

AWARD NUMBER: W81XWH-15-1-0072

TITLE: Development of Tethered Hsp90 Inhibitors Carrying Radioiodinated Probes to Specifically Discriminate and Kill Malignant Breast Tumor Cells

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7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Duke University 2200 W. Main St., Suite 710 Box 104010 Durham 27705		5e. TASK NUMBER
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14. ABSTRACT Hsp90 is a recognized oncogenic signal node whose up regulation/activation is associated with multiple forms of human cancer including 70% of all breast cancers. There are 17 Hsp9 inhibitors in clinical development for cancer, including SNX5422, developed by members of our team. In breast cancer, clinical studies have linked activation of Hsp90 and surface expression ectopic of Hsp90 (eHsp90) with poor outcomes in malignant breast tumors. We therefore hypothesized that selective targeting of eHsp90 on breast tumors presents an opportunity to not only discriminate indolent tumors from metastatic disease, but also offers a molecularly targeted radiotherapy approach for body wide tumor ablation with low normal tissue toxicity. To test this hypothesis, our laboratories developed a series of PET (Positron Emission Tomography) enabled tethered Hsp90inhibitors (Ti90s) that demonstrate exquisite selectivity in vivo for metastatic breast tumor cells. We also discovered that eHsp90 is rapidly internalized and can carry radionucleotide labeled Ti90 specifically into the breast cancer cell. In this proposal we will optimize conditions that enable two PET enabled Ti90s (HS-113 andHS-227) carrying radioiodine (124I and 131I) or 211Astatine to be effectively delivered to malignant breast tumors using mouse models of triple negative, ER+ and HER2+ breast cancer. These studies will provide essential proof of concept data to move one or more PET enabled Ti90 into first in human clinical trials. Ultimately, we envisage a process in which a patient, after standard of care breast exam, is first evaluated for malignancy vs. indolent disease by PET imaging using 124I-labeled Ti90. Then, in patients with malignancies detected in high contrast to normal tissues, targeted radiotherapy would be performed at patient-optimized doses of inhibitor labeled with the β -emitter 131I or the α -emitter 211At.		

15. SUBJECT TERMS					
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1. Introduction.

Our laboratories recently developed a series of optical and iodinated tethered Hsp90 inhibitors that have exquisite selectivity in vivo for metastatic breast tumors expressing ectopic (cell surface) Hsp90 (3). We also discovered that ectopically expressed Hsp90 is rapidly internalized and can carry these tethered inhibitors specifically into breast cancer cells. This work, in tandem with published clinical results, suggests that selective targeting of Hsp90 up regulated in malignancy may present an opportunity to offer a molecularly targeted radiotherapy with a very favorable therapeutic index. We proposed to develop a series of tethered Hsp90 inhibitors capable of selectively delivering radio labelled (^{124}I and ^{131}I) or ^{211}At to malignant tumor cells. We envisaged a process in which a patient facing either early stage or metastatic breast cancer is evaluated for Hsp90 overexpression on their tumor cells by positron emission tomography (PET) imaging of ^{124}I -labeled tethered inhibitors. Then, in patients with malignancies demonstrated to exhibit a sufficient level of ectopic Hsp90 expression, targeted radiotherapy would be preformed at patient-optimized doses of inhibitor labeled with the β -emitter ^{131}I or the α -emitter ^{211}At . This is an attractive strategy for breast cancer because the same molecule that detects ectopic Hsp90 can then be used for tumor selective radio ablation using our new compounds. In the same way Antibody Drug Conjugates can deliver chemotherapy selectively to HER2+ cells, our compounds would selectively deliver therapeutic doses of radiation to only malignant cells by manipulating the active uptake of ectopically expressed Hsp90 into cells.

2. Key Words.

Hsp90, Hsp70, PET, Pharmacokinetics, MTD, EGFR, ER, ^{124}I , ^{131}I , ^{211}At , Astatine, nrIR, SNX5422, Hsp90 inhibitor, Breast Cancer, Ti90, eHsp90.

Overall Project Summary.

We hypothesized that ^{124}I -labeled tethered Hsp90 inhibitors (Ti90s) and PET imaging will allow non-invasive discrimination of malignant breast tumors from more benign disease. Furthermore, we hypothesized that this will also permit determination on an individualized basis, of the optimal dose of versions labeled with the therapeutic radionuclides ^{131}I or ^{211}At to achieve tumor ablation with minimal normal tissue toxicity. Our objectives were to test these hypotheses in proof of concept studies using various mouse models of human breast cancer. We anticipated that if successful, these studies will provide the basis for moving our approach into clinical trial.

3. ACCOMPLISHMENTS.

What were the major goals of the project?

Milestone Aim 1 (**50% complete**). Identification of a minimum of 3 cell selective lead molecules based on both the SNX2112 and PU-H71 scaffolds. We also anticipate that these leads will be derived from a larger chemical series that can also be further developed if unanticipated problems arise in Aims 2-4.

Milestone 1 Aim 2 (**75% complete**). Identification by PET and necropsy biodistribution tethered inhibitors with appropriate PK for use as PET imaging agents with ^{124}I and targeted radiotherapy with ^{131}I and ^{211}At .

Milestone 2 Aim 2 (**50% complete**). Demonstration that tethered [^{124}I]PU-H71 shows greater tumor selectivity compared with untethered [^{124}I]PU-H71 in multiple mouse models of metastatic breast cancer.

Milestone Aim 3. (**50% complete**) Demonstration of successful targeted radiotherapy with one or more tethered Hsp90 inhibitor carrying either ^{131}I or ^{211}At in in multiple mouse models of metastatic breast cancer.

What was accomplished under these goals?

A major goal of the funded research was to develop Ti90s that were amenable for dual imaging modalities of PET and optical, such that they could be readily interfaced with current mammography practice. To this end, with this funding, we extended our existing tethered Hsp90 synthetic platform to develop Ti90 molecules that could first be used for histology with the nonradioactive iodine- containing analogs, and then for noninvasive whole body PET imaging with the same molecules labeled with ^{124}I , and ultimately, for selective tumor

ablation with analogs labeled with ^{131}I . We dubbed this strategy the “see and cure” approach. As discussed in the original narrative, in mammography for example, a patient who requires a biopsy receives a micro dose of a Ti90 carrying non-radioactive iodine. If fluorescence microscopy of the biopsy slides reveals uptake of the probe, and this concurs with the pathology diagnosis, the patient would be offered the PET enabled, ^{124}I -labeled version for whole body imaging to identify evidence of disease progression. This could then be followed by targeted radiotherapy with the ^{131}I version. Note, that the physicochemical properties of the molecule are the same at all stages, just the isotope changes. This is greatly enabling for the clinical development pathway because the initial safety studies (GMP/GLP) prior to PET imaging can be carried out with nonradioactive material. With these goals in mind, the Haystead lab developed a synthetic pathway to derive a series of fluorescent probe structures also carrying iodine that could subsequently stannylated. During the development of these probes, we discovered that placement of iodine within the ligand binding region inactivate the molecule completely and placement of the iodine distally and close to the fluorophore was essential to retain activity, both *in vitro* and *in vivo*. The original narrative outlined the synthetic pathway for two Ti90's we made, HS-113 and HS-227. The complete synthetic pathways of all our Ti90s containing an iodine (can be substituted with ^{124}I for PET) are described in recently issued US and worldwide patents (US9,738,643 B2, US 2015/0191471 A1, WO2014/025395 A1). In collaboration with the Zalutsky laboratory, we then developed a standard operating procedure for radioiodination of the tin precursors of HS-113 and HS-227 with ^{124}I . These methodologies will be disclosed in future submissions describing the utility of ^{124}I -labeled Ti90s for imaging tumors *in vivo*. Recent published work with non-radioactive fluorescent versions has enabled us to define the molecular mechanism by which PET imaging-amenable Ti90s recognize eHsp90 when it is expressed on the surface of multiple malignant breast tumor lines (*Barrott et al. Cell Chem. Biol.* 2013;20(9):1-11; *Crowe et al. ACS Chem. Biol.* 2017, 12, 1047–1055). Additionally, more recent independent studies by the Ramanujam lab demonstrated direct uptake of a non- radioactive analog of HS-113, synthesized in the Haystead lab (HS-27), into fresh needle biopsies obtained from women undergoing lumpectomy and diagnosed with either HER2+, ER+ or triple negative breast cancer (*Crouch et al. Sci Rep.* 2017;7(1):17487). These seminal studies provide evidence that aggressive forms of breast cancer selectively express eHsp90 in humans. Collectively, these studies demonstrate that Ti90s carrying a fluorescent probe have great utility for preclinical mechanistic studies as they can be detected by confocal microscopy, histological analysis and isolation of eHsp90+ve tumor cells by flow cytometry. In a recently completed DOD funded program, the Haystead lab successfully took a newly synthesized nrIR tethered Ti90 (HS-196) through the IND process (GMP manufacture, GLP animal toxicity and formulation). In 2017, the Lyerly group (Duke University) opened a 120 patient first in human solid tumor phase 1 study to evaluate the safety and utility of HS-196 to detect malignancies *in vivo* (<https://clinicaltrials.gov/ct2/show/NCT03333031>). Probes like HS-196 will have no utility for whole body imaging or treatment. Light scattering of the emitted fluorescence, limits detection to tissue depths of ~0.5-1.0cm. However, lessons learned with HS-196 will be invaluable in guiding the future preclinical and clinical development of PET imaging-amenable versions of Ti90s. In collaboration with the Zalutsky lab and the Small Animal Imaging (SAI) core and mouse phase 1 unit (MPIU) at UNC-Lineberger Comprehensive Cancer Center at the University of North Carolina, we tested our ability to image [^{124}I]HS-113 by PET in the MMTV mouse model of HER2+ breast cancer. MMTV mice produce spontaneous primary and secondary tumors that are thought to mimic the cellular heterogeneity of tumors observed in human disease, which are often a mixture of premalignant, malignant, normal stromal tissue and benign cells. The emergence of primary tumors is largely confined to mammary tissue, but secondary metastatic masses can be generated 60-70% of the time. Figure 1 highlights two separate studies in which we injected [^{124}I]HS-113 into MMTV mice bearing tumors (No DOD funds were used for these animal studies). In both examples, we observe discrete probe accumulation in secondary tumor masses consistent with prior published work suggesting a correlation between expression of eHsp90 and a metastatic phenotype. Figure 1C and 1D starkly highlight the specificity of [^{124}I] HS-113 relative to [^{18}F]FDG, the later showing broad uptake in metabolically active tissues such as kidney, heart and brain. These promising results also discriminate our PET-amenable Ti90s from that of a radioiodinated form of the Hsp90 inhibitor PU-H71 reported by Larson and colleagues (49). In their clinical PET/CT study, following injection of radioiodinated PU-H71, the drug showed non-specific whole-body distribution, consistent with a diffusible drug and ubiquitous expression of Hsp90 in most normal cell populations. However, after 48 h, as the radiotracer

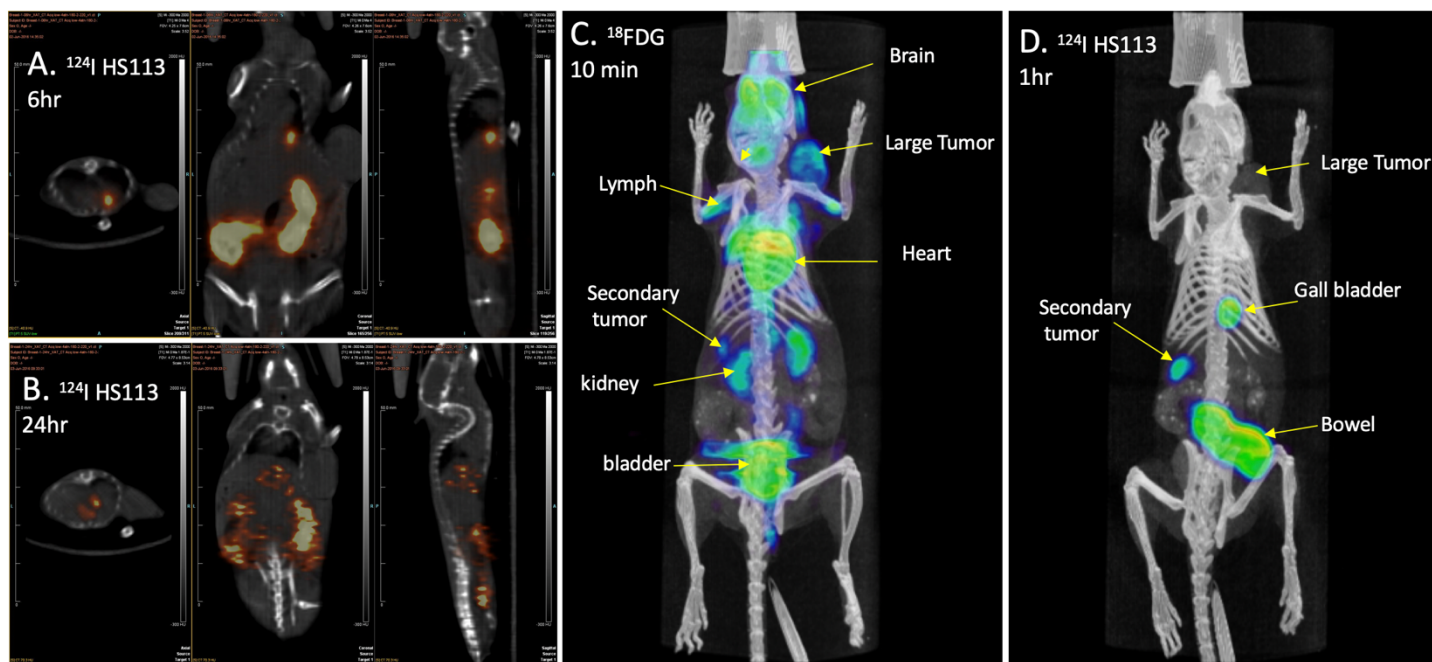


Figure 1. PET/CT images showing that ^{124}I HS-113 detects primary and secondary tumor masses in mouse models of HER2+ breast cancer. (A) and (B) shows a time course of a single mouse injected with a trace amount ^{124}I HS-113 and imaged by PET/CT at two time points. A mass is detected at 6 hours in the lung. The majority of radioactivity is cleared via the bowl by 24 hrs. In (C) an MMTV mouse bearing a primary tumor (neck region) was injected with ^{18}F FDG (5mM) and imaged by PET/CT. Uptake in all major organs are detected along with the primary tumor mass and a secondary mass in the mid region. In (D) the same animal was injected 24 hrs later with 1nmol of ^{124}I HS-113 and imaged by PET CT at 1 hr. The images shows discrete uptake in the secondary metastatic mass with more discrete uptake in the primary mass. No other organs exhibited uptake in contrast to ^{18}F FDG, illustrating the selectivity of ^{124}I HS-113. As with the example in (A) and (B), ^{124}I HS-113 was eliminated primarily through the bowel via the biliary system.

cleared systemically, tumor-specific contrast emerged. Although radio labelled PU-H71 might have value as diagnostic tool, its utility as a platform for developing a therapeutic ablative agent is limited because of its broad non-targeted distribution.

One of the most striking features of our studies with [^{124}I]HS-113 is its rapid and exclusive clearance through the biliary system (Fig. 2). Other Ti90s that we have examined such as HS-196 are primarily

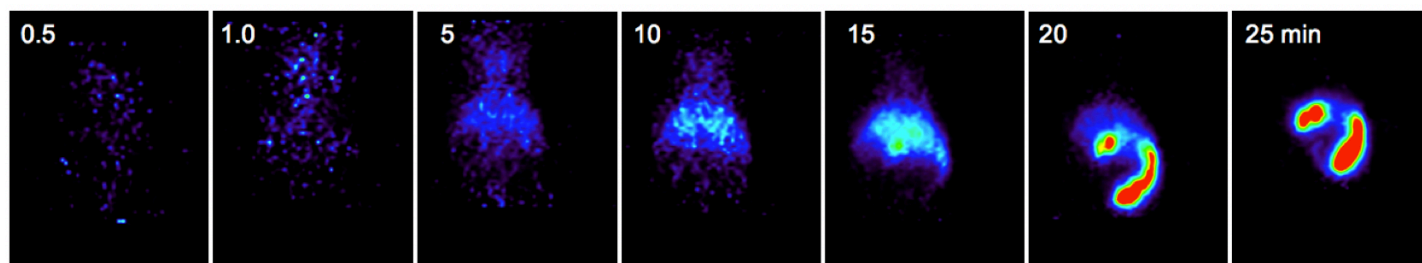


Figure 2. Time lapse sequence following the real time elimination of ^{124}I HS-113 by PET imaging in a mouse. The animal was injected with a tracer amount of ^{124}I HS-113 and the signal followed continuously over 30 min. Images show the free probe rapidly collecting in the liver and condensing into the biliary ductal network within 10 minutes, followed by concentration into the gall bladder (15 min). By 20 minutes the probe is emptied into the small intestine. Images are snap shots taken from a movie.

eliminated through the kidneys. In the proposed Aims, we will compare the PK properties of [^{124}I]HS-113 which carries a fluorescein moiety, with [^{124}I]HS-227, which carries a Cy5 far red dye. Our goal is to have a PET-amenable Ti90 that can be micro dosed with serum clearance times of 1-2 hrs. We also prefer to have the clearance through the biliary system and into the bowel. Elimination of the radiolabeled versions via the intestine may be desirable since the intestinal contents are likely to reduce irradiation of the gut wall. Rapid clearance via the liver is also desirable since this also limits any potential for secondary tissue damage if the radiolabeled Ti90s persisted for some time in the circulation and were more slowly eliminated.

To date, we have synthesized 68 different Ti90s carrying various cargoes (see patents US9,738,643 B2, US 2015/0191471 A1, WO2014/025395 A). These include a full range of fluorophores of varying excitation and

emission profiles. The full synthesis of these compounds is outlined in our patent filings as well as prior publications. With this funding, we identified at least two PET-amenable versions that we can reproducibly synthesize, stannylate and radioiodinate. If either of these lead compounds fail, we are well placed to draw upon the large pool of backup molecules that may perform better *in vivo* in future work. Our work revealed some unexpected aspects of our PET-amenable Ti90s that could be advantageous in the development of both diagnostic and radiotherapeutic agents. With additional funding we will focus specifically on defining optimal delivery conditions that ensure both efficient tumor loading as well as rapid normal tissue clearance of unbound probe. We believe this is a critical goal in order to move the development of these probes forward to first in human phase 1 studies.

What opportunities for training and professional development has the project provided?

Nothing to Report.

How were the results disseminated to communities of interest?

Two patents were filed that have issued describing the full synthesis of all Ti90s synthesized in this project US9,738,643 B2, US 2015/0191471 A1, WO2014/025395 A.

What do you plan to do during the next reporting period to accomplish the goals?

Nothing to Report.

4. IMPACT.

What was the impact on the development of the principal discipline(s) of the project?

Nothing to Report

What was the impact on other disciplines?

Nothing to Report

What was the impact on technology transfer?

Nothing to Report

What was the impact on society beyond science and technology?

Nothing to Report

5. CHANGES/PROBLEMS:

Nothing to Report.

6. PRODUCTS:

Two patents were filed that have issued describing the full synthesis of all Ti90s synthesized in this project US9,738,643 B2, US 2015/0191471 A1, WO2014/025395 A.

7. PARTICIPANTS & OTHER COLLABORATING ORGANIZATIONS

What individuals have worked on the project?

Name	Timothy Haystead
Project Role	PI
Researcher Identifier	0000-0001-7274-1519
Nearest person month worked:	15
Contribution to Project:	Oversaw study design
Project Role	David Carlson
Researcher Identifier	0000-0001-9910-5379
Nearest person month worked:	20
Contribution to Project:	Synthetic Chemist
Project Role	David Loiselle
Researcher Identifier	Not known
Nearest person month worked:	10
Contribution to Project:	Mass spectrometry

Has there been a change in the active other support of the PD/PI(s) or senior/key personnel since the last reporting period?

Nothing to report.

What other organizations were involved as partners?

Nothing to Report

8. SPECIAL REPORTING REQUIREMENTS.

COLLABORATIVE AWARDS:

My partnering POI was Dr. Michael Zalutsky (Duke University) who will be submitting his own final report.

9. APPENDICES:



US009738643B2

(12) **United States Patent**
Haystead et al.

(10) **Patent No.:** **US 9,738,643 B2**
(45) **Date of Patent:** **Aug. 22, 2017**

- (54) **SUBSTITUTED INDAZOLES FOR TARGETING HSP90**
- (71) Applicant: **Duke University**, Durham, NC (US)
- (72) Inventors: **Timothy Haystead**, Chapel Hill, NC (US); **Philip Floyd Hughes**, Chapel Hill, NC (US)
- (73) Assignee: **Duke University**, Durham, NC (US)
- (*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

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- (21) Appl. No.: **14/419,965**
- (22) PCT Filed: **Mar. 14, 2013**
- (86) PCT No.: **PCT/US2013/031614**
§ 371 (c)(1),
(2) Date: **Feb. 6, 2015**

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- (87) PCT Pub. No.: **WO2014/025395**
PCT Pub. Date: **Feb. 13, 2014**

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- (65) **Prior Publication Data**
US 2015/0191471 A1 Jul. 9, 2015

Related U.S. Application Data

- (60) Provisional application No. 61/680,107, filed on Aug. 6, 2012.

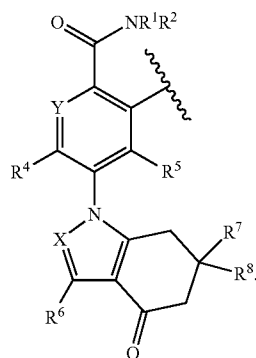
Primary Examiner — Douglas M Willis

(74) *Attorney, Agent, or Firm* — Michael Best & Friedrich LLP

- (51) **Int. Cl.**
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C07D 473/34 (2006.01)
C07D 405/14 (2006.01)
G01N 33/569 (2006.01)
G01N 33/574 (2006.01)
C07D 413/12 (2006.01)
C07D 473/24 (2006.01)
C07D 487/04 (2006.01)
C07D 493/10 (2006.01)

- (57) **ABSTRACT**

Described herein are substituted indazoles that may selectively bind to Hsp90, methods of using the compounds, and kits including the compounds. The compounds may include detection moieties such as fluorophores that may allow for selective detection of Hsp90 in a sample. Suitable compounds include those of formula (II):



- (52) **U.S. Cl.**
CPC **C07D 473/34** (2013.01); **C07D 231/54** (2013.01); **C07D 405/14** (2013.01); **C07D 413/12** (2013.01); **C07D 473/24** (2013.01); **C07D 487/04** (2013.01); **C07D 493/10** (2013.01); **G01N 33/56988** (2013.01); **G01N 33/574** (2013.01); **G01N 33/57496** (2013.01); **G01N 2333/47** (2013.01)

- (58) **Field of Classification Search**
CPC **C07D 231/54**
USPC **548/360.1**
See application file for complete search history.

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10 Claims, 23 Drawing Sheets

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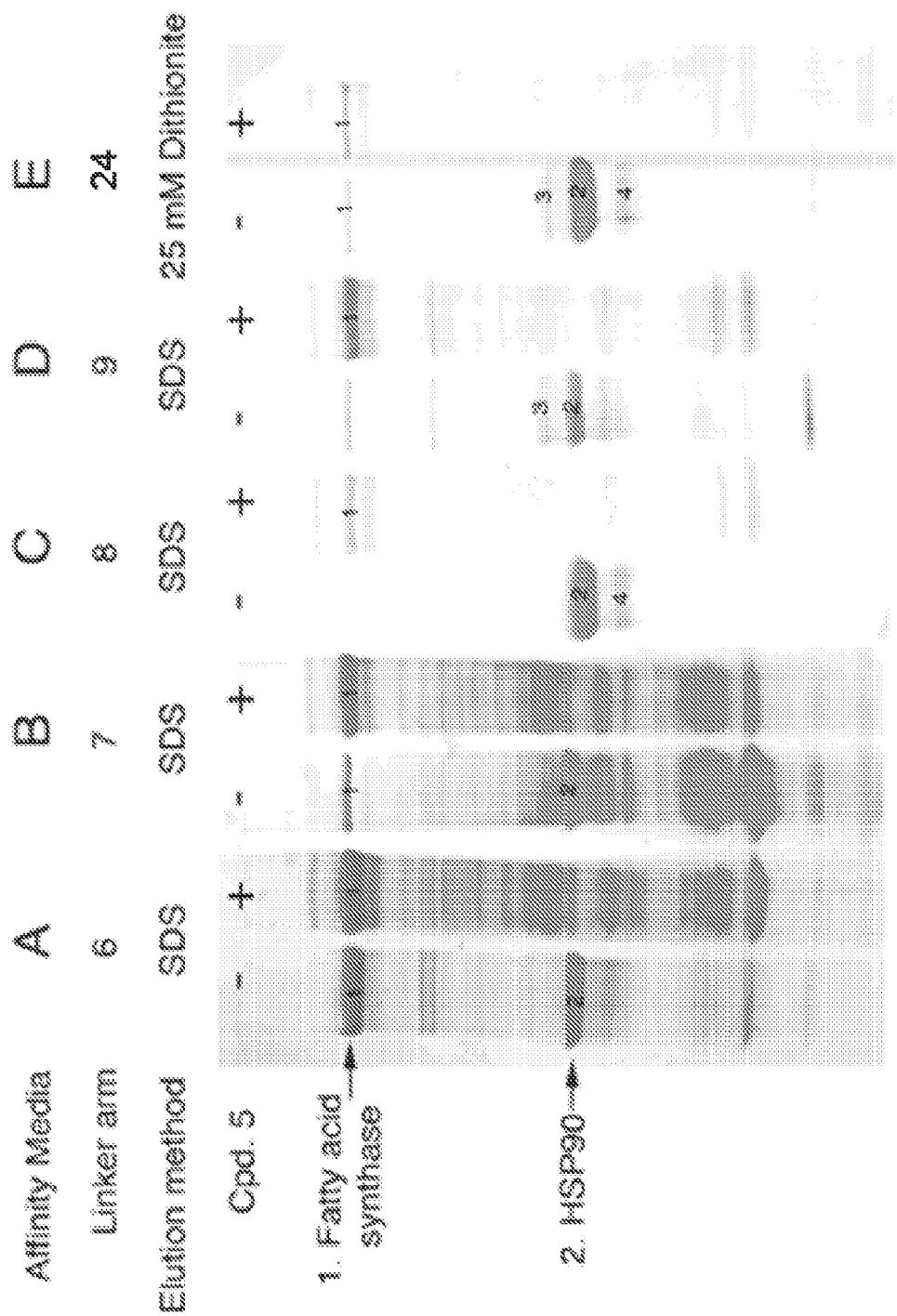


FIGURE 1

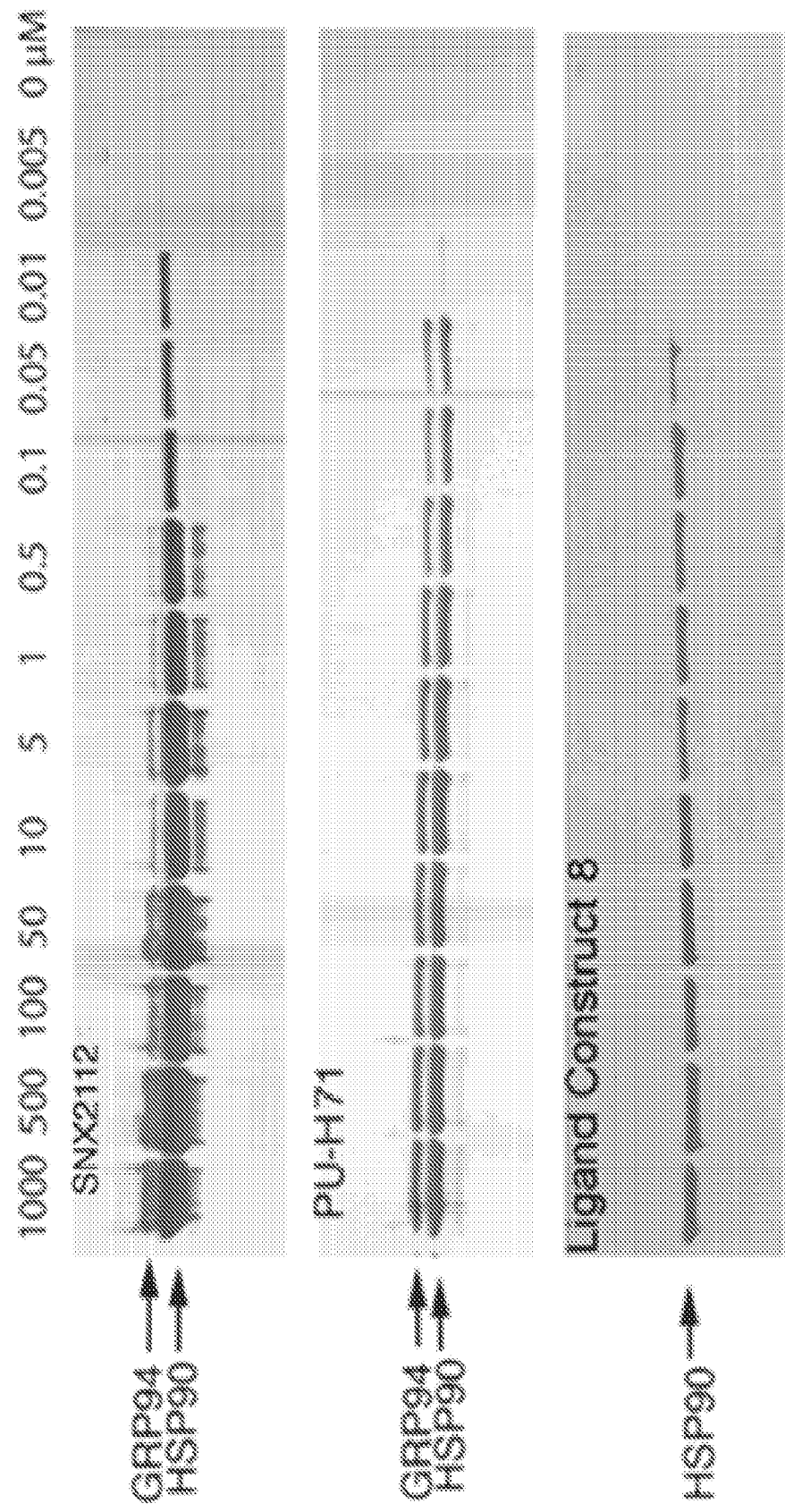


FIGURE 2

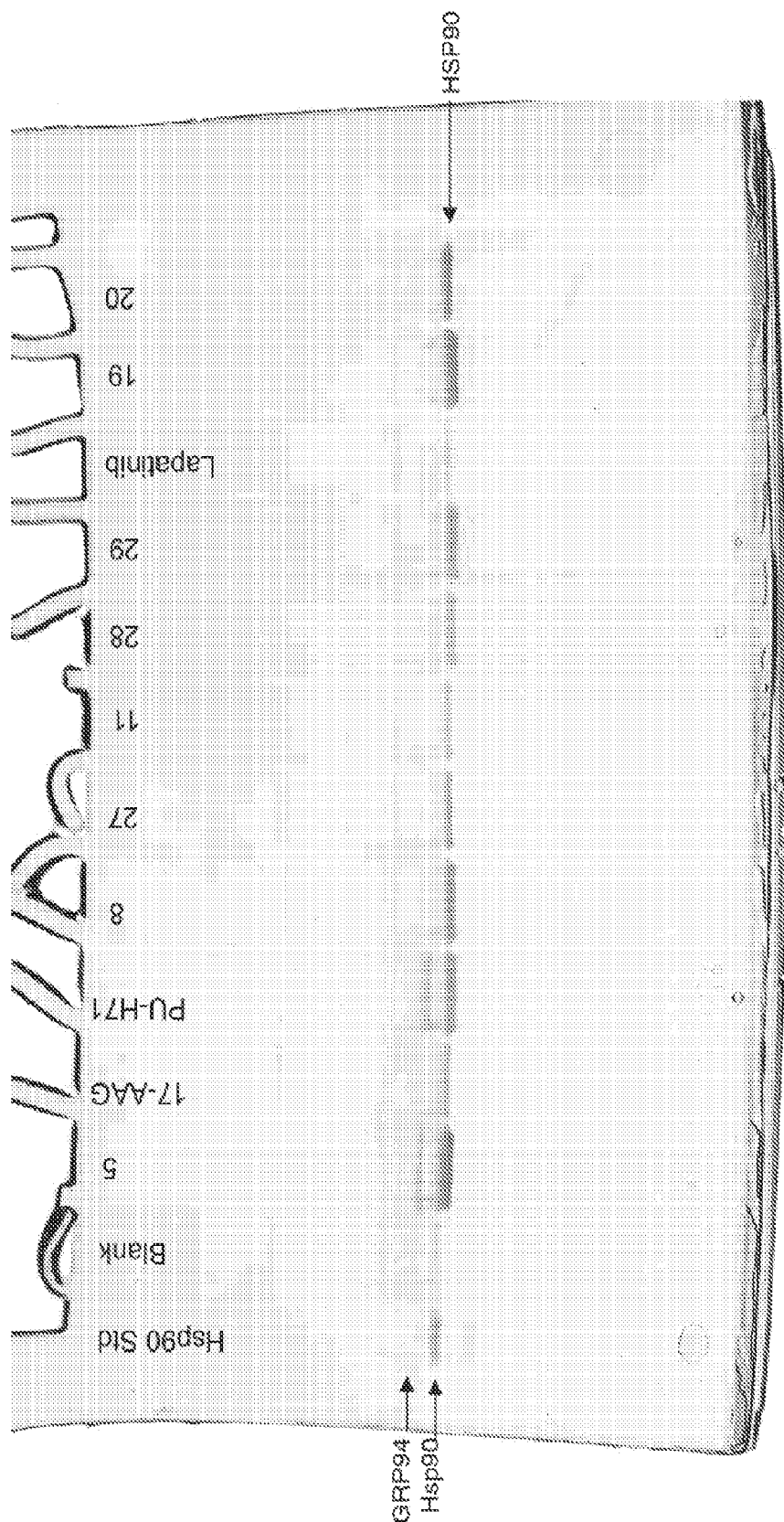


FIGURE 3

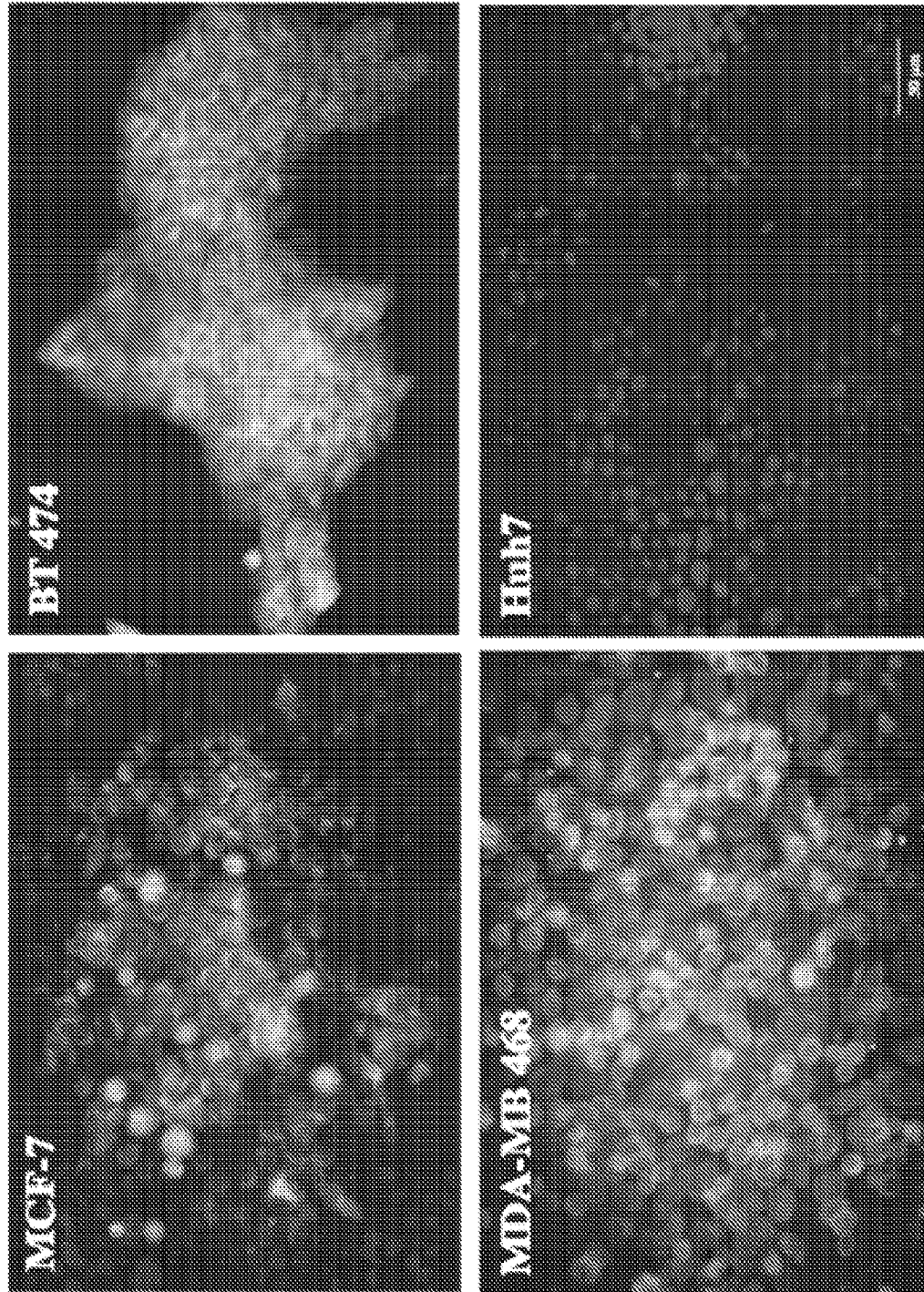


FIGURE 4

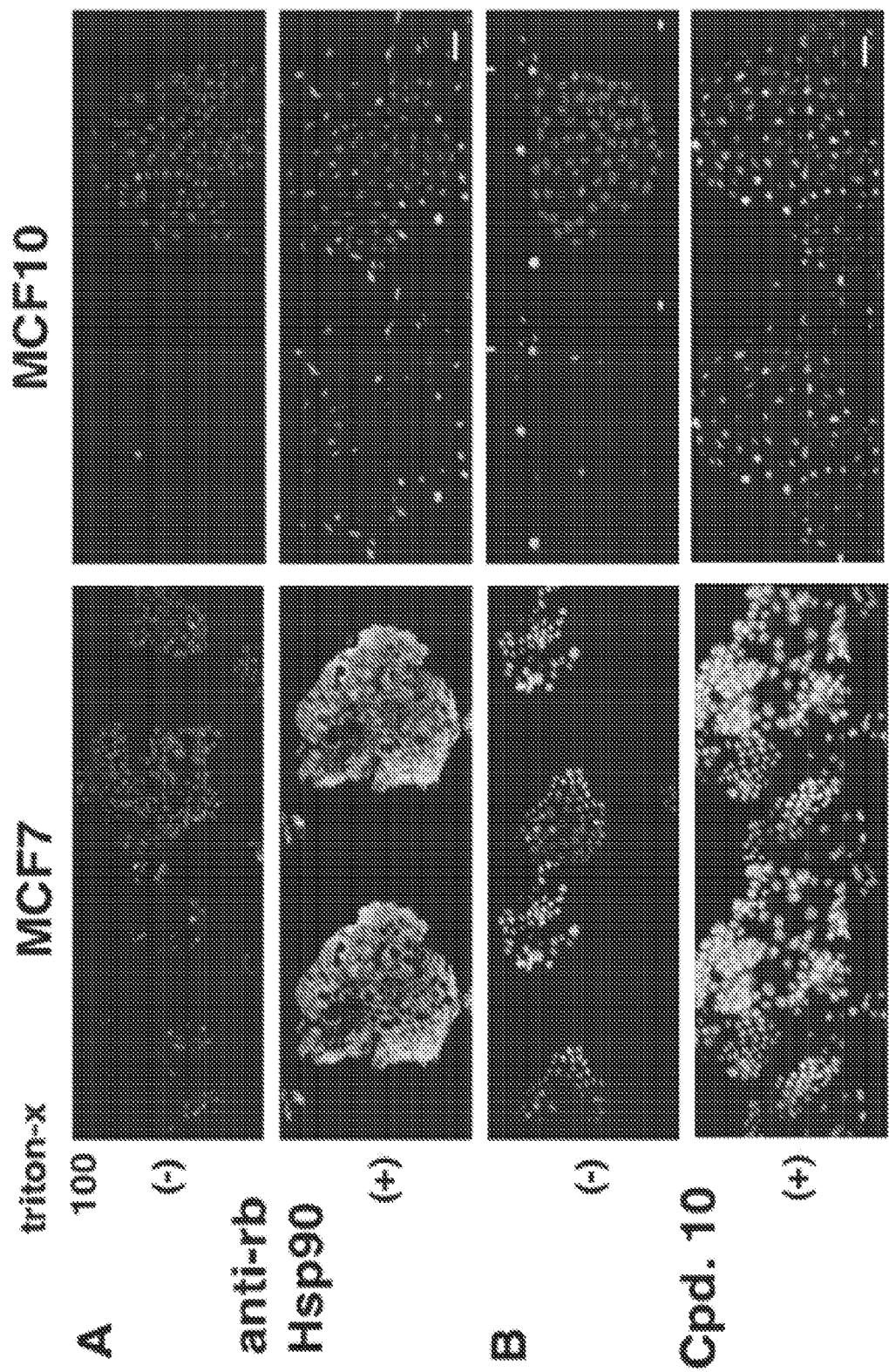


FIGURE 5

Entry of Compound 10 is mediated through
surface Hsp90 in MCF7 cells

anti-Hsp90
2° AF-568

Compound 10

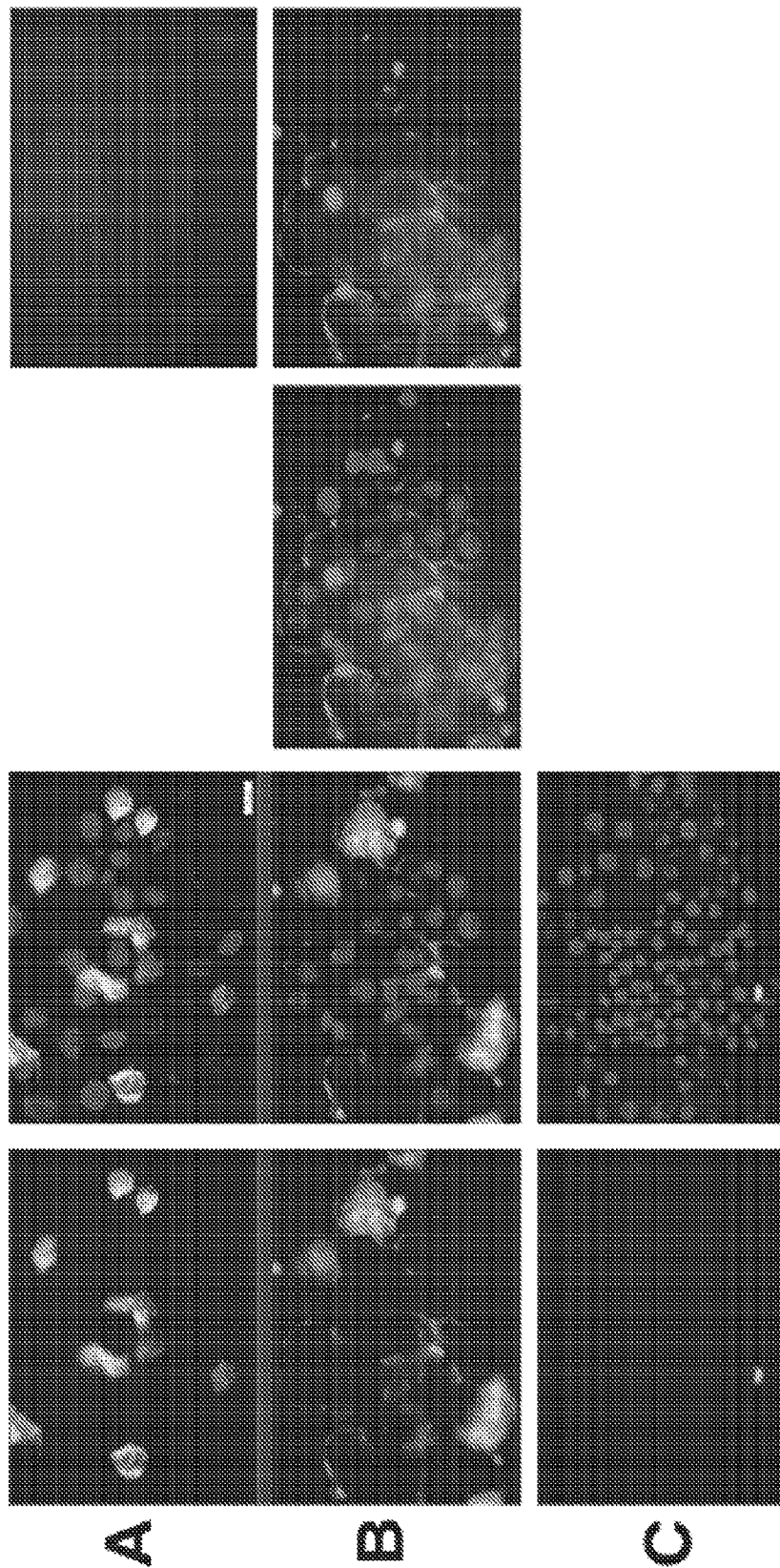
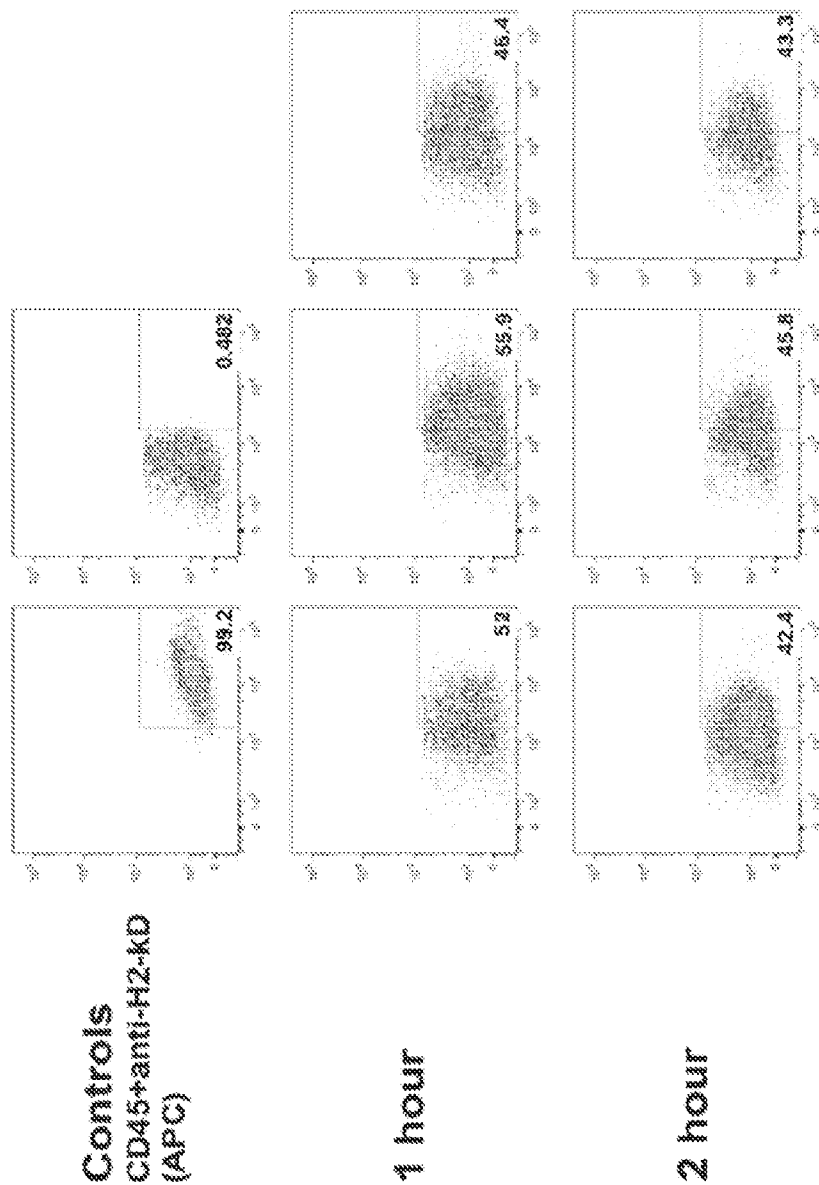


