FETs) fabricated on flexible materials without the need of masks, or markers, and in ambient conditions [13, 14]. Also, we have developed innovative methodologies for the time- and cost-effective scaling up of tSPL in combination with a biocompatible polymer, and we have replicated the exact topography of the bone tissue microenvironment with unprecedented sub-15 nm resolution over millimeter scale areas creating new possibilities for biomedical research and applications [15]. The grant partially supported one postdoctoral research fellow, and one graduate student.

### Significant results up to August 2020-January 2021:

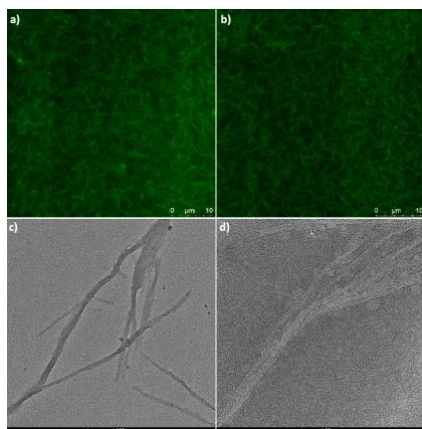
The final five months of the project were focused on completing and submitting manuscripts. Specifically, reviewers requested demonstration of the generality of the STED labeling approach and we could demonstrate labelling of arginine peptides (Table 1, entry 1), as well as nanoparticles functionalized with cationic peptides (entries 2-5) and correlate these with measured zeta potentials, as summarized in the following section. The paper was resubmitted and published in November 2020.

**Table 1:** Peptide sequences tested to expand the scope of Alexa-488 labeling of peptide nanostructures.

structure	sequence	Zeta potential (mV)	STED SRM imaging
1	FFALGLAGRR	10.3	yes
2	KPKGLRGDC-Au	18.1	yes
3	KKPKGLRGDC-Au	24.6	yes
4	KKKPKGLRGDC-Au	28.7	yes

Having demonstrated the nonspecific electrostatic interactions between the negatively charged sulfonate of Alexa-488 and positively charged lysines of peptides, we investigated the generality of the approach with a number of additional positively charged peptide nanostructures. We investigated if another positively charged

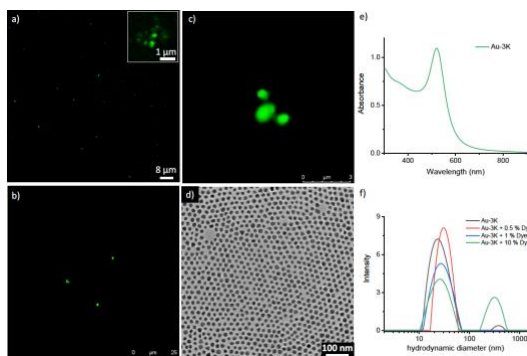
amino acid, arginine (R)-containing peptide nanostructures could be Alexa-488 labeled and subsequently imaged. Thus, we investigated the self-assembly of peptide **1**, which is analogous to the previously studied lysine peptide,



where the positive charge is provided by arginine (R) instead of lysine (K). We observe the formation of one-dimensional nanofibers that could also be labeled by Alexa-488 (Figure 1). Thus, we demonstrated the versatility of Alexa-488-based electrostatic fluorescent labeling of positively charged peptides containing lysine or arginine.

**Figure 1:** (a-b) STED SRM of peptide **1** labeled with Alexa-488 (0.5 mole % dye) and (c-d) TEM micrographs of peptide **1** without dye showing nanofiber network which could be imaged with STED SRM. Condition: 2.5 mM of **1** in 10 mM aq. phosphate buffer pH=7.4.

