

SYNTHESIS OF ^{13}C - AND ^{15}N -LABELED DNAN

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FOREWORD

The synthetic and analytical work described herein helps address a basic understanding of the toxicity and biodegradation potential of DNAN. Using two different isotopically labeled DNAN compounds will allow bioaccumulation factors and biodegradation mechanisms for DNAN, its metabolites, and its end-products to be determined. These data can then be applied to existing fate and effects models to determine the potential environmental risk that new insensitive munitions containing DNAN pose.



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TABLE OF CONTENTS

<i>Heading</i>	<i>Page</i>
FOREWORD	v
TABLE OF CONTENTS	vi
LIST OF FIGURES	vii
LIST OF TABLES	vii
LIST OF ABBREVIATIONS AND ACRONYMS	viii
ABSTRACT	1
INTRODUCTION	1
RESULTS & DISCUSSION	2
CONCLUSIONS	5
EXPERIMENTAL SECTION	5
SUPPORTING INFORMATION	10
ACKNOWLEDGMENTS	31
BIBLIOGRAPHY	31
DISTRIBUTION	32

LIST OF FIGURES

<i>Heading</i>	<i>Page</i>
Figure 1. DNAN, ^{13}C -Labeled DNAN, and ^{15}N -Labeled DNAN	1
Figure 2. Synthesis of DNAN	2
Figure 3. Dinitration of Chlorobenzene	6

LIST OF TABLES

<i>Heading</i>	<i>Page</i>
Table 1. Initial Trial Nitrations of Chlorobenzene	7

LIST OF ABBREVIATIONS AND ACRONYMS

DNAN	2,4-Dinitroanisole
Et ₂ O	Diethyl ether
IM	Insensitive Munitions
IMX-101	Insensitive Munition Explosive 101
IR	Infrared spectroscopy
HRMS	High resolution mass spectroscopy
NMR	Nuclear magnetic resonance spectroscopy
PAX-21	Picatinny Arsenal Explosive 21
R _f	Retention factor
T ₁	Longitudinal relaxation time
T ₂	Transverse relaxation time
T ₂ [*]	Observed transverse relaxation time
TNT	2,4,6-Trinitrotoluene
TLC	Thin layer chromatography

ABSTRACT

Isotopically labeled (^{13}C and ^{15}N) and unlabeled DNAN were synthesized in two steps from chlorobenzene in high purity and regioselectivity with an overall yield of 69% from readily available materials. The syntheses and full characterizations (^1H , ^{13}C , ^{14}N , ^{15}N NMR, IR, and HRMS) of labeled and unlabeled DNAN, as well as its synthetic precursor, 1-chloro-2,4-dinitrobenzene, are discussed. The use of isotopically labeled material and their spectra will enable the toxicity and biodegradation potential of DNAN to be more easily determined.

INTRODUCTION

The use of DNAN (2,4-dinitroanisole) has been undergoing a renaissance over the last decade. Once considered a poor substitute for TNT in terms of performance,¹ the insensitivity of DNAN makes it attractive for use in melt-castable insensitive munitions (IM), such as PAX-21 used in the 60-mm mortar (M720A1) and IMX-101 used in 155-mm artillery munitions (M795).² However, the environmental impact of DNAN at manufacturing facilities and training ranges, i.e., Picatinny Arsenal and Holston Army Ammunition Plant, is unclear. Of primary concern is the bioaccumulation of the highly toxic 2,4-dinitrophenol, a breakdown product readily formed via hydrolysis of DNAN, among other degradation products of interest.

In order to assist the determination of the biological degradation mechanisms and bioaccumulation data of DNAN and its metabolites, we have synthesized two different isotopically labeled DNAN compounds: one compound labeled with a ^{13}C -methoxy group and the other labeled with two ^{15}N -nitro groups (Fig. 1). This labeling strategy will allow the potential transformations of de-methylation, de-nitration, and nitro-reduction to be easily tracked by various mass spectroscopy and nuclear magnetic resonance techniques.

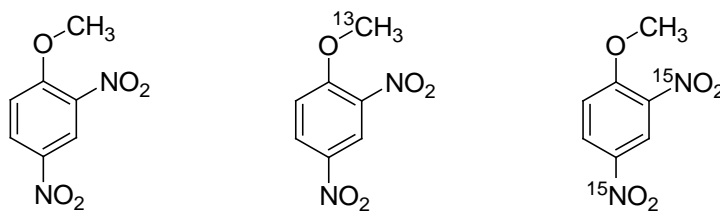


Figure 1. DNAN, ^{13}C -Labeled DNAN, and ^{15}N -Labeled DNAN

1) DNAN is approximately 10% less powerful than TNT (Fedoroff et al., 1960)

2) PAX-21 (36% RDX, 34% DNAN, 30% AP, <1% *N*-methyl nitroaniline) and IMX-101 (40% nitroguanidine, 40% DNAN, 20% NTO)

DNAN is synthesized in high yield and high purity by first dinitrating chlorobenzene to yield chlorodinitrobenzene, and then displacing chloride with methoxide to form DNAN (Fig. 2). Theoretically, dinitration of chlorobenzene can produce six possible regioisomers; in practice however, only two are predominantly formed: 1-chloro-2,4-dinitrobenzene and 1-chloro-2,6-dinitrobenzene (Urbański, 1964; Fedoroff and Sheffield, 1966). Regioselective nitration is best controlled via reaction temperature, with lower temperatures favoring the desired low *ortho*-to-*para* (σ/ρ) ratio. Whereas the first nitration can be conducted at low temperatures, higher temperatures are required for the second nitration of the more deactivated chloronitrobenzene, and some loss of regioselective control is unavoidable at this step. Thus, the regioselectivity of the first nitration is critical to synthesize the desired regioisomer of chlorodinitrobenzene.

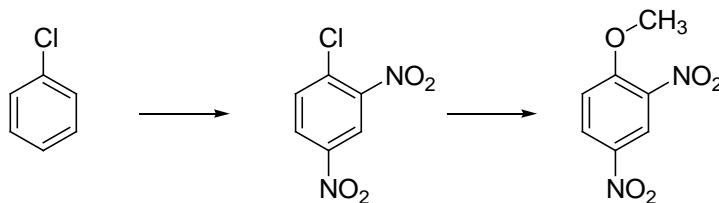


Figure 2. Synthesis of DNAN

RESULTS AND DISCUSSION

The industrial production of chlorodinitrobenzene uses 100% nitric, 96% sulfuric acid, and chlorobenzene, and has been extensively studied (Urbański, 1964; Fedoroff and Sheffield, 1966). To synthesize ^{15}N -labeled chlorodinitrobenzene in a cost effective manner, we adapted our procedure using ^{15}N -labeled sodium nitrate to generate ^{15}N -labeled nitric acid *in situ*, avoiding losses that may occur while distilling ^{15}N -labeled nitric acid. We modeled our molar ratio of sodium nitrate, sulfuric acid, and chlorobenzene on the 100-gram preparation of chlorodinitrobenzene (Davis, 1943), in which the molar ratio of sulfuric acid / nitric acid / chlorobenzene was 3.7 / 2.9 / 1. The sodium sulfate formed acted as a desiccant, absorbing the water produced during nitration (Fig. 3). Also since the sodium nitrate / sulfuric acid mixture was very heterogeneous, we decided to double the amount of sulfuric acid to improve both the mixing efficiency and the thermal control of the reaction.

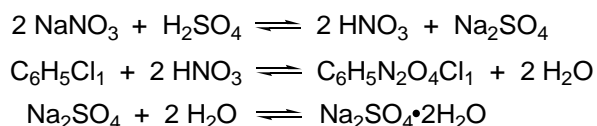


Figure 3. Dinitration of Chlorobenzene

The reaction conditions of our first trial nitration of chlorobenzene were not strong enough, as a significant amount of mono-nitro product (chloronitrobenzene) was observed in the product mixture by ^1H NMR analysis (Table 1). We increased the amount of sodium nitrate and sulfuric acid used in the second trial nitration, and then we increased the reaction time for the third trial nitration. ^1H and ^{13}C NMR analyses of the product mixture for the third trial nitration showed a mixture of the two regioisomers (1-chloro-2,4-dinitrobenzene and 1-chloro-2,6-dinitrobenzene), and failed to show any mono-nitro product. The yield of the third trial nitration was quantitative (ca. 99%).

Table 1. Initial Trial Nitrations of Chlorobenzene

Trial Nitration	Chlorobenzene	Sodium Nitrate	Sulfuric Acid	Conditions	Mono-nitro-product
1	0.51 mL (1)	1.29 g (3)	2.0 mL (7.2)	100 °C / 3 h	10%
2	0.51 mL (1)	1.72 g (4)	3.0 mL (10.8)	100 °C / 4 h	6%
3	0.51 mL (1)	1.72 g (4)	3.0 mL (10.8)	100 °C / 16 h	0%

^a Numbers in parentheses represent the molar ratios used.

We then scaled the nitration reaction 5-fold using the same reaction conditions of our third trial nitration. We improved the regioisomeric ratio of the desired 1-chloro-2,4-dinitrobenzene versus 1-chloro-2,6-dinitrobenzene from 85:15 to 95:5 by changing the heating regimen. Instead of heating the reaction mixture to 40 °C (external temperature), and then slowly raising the temperature to 100 °C over one hour, we introduced a one hour residence time at 40 °C before raising the temperature. This low temperature residence time directed the first nitration toward *para* instead of *ortho*, thus improving the regioselectivity of the final product mixture. Mixing efficiency during the initial nitration also seems to improve the regioselectivity. Again, the yield was quantitative (ca. 99%).

We synthesized DNAN from chlorodinitrobenzene by the simple displacement of chloride using methanol and sodium hydroxide at 60 °C (Fedoroff et al., 1960). Recrystallization from methanol gave needles in good yield (ca. 70%). To improve the yield, the remaining mass balance (ca. 30%) for this reaction could have been recovered during work-up³ and recrystallization. However our objective was to isolate DNAN in very high purity, and recrystallization improved the regioisomeric ratio of 2,4-dinitroanisole to 2,6-dinitroanisole from 95:5 to greater than 98:2, as determined by ^1H and ^{13}C NMR analyses. The use of ^{13}C -labeled methanol as a reagent readily allowed the synthesis of ^{13}C -labeled DNAN, and any possible scrambling of the ^{13}C -labeled carbon with the unlabeled methanol used in the recrystallization was not observed by HRMS analysis.

3) The yield might be increased by first removing some of the reaction solvent before water addition.

Remarkably, the ^1H and ^{13}C NMR spectra for unlabeled chlorodinitrobenzene and unlabeled DNAN were unknown until recently (Aridoss and Laali, 2011), and are reported herein. The mono-nitro product (1-chloro-4-nitrobenzene) and 1-chloro-2,6-dinitrobenzene each possess a plane of symmetry, and were readily identified via ^1H NMR analysis by the presence of a pair of coupled doublets and by the presence of a triplet coupled with a doublet, respectively. The nitro carbons of chlorodinitrobenzene and DNAN were readily identified in the ^{13}C NMR spectra since these chemical shifts are broad due to quadrupolar relaxation from the directly attached ^{14}N nucleus. The chemical shifts for these same nitro carbons of the ^{15}N -labeled compounds each becomes a sharpened doublet of doublets as the quadrupolar relaxation mechanism no longer exists without an ^{14}N nucleus present, allowing the one- and three-bond coupling ($^1J_{\text{N,C}}$ and $^3J_{\text{N,C}}$) between the ^{13}C and ^{15}N nuclei to be readily observed.

We collected the ^{15}N NMR spectra for ^{15}N -labeled chlorodinitrobenzene and DNAN, as well as their ^1H and ^{13}C NMR spectra. We observed two chemical shifts in the ^{15}N NMR spectra for both compounds, each in the characteristic region (10 to -30 ppm) of an aromatic nitro group (Berger et al., 1997). The longitudinal relaxation times (T_1) for these ^{15}N nitro groups were measured to be quite long (hundreds of seconds). Since ^{15}N NMR also suffers from low sensitivity and a negative gyromagnetic ratio, knowledge of T_1 allows one to optimize ^{15}N NMR pulse sequences of chlorodinitrobenzene and DNAN at natural abundance levels of nitrogen. Since we determined the T_1 values using de-oxygenated samples, one must keep in mind that the paramagnetic oxygen present in the typically prepared NMR solution will shorten the true T_1 reported herein.

We also collected the ^{14}N NMR spectra for natural abundance chlorodinitrobenzene, natural abundance DNAN, and ^{13}C -labeled DNAN. Whereas T_1 governs ^{15}N NMR, the transverse relaxation time (T_2) dictates ^{14}N NMR. Since the observed T_2 values (T_2^*) were extremely short (milliseconds), the spectra we collected show significant line broadening, and the NMR acquisition times used were only 26 milliseconds. The two nitro groups for both compounds are indistinguishable by ^{14}N NMR, and only one chemical shift is observed. Nevertheless, these ^{14}N NMR spectra clearly show that an aromatic nitro group is present (Berger et al., 1997).

We observed strong absorption bands in all the IR spectra that are characteristic for the symmetrical ($1360\text{-}1310\text{ cm}^{-1}$) and asymmetrical stretches ($1560\text{-}1490\text{ cm}^{-1}$) of the aromatic nitro group (Pretsch et al., 2000). The effect of isotopic substitution on the nitro stretches in the IR spectra is readily apparent as we observe the expected shift to lower vibrational frequencies for the ^{15}N -labeled compounds (ca. 25 cm^{-1} shift).

CONCLUSIONS

We synthesized chlorodinitrobenzene and ^{15}N -labeled chlorodinitrobenzene in high yield (ca. 99%) and high regioselectivity (ca. 95:5) via dinitration of chlorobenzene. Using sodium nitrate and sulfuric acid, we conducted the nitration first at 40 °C, and then slowly raised the reaction temperature to 100 °C. We determined the regioselectivity by ^1H NMR analysis, and we did not observe any chloronitrobenzene after our procedure had been optimized. The use of ^{15}N -labeled sodium nitrate readily allows the synthesis of the ^{15}N -labeled product. We also report the ^1H , ^{13}C , ^{14}N , and ^{15}N NMR, as well as the IR, and HRMS spectra for labeled and unlabeled chlorodinitrobenzene, as well as the ^{15}N NMR T_1 values.

We synthesized DNAN, ^{13}C -labeled DNAN, and ^{15}N -labeled DNAN from chlorodinitrobenzene by the simple displacement of chloride using methanol and sodium hydroxide at 60 °C. Recrystallization from methanol improved the purity, with a regioisomeric ratio of 2,4-dinitroanisole to 2,6-dinitroanisole greater than 98:2, yielding needles in high yield (ca. 70%). The use of ^{13}C -labeled methanol as a reagent readily allows the synthesis of ^{13}C -labeled DNAN. We also report the ^1H , ^{13}C , ^{14}N , and ^{15}N NMR, as well as the IR, and HRMS spectra for labeled and unlabeled DNAN, as well as the ^{15}N NMR T_1 values. We hope the use of these selectively labeled DNAN compounds will support future work in the determination of bioaccumulation factors and biodegradation mechanisms for DNAN.

EXPERIMENTAL SECTION

Chlorobenzene, sodium nitrate, and methanol were purchased from Aldrich Chemical Co. Sulfuric acid and sodium hydroxide were purchased from Fisher Scientific. The isotopic enrichment of ^{15}N -labeled sodium nitrate was 98.2%, and the isotopic enrichment of ^{13}C -labeled methanol was 99.1%; both were purchased from Cambridge Isotope Laboratories, Inc. Chlorodinitrobenzene and DNAN are toxic, and care was taken to avoid exposure to by working with gloves in a well-ventilated hood. Chlorodinitrobenzene and DNAN are also 1.1 explosives, and the appropriate safety precautions and disposal methods were taken.

Melting points are uncorrected and were measured using a Mel-Temp apparatus. ^1H , ^{13}C , ^{14}N , and ^{15}N NMR analyses (300.1, 75.5, 21.7, 30.4 MHz, respectively) were performed on a Varian Mercury Plus 300 instrument and on an Agilent DD2 300 instrument in samples dissolved in CDCl_3 . Downfield chemical shifts are recorded in positive δ units in ppm relative to TMS ($\delta = 0.00$) for ^1H NMR, to the residual solvent peak in CDCl_3 ($\delta = 77.0$) for ^{13}C NMR, and to external nitromethane ($\delta = 0.00$) for ^{14}N and ^{15}N NMR. Multiplicities are described as singlet (s), doublet (d), triplet (t), quartet (q), doublet of doublets (dd), doublet of doublet of doublets (ddd), multiplet (m), and broad (br). Proton decoupling (WALTZ-16) was used for acquiring the ^{13}C NMR spectra, and an inverse gated decoupling sequence was used for acquiring the ^{15}N NMR spectra. The line-widths at half height ($\Delta\nu_{1/2}$) reported for the ^{14}N NMR spectra

were processed without exponential multiplication; T_2^* -values were determined using solutions degassed of nitrogen⁴ (multiple freeze-pump-thaw cycles with J Young valves) from the equation: $\pi\Delta\nu_{1/2} = 1/T_2^*$. The T_1 -values reported for the ^{15}N NMR spectra were determined using 150 mM solutions degassed of oxygen (multiple freeze-pump-thaw cycles with J Young valves) via the inversion recovery method at 25 °C after sample calibration of the 90° pulse width, and fitted (<2.5% error) to an exponential plot. IR spectra were recorded on a Nicolet 750 FTIR spectrometer at 2 wavenumber resolution using a Diamond ATR accessory. High resolution (<10 ppm) mass spectra were acquired on a JEOL AccuTOF (time-of-flight) mass spectrometer using DART ionization conditions (250 °C, 20 eV), calibrated to PEG 600. Retention factors (R_f) for thin layer chromatography (TLC) were performed using Sigma-Aldrich 0.20 mm silica gel pre-coated aluminum plates; spots were visualized under UV light. All glassware used was oven dried for >2 h at 120 °C. All spectra are included in the Supporting Information section.

Synthesis of 1-Chloro-2,4-dinitrobenzene

A heterogeneous mixture of sodium nitrate (99%, 8.585 g, 100 mmol, 4 equiv) and sulfuric acid (96%, 15.0 mL, 270 mmol, 10.8 equiv) was stirred in a 50-mL round bottom flask in a room temperature oil bath at 900 rpm with a 20-mm egg-shaped stir bar for 30 min. Chlorobenzene (99.8%, 2.55 mL, 25.0 mmol, 1 equiv) was added dropwise over 2 min and a condenser was attached to the reaction flask. The oil bath was heated to 40 °C over 5 min, and the yellow reaction mixture was stirred for 1 h. Then the oil bath temperature was slowly raised to 100 °C over 1 h in 10 °C increments, and the reaction mixture was stirred for 16 h.

