

DTIC FILE COPY

(1)

AD-A191 654

USAARL Report No. 87-13



Dark Adaptation and Recovery from Light Adaptation: Smokers versus Nonsmokers

By
Roger W. Wiley

DTIC
ELECTE
S MAR 24 1988
D

Sensory Research Division
Visual Sciences Branch

September 1987

Approved for public release; distribution unlimited.

88 3 22 070

NOTICE

Qualified Requesters

Qualified requesters may obtain copies from the Defense Technical Information Center, Cameron Station, Alexandria, Virginia, 22314. Orders will be expedited if placed through the librarian or other person designated to request documents from the Defense Technical Information Center.

Change of Address

Organizations receiving reports from the U.S. Army Aeromedical Research Laboratory on automatic mailing lists should confirm correct address when corresponding about laboratory reports.

Disposition

Destroy this report when it is no longer needed. Do not return it to the originator.


Disclaimer


The views, opinions, and/or findings contained in this report are those of the authors and should not be construed as an official Department of the Army position, policy, or decision, unless so designated by other official documentation. Citation of trade names in this report does not constitute an official Department of the Army endorsement or approval of the use of such commercial items.


Human Use

Human subjects participated in these studies after giving free and informed voluntary consent. Investigators adhered to AR 70-25 and USAMRDC Reg 70-25 on Use of Volunteers in Research.

Reviewed:


BRUCE C. LEIBRECHT, Ph.D.
LTC, MS
Director, Sensory Research Division


J. D. LaMOTHE, Ph.D.
COL, MS
Chairman, Scientific Review
Committee


DUDLEY R. PRICE
Colonel, MC
Commanding

REPORT DOCUMENTATION PAGE				Form Approved OMB No. 0704-0188	
1a. REPORT SECURITY CLASSIFICATION Unclassified			1b. RESTRICTIVE MARKINGS		
2a. SECURITY CLASSIFICATION AUTHORITY			3. DISTRIBUTION / AVAILABILITY OF REPORT		
2b. DECLASSIFICATION / DOWNGRADING SCHEDULE					
4. PERFORMING ORGANIZATION REPORT NUMBER(S) USAARL Report No. 87-13			5. MONITORING ORGANIZATION REPORT NUMBER(S)		
6a. NAME OF PERFORMING ORGANIZATION U.S. Army Aeromedical Research Laboratory		6b. OFFICE SYMBOL (If applicable) SGRD-UAS-VS	7a. NAME OF MONITORING ORGANIZATION U.S. Army Medical Research and Development Command		
6c. ADDRESS (City, State, and ZIP Code) P.O. Box 577 Ft. Rucker, AL 36362-5292			7b. ADDRESS (City, State, and ZIP Code) Ft. Detrick Frederick, MD 21701-5012		
8a. NAME OF FUNDING / SPONSORING ORGANIZATION		8b. OFFICE SYMBOL (If applicable)	9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER		
8c. ADDRESS (City, State, and ZIP Code)			10. SOURCE OF FUNDING NUMBERS		
			PROGRAM ELEMENT NO. 62777A	PROJECT NO. A 3E162777879	TASK NO. BG
			WORK UNIT ACCESSION NO. 164		
11. TITLE (Include Security Classification) Dark Adaptation and Recovery from Light Adaptation: Smokers Versus Non-Smokers					
12. PERSONAL AUTHOR(S) Roger W. Wiley					
13a. TYPE OF REPORT		13b. TIME COVERED FROM _____ TO _____		14. DATE OF REPORT (Year, Month, Day) September 1967	
15. PAGE COUNT 25					
16. SUPPLEMENTARY NOTATION					
17. COSATI CODES			18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number)		
FIELD	GROUP	SUB-GROUP			
06/04					
09/05					
19. ABSTRACT (Continue on reverse if necessary and identify by block number) Since the published data concerning the effects of smoking on visual sensitivity at night are inconsistent, a new study was initiated to investigate this question. Thirty Army aviators between the ages of 19 and 39 volunteered to participate in this study. Of these subjects, 15 smoked and 15 were non-smokers. Each subject was seated in a light-controlled room and exposed to a standardized bright light for 5 minutes. Immediately after the bright light was extinguished, the subject's visual sensitivity was tested by gradually increasing the intensity of a test light until the subject could see it. This was continued over a period of 35 minutes by which time the subjects had reached their maximum light sensitivity. Each subject then wore a pair of AN/PVS-5 Night Vision Goggles for 5 minutes after which his visual sensitivity again was tested for 20 minutes. Our data do not show any differences in visual sensitivity between aviators who smoke and those who do not smoke. Blood samples were analyzed to compare serum levels of nicotine, cotinine and carboxyhemoglobin with the visual data. Again, no correlation exists between					
20. DISTRIBUTION / AVAILABILITY OF ABSTRACT <input type="checkbox"/> UNCLASSIFIED/UNLIMITED <input type="checkbox"/> SAME AS RPT. <input type="checkbox"/> DTIC USERS				21. ABSTRACT SECURITY CLASSIFICATION	
22a. NAME OF RESPONSIBLE INDIVIDUAL Roger W. Wiley			22b. TELEPHONE (Include Area Code) (205) 255-6810		22c. OFFICE SYMBOL SGRD-UAS-VS

19. ABSTRACT Continued

sensitivity and blood measures related to smoking. Aviators who smoke reach the same level of sensitivity to light as non-smokers and they do so in the same amount of time. Visual recovery after wearing the Night Vision Goggles also followed the same time course regardless of smoking history. The conclusion from these data is that light sensitivity, the ability to see the dimmest lights at night, is independent of smoking history.