To further demonstrate the general utility of our approach, we tested gold nanoparticles functionalized with cationic peptide ligands, as a relevant model system for therapeutic and diagnostic applications. We synthesized **Au-1K**, **Au-2K**, and **Au-3K** (zeta potential 18.1, 24.6, and 28.7 mV, respectively) containing increasing numbers of positively charged lysines. Interestingly, all of them could be visualized by our design of electrostatic labeling with Alexa-488 (Figure 2). We observed that the resolution of STED imaging was reduced for gold nanoparticles (approximately 300 nm) compared to peptide nanofibers, which could be due to partial quenching of the dye on nanoparticles. Indeed, it should be noted that fluorescence imaging of peptides attached to gold nanoparticles is not trivial because of the general tendency of gold nanoparticles to quench the emission of fluorophore covalently conjugated on its surface. Advantageously, although it requires further investigation, we demonstrate that the electrostatic labeling design introduced here enables imaging of peptide-conjugated gold nanoparticles without complete quenching of emission. Thus, we have shown the modular nature of Alexa-488 labeling design, which can be used for fluorescent imaging of gold nanoparticles conjugated with cationic

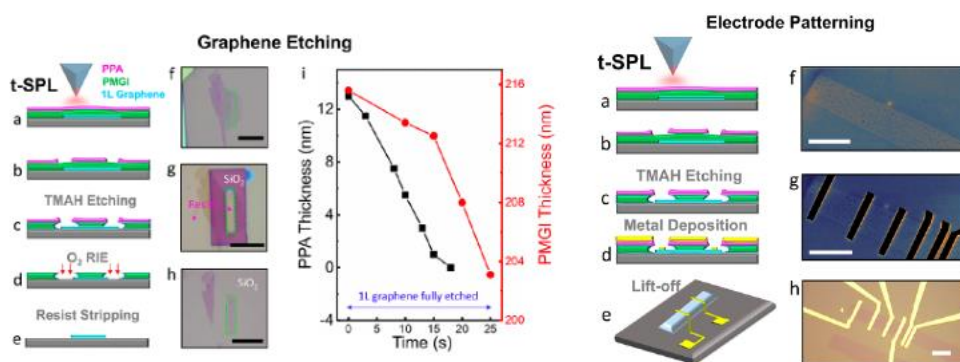


peptides.

**Figure 2:** (a-c) STED microscopy images of peptide functionalized gold nanoparticles (**Au-3K** *i.e.* KKKPKGLRGDC-Au) labeled with Alexa-488 (0.5 mole % with respect to the concentration of ligand present of nanoparticles surface. d) TEM image; e) UV-vis absorption spectrum and f) dynamic light scattering (DLS) data of the **Au-3K** nanoparticles without and with various mole% of dye labeling. In all the cases the solvent medium was 50 mM citrate buffer at pH 7.0.

While our research capabilities were reduced during lockdown, we also wrote a comprehensive review on the topic of *Peptide-Based Systems Chemistry* for ACS Chemical Reviews which covers the new findings gained in this project positioned within the context of the recent literature. This comprehensive review, which includes 350 reference paper has been reviewed and a revision is currently in preparation.

In this funding period, we finalized experiments and published two articles related to the use of tSPL as high-resolution nanofabrication tool. In the first work we demonstrate the possibility to apply thermal scanning probe lithography for the fabrication of graphene devices. In particular, we use tSPL to produce high performing graphene-based field effect transistors (FETs). The here described tSPL process includes the fabrication of high-quality metal contacts, as well as patterning and etching of graphene to define the active region of the device (see Fig. 3).