The reaction flask was removed from heat, and the warm reaction mixture was poured via a glass funnel into a 125-mL Erlenmeyer flask containing cold water (75 mL) that was sitting in an ice bath, along with the cold water (25 mL) rinsings of the reaction flask. After allowing the diluted reaction mixture to sit at 4 °C for 30 min, the resulting solid was isolated using vacuum filtration (Whatman #1) via a handheld pump, and the solid was washed with water (25 mL). The solid was dissolved in diethyl ether (50 mL) and transferred to a 125-mL separatory funnel, along with diethyl ether (25 mL total) rinsings of the reaction flask and the Erlenmeyer flask. The organic layer was washed with saturated sodium bicarbonate (3 x 25 mL), water (2 x 25 mL), and saturated brine (25 mL), and the solution was allowed to air-dry in a conductive container in the hood to yield a yellow solid (5.013 g, 99%). TLC showed one spot, $R_f = 0.65$ (Et_2O). ^1H NMR analysis determined the isomeric ratio of 1-chloro-2,4-dinitrobenzene to 1-chloro-2,6-dinitrobenzene to be ca. 95:5.

4) Molecular nitrogen was removed for aesthetic reasons. ^{14}N NMR (CDCl_3): δ -75 ppm.

1-Chloro-2,4-dinitrobenzene

mp: 44-46 °C

^1H NMR (CDCl_3): δ 8.77 (d, 2.6Hz, 1H), 8.40 (dd, 9.0Hz, 2.7Hz, 1H), and 7.80 (d, 8.8Hz, 1H) ppm.

^{13}C NMR (CDCl_3): δ 147.8 (br), 146.3 (br), 133.9, 133.2, 127.3, and 121.1 ppm.

^{14}N NMR (CDCl_3): δ -23 ($\Delta\nu_{1/2} = 52\text{Hz}$, $T_2^* = 6.1\text{ ms}$) ppm.

IR: 3108, 1604, 1590, 1542, 1532, 1458, 1346, 1304, 1246, 1158, 1140, 1046, 918, 902, 850, 836, 748, and 730 cm^{-1} .

HRMS: Calcd for $\text{C}_6\text{H}_4\text{N}_2\text{O}_4\text{Cl}_1$ $[\text{M}+\text{H}]^+$ 202.9854. Found 202.9853 (0.5 ppm).

1-Chloro-2,6-dinitrobenzene (^1H NMR chemical shift values for this isomer were determined directly from the isolated isomeric mixture.)

^1H NMR (CDCl_3): δ 8.00 (d, 8.1Hz, 2H) and 7.62 (t, 8.1Hz, 1H) ppm.

1-Chloro-4-nitrobenzene (^1H NMR chemical shift values were determined directly from the crude reaction mixture.)

^1H NMR (CDCl_3): δ 8.19 (d, 9.2Hz, 2H) and 7.54 (d, 9.2Hz, 2H) ppm.

1-Chloro-[2,4- $^{15}\text{N}_2$]dinitrobenzene

mp: 44-46 °C

^1H NMR (CDCl_3): δ 8.77 (dd, 4.8Hz, 2.6Hz, 1H), 8.40 (ddd, 8.8Hz, 2.6Hz, 1.8Hz, 1H), and 7.81 (d, 8.8Hz, 1H) ppm.

^{13}C NMR (CDCl_3): δ 147.8 (dd, 18Hz, 3Hz), 146.3 (dd, 17Hz, 3Hz), 133.9, 133.2 (t, 2Hz), 127.3 (d, 2Hz), and 121.1 (m*) ppm. *Apparent triplet with approximately 2Hz coupling.

^{15}N NMR (CDCl_3): δ -22.7 ($T_1 = 97\text{ s} \pm 2\text{ s}$) and -23.6 ($T_1 = 143\text{ s} \pm 3\text{ s}$) ppm.

IR: 3108, 1588, 1510, 1456, 1318, 1246, 1156, 1138, 1046, 914, 900, 850, 828, 748, and 720 cm^{-1} .

HRMS: Calcd for $\text{C}_6\text{H}_4^{15}\text{N}_2\text{O}_4\text{Cl}_1$ $[\text{M}+\text{H}]^+$ 204.9795. Found 204.9799 (2.0 ppm).

1-Chloro-[2,6-¹⁵N₂]dinitrobenzene (¹H NMR chemical shift values for this isomer were determined directly from the isolated isomeric mixture.)

¹H NMR (CDCl₃): δ 8.02 (dd, 8.1Hz, 1.8Hz, 2H), and 7.63 (t, 8.1Hz, 1H) ppm.

Synthesis of 2,4-Dinitroanisole (DNAN)

A heterogeneous mixture of freshly ground sodium hydroxide (97%, 619 mg, 15.0 mmol, 1.2 equiv) and methanol (99.8%, 4.06 mL, 100 mmol, 8 equiv) was stirred in a 25-mL round bottom flask in a room temperature water bath at 600 rpm with a 20-mm egg-shaped stir bar for 30 min. 1-Chloro-2,4-dinitrobenzene (2.532 g, 12.5 mmol, 1 equiv) was added in portions via a glass funnel over 10 min, and a condenser was attached to the reaction flask. The water bath was heated to 60 °C over 15 min, and the red reaction mixture was stirred for 1 h.

After the orange reaction mixture was removed from heat, water (10 mL) was added, and the stirred reaction mixture was allowed to cool to room temperature over 30 min. The crude solid was isolated using vacuum filtration (Whatman #1) via a handheld pump, and the solid was washed with water (4 x 10 mL). Recrystallization was achieved by stirring a heterogeneous mixture of the yellow solid and methanol (45 mL) in a 125-mL Erlenmeyer flask sitting in a room temperature water bath, and then heating the water bath to 85 °C. The resulting clear solution was poured into a 250-mL beaker, and allowed to cool to room temperature over 30 min with occasional gentle swirling. The product was isolated using vacuum filtration (Whatman #1) via a handheld pump, and then allowed to air-dry in a conductive container in the hood to yield yellow needles; successive second and third recrystallization crops also yielded yellow needles (1.734 g, 70%). TLC showed one spot, R_f = 0.38 (Et₂O).

2,4-Dinitroanisole

mp: 86-88 °C

¹H NMR (CDCl₃): δ 8.77 (d, 2.9Hz, 1H), 8.46 (dd, 9.2Hz, 2.9Hz, 1H), 7.23 (d, 9.2Hz, 1H), and 4.10 (s, 3H) ppm.

¹³C NMR (CDCl₃): δ 157.3, 140.2 (br), 138.9 (br), 129.1, 121.9, 113.6, and 57.5 ppm.

¹⁴N NMR (CDCl₃): δ -21 (Δν_{1/2} = 70Hz, T₂^{*} = 4.5 ms) ppm.

IR: 3098, 1600, 1520, 1488, 1442, 1416, 1344, 1318, 1278, 1186, 1152, 1136, 1070, 1002, 920, 830, 792, 762, and 742 cm⁻¹.

HRMS: Calcd for C₇H₇N₂O₅ [M+H]⁺ 199.0350. Found 199.0335 (7.5 ppm).

[2,4-¹⁵N₂]Dinitroanisol

mp: 86-88 °C

¹H NMR (CDCl₃): δ 8.77 (m, 4.8Hz, 2.6Hz, 1H), 8.46 (ddd, 9.2Hz, 2.6Hz, 1.8Hz, 1H), 7.23 (d, 9.2Hz, 1H), and 4.10 (s, 3H) ppm.

¹³C NMR (CDCl₃): δ 157.3, 140.1 (dd, 17Hz, 3Hz), 138.8 (dd, 17Hz, 3Hz), 129.2 (d, 2Hz), 121.9 (t, 2Hz), 113.6 (m*), and 57.5 ppm. *Apparent triplet with approximately 2Hz coupling.

¹⁵N NMR (CDCl₃): δ -21.1 (T₁ = 104 s ± 2 s) and -21.4 (T₁ = 135 s ± 2 s) ppm.

IR: 3114, 1602, 1498, 1442, 1414, 1328, 1310, 1276, 1186, 1152, 1136, 1068, 1002, 916, 828, 790, 750, and 730 cm⁻¹.

HRMS: Calcd for C₇H₇¹⁵N₂O₅ [M+H]⁺ 201.0290. Found 201.0302 (6.0 ppm).

[*methyl*-¹³C]2,4-Dinitroanisol

mp: 86-88 °C

¹H NMR (CDCl₃): δ 8.77 (d, 2.9Hz, 1H), 8.46 (dd, 9.2Hz, 2.9Hz, 1H), 7.23 (d, 9.2Hz, 1H), and 4.10 (d, 147.0Hz, 3H) ppm.

¹³C NMR (CDCl₃): δ 157.3, 140.2 (br), 138.8 (br), 129.1, 121.9, 113.6 (d, 5Hz), and 57.5 ppm.

¹⁴N NMR (CDCl₃): δ -21 (Δν_{1/2} = 68z, T₂^{*} = 4.7 ms) ppm.

IR: 3098, 1600, 1522, 1488, 1438, 1416, 1344, 1318, 1278, 1180, 1150, 1136, 1068, 990, 918, 830, 790, 762, and 742 cm⁻¹.

HRMS: Calcd for C₆¹³C₁H₇N₂O₅ [M+H]⁺ 200.0383. Found 200.0397 (7.0 ppm).

SUPPORTING INFORMATION

1-Chloro-2,4-dinitrobenzene

¹ H NMR	11
¹³ C NMR	12
¹⁴ N NMR	13
IR	14
HRMS	14

1-Chloro-[2,4-¹⁵N₂]dinitrobenzene

¹ H NMR	15
¹³ C NMR	16
¹⁵ N NMR	17
IR	18
HRMS	18

2,4-Dinitroanisole

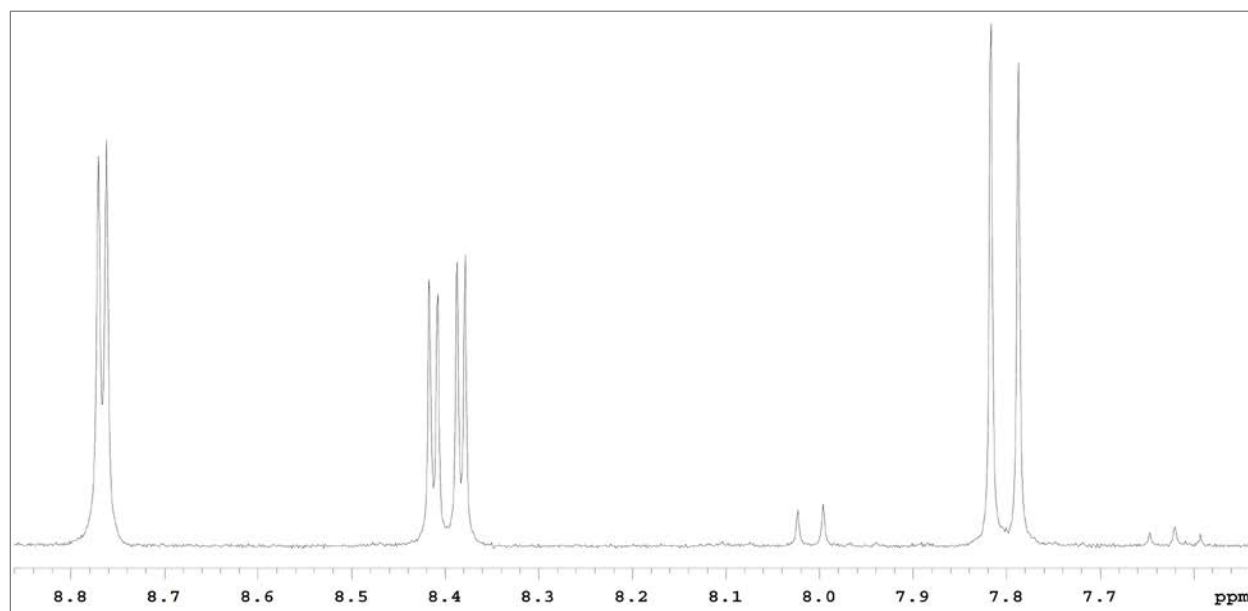
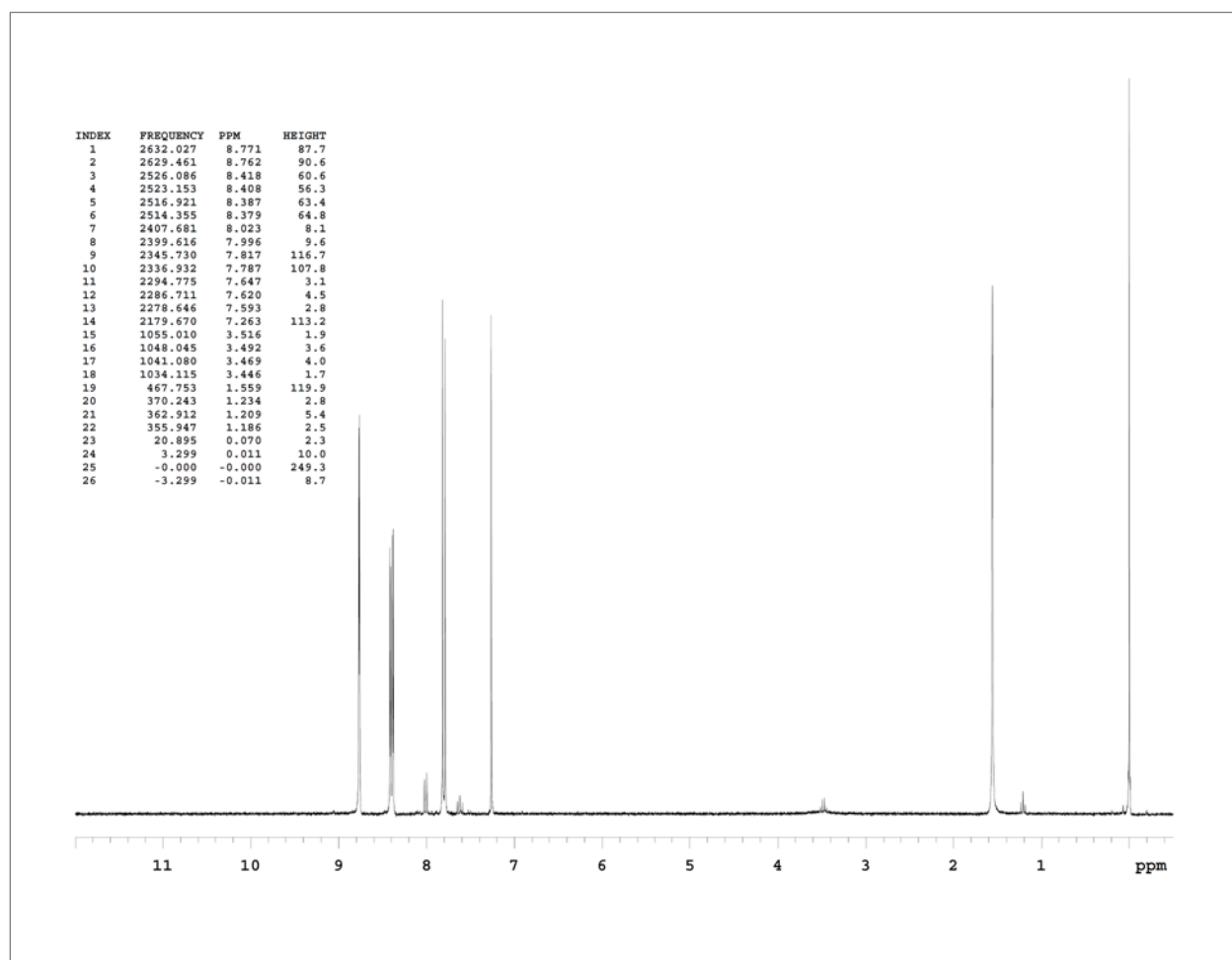
¹ H NMR	19
¹³ C NMR	20
¹⁴ N NMR	21
IR	22
HRMS	22

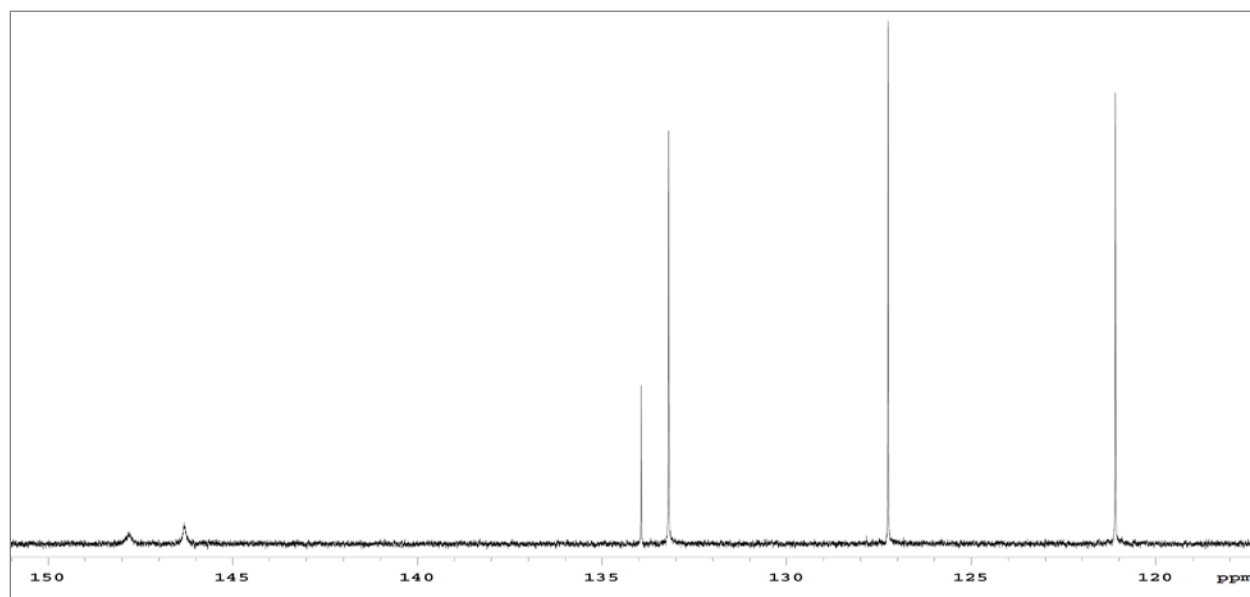
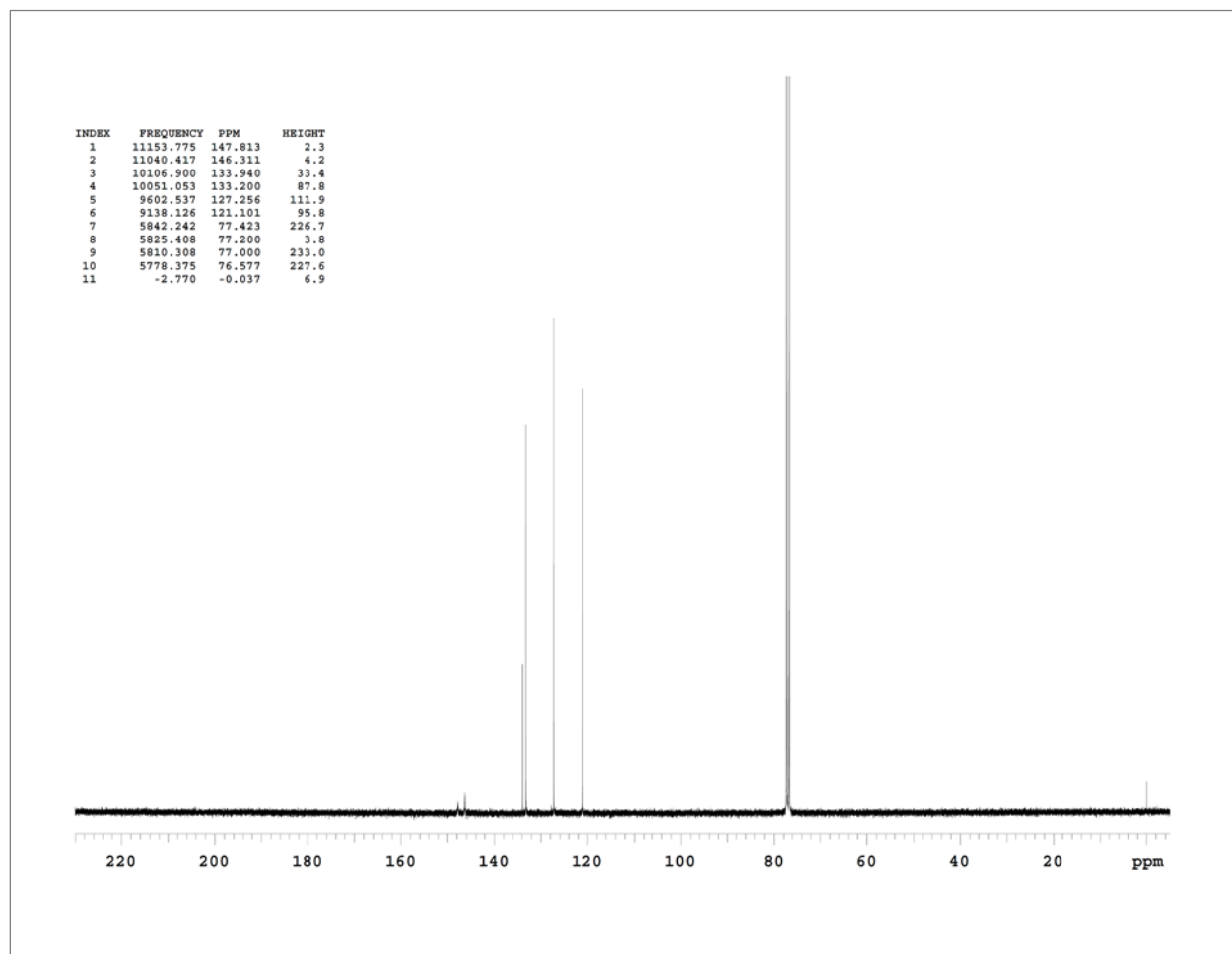
[2,4-¹⁵N₂]Dinitroanisole

