NONINVASIVE CARBON DIOXIDE MONITORING IN A PORCINE MODEL OF ACUTE LUNG INJURY DUE TO SMOKE INHALATION AND BURNS

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ABSTRACT In critically ill intubated patients, assessment of adequacy of ventilation relies on measuring partial pressure of arterial carbon dioxide (PaCO₂), which requires invasive arterial blood gas analysis. Alternative noninvasive technologies include transcutaneous CO₂ (tPCO₂) and end-tidal CO₂ (EtCO₂) monitoring. We evaluated accuracy of tPCO₂ and EtCO₂ monitoring in a porcine model of acute lung injury (ALI) due to smoke inhalation and burns. Eight anesthetized Yorkshire pigs underwent mechanical ventilation, wood-bark smoke inhalation injury, and 40% total body surface area thermal injury. tPCO₂ was measured with a SenTec system (SenTec AG, Therwil, Switzerland) and EtCO₂ with a Capnostream-20 (Oridion Medical, Jerusalem, Israel). These values were compared with PaCO₂ measurements from an arterial blood gas analyzer. Paired measurements of EtCO₂-PaCO₂ (n = 276) and tPCO₂-PaCO₂ (n = 250) were recorded in the PaCO₂ range of 25 to 85 mmHg. Overlapping data sets were analyzed based on respiratory and hemodynamic status of animals. Acute lung injury was defined as PaO₂/FIO₂ ≤ 200 mmHg; hemodynamic instability was defined as mean arterial pressure ≤ 60 mmHg. Before ALI, EtCO₂ demonstrated moderate correlation with PaCO₂ (R² = 0.45; P < 0.0001), which deteriorated after onset of ALI (R² = 0.12; P < 0.0001). Before ALI, tPCO₂ demonstrated moderate correlation (R² = 0.51, P < 0.0001), which was sustained after onset of ALI (R² = 0.78; P < 0.0001). During hemodynamic stability, EtCO₂ demonstrated moderate correlation with PaCO₂ (R² = 0.44; P < 0.0001). During hemodynamic instability, EtCO₂ did not correlate with PaCO₂ (R² = 0.03; P = 0.29). tPCO₂ monitoring demonstrated strong correlation with PaCO₂ during hemodynamic stability (R² = 0.80, P < 0.0001), which deteriorated under hemodynamically unstable conditions (R² = 0.39; P < 0.0001). Noninvasive carbon dioxide monitors are acceptable for monitoring trends in PaCO₂ under conditions of hemodynamic and pulmonary stability. Under unstable conditions, reevaluation of patient status and increased caution in the interpretation of results are required.

KEYWORDS Transcutaneous carbon dioxide, end-tidal carbon dioxide, blood gas analysis, acute lung injury, swine, burns, inhalation injury

INTRODUCTION

Measurement of the partial pressure of carbon dioxide in arterial blood (PaCO₂) remains the criterion standard for evaluating the adequacy of ventilation. In certain populations, such as trauma patients with head injury, providing appropriate ventilation (avoiding either overventilation or underventilation) has been shown to save lives (1–2). Obtaining blood gas samples, however, is invasive and requires special equipment, which is often impractical in prehospital settings. Thus, a noninvasive means of estimating PaCO₂ would be useful (3). Several such methods have been developed.

End-tidal carbon dioxide (EtCO₂) monitoring is performed with increasing frequency in injured patients (4,5). However, this method may become inaccurate in unstable trauma patients (6). The PaCO₂-EtCO₂ gradient is related to the physiologic dead space as described by Enghoff modification of the Bohr equation (7). PaCO₂-EtCO₂ gradient increases with increasing physiologic dead space, due to lung injuries or decreased pulmonary perfusion.