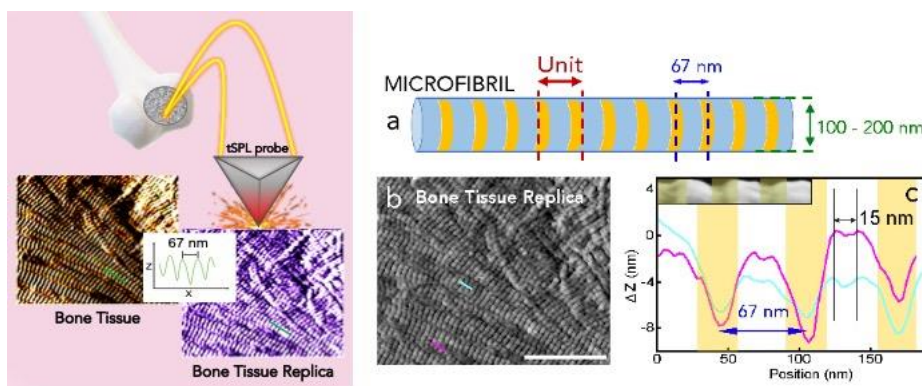
**Figure 3:** Left: Optimization of the t-SPL nano-patterning process for defining graphene regions. (a)–(e) Schematic illustration of the t-SPL process for patterning graphene active regions. (f) Optical microscope image of the starting graphene flake after coating with the PPA/PMGI resist. The green dotted box shows the target active region. (g) The same graphene flake after tSPL patterning and chemical etching of the PMGI layer. (h) Final rectangular graphene ribbon obtained after 25 s oxygen plasma etching. Scale bars are 10  $\mu\text{m}$ . (i) Etching rates of PPA and PMGI with oxygen plasma. Each data point represents one new substrate. Right: tSPL metal electrode patterning on graphene. (a)–(e) Schematic illustrations of the tSPL patterning process for the fabrication of metal electrodes for GFETs. (f) In situ tSPL imaging of monolayer graphene (rectangular ribbon with a length and width of 80  $\mu\text{m}$  and 6.7  $\mu\text{m}$ , respectively) after spin-coating the PPA/PMGI resist. (g) In situ tSPL imaging of the structure, showing the patterned electrode features in the PPA layer. (h) Example of the optical image of a backgated graphene device after lift-off. The spacings between electrodes are 0.6  $\mu\text{m}$ , 2.3  $\mu\text{m}$ , 4.3  $\mu\text{m}$ , 6.1  $\mu\text{m}$ , and 8.2  $\mu\text{m}$ . Scale bars are 10  $\mu\text{m}$ .

In a second work, we use tSPL to replicate, with nanometer resolution, the bone microenvironment in a thermo-sensitive and biocompatible poly(methacrylate) copolymer resist - poly((tetrahydropyran-2-yl N-(2-methacryloxyethyl) carbamate)-b-(methyl 4-(3-methacryloyloxypropoxy) cinnamate)) (PMCC), spin-coated on a transparent indium tin oxide (ITO) glass. To reproduce the bone tissue topography on the PMCC resist, we use an atomic force microscopy (AFM) image of a demineralized bone tissue section as a tSPL bitmap input image (Fig. 4). The results show that the depth (4–8 nm) and the periodic D-spacing (67 nm) of the replicated collagen fibrils are within the typical values reported for the bone tissue microenvironment. The quality of this replica confirms the capability of tSPL to pattern complex 3D tissue microenvironments with high fidelity and resolution (Fig. 4). In order to scale up the size of the replica we reproduce an array of replicas starting from the same bone