FIGURE 6

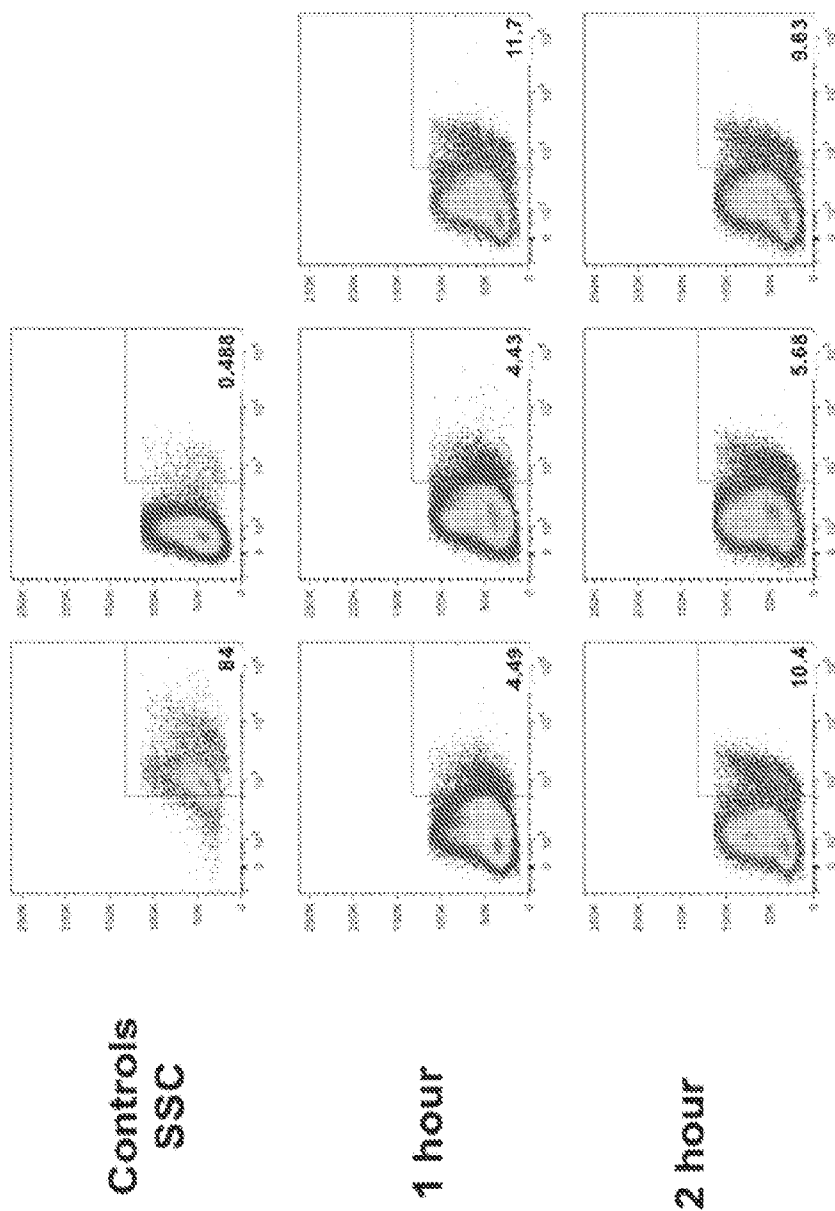
**Compound 10 Labeling of MDA-MB-468 tumor cells
in NOD/SCID mice**



Compound 10

FIGURE 7A

Mouse Splenocytes Compound 10 binding Gated
on Lymphocytes



Compound 10

FIGURE 7B

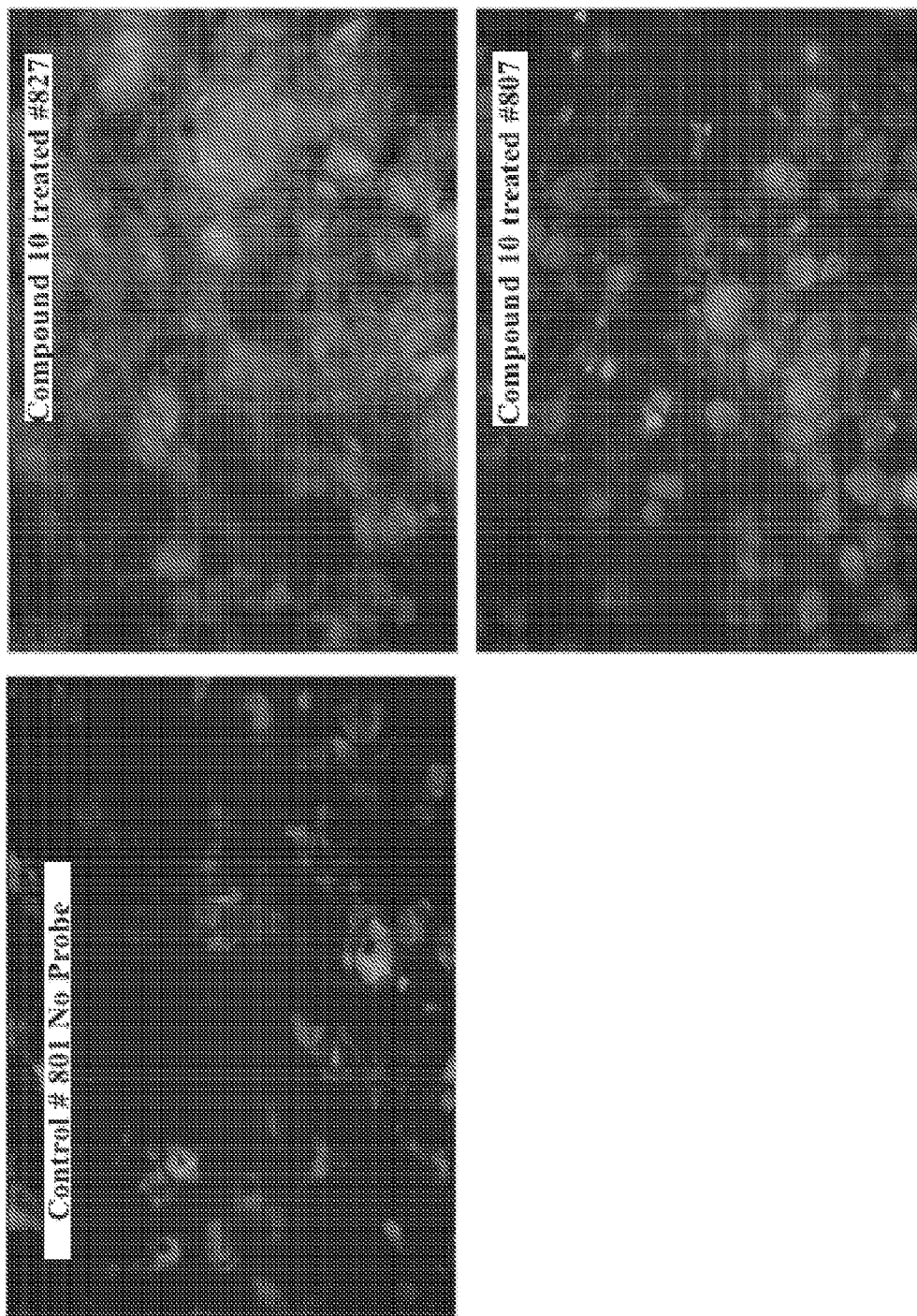


FIGURE 8

Competition of Signal with Free Drug (5)