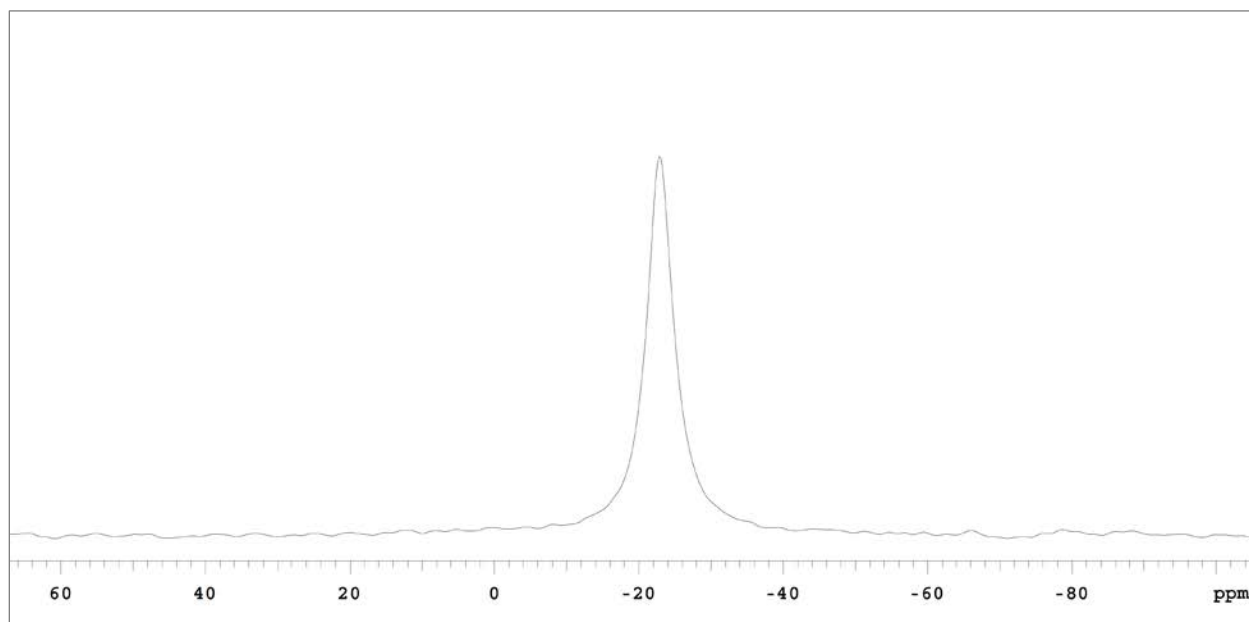
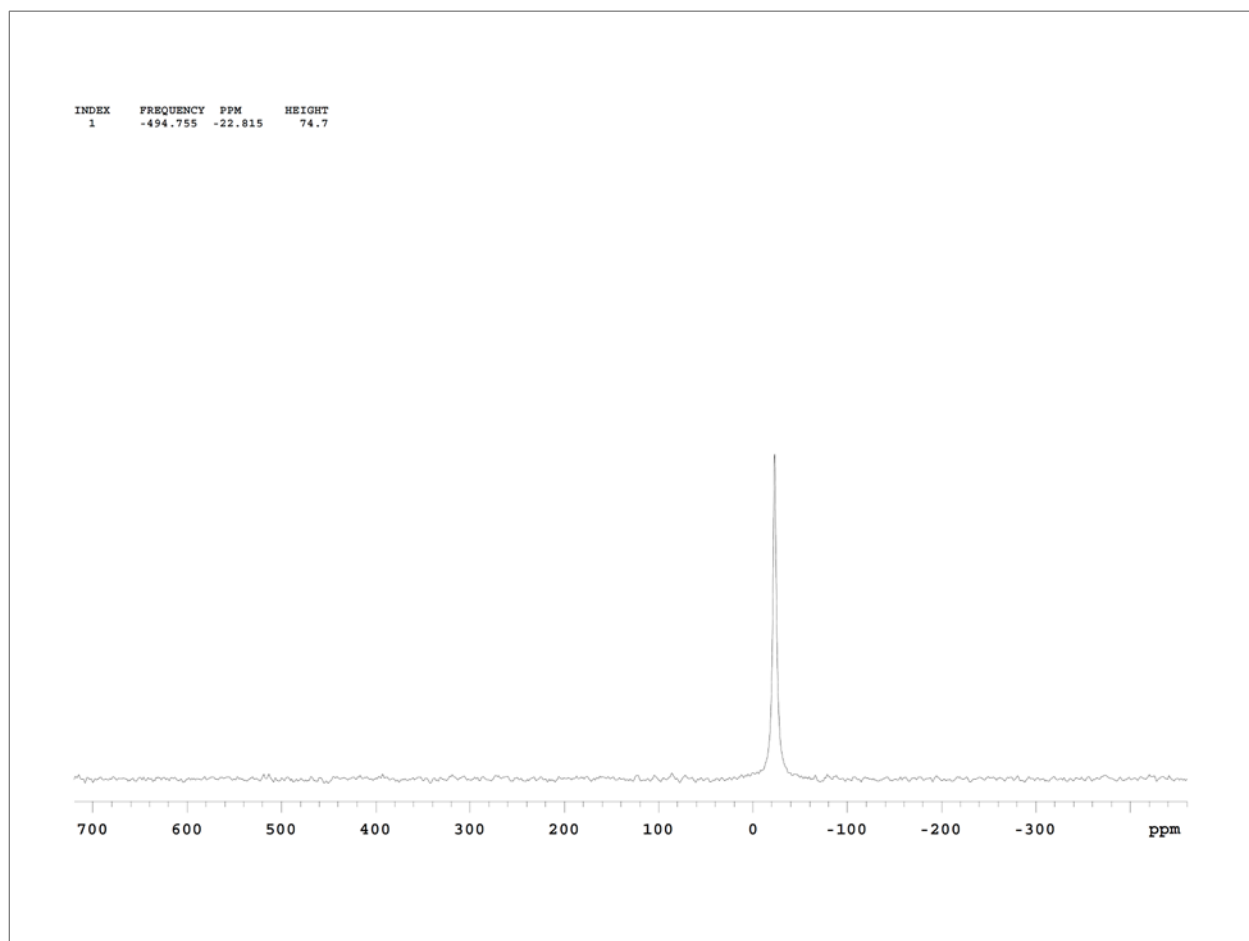
¹ H NMR	23
¹³ C NMR	24
¹⁵ N NMR	25
IR	26
HRMS	26

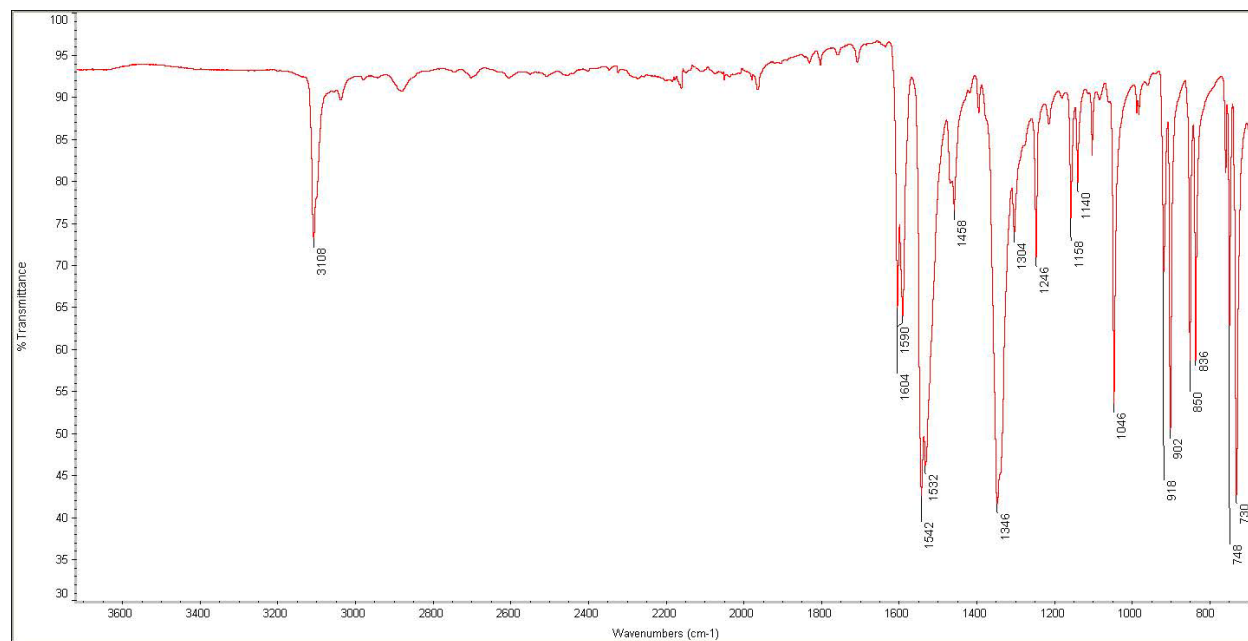
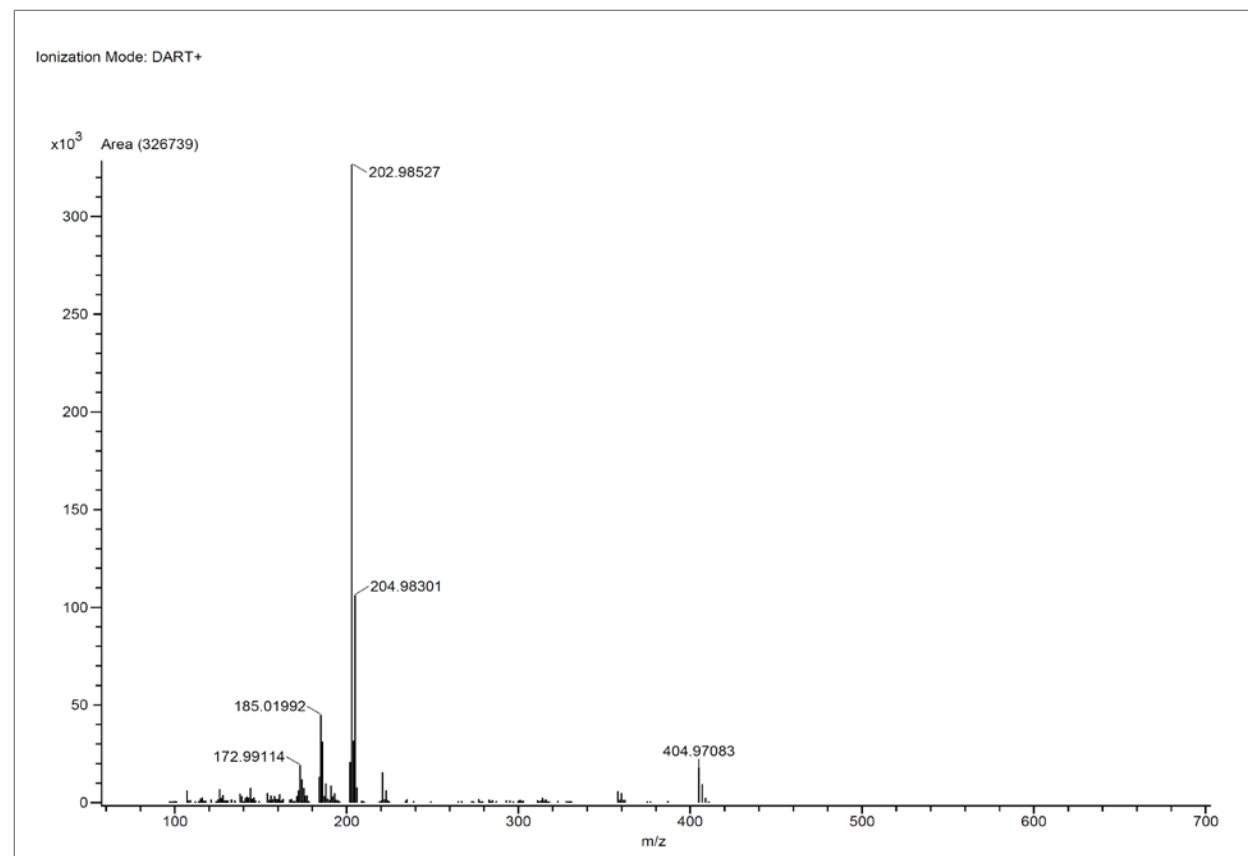
[*methyl*-¹³C]2,4-Dinitroanisole

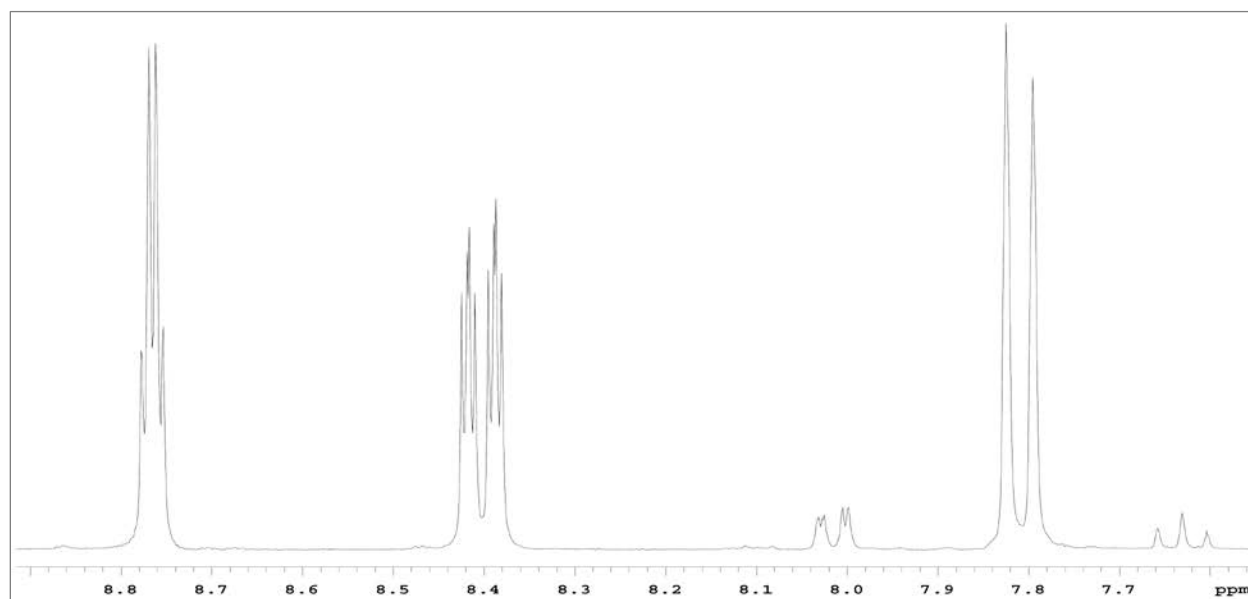
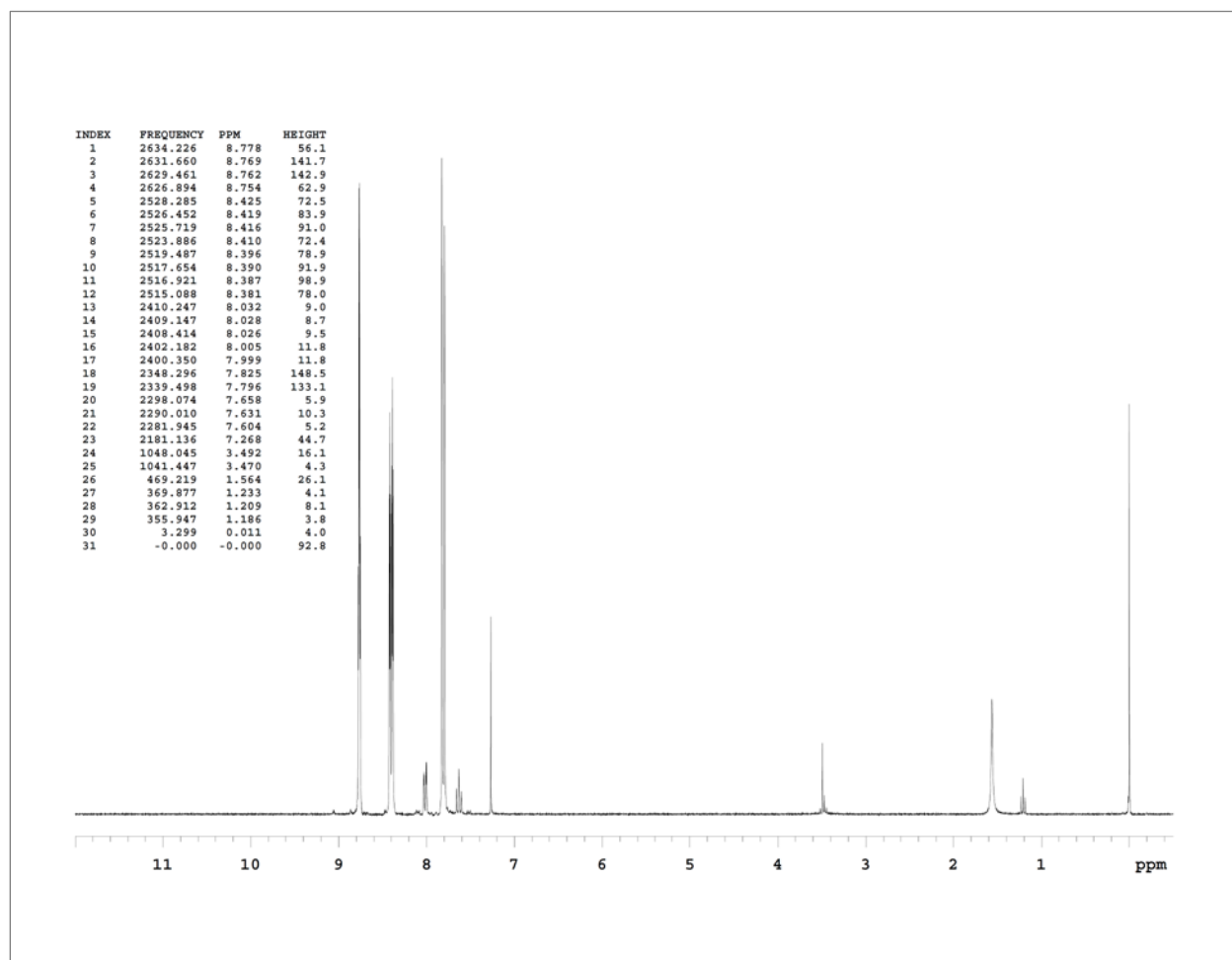
¹ H NMR	27
¹³ C NMR	28
¹⁴ N NMR	29
IR	30
HRMS	30