Another noninvasive technique that can be used as a surrogate for PaCO₂ is the measurement of the transcutaneous partial pressure of CO₂ (tPCO₂). This technique was pioneered by Severinghaus (8) in the 1960s using a temperature-stabilized heated electrode. Transcutaneous gas tension is a function of dermal capillary blood and in turn of arterial blood flow. Previous studies by Nishiyama et al. (9, 10) demonstrated good correlations between transcutaneous CO₂ measurements and PaCO₂ in adults undergoing general anesthesia. Several studies, performed in critically ill adults (11–14) as well as in adults undergoing noninvasive ventilation (15, 16), reported good correlation between tPCO₂ measurements and PaCO₂. Most notably, a study by Hinkelbein et al. (13) demonstrated the feasibility of tPCO₂ monitoring in critically ill adults during interhospital transport.
Noninvasive Carbon Dioxide Monitoring in a Porcine Model of Acute Lung Injury Due to Smoke Inhalation and Burns.

Belenkiy S., Ivey K. M., Batchinsky A. I., Langer T., Necsoiu C., Baker W., Salinas J., Cancio L. C.,
Nevertheless, there is a paucity of studies comparing EtCO$_2$ and tPCO$_2$ monitoring in settings of evolving acute lung injury (ALI). Therefore, we carried out an animal study to evaluate the usability and accuracy of these methods in a clinically relevant porcine model of ALI secondary to smoke inhalation and burns.

**MATERIALS AND METHODS**

This study was approved by the US Army Institute of Surgical Research Animal Care and Use Committee. It was conducted in compliance with the Animal Welfare Act and the implementing Animal Welfare Regulations and in accordance with the principles of the Guide for the Care and Use of Laboratory Animals.

**Animal preparation**

For this study, we selected a subgroup of eight female, nonpregnant, Yorkshire pigs from an ongoing study that investigates treatment of acute respiratory distress syndrome (ARDS) due to smoke inhalation and burns. Total intravenous anesthesia with ketamine HCL (20-30 mg/kg per hour), midazolam HCL (1.0-1.5 mg/kg per hour), and propofol (100 μg/kg per hour) was used during the experiment. All animals underwent tracheostomy and placement of central and arterial lines. After instrumentation, animals were allowed to recover for 2 to 3 h. After stabilization, they received inhalation injury with 22 breaths of pine bark smoke at room temperature (Table 1). This model of smoke inhalation injury has been described previously (17). Smoke injury was followed immediately by a 40% total body surface area, full thickness flame burn. Upon completion of smoke inhalation and burn, animals were transferred to the animal intensive care unit where they were continuously monitored for the duration of the experiment. Total intravenous anesthesia levels were adjusted to effect as needed to attain no response to painful stimuli. In addition, buprenorphine HCL (0.1 mg/kg) was administered intramuscularly every 6 h for analgesia for the duration of the study. Resuscitation with lactated Ringer’s solution was performed by means of a computerized bum resuscitation decision support system for the first 24 h after burn. Subsequently, the fluid rate was adjusted manually to achieve a urinary output of at least 0.5 mL/kg per hour. Controlled mechanical ventilation (CMV) was initiated, with a tidal volume of 10 mL/kg and a respiratory rate adjusted to maintain PaCO$_2$ between 35 and 45 mmHg. Before onset of ARDS, positive end expiratory pressure was set to 5 cm H$_2$O and peripheral oxygen saturation (SpO$_2$) at or above 90%. Fiberoptic bronchoscopy remained constant. Fraction of inspired oxygen (FIO$_2$) was titrated to achieve (1.0-1.5 mg/kg per hour), and propofol (100 μg/kg per hour) was used during the experiment. All animals underwent tracheostomy and placement of central and arterial lines. After instrumentation, animals were allowed to recover for 2 to 3 h. After stabilization, they received inhalation injury with 22 breaths of pine bark smoke at room temperature (Table 1). This model of smoke inhalation injury has been described previously (17). Smoke injury was followed immediately by a 40% total body surface area, full thickness flame burn. Upon completion of smoke inhalation and burn, animals were transferred to the animal intensive care unit where they were continuously monitored for the duration of the experiment. Total intravenous anesthesia levels were adjusted to effect as needed to attain no response to painful stimuli. 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After the onset of ARDS, animals were transitioned to low tidal volume CMV, and further ventilator changes, including adjustments of positive end expiratory pressure and FiO$_2$, were made in accordance with the ARDSnet protocol (18). The study was designed to continue for up to 7 days after injury. Experiments were terminated earlier if ARDS did not develop in 72 h after injury or if an animal reached terminal cardiopulmonary failure before 7 days. Animals were killed with intravenous injection of 20 mL Fatal Plus (Vortech Pharmaceuticals, Dearborn, Mich). Ratio of the partial pressure of oxygen in arterial blood (PaO$_2$) to the fraction of inspired oxygen (PFR) was maintained at baseline, 2 h after injury, and every 6 h thereafter. A single set of baseline measurements was recorded after surgery but before the induction of smoke inhalation and burns. Baseline duration generally was 2 to 3 h and was adequate to provide stabilization time for all vital signs after surgical preparation. We defined ALI as a sustained PFR less than 300 mmHg, and ARDS as a sustained PFR less than 200 mmHg. As an index of pulmonary instability, we recorded the frequency of changes in the respiratory rate (as set on the ventilator) and in the FiO$_2$. We defined hemo dynamic instability as mean arterial pressure (MAP) of 60 mmHg or less (or a requirement for norepinephrine infusion to maintain MAP >60 mmHg).