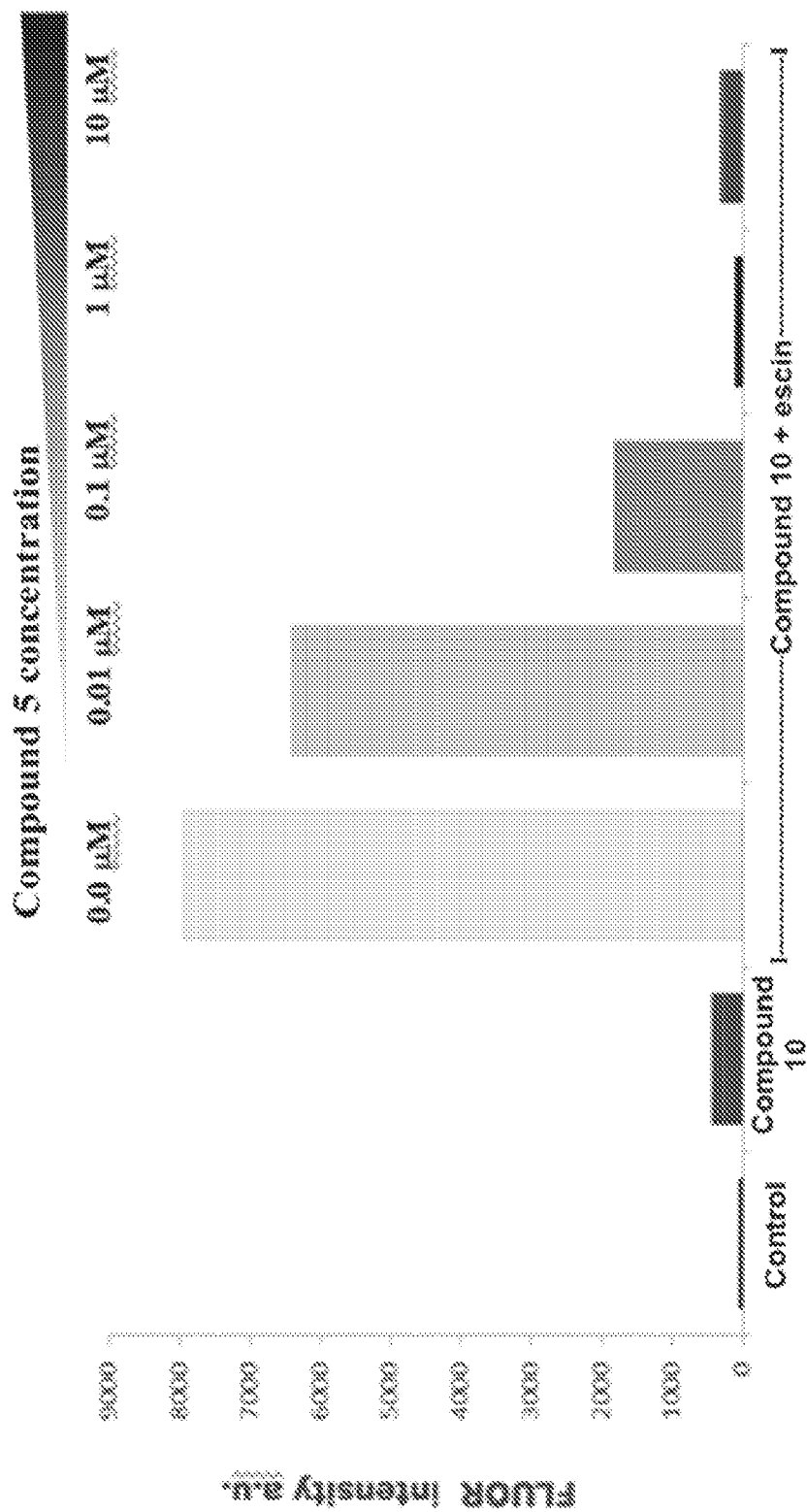


FIGURE 9

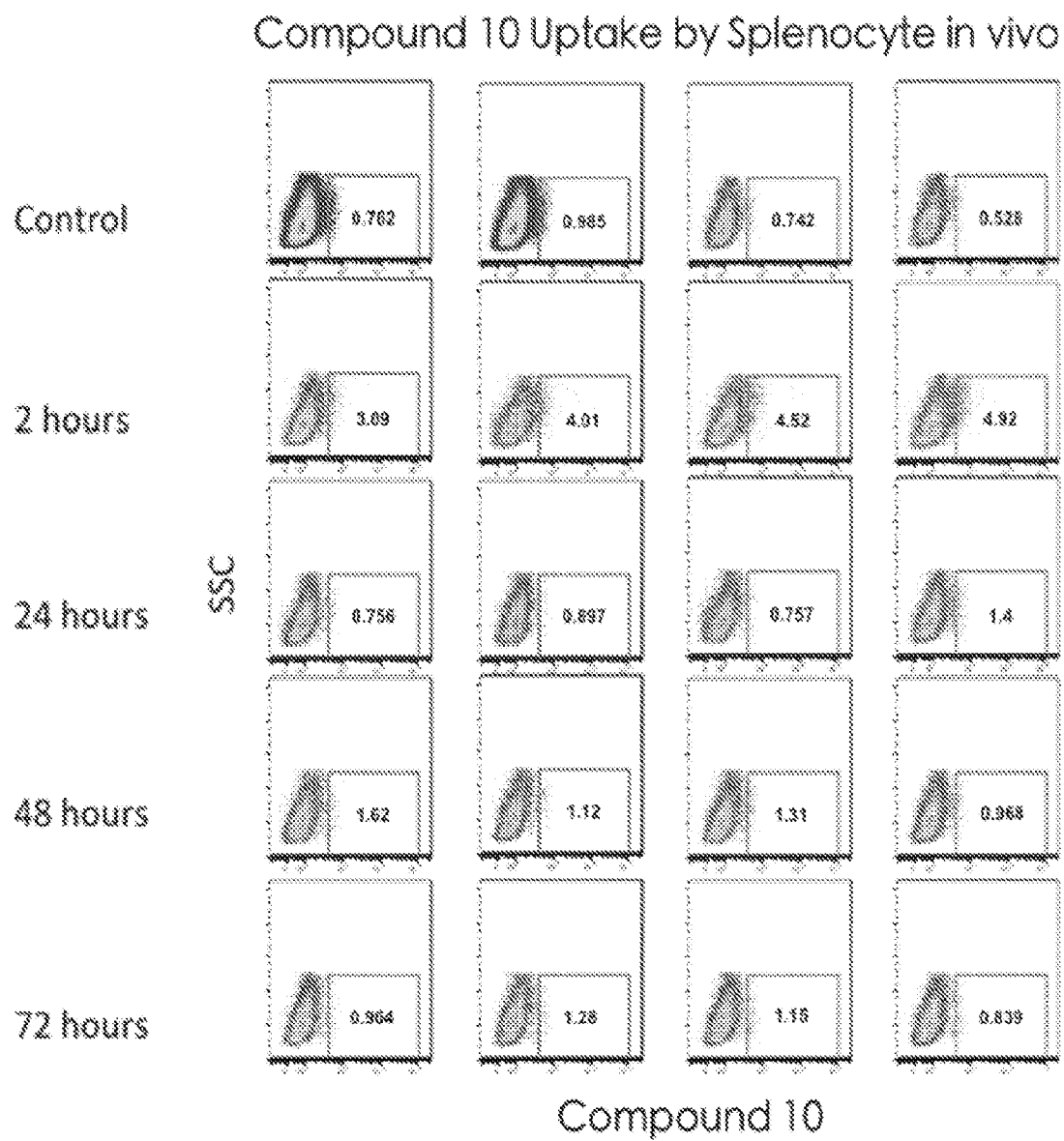


FIGURE 10A

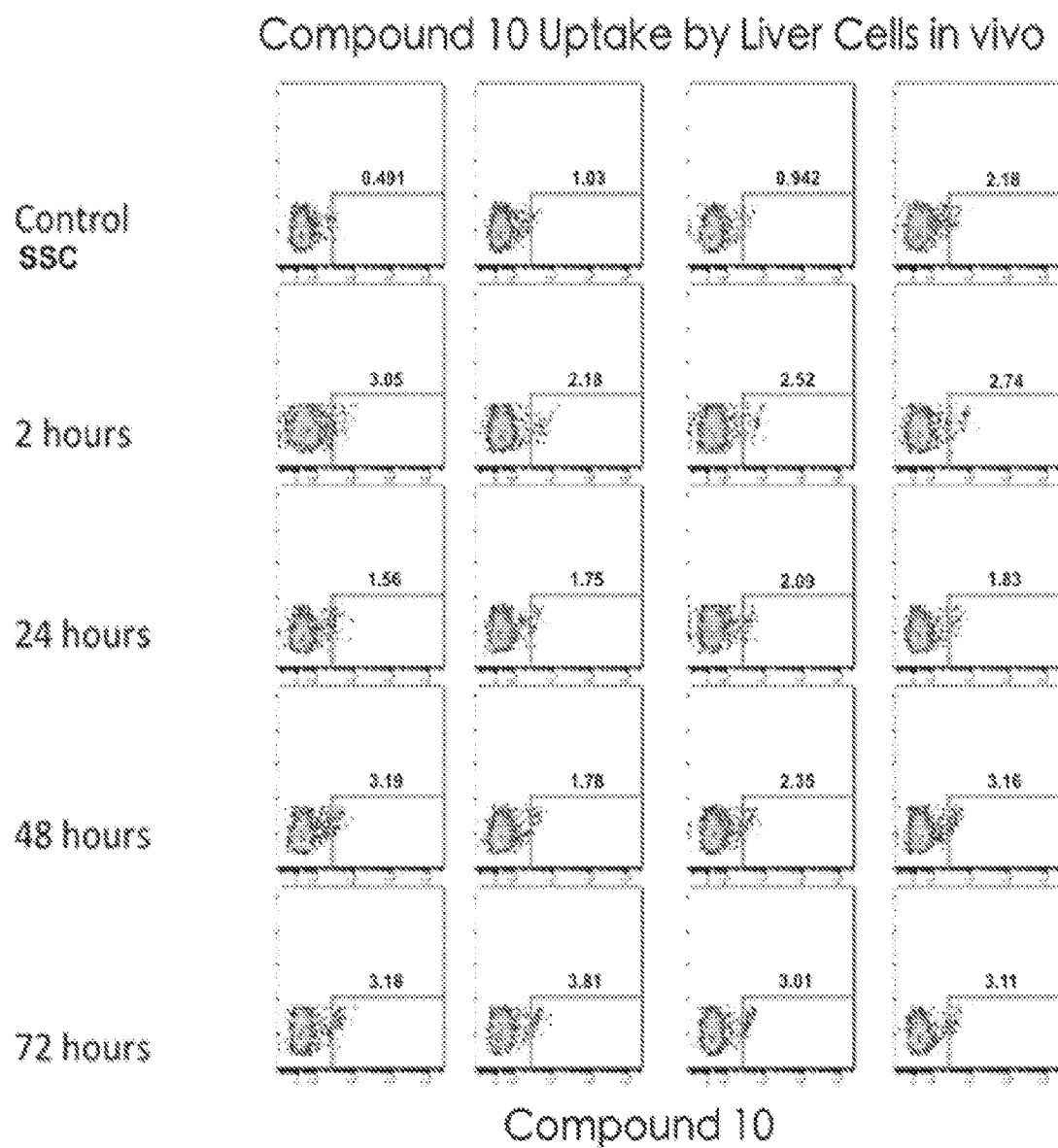


FIGURE 10B

Compound 10 Uptake by Tumor Cells in vivo

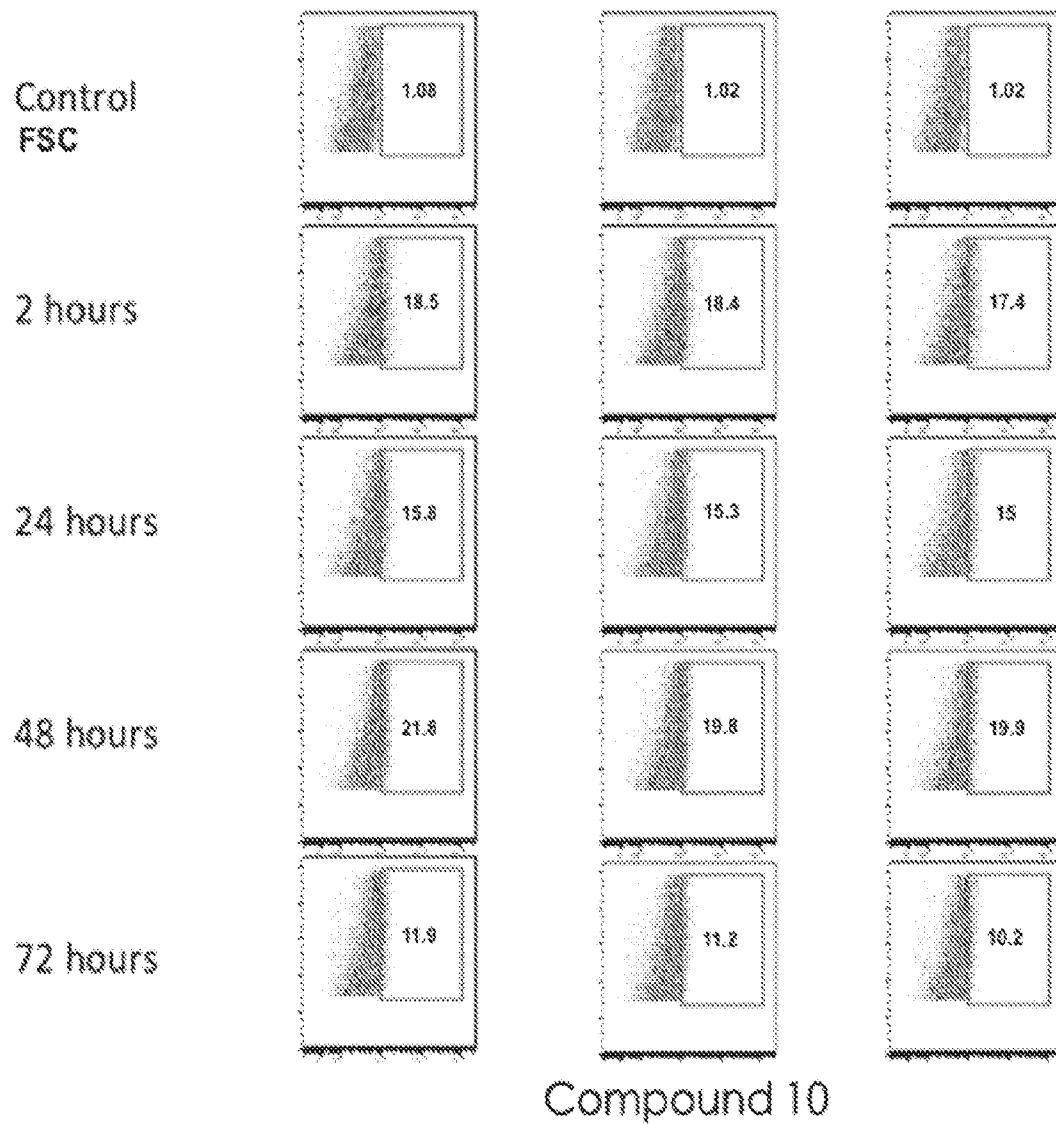


FIGURE 10C

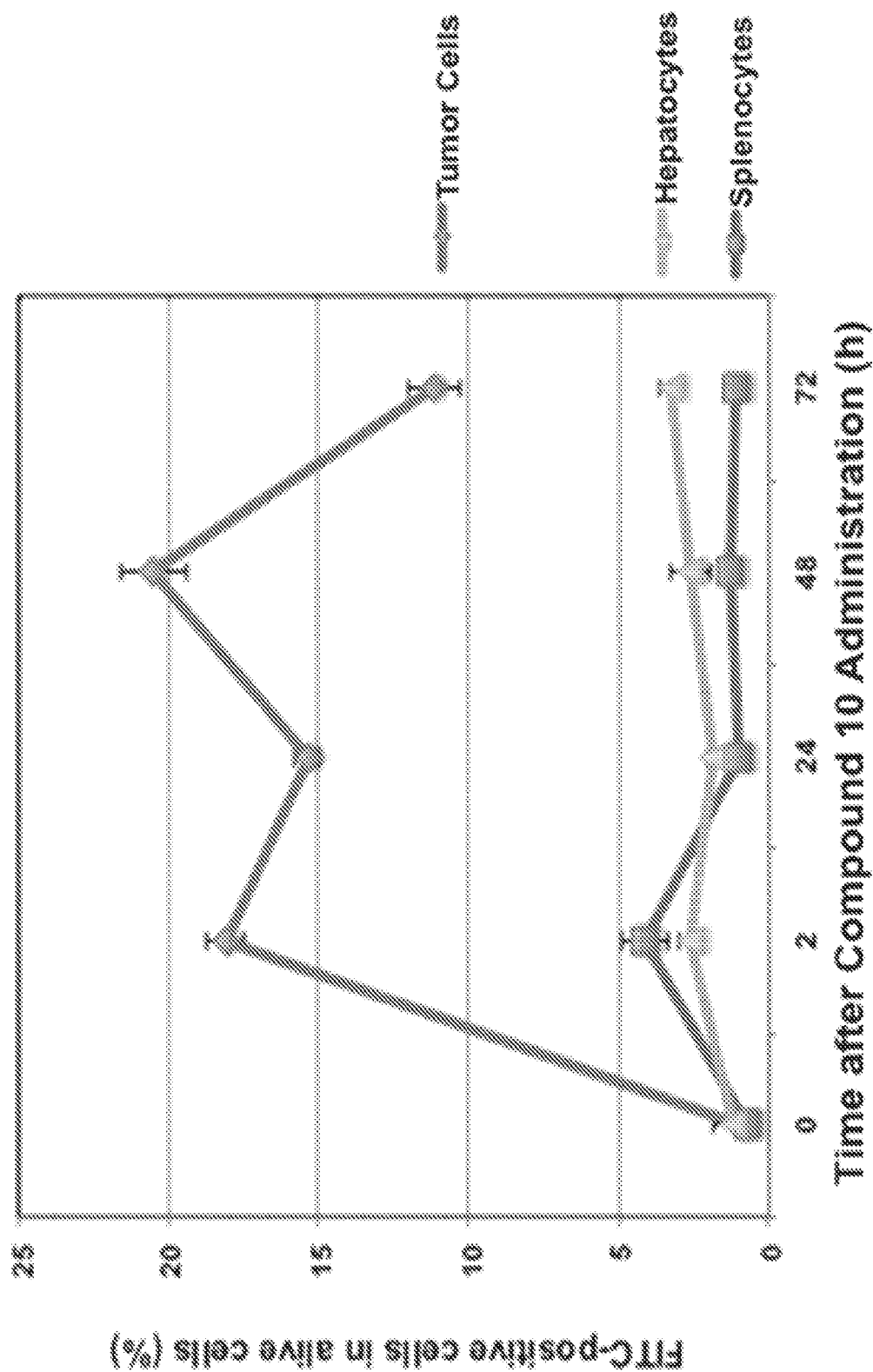


FIGURE 11

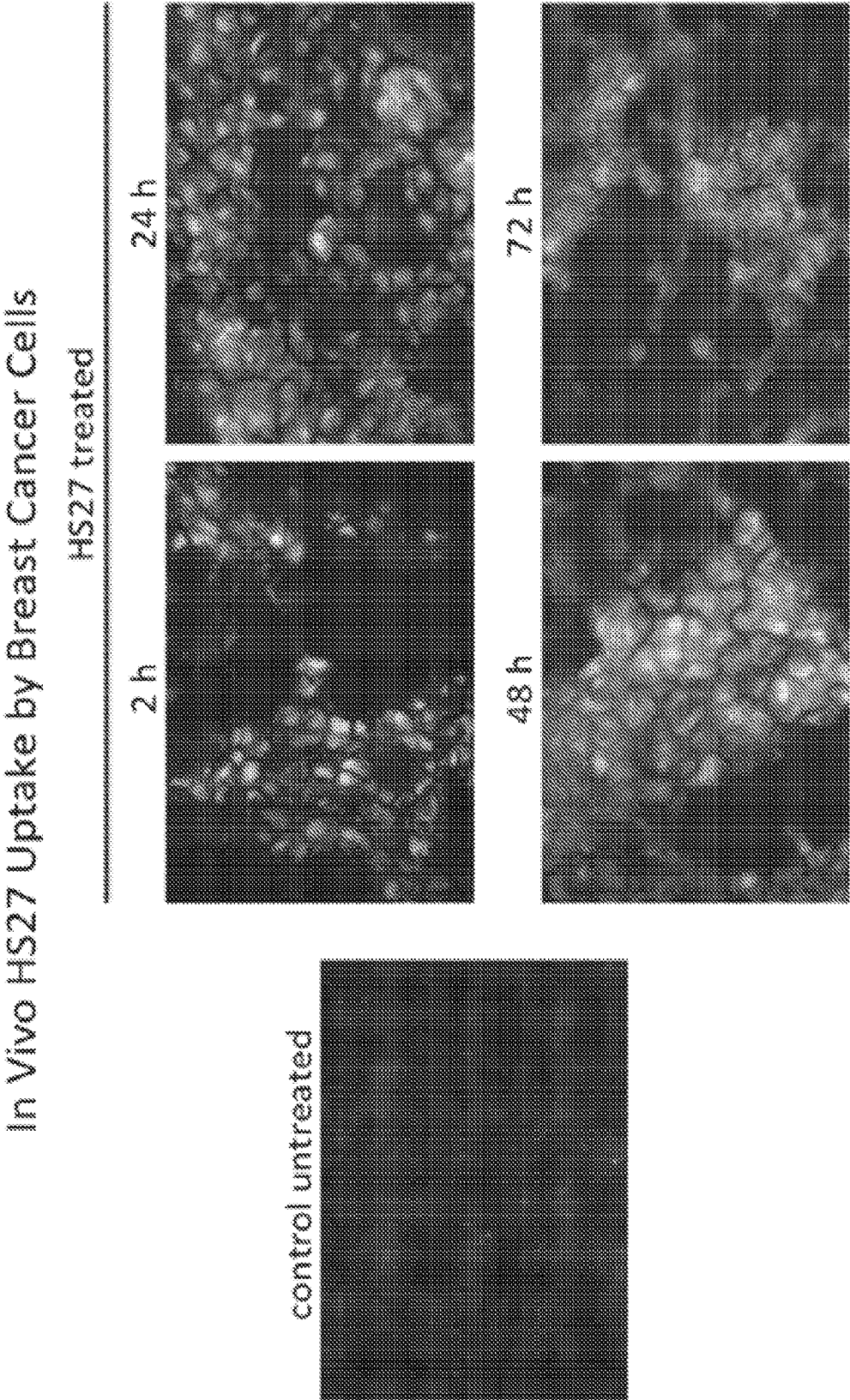


FIGURE 12

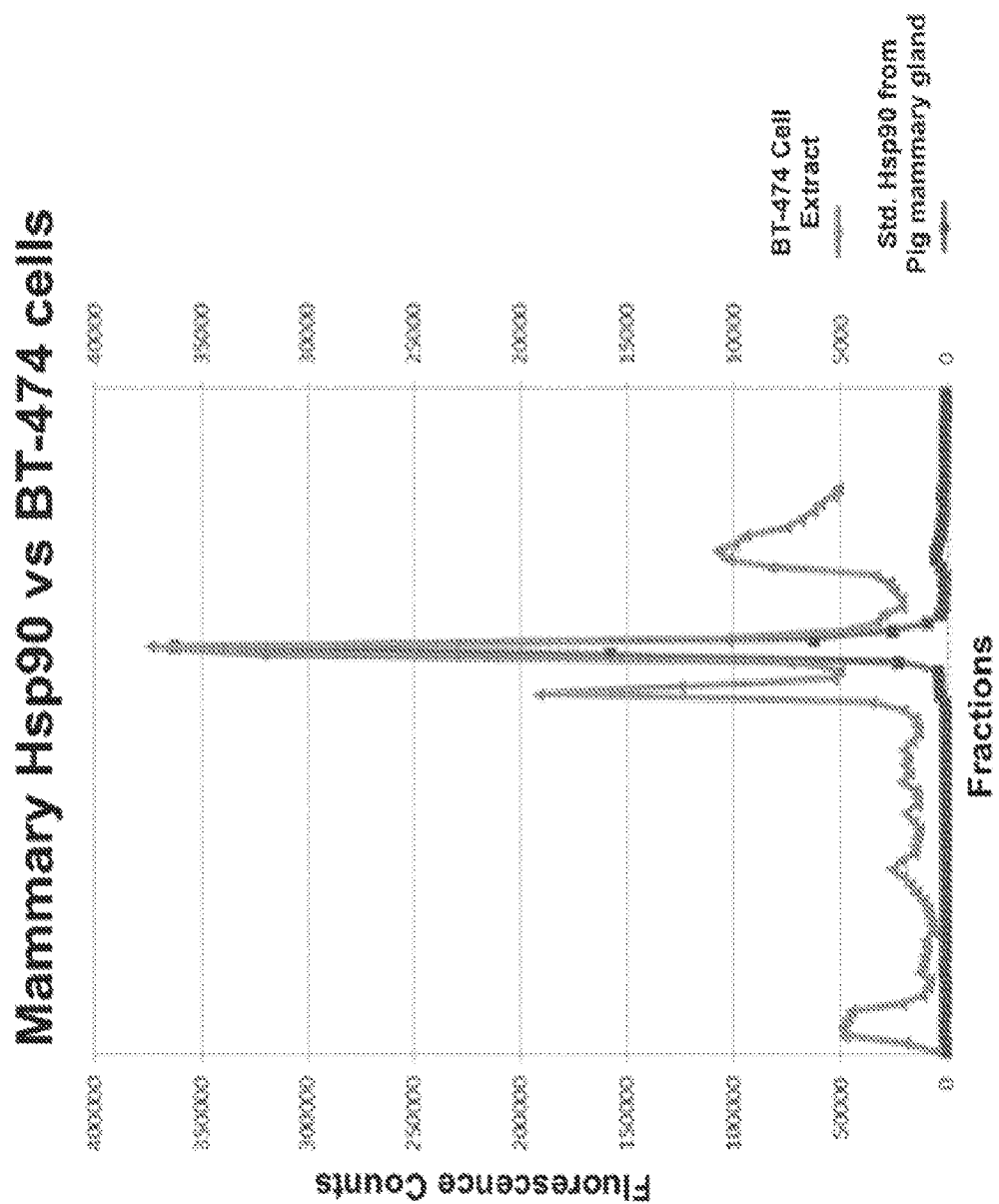


FIGURE 13A

MonoQ (anion exchange)
fractions of MCF-7 cells

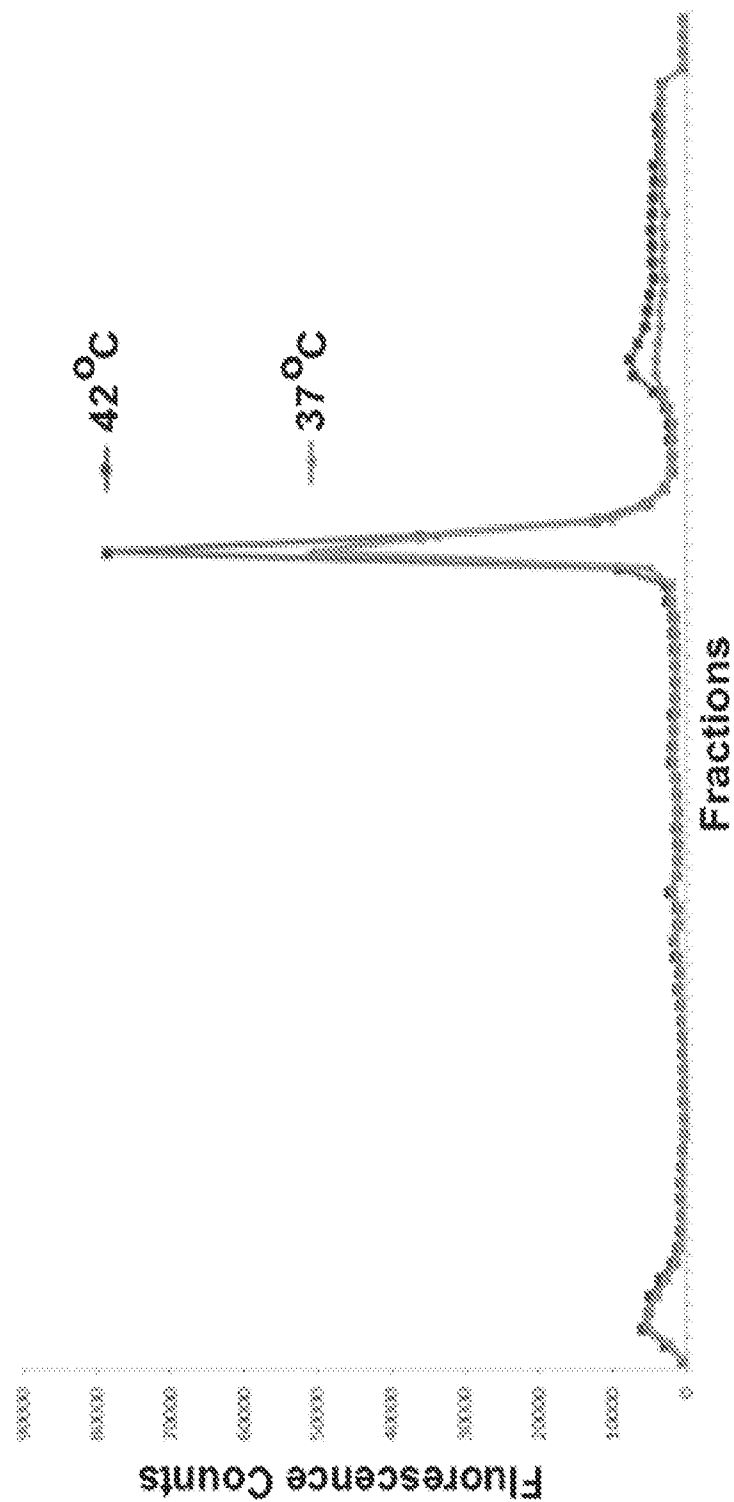


FIGURE 13B

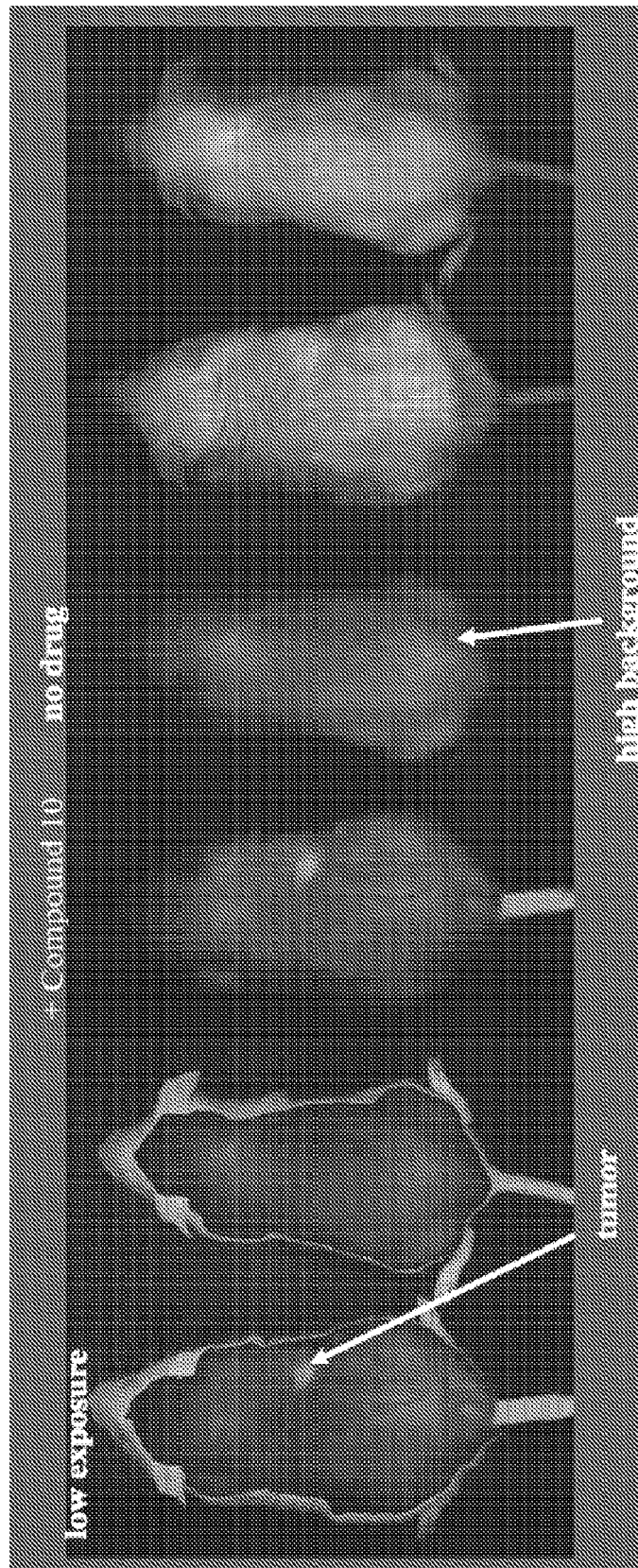


FIGURE 14A

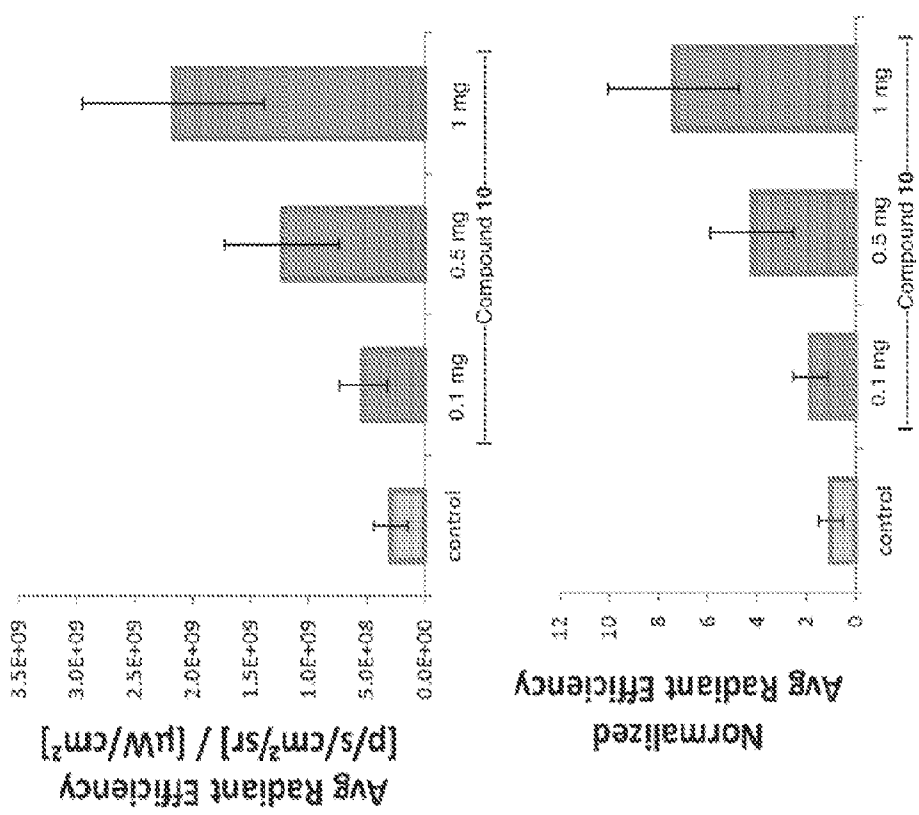
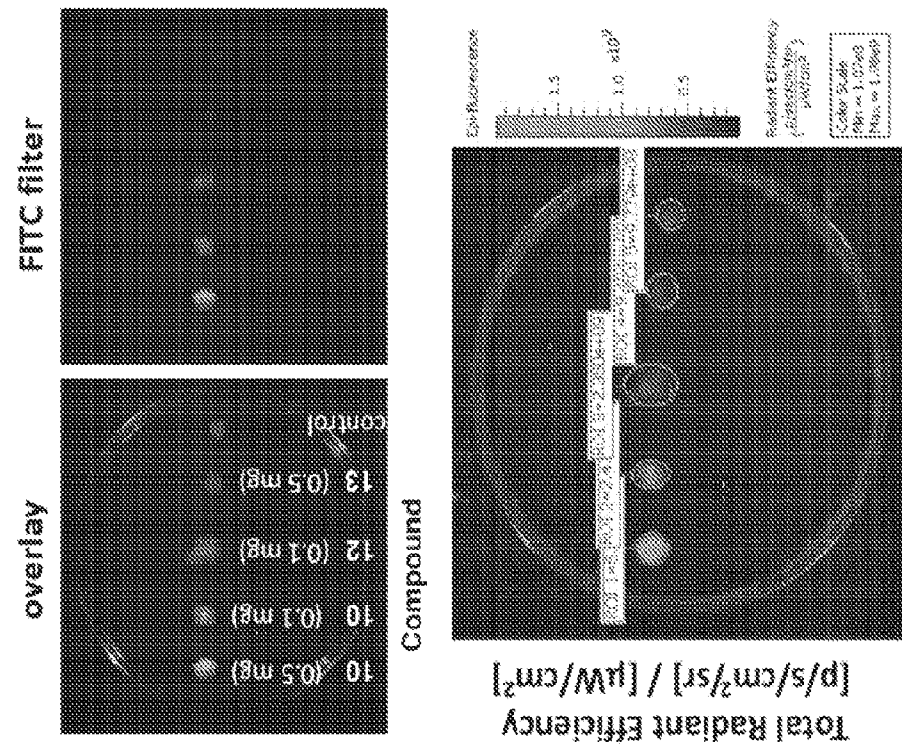


FIGURE 14B



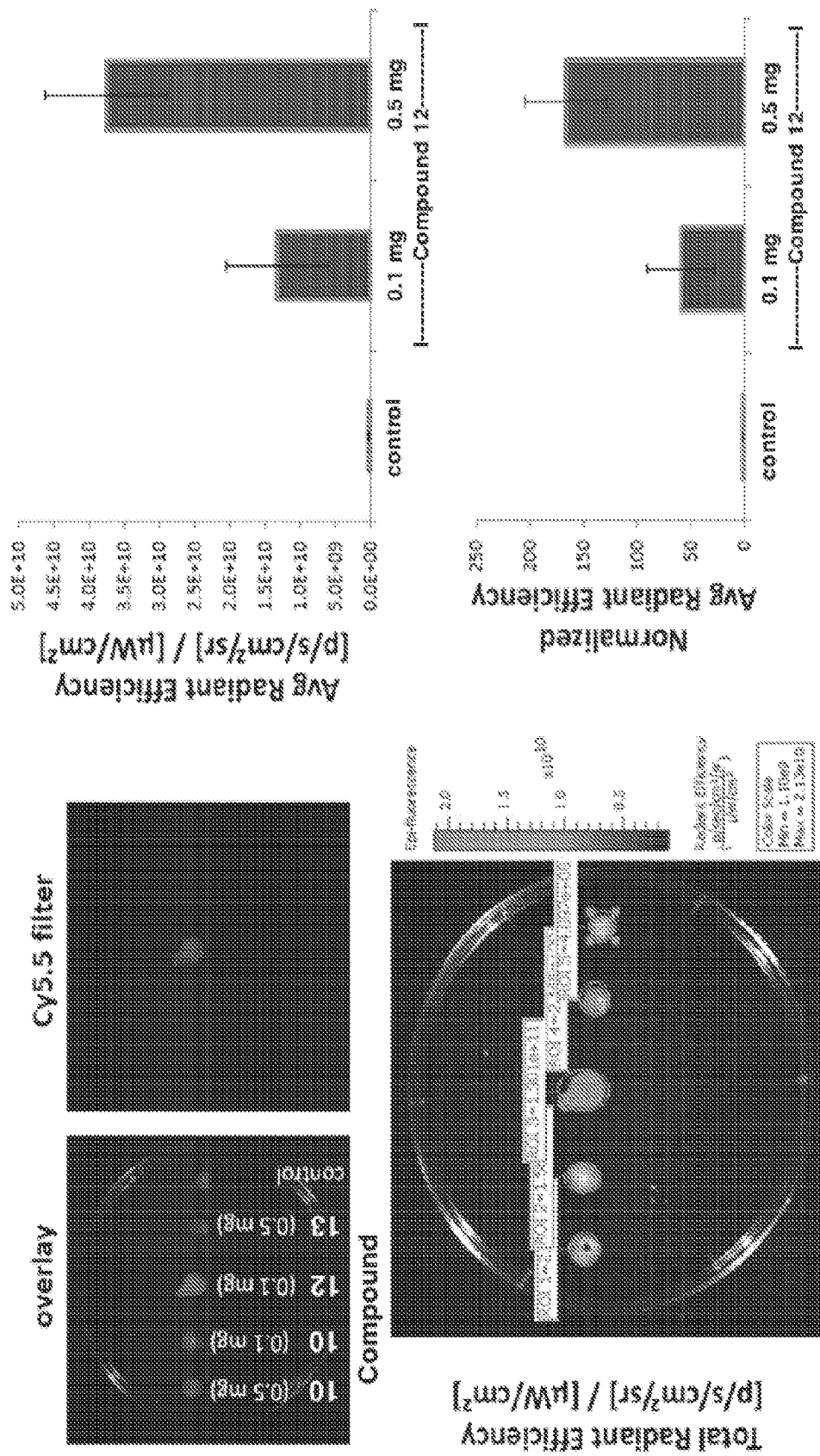


FIGURE 14C

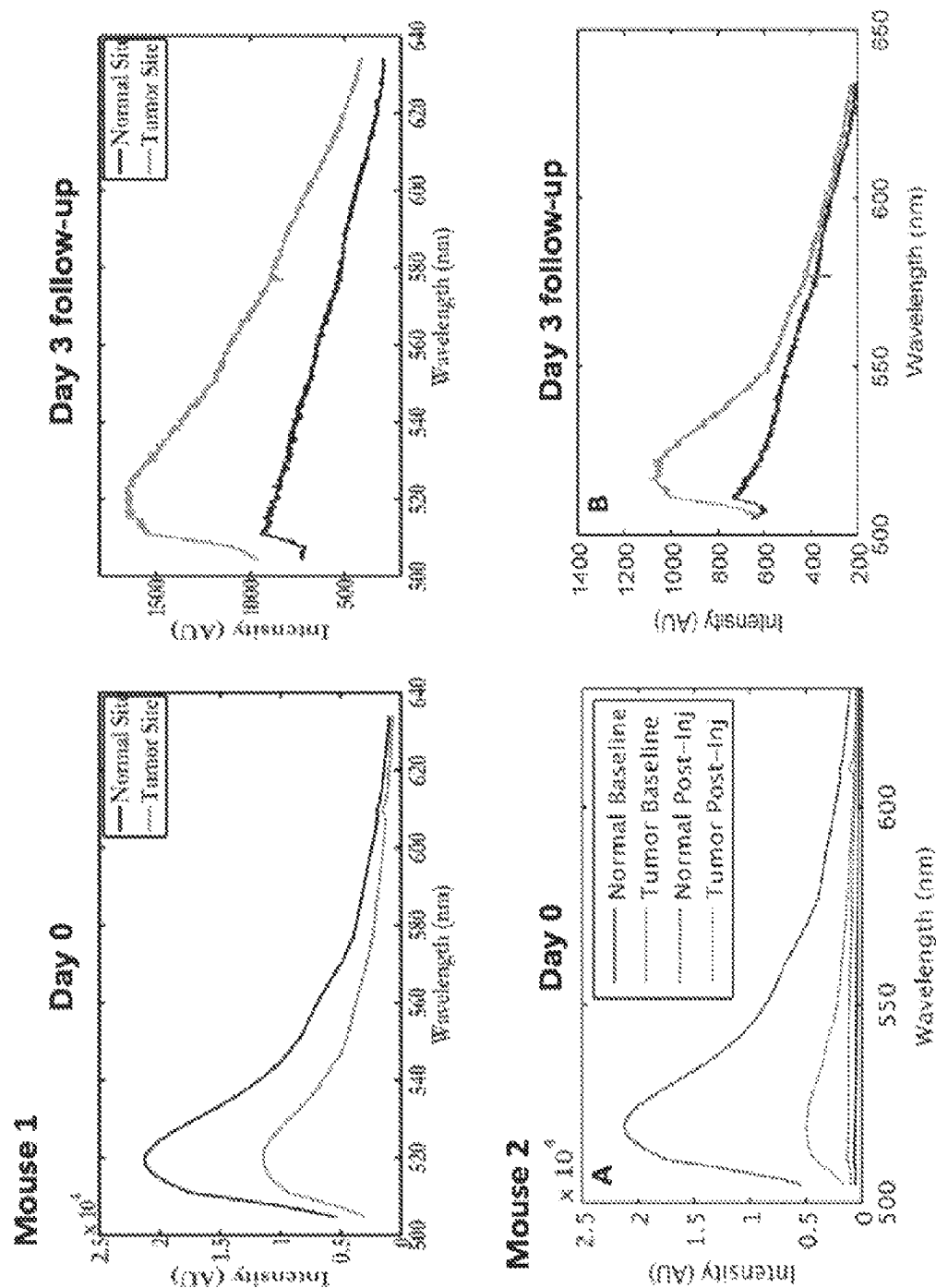


FIGURE 15

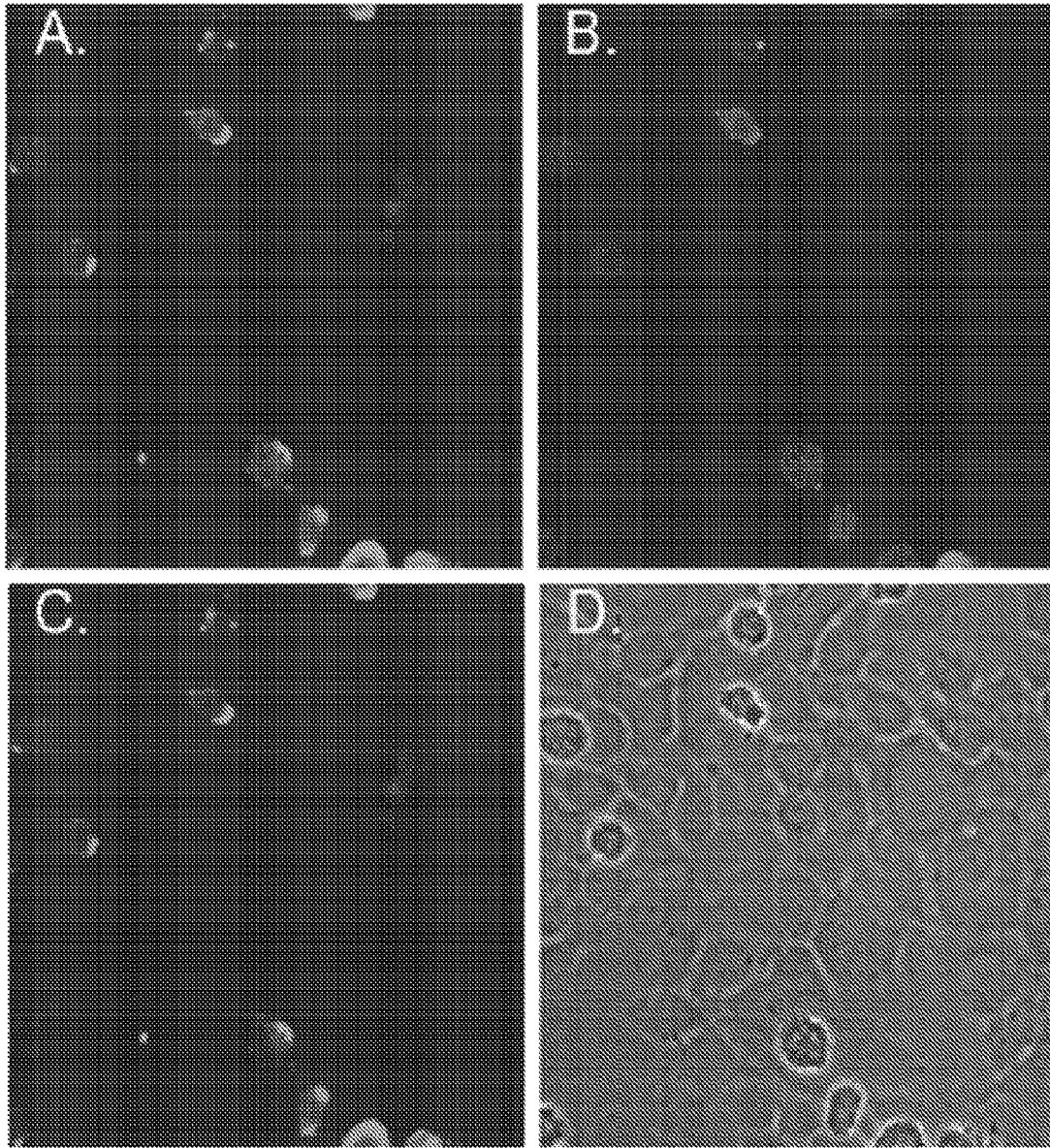


FIGURE 16

MDA tumor recovery Compound 10

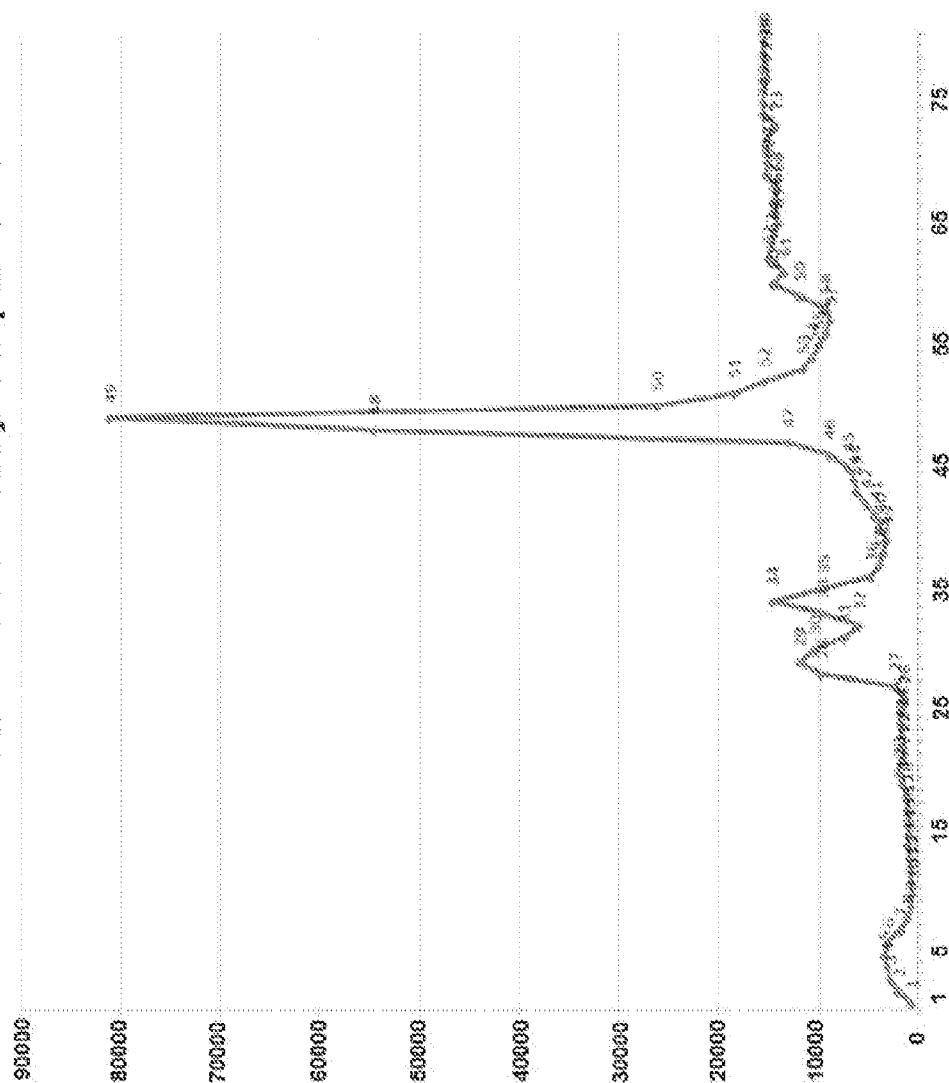
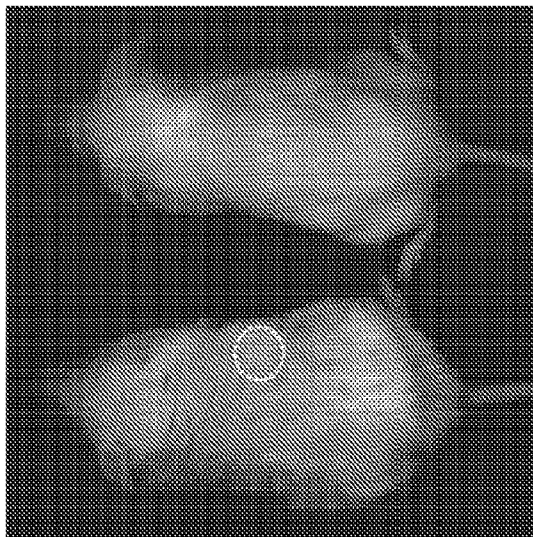


FIGURE 17



1

**SUBSTITUTED INDAZOLES FOR
TARGETING HSP90****CROSS-REFERENCE TO RELATED
APPLICATIONS**

This patent application is a national stage filing under 35 U.S.C. 371 of International Patent Application No. PCT/US2013/031614, filed Mar. 14, 2013, which claims priority to U.S. Provisional Patent Application No. 61/680,107, filed Aug. 6, 2012, the entire contents of which are incorporated by reference in their entireties.

**STATEMENT REGARDING FEDERALLY
SPONSORED RESEARCH**

This invention was made with U.S. Government support awarded by the National Institutes of Health, Grant Nos. 1R01-AI089526-01 and 1R01-AI090644-01. The U.S. Government has certain rights in this invention.

BACKGROUND

Heat shock protein 90 (Hsp90), one of the most abundant proteins expressed in cells, regulates cellular homeostasis by chaperoning protein folding and trafficking. Hsp90 is also highly upregulated in response to stress. The N-terminal domain of Hsp90 includes an ATP binding site, and ATPase activity is necessary for all of its cellular functions. To date, over 200 Hsp90 "client" proteins have been identified and many of these are involved in signal transduction.

Hsp90 has been implicated in diseases such as cancer, and its expression is up-regulated during oncogenesis. High expression is associated with poor prognosis for cancers, such as breast cancer, prostate cancer, non-small cell lung cancer, colorectal cancer and melanoma.

Proteins such as glucose-regulated protein 94 (GRP94) and tumor necrosis factor receptor-associated protein (TRAP1) share homology with Hsp90 and both proteins also possess ATPase activity. While inhibitors of Hsp90 have been shown to have antiproliferative and antitumor activities, current Hsp90 inhibitors may bind nonspecifically to GRP94 and TRAP1. There is a need for compounds that selectively bind to Hsp90.

SUMMARY

In one aspect, the disclosure provides a compound of formula (I):



wherein:

A is an Hsp90 binding component;
 X^1 is selected from the group consisting of $-\text{NH}-$, $-\text{O}-$, $-\text{S}-$, $-\text{C}(\text{O})-$ and $-\text{S}(\text{O})_2-$;
 X^2 is selected from the group consisting of $-\text{NR}-$, $-\text{O}-$, $-\text{S}-$, $-\text{C}(\text{O})-$ and $-\text{S}(\text{O})_2-$;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is selected from the group consisting of a detection moiety, an anti-cancer agent, and an Hsp90 binding component; and

R is selected from the group consisting of hydrogen and a detection moiety.

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In another aspect, the disclosure provides a method of detecting Hsp90 in a sample, comprising:

a) contacting the sample with a compound having the following formula (Ia):



wherein:

A is an Hsp90 binding component;

X^1 is selected from the group consisting of $-\text{NH}-$, $-\text{O}-$, $-\text{S}-$, $-\text{C}(\text{O})-$ and $-\text{S}(\text{O})_2-$;

X^2 is selected from the group consisting of $-\text{NR}-$, $-\text{O}-$, $-\text{S}-$, $-\text{C}(\text{O})-$ and $-\text{S}(\text{O})_2-$;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety; and

b) detecting a signal from the detection moiety.

In another aspect, the disclosure provides a method of detecting cancer in a subject, comprising:

a) contacting a biological sample from the subject with a compound having the following formula (Ia):



wherein:

A is an Hsp90 binding component;

X^1 is selected from the group consisting of $-\text{NH}-$, $-\text{O}-$, $-\text{S}-$, $-\text{C}(\text{O})-$ and $-\text{S}(\text{O})_2-$;

X^2 is selected from the group consisting of $-\text{NR}-$, $-\text{O}-$, $-\text{S}-$, $-\text{C}(\text{O})-$ and $-\text{S}(\text{O})_2-$;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety; and

b) detecting a signal from the detection moiety;

wherein cancer is detected in the sample when the signal from the detection moiety is higher relative to a signal from a reference sample.

In another aspect, the disclosure provides a method of treating cancer in a subject in need of treatment, comprising administering the subject a therapeutically effective amount of a compound having the following formula (Ib):



wherein:

A is an Hsp90 binding component;

X^1 and X^2 are each independently selected from the group consisting of $-\text{NH}-$, $-\text{O}-$, $-\text{S}-$, $-\text{C}(\text{O})-$ and $-\text{S}(\text{O})_2-$;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen; and

B is selected from the group consisting of an anti-cancer agent and an Hsp90 binding component.

In another aspect, the disclosure provides a method of detecting Human Immunodeficiency Virus (HIV) in a subject, the method comprising:

a) contacting a biological sample from the subject with a compound having the following formula (Ia):



wherein:

A is an Hsp90 binding component;

X¹ is selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

X² is selected from the group consisting of —NR—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety; and

b) detecting a signal from the detection moiety;

wherein HIV is detected in the sample when the signal from the detection moiety is higher relative to a signal from a reference sample.

In another aspect, the disclosure provides a pharmaceutical composition comprising a compound of formula (Ib):



wherein:

A is an Hsp90 binding component;

X¹ and X² are each independently selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen; and

B is selected from the group consisting of an anti-cancer agent and an Hsp90 binding component;

and a pharmaceutically acceptable carrier.

In another aspect, the disclosure provides a kit comprising a compound of formula (I) as described herein.

Other aspects and embodiments will become apparent in light of the following description and drawings.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is an SDS-PAGE silver stain showing the effect of different linker structures on Hsp90 recovery and recovery of non-specifically bound proteins. Selectivity towards Hsp90 was demonstrated by including 1 mM compound 5 in the tissue extract prior to mixing.

FIG. 2 is an SDS-PAGE silver stain showing elution of pig mammary gland proteins from ATP-Sepharose media with a compound described herein (8) and two known Hsp90 inhibitors.

FIG. 3 is an SDS-PAGE silver stain showing elution of pig mammary gland proteins from ATP-Sepharose media with various compounds.

FIG. 4 represents fluorescence microscopy images of cancer cell lines treated with a compound described herein (10).

FIG. 5 shows fluorescence microscopy images of metastatic cancer cells and benign cells treated with: (A) an anti-Hsp90 antibody; and (B) compound 10.

FIG. 6 shows fluorescence microscopy images of MCF7 cells treated with compound 10, in the presence or absence of an anti-Hsp90 antibody, and an Hsp90-binding compound.

FIGS. 7A and B show labeling of MDA-MB-468 tumor cells in NOD/SCID mice.

FIG. 8 shows fluorescence microscopy images of xenografts harvested one hour after injection (IP) with compound 10.

FIG. 9 shows the results of competitive binding experiments between compound 10 and the free parent drug compound 5.

FIG. 10 shows the distribution of compound 10 in mice in (A) splenocytes; (B) liver cells; and (C) tumor cells.

FIG. 11 shows quantitative analysis of the distribution of compound 10 in tumor cells, splenocytes and hepatocytes.

FIG. 12 shows fluorescence microscopy images of compound 10 uptake in breast cancer cells.

FIG. 13 shows chromatograms of Hsp90 purified from: (A) pig mammary gland and BT474 cells, and (B) MCF7 cells, treated with compound 10.

FIG. 14 shows: (A) Fluorescence images of HSP90 expression in mouse xenografts; (B) quantitative analysis of compound 10 accumulation in tumors isolated from mice; and (C) accumulation of compound 12 within tumors isolated from mice.

FIG. 15 shows in vivo detection of compound 10 with optical spectroscopy.

FIG. 16 shows images of cells infected with human immunodeficiency virus, treated with compound 10 and an HIV-specific antibody.

FIG. 17 shows recovery of compound 10-bound protein from mice.

DETAILED DESCRIPTION

Described herein are compounds that may selectively bind to Hsp90. The compounds include an Hsp90 binding component, a linker, and an active moiety such as a detection group (e.g., a fluorophore or a radioisotope), an anti-cancer agent, or a second Hsp90 binding component. The compounds described herein may selectively bind to Hsp90 and may have significantly higher affinities for Hsp90 than for proteins with homology to Hsp90, such as GRP94 and TRAP1. The selective nature of the compounds may make them useful probes of Hsp90 in samples, and may allow for the selective targeting of Hsp90. For example, compounds described herein may be used to selectively detect Hsp90 in samples, and to selectively deliver anti-cancer agents to cells expressing high levels of Hsp90.

1. DEFINITIONS

The terminology used herein is for the purpose of describing particular embodiments only and is not intended to be limiting. As used in the specification and the appended claims, the singular forms “a,” “and” and “the” include plural references unless the context clearly dictates otherwise.

Section headings as used in this section and the entire disclosure herein are not intended to be limiting.

For the recitation of numeric ranges herein, each intervening number there between with the same degree of precision is explicitly contemplated. For example, for the range 6-9, the numbers 7 and 8 are contemplated in addition to 6 and 9, and for the range 6.0-7.0, the numbers 6.0, 6.1, 6.2, 6.3, 6.4, 6.5, 6.6, 6.7, 6.8, 6.9 and 7.0 are explicitly contemplated.

As used herein, the term “about” is used synonymously with the term “approximately.” Illustratively, the use of the term “about” indicates that values slightly outside the cited values, namely, plus or minus 10%. Such values are thus encompassed by the scope of the claims reciting the terms “about” and “approximately.”

The terms “administer”, “administering”, “administered” or “administration” refer to any manner of providing a

compound or a pharmaceutical composition (e.g., one described herein), to a subject or patient. Routes of administration can be accomplished through any means known by those skilled in the art. Such means include, but are not limited to, oral, buccal, intravenous, subcutaneous, intramuscular, transdermal, by inhalation and the like.

"Contacting" as used herein, e.g., as in "contacting a sample" refers to contacting a sample directly or indirectly in vitro, ex vivo, or in vivo (i.e. within a subject as defined herein). Contacting a sample may include addition of a compound to a sample (e.g., a sample comprising cells that contain Hsp90), or administration to a subject. Contacting encompasses administration to a solution, cell, tissue, mammal, subject, patient, or human. Further, contacting a cell includes adding an agent to a cell culture.

"Detection moiety" as used herein includes one or more groups that are detectable, either directly or indirectly, by methods such as spectroscopic, photochemical, biochemical, chemical, or other methods. For example, useful detectable moieties or labels include chromophores, fluorophores, biotin, radioactive compounds, and the like. The detection moiety often generates a measurable signal, such as a radioactive, chromogenic, luminescent, or fluorescent signal, which can be used to quantitate the amount of the detection moiety in a sample. In some embodiments a detection moiety may include more than one detectable group, e.g., a fluorophore and a radioactive moiety.

"Effective amount," as used herein, refers to a dosage or an amount of a compound or a composition effective for eliciting a desired effect. This term as used herein may also refer to an amount effective at bringing about a desired in vivo effect in an animal, e.g., a mammal, e.g., a human. For example, in methods of treating cancer, an effective amount may be an amount sufficient to treat the disorder.

"Member atom" as used herein refers to a polyvalent atom (e.g., a C, O, N, or S atom) in a chain or ring system that constitutes a part of the chain or ring. For example, in pyridine, five carbon atoms and one nitrogen atom are member atoms of the ring. In diethyl ether, four carbon atoms and one oxygen atom are member atoms of the chain. Member atoms will be substituted up to their normal valence. For example, in pyridine, the five carbon atoms will each be further substituted with a hydrogen or another substituent.

As used herein, the term "subject" is intended to include human and non-human animals. Exemplary human subjects include a human patient having a disorder, e.g., cancer, or a normal subject. The term "non-human animals" includes all vertebrates, e.g., non-mammals (such as chickens, amphibians, reptiles) and mammals, such as non-human primates, domesticated and/or agriculturally useful animals (such as sheep, dogs, cats, cows, pigs, etc.), and rodents (such as mice, rats, hamsters, guinea pigs, etc.).

As used herein, the term "treat" or "treating" a subject having a disorder refers to administering a compound or a composition described herein to the subject, such that at least one symptom of the disorder is cured, healed, alleviated, relieved, altered, remedied, ameliorated, or improved. Treating includes administering an amount effective to alleviate, relieve, alter, remedy, ameliorate, cure, improve or affect the disorder or the symptoms of the disorder. The treatment may inhibit deterioration or worsening of a symptom of a disorder.

Definitions of specific functional groups and chemical terms are described in more detail below. For purposes of this disclosure, the chemical elements are identified in accordance with the Periodic Table of the Elements, CAS

version, Handbook of Chemistry and Physics, 75th Ed., inside cover, and specific functional groups are generally defined as described therein. Additionally, general principles of organic chemistry, as well as specific functional moieties and reactivity, are described in *Organic Chemistry*, Thomas Sorrell, University Science Books, Sausalito, 1999; Smith and March *March's Advanced Organic Chemistry*, 5th Edition, John Wiley & Sons, Inc., New York, 2001; Larock, *Comprehensive Organic Transformations*, VCH Publishers, Inc., New York, 1989; Carruthers, *Some Modern Methods of Organic Synthesis*, 3rd Edition, Cambridge University Press, Cambridge, 1987; the entire contents of each of which are incorporated herein by reference.

The term "acyl" refers to an alkylcarbonyl, cycloalkylcarbonyl, heterocyclylcarbonyl, arylcarbonyl or heteroarylcarbonyl substituent, any of which may be further substituted (e.g., with one or more substituents).

The term "alkyl" refers to a straight or branched saturated hydrocarbon chain. Alkyl groups may include a specified number of carbon atoms. For example, C₁-C₁₂ alkyl indicates that the alkyl group may have 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 or 12 carbon atoms. An alkyl group may be, e.g., a C₁-C₁₂ alkyl group, a C₁-C₁₀ alkyl group, a C₁-C₈ alkyl group, a C₁-C₆ alkyl group or a C₁-C₄ alkyl group. For example, exemplary C₁-C₄ alkyl groups include methyl, ethyl, n-propyl, isopropyl, n-butyl, sec-butyl, isobutyl and tert-butyl groups. An alkyl group may be optionally substituted with one or more substituents.

The term "alkylenyl" refers to a divalent alkyl group, examples of which include but are not limited to —CH₂—, —CH₂CH₂—, —CH₂CH₂CH₂— and —CH₂CH(CH₃)CH₂—. An alkylenyl group may be optionally substituted with one or more substituents.

The term "alkenyl" refers to a straight or branched hydrocarbon chain having one or more double bonds. Alkenyl groups may include a specified number of carbon atoms. For example, C₂-C₁₂ alkenyl indicates that the alkenyl group may have 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 or 12 carbon atoms. An alkenyl group may be, e.g., a C₂-C₁₂ alkenyl group, a C₂-C₁₀ alkenyl group, a C₂-C₈ alkenyl group, a C₂-C₆ alkenyl group or a C₂-C₄ alkenyl group. Examples of alkenyl groups include but are not limited to allyl, propenyl, 2-butenyl, 3-hexenyl and 3-octenyl groups. One of the double bond carbons may optionally be the point of attachment of the alkenyl substituent. An alkenyl group may be optionally substituted with one or more substituents.

The term "alkenylenyl" refers to a divalent alkenyl group, examples of which include but are not limited to —CH=CH—, —CH=CH—CH₂—, —CH=CH—CH₂—CH₂— and —CH₂—CH=CH—CH₂—. An alkenylenyl group may be optionally substituted with one or more substituents.

The term "alkynyl" refers to a straight or branched hydrocarbon chain having one or more triple bonds. Alkynyl groups may include a specified number of carbon atoms. For example, C₂-C₁₂ alkynyl indicates that the alkynyl group may have 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 or 12 carbon atoms. An alkynyl group may be, e.g., a C₂-C₁₂ alkynyl group, a C₂-C₁₀ alkynyl group, a C₂-C₈ alkynyl group, a C₂-C₆ alkynyl group or a C₂-C₄ alkynyl group. Examples of alkynyl groups include but are not limited to ethynyl, propargyl, and 3-hexynyl. One of the triple bond carbons may optionally be the point of attachment of the alkynyl substituent. An alkynyl group may be optionally substituted with one or more substituents.

The term "alkynylenyl" refers to a divalent alkynyl group, examples of which include but are not limited to —C≡C—,

—C≡C—CH₂—, —C≡C—CH₂—CH₂— and —CH₂—C≡C—CH₂—. An alkynylene group may be optionally substituted with one or more substituents.

The term “aryl” refers to an aromatic monocyclic, bicyclic, or tricyclic hydrocarbon ring system, wherein any ring atom capable of substitution can be substituted (e.g., with one or more substituents). Examples of aryl moieties include but are not limited to phenyl, naphthyl, and anthracenyl. Aryl groups may be optionally substituted with one or more substituents.

The term “arylalkyl” refers to an alkyl moiety in which at least one alkyl hydrogen atom is replaced with an aryl group. Arylalkyl includes groups in which more than one hydrogen atom has been replaced with an aryl group. Examples of arylalkyl groups include but are not limited to benzyl, 2-phenylethyl, 3-phenylpropyl, 9-fluorenyl, benzhydryl, and trityl groups. Arylalkyl groups may be optionally substituted with one or more substituents, on either the aryl moiety or the alkyl moiety.

The term “cycloalkyl” as used herein refers to non-aromatic, saturated or partially unsaturated cyclic, bicyclic, tricyclic or polycyclic hydrocarbon groups having 3 to 12 carbons. Any ring atom can be substituted (e.g., with one or more substituents). Cycloalkyl groups can contain fused rings. Fused rings are rings that share one or more common carbon atoms. Examples of cycloalkyl groups include but are not limited to cyclopropyl, cyclobutyl, cyclopentyl, cyclohexyl, cyclohexenyl, cyclohexadienyl, methylcyclohexyl, adamantyl, norbornyl, norbornenyl, tetrahydronaphthalenyl and dihydroindenyl. Cycloalkyl groups may be optionally substituted with one or more substituents.

The term “cycloalkylalkyl”, as used herein, refers to an alkyl group in which at least one hydrogen atom is replaced with a cycloalkyl group. Cycloalkylalkyl groups include those in which more than one hydrogen atom of the alkyl group is replaced with a cycloalkyl group. Examples of cycloalkylalkyl groups include but are not limited to cyclohexylmethyl, cyclopentylmethyl, cyclobutylmethyl and cyclopropylmethyl. Cycloalkylalkyl groups can be optionally substituted with one or more substituents, on either the cycloalkyl moiety or the alkyl moiety.

The term “halo” or “halogen” as used herein refers to any radical of fluorine, chlorine, bromine or iodine.