Table of Contents

Introduction	3
Materials and methods.	4
Results.	6
Discussion	10
Conclusions.	12
References	13



Accession For	
NTIS CRA&I	<input checked="" type="checkbox"/>
DTIC TAB	<input type="checkbox"/>
Unannounced	<input type="checkbox"/>
Justification	
By	
Distribution /	
Availability Codes	
Dist	Avail and/or Special
A-1	

This page left blank intentionally.

Introduction

The possible biological effects of smoking tobacco products have been studied intensively from a variety of different aspects. However, despite many efforts, the reported changes in visual sensory functions are inconsistent. The earlier reports indicated subjects who smoke cigarettes demonstrated reduced visual thresholds. For example, McFarland and coworkers (1944, 1953, 1970) reported a loss in visual sensitivity associated with smoking cigarettes. They used, as a measure of visual sensitivity, discrimination thresholds, i.e., the ability to detect light stimuli presented against backgrounds of various brightnesses. These authors stated that they could detect a change in the discrimination threshold if the subject smoked just one cigarette. Since no change in threshold was reported if the subjects did not inhale the smoke, the authors concluded that carboxyhemoglobin saturation was the cause of reduced visual sensitivity. They reached this conclusion after considering that nicotine still reaches the blood even without inhaling the smoke while carboxyhemoglobin saturation is not present without inhalation. In partial support of this conclusion, Sheard (1946) reported that the immediate effect of inhaling smoke was a reduction of from 0.25 to 0.75 log units in absolute light sensitivity. However, he ascribed his results to nicotine since his data indicated no effect if the nicotine was filtered from the smoke. These early reports have not remained unchallenged.

In contrast to the previous reports, Troemel, Davis and Hendley (1951) found that nicotine actually facilitated the course of dark adaptation in their subjects. Johansson and Jansson (1964) used a visual discrimination threshold and a repeated measures design to assess smoking effects and failed to show any change in thresholds after their subjects smoked cigarettes. Calissendorff (1977) also used a repeated measures design and reported a slight reduction in mesopic, but not scotopic, light sensitivity when measured after his subjects smoked cigarettes. Durazzini, Zazo, and Bertoni (1975) attempted to correlate the presence of thiocyanates in the urine secondary to smoking with several measures of visual function. These authors reported about half of their subjects demonstrated a reduction in absolute visual sensitivity thresholds after smoking cigarettes. In comparison to these investigations which have addressed primarily scotopic or mesopic function, Fine and Kobrick (1987) studied the effects of smoking on visual contrast sensitivity which is primarily subserved by the photopic system. No differences in contrast sensitivity were found in their test subjects pre- and postcigarette smoking. However, habitual smokers had slightly lower contrast sensitivities to certain spatial frequencies.

While many of the above investigations have used only a limited number of subjects or examined either the scotopic or photopic system using psychophysical procedures, Luria and McKay (1979a, b) used both psychophysical and electrophysiological techniques to assess the effects of carbon monoxide exposure on smokers and nonsmokers. Using age-matched subjects (40 smokers and 40 nonsmokers), they tested scotopic sensitivity, reaction time, color vision, visually evoked cortical potentials, and EEGs. As a group, the smokers had a poorer scotopic sensitivity score and a slower reaction time. The remaining tests in their battery did not show any differences between the two groups. Further, their results did not demonstrate any trends to indicate that a history of smoking caused a cumulative decrement in visual sensory function.

There is a growing body of evidence to indicate that smoking cigarettes can cause many different physical infirmities (US Surgeon General Report, 1979). However, the effects of smoking on visual sensory function are equivocal. A review of the published evidence presents a confusing picture. Cigarette smoking does, or possibly does not, cause a change in visual perceptual processes; if visual processes are changed, they might be enhanced or reduced. Finally, if a change occurs, the photopic, mesopic, and/or scotopic systems might be affected.

The objective of the present investigation is to determine if there are changes in scotopic sensitivity and its recovery which possibly could be attributable to chronic tobacco use. To assess this, we measured absolute scotopic sensitivity using standardized clinical testing procedures in a group of Army aviators who smoke cigarettes and compared those results with an age-matched group of aviators who do not smoke. Additional data were obtained to determine if differences exist between the two groups in recovery of absolute scotopic sensitivity after viewing a military electro-optical device (AN/PVS-5 night vision goggles).

Materials and methods

Subjects

Thirty Army aviator volunteers served as subjects for this study. Of these, 15 subjects did not smoke or use any tobacco products and 15 smoked cigarettes. All among the smoking group had smoked for more than 1 year with 11 of them having smoked for more than 10 years. Daily usage ranged from about 10 cigarettes to more than 40. The ages among the smokers ranged between 28 and 38 years (mean = 32.87 years) and among the nonsmokers between 19 and 39 years (mean = 30.20 years).

Procedures

Since the purpose of this study was to investigate the cumulative rather than immediate effects of cigarettes, no attempt was made to control the subjects' smoking prior to data collection. However, the testing procedures required approximately 2 hours during which the subjects were not allowed to smoke. The testing schedule required complete data collection on two subjects daily, and subjects from the smoking and nonsmoking groups were interspersed.