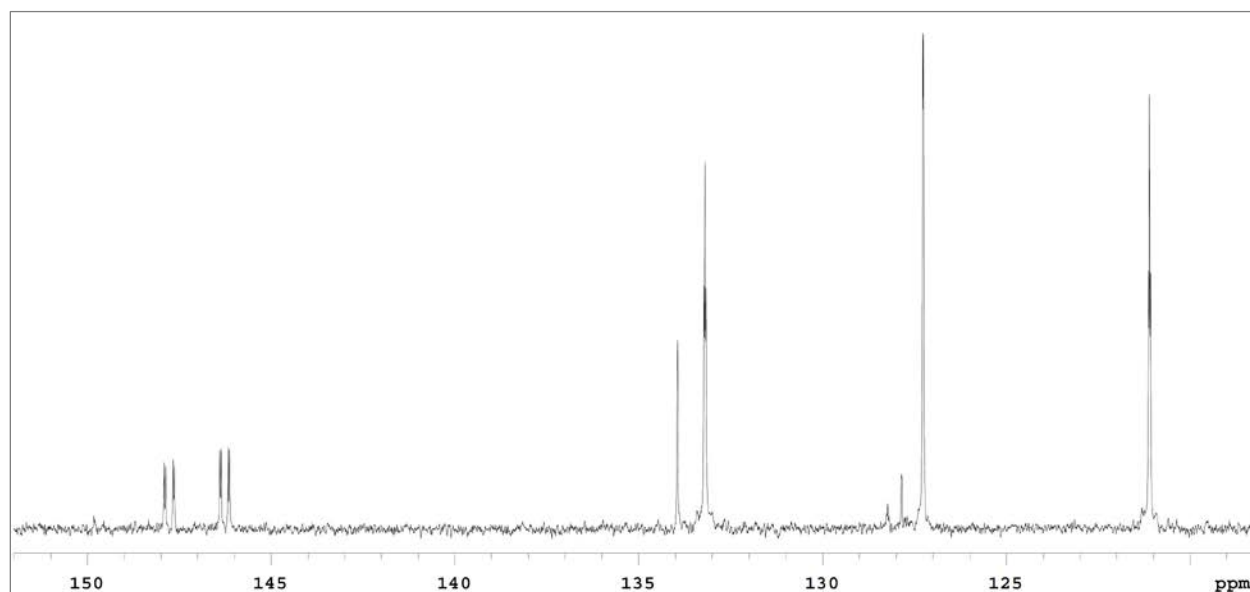
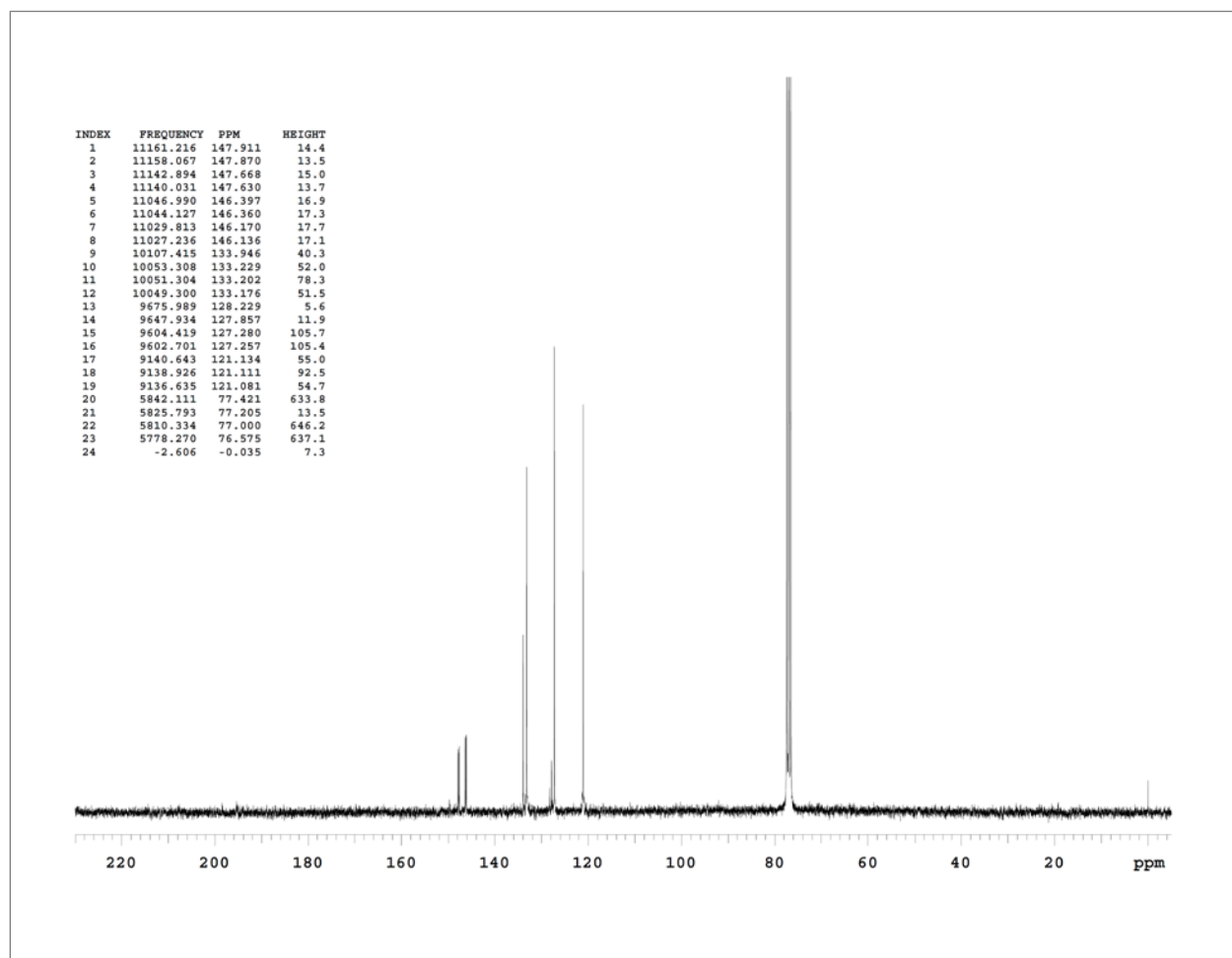
1-Chloro-2,4-dinitrobenzene / ^1H NMR

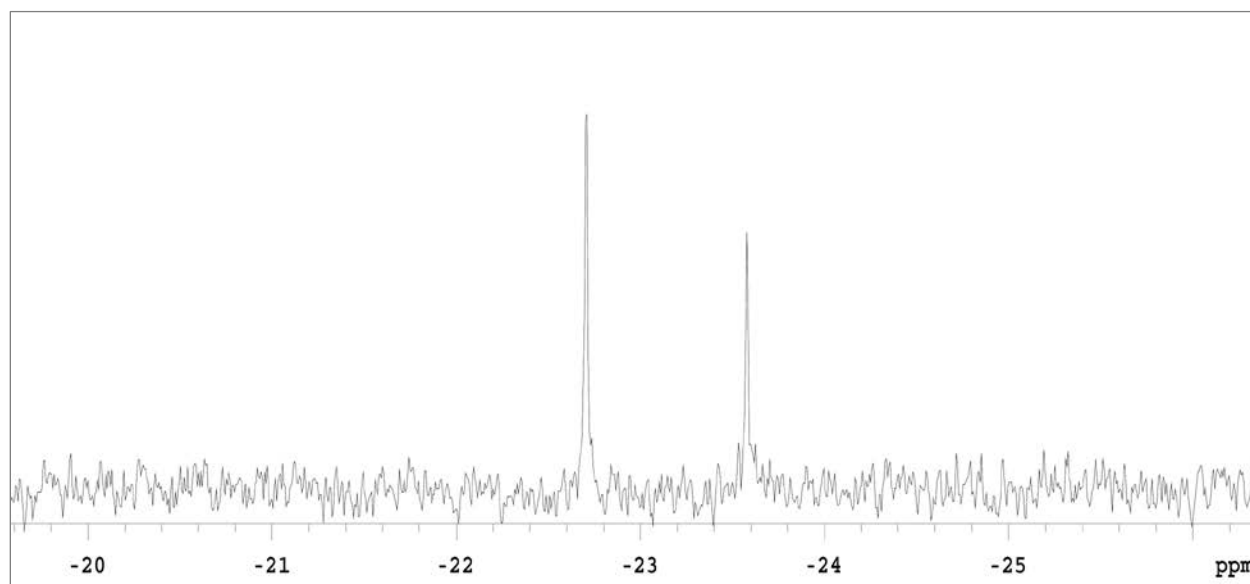
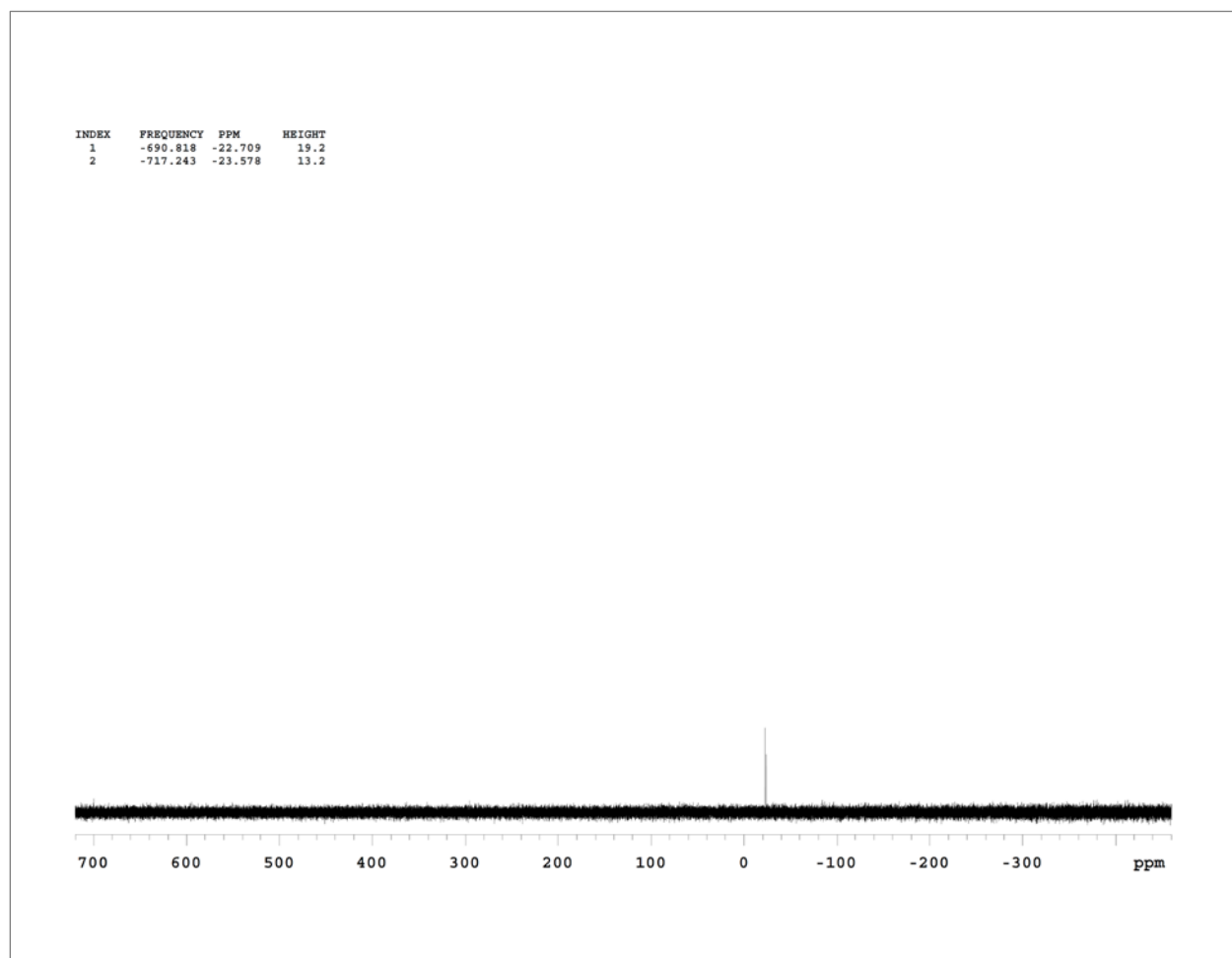
1-Chloro-2,4-dinitrobenzene / ^{13}C NMR

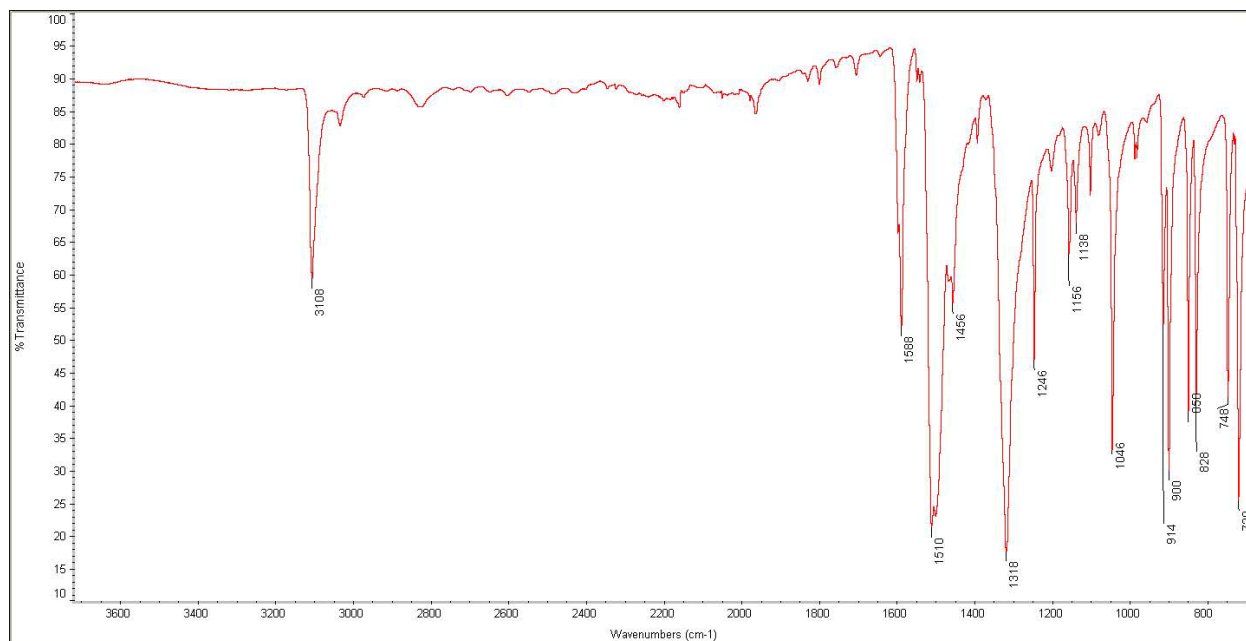
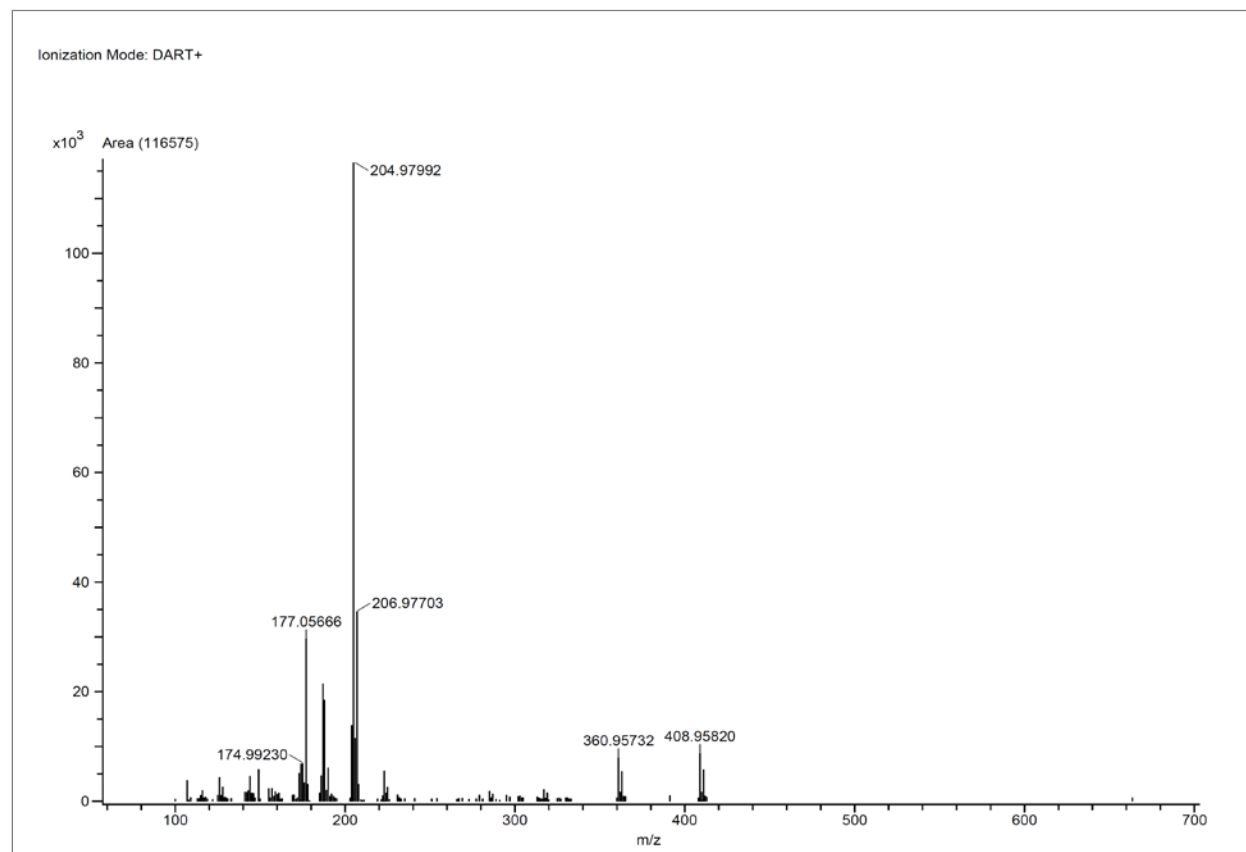
1-Chloro-2,4-dinitrobenzene / ^{14}N NMR