The term “haloalkyl” as used herein refers to an alkyl group as defined herein, in which one or more hydrogen atoms are replaced with halogen atoms, and includes alkyl moieties in which all hydrogens have been replaced with halogens (e.g., perfluoroalkyl such as CF₃).

“Heteroalkyl” refers to an alkyl, alkenyl or alkynyl group as defined herein, wherein at least one carbon atom of the alkyl group is replaced with a heteroatom. Heteroalkyl groups may contain from 1 to 18 non-hydrogen atoms (carbon and heteroatoms) in the chain, or 1 to 12 atoms, or 1 to 6 atoms, or 1 to 4 atoms. Heteroalkyl groups may be straight or branched, and saturated or unsaturated. Unsaturated heteroalkyl groups have one or more double bonds and/or one or more triple bonds. Heteroalkyl groups may be unsubstituted or substituted. Exemplary heteroalkyl groups include but are not limited to alkoxyalkyl (e.g., methoxymethyl), and aminoalkyl (e.g., alkylaminoalkyl and dialkylaminoalkyl). Heteroalkyl groups may be optionally substituted with one or more substituents.

The term “heteroalkylene” refers to a divalent heteroalkyl group, examples of which include but are not limited to —CH₂OCH₂—, —CH₂NHCH₂—, polyethyleneglycol groups (e.g., —(CH₂CH₂O)_n—), polyethyleneimine groups

(e.g., —(CH₂CH₂NH)_n—), and the like. A heteroalkylene group may be optionally substituted with one or more substituents.

The term “heteroaryl” as used herein refers to an aromatic 5-8 membered monocyclic, 8-12 membered bicyclic, or 11-14 membered tricyclic ring system having 1-3 heteroatoms if monocyclic, 1-6 heteroatoms if bicyclic, or 1-9 heteroatoms if tricyclic, said heteroatoms independently selected from O, N, S, P and Si (e.g., carbon atoms and 1-3, 1-6, or 1-9 heteroatoms independently selected from O, N, S, P and Si if monocyclic, bicyclic, or tricyclic, respectively). Any ring atom can be substituted (e.g., with one or more substituents). Heteroaryl groups can contain fused rings, which are rings that share one or more common atoms. Examples of heteroaryl groups include but are not limited to radicals of pyridine, pyrimidine, pyrazine, pyridazine, pyrrole, imidazole, pyrazole, oxazole, isoxazole, furan, thiazole, isothiazole, thiophene, quinoline, isoquinoline, quinoxaline, quinazoline, cinnoline, indole, isoindole, indolizine, indazole, benzimidazole, phthalazine, pteridine, carbazole, carboline, phenanthridine, acridine, phenanthroline, phenazine, naphthyridines and purines. Heteroaryl groups may be optionally substituted with one or more substituents.

The term “heteroarylalkyl” refers to an alkyl moiety in which at least one alkyl hydrogen atom is replaced with a heteroaryl group. Heteroarylalkyl includes groups in which more than one hydrogen atom has been replaced with a heteroaryl group. Examples of heteroarylalkyl groups include but are not limited to imidazolylmethyl (e.g., 1H-imidazol-2-ylmethyl and 1H-imidazol-4-ylmethyl), pyridinylmethyl (e.g., pyridin-3-ylmethyl and pyridin-4-ylmethyl), pyrimidinylmethyl (e.g., pyrimidin-5-ylmethyl), furylmethyl (e.g., fur-2-ylmethyl and fur-3-ylmethyl), and thienylmethyl (e.g., thien-2-ylmethyl and thien-3-ylmethyl) groups. Heteroarylalkyl groups may be optionally substituted with one or more substituents, on either the heteroaryl moiety or the alkyl moiety.

The term “heteroatom”, as used herein, refers to a non-carbon or hydrogen atom such as a nitrogen, sulfur, oxygen, silicon or phosphorus atom. Groups containing more than one heteroatom may contain different heteroatoms.

The term “heterocyclyl”, as used herein, refers to a nonaromatic, saturated or partially unsaturated 3-10 membered monocyclic, 8-12 membered bicyclic, or 11-14 membered tricyclic ring system having 1-3 heteroatoms if monocyclic, 1-6 heteroatoms if bicyclic, or 1-9 heteroatoms if tricyclic, said heteroatoms selected from O, N, S, Si and P (e.g., carbon atoms and 1-3, 1-6, or 1-9 heteroatoms of O, N, S, Si and P if monocyclic, bicyclic, or tricyclic, respectively). Any ring atom can be substituted (e.g., with one or more substituents). Heterocyclyl groups can contain fused rings, which are rings that share one or more common atoms. Examples of heterocyclyl groups include but are not limited to radicals of tetrahydrofuran, tetrahydrothiophene, tetrahydropyran, oxetane, piperidine, piperazine, morpholine, pyrrolidine, pyrrolidine, indoline, tetrahydropyridine, dihydropyran, thianthrene, pyran, benzopyran, xanthene, phenoxathiin, phenothiazine, furazan, lactones, lactams such as azetidinones and pyrrolidinones, sultams, sultones, and the like. Heterocyclyl groups may be optionally substituted with one or more substituents.

The term “heterocyclylalkyl” refers to an alkyl moiety in which at least one alkyl hydrogen atom is replaced with a heterocyclyl group. Heterocyclylalkyl includes groups in which more than one hydrogen atom has been replaced with a heterocyclyl group. Examples of heterocyclylalkyl groups

include but are not limited to oxetanylmethyl, morpholinomethyl, and pyrrolidinylmethyl groups, and the like. Heterocyclalkyl groups may be optionally substituted with one or more substituents, on either the heterocyclalkyl moiety or the alkyl moiety.

The term "hydroxy" refers to an —OH radical. The term "alkoxy" refers to an —O— alkyl radical. The term "aryloxy" refers to an —O-aryl radical.

The term "oxo" refers to an oxygen atom, which forms a carbonyl when attached to carbon, an N-oxide when attached to nitrogen, and a sulfoxide or sulfone when attached to sulfur (i.e. =O).

The term "mercapto" or "thiol" refers to an —SH radical. The term "thioalkoxy" or "thioether" refers to an —S-alkyl radical. The term "thioaryloxy" refers to an —S-aryl radical.

The term "substituents" refers to a group "substituted" on an alkyl, alkenyl, alkynyl, cycloalkyl, heterocyclalkyl, aryl, arylalkyl, heteroaryl or heteroarylalkyl group at any atom of that group. Any atom can be substituted. Suitable substituents include, without limitation: acyl, acylamido, acyloxy, alkoxy, alkyl, alkenyl, alkynyl, amido, amino, carboxy, cyano, ester, halo, hydroxy, imino, nitro, oxo (e.g., C=O), phosphonate, sulfinyl, sulfonyl, sulfonate, sulfonamino, sulfonamido, thioamido, thiol, thiooxo (e.g., C=S), and ureido. In embodiments, substituents on a group are independently any one single, or any combination of the aforementioned substituents. In embodiments, a substituent may itself be substituted with any one of the above substituents.

The above substituents may be abbreviated herein. For example, the abbreviations Me, Et, Ph and Bn represent methyl, ethyl, phenyl and benzyl, respectively. A more comprehensive list of standard abbreviations used by organic chemists appears in a table entitled Standard List of Abbreviations of the Journal of Organic Chemistry. The abbreviations contained in said list are hereby incorporated by reference.

For compounds described herein, groups and substituents thereof may be selected in accordance with permitted valence of the atoms and the substituents, and such that the selections and substitutions result in a stable compound, e.g., a compound that does not spontaneously undergo transformation such as by rearrangement, cyclization, elimination, etc.

Where substituent groups are specified by their conventional chemical formulae, written from left to right, they optionally encompass substituents resulting from writing the structure from right to left, e.g., —CH₂O— optionally also recites —OCH₂—.

In accordance with a convention used in the art, the group:



is used in structural formulae herein to depict the bond that is the point of attachment of the moiety or substituent to the core or backbone structure.

2. COMPOUNDS

Compounds that may selectively bind to Hsp90 include compounds of formula (I):



wherein:

A is an Hsp90 binding component;

X¹ is selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

X² is selected from the group consisting of —NR—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is selected from the group consisting of a detection moiety, an anti-cancer agent, and an Hsp90 binding component; and

R is selected from the group consisting of hydrogen and a detection moiety.

Compounds that may be used in methods described herein also include compounds of formula (Ia):



wherein:

A is an Hsp90 binding component;

X¹ is selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

X² is selected from the group consisting of —NR—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety.

Compounds that may be used in methods described herein also include compounds of formula (Ib):



wherein:

A is an Hsp90 binding component;

X¹ and X² are each independently selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen; and

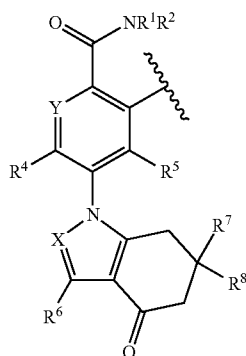
B is selected from the group consisting of an anti-cancer agent and an Hsp90 binding component.

a. Hsp90 Binding Components

In the compounds of formula (I), A is an Hsp90 binding component. The compounds of formula (I) include at least one Hsp90 binding component, and may include two Hsp90 binding components in embodiments in which B is also an Hsp90 component. Any molecule that binds to Hsp90 can be used as the basis for the Hsp90 binding component. Based on the complete structure of the compounds of formula (I), it will be understood that the Hsp90 binding component corresponds to an Hsp90 binding compound in which an atom or a group of atoms is removed, to provide a point of attachment to the —X¹-L-X²-B moiety of the remainder of the compound of formula (I) (or the —X²-L-X¹-A moiety in embodiments in which B is an Hsp90 binding component).

In embodiments, an Hsp90 binding component has the following formula (II):

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wherein:

R¹ and R² are independently selected from the group consisting of —H and —C₁₋₈-alkyl;

Y is CR³ or N;

R³ is —H, —F or —OCH₃;

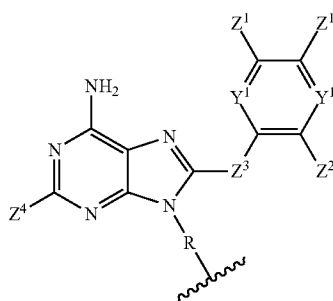
R⁴ and R⁵ are independently selected from the group consisting of —H, —F and —OCH₃;

R⁶ is —C₁₋₈-alkyl, —C₂₋₈-alkenyl, —C₂₋₈-alkynyl, —C₃₋₈-cycloalkenyl, —C₃₋₈-cycloalkenyl-C₁₋₈-alkyl, —C₃₋₈-cycloalkyl, —C₃₋₈-cycloalkyl-C₁₋₈-alkyl, aryl, aryl-C₁₋₈-alkyl, halo-C₁₋₈-alkyl, heteroaryl, heteroaryl-C₁₋₈-alkyl, heterocyclyl, heterocyclyl-C₁₋₈-alkyl, or hydroxy-C₁₋₈-alkyl;

R⁷ and R⁸ are independently selected from the group consisting of —H and —C₁₋₈-alkyl; or R⁷ and R⁸ taken together with the carbon atom to which they are attached form a C₃₋₈-cycloalkyl group;

X is N or CR⁹; and

R⁹ is —H or —C₁₋₈-alkyl; and



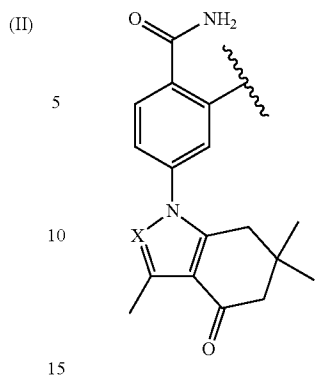
represents the point of attachment to the —X¹-L-X²—B moiety of the compound of formula (I).

In embodiments of formula (II), R¹ and R² are each —H, Y is CR³, R³ is —H, R⁴ and R⁵ are each —H, R⁶ is —C₁₋₈-alkyl (e.g., methyl) or halo-C₁₋₈-alkyl (e.g., trifluoromethyl), R⁷ and R⁸ are independently —C₁₋₈-alkyl (e.g., methyl), and X is N. In embodiments, formula (II) is the following:



In embodiments, an Hsp90 binding component has the following formula (III):

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wherein:

each Y¹ is independently selected from the group consisting of CH and N;

R is alkylenyl or heteroalkylenyl;

each Z¹ is taken together with the carbon atoms to which they are attached to form a heterocyclic ring;

Z² is hydrogen or halo;

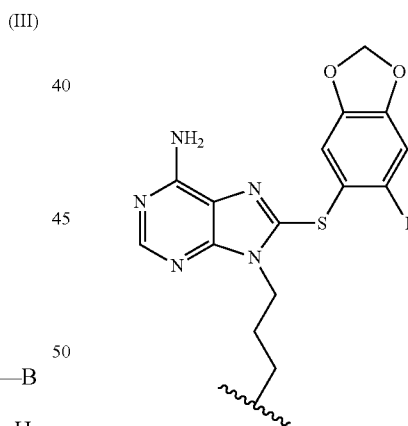
Z³ is CH₂, S, O, or NH;

Z⁴ is hydrogen or halo; and



represents the point of attachment to the remainder of the compound of formula (I), e.g., to the —X¹-L-X²—B moiety, or to the X²-L-X¹-A moiety when B is an Hsp90 binding component having the formula (III).

In embodiments, a group of formula (III) is the following:



The Hsp90 binding component may comprise any known Hsp90 binding compound or a portion thereof that retains affinity for Hsp90. Other compounds that bind to Hsp90 are known in the art, and include, for example, geldanamycin, herbimycin, radicicol, deguelin, derrubone and macebecin. Other compounds that bind to Hsp90 may include but are not limited to those described in U.S. Pat. Nos. 7,358,370, 7,678,803, 7,906,529 and 7,928,135, U.S. Patent Publication No. 2011/0183977, and International Patent Publication Nos. WO2008/130879 and WO2006/084030.

b. Linker

In the compounds of formula (I), L is a divalent linker comprising at least twelve member atoms independently

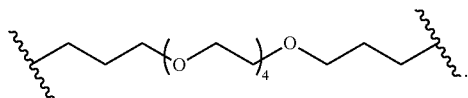
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selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen. In embodiments, L is a heteroalkylenyl group comprising at least twelve member atoms. In some embodiments, L comprises 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49 or 50 atoms. In some embodiments, the heteroalkylenyl group may be optionally substituted (e.g., a carbon may be substituted with an oxo group).

In embodiments, L is a straight-chain heteroalkylenyl group. In embodiments, L includes at least one monomer of a polyethylene glycol group (i.e. $-(CH_2CH_2O)-$), or it may include at least two, three, four, five, six or more such monomers.

In some embodiments, L has the formula $-(CH_2)_m-(OCH_2CH_2)_n-O-(CH_2)_p-$, wherein m is 2 or 3, n is 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20, and p is 2 or 3. In embodiments, m is 2. In embodiments, m is 3. In embodiments, n is 4, 5, 6, 7, 8, 9 or 10. In embodiments, n is 4. In embodiments, p is 2. In embodiments, p is 3.

A suitable linker has the following formula:



c. X^1 and X^2

The groups X^1 and X^2 in the compounds formula (I) are each independently selected from the group consisting of $-NH-$, O , S , $C(O)-$ and $-S(O)_2-$. These groups serve as bridges between the linker and the A and B groups of the compound of formula (I). During the synthesis of these molecules (described in further detail below), the X^1 and X^2 groups may be derived from a linker precursor compound. For example, in some embodiments X^1 and X^2 may be $-NH-$. In such embodiments, a suitable linker precursor compound may have the formula $H_2N-Linker-NH_2$. In this case, the terminal amino groups of the linker precursor compound provide the source of the X^1 and X^2 groups.

In some embodiments, X^2 may be NR , wherein R is selected from the group consisting of hydrogen and a detection moiety. Such compounds may be synthesized, for example, by first synthesizing a group in which X^2 is NH and then further derivatizing the NH group with a compound comprising an additional detection moiety.

d. Detection Moieties

In embodiments of the compounds of formula (I), B is a detection moiety. The detection moiety comprises one or more groups that are, either directly or indirectly, detectable via a method such as a spectroscopic method. Exemplary detection moieties may comprise, for example, a chromophore, a fluorophore, a luminescent moiety or a radioisotope. In some embodiments, B comprises one or more detection moieties, such as, for example, a fluorophore and a radioisotope.

In embodiments, the detection moiety comprises a fluorophore. Suitable fluorophores include fluoresceins, rhodamines, coumarins, cyanines, and boron-dipyrromethenes (also known as BODIPYs). The fluorophores may be attached to the remainder of a compound of formula (I), for example, by using a reagent comprising a fluorophore and a reactive group such as a carboxylic acid, an isothiocyanate, a maleimide, or an ester such as a succinimidyl, pentafluorophenyl or tetrafluorophenyl ester. Such groups may react

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with a group present on a linker precursor compound, such as an amine, to attach the fluorophore to the remainder of the molecule of formula (I).

Suitable reagents comprising fluorophores, which may be used to prepare compounds of formula (I), are known in the art, and include but are not limited to fluoresceins, rhodamines, coumarins, cyanines and boron-dipyrromethenes. For example, reagents comprising fluorophores that are commercially available include but are not limited to: 5- and 6-carboxyfluoresceins and esters thereof; fluorescein-5-isothiocyanate and fluorescein-6-isothiocyanate; BODIPY® dyes commercially available from Molecular Probes; Alexa Fluor® dyes commercially available from Molecular Probes; CyDye fluors commercially available from GE Healthcare Biosciences, including but not limited to Cy3, Cy 5, Cy5.5 and Cy 7 esters; and VivoTag™ fluorophores available from PerkinElmer, including but not limited to VivoTag 645, VivoTag 645-MAL, VivoTag 680, VivoTag 680-MAL, VivoTag 680 XL, VivoTag-S 680, VivoTag 750, VivoTag-S 750, VivoTag 750-MAL, and VivoTag 800.

It will be understood by the skilled artisan that when B is a detection moiety, e.g., a detection moiety comprising a fluorophore, the detection moiety may include the detectable moiety as well as additional atoms or groups of atoms. For example, as will be further described in the synthesis section, a fluorophore may be attached to the remainder of a compound of formula (I), for example, by reacting a compound of the formula $A-X_1-L-NH_2$ with a reagent comprising a fluorophore and a reactive group, such as an isothiocyanate or N-succinimidyl ester. A reaction with an isothiocyanate will produce a compound $A-X_1-L-NH-C(S)-NH-fluorophore$. In such instances, it is understood that the $-NH-$ group attached to $-L$ corresponds to the $-X_2-$ group of formula (I), while the group $-B$ corresponding to the “detection moiety” includes not only the fluorophore but also the $-C(S)-NH-$ linking atoms.

In other embodiments, the detection moiety may comprise a radioisotope. Such detection moieties may be useful for detection in samples by scintillation counting, or for radioimaging applications. Suitable radioisotopes include, but are not limited to, tritium (i.e. hydrogen-3), carbon-14, nitrogen-15, phosphorus-32, iodine-125 and iodine-131. For example, compounds comprising iodine-131 may be useful as radiopharmaceuticals.

e. Anti-Cancer Agents

In embodiments of the compounds of formula (I), B comprises an anti-cancer agent. Exemplary anti-cancer/chemotherapeutic agents include, but are not limited to, the following:

alkylating agents (including, without limitation, nitrogen mustards, ethylenimine derivatives, alkyl sulfonates, nitrosoureas and triazenes): uracil mustard (Aminouracil Mustard®, Chlorethaminacil®, Demethylodopan®, Desmethyldopan®, Haemanthamine®, Nordopan®, Uracil nitrogen Mustard®, Uracillost®, Uracilmostaza®, Uramustin®, Uramustine®), bendamustine (Treakisym®, Ribomustin®, Treand®) chlormethine (Mustargen®), cyclophosphamide (Cytosan®, Neosar®, Clafen®, Endoxan®, Procytox®, Revimmune™), ifosfamide (Mitoxana®), melphalan (Alkeran®), Chlorambucil (Leukeran®), pipobroman (Amedel®, Vercyte®), triethylenemelamine (Hemel®, Hexylen®, Hexastat®), triethylenethiophosphoramine, Temozolomide (Temodar®), thiotepa (Thiopex®, busulfan (Busilvex®, Myleran®), carmustine (BiCNU®), lomustine (CeeNU®), streptozocin (Zanosar®), estramustine (Emcynt®, Estracit®), fotemustine, ifofulven, mannosulfan,

mitobronitol, nimustine, procarbazine, ranimustine, semustine, triaziquone, treosulfan, and Dacarbazine (DTIC-Dome®).

anti-EGFR antibodies (e.g., cetuximab (Erbix®), panitumumab (Vectibix®), and gefitinib (Iressa®)).

anti-Her-2 antibodies (e.g., trastuzumab (Herceptin®) and other antibodies from Genentech).

antimetabolites (including, without limitation, folic acid antagonists (also referred to herein as antifolates), pyrimidine analogs, purine analogs and adenosine deaminase inhibitors): methotrexate (Rheumatrex®, Trexall®), 5-fluorouracil (Adrucil®, Efudex®, Fluoroplex®), floxuridine (FUDF®), carmofur, cytarabine (Cytosar-U®, Tarabine PFS), 6-mercaptopurine (Puri-Nethol®), 6-thioguanine (Thioguanine Tabloid®), fludarabine phosphate (Fludara®), pentostatin (Nipent®), pemetrexed (Alimta®), raltitrexed (Tomudex®), cladribine (Leustatin®), clofarabine (Clofarex®, Clolar®), mercaptopurine (Puri-Nethol®), capecitabine (Xeloda®), nelarabine (Arranon®), azacitidine (Vidaza®), decitabine (Dacogen®), enocitabine (Sunrabin®), sapacitabine, tegafur-uracil, tiazofurine, tioguanine, trofosfamide, and gemcitabine (Gemzar®).

vinca alkaloids: vinblastine (Velban®, Velsar®), vincristine (Vincasar®, Oncovin®), vindesine (Eldisine®), vinorelbine (Navelbine®), vinflunine (Jaylor®).

platinum-based agents: carboplatin (Paraplat®, Paraplatin®), cisplatin (Platinol®), oxaliplatin (Eloxatin®), nedaplatin, satraplatin, triplatin.

anthracyclines: daunorubicin (Cerubidine®, Rubidomycin®), doxorubicin (Adriamycin®), epirubicin (Ellence®), idarubicin (Idamycin®), mitoxantrone (Novantrone®), valrubicin (Valstar®), aclarubicin, amrubicin, liposomal doxorubicin, liposomal daunorubicin, pirarubicin, pixantrone, zorubicin.

topoisomerase inhibitors: topotecan (Hycamtin®), irinotecan (Camptosar®), etoposide (Toposar®, VePesid®), teniposide (Vumon®), lamellarin D, SN-38, camptothecin (e.g., IT-101), belotecan, rubitecan.

taxanes: paclitaxel (Taxol®), docetaxel (Taxotere®), larotaxel, cabazitaxel, ortataxel, tesetaxel.

antibiotics: actinomycin (Cosmegen®), bleomycin (Blenoxane®), hydroxyurea (Droxia®, Hydrea®), mitomycin (Mitozytrex®, Mutamycin®).

immunomodulators: lenalidomide (Revlimid®), thalidomide (Thalomid®).

immune cell antibodies: alemtuzumab (Campath®), gemtuzumab (Myelotarg®), rituximab (Rituxan®), tositumomab (Bexxar®).

interferons (e.g., IFN-alpha (Alferon®, Roferon-A®, Intron®-A) or IFN-gamma (Actimmune®)).

interleukins: IL-1, IL-2 (Proleukin®), IL-24, IL-6 (Sigosix®), IL-12.

HSP90 inhibitors (e.g., geldanamycin or any of its derivatives). In certain embodiments, the HSP90 inhibitor is selected from geldanamycin, 17-alkylamino-17-desmethoxygeldanamycin ("17-AAG") or 17-(2-dimethylaminoethyl)amino-17-desmethoxygeldanamycin ("17-DMAG").

anti-androgens which include, without limitation nilutamide (Nilandron®) and bicalutamide (Caxodex®).

antiestrogens which include, without limitation tamoxifen (Nolvadex®), toremifene (Fareston®), letrozole (Femara®), testolactone (Teslac®), anastrozole (Arimidex®), bicalutamide (Casodex®), exemestane (Aromasin®), flutamide (Eulexin®), fulvestrant (Faslodex®), raloxifene (Evista®, Keoxifene®) and raloxifene hydrochloride.

anti-hypercalcaemia agents which include without limitation gallium (III) nitrate hydrate (Ganite®) and pamidronate disodium (Aredia®).

apoptosis inducers which include without limitation ethanol, 2-[[[3-(2,3-dichlorophenoxy)propyl]amino]-(9CI), gambogic acid, elesclomol, embelin and arsenic trioxide (Trisenox®).

Aurora kinase inhibitors which include without limitation binucleine 2.

Bruton's tyrosine kinase inhibitors which include without limitation terreic acid.

calcineurin inhibitors which include without limitation cypermethrin, deltamethrin, fenvalerate and tyrphostin 8.

CaM kinase II inhibitors which include without limitation 5-Isoquinolinesulfonic acid, 4-[[{2S)-2-[(5-isoquinoliny)sulfonyl]methylamino]-3-oxo-3-{4-phenyl-1-piperazinyl}propyl]phenyl ester and benzenesulfonamide.

CD45 tyrosine phosphatase inhibitors which include without limitation phosphonic acid.

CDC25 phosphatase inhibitors which include without limitation 1,4-naphthalene dione, 2,3-bis[(2-hydroxyethyl)thio]-(9CI).

CHK kinase inhibitors which include without limitation debromohymenialdisine.

cyclooxygenase inhibitors which include without limitation 1H-indole-3-acetamide, 1-(4-chlorobenzoyl)-5-methoxy-2-methyl-N-(2-phenylethyl)-(9CI), 5-alkyl substituted 2-arylamino-phenylacetic acid and its derivatives (e.g., celecoxib (Celebrex®), rofecoxib (Vioxx®), etoricoxib (Arcoxia®), lumiracoxib (Prexige®), valdecoxib (Bextra®) or 5-alkyl-2-arylamino-phenylacetic acid).

cRAF kinase inhibitors which include without limitation 3-(3,5-dibromo-4-hydroxybenzylidene)-5-iodo-1,3-dihydroindol-2-one and benzamide, 3-(dimethylamino)-N-[3-[(4-hydroxybenzoyl)amino]-4-methylphenyl]-(9CI).

cyclin dependent kinase inhibitors which include without limitation olomoucine and its derivatives, purvalanol B, roscovitine (Seliciclib®), indirubin, kenpaullone, purvalanol A and indirubin-3'-monooxime.

cysteine protease inhibitors which include without limitation 4-morpholinecarboxamide, N-[(1S)-3-fluoro-2-oxo-1-(2-phenylethyl)propyl]amino]-2-oxo-1-(phenylmethyl)ethyl]-(9CI).

DNA intercalators which include without limitation pliamycin (Mithracin®) and daptomycin (Cubicin®).

DNA strand breakers which include without limitation bleomycin (Blenoxane®).

E3 ligase inhibitors which include without limitation N-[(3,3,3-trifluoro-2-trifluoromethyl)propionyl)sulfanilamide].

EGF Pathway Inhibitors which include, without limitation tyrphostin 46, EKB-569, erlotinib (Tarceva®), gefitinib (Iressa®), lapatinib (Tykerb®) and those compounds that are generically and specifically disclosed in WO 97/02266, EP 0 564 409, WO 99/03854, EP 0 520 722, EP 0 566 226, EP 0 787 722, EP 0 837 063, U.S. Pat. No. 5,747,498, WO 98/10767, WO 97/30034, WO 97/49688, WO 97/38983 and WO 96/33980.

farnesyltransferase inhibitors which include without limitation a-hydroxyfarnesylphosphonic acid, butanoic acid, 2-[(2S)-2-[[[(2S,3S)-2-[[[(2R)-2-amino-3-mercaptopropyl]amino]-3-methylpent-yl]oxy]-1-oxo-3-phenylpropyl]amino]-4-(methylsulfonyl)-1-methylethylester (2S)-(9CI), tipifarnib (Zarnestra®), and manumycin A.

Flk-1 kinase inhibitors which include without limitation 2-propenamide, 2-cyano-3-[4-hydroxy-3,5-bis(1-methylethyl)phenyl]-N-(3-phenylpropyl)-(2E)-(9CI).

glycogen synthase kinase-3 (GSK3) inhibitors which include without limitation indirubin-3'-monooxime.

histone deacetylase (HDAC) inhibitors which include without limitation suberoylanilide hydroxamic acid (SAHA), [4-(2-amino-phenylcarbamoyl)-benzyl]-carbamic acid pyridine-3-ylmethylester and its derivatives, butyric acid, pyroxamide, trichostatin A, oxamflatin, apicidin, depsipeptide, depudecin, trapoxin, vorinostat (Zolinza®), and compounds disclosed in WO 02/22577.

I-kappa B-alpha kinase inhibitors (IKK) which include without limitation 2-propenenitrile, 3-[(4-methylphenyl)sulfonyl]-(2E)-(9CI).

imidazotetrazinones which include without limitation temozolomide (Methazolastone®, Temodar® and its derivatives (e.g., as disclosed generically and specifically in U.S. Pat. No. 5,260,291) and Mitozolomide.

insulin tyrosine kinase inhibitors which include without limitation hydroxyl-2-naphthalenylmethylphosphonic acid.

c-Jun-N-terminal kinase (JNK) inhibitors which include without limitation pyrazoleanthrone and epigallocatechin gallate.

mitogen-activated protein kinase (MAP) inhibitors which include without limitation benzenesulfonamide, N-[2-[[[3-(4-chlorophenyl)-2-propenyl]methyl]amino]methyl]phenyl]-N-(2-hydroxyethyl)-4-methoxy-(9CI).

MDM2 inhibitors which include without limitation trans-4-iodo, 4'-boranyl-chalcone.

MEK inhibitors which include without limitation butanedinitrile, bis[amino[2-aminophenyl]thio]methylene]-(9CI).

MMP inhibitors which include without limitation Actinonin, epigallocatechin gallate, collagen peptidomimetic and non-peptidomimetic inhibitors, tetracycline derivatives marimastat (Marimastat®), prinomastat, incyclinide (Metastat®), shark cartilage extract AE-941 (Neovastat®), Tanomastat, TAA211, MMI270B or AAJ996.

mTor inhibitors which include without limitation rapamycin (Rapamune®), and analogs and derivatives thereof, AP23573 (also known as ridaforolimus, deforolimus, or MK-8669), CCI-779 (also known as temsirolimus) (Torisel®) and SDZ-RAD.

NGFR tyrosine kinase inhibitors which include without limitation tyrphostin AG 879.

p38 MAP kinase inhibitors which include without limitation Phenol, 4-[4-(4-fluorophenyl)-5-(4-pyridinyl)-1H-imidazol-2-yl]-(9CI), and benzamide, 3-(dimethylamino)-N-[3-[(4-hydroxybenzoyl)amino]-4-methylphenyl]-(9CI).

p56 tyrosine kinase inhibitors which include without limitation damnacanthol and tyrphostin 46.

PDGF pathway inhibitors which include without limitation tyrphostin AG 1296, tyrphostin 9, 1,3-butadiene-1,1,3-tricarbonitrile, 2-amino-4-(1H-indol-5-yl)-(9CI), imatinib (Gleevec®) and gefitinib (Iressa®) and those compounds generically and specifically disclosed in European Patent No.: 0 564 409 and PCT Publication No.: WO 99/03854.

phosphatidylinositol 3-kinase inhibitors which include without limitation wortmannin, and quercetin dihydrate.

phosphatase inhibitors which include without limitation cantharidic acid, cantharidin, and L-leucinamide.

protein phosphatase inhibitors which include without limitation cantharidic acid, cantharidin, L-P-bromotetramisole oxalate, 2(5H)-furanone, 4-hydroxy-5-(hydroxymethyl)-3-(1-oxohexadecyl)-(5R)-(9CI) and benzylphosphonic acid.

PKC inhibitors which include without limitation 1-H-pyrrolo-2,5-dione, 3-[1-3-(dimethylamino)propyl]-1H-indol-3-yl]-4-(1H-indol-3-yl)-(9CI), Bisindolylmaleimide IX, Sphingosine, staurosporine, and Hypericin.

PKC delta kinase inhibitors which include without limitation rottlerin.

polyamine synthesis inhibitors which include without limitation DMFO.

PTP1B inhibitors which include without limitation L-leucinamide.

protein tyrosine kinase inhibitors which include, without limitation tyrphostin Ag 216, tyrphostin Ag 1288, tyrphostin Ag 1295, geldanamycin, genistein and 7H-pyrrolo[2,3-d]pyrimidine derivatives as generically and specifically described in PCT Publication No.: WO 03/013541 and U.S. Publication No.: 2008/0139587.

SRC family tyrosine kinase inhibitors which include without limitation PP1 and PP2.

Syk tyrosine kinase inhibitors which include without limitation piceatannol.

Janus (JAK-2 and/or JAK-3) tyrosine kinase inhibitors which include without limitation tyrphostin AG 490 and 2-naphthyl vinyl ketone.

retinoids which include without limitation isotretinoin (Accutane®, Amnesteem®, Cistane®, Claravis®, Sotret®) and tretinoin (Aberel®, Aknoten®, Avita®, Renova®, Retin-A®, Retin-A MICRO®, Vesanoid®).

RNA polymerase II elongation inhibitors which include without limitation 5,6-dichloro-1-beta-D-ribofuranosylbenzimidazole.

serine/Threonine kinase inhibitors which include without limitation 2-aminopurine.

sterol biosynthesis inhibitors which include without limitation squalene epoxidase and CYP2D6.

VEGF pathway inhibitors, which include without limitation anti-VEGF antibodies, e.g., bevacizumab, and small molecules, e.g., sunitinib (Sutent®), sorafenib (Nexavar®), ZD6474 (also known as vandetanib) (Zactima™), SU6668, CP-547632 and AZD2171 (also known as cediranib) (Recentin™).

Examples of chemotherapeutic agents are also described in the scientific and patent literature, see, e.g., Bulinski (1997) J. Cell Sci. 110:3055-3064; Panda (1997) Proc. Natl. Acad. Sci. USA 94:10560-10564; Muhlradt (1997) Cancer Res. 57:3344-3346; Nicolaou (1997) Nature 387:268-272; Vasquez (1997) Mol. Biol. Cell. 8:973-985; Panda (1996) J. Biol. Chem. 271:29807-29812.

Other exemplary anti-cancer agents include alitretinon, altretamine, aminopterin, aminolevulinic acid, amsacrine (Amsidine®), asparaginase (crisantaspase, Erwinase®), atrasentan, bexarotene (Targretin®), carboquone, demecolcine, efaproxiral, elsamitrucin, etoglucid, ferrocene, Gliadel implants, hydroxycarbamide, leucovorin, lonidamine, lucanthone, masoprocol, methyl aminolevulinate, mitoguanzone, mitotane (Lysodren®), oblimersen, omacetaxine (Genasense®), pegaspargase (Oncaspar®), porfimer sodium (Photofrin®), prednimustine, sitimagene ceradenovec (Cerepro®), talaporfin, temoporfin, trabectedin (Yondelis®), and verteporfin.

It will be understood by the skilled artisan that when B comprises an anti-cancer agent, the anti-cancer agent may be attached to the linker group of the compound of formula (I) by any suitable linkage. For example, if the anti-cancer agent has a reactive moiety such as a hydroxyl group, an amine group, or the like, such groups may be used to attach the anti-cancer agent to the remainder of the compound of formula (I). Alternatively, an anti-cancer agent may be further derivatized with an atom or a group of atoms in order to provide a reactive group. In such instances, it is understood that the group —B corresponding to the anti-cancer agent includes not only the anti-cancer agent itself but also the linking atoms.

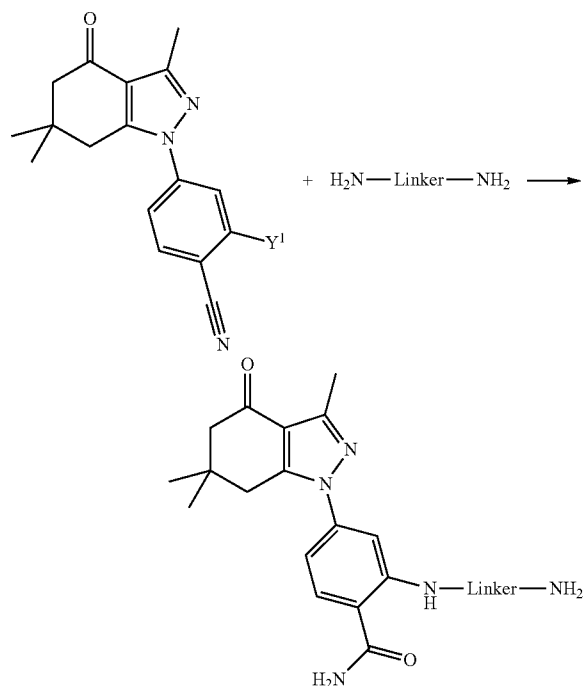
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f. Preparation of Compounds

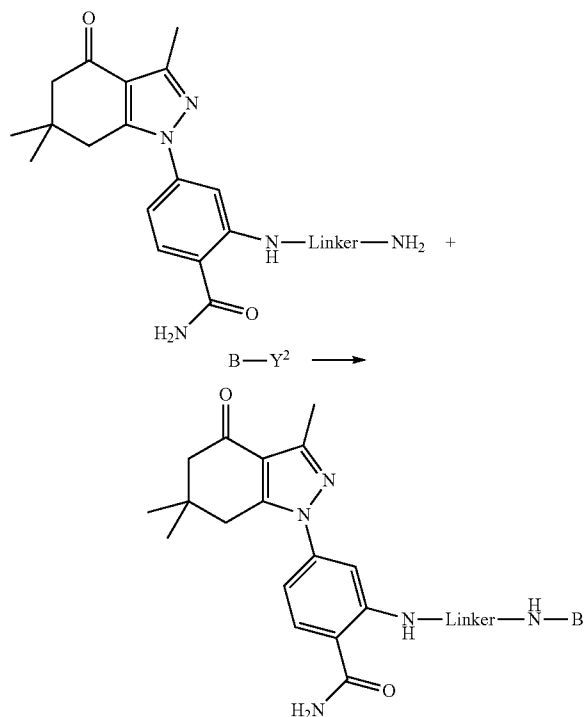
Compounds described herein may be prepared according to a variety of methods. A representative synthesis of exemplary compounds of formula (I) is illustrated in Scheme 1.

Scheme 1. Exemplary Synthesis

1)



2)



In reaction 1 of Scheme 1, the starting material includes an exemplary Hsp90-binding moiety precursor, and the

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group Y^1 is a leaving group or a reactive group. For example and as illustrated in Scheme 1, Y^1 may be leaving group such as a halogen, such that the compound $\text{H}_2\text{N}-\text{Linker}-\text{NH}_2$ may react with the compound via a reaction such as nucleophilic aromatic substitution. The nitrile can then be subsequently hydrolyzed to produce the Hsp90-binding moiety.

In reaction 2 of Scheme 1, the product of the initial reaction is further reacted with a compound $\text{B}-\text{Y}^2$, where B comprises a detection moiety, an anti-cancer agent or an Hsp90 binding component, and Y^2 is a leaving group or a reactive group. For example, Y^2 may be reactive group such as an isothiocyanate or a maleimide.

As can be appreciated by the skilled artisan, further methods of synthesizing the compounds of the formulae herein will be evident to those of ordinary skill in the art. Additionally, the various synthetic steps may be performed in an alternate sequence or order to give the desired compounds. Synthetic chemistry transformations and protecting group methodologies (protection and deprotection) useful in synthesizing the compounds described herein are known in the art and include, for example, those such as described in R. Larock, *Comprehensive Organic Transformations*, VCH Publishers (1989); T. W. Greene and P. G. M. Wuts, *Protective Groups in Organic Synthesis*, 2d. Ed., John Wiley and Sons (1991); L. Fieser and M. Fieser, *Fieser and Fieser's Reagents for Organic Synthesis*, John Wiley and Sons (1994); and L. Paquette, ed., *Encyclopedia of Reagents or Organic Synthesis*, John Wiley and Sons (1995), and subsequent editions thereof.

g. Evaluation of Compounds

Compounds can be evaluated by determining their ability to bind to Hsp90. The selectivity for Hsp90 over other proteins, such as GRP94 and TRAP1, may also be determined

For example, prior to attachment of a detection moiety, an A-Linker- NH_2 compound can be bound to Sepharose resin. Such a resin can be exposed to a pig mammary gland extract, which is a tissue known to be high in ATP binding proteins including native forms of Hsp90, GRP94 and TRAP1. Media can be incubated with extract, washed stringently, and bound proteins removed with SDS and subsequently characterized. Preincubation of the resin with a known Hsp90 inhibitor should selectively block binding of Hsp90 and associated proteins, while those recovered in the presence of such a quenching agent are likely to be non-specifically bound.

Compounds of formula (I) may have increased selectivity for Hsp90 compared to proteins that share sequence homology with Hsp90. For example, the compounds may bind to Hsp90 with an affinity that is at least about 100 to 1000 fold higher than the affinity for GRP94, a molecular chaperone that functions in the processing and transport of secreted proteins. The Hsp90 affinity probe may bind to Hsp90 with an affinity that is at least about 100 fold to 1000-fold higher than the affinity for TRAP1, (TNF receptor-associated protein 1), a heat shock protein of 75 kDa. This increased selectivity may lead to more accurate detection and targeting of Hsp90.

3. METHODS OF USE

Compounds of formula (I) may be used in a variety of methods, such as methods of detecting Hsp90 in a sample, methods of detecting cancer in a subject, and methods of treating cancer in a subject.

a. Methods of Detecting Hsp90

In some embodiments, compounds described herein can be used in methods of detecting Hsp90 in a sample, the methods comprising:

a) contacting the sample with a compound having the following formula (Ia):



wherein:

A is an Hsp90 binding component;

X^1 is selected from the group consisting of $-NH-$, $-O-$, $-S-$, $-C(O)-$ and $-S(O)_2-$;

X^2 is selected from the group consisting of $-NR-$, $-O-$, $-S-$, $-C(O)-$ and $-S(O)_2-$;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety; and

b) detecting a signal from the detection moiety.

In some embodiments, the sample is an in vitro sample, such as a cell or tissue extract. In some embodiments, the sample is a cell culture. For example, the sample may be a culture of cells such as cancer cells.