An identical test procedure was followed on every subject. When the individual arrived at the laboratory, he was thoroughly briefed on the purpose of the experiment and trained on the observations required of him. He then sat in a dimly illuminated room (5.12 footcandles) for 5 minutes. Following this period, all lights were extinguished in the specially prepared dark room and the subject remained in the dark for 3 minutes. During this time, his left eye was occluded, and he positioned himself comfortably in front of the hemispherical ganzfeld of a clinical Goldmann/Weekers Adaptometer. The instrument then was turned on and the subject was light adapted by staring at the uniformly illuminated hemisphere having a brightness of 312 footlamberts. In accordance with standard clinical testing procedure, this period of light adaptation lasted for 5 minutes, after which the hemisphere lighting was extinguished and the fixation light became visible. Testing light sensitivity thresholds started immediately.

An ascending method of limits was used to measure the threshold with the subject indicating when the test stimulus became visible by tapping on the instrument table. The angular subtense of the test stimulus was 10 degrees and it stimulated a portion of the retina approximately 10 degrees below the fovea. During the first 15 minutes of dark adaptation, the threshold was measured every 15 seconds. Measurements were made every 30 seconds during the remainder of the 35 minutes.

After completing the 35 minutes of threshold testing, the subject donned a pair of AN/PVS-5 night vision goggles (NVGs). He was instructed simply to observe objects in the darkened test room using the infra-red source incorporated into the NVGs to illuminate them. The goggle output tubes provide a brightness of 0.098 footlambert and the subject was exposed to this brightness for 5 minutes. Following the 5 minute NVG exposure, the subject immediately positioned himself in the Dark Adaptometer again and threshold testing was resumed for an additional 20 minutes to assess the speed with which he recovered his absolute sensitivity.

When the psychophysical testing had been completed, the

subject was allowed to light adapt and a medical technician, using standard medical laboratory technique, took two venous blood samples. A 15 ml sample was forwarded to the Alabama Reference Laboratory which had been contracted to analyze each sample for nicotine and cotinine levels. A 7 ml sample was analyzed immediately to determine the percentage of carboxyhemoglobin (COHb).

Results

The primary results from this study are shown in Figure 1. In this figure, the changing threshold light sensitivity is graphed as a function of time in the dark. The averaged data obtained from the smoking and nonsmoking groups are practically identical. Both groups started at the same level of desensitization following the pretest bleaching exposure and achieved an approximate 4 log unit increase in visual sensitivity, demonstrating an average time to absolute sensitivity of about 28 minutes. There was an intermediate window of time (9 minutes to 24 minutes) during which the averaged thresholds from the nonsmoking group showed a very slight, and statistically insignificant, greater sensitivity.

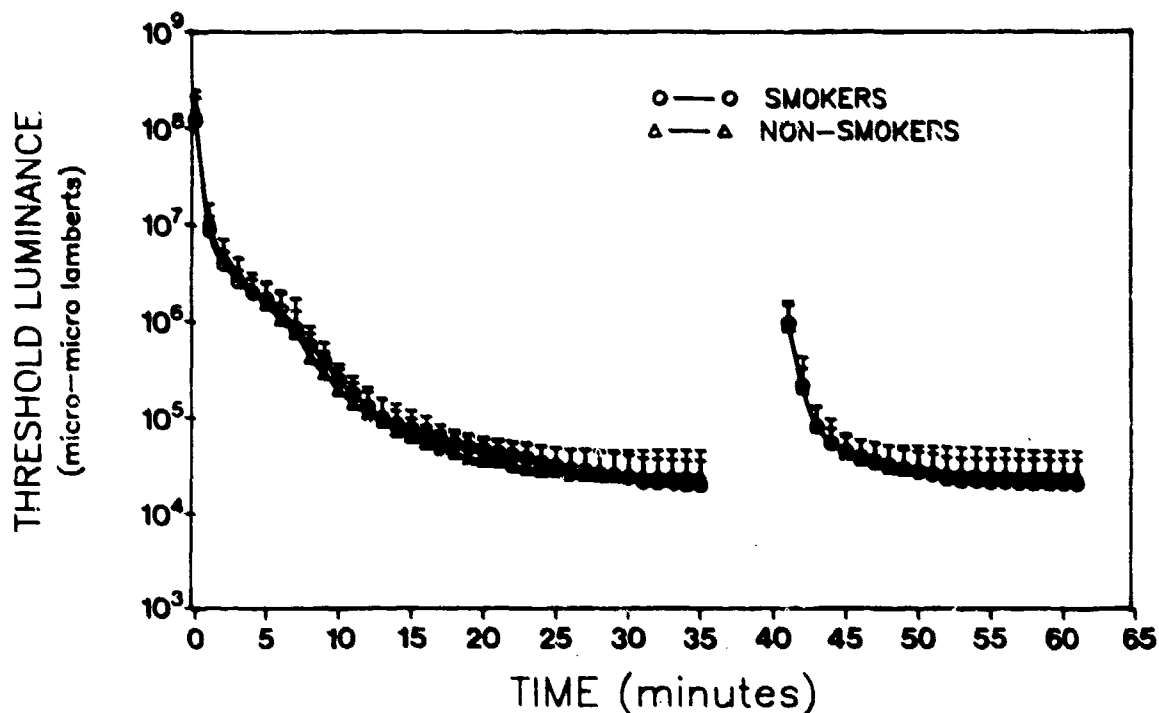


Figure 1. Average threshold luminance for smokers (circles) and nonsmokers (triangles) following white light bleach or night vision goggle exposure. Brackets indicate +1 standard deviation.