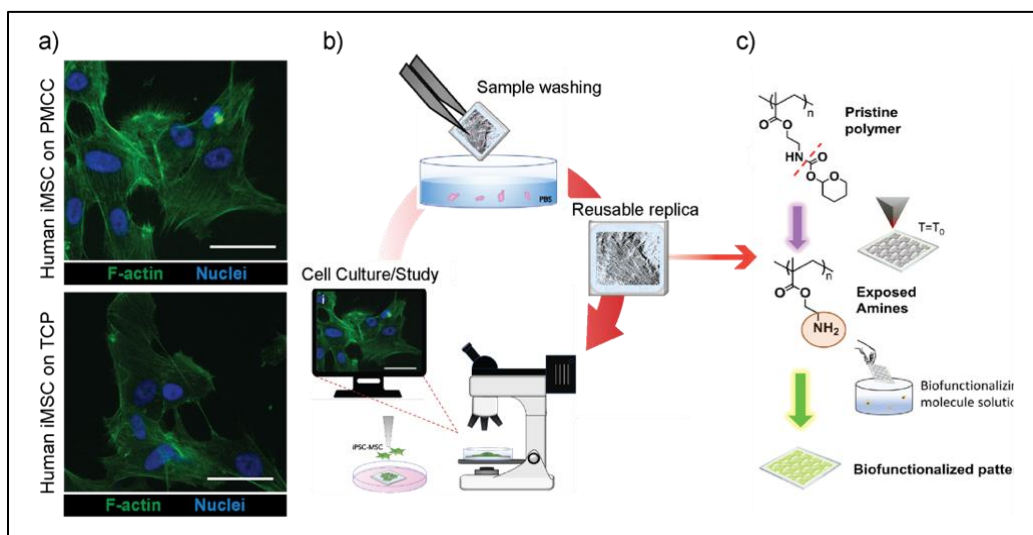


input AFM images. Furthermore, we develop a probe washing procedure, based on cleaning the probe in chloroform.

Very importantly, we find that the PMCC resist supports the culture of human iMSCs similarly to standard tissue culture plasticware (TCP). The nanopatterned replicas are tested for reusability after a cell culture experiment by removing the cells and cleaning the sample, and we obtained some promising results (Fig. 5). The ability to simultaneously pattern topographical and chemical features of biological tissues creates unprecedented opportunities for the production of biomimetic materials and surfaces that control cell behavior.



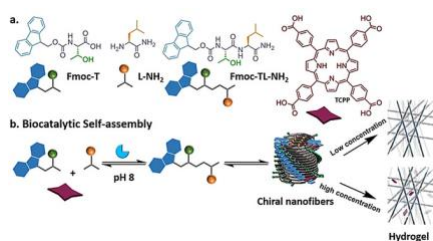
**Figure 4.** Left: AFM images of the bone tissue (bottom left) and the replica fabricated by tSPL (bottom right). Right: image presenting (a) a schematics of a bone microfibril; (b) a bone tissue replica fabricated by tSPL in the PMCC resist. Scale bar: 1  $\mu\text{m}$ ; (c) cross-sections of two segments shown in (b), highlighting the presence of  $\sim 20$ -30 nm gaps and twin-peak fibril bumps, and the characteristic 67 nm distance between the gaps, as in type I collagen microfibrils (in inset).



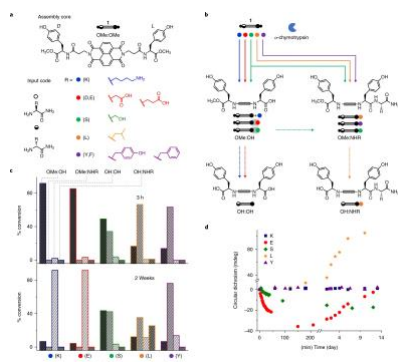
**Figure 5.** (a) Fluorescence micrographs of human iPSC-MP cells attached to PMCC and TCP 1 day after seeding. Scale bar 50  $\mu\text{m}$ ; (b) schematic showing the reusability of the bone replica where Cell cultured on the large-scale bone tissue replicas are removed and the sample is washed and reuse for cell culture studies; (c) Schematic illustration of the tSPL patterning process and pattern biofunctionalization exploiting the PMCC carbamate block chemical structure with protected amine which are exposed by tSPL and used for the biofunctionalization. On the right the fluorescence microscopy image of an array of bone tissue replicas patterned using different probe temperatures and functionalized with Alexa 488 dye. Scale bar: 5  $\mu\text{m}$ .

## Summary of overall accomplishments during the project duration.

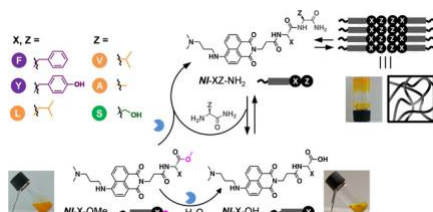
The detailed achievements in each year have been reported in our annual reports. Here we provide an overview of the most significant achievements and publications during the duration of this project.