In some embodiments, the sample is a biological sample from a subject, such as a human. In some embodiments, the biological sample is selected from the group consisting of a tissue sample, bodily fluid, whole blood, plasma, serum, urine, bronchoalveolar lavage fluid, and a cell culture suspension or fraction thereof. In embodiments in which Hsp90 is detected in a biological sample from a subject, the methods may further involve providing or obtaining a biological sample from the subject, which can be obtained by any known means including needle stick, needle biopsy, swab, and the like. In an embodiment of such methods, the biological sample is a blood sample, such as a blood plasma or serum sample, which may be obtained by any standard technique such as, for example, by venipuncture. Biological samples used in the methods may be stored or banked under suitable tissue storage conditions, or can be accessed from samples that have been previously stored or banked under suitable conditions.

Following contacting the sample with a compound, the method further includes detecting a signal from the detection moiety. A signal may be detected by any suitable means appropriate for the particular detection moiety being used. For example, when the detection moiety comprises a fluorophore, a signal may be detected using a fluorometer or a fluorescence plate reader, or by using fluorescence techniques such as fluorescence microscopy, fluorescence resonance energy transfer, flow cytometry and fluorescence-activated cell sorting. In embodiments in which the detection moiety comprises a radioisotope, a signal may be detected using scintillation counting or radioimaging techniques.

A signal from the detection moiety may be quantitated, for example, by comparing the quantity of the signal to that of a reference sample.

b. Methods of Detecting Cancer

In some embodiments, compounds described herein can be used in a method of detecting cancer in a subject, the method comprising:

a) contacting a biological sample from the subject with a compound having the following formula (Ia):



wherein:

A is an Hsp90 binding component;

X^1 is selected from the group consisting of $-NH-$, $-O-$, $-S-$, $-C(O)-$ and $-S(O)_2-$;

X^2 is selected from the group consisting of $-NR-$, $-O-$, $-S-$, $-C(O)-$ and $-S(O)_2-$;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety; and

b) detecting a signal from the detection moiety;

wherein cancer is detected in the sample when the signal from the detection moiety is higher relative to a signal from a reference sample.

In some embodiments, the method further comprises obtaining the biological sample from the subject, such as a biological sample described herein, according to methods described herein. In some embodiments, the subject is a human.

A reference sample may be a sample from a healthy subject, i.e. a subject having no clinical signs or symptoms of cancer. Suitably, the healthy subject may be clinically evaluated for otherwise undetected signs or symptoms of cancer, which evaluation may include routine physical examination and/or laboratory testing.

In embodiments, the cancer may be any type of cancer, such as a cancer recognized by the National Cancer Institute. In embodiments, the cancer may be a type of cancer associated with elevated levels of Hsp90. Exemplary types of cancers include the following:

Digestive/gastrointestinal cancers such as anal cancer; bile duct cancer; extrahepatic bile duct cancer; appendix cancer; carcinoid tumor, gastrointestinal cancer; colon cancer; colorectal cancer including childhood colorectal cancer; esophageal cancer including childhood esophageal cancer; gallbladder cancer; gastric (stomach) cancer including childhood gastric (stomach) cancer; hepatocellular (liver) cancer including adult (primary) hepatocellular (liver) cancer and childhood (primary) hepatocellular (liver) cancer; pancreatic cancer including childhood pancreatic cancer; sarcoma, rhabdomyosarcoma; islet cell pancreatic cancer; rectal cancer; and small intestine cancer;

Breast cancer, including childhood breast cancer, male breast cancer and breast cancer during pregnancy;

Genitourinary cancers such as bladder cancer including childhood bladder cancer; renal cell (kidney) cancer; ovarian cancer including childhood ovarian cancer; ovarian epithelial cancer; ovarian low malignant potential tumor; penile cancer; prostate cancer; renal cell cancer including childhood renal cell cancer; renal pelvis and ureter, transitional cell cancer; testicular cancer; urethral cancer; vaginal cancer; vulvar cancer; cervical cancer; Wilms tumor and other childhood kidney tumors; endometrial cancer; and gestational trophoblastic tumor;

Lung cancer such as non-small cell lung cancer; and small cell lung cancer;

Respiratory cancers such as malignant mesothelioma, adult; malignant mesothelioma, childhood; malignant thymoma; childhood thymoma; thymic carcinoma; bronchial adenomas/carcinoids including childhood bronchial adenomas/carcinoids; pleuropulmonary blastoma; non-small cell lung cancer; and small cell lung cancer; and

Skin cancers such as Kaposi's sarcoma; Merkel cell carcinoma; melanoma; and childhood skin cancer.

In suitable embodiments, the cancer may be a cancer that is associated with increased levels of Hsp90, including but not limited to breast cancer, prostate cancer, non-small cell lung cancer, colorectal cancer, pancreatic cancer, ovarian cancer and melanoma. In particular embodiments, the cancer is breast cancer or prostate cancer.

c. Methods of Treating Cancer

In some embodiments, compounds described herein can be used in a method of treating cancer in a subject in need of treatment. Such methods comprise administering the subject a therapeutically effective amount of a compound having the following formula (Ib):



wherein:

A is an Hsp90 binding component;

X^1 and X^2 are each independently selected from the group consisting of $-NH-$, $-O-$, $-S-$, $-C(O)-$ and $-S(O)_2-$;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen; and

B is selected from the group consisting of an anti-cancer agent and an Hsp90 binding component.

In embodiments, the cancer is a cancer described herein. In embodiments, the cancer is selected from the group consisting of breast cancer, prostate cancer, non-small cell lung cancer, colorectal cancer, pancreatic cancer, ovarian cancer and melanoma. In particular embodiments, the cancer is breast cancer or prostate cancer.

In embodiments, the anti-cancer agent is an anti-cancer agent described herein. In embodiments, B is an anti-cancer agent selected from the group consisting of a protein kinase inhibitor, a protein phosphatase inhibitor and a histone deacetylase inhibitor.

In the methods of treating cancer, a compound, or a pharmaceutical composition comprising the compound, may be administered to the subject by any convenient route of administration, whether systemically/peripherally or at the site of desired action, including but not limited to, oral (e.g. by ingestion); topical (including e.g. transdermal, intranasal, ocular, buccal, and sublingual); pulmonary (e.g. by inhalation or insufflation therapy using, e.g. an aerosol, e.g. through mouth or nose); rectal; vaginal; parenteral, for example, by injection, including subcutaneous, intradermal, intramuscular, intravenous, intraarterial, intracardiac, intrathecal, intraspinal, intracapsular, subcapsular, intraorbital, intraperitoneal, intratracheal, subcuticular, intraarticular, subarachnoid, and intrasternal; by implant of a depot, for example, subcutaneously or intramuscularly. Additional modes of administration may include adding the compound and/or a composition comprising the compound to a food or beverage, including a water supply for an animal, to supply the compound as part of the animal's diet.

While it is possible for the compound to be administered alone, in some embodiments the compound may be presented as a pharmaceutical composition (e.g., formulation) comprising at least one compound, as defined above, together with one or more pharmaceutically-acceptable carriers, adjuvants, excipients, diluents, fillers, buffers, stabilizers, preservatives, lubricants, or other materials well known to those skilled in the art and optionally other therapeutic or prophylactic agents.

Thus, the disclosure further provides pharmaceutical compositions, as defined above, and methods of making a pharmaceutical composition comprising admixing at least one compound, as defined above, together with one or more pharmaceutically acceptable carriers, excipients, buffers, adjuvants, stabilizers, or other materials, as described herein.

Suitable carriers, excipients, etc. can be found in standard pharmaceutical texts, for example Remington's Pharmaceutical Sciences, 18th edition, Mack Publishing Company, Easton, Pa., 1990.

The formulations may conveniently be presented in unit dosage form and may be prepared by any method well known in the art of pharmacy. Such methods include the step of bringing into association the compound with the carrier which constitutes one or more accessory ingredients. In general, the formulations may be prepared by uniformly and intimately bringing into association the compound with liquid carriers or finely divided solid carriers or both, and then if necessary shaping the product.

Formulations may be in the form of liquids, solutions, suspensions, emulsions, elixirs, syrups, tablets, lozenges, granules, powders, capsules, cachets, pills, ampoules, suppositories, pessaries, ointments, gels, pastes, creams, sprays, mists, foams, lotions, oils, boluses, electuaries, or aerosols.

Formulations suitable for oral administration (e.g., by ingestion) may be presented as discrete units such as capsules, cachets or tablets, each containing a predetermined amount of the compound; as a powder or granules; as a solution or suspension in an aqueous or non-aqueous liquid; or as an oil-in-water liquid emulsion or a water-in-oil liquid emulsion; as a bolus; as an electuary; or as a paste.

A tablet may be made by conventional means, e.g., compression or molding, optionally with one or more accessory ingredients. Compressed tablets may be prepared by compressing in a suitable machine the compound in a free-flowing form such as a powder or granules, optionally mixed with one or more binders (e.g., povidone, gelatin, acacia, sorbitol, tragacanth, hydroxypropylmethyl cellulose); fillers or diluents (e.g., lactose, microcrystalline cellulose, calcium hydrogen phosphate); lubricants (e.g., magnesium stearate, talc, silica); disintegrants (e.g., sodium starch glycolate, cross-linked povidone, cross-linked sodium carboxymethyl cellulose); surface-active or dispersing or wetting agents (e.g., sodium lauryl sulfate); and preservatives (e.g., methyl p-hydroxybenzoate, propyl p-hydroxybenzoate, sorbic acid). Molded tablets may be made by molding in a suitable machine a mixture of the powdered compound moistened with an inert liquid diluent. The tablets may optionally be coated or scored and may be formulated so as to provide slow or controlled release of the compound therein using, for example, hydroxypropylmethyl cellulose in varying proportions to provide the desired release profile. Tablets may optionally be provided with an enteric coating, to provide release in parts of the gut other than the stomach.

Formulations suitable for topical administration (e.g., transdermal, intranasal, ocular, buccal, and sublingual) may be formulated as an ointment, cream, suspension, lotion, powder, solution, past, gel, spray, aerosol, or oil. Alternatively, a formulation may comprise a patch or a dressing such as a bandage or adhesive plaster impregnated with compounds and optionally one or more excipients or diluents. In addition, a formulation may be added to a conventional bandage, e.g. to a gauze portion that contacts a wound, as an antimicrobial agent.

Formulations suitable for topical administration in the mouth include lozenges comprising the compound in a flavored basis, usually sucrose and acacia or tragacanth; pastilles comprising the compound in an inert basis such as gelatin and glycerin, or sucrose and acacia; and mouthwashes comprising the compound in a suitable liquid carrier.

Formulations suitable for topical administration to the eye also include eye drops wherein the compound is dissolved or suspended in a suitable carrier, especially an aqueous solvent for the compound.

Formulations suitable for nasal administration, wherein the carrier is a solid, include a coarse powder having a

particle size, for example, in the range of about 20 to about 500 microns which is administered in the manner in which snuff is taken, i.e., by rapid inhalation through the nasal passage from a container of the powder held close up to the nose. Suitable formulations wherein the carrier is a liquid for administration as, for example, nasal spray, nasal drops, or by aerosol administration by nebulizer, include aqueous or oily solutions of the compound.

Formulations suitable for administration by inhalation include those presented as an aerosol spray from a pressurized pack, with the use of a suitable propellant, such as dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide, or other suitable gases.

Formulations suitable for topical administration via the skin include ointments, creams, and emulsions. When formulated in an ointment, the compound may optionally be employed with either a paraffinic or a water-miscible ointment base. Alternatively, the compounds may be formulated in a cream with an oil-in-water cream base. If desired, the aqueous phase of the cream base may include, for example, at least about 30% w/w of a polyhydric alcohol, i.e., an alcohol having two or more hydroxyl groups such as propylene glycol, butane-1,3-diol, mannitol, sorbitol, glycerol and polyethylene glycol and mixtures thereof. The topical formulations may desirably include a compound which enhances absorption or penetration of the compound through the skin or other affected areas. Examples of such dermal penetration enhancers include dimethylsulfoxide and related analogues.

When formulated as a topical emulsion, the oily phase may optionally comprise merely an emulsifier (otherwise known as an emulgent), or it may comprise a mixture of at least one emulsifier with a fat or an oil or with both a fat and an oil. Preferably, a hydrophilic emulsifier is included together with a lipophilic emulsifier which acts as a stabilizer. It is also preferred to include both an oil and a fat. Together, the emulsifier(s) with or without stabilizer(s) make up the so-called emulsifying wax, and the wax together with the oil and/or fat make up the so-called emulsifying ointment base which forms the oily dispersed phase of the cream formulations.

Suitable emulgents and emulsion stabilizers include Tween 60, Span 80, cetostearyl alcohol, myristyl alcohol, glyceryl monostearate and sodium lauryl sulfate. The choice of suitable oils or fats for the formulation is based on achieving the desired cosmetic properties, since the solubility of the compound in most oils likely to be used in pharmaceutical emulsion formulations may be very low. Thus the cream should preferably be a non-greasy, non-staining and washable product with suitable consistency to avoid leakage from tubes or other containers. Straight or branched chain, mono- or dibasic alkyl esters such as diisoadipate, isocetyl stearate, propylene glycol diester of coconut fatty acids, isopropyl myristate, decyl oleate, isopropyl palmitate, butyl stearate, 2-ethylhexyl palmitate or a blend of branched chain esters known as Crodamol CAP may be used, the last three being preferred esters. These may be used alone or in combination depending on the properties required. Alternatively, high melting point lipids such as white soft paraffin and/or liquid paraffin or other mineral oils can be used.

Formulations suitable for rectal administration may be presented as a suppository with a suitable base comprising, for example, cocoa butter or a salicylate.

Formulations suitable for vaginal administration may be presented as pessaries, tampons, creams, gels, pastes, foams

or spray formulations containing in addition to the compound, such carriers as are known in the art to be appropriate.

Formulations suitable for parenteral administration (e.g., by injection, including cutaneous, subcutaneous, intramuscular, intravenous and intradermal), include aqueous and non-aqueous isotonic, pyrogen-free, sterile injection solutions which may contain anti-oxidants, buffers, preservatives, stabilizers, bacteriostats in addition to the compound, and solutes which render the formulation isotonic with the blood of the intended recipient; and aqueous and non-aqueous sterile suspensions which may include suspending agents and thickening agents, and liposomes or other microparticulate systems which are designed to target the compound to blood components or one or more organs. Examples of suitable isotonic vehicles for use in such formulations include Sodium Chloride Injection, Ringer's Solution, or Lactated Ringer's Injection. Typically, the concentration of the compound in the solution is from about 1 ng/ml to about 1 µg/ml, although other concentrations are possible and are encompassed within the invention. The formulations may be presented in unit-dose or multi-dose sealed containers, for example, ampoules and vials, and may be stored in a freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid carrier, for example water for injections, immediately prior to use. Extemporaneous injection solutions and suspensions may be prepared from sterile powders, granules, and tablets. Formulations may be in the form of liposomes or other microparticulate systems which are designed to target the compound to blood components or one or more organs.

It will be appreciated that appropriate dosages of the compounds, and compositions comprising the compounds, can vary from patient to patient. Determining the optimal dosage will generally involve the balancing of the level of therapeutic benefit against any risk or deleterious side effects of the treatments of the present invention. The selected dosage level will depend on a variety of factors including, but not limited to, the activity of the particular compound, the route of administration, the time of administration, the rate of excretion of the compound, the duration of the treatment, other drugs, compounds, and/or materials used in combination, and the age, sex, weight, condition, general health, and prior medical history of the patient. The amount of compound and route of administration will ultimately be at the discretion of the physician, although generally the dosage will be to achieve local concentrations at the site of action which achieve the desired effect without causing substantial harmful or deleterious side-effects.

Administration in vivo can be effected in one dose, continuously or intermittently (e.g., in divided doses at appropriate intervals) throughout the course of treatment. Methods of determining the most effective means and dosage of administration are well known to those of skill in the art and will vary with the formulation used for therapy, the purpose of the therapy, the target cell being treated, and the subject being treated. Single or multiple administrations can be carried out with the dose level and pattern being selected by the treating physician. In general, a suitable dose of the compound is in the range of about 100 µg to about 250 mg per kilogram body weight of the subject per day.

The composition may be administered once, on a continuous basis (e.g. by an intravenous drip), or on a periodic/intermittent basis, including about once per hour, about once per two hours, about once per four hours, about once per eight hours, about once per twelve hours, about once per day, about once per two days, about once per three days,

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about twice per week, about once per week, and about once per month. The composition may be administered until a desired reduction of symptoms is achieved.

The present compounds, compositions, and methods may be administered as part of a therapeutic regimen along with other treatments appropriate for the particular injury or disease being treated.

d. Methods of Detecting HIV

In some embodiments, compounds described herein can be used in a method of detecting Human Immunodeficiency Virus (HIV) in a subject, the method comprising:

a) contacting a biological sample from the subject with a compound having the following formula (Ia):



wherein:

A is an Hsp90 binding component;

X^1 is selected from the group consisting of $-NH-$, $-O-$, $-S-$, $-C(O)-$ and $-S(O)_2-$;

X^2 is selected from the group consisting of $-NR-$, $-O-$, $-S-$, $-C(O)-$ and $-S(O)_2-$;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety; and

b) detecting a signal from the detection moiety; wherein HIV is detected in the sample when the signal from the detection moiety is higher relative to a signal from a reference sample.

4. KITS

In another aspect, the disclosure provides a kit, which may be used for detecting Hsp90 in a sample, for detecting cancer in a sample, or for treating cancer in a subject.

A kit will include a compound of formula (I) as described herein. A kit may also include instructions for use of the compound of formula (I). Instructions included in kits can be affixed to packaging material or can be included as a package insert. While the instructions are typically written or printed materials they are not limited to such. Any medium capable of storing such instructions and communicating them to an end user is contemplated by this disclosure. Such media include, but are not limited to, electronic storage media (e.g., magnetic discs, tapes, cartridges, chips), optical media (e.g., CD, DVD), and the like. As used herein, the term "instructions" can include the address of an internet site that provides the instructions.

In one embodiment, the disclosure provides a kit for detecting Hsp90 in a sample. The kit comprises at least one compound of formula (I), and instructions for assaying the test sample for Hsp90. For example, the kit can comprise instructions for assaying the test sample for Hsp90 by fluorescence detection. The kit may further comprise a calibrator or control, e.g., purified, and optionally lyophilized, (e.g., Hsp90), and/or at least one container (e.g., tube, microtiter plates or strips) for conducting the assay, and/or a buffer, such as an assay buffer or a wash buffer, either one of which can be provided as a concentrated solution. Preferably, the kit comprises all components, i.e., reagents, standards, buffers, diluents, etc., which are necessary to perform the assay. The instructions also can include instructions for generating a standard curve or a reference standard for purposes of quantifying Hsp90.

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The kit can also optionally include other reagents required to conduct a diagnostic assay or facilitate quality control evaluations, such as buffers, salts, enzymes, enzyme cofactors, substrates, detection reagents, and the like. Other components, such as buffers and solutions for the isolation and/or treatment of a test sample (e.g., pretreatment reagents), also can be included in the kit. The kit can additionally include one or more other controls. One or more of the components of the kit can be lyophilized, in which case the kit can further comprise reagents suitable for the reconstitution of the lyophilized components.

The various components of the kit optionally are provided in suitable containers as necessary, e.g., a microtiter plate. The kit can further include containers for holding or storing a sample (e.g., a container or cartridge for a blood sample). Where appropriate, the kit optionally also can contain reaction vessels, mixing vessels, and other components that facilitate the preparation of reagents or the test sample. The kit can also include one or more instrument for assisting with obtaining a test sample, such as a syringe, pipette, forceps, measured spoon, or the like.

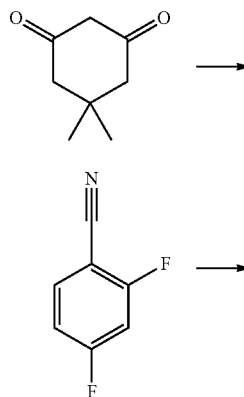
It will be readily apparent to those skilled in the art that other suitable modifications and adaptations of the compounds and methods of the present disclosure described herein are readily applicable and appreciable, and may be made using suitable equivalents without departing from the scope of the present disclosure or the aspects and embodiments disclosed herein. Having now described the present disclosure in detail, the same will be more clearly understood by reference to the following examples which are merely intended only to illustrate some aspects and embodiments of the disclosure, and should not be viewed as limiting to the scope of the disclosure. The disclosures of all journal references, U.S. patents and publications referred to herein are hereby incorporated by reference in their entireties.

EXAMPLES

General Experimental and Analytical Details

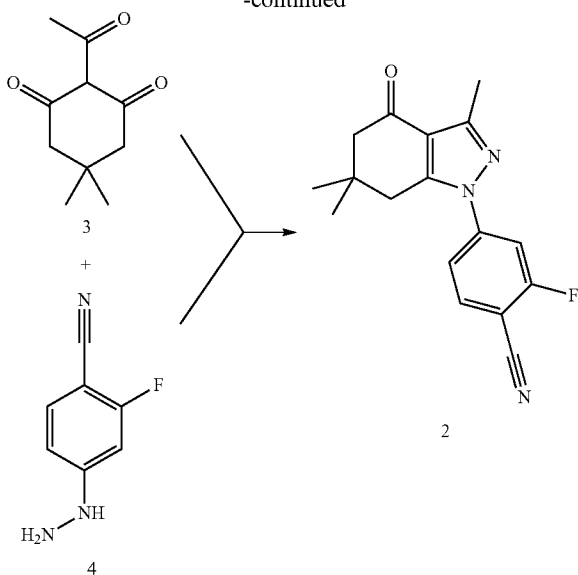
Reagents were obtained from commercial sources and used without further purification. 1,19-Diamino-4,7,10,13,16-pentaoxanonadecane was obtained from Berry and Associates. Proton NMR spectra were obtained on Varian 400 and 500 MHz spectrometers. LC/MS were obtained on an Agilent ion-trap LC/MS system. HRMS results were obtained on an Agilent 6224 LCMS-TOF and are reported as an average of four runs.

Example 1. Synthesis of 2-Fluoro-4-(3,6,6-trimethyl-1-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzonitrile (2)

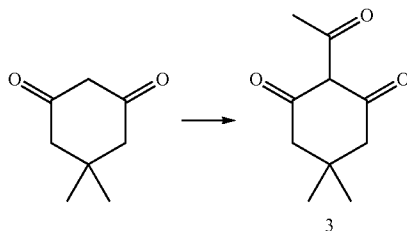


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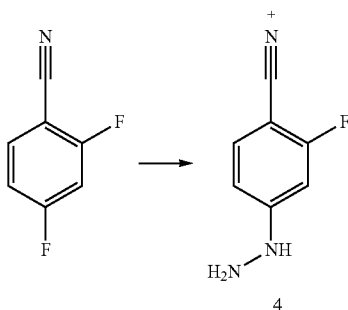


2-Acetyl-5,5-dimethylcyclohexane-1,3-dione (3)



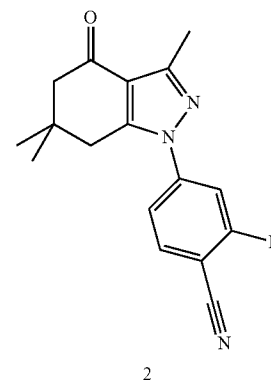
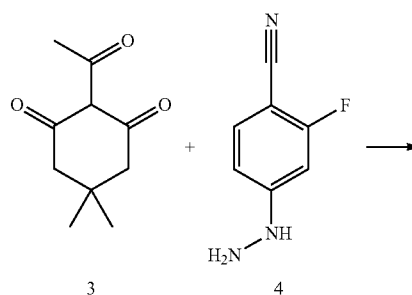
Dimedone (10 g, 71.3 mmol) was dissolved in methylene chloride (200 mL) and treated with Hunig's base (9.7 g, 74.9 mmol) and DMAP (440 mg, 3.6 mmol) followed by slow addition of acetic anhydride (7.65 g, 74.9 mmol). After 24 h, the mixture was concentrated, partitioned between hexanes (150 mL) and 1 N HCl (70 mL), then washed with brine (50 mL), treated with Norit A and then dried (MgSO₄), filtered and concentrated to give 2-acetyl-5,5-dimethylcyclohexane-1,3-dione 3 (~11 g) as a yellow oil. The entire product was used in the next step.

2-Fluoro-4-hydrazinylbenzonitrile (4)

**30**

2,4-difluorobenzonitrile (10 g, 72 mmol) was dissolved in methanol (100 mL) and treated drop wise with hydrazine hydrate (18 g, 0.36 mol) and stirred at RT. After 16 h, the reaction mixture, containing a mixture of isomers, was concentrated then partitioned between ethyl acetate (100 mL), water (70 mL) and 1 N NaOH (30 mL). The organic layer was washed with brine (40 mL) then concentrated to give 2,4-difluorobenzonitrile (4) in a mixture of isomers as a white solid.

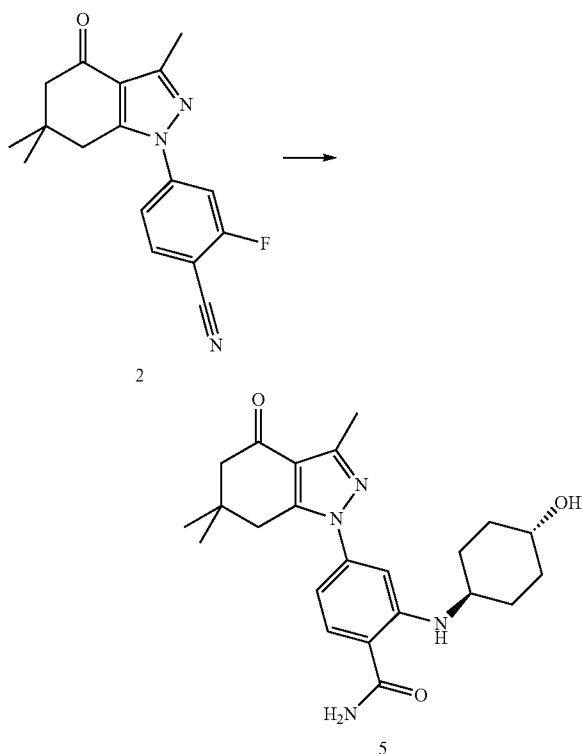
2-Fluoro-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzonitrile (2)



All of 3 (assume 13 g, 71.8 mmol) and 4 (assume 10.9 g, 71.8 mmol) were combined, then dissolved in methanol (40 mL) and treated with acetic acid (1 mL) and stirred at RT for 3 days. The mixture was concentrated and then dissolved in methylene chloride (20 mL). The mixture was chromatographed in a number of batches, with the best approach to load a DCE solution onto a dry column, wait a bit (maybe 15 to 20 min) then elute with 10% ethyl acetate in hexanes to remove the yellow (isomeric product) followed by elution with 20% to get the product. The chromatographed products were recrystallized from ethyl acetate/hexanes to give 2 (10.4 g, 49%). TLC (hexane/EtOAc: 60/40) R_f=0.47; ¹H NMR (CDCl₃) δ 7.73 (dd, J=7, 8.5 Hz, 1H), 7.49 (dd, J=2, 9.8 Hz, 1H), 7.45 (dd, J=2, 8.5 Hz, 1H), 2.90 (s, 2H) 2.54 (s, 3H) 2.43 (s, 2H), 1.14 (s, 6H); MS (ESI): m/z 298.2 [M+H]⁺.

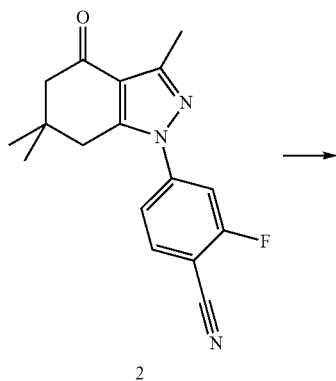
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Example 2. Synthesis of 2-(((1r,4r)-4-Hydroxycyclohexyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (5)



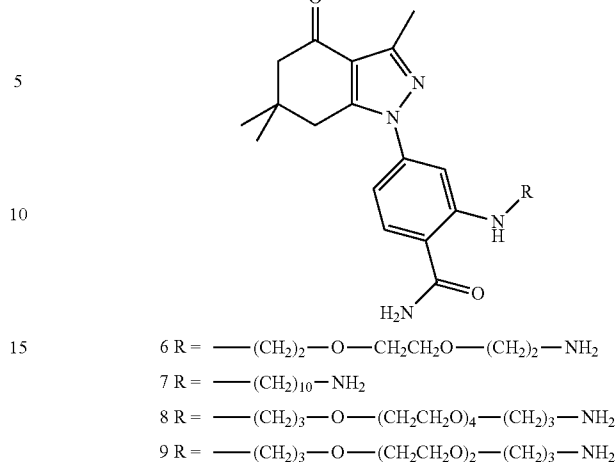
A mixture of nitrile 2 (200 mg, 0.67 mmol) and trans-4-aminocyclohexanol (232 g, 2.0 mmol), Hunig's base (117 μ L) and DMSO (300 μ L) were heated to 90° C. for 30 m. The mixture diluted with ethanol (2 mL) and treated with 50% NaOH (5 drops) and then, very slowly, a drop at a time, with hydrogen peroxide. After each drop, the reaction foamed up a bit. After adding 5 drops over 10 m, the mixture was diluted with water (18 mL) and allowed to cool slowly with rapid stirring. After stirring overnight, the solid was filtered off to give the product 5 (251 mg, 91%) as a white powder. TLC (EtOAc) R_f =0.25; ^1H NMR (CDCl_3) δ 8.07 (d, J =7.2 Hz), 7.44 (d, J =8.6 Hz, 1H), 6.75 (d, J =2 Hz, 1H), 6.60 (dd, J =2, 8.6 Hz, 1H), 5.6 (br s, 2H), 3.71 (m, 1H), 3.35 (m, 1H), 2.80 (s, 2H) 2.53 (s, 3H) 2.38 (s, 2H), 2.13 (m, 2H), 2.02 (m, 2H), 1.64 (br s, 1H), 1.41 (m, 4H), 1.09 (s, 6H); MS (ESI): m/z 411.3 $[\text{M}+\text{H}]^+$.

Example 3. Attachment of Linkers

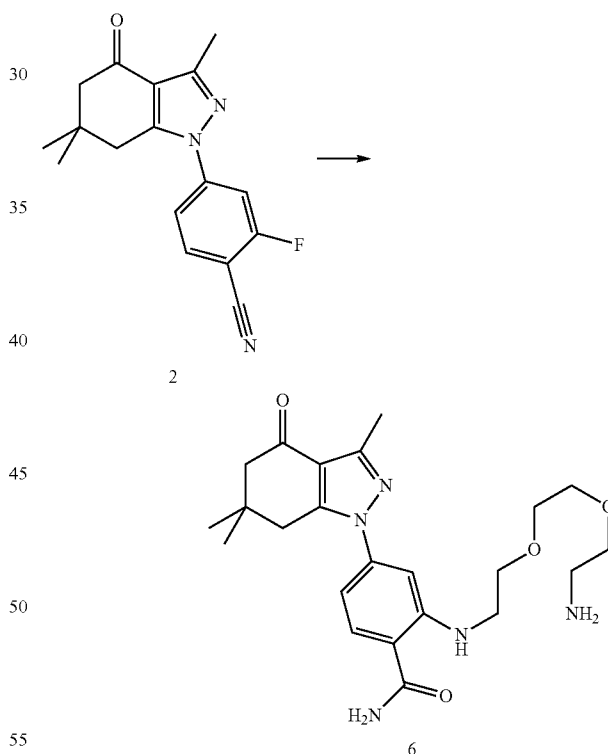


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2-((2-(2-(2-aminoethoxy)ethoxy)ethyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (6)

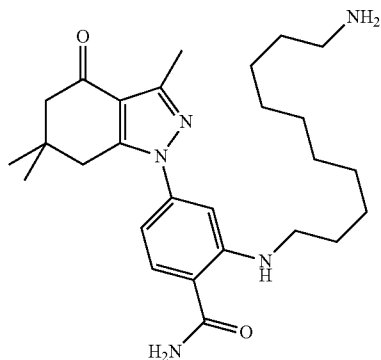


A mixture of 2 (500 mg, 1.7 mmol) and 2,2'-(ethylene-dioxy)bis(ethylamine) (1.2 g, 8.4 mmol) in DMSO (1 mL) were and heated to 90° C. for 30 minutes. The mixture was diluted with ethanol (2 mL) and, still at 90° C., treated with 50% NaOH (20 drops) and very slowly with 30% hydrogen peroxide (40 drops). The reaction mixture was diluted with methylene chloride and methanol and adsorbed onto silica gel. The mixture was chromatographed (silica gel, 1.5 cm \times 20 cm) and eluted with 9/1 $\text{CH}_2\text{Cl}_2/\text{MeOH}$, followed by 9/1/0.1 $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$ to give 6 (670 mg, 89%) as a

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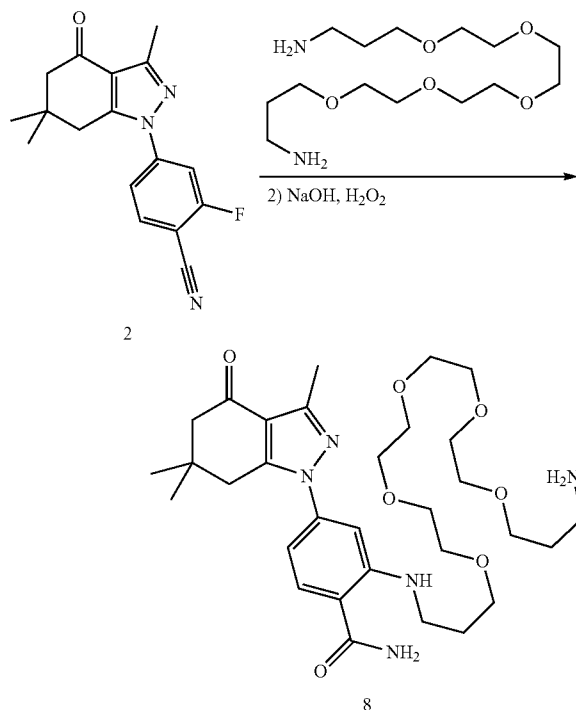
clear glass. TLC (4/1/0.1 CH₂Cl₂/MeOH/NH₃) R_f=0.14; ¹H NMR (CDCl₃) δ 8.17 (t, J=5 Hz), 7.47 (d, J=8.4 Hz, 1H), 6.77 (d, J=2 Hz, 1H), 6.60 (dd, J=2, 8.4 Hz, 1H), 6.1 (br s, 2H), 3.73 (t, 3H), 3.65 (m, 4H), 3.50 (t, 2H), 3.36 (m, 2H), 2.85 (t, 2H), 2.77 (s, 2H) 2.51 (s, 3H) 2.36 (s, 2H), 1.07 (s, 6H); HRMS (ESI) [M+H]⁺ calcd for C₂₃H₃₄N₅O₄, 444.2605; found 444.2607.

2-((10-aminodecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (7)



Compound 7 was made in an analogous manner to 6. HRMS (ESI) [M+H]⁺ calcd for C₂₂H₄₂N₅O₂, 468.3333; found 468.3342.

2-((19-Amino-4,7,10,13,16-pentaoxononadecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (8)

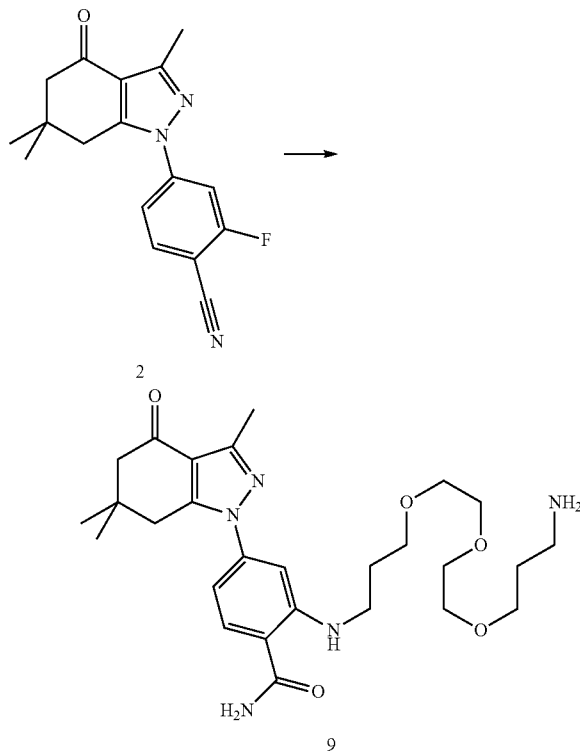


A mixture of 2 (482 mg, 1.62 mmol) and 1,19-diamino-4,7,10,13,16-pentaoxononadecane (1 g, 3.24 mol), diisopropylethylamine (628 mg, 4.8 mmol) and DMSO (1 mL) were heated to 90° C. for 20 m. Still at 90° C., the mixture diluted

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with ethanol (2 mL) and treated with 50% NaOH (10 drops) and then, very slowly, a drop at a time, with hydrogen peroxide. After each drop, the reaction foamed up substantially. After about 10 drops over 10 m, the reaction mixture was diluted with ethanol and added to silica gel (6 g) and left overnight. The next day, the slurry was concentrated to a powder, added to a silica gel column (2.5×20 cm) and chromatographed with CH₂Cl₂ (300 mL), CH₂Cl₂/MeOH/NH₃ 19/0.9/0.1 (300 mL), 9/0.9/0.1 (300 mL) and 4/0.9/0.1 (500 mL). Fractions containing two by-products were set aside (see below). The cleanest fractions were combined to give 8 (600 mg, 61%) as a lightly yellow glass. TLC (4/1/0.1 CH₂Cl₂/MeOH/NH₃) R_f=0.30; ¹H NMR (CDCl₃) δ 7.98 (t, J=4 Hz), 7.47 (d, J=8.4 Hz, 1H), 6.77 (d, J=2 Hz, 1H), 6.60 (dd, J=2, 8.4 Hz, 1H), 6.0 (br s, 2H), 3.61 (m, 16H), 3.28 (m, 2H), 2.85 (t, 2H), 2.79 (s, 2H), 2.52 (s, 3H) 2.37 (s, 2H), 2.28 (br s, 2H), 1.94 (m, 2H), 1.76 (m, 2H), 1.07 (s, 6H); HRMS (ESI) [M+H]⁺ calcd for C₃₁H₅₀N₅O₇, 604.3705; found 604.3715.

2-((3-(2-(2-(3-aminopropoxy)ethoxy)ethoxy)propyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (9)

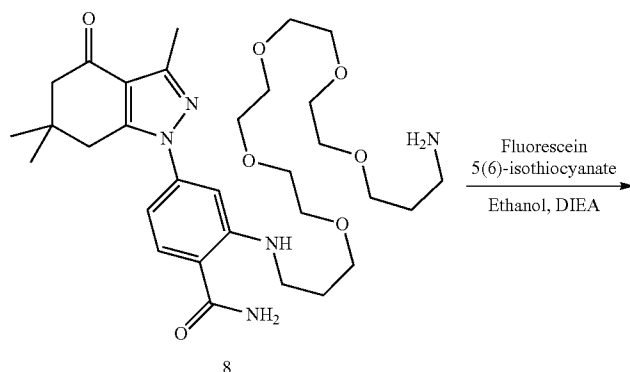


Compound 9 was prepared in the same way as compound 6. TLC (4/1/0.1 CH₂Cl₂/MeOH/NH₃) R_f=0.39; ¹H NMR (CDCl₃) δ 8.00 (t, J=5 Hz, 1H), 7.46 (d, J=8.2 Hz, 1H), 6.77 (s, 1H), 6.60 (d, J=8.4 Hz, 1H), 6.1 (br s, 2H), 3.59 (m, 8H), 3.29 (m, 2H), 2.82 (t, 2H), 2.79 (s, 2H) 2.59 (br s, 2H), 2.51 (s, 3H), 2.37 (s, 2h), 1.93 (M, 2h), 1.73 (m, 2H), 1.07 (s, 6H); HRMS (ESI) [M+H]⁺ calcd for C₂₇H₄₂N₅O₅, 516.3180; found 516.3191.

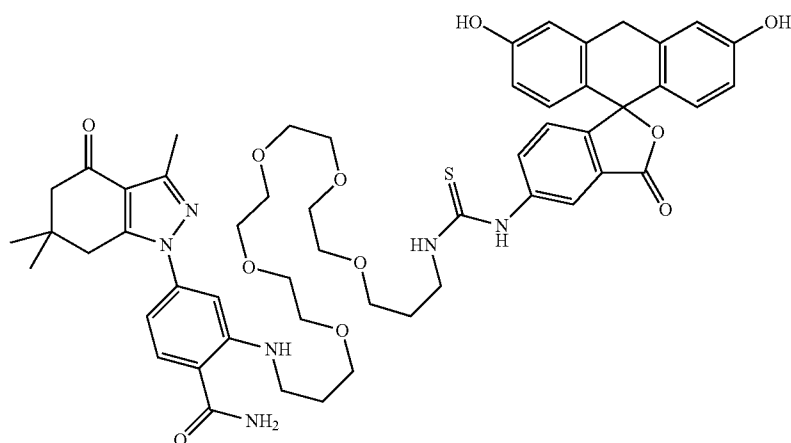
Example 4. Synthesis of Probe Compounds

2-((1-((3',6'-dihydroxy-3-oxo-3H-spiro[isobenzofuran-1,9'-xanthen]-5-yl)amino)-1-thioxo-6,9,12,15,18-pentaoxa-2-azahenicosan-21-yl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (10)

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2-((1-((3',6'-dihydroxy-3-oxo-3H-spiro[isobenzofuran-1,9'-xanthen]-5-yl)amino)-1-thioxo-6,9,12,15,18-pentaoxa-2-azahenicosan-21-yl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (10). Fluorescein 5(6)-isothiocyanate (35 mg, 90 μ mol) was dissolved in ethanol (10 mL) and treated with amine 8 (54.2 mg, 90 μ mol) followed by Hunig's base (35 mg, 270 μ mol) and stored in a drawer overnight. TLC ($\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{AcOH}$: 4/1/0.1) showed formation of a new product. The reaction mixture was concentrated to a glass and dissolved in DMSO/water (4/1, 2.5 mL). About half of the product was purified in two injections by chromatography (5 to 100% MeOH, Agilent C-18 21.2 \times 250 mm) to give the product (45 mg) as a yellow solid. TLC (4/0.9/0.1 $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$) R_f =0.3; ^1H NMR (DMSO- d_6) δ 10.2 (br s, 2H), 8.40 (br t, 1H), 8.34 (br s, 1H), 8.23 (s, 1H), 7.92 (b s, 1H), 7.73 (d, J=8.1 Hz, 2H), 7.26 (br s, 1H), 7.15 (d, J=8.1 Hz, 1H), 6.76 (s, 1H), 6.67 (s, 1H), 6.65 (br s, 2H), 6.59 (d, J=8.5 Hz, 2H), 6.54 (d, J=8.5 Hz, 2H), 3.47 (m, 20H), 3.19 (m, 2H), 2.91 (s, 2H), 2.38 (s, 3H), 2.31 (s, 2H), 1.79 (m, 4H), 1.00 (s, 6H); MS (ESI): m/z 993.5 $[\text{M}+\text{H}]^+$.