After 35 minutes of testing, the subjects used the AN/PVS-5 night vision goggles (NVGs) to view randomly around the darkened test room. The infrared light emitting diode provided in the NVGs was used as the illumination source. By doing this, the output phosphor (S20) screen had a luminance of 0.098 foot-lambert to which the subjects were exposed for five minutes. Data showing the visual sensitivity recovery from this exposure also are shown in Figure 1. Again, no differences between the smokers and nonsmokers were revealed, the two averaged curves being practically identical. After viewing with the NVGs, the subjects were reduced to about the same level of sensitivity which they previously had demonstrated at the 6-minute point during the initial testing following a more intense bleaching exposure. However, recovery back to baseline sensitivity following the NVGs exposure was much more rapid. This is shown in Figure 2 and has been reported previously (Glick, et al., 1975). Since the two groups' data were almost identical, only

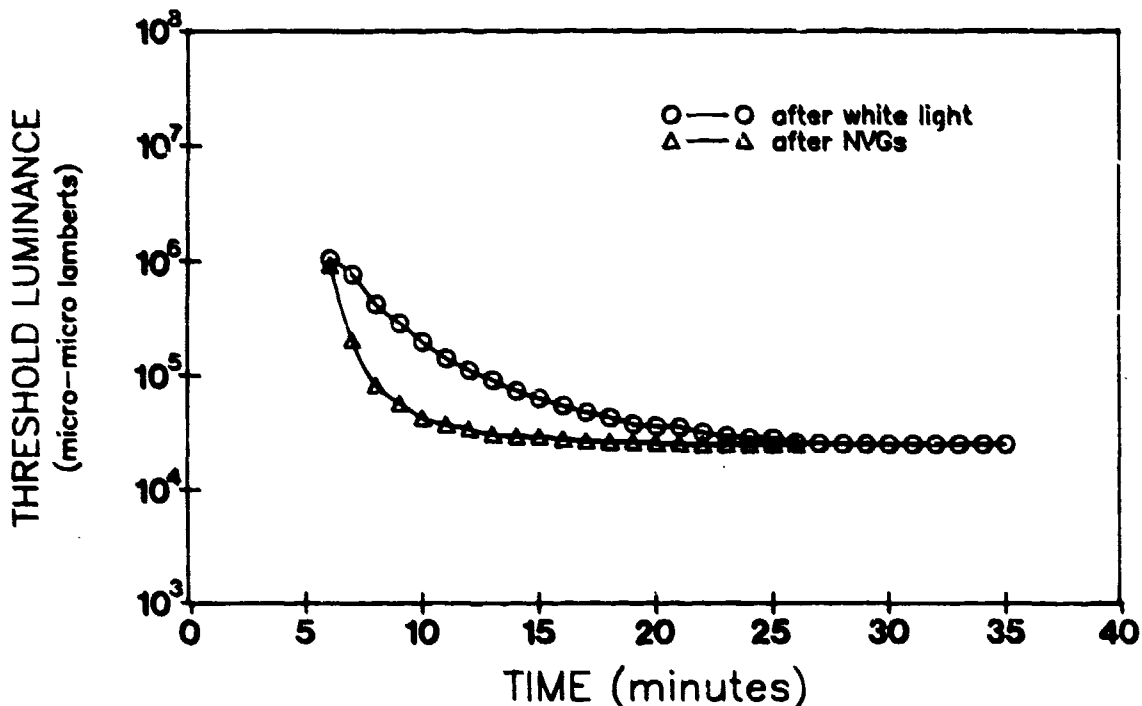


Figure 2. Comparison of recovery of light sensitivity following white light bleach (circles) or night vision goggle exposure (triangles). The abscissa values (minutes) relate to the time following the white light bleach. To allow comparison, the curve obtained after night vision goggle exposure has been shifted laterally along the abscissa so that initial sensitivity is the same for both conditions.

the smokers' data are shown. In Figure 2, the initial threshold data are plotted from 6 minutes until 32 minutes. As stated previously, maximum sensitivity was reached by the 28th minute of testing. For comparison, the threshold recovery data following exposure with the NVGs also are shown in the figure. In this latter condition, threshold recovery is much more rapid, approaching the maximum sensitivity within 5 minutes after removing the NVGs.

Since the curves shown in Figure 1 represent grouped data which conceivably could mask subtle individual effects, the absolute sensitivity thresholds for each of the subjects who smoked were plotted with the results from their respective blood analyses. Figures 3A, B, and C show these thresholds plotted against the blood nicotine, cotinine, and carboxyhemoglobin results from each of the subjects. These truly are scattergrams, showing no correlation or even gross trends between visual threshold and the several physiological byproducts which presumably are related to smoking history.