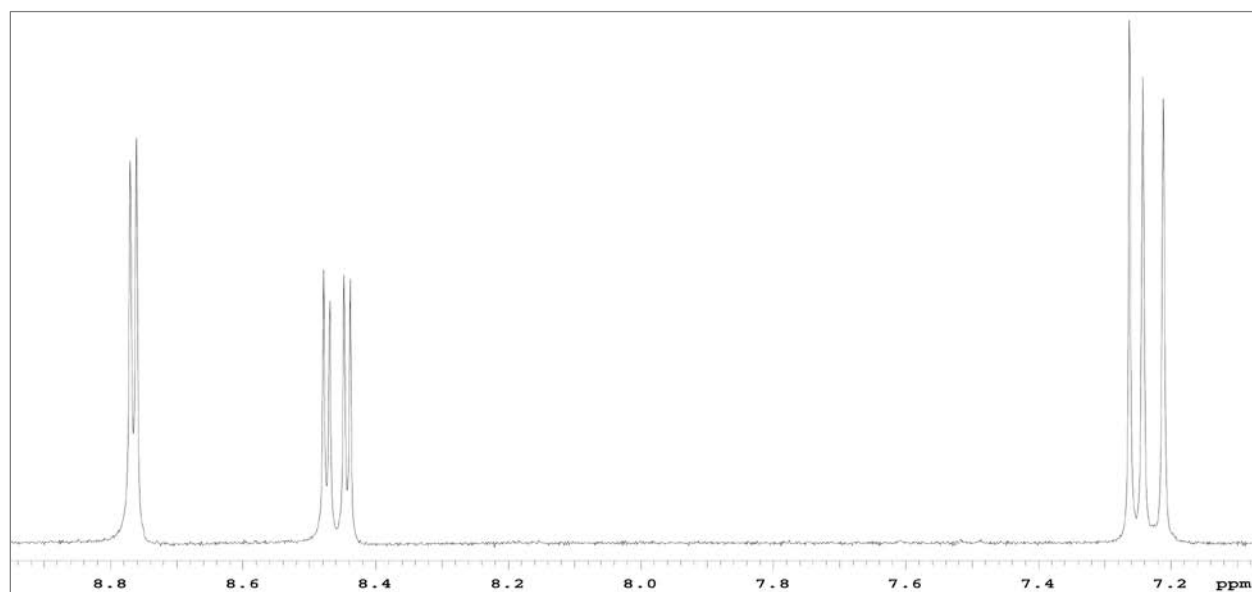
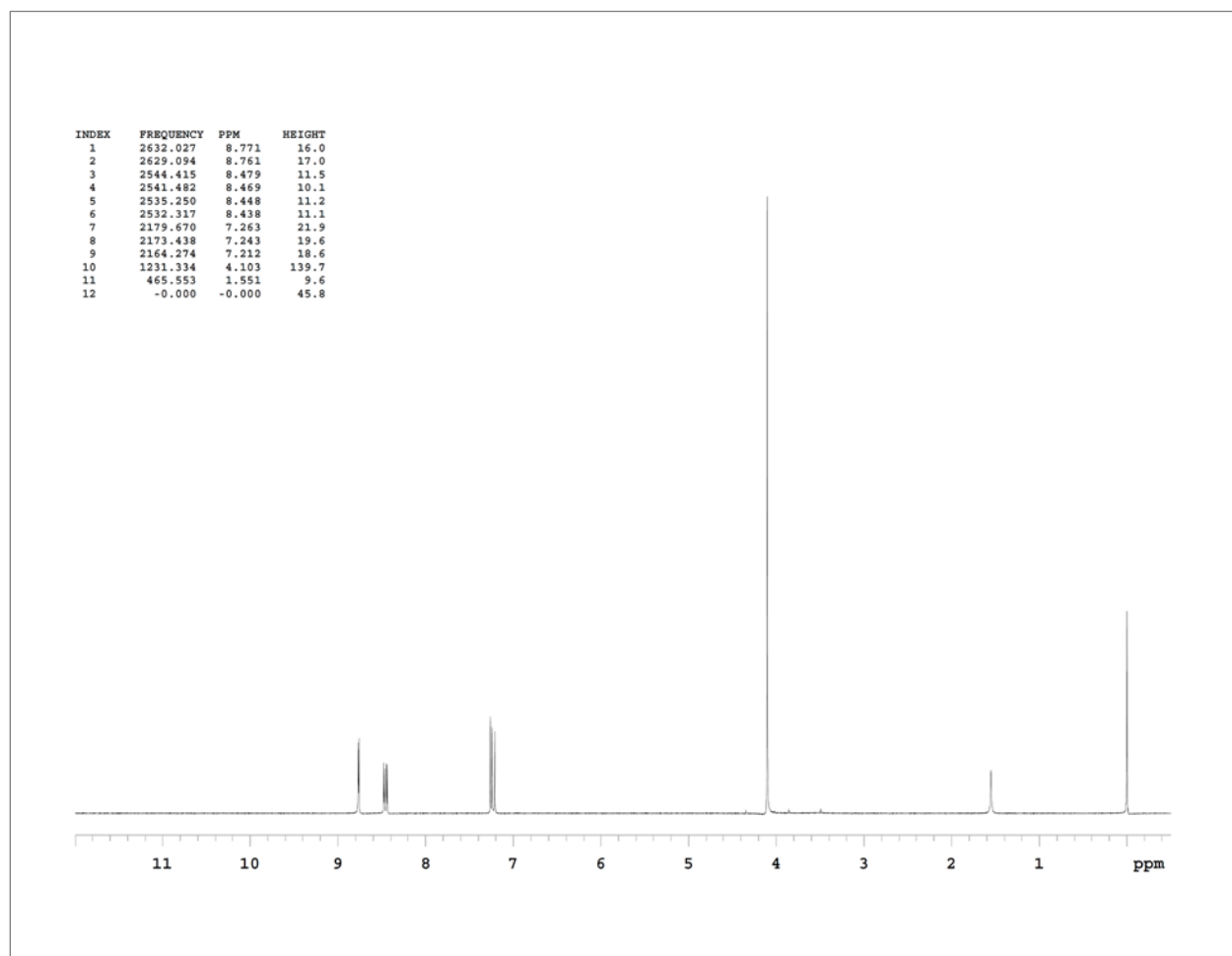
1-Chloro-2,4-dinitrobenzene / IR**1-Chloro-2,4-dinitrobenzene / HRMS**

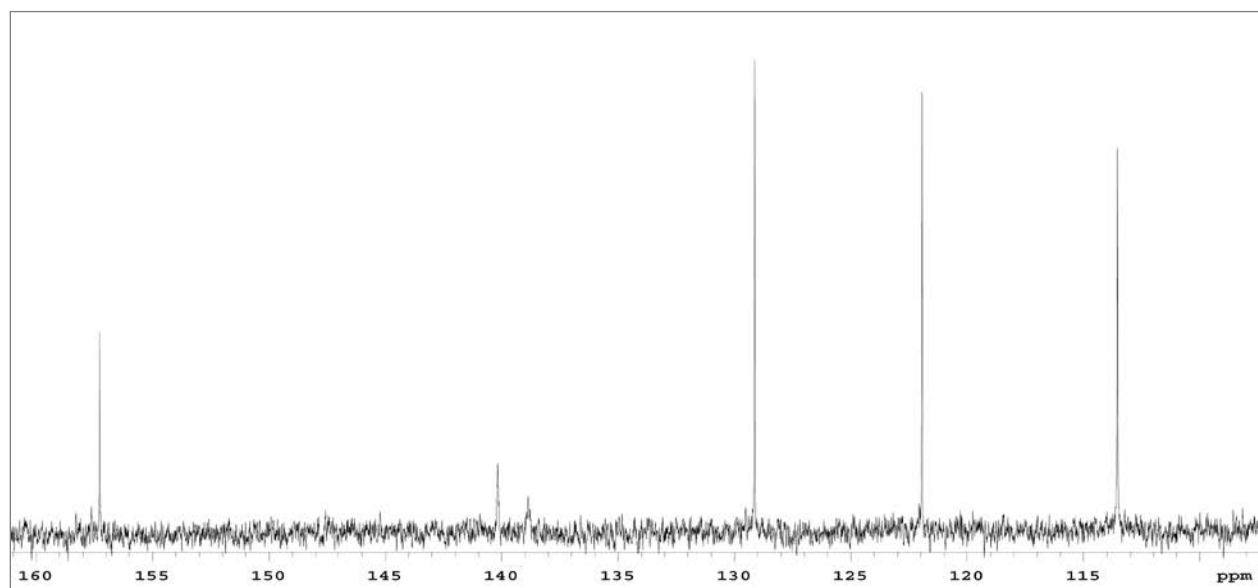
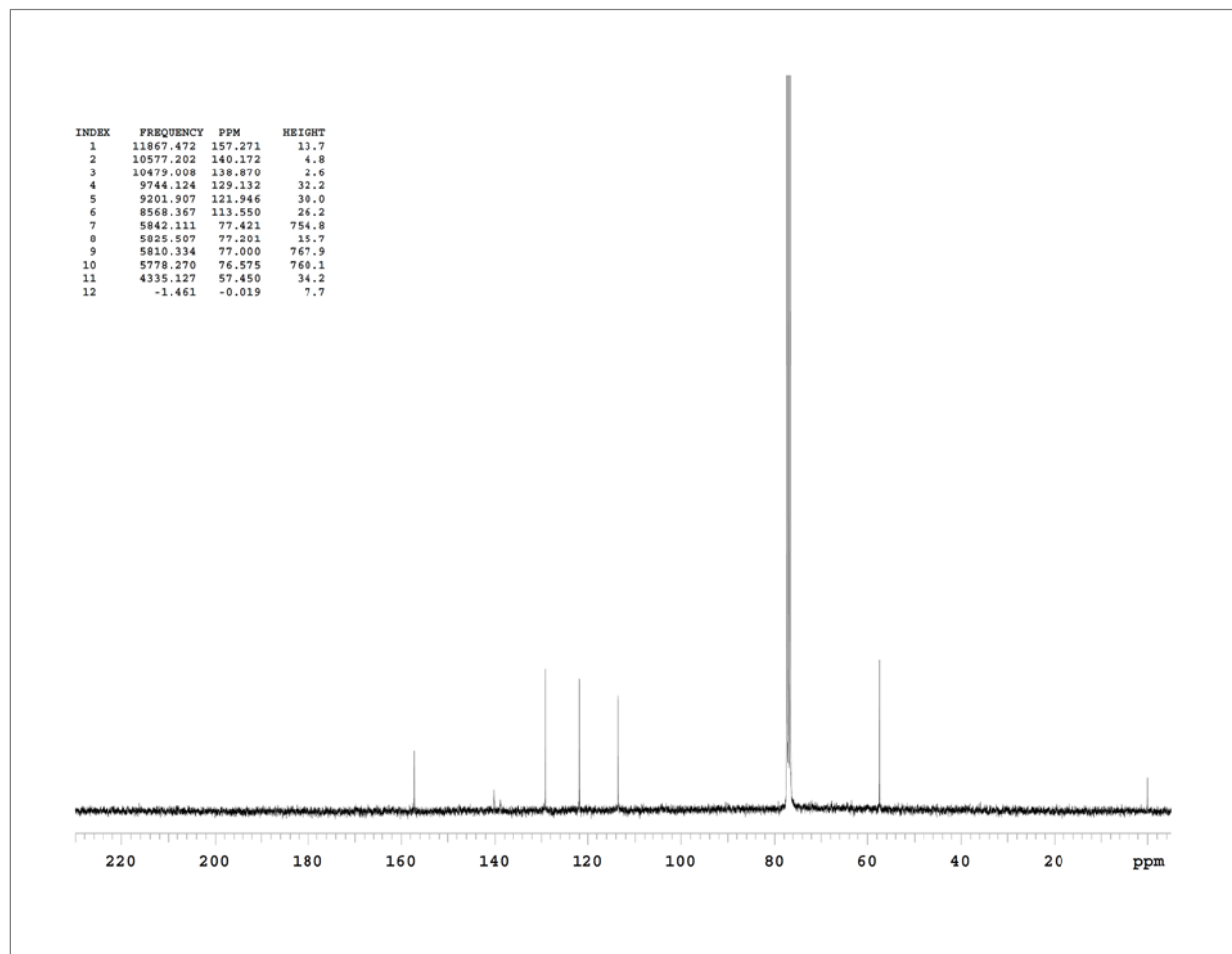
1-Chloro-[2,4-¹⁵N₂]dinitrobenzene / ¹H NMR

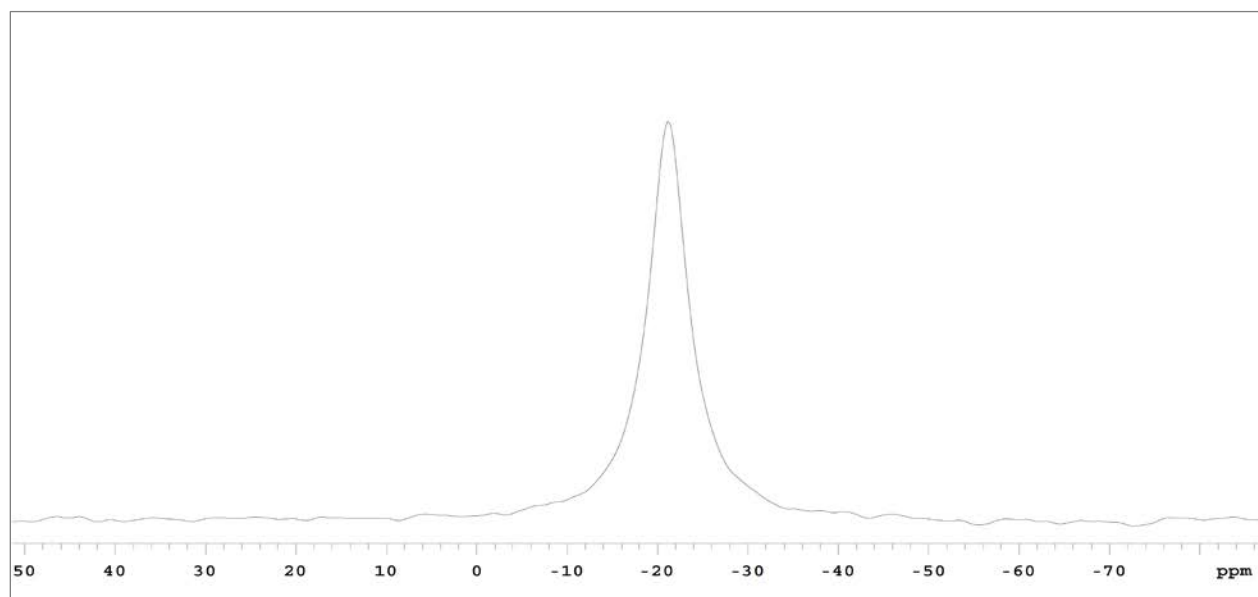
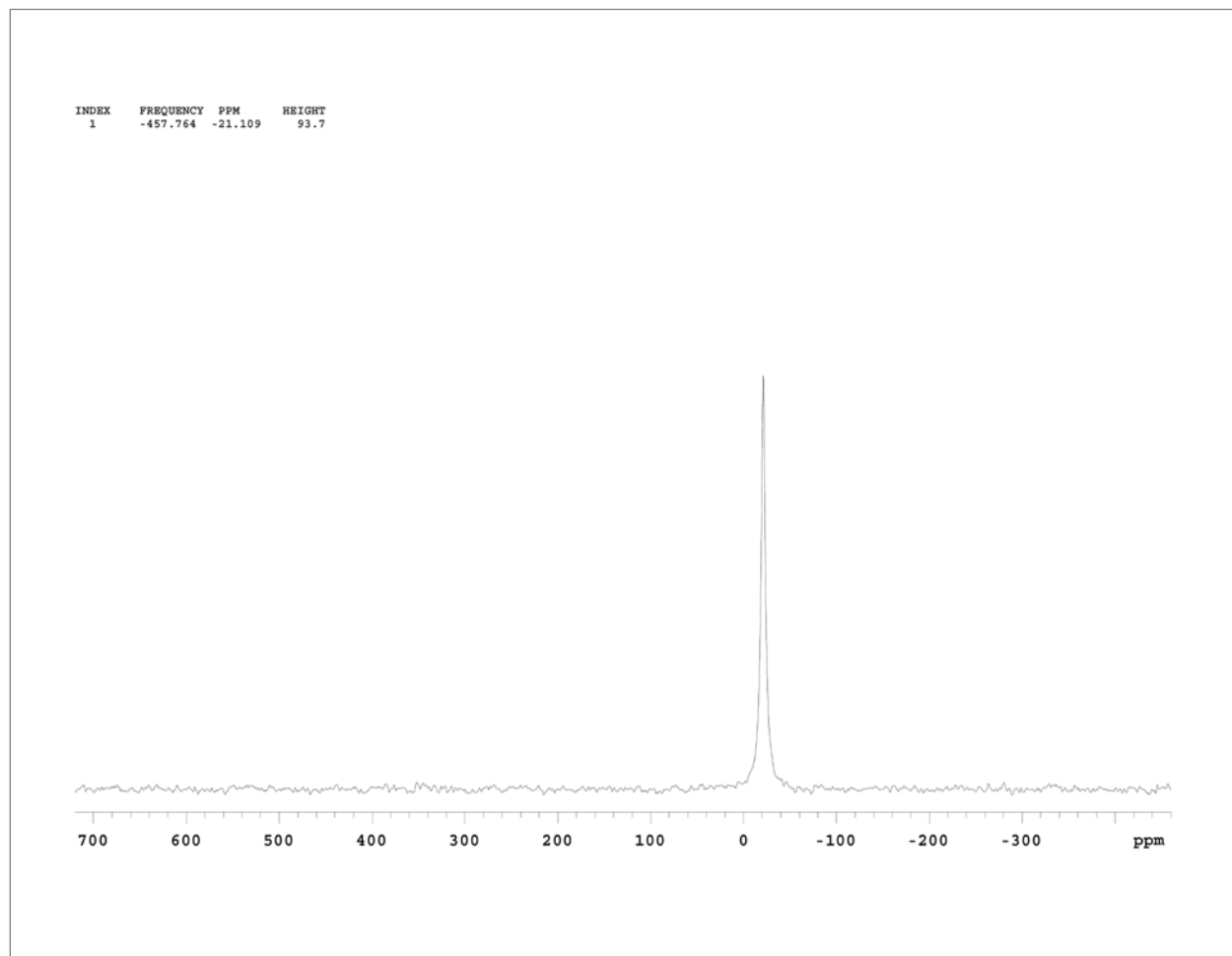
1-Chloro-[2,4-¹⁵N₂]dinitrobenzene / ¹³C NMR