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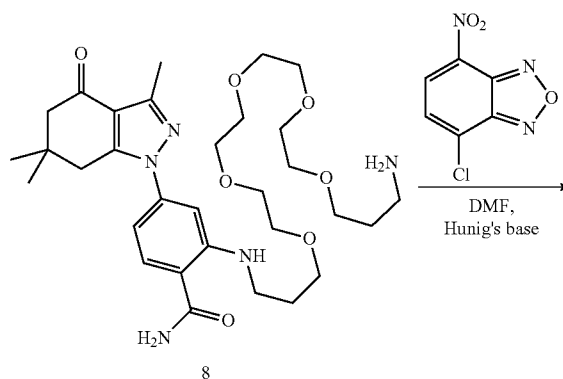
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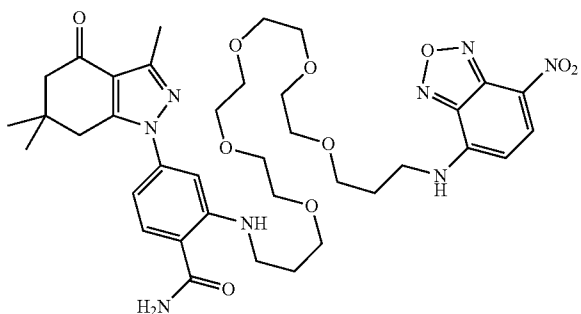
2-((19-((7-nitrobenzo[c][1,2,5]oxadiazol-4-yl)amino)-4,7,10,13,16-pentaoxononadecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (11)



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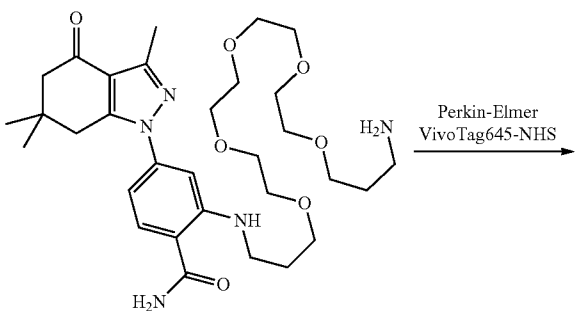
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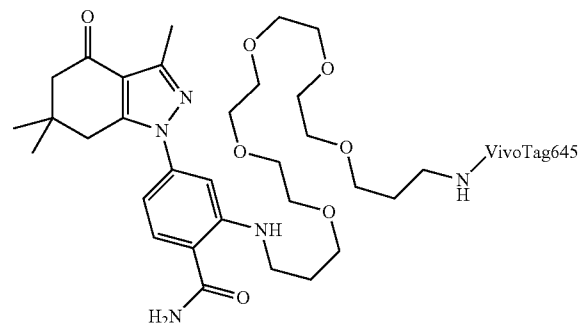
11

2-((19-((7-nitrobenzo[c][1,2,5]oxadiazol-4-yl)amino)-4,7,10,13,16-pentaoxonadecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (11). A mixture of Amine 8 (100 mg, 165 μmol) and 4-chloro-7-nitro-1,2,3-benzoxadiazole (40 mg, 200 μmol), Hunig's base (87 μL) and DMF (1 mL) were mixed. The solution turned immediately much darker. TLC (9/1: $\text{CH}_2\text{Cl}_2/\text{MeOH}$) showed no starting materials and a major new yellow product, which changed color under UV light. The mixture was concentrated and then chromatographed with CH_2Cl_2 (200 mL), $\text{CH}_2\text{Cl}_2/\text{MeOH}$ 9/1 (300 mL), 9/1 (300 mL). The cleanest fractions were combined and then rechromatographed with CH_2Cl_2 (200 mL), $\text{CH}_2\text{Cl}_2/\text{MeOH}$ 9/1 (500 mL), 9/1 (300 mL), to give the product (11) (52 mg, 41%) as a dark brown oil. MS (ESI): m/z 767.4 $[\text{M}+\text{H}]^+$.

VivoTag 645 Compound (12)



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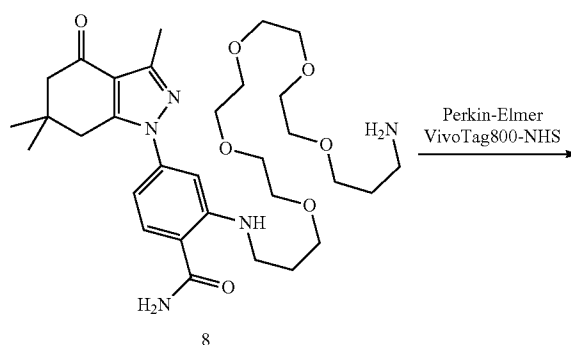


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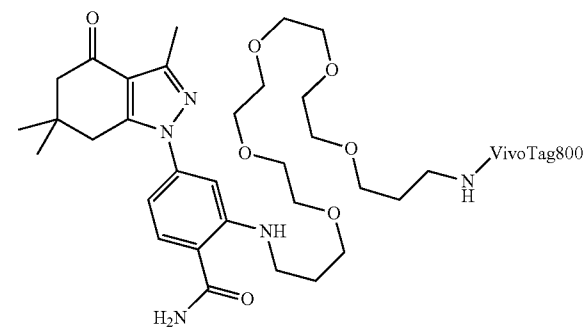
Amine 8 (4.5 mg, 7.45 μmol) was dissolved in DMSO (200 μL) and added to VivoTag 645 (Perkin Elmer, 2.5 mg, 1.8 μmol) in its original container and placed back into the freezer overnight. The sample was purified in 3 portions by reverse-phase chromatography and concentrated to give VivoTag 645 compound (12) (~ 3 mg) as a blue solid. MS (ESI): m/z 1577.5 $[\text{M}]^-$.

VivoTag 800 Compound (13)



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Perkin-Elmer
VivoTag800-NHS



14

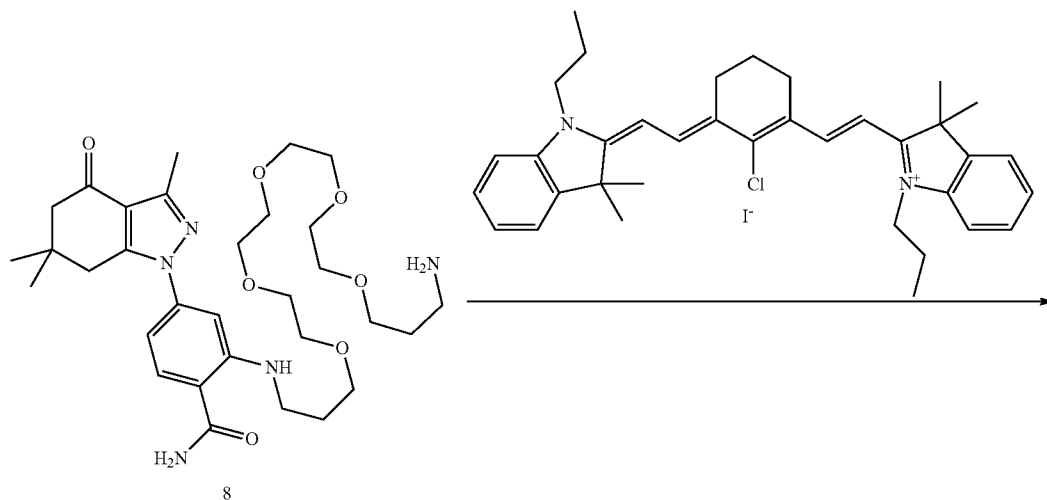
VivoTag 800 compound (13). Amine 8 (4.3 mg, 7.2 μmol) was dissolved in DMSO (100 μL) and treated with Hunig's base (10 μL) and added to VivoTag 800 (Perkin Elmer, 5 mg, 3.42 μmol) in its original container. The starting material vial was washed twice with DMSO (100 μL), which was added to the reaction mixture. The mixture was left in an aluminum bag at RT for 30 m, then placed back into the freezer for the 3 d. The sample was purified in 3 portions by reverse-phase chromatography and concentrated to give VivoTag 800 compound (13) (~ 9 mg) as a blue solid. MS (ESI): m/z 823.7 $[\text{M}-2]^{2-}$.

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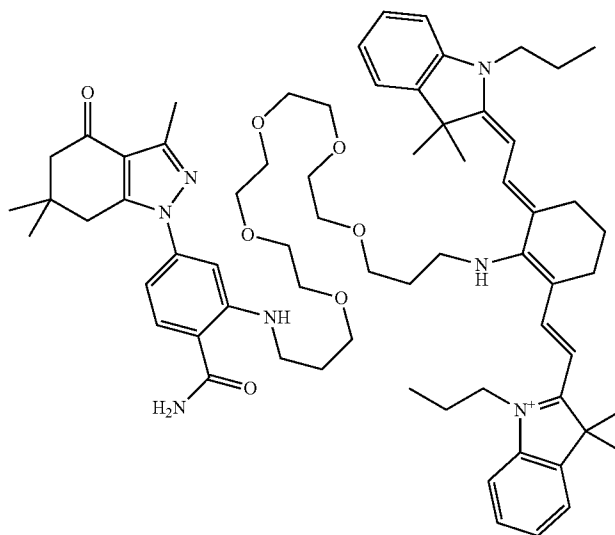
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2-((E)-2-((E)-2-((19-((2-carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-4,7,10,13,16-pentaoxanonadecyl)amino)-3-((E)-2-(3,3-dimethyl-1-propylindolin-2-ylidene)ethylidene)cyclohex-1-en-1-yl)vinyl)-3,3-dimethyl-1-propyl-3H-indol-1-ium (14)

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8



14

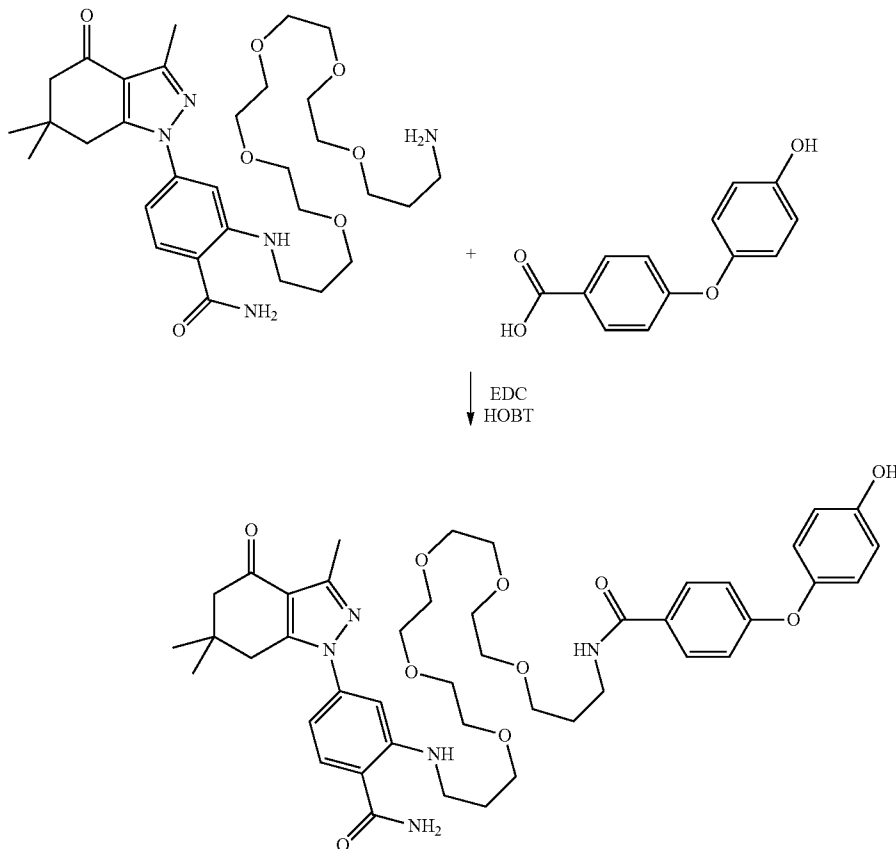
Amine 8 (12.1 mg, 20 μmol) was dissolved in DMSO (200 μL) and added to IR-780 iodide (7 mg, 10 μmol) and treated with Hunig's base (15 μL) followed by ethanol (100 μL). The sample was heated for 3 h at 70° C. The entire sample was added to a silica gel column (2.5x20 cm) and chromatographed with CH_2Cl_2 (200 mL), $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$ 39/0.9/0.1 (240 mL), 19/0.9/0.1 (240 mL) and 12.3/0.9/0.1 (240 mL). The blue band was concentrated to give product 14 (2.4 mg, 22%) as a blue solid. MS (ESI): m/z 554.1 $[\text{M}+\text{H}]^{2+}$.

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N-(19-((2-Carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-4,7,10,13,16-pentaoxanonadecyl)-4-(4-hydroxyphenoxy)benzamide (15)

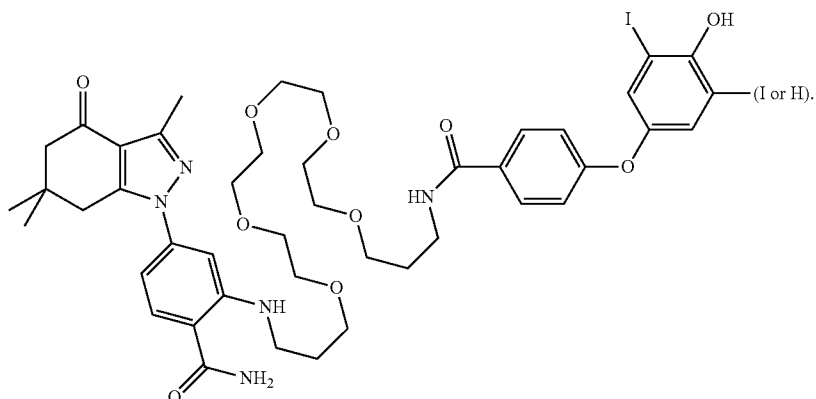


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Amine 8 (90 mg, 149 μmol), 4-(4-hydroxyphenoxy)benzoic acid (38 mg, 164 μmol), EDC (43 mg, 224 μmol), HOBT (21 mg, 156 μmol) and 1 chip of DMAP were dissolved in DMF (1 mL) and stirred at RT. Additional acid and EDC were added until starting amine disappeared (by TLC analysis). The mixture was then treated with 1N HCl (5

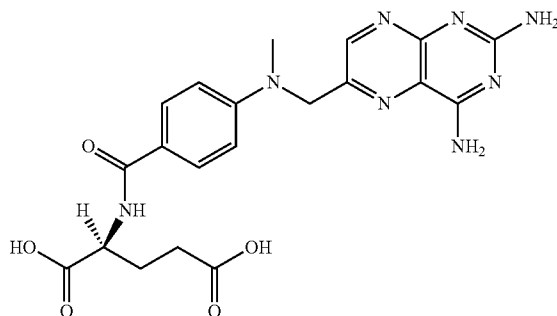
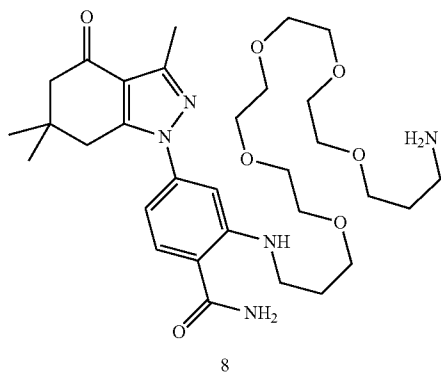
25 cm). The product peak was concentrated to give 15 (44 mg, 36%) as a clear glass. (ESI): m/z 816.6 $[\text{M}+\text{H}]^+$.

45 The phenol moiety of compound 15 will be iodinated using standard methods to produce the following compounds:

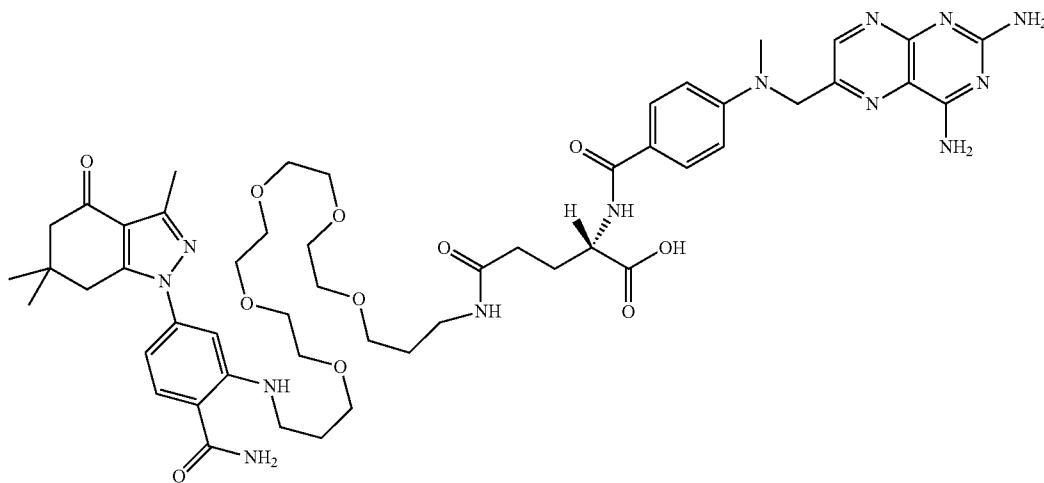


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(S)-1-((2-Carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-24-(4-(((2,4-diaminopteridin-6-yl)methyl)(methyl)amino)benzamido)-21-oxo-4,7,10,13,16-pentaoxa-20-azapentacosan-25-oic acid (16)



↓ EDC
HOBT



Amine 8 (36 mg, 60 μ mol) and methotrexate (MTX, 30 mg, 65 μ mol) were dissolved in DMF (0.5 mL) and then treated with EDC (12 mg, 62 μ mol) in DMF (150 μ L) quickly and stirred at room temperature. After one day, more MTX (6 mg) was added to the mixture followed by more EDC (1.2 mg). After another 2 h the reaction mixture was shot directly onto a prep HPLC (Agilent Prep C-18, 2.5 \times 25 cm, 5 to 100% MeOH w/2% formic acid) and the product collected and concentrated to give product 16 (40 mg, 64%) as a yellow glass.

2-((1-hydroxy-3,6,9,12-tetraoxapentadecan-15-yl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (19) and 2,2'-(4,7,10,13,16-pentaoxanonadecane-1,19-diylbis(azanediyl))bis

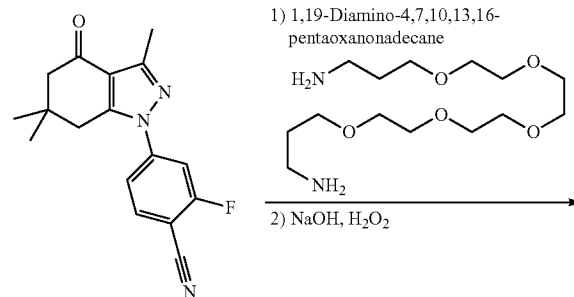
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(4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide) (20)

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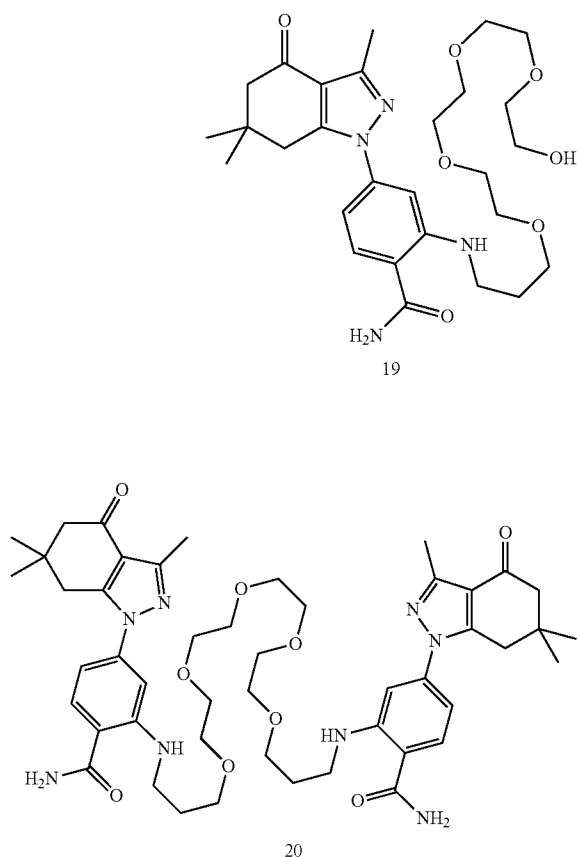
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-continued

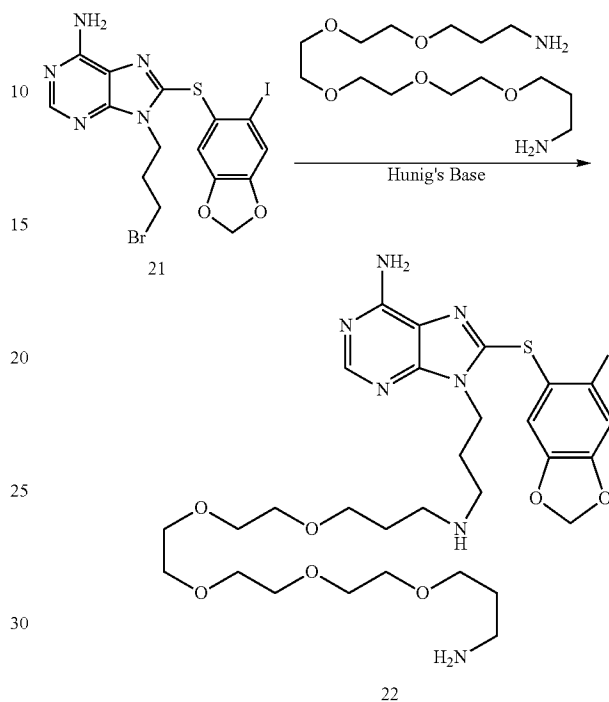


Using the synthesis of Amine 8, two by-products resulting from bis-coupling and a diamine impurity were recovered. The mixture (~100 mg) was dissolved in DMSO (1 mL) and injected on a prep HPLC (PFH-002_078 Prep, 5 to 100% methanol, Agilent C-18, 21.1×25 cm). The early eluting peaks was concentrated to give alcohol 19 (48 mg). (ESI): m/z 547.5 $[M+H]^+$. The later eluting peak was concentrated to give the symmetrical compound 20 (34 mg). (ESI): m/z 899.7 $[M+H]^+$.

46

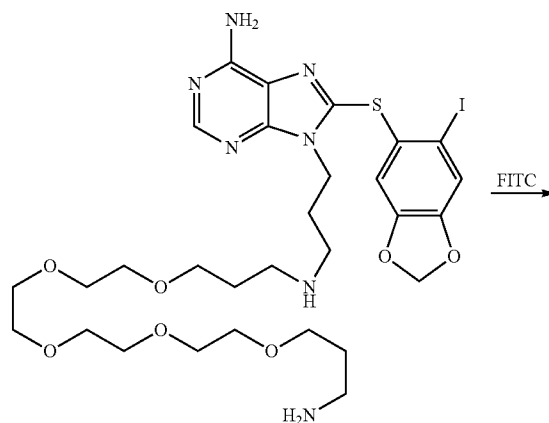
Example 5. Synthesis of PU-H71 Derivatives

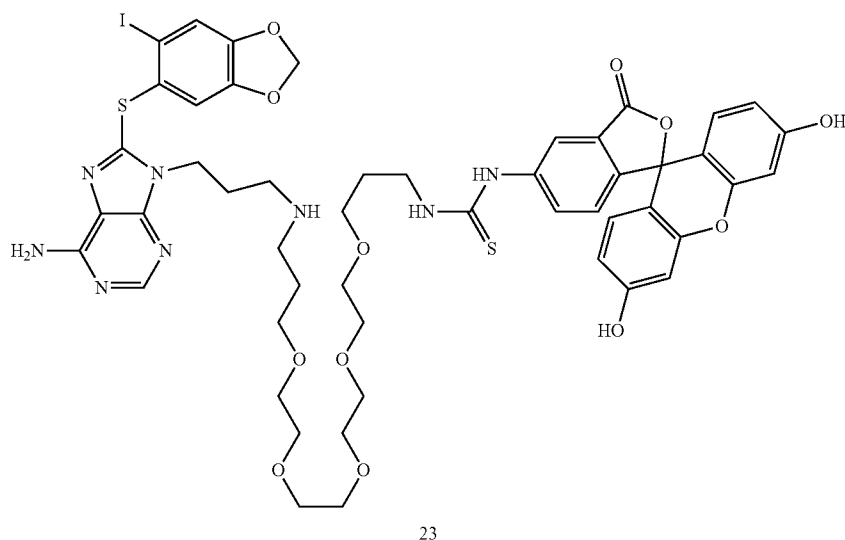
N1-(3-(6-amino-8-((6-iodobenzo[d][1,3]dioxol-5-yl)thio)-9H-purin-9-yl)propyl)-4,7,10,13,16-pentaoxanonadecane-1,19-diamine (22)



9-(3-Bromopropyl)-8-((6-iodobenzo[d][1,3]dioxol-5-yl)thio)-9H-purin-6-amine (21, prepared according to *Bioorg. Med. Chem.* 19, 2603-2614 (2011)) (14 mg, 26 μ mol) was treated with 1,9-diamino-4,7,10,13,16-pentaoxanonadecane (24 mg, 79 μ mol) in ethanol (1 mL) and DMSO (400 μ L). The ethanol was removed under vacuum and the mixture stirred at RT for 1 day. The mixture was purified by preparative HPLC (5 to 100% methanol, 20 mL/m, Agilent C-18, 21.1×25 cm) to give the product 22 (14 mg, 70%) as a clear oil. (ESI): m/z 762.2 $[M+H]^+$.

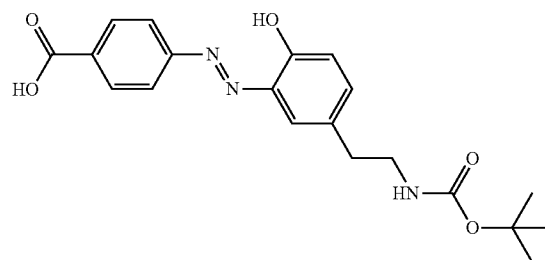
1-(23-(6-amino-8-((6-iodobenzo[d][1,3]dioxol-5-yl)thio)-9H-purin-9-yl)-4,7,10,13,16-pentaoxa-20-azatricosyl)-3-(3',6'-dihydroxy-3-oxo-3H-spiro[isobenzofuran-1,9'-xanthen]-5-yl)thiourea (23)





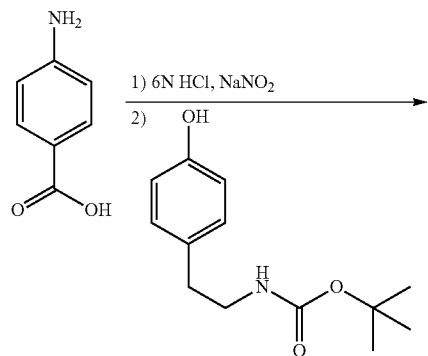
Amine (22) (all 14 mg, 18 μ mol) was dissolved in ethanol (1 mL) and added to FITC (10 mg, 25.7 μ mol). Additional solid persisted so more ethanol (250 μ L) was added, followed by DMSO (250 μ L) and Hunig's base (25 μ L). The mixture was concentrated and purified by preparative HPLC (PFH-003-027Prep, 5 to 100% methanol, 20 mL/m, Agilent C-18, 21.1 \times 25 cm) to give the fluorescent product (23) (16 mg, 75%) as a yellow glass. (ESI): m/z 1151.3 $[M+H]^+$.

-continued



Example 6. Synthesis of Compound with a Cleavable Linker

(E)-4-((5-(2-((tert-butoxycarbonyl)amino)ethyl)-2-hydroxyphenyl)diazenyl)benzoic acid (17)



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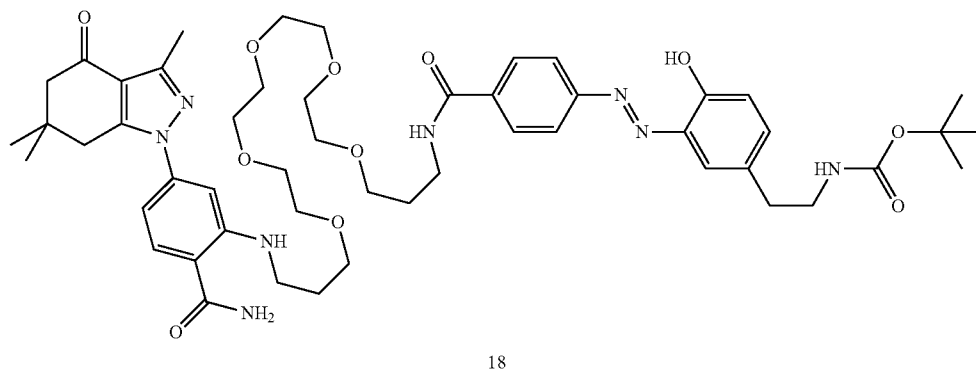
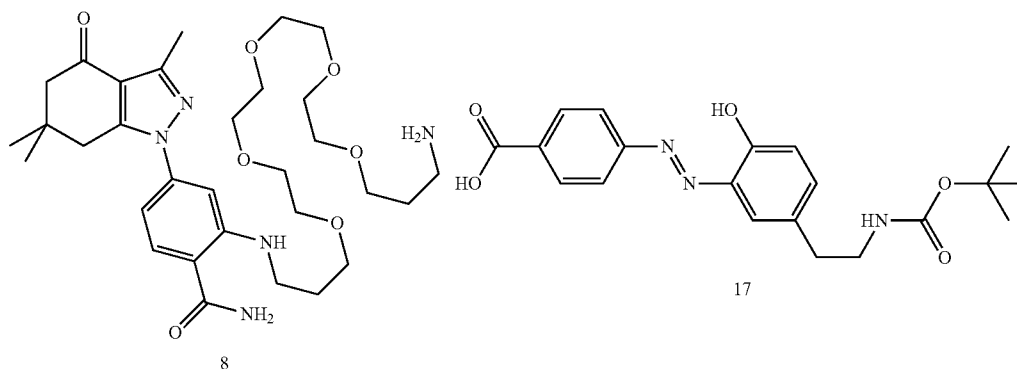
65

4-Aminobenzoate (500 mg, 3.6 mmol) was slurried in 6N HCl (10 mL), cooled to 0° C. and treated slowly with sodium nitrite (629 mg, 9.11 mmol). After stirring for 20 m, the mixture was added slowly to an ice-cooled solution of N-Boc tyramine (865 mg, 3.6 mmol) in saturated sodium bicarbonate solution (40 mL) with added sodium bicarbonate (4 g) and a bit of acetone (~5 mL). The orange reaction slurry was left to stir overnight. The reaction mixture was treated 1N HCl (100 mL) until acidic and then stirred an additional 2 h. The solids were filtered off, washed with water and air dried, then dried under vacuum to give 17 (1.03 g, 73%) as a reddish-orange solid. The product was used as is though it contained a minor impurity by NMR. TLC (9/1 CH_2Cl_2 /MeOH) R_f =0.20; 1H NMR (DMSO- d_6) δ 13.19 (br s, 1H), 10.82 (s, 1H), 8.11 (d, J =8.5 Hz, 2H), 8.05 (d, J =8.5 Hz, 2H), 7.55 (s, 1H), 7.28 (d, J =8.3 Hz, 1H), 7.00 (d, J =8.3 Hz, 1H), 6.88 (t, 1H), 3.12 (m, 2H), 2.67 (t, J =6.8 Hz, 2H), 1.33 (s, 9H); MS (ESI): m/z 384.2 $[M]^-$, 791.4 $[2M+Na]^-$; HRMS (ESI) $[M+Na]^+$ calcd for $C_{20}H_{23}N_3O_5Na$, 408.1536; found 408.1518.

49

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(E)-tert-butyl 3-((4-((19-((2-carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-4,7,10,13,16-pentaoxanonadecyl)carbamoyl)phenyl)diazenyl)-4-hydroxyphenethylcarbamate (18)



18

Amine 8 (483 mg, 800 μ mol), acid 17 (308 mg, 800 μ mol), EDC (161 mg, 840 μ mol) and HOBT (113 mg, 840 μ mol) and 2 chips of DMAP and were dissolved in CH_2Cl_2 (10 mL) and stirred at RT for 2 h. The reaction mixture was added to a column and chromatographed (2.5 \times 20 cm, silica gel, CH_2Cl_2 (300 mL), 9/1 $\text{CH}_2\text{Cl}_2/\text{MeOH}$ (300 mL), 4/1 $\text{CH}_2\text{Cl}_2/\text{MeOH}$ (300 mL). The active fractions were combined and concentrated to a frothy glass to give 18 (560 mg, 72%). The hard glass was scraped out to give an orange

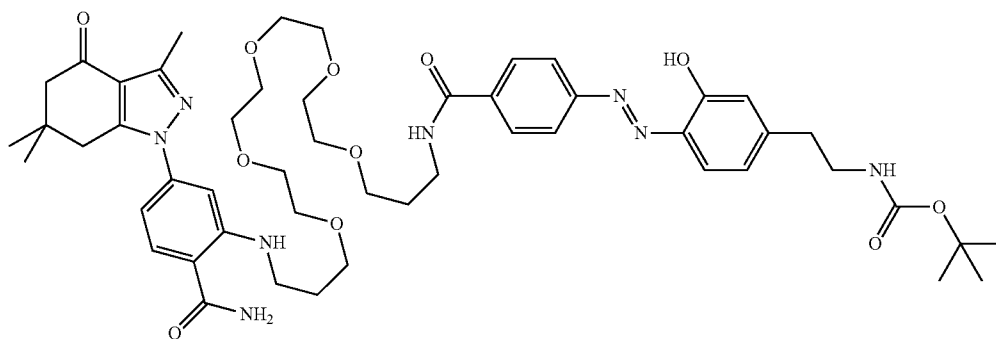
powder (460 mg). TLC (9/0.9/0.1 $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$) R_f =0.44; ^1H NMR ($\text{DMSO}-d_6$) δ 10.85 (s, 1H), 8.60 (br t, 1H), 8.40 (br t, 1H), 8.02 (br s, 4H), 7.92 (b s, 1H), 7.73 (d, 60 J =8.1 Hz, 1H), 7.55 (s, 1H), 7.27 (d, J =8.1 Hz, 1H), 7.00 (d, J =8.1 Hz, 1H), 6.88 (br t, 1H), 6.76 (s, 1H), 6.66 (d, J =8.1 Hz, 1H), 3.47 (m, 20H), 3.19 (m, 1H), 3.12 (m, 1H), 2.90 (s, 2H), 2.67 (t, 2H), 2.52 (m under DMSO, 2H), 2.38 (s, 3H), 65 2.31 (s, 2H), 1.77 (m, 4H), 1.33 (s, 9H), 0.99 (s, 6H); HRMS (ESI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{51}\text{H}_{71}\text{N}_8\text{O}_{11}$, 971.5237; found 971.523575.

51

Example 7. Preparation of Affinity Chromatography Resin with a Cleavable Linker (E)

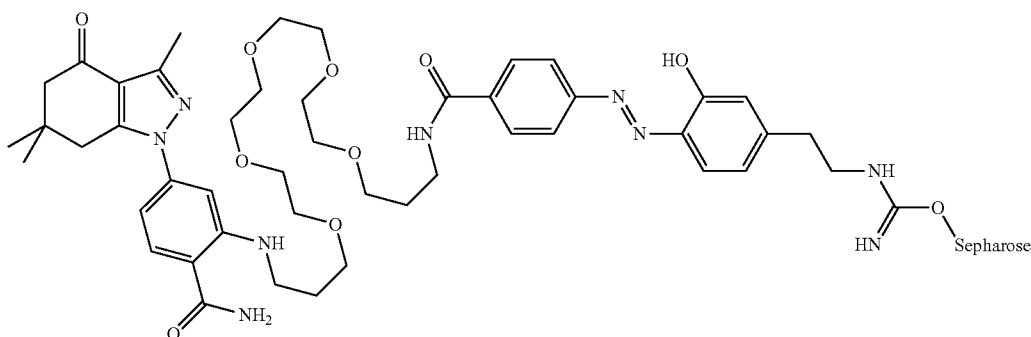
52

High buffer 0.1 M TRIS-HCl, 0.5 M NaCl pH=8
Storage buffer 0.1M KH_2PO_4 , pH=7.4 w/200 mg NaN_3/L



18

- 1) TFA
- 2) CNBr-activated Sepharose™ 4B



E

CNBr-activated Sepharose™ 4B was obtained from GE Healthcare Life Sciences. The following procedure generally followed GE Healthcare Instructions 71-7086-00 AFA.

Buffers and Solutions:

Swelling solution 1 mM HCl

Coupling buffer 0.1 M NaHCO₃, 0.5 M NaCl, pH=8.3

Capping solution 1 M ethanolamine

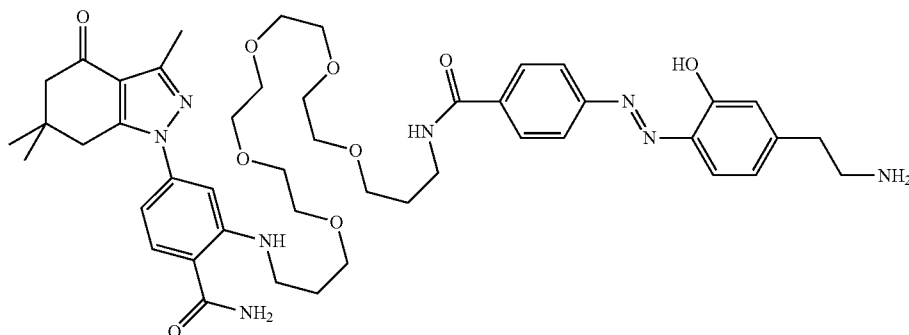
Low buffer 0.1 M AcOH/NaAcOH, 0.5 M NaCl pH=4

Ligand 18 (25 mg 25.74 μmol) was dissolved in trifluoroacetic acid (1 mL). TLC (9/1/0.1: $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$) showed loss of starting material and formation of a lower product, corresponding to compound 24 (HRMS (ESI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{46}\text{H}_{63}\text{H}_8\text{O}_9$, 871.4713; found 871.4696). The mixture was concentrated, then dissolved in ethanol (5 mL) and concentrated again. The residue was then dissolved in ethanol (5 mL) for addition to the resin.

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24

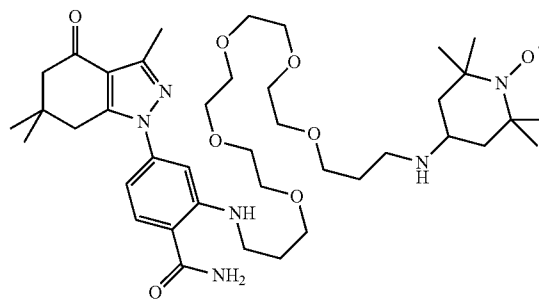


In a big 275 mL column, CNBr-activated Sepharose™ 4B (25 g) was swelled in 1 mM HCl (450 ml) and then washed with 1 mM HCl (5 L). The resin was washed with coupling buffer (125 mL) and then slurried with coupling buffer (125 mL). The mixture was then treated with the compound described above. The mixture was tumbled at RT for 4 h. The resin was then drained and washed with coupling buffer (5×125 mL), diluted with more coupling buffer (~125 mL) and treated with capping solution (2 mL) and rotated at RT for 2 h. The solution was drained and washed with 3 rounds of high buffer/low buffer (250 mL ea.) and finally washed with water (250 mL) and transferred in storage buffer (125 mL) to a bottle and stored at 4° C.

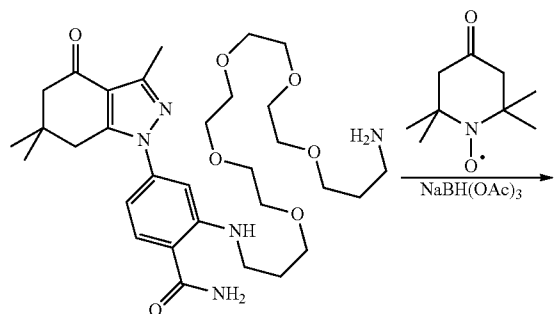
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Example 8. Synthesis of Other Compounds for Hsp90 Binding and Cell Penetration Comparisons

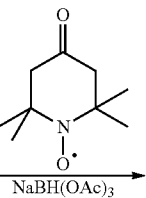
2-((19-(((1-oxo-2,2,6,6-tetramethylpiperidin-4-yl)amino)-4,7,10,13,16-pentaoxanonadecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (25)



25
C₄₀H₆₅N₆O₈⁺
757.98



8
C₃₁H₄₉N₅O₇
603.75



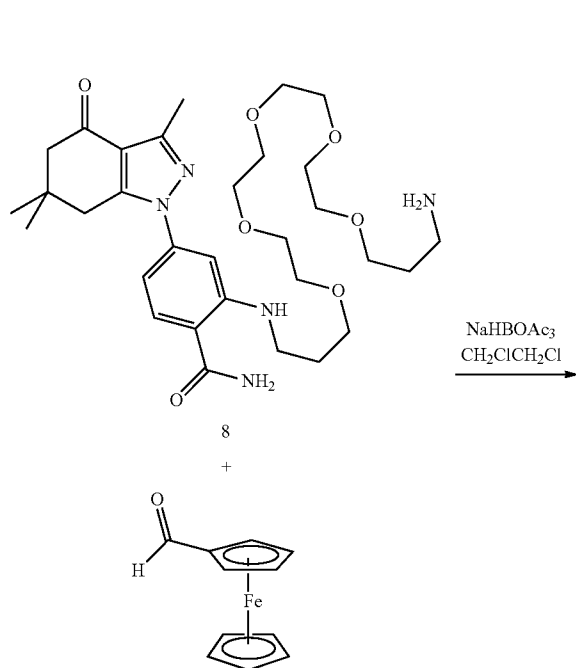
NaBH(OAc)₃

Amine 8 (50 mg, 83 μmol) was dissolved in dichloroethane (2 mL) and treated with 4-oxo-TEMPO (14 mg, 83 μmol) followed by sodium triacetoxyborohydride (23 mg, 107 μmol) and stirred at RT overnight. TLC (CH₂Cl₂/MeOH/AcOH: 4/1/0.1) showed primarily a new product. The mixture was concentrated then dissolved in DMSO/water and chromatographed (5 to 100% MeOH, Agilent C-18 21.2×250 mm) to give product 25 (39 mg, 63%) as a glass. TLC (9/0.9/0.1 CH₂Cl₂/MeOH/NH₃) R_f=0.55; ¹H NMR (DMSO-d₆) δ spectra not assigned due to broadening by TEMPO; MS (ESI): m/z 758.5 [M+H]⁺.