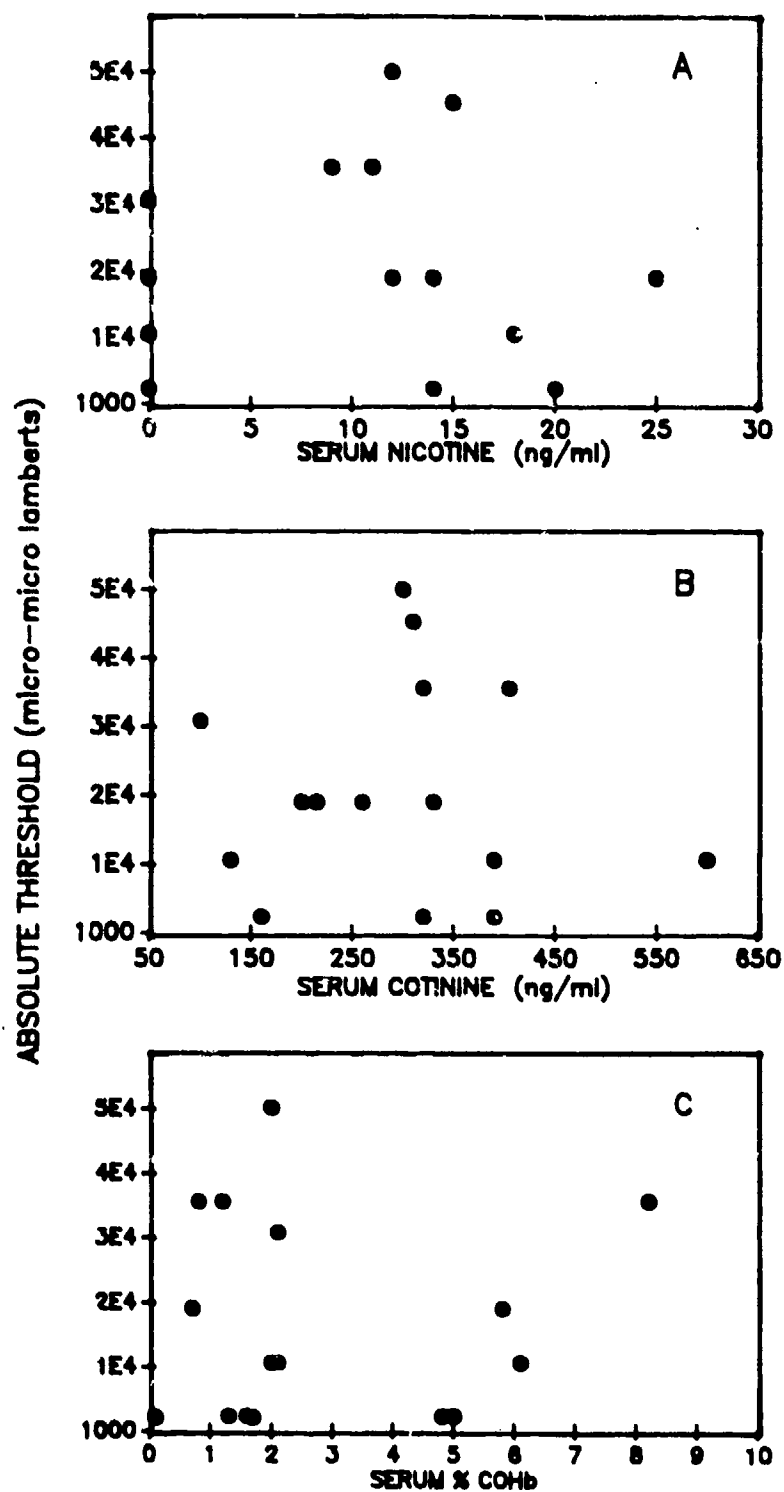


Figure 3. Absolute threshold sensitivities as a function of serum nicotine (A), serum cotinine (B), and serum carboxyhemoglobin (C).

Discussion

The impetus for this investigation has been provided by considerations at the Department of the Army staff level to broaden the restrictions on smoking among Army aviators. At present, smoking is not allowed in Army aircraft during flight. A further restriction under consideration would be to not allow any smoking by Army aviators at any time, both official and personal. By this restriction, smoking tobacco could be the basis for nonselection for aviation training or removal from flight status if already rated.

A restriction on tobacco products would significantly impact the personal lives and professional careers of the affected aviators. Such a restriction should not be taken precipitously without clear indications that the use of tobacco products negatively affect military performance or endanger mission accomplishment. There are many precedents for prohibition based upon potentially compromising performance. Almost simultaneous with the dawn of aviation, the use of alcohol along with or prior to operating an aircraft has been forbidden. However, the adverse sensory and motor effects of alcohol are well-documented (Collins, et al., 1987). That is not the case with tobacco. As discussed previously, the visual sensory effects of tobacco are contradictory. Several investigations have reported a reduction in absolute light sensitivity with smoking while others have failed to show any change in threshold or even showed a facilitation. Among the investigations which have reported a visual change, the visual change has been variously photopic, mesopic, or scotopic and the effect has been attributed to nicotine or carboxyhemoglobin.

The results from the present investigation which are shown in Figure 1 support previous reports of no change in visual threshold secondary to tobacco use. The average sensitivity profiles are practically identical over the course of dark adaptation for the two test groups. In addition, exposure by the AN/PVS-5 NVGs subsequent to reaching absolute light sensitivity caused the same average visual desensitization in the smoking and non-smoking groups and the measured recovery of sensitivity occurred at the same rate. The data shown in Figure 2 are similar to comparison curves reported previously by Glick, et al. (1974).

As shown in Figure 2, recovery of visual threshold following exposure to the light output from the NVGs is much faster than recovery following the white light initial bleaching. As mentioned in the earlier report, the more rapid recovery from NVG exposure possibly can be attributed to the narrower wavelength

band of the NVG output (S20 phosphor). While this would not affect rod function, separate cone populations might be differentially influenced. Although the NVG output is quite dim (0.098 footlambert), it is definitely photopic as evidenced by the green color perception resulting. However, an equally acceptable explanation is provided by a consideration of neural versus photochemical adaptation. It is possible that the visual desensitization after exposure to the dim NVG tube is caused by a change in the neural gain of the visual system rather than a change in the bleached versus unbleached retinal photopigments. By this reasoning, the recovery would be faster because of the more rapid neural recovery rather than a change in photopigment state.