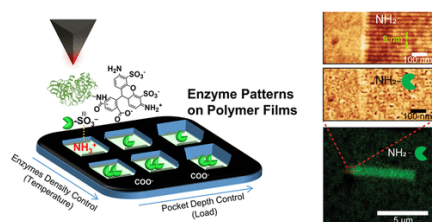
We demonstrated, in a paper published in *Chemistry European Journal*, the biocatalytic co-assembly of peptides and porphyrins in aqueous medium to generate and modulate an energy transfer hydrogel. Depending on the concentrations of porphyrin used, we observed distinct regions of self-assembly behavior: integration of the porphyrin into nanostructures to produce two-component co-assembly fibers, or heterogeneous self-aggregation. The mode of assembly directly impacts on the energy transfer efficiency of these nanostructures.



In a front cover article published in *Nature Chemistry* [7], we demonstrated the use of biocatalytic reactions to covalently or non-covalently modify a self-assembling core molecule, thereby controlling its amphiphilicity and consequent mode of assembly. Taking advantage of the semiconducting nature of the naphthalene diimide core, electronic wires could be formed and subsequently degraded, giving rise to temporally regulated electro-conductivity.

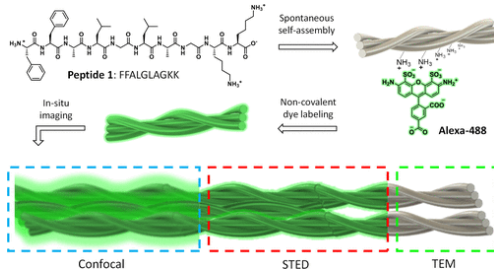


Following from this work, we focused on time-programmable, transient supramolecular naphthalimide-dipeptide nanofibers, as reported in *Chemistry European Journal*, where the lifetimes of which are predictably variable, demonstrated through variation of the self-assembly propensity of their amino acid precursors. The work shows that a thermodynamic parameter dictates kinetic lifetimes in transient fibers formed by competing biocatalytic self-assembly and disassembly of aromatic peptide amphiphiles.

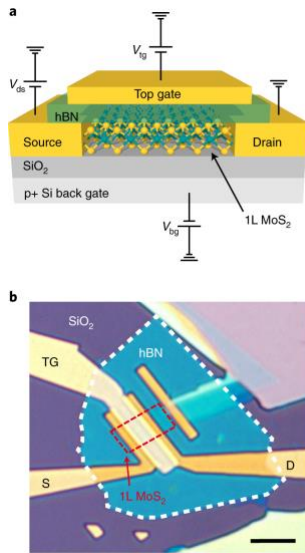


The ability to precisely control the localization of enzymes on a surface is critical for several applications including biosensing, bio-nanoreactors, and single molecule studies. Despite recent advances, fabrication of enzyme patterns with resolution at the single enzyme level is limited by the lack of lithography methods that combine high resolution, compatibility with soft, polymeric structures, ease of fabrication, and high throughput. In papers published in *ACS Advanced Materials Interfaces*, and *Faraday Discussions*, we introduced a method to generate enzyme nanopatterns (using thermolysin as a model system) on a polymer surface is demonstrated using thermochemical scanning probe lithography (tSPL). We were

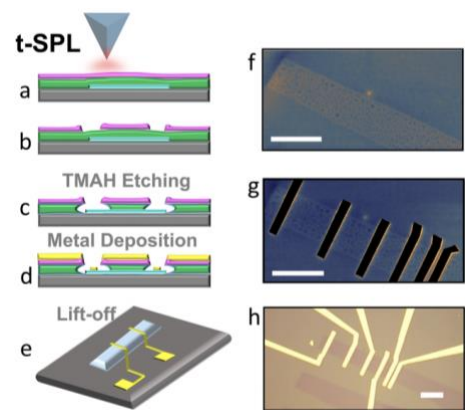
able to achieve combined single-enzyme resolution over  $\text{mm}^2$  areas and the possibility of fabricating enzymes nanogradient.