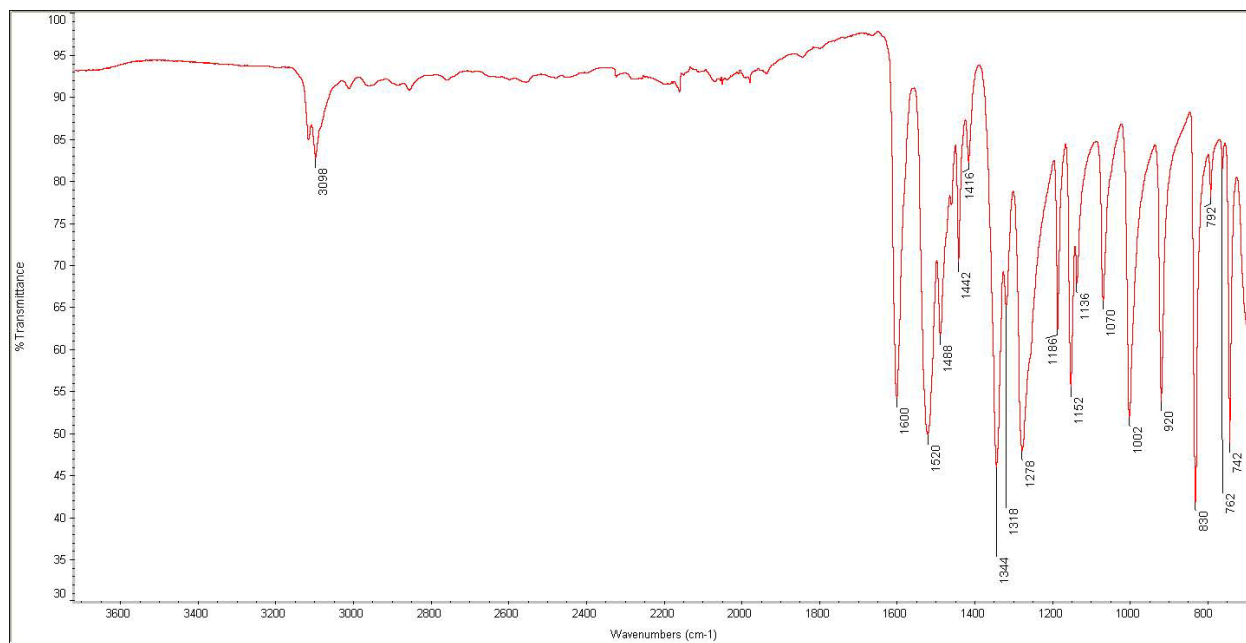
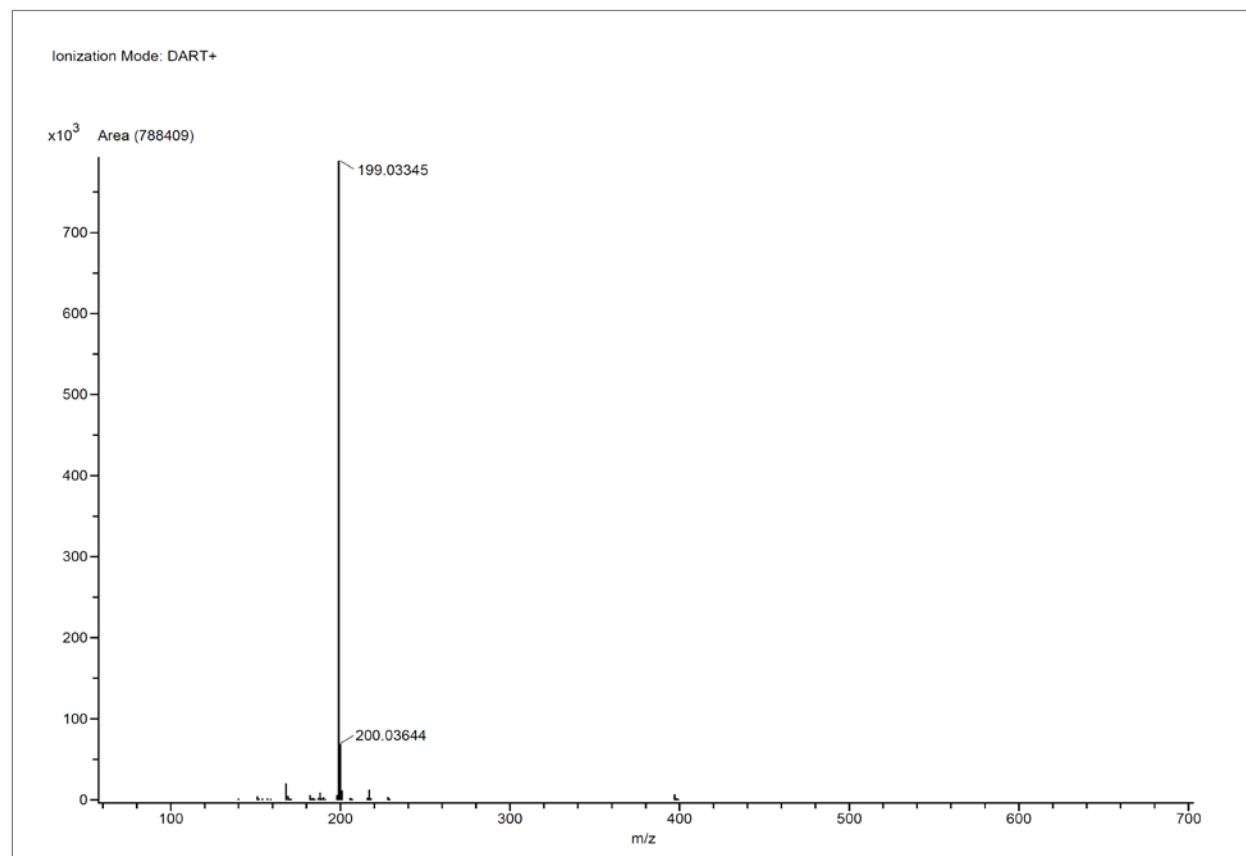
1-Chloro-[2,4-¹⁵N₂]dinitrobenzene / ¹⁵N NMR

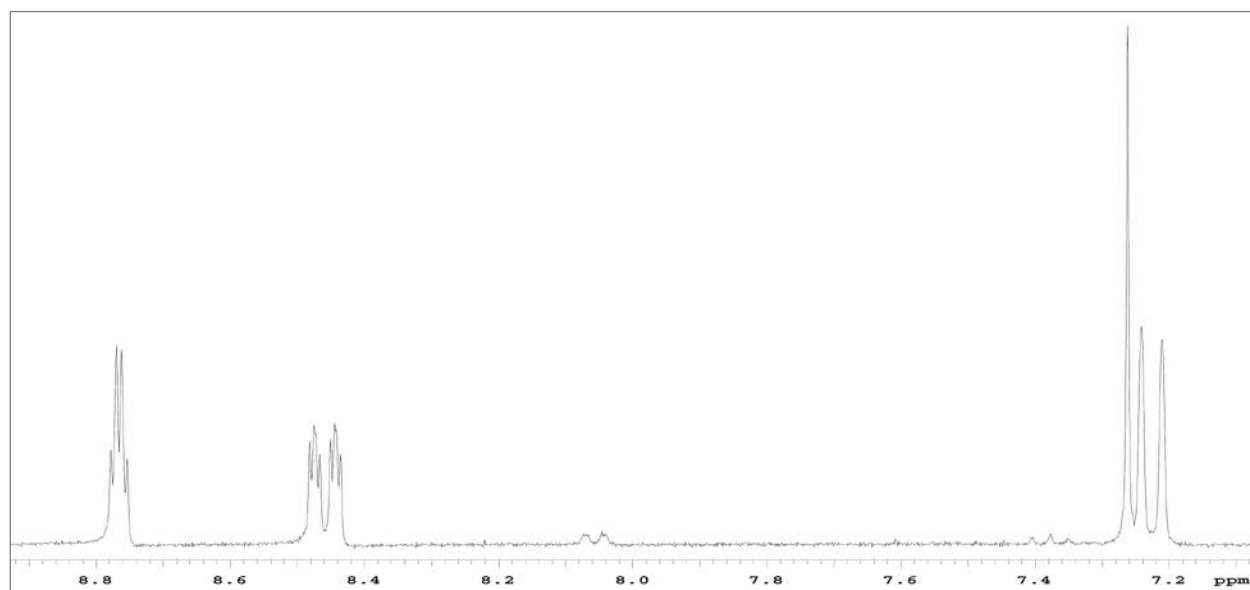
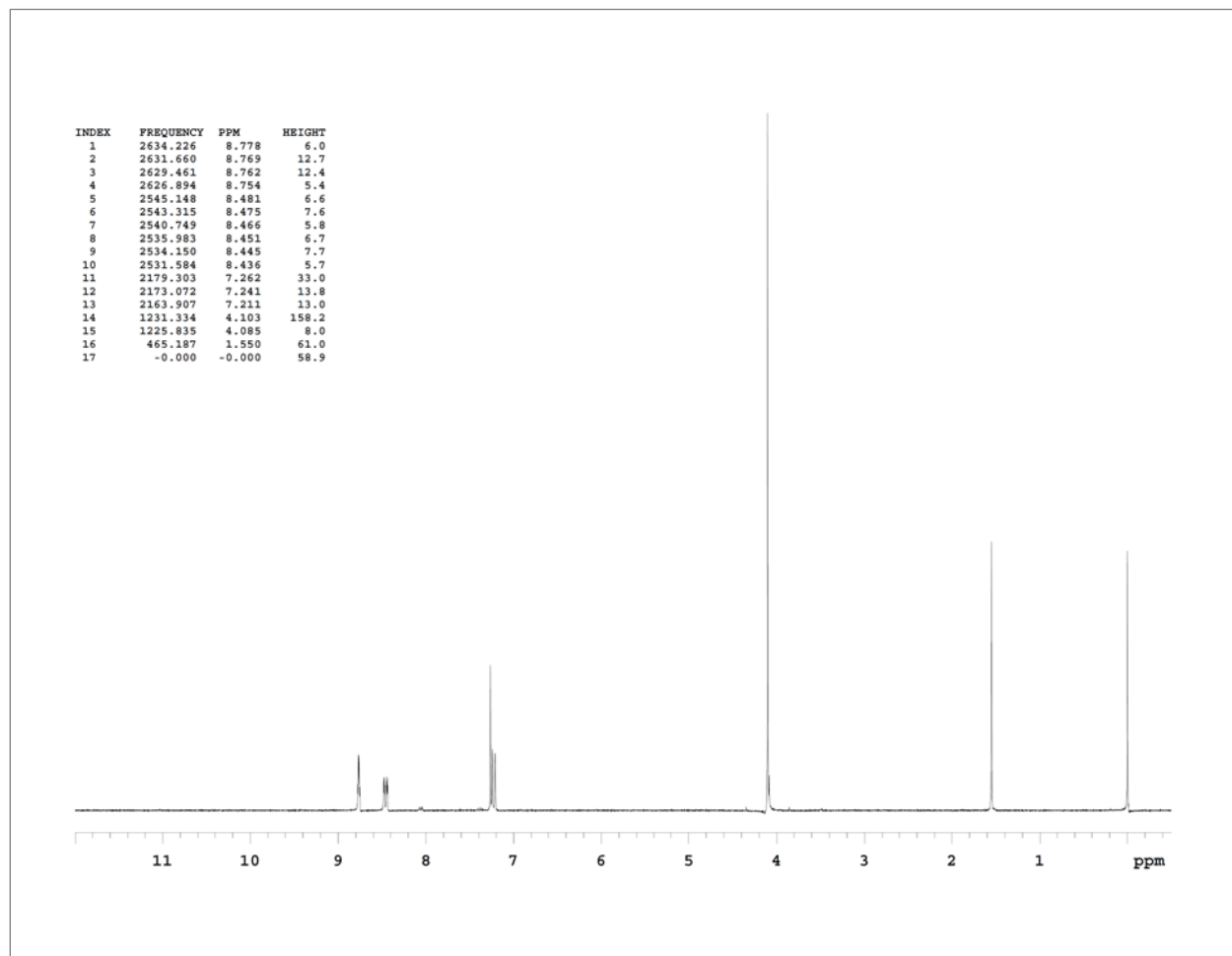
1-Chloro-[2,4- $^{15}\text{N}_2$]dinitrobenzene / IR**1-Chloro-[2,4- $^{15}\text{N}_2$]dinitrobenzene / HRMS**

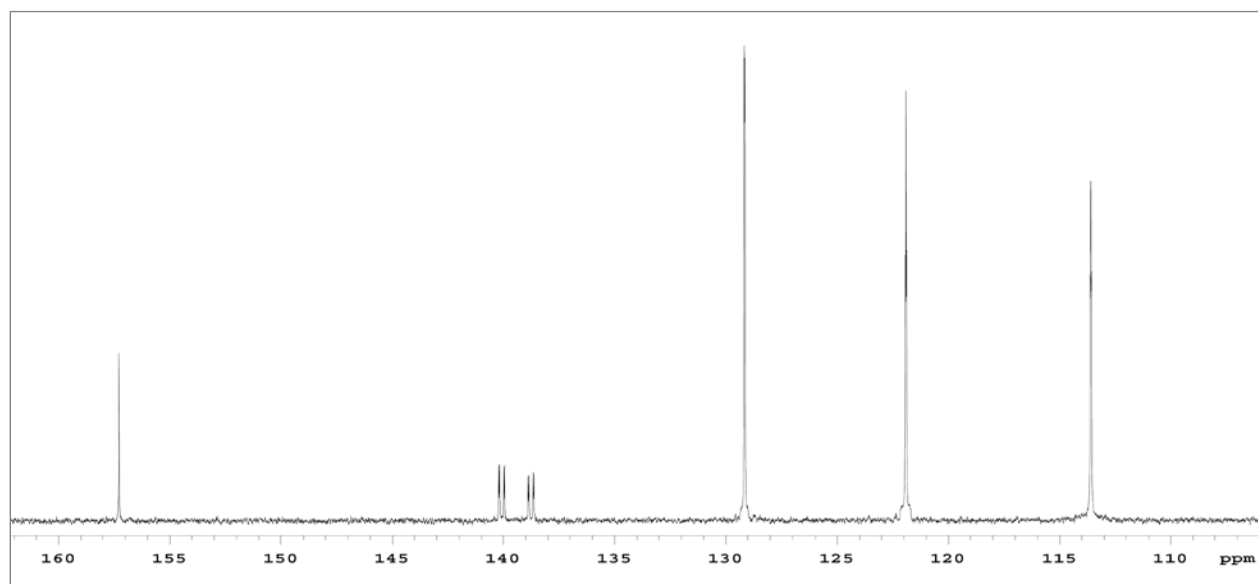
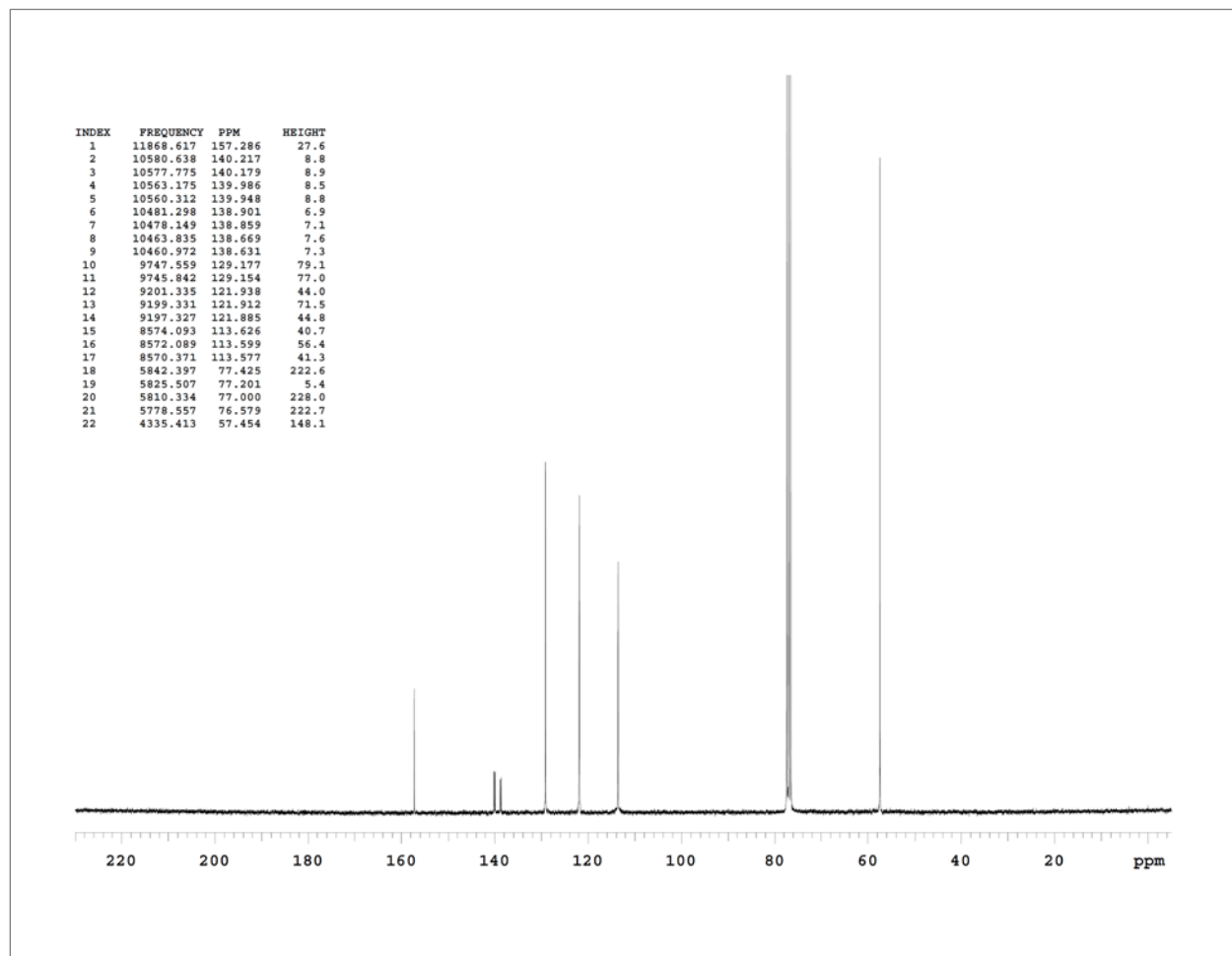
2,4-Dinitroanisole / ^1H NMR