Realizing that grouped data analyses might fail to reveal subtle threshold changes among the smokers and that self-reports of smoking history would not be sufficiently reliable or quantitative, venous blood samples were taken from each subject. These samples were used to analyze sera concentrations of several contaminants resulting from tobacco use. Both carboxyhemoglobin and nicotine previously have been considered to be implicated in changes in visual function. Unfortunately, both of these have relatively short plasma half-lives and our subjects were prevented from smoking for at least two hours during the study. Therefore, we also measured cotinine, a major metabolite of nicotine, which has a much longer life (Pojer, et al., 1984). However, the results shown in Figure 3 indicate that there was no correlation between any of these products and absolute threshold in our subjects.

Conclusions

Our data indicate that there is no difference in visual function between smokers and nonsmokers when the measures of visual function are absolute light sensitivity and rate of recovery of sensitivity after light exposure. There is a growing body of evidence that use of tobacco products has a variety of negative health effects. Also, the immediate physiological consequences of smoking may or may not degrade visual perception. The present data show that there are no cumulative effects of smoking which degrades light sensitivity. Therefore, changes in visual function related to chronic cigarette smoking do not appear to provide a useful basis for prohibiting cigarette use.

REFERENCES

- Calissendorff, B. 1977. Effects of repeated smoking on dark adaptation. Acta Ophthalmologica. 55:261-268.
- Collins, W.E., Mertens, H.W., and Higgins, E.A. 1987. Some effects of alcohol and simulated altitude on complex performance scores and breathalyzer readings. Aviation, Space, and Environmental Medicine. 58:328-332.
- Durazzini, G., Zazo, F., and Bertoni, G. 1975. The importance of the dosage of thiocyanates in urine and blood of flying personnel for the prevention of diseases of visual function. In G. Perdriel (Ed), Medical requirements and examination procedures in relation to the tasks of today's aircrew, p.A7.1-5. London: NATO Advisory Group for Aerospace Research and Development.
- Fine, B.J. and Kobrick, J.L. 1987. Cigarette smoking, field-dependence and contrast sensitivity. Aviation, Space, and Environmental Medicine. 58:777-782.
- Glick, D.D., Wiley, R.W., Moser, C.E., Park, C.K. 1974. Dark adaptation changes associated with use of the AN/PVS-5 Night Vision Goggle. USAARL-LR-75-2-7-2. US Army Aeromedical Research Laboratory, Fort Rucker, Alabama.
- Luria, S.M. and McKay, C.L. 1979a. Effects of low levels of carbon monoxide on vision of smokers and nonsmokers. Archives of Environmental Health. 34:38-44.
- Luria, S.M. and McKay, C.L. 1979b. Visual processes of smokers and nonsmokers at different ages. Archives of Environmental Health. 34:449-454.
- McFarland, R.A., Roughton, F.J.W., Halperin, M.H. and Niven, J.I. 1944. The effects of carbon monoxide and altitude on visual thresholds. Journal of Aviation Medicine. 15:381-394.
- McFarland, R.A. 1953. Tobacco and efficiency. In Human factors in air transportation, occupational health and safety, p.299-307. McGraw-Hill Book Company, New York.
- McFarland, R.A. 1970. The effects of exposure to small quantities of carbon monoxide on vision. Annals of the New York Academy of Science. 174:301-312.

Pojer, R., Whitfield, J.B., Poulos, V., Eckhard, I.F., Richmond, R., and Hensley, W.J. 1984. Carboxyhemoglobin, cotinine, and thiocyanate assay compared for distinguishing smokers from nonsmokers. Clinical Chemistry. 30:1377-1380.

Sheard, C. 1946. The effects of smoking on the dark adaptation of rods and cones. Federation Proceedings. 5:94.

Troemel, R.G., Davis, R.T., and Hendley, C.D. 1951. Dark adaptation as a function of caffeine and nicotine administration. Proceedings of the South Dakota Academy of Sciences. 30:979-985.

US Surgeon General Report. 1979. Smoking and Health. US Government Printing Office. Washington, D.C.

Initial distribution

**Commander
US Army Natick Research and Development Center
ATTN: Documents Librarian
Natick, MA 01760**

**Commander
US Army Research Institute of Environmental Medicine
Natick, MA 01760**

**Naval Submarine Medical Research Laboratory
Medical Library, Naval Sub Base
Box 900
Groton, CT 05340**

**US Army Avionics Research and Development Activity
ATTN: SAVAA-P-TP
Fort Monmouth, NJ 07703-5401**

**Commander/Director
US Army Combat Surveillance and Target Acquisition Laboratory
ATTN: DELCS-D
Fort Monmouth, NJ 07703-5304**

**US Army Research and Development Support Activity
Fort Monmouth, NJ 07703**

**Commander
10th Medical Laboratory
ATTN: Audiologist
APO NEW YORK 09180**

**Chief, Benet Weapons Laboratory
LCWSL, USA ARRADCOM
ATTN: DRDAR-LCB-TL
Watervliet Arsenal, NY 12189**

**Commander
Naval Air Development Center
Biophysics Lab
ATTN: G. Kydd
Code 60B1
Warminster, PA 18974**

Commander
Man-Machine Integration System
Code 602
Naval Air Development Center
Warminster, PA 18974

Naval Air Development Center
Technical Information Division
Technical Support Detachment
Warminster, PA 18974

Commander
Naval Air Development Center
ATTN: Code 6021 (Mr. Brindle)
Warminster, PA 18974