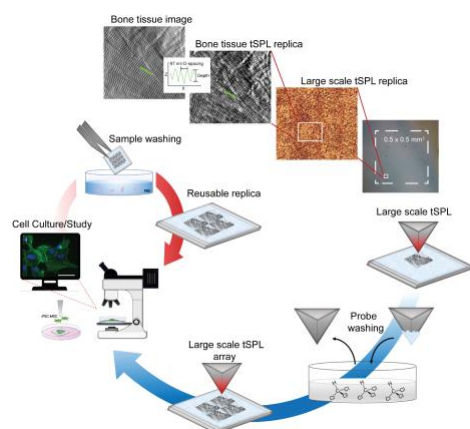
For optimum utilization of these dynamic self-assembled materials, it is important to visualize and understand their structures at the nanoscale, in solution and in real time. We demonstrate a noncovalent fluorescent labeling design for STED-based super-resolution imaging of self-assembling peptides. This is achieved by in situ, electrostatic binding of anionic sulfonates of Alexa-488 dye to the cationic sites of lysine (or arginine) residues exposed on the peptide nanostructure surface. Overall, our approach presents a general and simple method for the electrostatic fluorescent labeling of cationic peptide nanostructures for nanoscale imaging under physiological conditions and probe dynamic processes in real time and in situ.



Two-dimensional semiconductors, such as molybdenum disulfide (MoS<sub>2</sub>), exhibit a variety of properties that could be useful in the development of novel electronic devices. In this paper published on *Nature Electronics* [13], we show that thermal scanning probe lithography can be used to pattern metal electrodes with high reproducibility, sub-10 nm resolution, and high throughput. The approach, which offers simultaneous in situ imaging and patterning, does not require a vacuum, high energy, or charged beams, in contrast to electron beam lithography. Using this technique, we pattern metal electrodes in direct contact with monolayer MoS<sub>2</sub> for top-gate and back-gate field-effect transistors. These devices exhibit exceptional performances, including vanishing Schottky barrier heights (around 0 meV), on/off ratios of  $10^{10}$ , no hysteresis, and subthreshold swings as low as 64 mV per decade without using negative capacitors or hetero-stacks.



As reported in *APL Materials* [14] we demonstrate the possibility to apply thermal scanning probe lithography (tSPL) for the fabrication of graphene devices. In particular, we use tSPL to produce high performing graphene-based field effect transistors (FETs). The here described tSPL process includes the fabrication of high-quality metal contacts, as well as patterning and etching of graphene to define the active region of the device. The electrical measurements on the tSPL fabricated FETs indicate a symmetric conductance at the Dirac point and a low specific contact resistance without the use of any contact engineering strategy. The entire tSPL nanofabrication process is performed without the need for masks, and in ambient conditions. Furthermore, thanks to the tSPL in situ simultaneous patterning and imaging capability, no markers are required. These features substantially decrease fabrication time and cost.



In this cover article published in *Advance Functional Materials* [15] we replicated with sub-15 nm resolution the bone tissue structure over large areas, by scaling up and adapting to cell studies thermal scanning probe lithography. By introducing cell-culture compatible reusable materials and novel writing strategies, we increase throughput and reduce cost by orders of magnitude, thus opening up unprecedented possibilities for pioneering new stem cell studies and biomedical applications.