**RESULTS**

A total of 454 h of animal intensive care unit care were provided during this study. We recorded 276 paired measurements of Paco$_2$ and EtCO$_2$, and 250 paired measurements of PaCO$_2$ and tPCO$_2$, from eight animals. The average duration of monitoring was 56.8 ± 40.9 h. EtCO$_2$ and tPCO$_2$ measurements were made across a PaCO$_2$ range of 25 to 85 mmHg. Animal data are presented in Table 1.

It is important to highlight that our injury model was fluid rather than static with rapidly declining respiratory status. The period between smoke/burn injury and onset of ALI was marked by pulmonary instability while animals progressed from healthy baseline conditions to ALI over 22.5 ± 10.6 h. The mean number of ventilator changes per hour was greater before onset of ALI (3.07 ± 1.53) than during ALI (1.84 ± 1.68, P = 0.02). Similarly, the average decrease in PFR in the first 24 h after injury was 199; in the second 24-h period, it was 100 (Fig. 1). There was a statistically significant decrease in PFR between baseline and 24 h, as well as between 12 and 24 h (P < 0.05), but not between 24 and 48 h (P > 0.05). During the study, two animals required norepinephrine infusion due to hypotension. During the study, we observed no thermal complications due to the placement of a heated tPCO$_2$ sensor on the auricle.

**TABLE 1. Animal data**

| Weight, kg | 45.0 ± 2.8 |
| Total smoke volume, L | 27.6 ± 4.9 |
| Post-smoke-inhalation arterial COHb, % | 88.3 ± 4.1 |
| Time to ALI, h | 22.5 ± 10.6 |
| Survival time, h | 56.8 ± 40.9 |

COHb indicates carboxyhemoglobin.

**Statistical analysis**

SigmaPlot for Windows version 12.0 (Systat Software, San Jose, Calif) was used. Overlapping sets of data were analyzed based on: (i) respiratory status (before vs. after the onset of ALI) and (ii) hemodynamic status (stable vs. unstable). All results are expressed as mean ± SD. P < 0.05 was considered significant.

Linear regression was performed for pairs of PaCO$_2$ and tPCO$_2$ as well as for pairs of Paco$_2$ and EtCO$_2$ data. Bias and 95% limits of agreement were calculated using Bland Altman analysis (25). Student t tests and analyses of variance with post hoc Holm Sidak multiple comparison tests were used.
Two partially overlapping sets of data were analyzed based on (i) pulmonary status (before vs. after the onset of ALI) and (ii) cardiovascular status (hemodynamically stable vs. unstable). The results of this analysis, describing the EtCO₂-Paco₂ and tPCO₂-Paco₂ relationships, are presented in Table 2 and are described in the following sections.

**Before versus after the onset of ALI**

Linear regression analysis of 105 EtCO₂-Paco₂ pairs, recorded before the onset of ALI (Fig. 2A), demonstrated moderate correlation: Paco₂ = 13.45 + 0.68 × EtCO₂ (R² = 0.45, P < 0.0001). There was systemic underreading by capnography (P = 0.02). Bland-Altman analysis revealed a mean bias of 0.91 ± 3.77 mmHg (Fig. 2B).