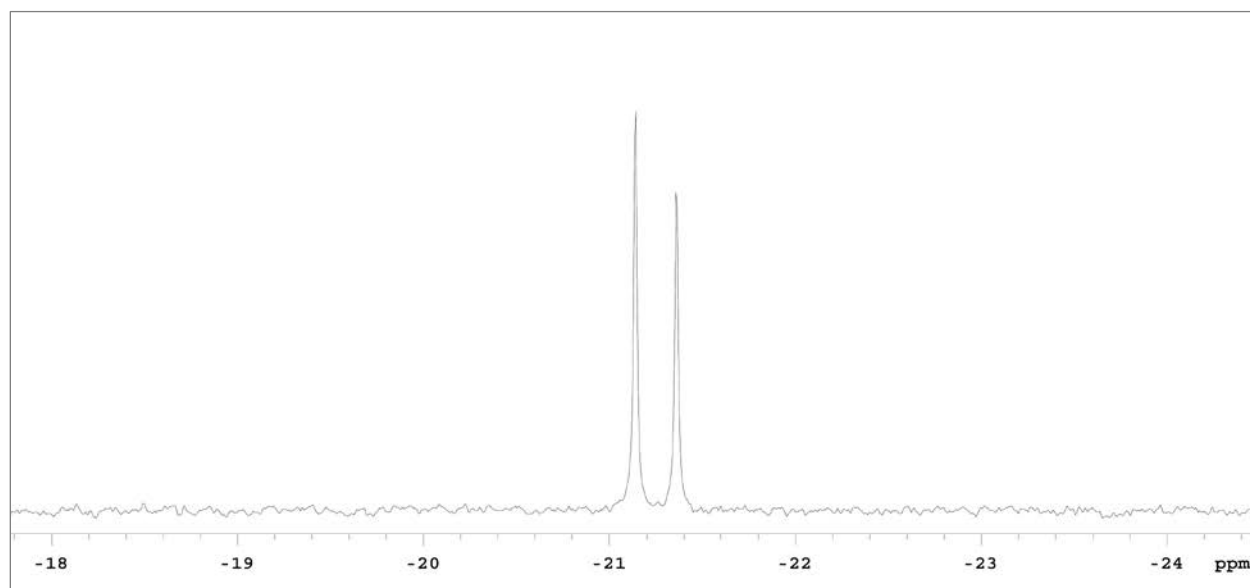
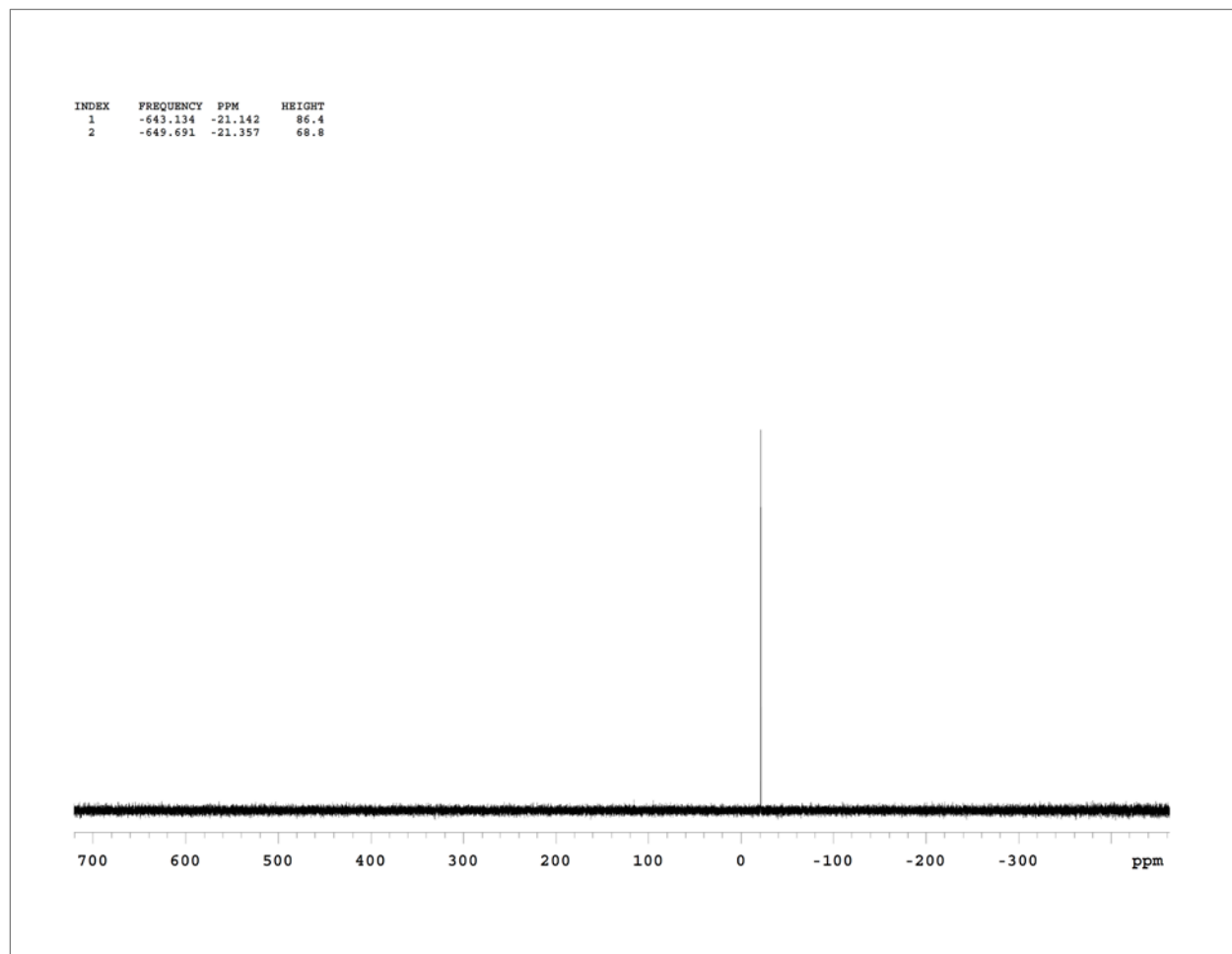
2,4-Dinitroanisole / ^{13}C NMR

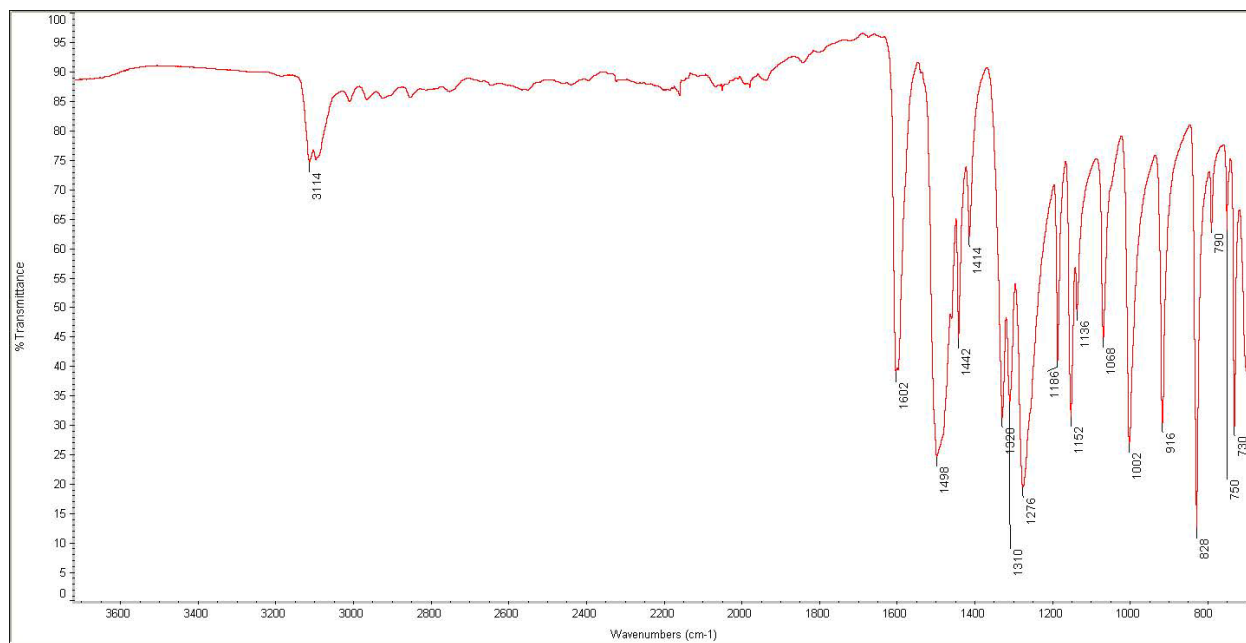
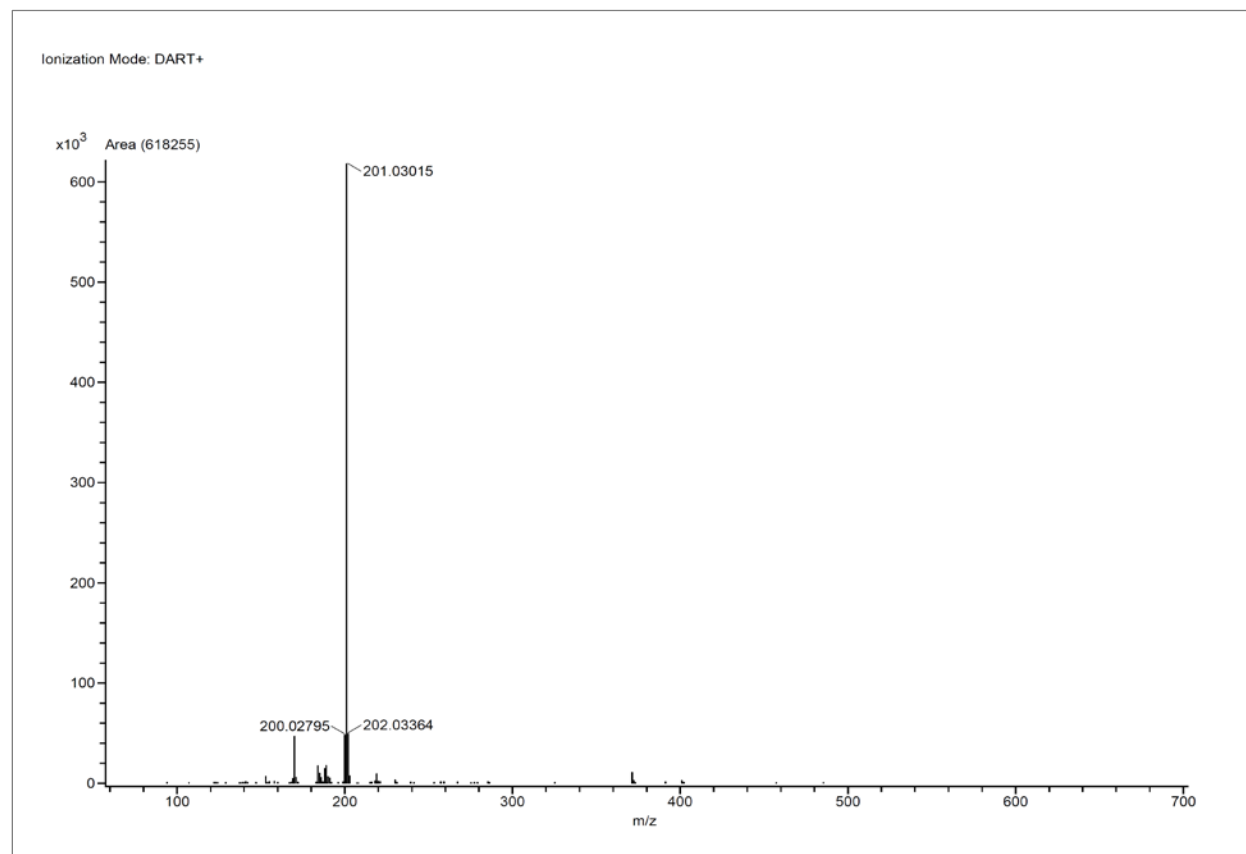
2,4-Dinitroaniso / ^{14}N NMR

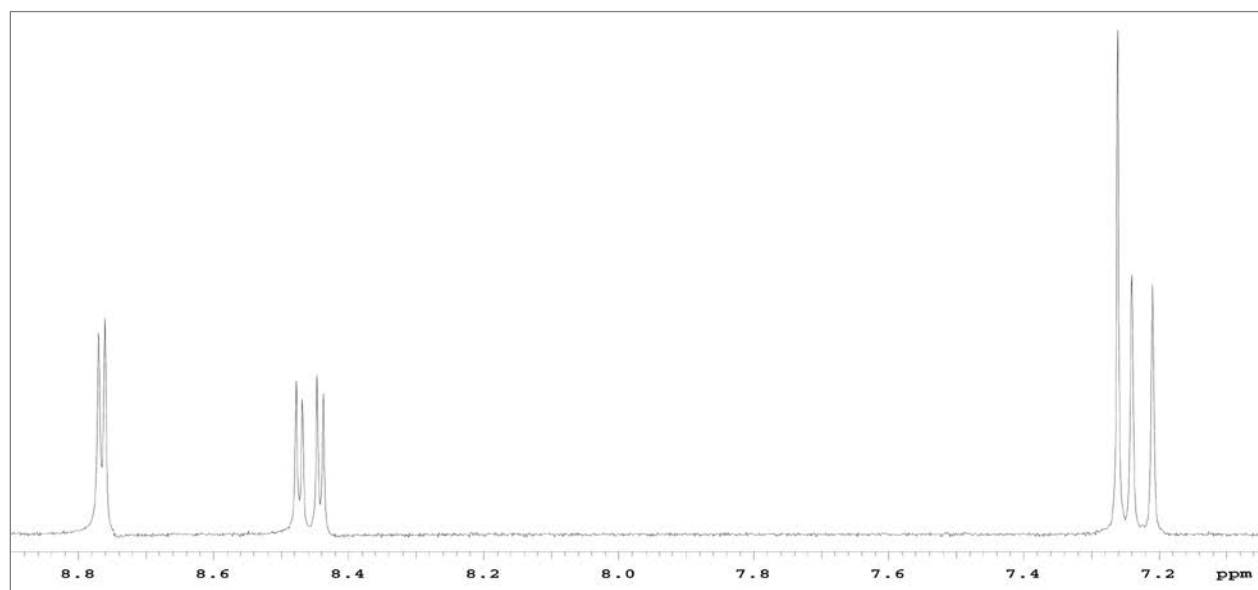
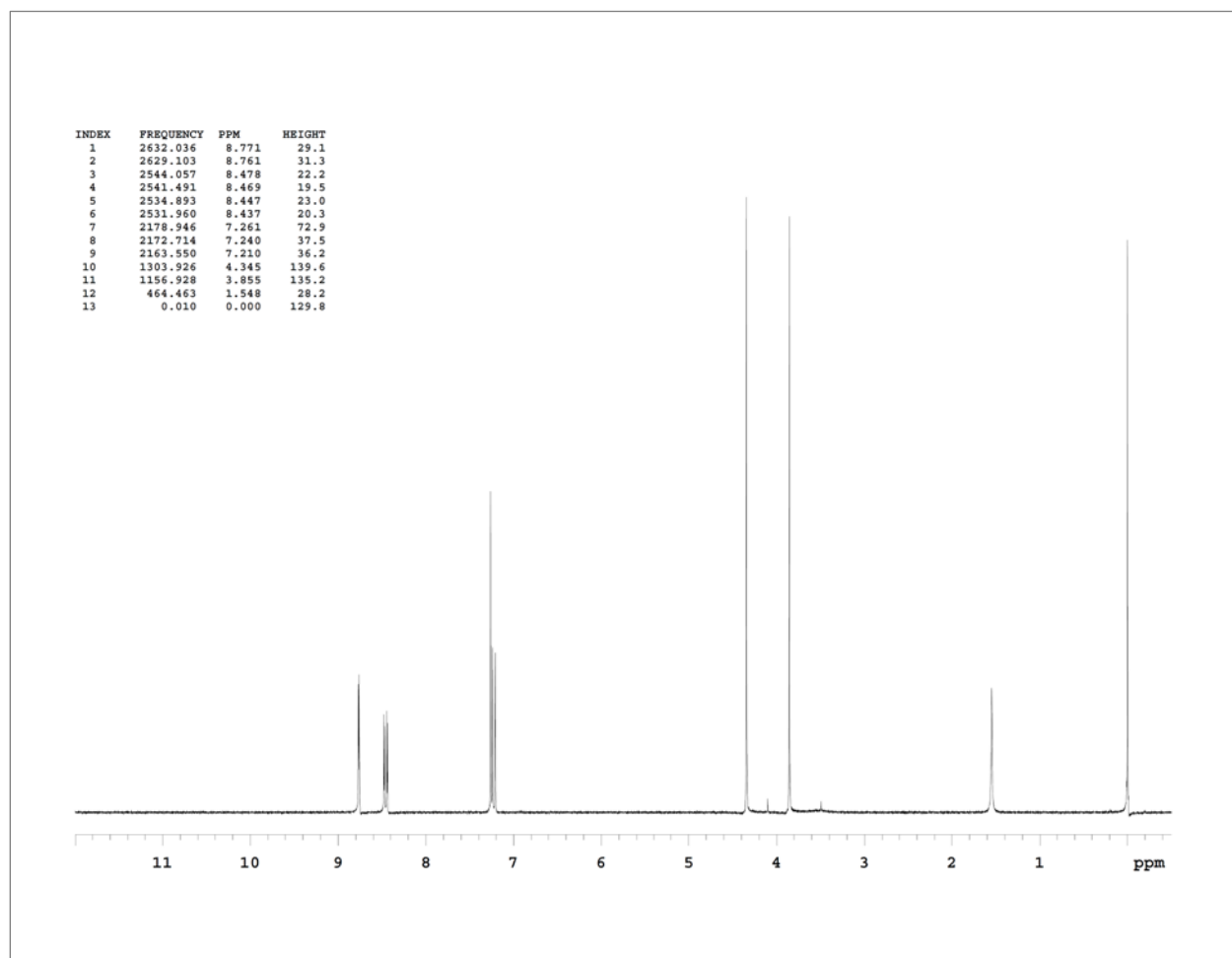
2,4-Dinitroanisole / IR**2,4-Dinitroanisole / HRMS**