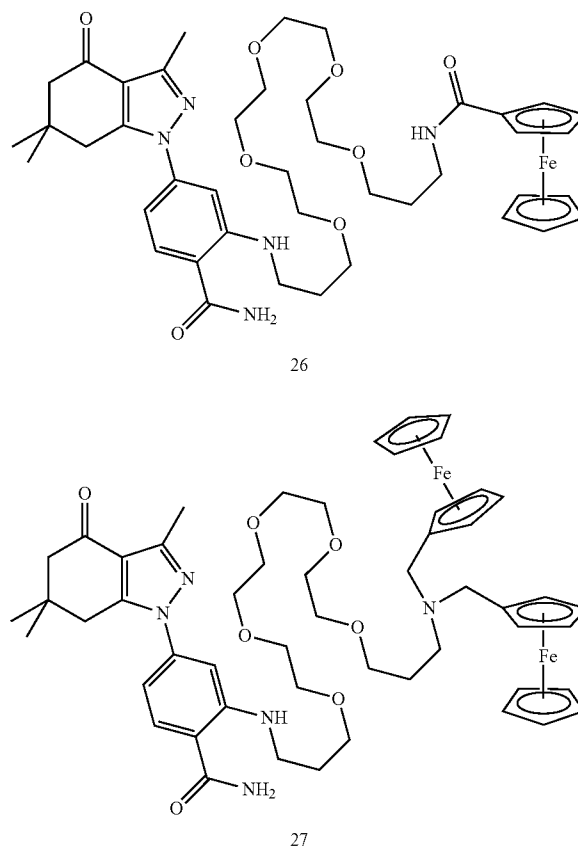
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2-((19-(ferrocenemethylamino)-4,7,10,13,16-pentaoxonadecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (26),
 2-((19-di(ferrocenemethylamino)-4,7,10,13,16-pentaoxonadecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (27)

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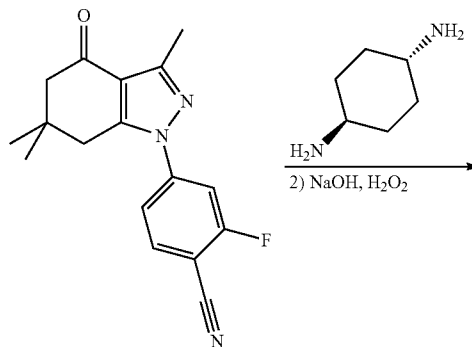


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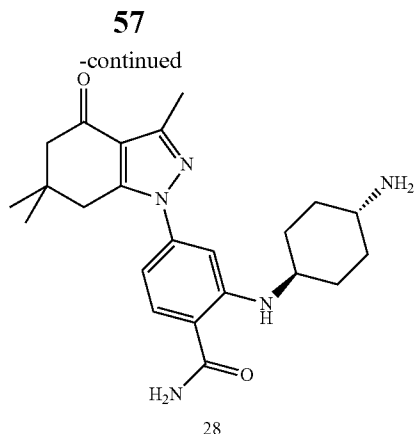


Amine 8 (673 mg, 1.11 mmol) and ferrocene carboxaldehyde (239 mg, 1.11 mmol) were dissolved in dichloroethane (5 mL) and treated with sodium triacetoxy-borohydride (283.5 mg, 1.34 mmol) followed by acetic acid (77 μ L). The reaction mixture was stirred over the weekend. The mixture was adsorbed onto silica (3 g), added to a dry column (1.5 \times 12 cm silica gel) and eluted with 100%, 19/1/0.1, then 9/1/0.1: $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_4\text{OH}$ (300 mL ea.) to give the less polar product, 18 (~180 mg) and the more polar product (~650 mg), both as yellow glasses. The products were separately dissolved in DMSO and chromatographed by HPLC (Agilent Prep C-18, 2.5 \times 25 cm, 5 to 100% MeOH w/2% formic acid) to give 26 (490 mg, 54%) and 27 (200 mg, 17%) as viscous yellow glasses. 26. TLC (9/0.9/0.1 $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$) $R_f=0.23$; ^1H NMR (CDCl_3) δ 8.52 (s, 1H), 7.98 (br s, 1H), 7.51 (d, $J=8.1$ Hz, 1H), 6.78 (d, $J=1.6$ Hz, 1H), 6.59 (dd, $J=1.6, 8.1$ Hz, 1H), 6.44 (br s, 1H), 4.3 (br s, 2H), 4.16 (br s, 2H), 4.11 (s, 5H), 3.91 (br s, 2H), 3.57 (br m, 20H), 3.28 (br s, 2H), 2.97 (br t, 2H), 2.79 (s, 2H), 5.52 (s, 3H), 2.37 (s, 2H), 1.91 (br m, 4H), 1.08 (s, 6H); MS (ESI): m/z 802.4 $[\text{M}+\text{H}]^+$. 27 TLC (9/0.9/0.1 $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$) $R_f=0.40$; MS (ESI): m/z 1000.5 $[\text{M}+\text{H}]^+$.

2-(((1r,4r)-4-aminocyclohexyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (28)

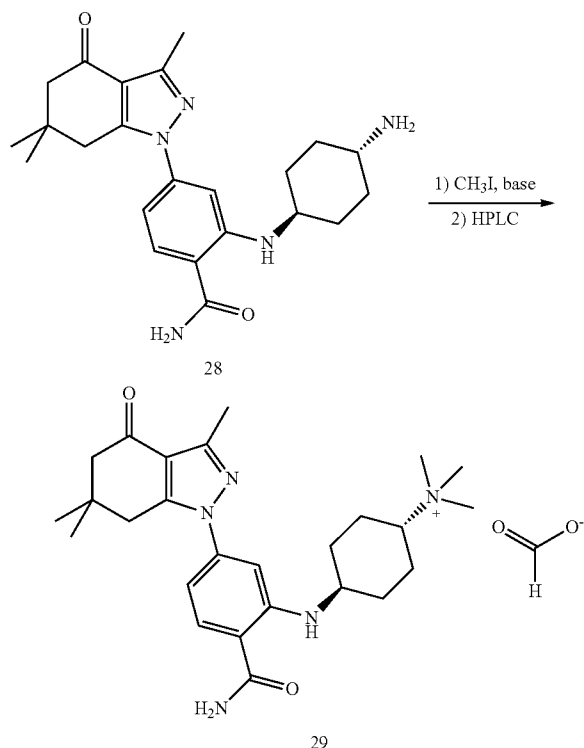


2



A mixture of 2 (500 mg, 1.68 mmol) and trans-1,4-diaminocyclohexane (1.15 g, 10 mmol) and DMSO (1.5 ml) were stirred at RT for 3 days. The mixture was then diluted with ethanol (3 mL), heated to 90° C. and treated with 50% NaOH (12 drops) and then treated, very slowly, a drop at a time, with 30% hydrogen peroxide. After each drop, the reaction foamed up a bit. After 12 drops the reaction mixture was diluted with water (20 mL) and stirred rapidly overnight. A lot of powder fell out and was filtered off, washed with water and air dried to give 28 (689 mg, 100%) as an off-white powder. MS (ESI): m/z 410.3 [M+H]⁺.

(1*r*,4*r*)-4-((2-carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-N,N,N-trimethylcyclohexanaminium (29)



A mixture of 28 (100 mg, 244 μ mol) and methyl iodide (208 mg, 1.5 mmol) were dissolved in methylene chloride (2 mL) containing solid sodium bicarbonate (123 mg, 1.5

58

mmol) and stirred at RT. With monitoring by LC/MS, additional methyl iodide (total 1.6 mL), sodium bicarbonate (123 mg) and DMF (500 μ L) were added over 4 days until some tetra-methylation was detected. The sample was concentrated then diluted with DMSO and water to give a milky solution which was injected on the Prep HPLC (PFH-003_002 Prep, 5 to 100% methanol (0.2% formic acid), 20 mL/m, Agilent C-18, 21.1 \times 25 cm). The active fractions were combined and stirred in ethanol to yield a powder which was filtered off to give 29 (28 mg, 23%) as a white powder. MS (ESI): m/z 452.5 [M+H]⁺.

Example 9. Selectivity Studies

Several media were prepared as described in Example 7, using compounds 6, 7, 8 and 9. These resins were termed A, B, C and D respectively. To prepare the resin in each case, the appropriate compound was added at 1-10 μ mol/gram of resin in minimal methanol.

The ability of each resin to selectively capture Hsp90 from pig mammary gland extract, a tissue shown to be high in ATP binding proteins including native forms of Hsp90, GRP94 and TRAP1, was evaluated. The resin was incubated in the protein solution then washed with a high salt buffer. The bound proteins were removed via an SDS boil procedure, separated by SDS-PAGE electrophoresis, located by silver staining and identified using mass spectrometry (MS) sequencing. Selectivity towards Hsp90 was demonstrated by inclusion of 1 mM compound 5 in the tissue extract prior to mixing with the affinity resin. In the case of resin E (Example 7), bound proteins were eluted with 25 mM sodium dithionite in phosphate-buffered saline.

Results are illustrated in FIG. 1. Numbers indicate bands that were sequenced by MS: (1) Fatty acid synthase. (2) Hsp90 (3) Hsp90 (4) Hsp90 proteolytic fragments.

A large number of proteins, including Hsp90, were retained on some resins. The competition experiment, performed by pre-incubating the protein solution with known ligand compound 5, showed clean exclusion of Hsp90 but not other proteins, demonstrating that proteins other than Hsp90 are not client proteins.

Resins A and B showed significant non-specific binding as illustrated in FIG. 1. Resin C showed an intense band of Hsp90, and quenching with compound showed complete blocking of Hsp90 binding as well as several N-terminal fragments. MS analysis demonstrated that all of the recovered proteins were Hsp90 or proteolytic fragments of the protein, suggesting that this resin is selective for Hsp90 over GRP94 and TRAP-1. Resin D was more selective than A and B, although some non-specific binding was observed.

Following exposure of resin E to pig mammary gland extract and treatment with 25 mM sodium dithionite, the eluted proteins were identified by MS. The retained protein profile (FIG. 1, lane E) was essentially identical to resin C, except that the proteins were recovered using non-denaturing conditions.

Example 10. Additional Selectivity Studies

To further explore the selectivity, the elution of pig mammary gland proteins from gamma phosphate linked ATP Sepharose was studied with compound 8, as well as known potent Hsp90 inhibitors PU-H71 (Caldas-Lopes et al. *Proc. Natl. Acad. Sci. USA* 106(20), 8368 (2009)) and SNX-2112 (Chandarlapaty et al. *Clin. Cancer Res.* 14(1) 240 (2008)). This ATP resin has been used previously to study purine-binding proteins (Graves et al. *Mol. Pharma-*

col. 2002, 62, 1364) and to discover novel Hsp90 inhibitors (Huang et al. *J. Med. Chem.* 2009, 52, 4288.)

The ATP resin was charged with the pig mammary gland extract and aliquots were distributed into individual wells. Proteins eluted from the wells with increasing amounts of indicated compound were analyzed as described above; results are illustrated in FIG. 2. Compound PU-H71 demonstrates strong potency towards both Hsp90 and GRP94, whereas SNX-2112 shows weaker affinity for GRP94. Compound 8 shows essentially no elution of GRP94.

Example 11. Additional Selectivity Studies

The purinome was captured from pig mammary gland extract as described (Hughes et al., *Bioorg Med Chem.* 2012, 20(10):3298-3305) on gamma phosphate linked ATP Sepharose. Following extensive washing with physiological buffers to remove non-specifically associated proteins, the resin was divided equally and interrogated with the indicated compounds in parallel as described (Hughes et al 2012). All eluted proteins were characterized by SDS-PAGE and silver staining, and then identified by mass spectrometry. Results are illustrated in FIG. 3. The experiment shows that compounds 5, 17-AAG (Schulte et al. *Cancer Chemoth. Pharm.* 42(4), 273 (1998)) and PU-H71 have specificity for both Hsp90 and GRP94, whereas compounds 8, 27, 11, 28, 29, 19, 20 only recognize Hsp90. Key: GRP94, glucose regulated protein 94; Hsp90, heat shock protein 90; LAPT, lapatinib; Hsp90 STD, purified Hsp90; 17-AAG, derivatized form of geldanamycin, BLK, vehicle control consisting of buffer containing 10% DMSO used to solubilize all compounds included in the test.

Example 12. Evaluation of Compound 10 in Cancer Cells

In one experiment, the fluorescein compound 10 was applied at micromolar concentrations to four types of tumor cells. As illustrated in FIG. 4, the cell lines MCF-7, BT474 and MDA-MB 468, which are metastatic forms of human breast cancer, take up the probe readily. The Huh7 cell line, a human liver non-metastatic cell line, does not absorb the probe. The probe (10) fluoresces to give green staining in images obtained; in FIG. 4, the green staining appears white. DAPI staining shows the nucleus in blue in images obtained; in FIG. 4, the blue staining appears gray.

In another experiment, MCF7 (metastatic) and MCF10 (benign) cells were treated with an Hsp90 antibody±membrane permeabilization with Triton X 100. As shown in FIG. 5A, in non-permeabilized cells the antibody recognizes surface Hsp90 in MCF7 cells but not in MCF10 cells. After permeabilization the antibody recognizes intracellular Hsp90. Significantly more Hsp90 staining is observed in the MCF7 line. The experiment was also performed using fluorescein compound 10. As shown in FIG. 5B, the probe enters and detects Hsp90 in MCF7 cells, but not in MCF10 cells. The probe also detects more intracellular Hsp90 in MCF7s following permeabilization compared with MCF10s consistent with increased expression of Hsp90. These results suggest up-regulation of Hsp90 in metastatic cells only. In FIG. 5 the green staining from compound 10 fluorescence appears white, while blue from the DAPI-stained nuclei appears gray.

As shown in FIG. 6A, compound 10 was added to MCF7 cells alone. In FIG. 6B, the compound was added in the presence of an antibody to Hsp90. The antibody prevents internalization of surface 90 causing the probe to accumulate

at the membrane. In FIG. 6C, compound 5 competes away the fluorescence signal. DAPI (blue) highlights the nucleus of the cell. In FIG. 6 the green staining from compound 10 fluorescence appears white, while blue from the DAPI-stained nuclei appears gray.

Example 13. Labeling of MDA-MB-468 Tumor Cells in NOD/SCID Mice

MDA-MB-468 tumor-bearing NOD-SCID mice were ip injected with compound 10 (1 mg/mouse, in 100 μ L of DMSO). After 1 and 2 hours, mice were sacrificed and tumors and spleens were harvested. Tumors were digested with collagenase/hyaluronidase/Dnase for 1 hour, and ficoll, and washed, then stained with anti-mouse CD45+ anti-H2-kd) (APC). Tumor cells were analyzed by flow cytometry, acquired by LSRII for FITC color compound 10 and mouse lineage expression. Lineage negative cells were analyzed for compound 10 labeling (% positivity is shown in each dot plot). Results are illustrated in FIG. 7A.

Spleen were minced and passed through the strainer. Splenocytes were acquired as described above to analyze compound 10 labeling. Results are illustrated in FIG. 7B.

Example 14. Xenografts

FIG. 8 shows images of xenografts (MDA-MB 468) harvested one hour after injection (IP) with compound 10. Tissue slices were prepared from the harvested xenografts and prepared for analysis by confocal analysis. The presence of compound 10 was detected within the tumor cells confirming uptake of the probe in vivo. A tissue slice from the control experiment is shown in which the animal was injected with vehicle alone (DMSO). Only background fluorescence is detected. In FIG. 8 the green staining from compound 10 fluorescence appears white or light gray, while blue from the DAPI-stained nuclei appears dark gray.

Example 15. Competition Experiments

BT474 cells were incubated with varying doses of compound 5 (0 μ M, 0.01 μ M, 0.1 μ M, 1 μ M and 10 μ M) in the presence of a fixed concentration of compound 10 (10 μ M) and a cell permeabilizing agent (escin) for 30 minutes. The cells were then washed to remove the free probe and cell extracts prepared from the homogenized cells. The amount of drug uptake was then measured by fluorescence (Ex488/Em522 nm) in a micro plate fluorescence reader. FIG. 9 shows that the signal from compound 10 can be effectively competed in vivo with the free parent drug, compound 5, in a dose-dependent manner. This data demonstrates that the fluorescence signal measured in cell and xenograft studies is derived from the probe bound to Hsp90.

Example 16. Measuring the Biodistribution of Compound 10 In Vivo in Mouse Xenografts

MDA-MB-468 cells (5M cells/mouse) were injected to the flank of NOD/SCID mice. When tumors reached 5-10 mm in diameter, tumor-bearing mice were intraperitoneally injected with compound 10 (1 mg/mouse, in 100 μ L of DMSO). After 2, 24, 48 or 72 hours, mice were sacrificed and tumors, livers and spleens were harvested. Tumors and livers were digested with collagenase/hyaluronidase/DNase for 1 hour at 37° C., and ficoll to eliminate debris and dead cells, and then washed. Cells in tumor digest were labeled with anti-mouse CD45-biotin+anti-H2-kD-biotin for 15 min

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on ice, and then 15 min with streptavidin-APC. Tumor cells were analyzed by flow cytometry to detect fluorescence from compound 10. Mouse lineage negative cells were gated for the analysis. Spleen were minced and passed through the strainer. Splenocytes, liver cells and tumor cells were analyzed by flow cytometry to detect fluorescence from compound 10.

Spleen were minced and passed through a strainer to get rid of aggregations/membranous tissue. Cells were stained with violet viability dye. Viable cells were analyzed for their compound 10 uptake. FIG. 10A shows little accumulation of compound 10 in splenocytes.

Liver tissues were minced with blades and digested with triple enzyme buffer (collagenase III, hyaluronidase, DNase) for 1 h at 37° C., and cells were passed through a strainer to get rid of aggregations. Cells were stained with violet viability dye. Viable cells were analyzed for their compound 10 uptake. FIG. 10B shows little accumulation of compound 10 in liver cells.

Tumor tissues were minced with blades and digested with triple enzyme buffer (collagenase III, hyaluronidase, DNase) for 1 h at 37° C., and cells were passed through a strainer to get rid of aggregations. Cells were stained with anti-mouse CD45-biotin/anti-H2-kd-biotin and then streptavidin-APC and violet viability dye. Viable and CD45/H2-kd negative cells were analyzed for their compound 10 uptake. FIG. 10C shows a high degree of accumulation of compound 10 in tumor cells.

FIG. 11 shows quantitation of compound 10 accumulation in vivo following harvest of the indicated tissues from mice injected with the probe at the indicated time points (5 mice/time point).

Example 17. Tumor Labeling Experiment

MDA-MB-468 tumor cells (5 M cells/mouse) were injected to the flank of NOD/SCID mice. After 3 weeks, tumor volumes reached 400-450 mm³. Mice were treated/untreated with intraperitoneal injection of compound 10 (1 mg/mouse). After 2, 24, 48 and 72 hours, mice were sacrificed, and tumor tissues were frozen in OCT compound. FITC(+) cells in tumor tissues (frozen sections) were analyzed under fluorescent microscope (Axio Observer). From 2 to 48 h, tumor cells showed intense FITC signal, as shown in FIG. 12. In FIG. 12, the green staining from compound 10 fluorescence appears white or light gray.

Example 18. Recovery of Compound 10 Bound to Hsp90 In Vitro and In Vivo

Hsp90 was purified from pig mammary gland as described in Hughes et al., *Bioorg Med Chem.* 2012, 20(10): 3298-3305. The purified protein (0.3 µg) was mixed with an excess of compound 10. The unbound probe was removed by dialysis and the labeled protein subjected to microanion exchange chromatography using a Pharmacia SMART system. Column fractions (100 µL) were collected and analyzed for fluorescence in a plate reader. Chromatographs are shown in FIG. 13A, showing a single peak was recovered by fluorescence that correlated with elution of Hsp90 as determined by Western blot and mass spectrometry. The chromatography experiment was then repeated on cell extracts prepared from MCF7 and BT474 cells treated with compound 10. In both cases a peak of fluorescence was recovered at 49 minutes that co-elutes with Hsp90.

MCF cells were heated to 42° C. or maintained at 37° C. for 60 minutes. Compound 10 was added (10 µM) and after 60 minutes the cells harvested. Extracts were prepared and

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separated by micro anion-exchange chromatography. Column fractions were analyzed for the presence of compound 10. FIG. 13B shows that the major peak of fluorescence correlates with elution of Hsp90 and that heat treatment induces activation of the protein as reflected by increased uptake of compound 10.

Collectively these data demonstrate that compound 10 binds to Hsp90 in tumor cells and is not associated with any other cellular protein, and that the probe can be recovered bound to the protein from cell extracts.

Example 19. Imaging of HSP90 Expression in Mouse Xenografts by Fluorescence Imaging

Mice with MDA MB 468 xenografts were injected with either 0.1, 0.5 or 1 mg of compound 10 intravenously. The animals were then immediately imaged in real time in an IVIS Kinetic imager (Caliper Life Science) for the presence of fluorescence at 465ex/520em. After two hours the tumors were excised and examined for the presence of compound 10.

FIG. 14A shows rapid accumulation of compound 10 in the xenograft following tail vein injection. Red image (false, 1 vs. 2) clearly demarks the tumor over background signal (appearing very dark gray in FIG. 14A). Green image (actual, 3 vs. 4) shows clear demarcation of the tumor over and above the natural background fluorescence of the animal (with the tumor appearing bright white and background fluorescence appearing gray).

FIG. 14B shows quantitative analysis of compound 10 accumulation in tumors isolated from mice treated with the indicated doses of the compound. FIG. 14B shows the probe accumulates selectively within the tumor mass in a dose dependent manner. Measurements were made in the IVIS imager or by preparing cells extracts from the tumors and measuring their fluorescence in a microtitre plate reader.

FIG. 14C shows accumulation of compound 12 within tumors isolated from mice injected with the indicated concentrations of the compound. FIG. 14C shows the probe accumulates selectively within the tumor mass in a dose dependent manner. Measurements were made in the IVIS imager or by preparing cells extracts from the tumors and measuring their fluorescence in a microtitre plate reader.

Example 20. In Vivo Detection of Compound 10 with Optical Spectroscopy

MDA-MB-468 tumor cells (1 M cells/mouse) were injected to the flank of SCID mice. Three weeks later, when tumor size reached 5-8 mm in diameter, the experiment was performed. Baseline autofluorescence levels were measured for tumor and normal tissues (skin adjacent to tumor). 1 mg or 0.5 mg of compound 10 was injected via tail vein. After drug injection, fluorescence signals were measured with the optical spectroscopy (~30 min) Two days after injection, measurement of fluorescence signal was repeated.

FIG. 15 shows results from two separate mice injected with compound 10 (1 mg) and the presence of the compound detected tumor by optical spectroscopy at time 0 and then 3 days later. Normal: Normal skin adjacent to tumor. All data are normalized to peak fluorescence intensity to compare fluorescence line shapes. In both mice at Day 0 Normal shows higher signal than tumor after compound 10 injection (due to poor vasculature in tumor). Measurements were made within a few minutes of injection. On Day 3, normal site has declined to pre-injection baseline level whereas the tumor still retains FITC signal in both cases. These data again demonstrate accumulation of compound 10 in tumors only and not other tissues.

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Example 21. Compound 10 Recognizes Cells Infected with HIV-1

TZM-bl (Hela derived) cells were added on glass cover-slips in 24-well plates (1×10⁵ cells/well) and were allowed to attach to cover slips overnight. For infection NL4.3 virus (HIV-1) was added to medium to achieve 10 ng/mL P24 protein concentration along with DEAE dextran at a final concentration 15 µg/mL. After 48 hours of infection, compound 10 was added to the medium and incubated for 1 hour at 37° C. Later the infected cells were washed twice with phosphate-buffered saline (PBS) and fixed for 20 min in 4% paraformaldehyde. They were washed twice with PBS and permeabilized for 30 min in 50% ice cold methanol on ice. Blocking was carried out using 1% FBS for 30 minutes at room temperature after two washes with PBS. Cells were then incubated for 2 hours at room temperature with antibody against HIV-1 capsid protein P24 at 1:100 dilutions in PBS. The antibody was obtained from the AIDS Reference and Reagent Program (HIV-1 p24 Monoclonal Antibody-183-H12-5C, Catalogue number: 3537). The cells were washed three times in PBS before and after 1 h of incubation with anti-mouse Texas Red secondary antibody (1:250). (Texas Red® Goat Anti-Mouse IgG (H+L), catalogue number: T682, Life technologies). Cover slips were mounted on the slides using Prolong gold with DAPI (ProLong® Gold Antifade Reagent with DAPI Catalog Number P36935, life technologies) and observed by confocal microscopy (Fluoview FV10i Olympus).

As shown in FIG. 16, panel A is an overlay of B and C; panel B shows the fluorescence from compound 10 (where green staining appears white or light gray); panel C shows fluorescence from the antibody to SP4 viral capsid protein (where red staining appears white or light gray); D shows an optical image of the entire field, showing that the probe and antibody do not recognize uninfected cells. These data demonstrate that compound 10 can be used to detect cells infected with HIV.

Example 22. Recovery of Drug-Bound Protein from Mice

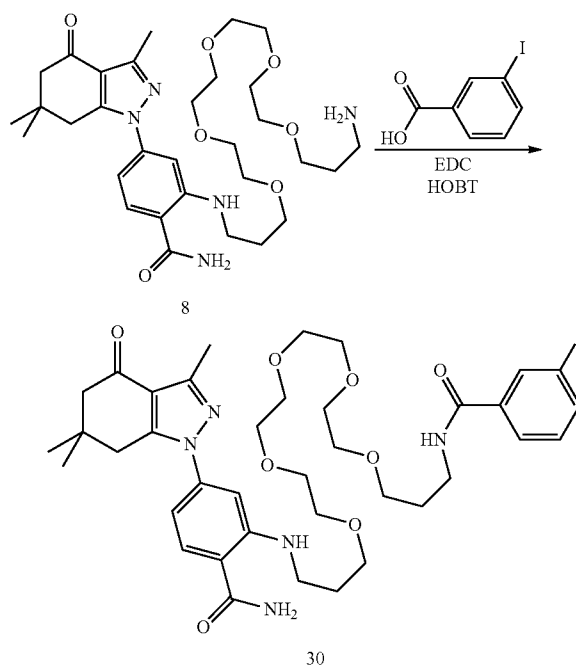
Mice were injected (IV) with 1 mg of compound 10 and the tumors were excised from the animal after 2 hours. Tissue extracts were prepared and characterized by micro anion-exchange chromatography. Column fractions were analyzed for the presence of compound 10 by fluorescence and mass spectrometry. Results are shown in FIG. 17. The major peak of fluorescence (fractions 49-53) correlates with elution of full length Hsp90. The two minor peaks (fractions

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27-35) were attributable to binding to degraded forms of the N terminal domain of Hsp90.

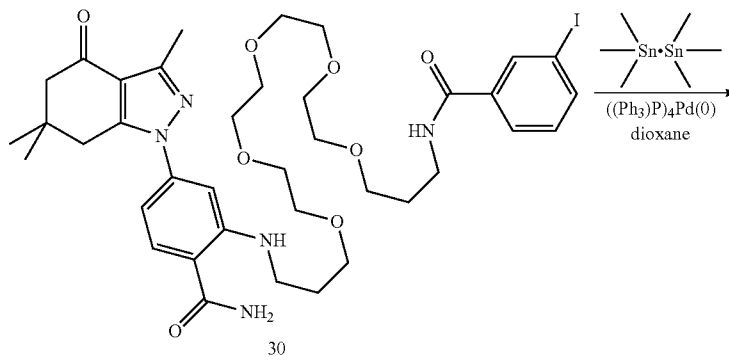
Example 23. Synthesis of Iodine-Containing Compounds

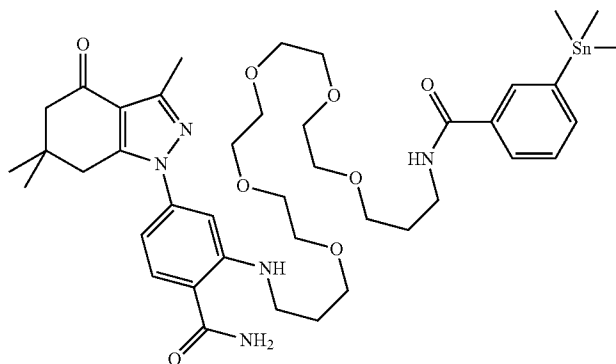
N-(19-((2-carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-4,7,10,13,16-pentaoxonadecyl)-3-iodobenzamide (30)



Amine (8) (165 mg, 273 µmol), 3-iodobenzoic acid (68 mg, 273 µmol), EDC (79 mg, 410 µmol) and HOBT (37 mg, 227 µmol) and were dissolved in methylene chloride (4 mL) and stirred at room temperature for 1 day. The reaction mixture was concentrated then dissolved in DMSO (1.5 mL) and purified by prep HPLC (30 to 100% methanol, 20 mL/m, Agilent C-18, 21.1×25 cm) to give 30 (155.7 mg, 68%) as a clear glass. MS (ESI): *m/z* 834.3 [M+H]⁺.

N-(19-((2-carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-4,7,10,13,16-pentaoxonadecyl)-3-(trimethylstannyl)benzamide (31)

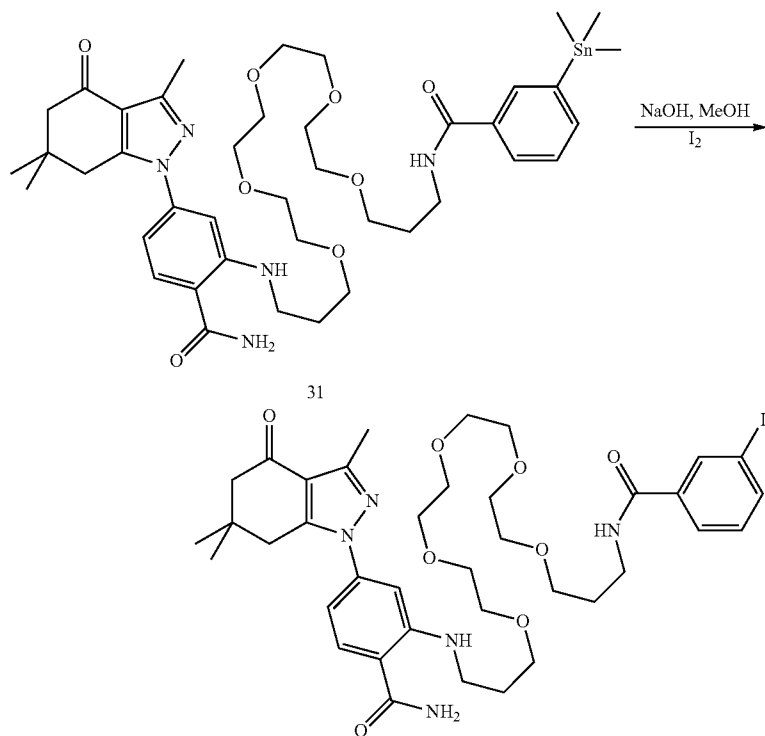




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From *J. Org. Chem.* 2012, 77, 1931-1938 and *Appl. Radiat. Isot.* Vol. 49, No. 8, pp. 955-959, 1998. Iodide (30) (92.7 mg, 111 μmol), hexamethylditin (40 mg, 122 μmol) and tetrakis triphenylphosphine palladium(0) (2.6 mg 2.22 μmol) were slurried in dioxane (2 mL), purged with nitrogen and heated to 100° C. for 1 h then 80° C. for 16 h. The reaction mixture was concentrated and chromatographed (silica gel, 2.5×25, CH_2Cl_2 (100 mL), $\text{CH}_2\text{Cl}_2/\text{MeOH}$: 19/1 (250 mL), $\text{CH}_2\text{Cl}_2/\text{MeOH}$: 9/1 (250 mL)) to give 31 (92 mg, 95%) as a slightly yellowish oil. The product was further purified by prep HPLC (30 to 100% methanol, 20 mL/m, Agilent C-18, 21.1×25 cm) to give a clear glass. MS (ESI): base peak m/z 872.4 $[\text{M}+\text{H}]^+$. Development of Method for Iodine Incorporation from I_2

20 Tin compound 31 (15.4 mg, 17.7 μmol) was dissolved in methanol (1 mL) and treated with 1N NaOH (53 μL of 1N solution) followed by Iodine (194 μL of 0.1 M solution). The iodine color went away almost immediately. After stirring for 20 m, an aliquot was removed for LC/MS analysis. LC/MS shows clean conversion to the iodide, about 60% there. Additional iodine (194 μL) was added to test for over iodination. After a day, LC/MS showed complete and clean conversion to the mono-iodo compound 30. MS (ESI): m/z 834.3 $[\text{M}+\text{H}]^+$.

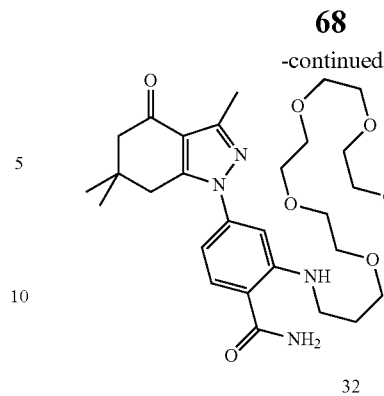
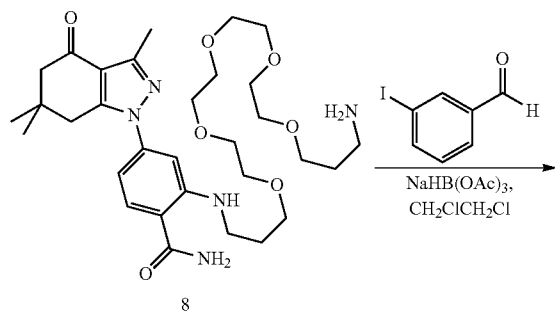


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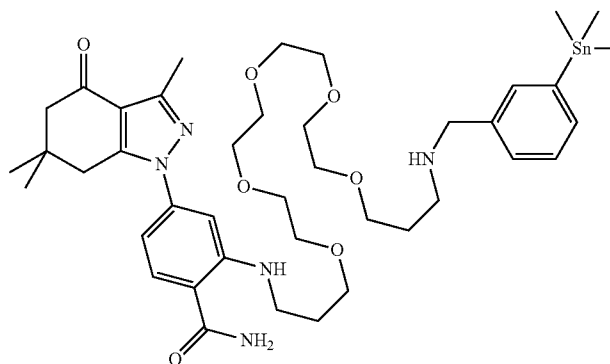
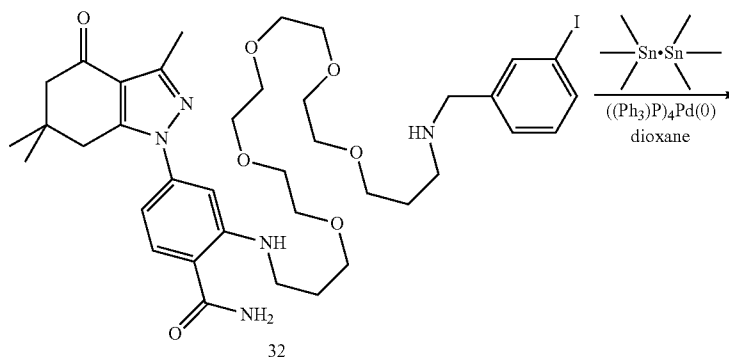
Example 24. Synthesis of Dual-Function Compounds

2-((1-(3-iodophenyl)-6,9,12,15,18-pentaoxa-2-azahenicosan-21-yl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (32)



Amine 8 (100 mg, 166 μ mol) and 3-iodobenzaldehyde (38 mg, 166 μ mol) were dissolved in dichloroethane (2 mL) and treated with sodium triacetoxyborohydride (60 mg, 282 μ mol) and stirred at room temperature for 16 h. The sample was then loaded onto a column and chromatographed (silica gel, 2.5 \times 15 cm, CH₂Cl₂ (100 mL), then 19/0.9/0.1: CH₂Cl₂/MeOH/NH₃ (200 mL), then 9/0.9/0.1: CH₂Cl₂/MeOH/NH₃ (450 mL)) to give 32 (108 mg, 63%) as a clear oil. MS (ESI): m/z 820.3 [M+H]⁺.

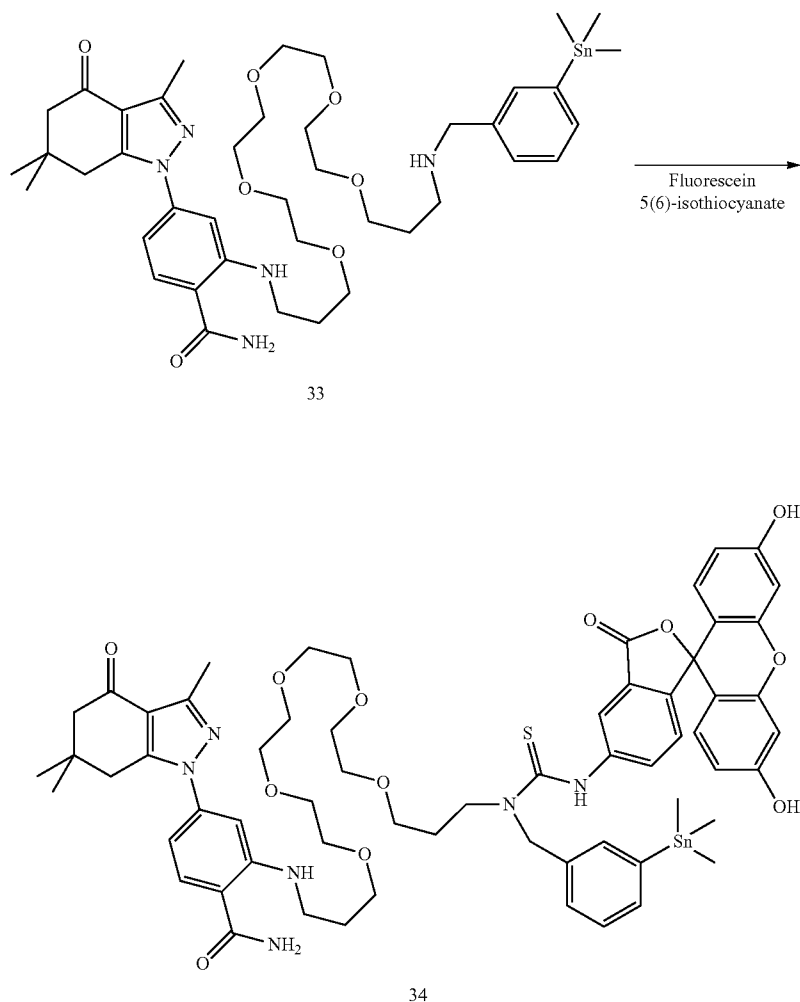
4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)-2-((1-(3-(trimethylstannyl)phenyl)-6,9,12,15,18-pentaoxa-2-azahenicosan-21-yl)amino)benzamide (33)



Iodide 32 (230 mg, 280 μmol), hexamethylditin (101 mg, 308 μmol) and tetrakis triphenylphosphine palladium(0) (6.48 mg 5.61 μmol) were slurried in dioxane (5 mL), purged with nitrogen and heated to 100° C. for 45 m. The reaction mixture was concentrated then added to a column and chromatographed (silica gel, 2.5×25, CH_2Cl_2 (100 mL), $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$: 19/0.9/0.1 (250 mL), $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$: 9/0.9/0.1 (500 mL)) to give 33 (213 mg, 89%) as a dark glass. MS (ESI): base peak m/z 858.4 $[\text{M}+\text{H}]^+$.

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2-((1-((3',6'-dihydroxy-3-oxo-3H-spiro[isobenzofuran-1,9'-xanthen]-5-yl)amino)-1-thioxo-2-(3-(trimethylstannyl)benzyl)-6,9,12,15,18-pentaoxa-2-azahenicosan-21-yl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (34)

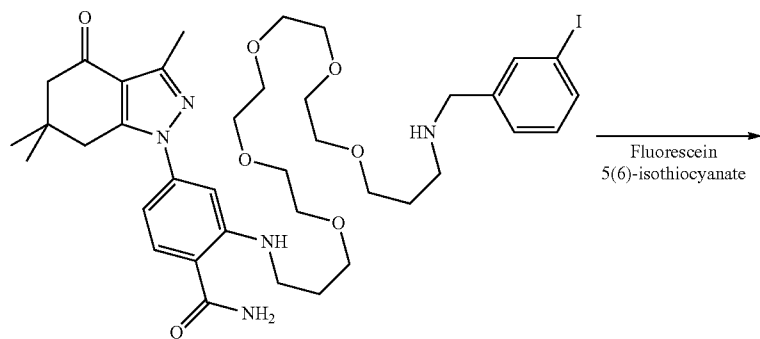


Tin compound 33 (116 mg, 135 μmol) was dissolved in DMSO (200 μL) and treated with FITC (53 mg, 135 μmol) dissolved in DMSO (200 μL) followed by Hunig's base (around 35 mg, 270 μmol). The mixture was purified by prep HPLC (30 to 100% methanol, 20 mL/m, Agilent C-18, 21.1×25 cm) to give 34 (68 mg, 40%) as a yellow solid. MS (ESI): base peak m/z 1247.4 $[\text{M}]^-$.