Dr. E. Hendler
Human Factors Applications, Inc.
295 West Street Road
Warminster, PA 18974

Commanding Officer
Naval Medical Research and Development Command
National Naval Medical Center
Bethesda, MD 20014

Under Secretary of Defense for Research and Engineering
ATTN: Military Assistant for Medical and Life Sciences
Washington, DC 20301

Director
Army Audiology and Speech Center
Walter Reed Army Medical Center
Washington, DC 20307-5001

COL Franklin H. Top, Jr., MD
Walter Reed Army Institute of Research
Washington, DC 20307-5100

Commander
US Army Institute of Dental Research
Walter Reed Army Medical Center
Washington, DC 20307-5300

HQ DA (DASG-PSP-0)
Washington, DC 20310

Naval Air Systems Command
Technical Air Library 950D
Rm 278, Jefferson Plaza II
Department of the Navy
Washington, DC 20361

Naval Research Laboratory Library
Code 1433
Washington, DC 20375

Naval Research Laboratory Library
Shock and Vibration Information Center
Code 5804
Washington, DC 20375

Harry Diamond Laboratories
ATTN: Technical Information Branch
2800 Powder Mill Road
Adelphi, MD 20783-1197

Director
US Army Human Engineering Laboratory
ATTN: Technical Library
Aberdeen Proving Ground, MD 21005-5001

US Army Materiel Systems Analysis Agency
ATTN: Reports Processing
Aberdeen Proving Ground, MD 21005-5017

Commander
US Army Test and Evaluation Command
ATTN: AMSTE-AD-H
Aberdeen Proving Ground, MD 21005-5055

US Army Ordnance Center and School Library
Bldg 3071
Aberdeen Proving Ground, MD 21005-5201

Director (2)
US Army Ballistic Research Laboratory
ATTN: DRXBR-OD-ST Tech Reports
Aberdeen Proving Ground, MD 21005-5066

US Army Environmental Hygiene Agency Laboratory
Bldg E2100
Aberdeen Proving Ground, MD 21010

Commander
US Army Medical Research Institute of Chemical Defense
ATTN: SGRD-UV-AO
Aberdeen Proving Ground, MD 21010-5425

Technical Library
Chemical Research and Development Center
Aberdeen Proving Ground, MD 21010-5423

Commander (5)
US Army Medical Research and Development Command
ATTN: SGRD-RMS (Mrs. Madigan)
Fort Detrick, Frederick, MD 21701-5012

Commander
US Army Medical Research Institute of Infectious Diseases
Fort Detrick, Frederick, MD 21701

Commander
US Army Medical Bioengineering Research and Development Laboratory
ATTN: SGRD-UBZ-I
Fort Detrick, Frederick, MD 21701

Director, Biological Sciences Division
Office of Naval Research
600 North Quincy Street
Arlington, VA 22217

Defense Technical Information Center
Cameron Station
Alexandria, VA 22314

Commander
US Army Materiel Command
ATTN: AMCDE-3 (CPT Broadwater)
5001 Eisenhower Avenue
Alexandria, VA 22333

US Army Foreign Science and Technology Center
ATTN: MTZ
220 7th Street, NE
Charlottesville, VA 22901-5396

Commandant
US Army Aviation Logistics School
ATTN: ATSQ-TDN
Fort Eustis, VA 23604

Director, Applied Technology Laboratory
USARTL-AVSCOM
ATTN: Library, Bldg 401
Fort Eustis, VA 23604

US Army Training and Doctrine Command
ATTN: ATCD-ZX
Fort Monroe, VA 23651

US Army Training and Doctrine Command (2)
ATTN: Surgeon
Fort Monroe, VA 23651-5000

Structures Laboratory Library
USARTL-AVSCOM
NASA Langley Research Center
Mail Stop 266
Hampton, VA 23665

Aviation Medicine Clinic
TMC #22, SAAF
Fort Bragg, NC 28305

Naval Aerospace Medical Institute Library
Bldg 1953, Code 102
Pensacola, FL 32508

US Air Force Armament Development and Test Center
Eglin Air Force Base, FL 32542

Command Surgeon
US Central Command
MacDill Air Force Base, FL 33608

US Army Missile Command
Redstone Scientific Information Center
ATTN: Documents Section
Redstone Arsenal, AL 35898-5241

Air University Library
(AUL/LSE)
Maxwell AFB, AL 36112

US Army Research and Technology Laboratories (AVSCOM)
Propulsion Laboratory MS 302-2
NASA Lewis Research Center
Cleveland, OH 44135

AFAMRL/HEX
Wright-Patterson AFB, OH 45433

US Air Force Institute of Technology
(AFIT/IDE)
Bldg 640, Area B
Wright-Patterson AFB, OH 45433

University of Michigan
NASA Center of Excellence in Man-Systems Research
ATTN: R.G. Snyder, Director
Ann Arbor, MI 48109

Henry L. Taylor
Director, Institute of Aviation
University of Illinois--Willard Airport
Savoy, IL 61874

John A. Dellinger, MS, ATP
University of Illinois--Willard Airport
Savoy, IL 61874

Commander
US Army Aviation Systems Command
ATTN: DRSV-WS
4300 Goodfellow Blvd
St Louis, MO 63120-1798

Project Officer
Aviation Life Support Equipment
ATTN: AMCP-ALSE
4300 Goodfellow Blvd
St Louis, MO 63120-1798

Commander
US Army Aviation Systems Command
ATTN: SGRD-UAX-AL (MAJ Lacy)
Bldg 105, 4300 Goodfellow Blvd
St Louis, MO 63120