## REFERENCES:

1. X. Liu, M. Kumar, A. Calò, E. Albisetti, X. Zheng, K. Manning, E. Elacqua, M. Weck, R.V. Ulijn, and E. Riedo, Sub-10 nm Resolution Patterning of Pockets for Enzymes Immobilization with Independent Density and Quasi-3D Topography Control, *ACS Appl. Mater. Interfaces*, **2019**, 11, 41780-41790.
2. X. Liu, M. Kumar, A. Calò, E. Albisetti, X. Zheng, K.B. Manning, E. Elacqua, M. Weck, R.V. Ulijn and E. Riedo, High-throughput Protein Nanopatterning, *Faraday Discuss.*, **2019**, 219, 33-43.
3. M. Kumar, D. Sementa, V. Narang, E. Riedo, and R.V. Ulijn, Self-Assembly Propensity Dictates Lifetimes in Transient Naphthalimide-Dipeptide Nanofibers, *Chem. Eur J.*, **2020**, 26, 8372-8376.
4. C.G. Pappas, N.K. Wijerathne, J.K. Sahoo, A. Jain, D. Kroiss, I.R. Sasselli, A.S. Pina, A. Lampel, and R.V. Ulijn, Spontaneous Aminolytic Cyclization and Self-Assembly of Dipeptide Methyl Esters in Water, *ChemSystemsChem*, **2020**, 2, e2000013.

5. R.V. Ulijn, and A. Lampel, Order/Disorder in Protein and Peptide-Based Biomaterials, *Isr. J. Chem.*, **2019**, 59, 1-9.
6. N.K. Wijerathne, M. Kumar, and R.V. Ulijn, Fmoc-dipeptide/Porphyrin Molar Ratio Dictates Energy Transfer Efficiency in Nanostructures Produced by Biocatalytic Coassembly, *Chem. Eur. J.*, **2019**, 25, 11847-11851.
7. M. Kumar, N.L. Ing, V. Narang, N.K. Wijerathne, A.I. Hochbaum and R.V. Ulijn, Amino Acid-Encoded Biocatalytic Self-Assembly Enables the Formation of Transient Conducting Nanostructures, *Nature Chemistry*, **2018**, 10, 696-703. (front cover)
8. C. Zhang, R. Shafi, A. Lampel, D. MacPherson, C.G. Pappas, V. Narang, T. Wang, C. Madarelli and R.V. Ulijn, Switchable Hydrolase Based on Reversible Formation of Supramolecular Catalytic Site Using a Self-Assembling Peptide, *Angew. Chem. Int. Ed.*, **2017**, 56, 14511-14515.
9. A. Brito, P.M.R. Pereira, D. Soares da Costa, R.L. Reis, R.V. Ulijn, J.S. Lewis, R.A. Pires, and I. Pashkuleva, Inhibiting Cancer Metabolism by Aromatic Carbohydrate Amphiphiles that Act as Antagonists of the Glucose Transporter GLUT1, *Chem. Sci.*, **2020**, 11, 3737-3744.
10. M.P. Conte, J.K.Sahoo, Y.M. Abul-Haija, K.H.A. Lau and R.V. Ulijn, Biocatalytic Self-Assembly on Magnetic Nanoparticles, *ACS Appl. Mater. Interfaces*, **2018**, 10, 3069-3075.
11. M.P. Conte, K.H.A. Lau and R.V. Ulijn, Biocatalytic Self-assembly Using Reversible and Irreversible Enzyme Immobilization, *ACS Appl. Mater. Interfaces*, **2017**, 9, 3266.
12. M. Kumar, J. Son, R.H. Huang, D. Sementa, M. Lee, S. O'Brien, R.V. Ulijn, In situ, non-covalent labeling and super-resolution STED imaging of supramolecular peptide nanostructures, *ACS Nano*, **2020**, accepted.
13. X. Zheng, A. Calò, E. Albisetti, X. Liu, A.S.M. Alharbi, G. Arefe, X. Liu, M. Spieser, W.J. Yoo, T. Taniguchi, K. Watanabe, C. Aruta, A. Ciarrocchi, A. Kis, B.S. Lee, M. Lipson, J. Hone, D. Shahrjerdi, E. Riedo, Patterning metal contacts on monolayer MoS<sub>2</sub> with vanishing Schottky barriers using thermal nanolithography. *Nature Electronics*, **2019**, 2(1), 17-25.
14. X. Liu, Z. Huang, X. Zheng, D. Shahrjerdi, E. Riedo, Nanofabrication of graphene field-effect transistors by thermal scanning probe lithography, *APL Materials*, **2021** 9(1), 011107.
15. X. Liu, A. Zanut, M. Sladkova-Faure, L. Xie, M. Weck, X. Zheng, E. Riedo, G.M. de Peppo, Cost and Time Effective Lithography of Reusable Millimeter Size Bone Tissue Replicas With Sub-15 nm Feature Size on A Biocompatible Polymer, *Adv. Func. Materials*, **2021**, 31(19), 2008662. (Cover).