Analysis of 171 EtCO₂-Paco₂ pairs, recorded after ALI (Fig. 3A), demonstrated low correlation: Paco₂ = 41.51 + 0.34 × EtCO₂ (R² = 0.12, P < 0.0001). There was systemic underreading by capnography (P < 0.001). Mean bias increased to 14.84 ± 11.76 mmHg (Fig. 3B).

Analysis of 88 tPCO₂-Paco₂ pairs, recorded before the onset of ALI (Fig. 2A), revealed moderate correlation: Paco₂ = 17.01 + 0.56 × tPCO₂ (R² = 0.51, P < 0.0001). No systemic difference was observed (P = 0.85). Mean bias was 0.09 ± 4.56 mmHg (Fig. 2C).

Analysis of 140 tPCO₂-Paco₂ pairs, recorded after the onset of ALI (Fig. 3A), revealed strong correlation: Paco₂ = 7.74 + 0.83 × tPCO₂ (R² = 0.78, P < 0.0001). Slight systemic underreading was present (P = 0.026). Mean bias was 0.03 ± 5.44 mmHg (Fig. 3C).

**Hemodynamically stable versus unstable**

We repeated the analysis based on hemodynamic status. Linear regression of 233 EtCO₂-Paco₂ pairs under hemodynamically stable conditions (Fig. 4A) demonstrated moderate correlation: Paco₂ = 5.32 + 1.03 × EtCO₂ (R² = 0.44, P < 0.0001). There was systemic underreading by capnography. The mean bias was 6.74 ± 8.26 mmHg (Fig. 4B).

Analysis of 41 EtCO₂-Paco₂ pairs under hemodynamically unstable conditions (Fig. 5A) did not demonstrate a linear relationship: Paco₂ = 61.4 - 0.16 × EtCO₂ (R² = 0.03, P = 0.29). Systemic underreading was present (P < 0.001). Mean bias was 25.59 ± 15.30 mmHg (Fig. 5B).

Analysis of 212 tPCO₂-Paco₂ pairs under hemodynamically stable conditions (Fig. 4A) demonstrated strong correlation: Paco₂ = 7.86 + 0.82 × tPCO₂ (R² = 0.80, P < 0.0001). There was no systemic difference between the two techniques (P = 0.29). Mean bias was −0.35 ± 4.75 mmHg (Fig. 4C).

Finally, analysis of 38 tPCO₂-Paco₂ pairs under hemodynamically unstable conditions (Fig. 5A) demonstrated poor correlation: Paco₂ = 33.60 + 0.3 × tPCO₂ (R² = 0.39, P < 0.0001). There was systemic overreading (P < 0.001). Mean bias was −11.32 ± 14.87 mmHg (Fig. 5C).

**DISCUSSION**

In this study, we evaluated the utility of two commonly used noninvasive carbon dioxide monitoring technologies in a severely injured, mechanically ventilated porcine model of ALI due to smoke inhalation and burns. Our principal findings were (i) the period of time between smoke/burn injury and the onset of ALI was characterized by pulmonary instability, manifested by a steady decrease in the PFR and by frequent ventilator changes and other interventions; (ii) both EtCO₂ and tPCO₂ were moderately correlated with Paco₂ during this period; (iii) after

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**Table 2. EtCO₂-Paco₂ and tPCO₂-Paco₂ relationships**