Commander
US Army Aviation Systems Command
ATTN: DRSV-ED
4300 Goodfellow Blvd
St Louis, MO 63120

US Army Aviation Systems Command
Library and Information Center Branch
ATTN: DRSV-DIL
4300 Goodfellow Blvd
St Louis, MO 63120

Commanding Officer
Naval Biodynamics Laboratory
P.O. Box 24907
New Orleans, LA 70189

Federal Aviation Administration
Civil Aeromedical Institute
CAMI Library AAC 64D1
P.O. Box 25082
Oklahoma City, OK 73125

US Army Field Artillery School
ATTN: Library
Snow Hall, Room 14
Fort Sill, OK 73503

Commander
US Army Academy of Health Sciences
ATTN: Library
Fort Sam Houston, TX 78234

Commander
US Army Health Services Command
ATTN: HSOP-SO
Fort Sam Houston, TX 78234-6000

Commander
US Army Institute of Surgical Research
ATTN: SGRD-USM (Jan Duke)
Fort Sam Houston, TX 78234-6200

Director of Professional Services
AFMSC/GSP
Brooks Air Force Base, TX 78235

US Air Force School of Aerospace Medicine
Strughold Aeromedical Library
Documents Section, USAFSAM/TSK-4
Brooks Air Force Base, TX 78235

US Army Dugway Proving Ground
Technical Library
Bldg 5330
Dugway, UT 84022

Dr. Diane Damos
Department of Human Factors
ISSM, USC
Los Angeles, CA 90089-0021

US Army Yuma Proving Ground
Technical Library
Yuma, AZ 85364

US Army White Sands Missile Range
Technical Library Division
White Sands Missile Range, NM 88002

US Air Force Flight Test Center
Technical Library, Stop 238
Edwards Air Force Base, CA 93523

US Army Aviation Engineering Flight Activity
ATTN: SAVTE-M (Tech Lib) Stop 217
Edwards Air Force Base, CA 93523-5000

Commander
Code 3431
Naval Weapons Center
China Lake, CA 93555

US Army Combat Developments Experimental Center
Technical Information Center
Bldg 2925
Fort Ord, CA 93941-5000

Aeromechanics Laboratory
US Army Research and Technical Laboratories
Ares Research Center, M/S 215-1
Moffett Field, CA 94035

Commander
Letterman Army Institute of Research
ATTN: Medical Research Library
Presidio of San Francisco, CA 94129

Sixth US Army
ATTN: SMA
Presidio of San Francisco, CA 94129

Director
Naval Biosciences Laboratory
Naval Supply Center, Bldg 844
Oakland, CA 94625

Commander
US Army Aeromedical Center
Fort Rucker, AL 36362

Commander
US Army Aviation Center and Fort Rucker
ATTN: ATZQ-CDR
Fort Rucker, AL 36362

Directorate of Combat Developments
Bldg 507
Fort Rucker, AL 36362

Directorate of Training Development
Bldg 502
Fort Rucker, AL 36362

Chief
Army Research Institute Field Unit
Fort Rucker, AL 36362

Chief
Human Engineering Laboratory Field Unit
Fort Rucker, AL 36362

Commander
US Army Safety Center
Fort Rucker, AL 36362

Commander
US Army Aviation Center and Fort Rucker
ATTN: ATZQ-T-ATL
Fort Rucker, AL 36362

US Army Aircraft Development Test Activity
ATTN: STEBG-MF-QA
Cairns AAF, Fort Rucker, AL 36362

President
US Army Aviation Board
Cairns AAF, Fort Rucker, AL 36362

Distribution to foreign addressees

**Chief
Defence and Civil Institute of Environmental Medicine
P.O. Box 2000
ATTN: Director MLSD
Downsview, Ontario Canada M3M 3B9**

**USDAO-AMLO, US Embassy
Box 36
FPO New York 09510**

**Staff Officer, Aerospace Medicine
RAF Staff, British Embassy
3100 Massachusetts Avenue, NW
Washington, DC 20008**

**Canadian Society of Aviation Medicine
c/o Academy of Medicine, Toronto
ATTN: Ms. Carmen King
288 Bloor Street West
Toronto, Canada M5S 1V8**

**Canadian Airline Pilot's Association
MAJ (Retired) J. Soutendam
1300 Steeles Avenue East
Brampton, Ontario, Canada L6T 1A2**

**Canadian Forces Medical Liaison Officer
Canadian Defence Liaison Staff
2450 Massachusetts Avenue, NW
Washington, DC 20008**

**Commanding Officer
404 Squadron CFB Greenwood
Greenwood, Nova Scotia, Canada B0P 1N0**

**Officer Commanding
School of Operational and Aerospace Medicine
DCIEM P.O. Box 2000
1133 Sheppard Avenue West
Downsview, Ontario, Canada M3M 3B9**

**National Defence Headquarters
101 Colonel By Drive
ATTN: DPM
Ottawa, Ontario, Canada K1A 0K2**

Commanding Officer
Headquarters, RAAF Base
Point Cook Victoria, Australia 3029

Canadian Army Liaison Office
Bldg 602
Fort Rucker, AL 36362

Netherlands Army Liaison Office
Bldg 602
Fort Rucker, AL 36362

German Army Liaison Office
Bldg 602
Fort Rucker, AL 36362

British Army Liaison Office
Bldg 602
Fort Rucker, AL 36362

French Army Liaison Office
Bldg 602
Fort Rucker, AL 36362