## Honors

Ulijn was awarded a Vannevar Bush Faculty Fellowship from ONR (2021).

Ulijn was elected co-Chair of the editorial board of *ChemSystemsChem* (Wiley) (2020).

Ulijn was elected to **World Science University Faculty** for the World Science Festival's global science scholar program (2020).

Ulijn awarded **Batsheva de Rothschild Fellowship**, personal travel fellowship to support lecture tour and University visits in Israel (Tel Aviv, Weizmann Institute, Ben Gurion, Hebrew University) (2020).

AFOSR DURIP ( Rein Ulijn Co-PI) *Dual Source Single Crystal X-ray Diffractometer for the Characterization of Small Molecules and Materials*, \$399,000 (PI Share-\$0), 7/15/20-7/14/21

Ulijn (in his role chair) received sponsorship from ARO for the Gordon Research Conference on Systems Chemistry (June 2020). The conference was postponed to 2022 due to COVID.

Riedo was invited speaker at the *Faraday Discussions* on "Nanolithography of Biointerfaces" (ARO UK office program managers among the participants, Hollie Pietsch CCDC Atlantic).

Riedo became Editorial Board Member of *Nature 2D Materials and Applications*

Riedo Invited Speaker at the 2021 TMS Meeting

Riedo Invited Speaker at the 2020 Fall MRS Meeting

Riedo Invited Speaker at the Workshop on 2D and Quantum Materials at the 2020 CFN/NSLS-II Brookhaven Meeting

Riedo's work on tSPL selected to appear in the *National Nanotechnology Initiative* (NNI) Supplement to the President's 2020 Budget (which also serves as the annual report to Congress for the NNI).

ARO Materials Science (Elisa Riedo, Single PI) *Novel properties of pressure activated 2D materials*, \$569,887, 08/01/20-07/31/23

Grad Student Naxhije Berisha received Tow Foundation Fellowship.

Post-Doc Zanut received the NYU Provost Fellowship.

Ph.D. Xiangyu Liu won a Conference Award for the World Congress on Micro and Nano Manufacturing.

## **Tech Transfer**

Ulijn: Virtual seminar at Soldier Center, US Army Future Command, Natick MA (May 2020).

Riedo: Provisional Patent Application: "Biomimetic Tissue And Method Of Use Thereof", NYU and NYSCF (2021)

Riedo: new collaboration with Thomas J. Kiel, Armaments Center, CCDC U.S. Army Futures Command Picatinny Arsenal

Riedo: ongoing partnership with SwissLitho and IBM for commercialized tSPL.

Visit by Wendy Mills and Dawanne Poree to CUNY ASRC (October 2018).

Visit by Rajesh Naik (AFoSR) to CUNY ASRC as part of 5-year review (February 2019).

Visit by Thomas J. Kiel, U.S. Army Futures Command Picatinny Arsenal to NYU Tandon Engineering (May 2019)

Riedo participated in a virtual meeting with Dr. Daniel Cole, Dr. Dan Magagnosc, and Dr. Matt Guziewski from ARL to discuss possible interactions and collaborations.

## **Students**

Number of students receiving STEM degrees during the reporting period: 3

Number of undergraduate and graduate STEM participants during the reporting period: 3