<table>
<thead>
<tr>
<th>n (No. of pairs)</th>
<th>Variable compared with Paco₂</th>
<th>Condition</th>
<th>R²</th>
<th>P</th>
<th>Bias, mean ± SD (95% limits of agreement), mmHg</th>
<th>Student t test, P</th>
</tr>
</thead>
<tbody>
<tr>
<td>105</td>
<td>EtCO₂</td>
<td>Pre-ALI</td>
<td>0.45</td>
<td>&lt;0.0001</td>
<td>0.91 ± 3.77 (6.48 to 8.31)</td>
<td>0.02</td>
</tr>
<tr>
<td>171</td>
<td>EtCO₂</td>
<td>ALI</td>
<td>0.12</td>
<td>&lt;0.0001</td>
<td>14.84 ± 11.76 (8.21 to 37.89)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>88</td>
<td>tPCO₂</td>
<td>Pre-ALI</td>
<td>0.51</td>
<td>&lt;0.0001</td>
<td>0.09 ± 4.56 (8.95 to 9.03)</td>
<td>0.85</td>
</tr>
<tr>
<td>140</td>
<td>tPCO₂</td>
<td>ALI</td>
<td>0.78</td>
<td>&lt;0.0001</td>
<td>0.03 ± 5.44 (10.64 to 10.70)</td>
<td>0.026</td>
</tr>
<tr>
<td>233</td>
<td>EtCO₂</td>
<td>HD stable</td>
<td>0.44</td>
<td>&lt;0.0001</td>
<td>6.74 ± 8.26 (9.45 to 22.95)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>41</td>
<td>EtCO₂</td>
<td>HD unstable</td>
<td>0.03</td>
<td>0.29</td>
<td>25.59 ± 15.30 (4.39 to 55.57)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>212</td>
<td>tPCO₂</td>
<td>HD stable</td>
<td>0.80</td>
<td>&lt;0.0001</td>
<td>0.35 ± 4.75 (9.67 to 8.97)</td>
<td>0.29</td>
</tr>
<tr>
<td>38</td>
<td>tPCO₂</td>
<td>HD unstable</td>
<td>0.39</td>
<td>&lt;0.0001</td>
<td>11.32 ± 14.87 (40.46 to 17.82)</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

HD indicates hemodynamically stable.
the onset of ALI. EtcO2 became relatively inaccurate, whereas tPco2 did not; (iv) hemodynamic instability caused EtcO2 values to lose their linear relationship with Paco2. Based on these observations, we conclude that both methods can be useful in the monitoring of patients with developing severe lung injury, but that caution should be used in the interpretation of results when patients are changing rapidly.

Good correlation between EtcO2 and Paco2 across all ranges of dead space was reported by McSwain et al. (7). Our previous study (26) demonstrated close correlation between EtcO2 and Paco2 in a porcine model of severe chest injury during periods of hemodynamic stability and during settings of hyperventilation and hyperventilation induced by varying tidal volumes and different respiratory rates in healthy swine. At the same time, results were not as promising when accuracy of capnography was evaluated in trauma patients (6). In addition, several studies compared accuracy of end-tidal and tPco2 monitoring in human patients (13, 27, 28), favoring tPco2. In our study, EtcO2 systematically underestimated Paco2. Presence of systemic bias was confirmed by paired Student t test results (Table 2). This is to be expected, particularly in patients with developing ALI, in whom dead space progressively increases, thereby leading to an increase in Paco2-EtcO2 gradient.

There was moderate correlation between tPco2 and Paco2 ($R^2 = 0.51$) before ALI. This correlation became more linear ($R^2 = 0.78$) after onset of ALI. We expected better correlation during pre-ALI stage, and such findings were surprising. We speculate that lower tPco2-Paco2 correlation before onset of ALI was possibly due to more frequent ventilator changes; approximately $3.07 \pm 1.53$ changes per hour in the initial phase of the experiment, compared with $1.84 \pm 1.68$ changes per hour after the onset of ALI. Also, more frequent suctioning was required in the first 24 hr after injury due to copious secretions. Because it can cause alveolar collapse and loss of recruitment, repeated suctioning may have had a destabilizing effect on Paco2 correlation. In addition, the tPco2 sensor was more frequently disconnected and removed from the animals during the first 24 hr of the experiment to accommodate bronchoscopies.