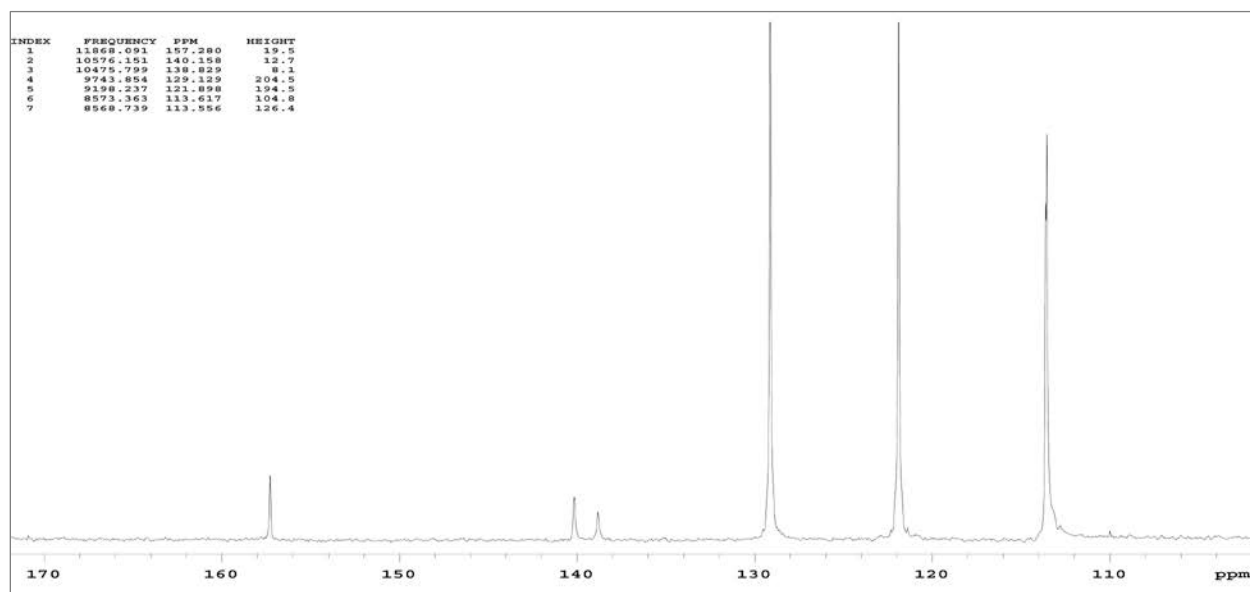
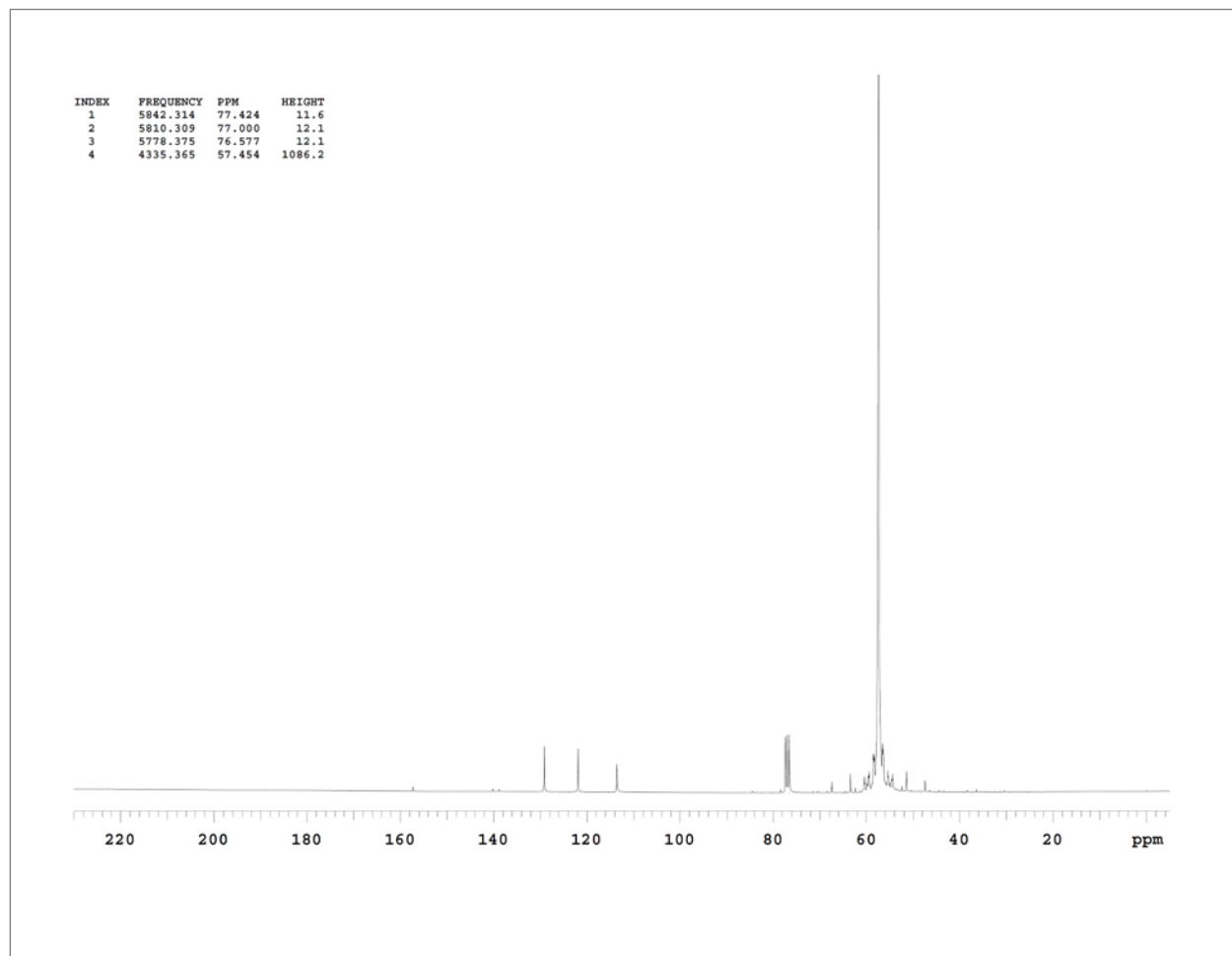
[2,4-¹⁵N₂]Dinitroanisole / ¹H NMR

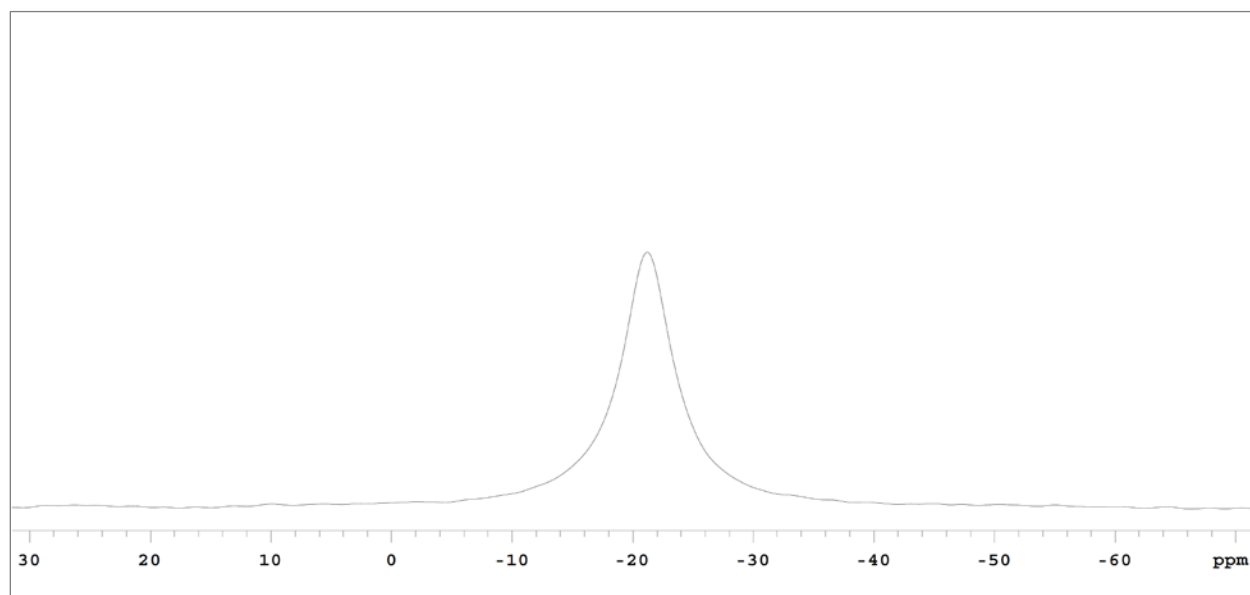
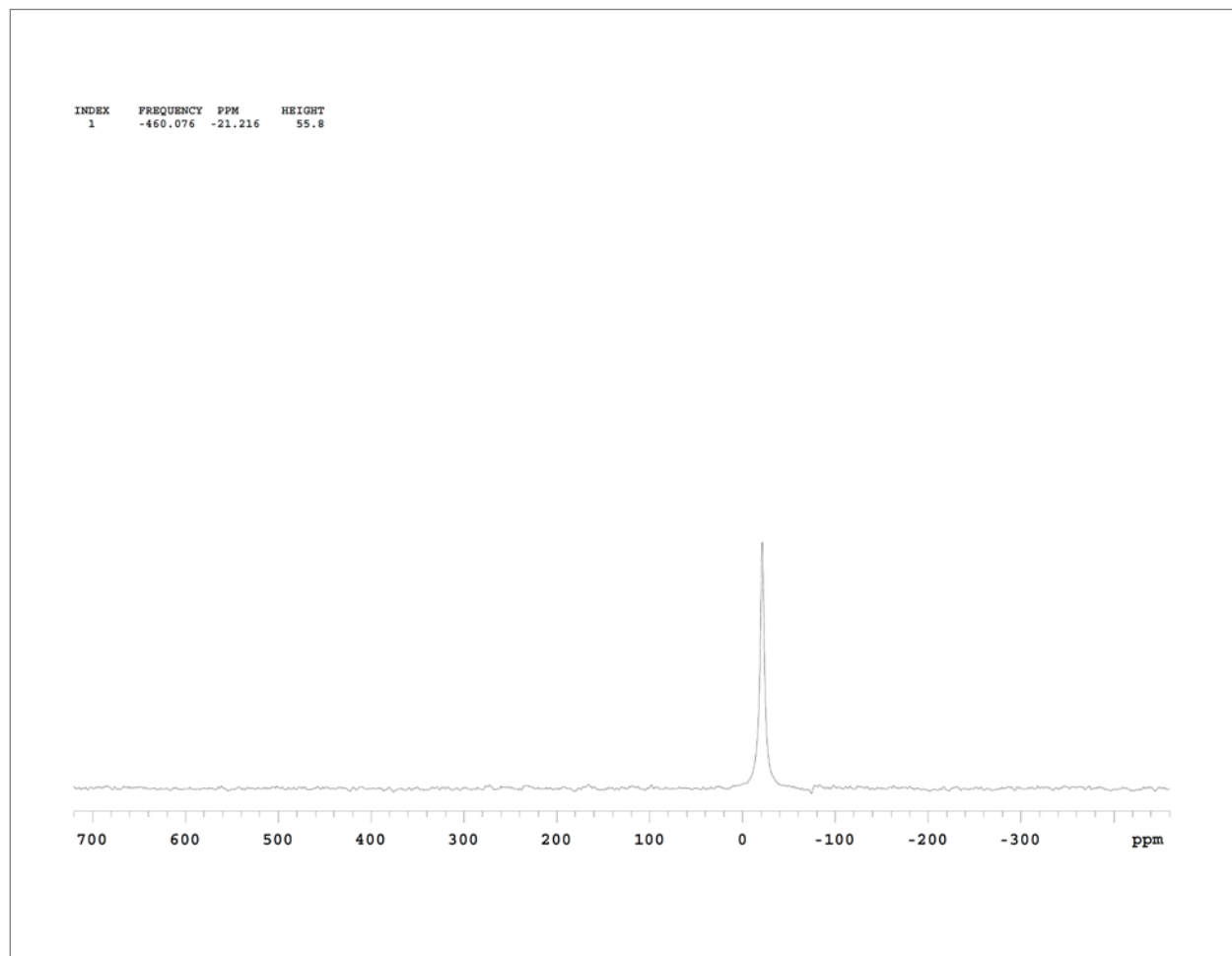
[2,4-¹⁵N₂]Dinitroanisole / ¹³C NMR

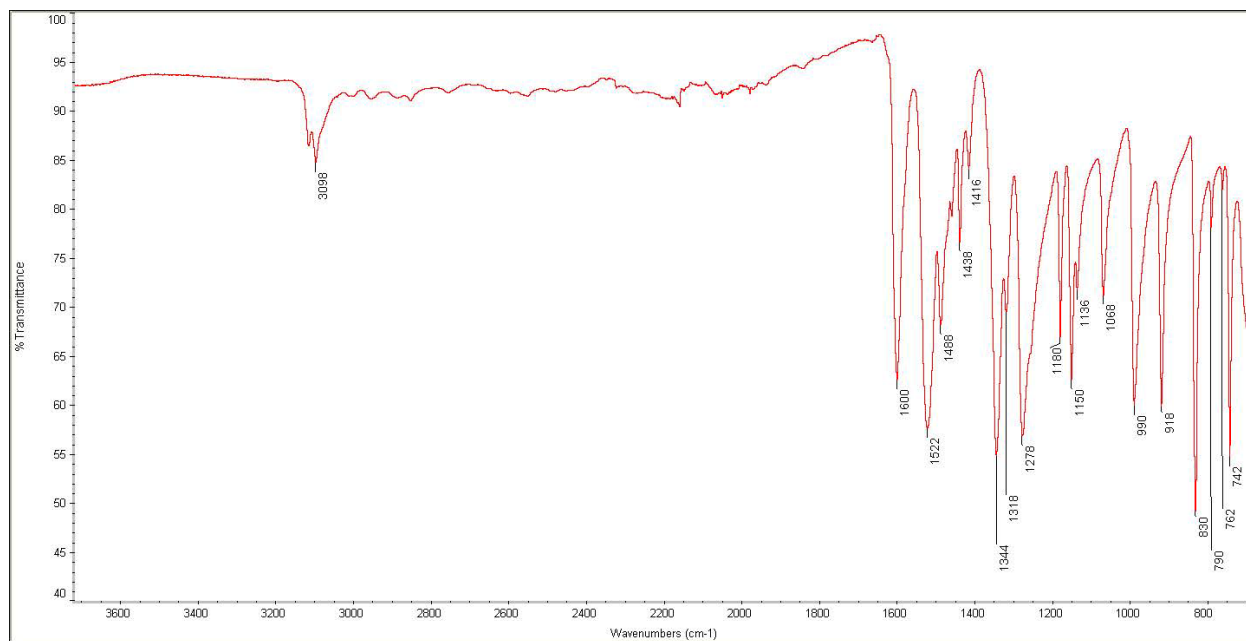
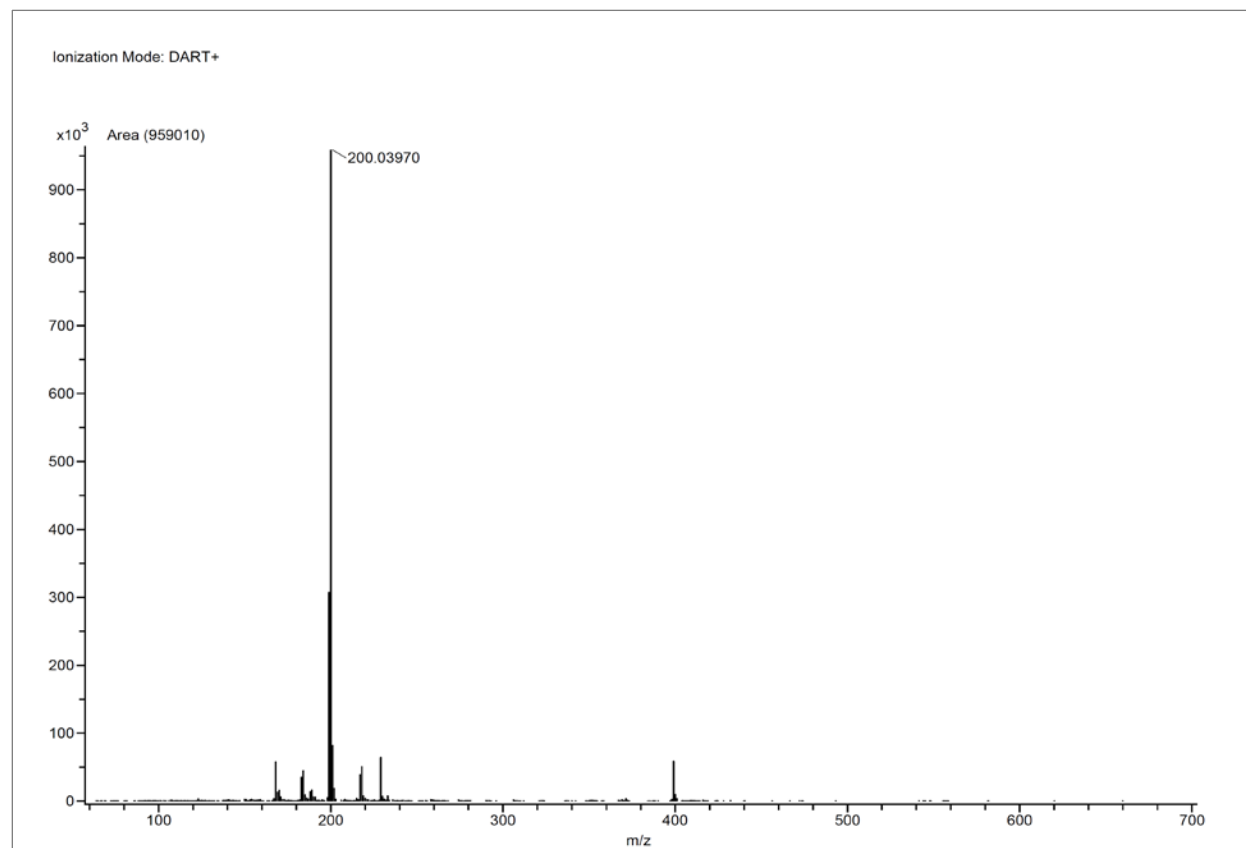
[2,4- $^{15}\text{N}_2$]Dinitroanisole / ^{15}N NMR

[2,4-¹⁵N₂]Dinitroanisole / IR**[2,4-¹⁵N₂]Dinitroanisole / HRMS**

[methyl- ^{13}C]2,4-Dinitroanisole / ^1H NMR

[methyl- ^{13}C]2,4-Dinitroanisole / ^{13}C NMR

[methyl- ^{13}C]2,4-Dinitroanisole / ^{14}N NMR

[methyl-¹³C]2,4-Dinitroanisole / IR**[methyl-¹³C]2,4-Dinitroanisole / HRMS**

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I would like to thank Dr. Sandra Brasfield of the U.S. Army Engineer Research and Development Center (ERDC) Environmental Laboratory, Vicksburg, MS, for her financial support in this effort. I want to acknowledge Dr. Victor Bellitto of the Naval Surface Warfare Center, Indian Head Explosive Ordnance Disposal Technology Division (NSWC IHEODTD) for the Infrared spectra he provided. I would also like to acknowledge Dr. Rebecca Wilson of NSWC IHEODTD for her help and support to maintain the NMR and Mass Spectrometry instruments. Finally, I want to thank Drs. Jason Jouet, Gerardo Pangilinan, and Ruth Doherty of NSWC IHEODTD for their generous support, guidance, and patience.

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