When the tPco2-Paco2 relationship was examined based on hemodynamic status, it was strongly linear ($R^2 = 0.80$) with
minimal bias (−0.35 ± 4.75 mmHg) as long as MAP remained greater than 60 mmHg. Our results confirm those reported previously in human studies (11, 13, 20, 27, 28) that demonstrated similar correlation. tPCO2 had a less linear correlation to Paco2 during hemodynamic instability ($R^2 = 0.39$). In addition, transcutaneous monitoring tended to overestimate Paco2 under these conditions. Previous reporting (29, 30) indicated that vasoactive medications did not have a significant effect on tPCO2 monitoring accuracy. However, our results were different, possibly because of the sensor location on the right auricle. Although the animals were hypotensive, their ears were cold to touch, and there was a visible area of vascular congestion. Preferred locations for tPCO2 monitoring have been reported in humans (9, 10), but no such location has been defined in pigs.

Because we used a subset of animals for this work from a larger study, which pursued treatment of ARDS due to smoke inhalation and burns, the samples we obtained were convenience samples. This limited our ability to further investigate effects of suctioning, ventilator changes, and sensor disconnections on stabilizations/steady state times of the measured variables. Future studies designed specifically to address these limitations may be warranted.

Based on our data, transcutaneous capnometry is an acceptable trend-monitoring tool in settings of lung injury in hemodynamically stable patients and may be useful in a prehospital environment. However, the current generation of tPCO2 sensors has some limitations. First, the tPCO2 sensor requires stabilization time after placement. Several articles (31, 32) recommended a 20-min stabilization time while using the SenTec Monitor. Second, to maintain accuracy, the sensor has to be regularly recalibrated in vitro. Recalibration frequency appears to depend on sensor temperature. At 43.5°C, recalibrations were required every 6 to 8 h. On average, recalibration can be completed in approximately 3 min. Afterward, the sensor is repositioned on a patient and again requires stabilization time. In our experience, about 25 to 30 min was spent on moving, cleaning, recalibrating, replacing, and waiting for the sensor to stabilize. In this study, because of previously described limitations, we could not establish the precise stabilization timing necessary for improved accuracy; further study should be considered to

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**Fig. 4.** A, Scatter plot and linear regression analysis between EtcO2 and PacO2 as well as tPCO2 and PacO2 during hemodynamic stability (HD stable). Solid line represents linear regression between tPCO2 and PacO2. Dashed line represents linear regression between EtcO2 and PacO2. B and C demonstrate Bland Altman analysis between EtcO2, PacO2 and tPCO2, PacO2 pairs; middle solid line indicates the mean difference (bias), and outer solid lines represent limits of agreement (mean ± 1.96 SD) between two methods.

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**Fig. 5.** A, Scatter plot and linear regression analysis between EtcO2 and PacO2 as well as tPCO2 and PacO2 during hemodynamic instability (HD unstable). Solid line represents linear regression between tPCO2 and PacO2. Dashed line represents linear regression between EtcO2 and PacO2. B and C demonstrate Bland Altman analysis between EtcO2, PacO2 and tPCO2, PacO2 pairs; middle solid line indicates the mean difference (bias), and outer solid lines represent limits of agreement (mean ± 1.96 SD) between two methods.
address this issue. Also, we should point out that a prolonged stabilization time may limit utilization of tPCO2 monitors in prehospital environments. Finally, the sensor membrane requires replacement every 42 days and between patients. Currently, several companies are developing next-generation solid-state tPCO2 sensors that may overcome these shortcomings. Given promising results obtained from the use of a transcutaneous CO2 sensor, these new developments will be a welcomed addition to critical care monitoring armamentarium as well as potentially opening new possibilities for servo controlling mechanical ventilators and extracorporeal life support devices.

CONCLUSIONS

In a porcine model of ALI due to smoke inhalation and burns, transcutaneous CO2 monitoring is an acceptable noninvasive surrogate for PACO2 under hemodynamically stable conditions and can be useful as a trend monitoring tool. However, tPCO2 readings should be correlated with PACO2 with increased frequency when a patient’s condition is dynamically changing, for example, during periods of hemodynamic instability. End-tidal CO2 monitoring offers an ease-of-use advantage over the current generation of transcutaneous CO2 monitors. However, EtCO2 readings should be correlated with PACO2 with increased frequency during evolution of lung injury. 

ACKNOWLEDGMENTS

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