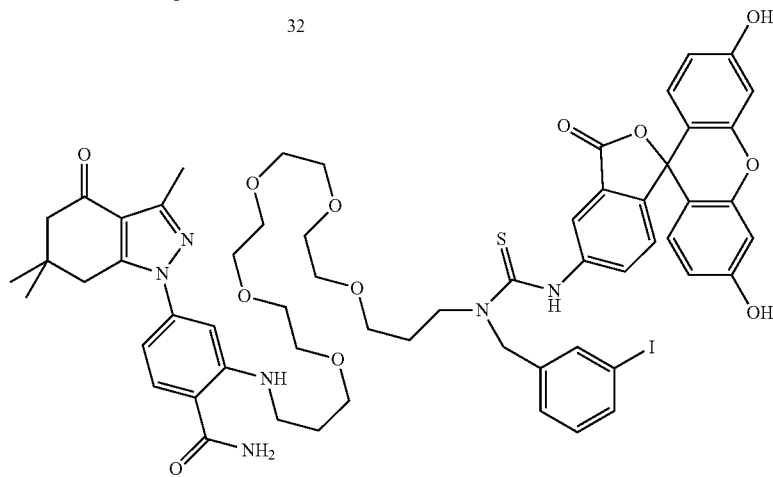
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Synthesis of Iodide Standard: 2-((1-((3',6'-dihydroxy-3-oxo-3H-spiro[isobenzofuran-1,9'-xanthen]-5-yl)amino)-2-(3-iodobenzyl)-1-thioxo-6,9,12,15,18-pentaoxa-2-azahenicosan-21-yl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (35)



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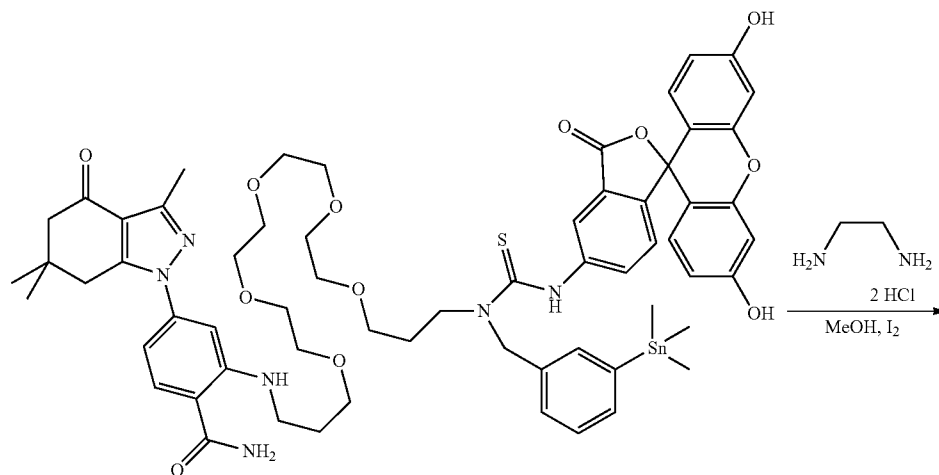


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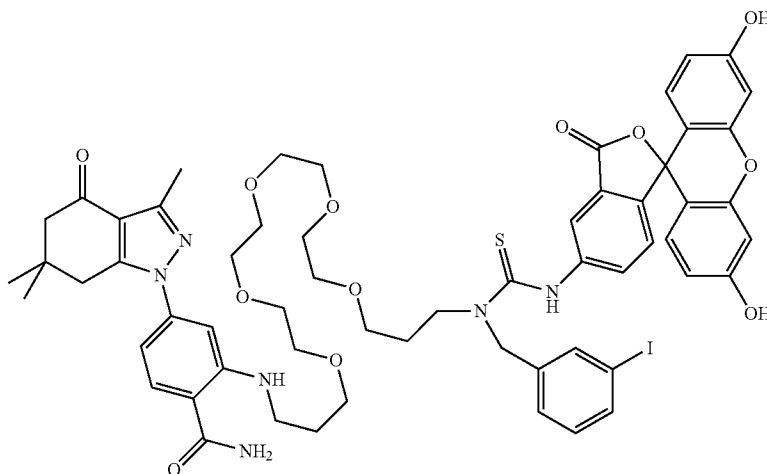
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Iodide 32 (31 mg, 38 μmol) was dissolved in DMSO (200 μL) and treated with FITC (15 mg, 38 μmol) dissolved in DMSO (200 μL) and Hunig's base (10 mg, 76 μmol) and stirred for 1 h. The sample was then loaded onto a column and chromatographed (2.5 \times 15 cm, CH_2Cl_2 (100 mL), then 19/1/0.1: $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{AcOH}$ (1500 mL), then 9/1/0.1: $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{AcOH}$ (1500 mL)) to give 35 (28 mg, 62%) as a yellow glass. MS (ESI): m/z 1207.4 $[\text{M}]^-$.

Iodination Test



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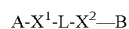
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Tin compound 34 (1 mg, 0.8 μmol) was dissolved in methanol (1 mL) and treated with ethylenediamine diHCl (1 mg in 10 μL of water) followed by iodine (8 μL of 0.1 M solution). The color dissipated instantly. After stirring for 1 day, LC/MS showed clean formation of the iodide. MS (ESI): m/z 1207.4 $[\text{M}]^-$.

The iodination can be repeated using a radiolabelled iodine source to provide a dual function compound.

The invention claimed is:

1. A compound of formula (I):



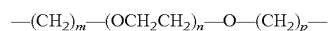
wherein:

A is a heat shock protein 90 binding component;

X^1 is selected from the group consisting of $-\text{NH}-$, $-\text{O}-$, $-\text{S}-$, $-\text{C}(\text{O})-$ and $-\text{S}(\text{O})_2-$;

X^2 is selected from the group consisting of $-\text{NR}-$, $-\text{O}-$, $-\text{S}-$, $-\text{C}(\text{O})-$ and $-\text{S}(\text{O})_2-$;

L is a divalent linker of the following formula:



wherein:

m is 2 or 3;

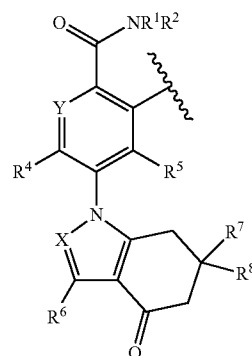
n is 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20; and

p is 2 or 3;

B is selected from the group consisting of a detection moiety, an anti-cancer agent, and a heat shock protein 90 binding component; and

R is selected from the group consisting of $-\text{H}$ and a detection moiety;

wherein each heat shock protein 90 binding component independently has the following formula (II):



wherein:

R^1 and R^2 are independently selected from the group consisting of $-\text{H}$ and $-\text{C}_{1-8}\text{-alkyl}$;

Y is CR^3 or N;

R^3 is $-\text{H}$, $-\text{F}$ or $-\text{OCH}_3$;

R^4 and R^5 are independently selected from the group consisting of $-\text{H}$, $-\text{F}$ and $-\text{OCH}_3$;

R^6 is $-\text{C}_{1-8}\text{-alkyl}$, $-\text{C}_{2-8}\text{-alkenyl}$, $-\text{C}_{2-8}\text{-alkynyl}$, $-\text{C}_{3-8}\text{-cycloalkenyl}$, $-\text{C}_{3-8}\text{-cycloalkenyl-C}_{1-8}\text{-alkyl}$, $-\text{C}_{3-8}\text{-cycloalkyl}$, $-\text{C}_{3-8}\text{-cycloalkyl-C}_{1-8}\text{-alkyl}$, aryl, aryl- $\text{C}_{1-8}\text{-alkyl}$, halo- $\text{C}_{1-8}\text{-alkyl}$, heteroaryl, heteroaryl- $\text{C}_{1-8}\text{-alkyl}$, heterocyclyl, heterocyclyl- $\text{C}_{1-8}\text{-alkyl}$, or hydroxy- $\text{C}_{1-8}\text{-alkyl}$;

R^7 and R^8 are independently selected from the group consisting of $-\text{H}$ and $-\text{C}_{1-8}\text{-alkyl}$;

X is N;

and



65 represents the point of attachment in formula (I);

wherein each detection moiety independently comprises a fluorophore or a radioactive compound;

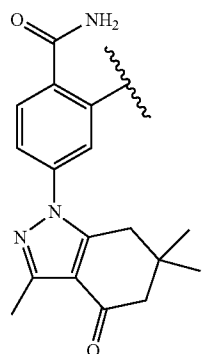
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wherein the fluorophore is a fluorescein, a rhodamine, a coumarin, a cyanine, or a boron-dipyrromethene; wherein the radioactive compound is a radioisotope; and wherein the anti-cancer agent is an alkylating agent, an anti-epidermal growth factor receptor antibody, an anti-Her-2 antibody, an antimetabolite, a vinca alkaloid, an anthracycline, a platinum-based agent, a topoisomerase inhibitor, a taxane, an anti-cancer antibiotic, an immune cell antibody, an interferon, an interleukin, a heat shock protein 90 inhibitor, an anti-androgen, an anti-estrogen, an antihypercalcemia agent, an apoptosis inducer, an aurora kinase inhibitor, a Bruton's tyrosine kinase inhibitor, a calcineurin inhibitor, a Ca^{2+} -calmodulin-dependent protein kinase II inhibitor, a CD45 tyrosine phosphatase inhibitor, a cell division cycle 25 phosphatase inhibitor, a checkpoint kinase inhibitor, a cyclooxygenase inhibitor, a cRAF kinase inhibitor, a cyclin dependent kinase inhibitor, a cysteine protease inhibitor, a deoxyribonucleic acid intercalator, a deoxyribonucleic acid strand breaker, an E3 ligase inhibitor, an epidermal growth factor pathway inhibitor, a farnesyltransferase inhibitor, a fetal liver kinase-1 inhibitor, a glycogen synthase kinase-3 inhibitor, a histone deacetylase inhibitor, an I-kappa B-alpha kinase inhibitor, an imidazotetrazinone, an insulin tyrosine kinase inhibitor, a c-Jun N-terminal kinase inhibitor, a mitogen-activated protein kinase inhibitor, a mouse double minute 2 inhibitor, an MEK inhibitor, a matrix metalloproteinase inhibitor, a mammalian target of rapamycin inhibitor, a nerve growth factor receptor tyrosine kinase inhibitor, a p38 mitogen-activated protein kinase inhibitor, a p56 tyrosine kinase inhibitor, a platelet-derived growth factor pathway inhibitor, a phosphatidylinositol 3-kinase inhibitor, a phosphatase inhibitor, a protein phosphatase inhibitor, a protein kinase C inhibitor, a protein kinase C delta kinase inhibitor, a polyamine synthesis inhibitor, a protein tyrosine phosphatase 1B inhibitor, a protein tyrosine kinase inhibitor, an SRC family tyrosine kinase inhibitor, a spleen tyrosine kinase inhibitor, a Janus tyrosine kinase inhibitor, a retinoid, a ribonucleic acid polymerase II elongation inhibitor, a serine/threonine kinase inhibitor, a sterol biosynthesis inhibitor, a vascular endothelial growth factor pathway inhibitor, alitretinon, altret-

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amine, aminopterin, aminolevulinic acid, amsacrine, asparaginase, atrasentan, bexarotene, carboquone, demecolcine, efaproxiral, elsamitrucin, etoglucid, a Gliadel implant, hydroxycarbamide, leucovorin, lonidamine, lucanthone, masoprocol, methyl aminolevulinate, mitoguazone, mitotane, oblimersen, omacetaxine, pegaspargase, porfimer sodium, prednimustine, sitimagene ceradenovec, talaporfin, temoporfin, trabectedin or verteporfin.

2. The compound of claim 1, wherein A has the formula:



3. The compound of claim 1, wherein B is a detection moiety.
4. The compound of claim 3, wherein B is a fluorophore.
5. The compound of claim 1, wherein B is an anti-cancer agent.
6. The compound of claim 1, wherein B is a heat shock protein 90 binding component.
7. The compound of claim 1, wherein:
- R^1 and R^2 are independently $-\text{H}$;
 - Y is CR^3 ;
 - R^3 is $-\text{H}$;
 - R^4 and R^5 are independently $-\text{H}$;
 - R^6 is $-\text{C}_{1-8}\text{-alkyl}$ or halo- $\text{C}_{1-8}\text{-alkyl}$; and
 - R^7 and R^8 are independently $-\text{C}_{1-8}\text{-alkyl}$.
8. The compound of claim 1, wherein m is 3 and p is 3.
9. The compound of claim 1, wherein n is 4.
10. A kit comprising a compound of claim 1.

* * * * *



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(12) **United States Patent**
Haystead et al.

(10) **Patent No.:** **US 10,112,947 B2**
(45) **Date of Patent:** **Oct. 30, 2018**

(54) **SUBSTITUTED 6-AMINOPURINES FOR
TARGETING HSP90**

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Hill, NC (US)

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(60) Provisional application No. 61/680,107, filed on Aug.
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(51) **Int. Cl.**

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C07D 231/54 (2006.01)
C07D 405/14 (2006.01)
C07D 487/04 (2006.01)

(52) **U.S. Cl.**

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(2013.01); **C07D 405/14** (2013.01); **C07D**
413/12 (2013.01); **C07D 473/24** (2013.01);
C07D 487/04 (2013.01); **C07D 493/10**
(2013.01); **G01N 33/56988** (2013.01); **G01N**
33/574 (2013.01); **G01N 33/57496** (2013.01);
G01N 2333/47 (2013.01)

(58) **Field of Classification Search**

CPC **C07D 473/34**
USPC **544/277**
See application file for complete search history.

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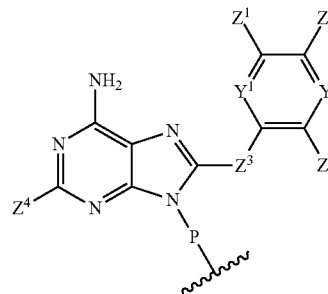
Primary Examiner — Douglas M Willis

(74) *Attorney, Agent, or Firm* — Michael Best &
Friedrich LLP

(57) **ABSTRACT**

Described herein are 6-aminopurine compounds comprising
formula (III) that may selectively bind to Hsp90, methods of
using the compounds, and kits including the compounds.
Formula (III) may link to detection moieties such as fluo-
rophores that may allow for selective detection of Hsp90 in
a sample.

(III)



(56)

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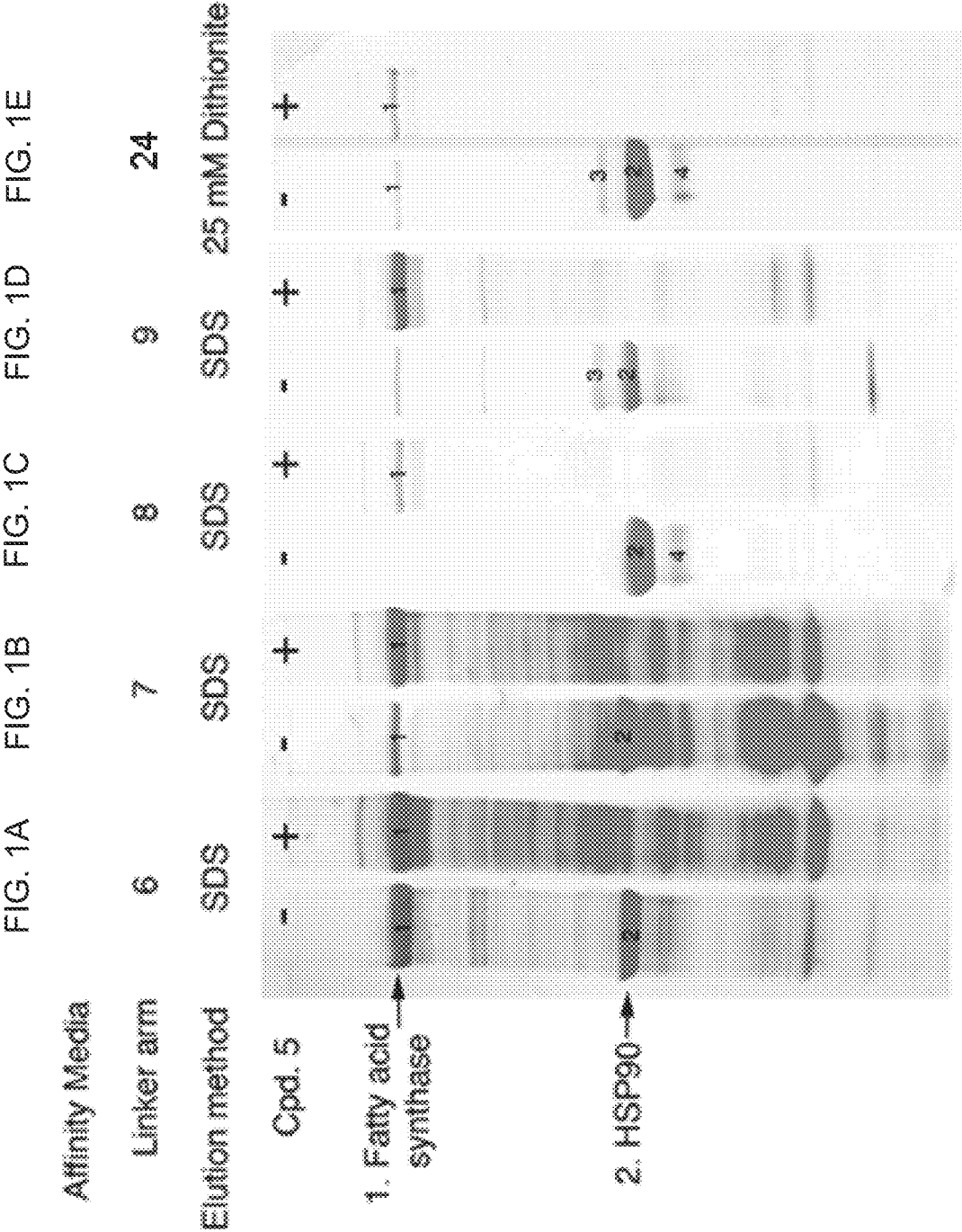
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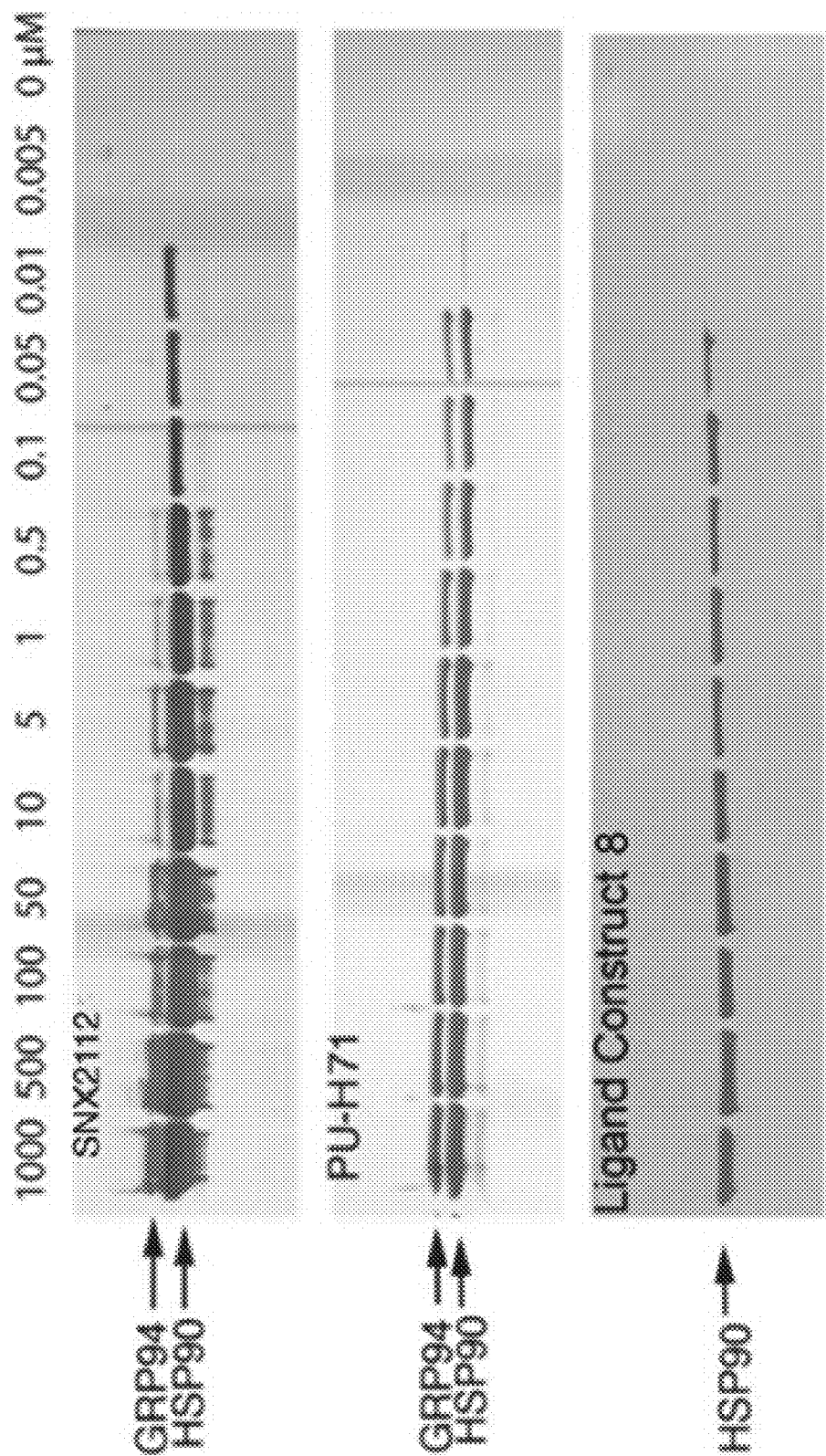


FIG. 2

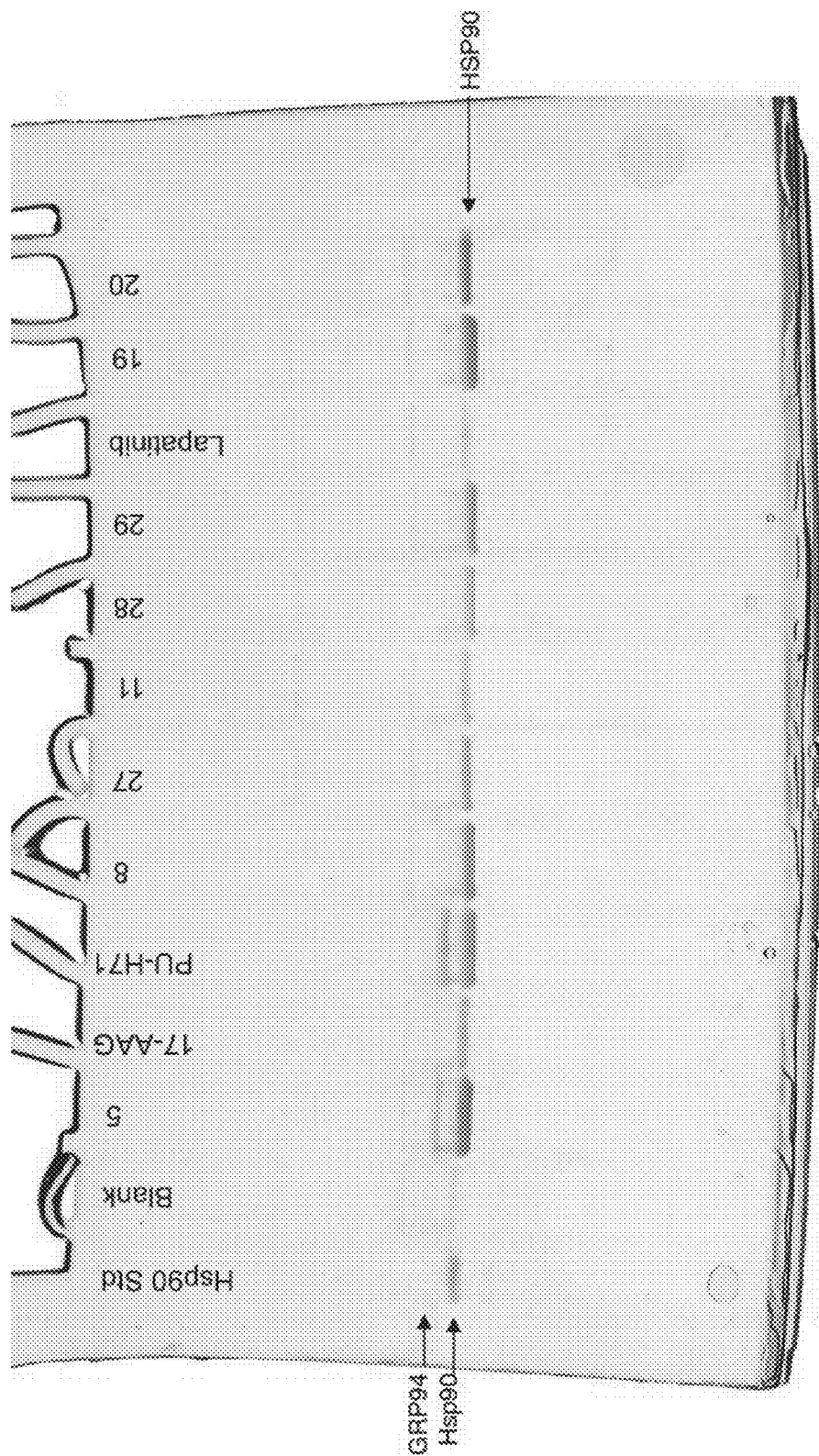


FIG. 3

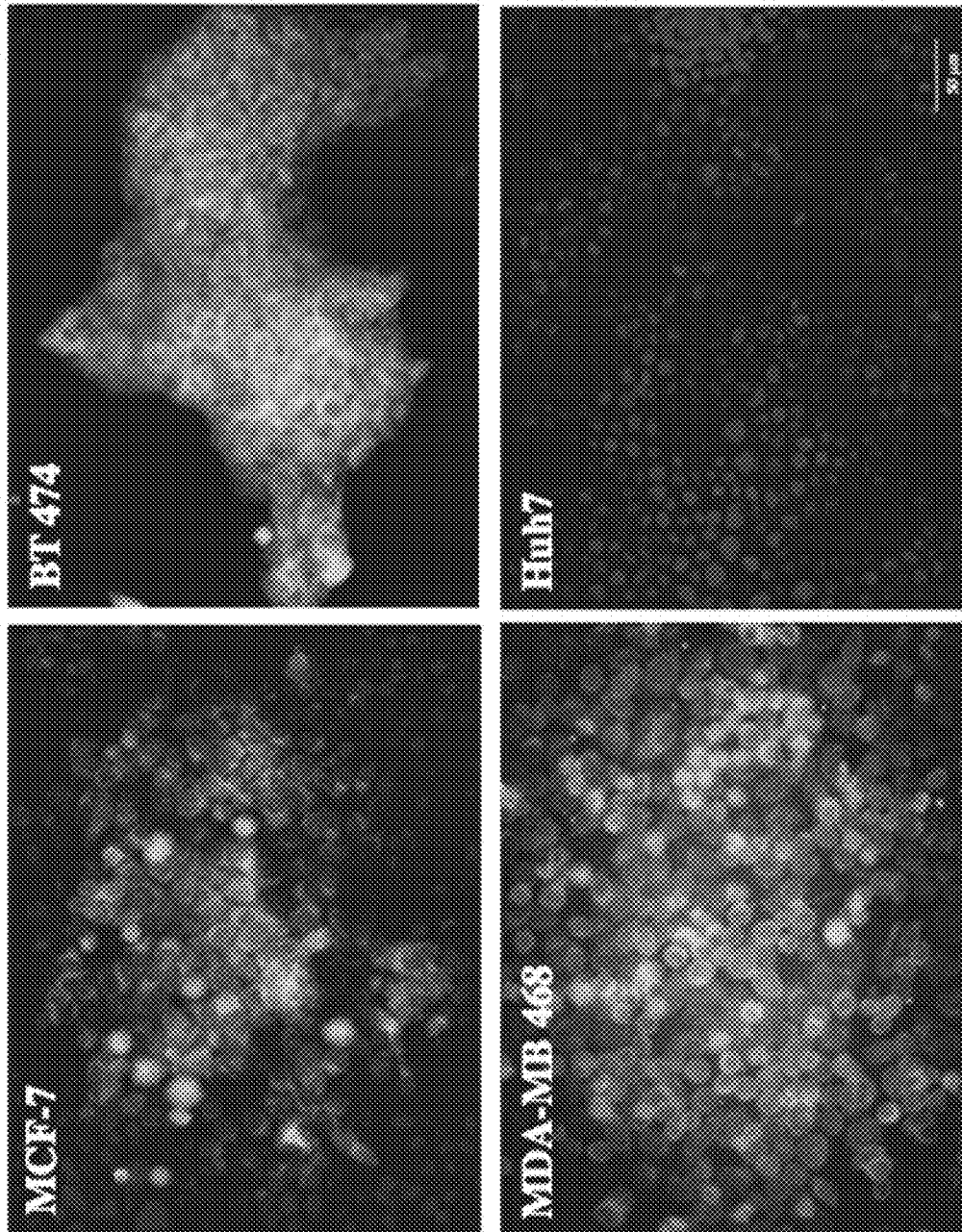
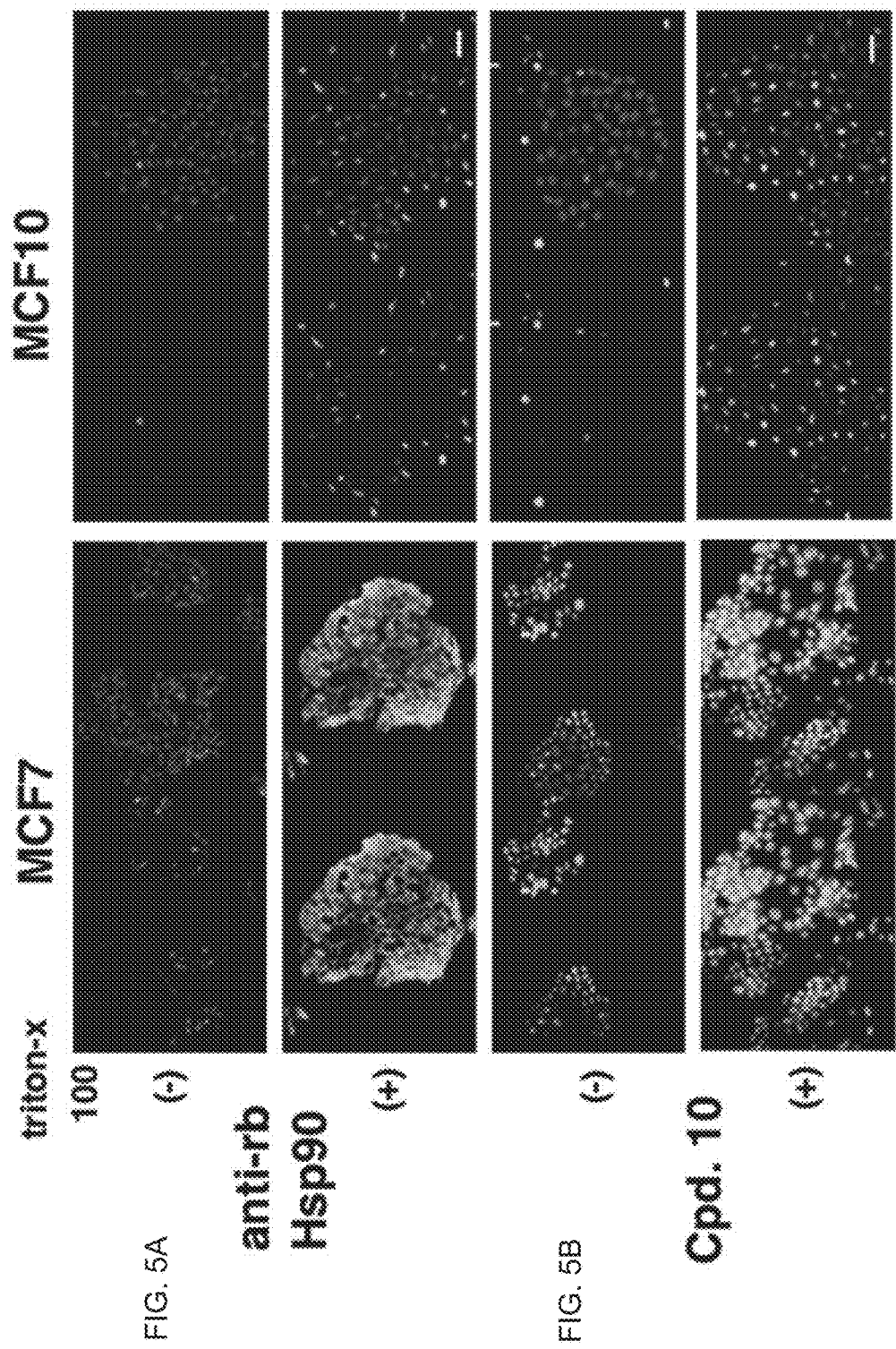


FIG. 4



Entry of Compound 10 is mediated through
surface Hsp90 in MCF7 cells

anti-Hsp90
2° AF-568

Compound 10

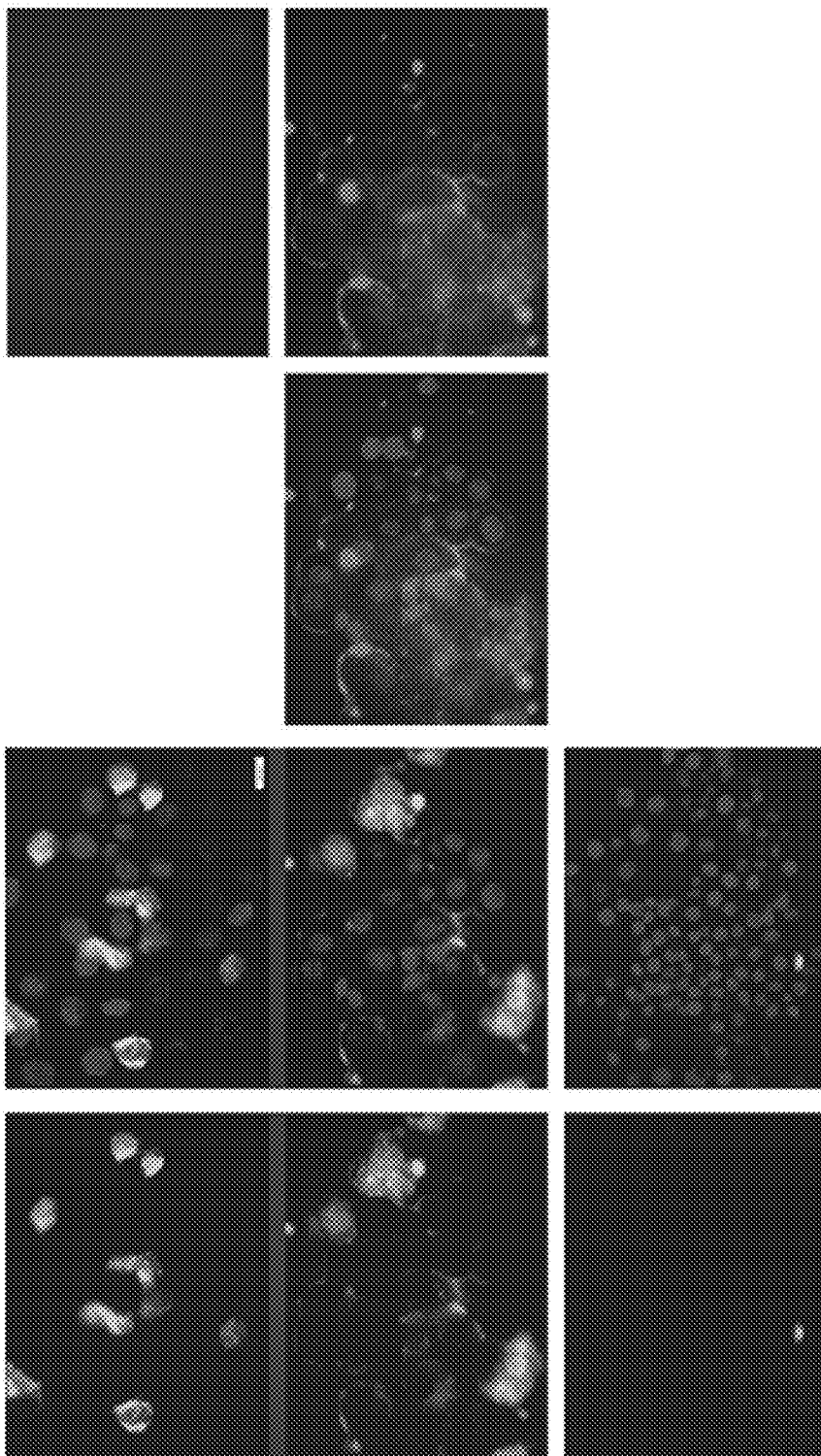
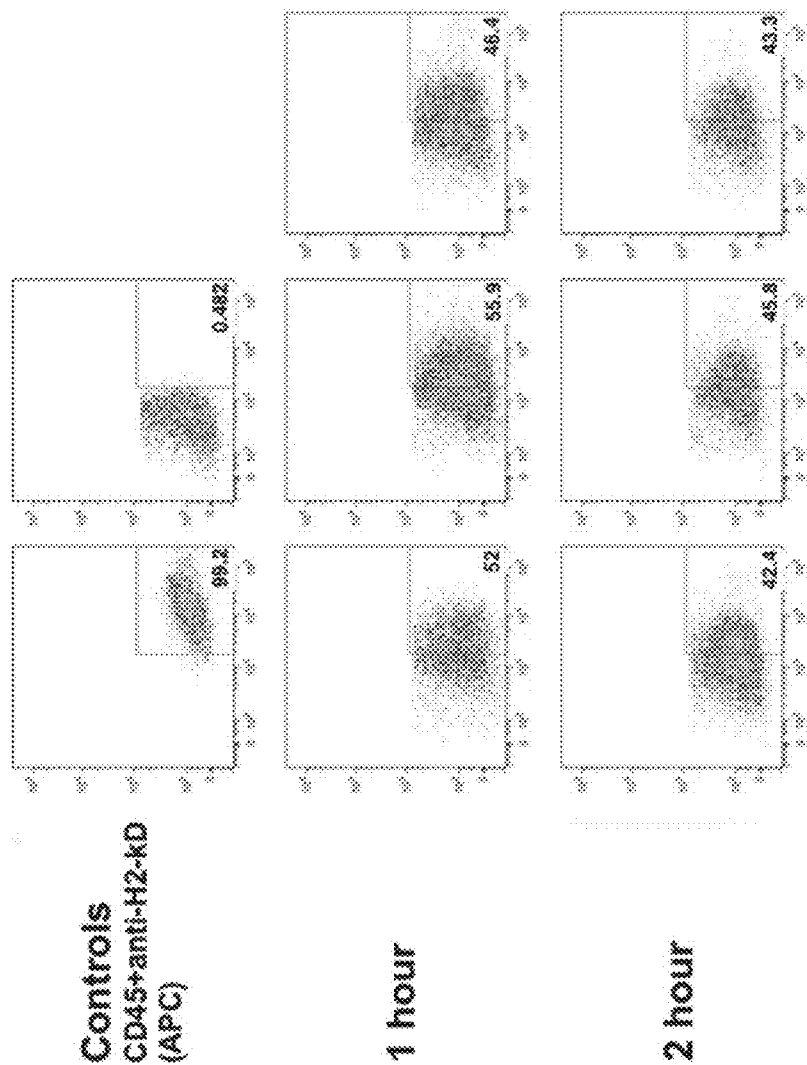


FIG. 6A

FIG. 6B

FIG. 6C

Compound 10 Labeling of MDA-MB-468 tumor cells
in NOD/SCID mice



Compound 10

FIG. 7A

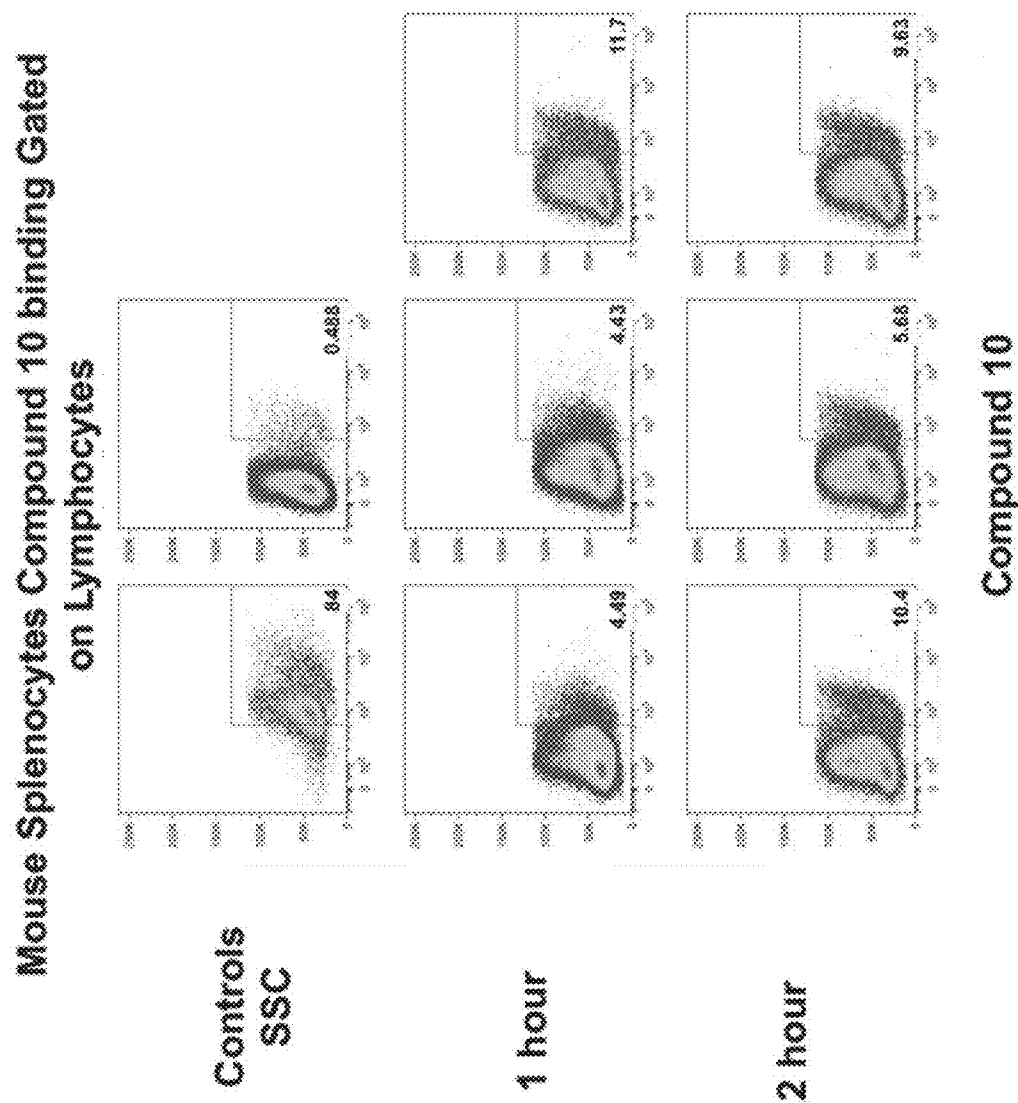


FIG. 7B

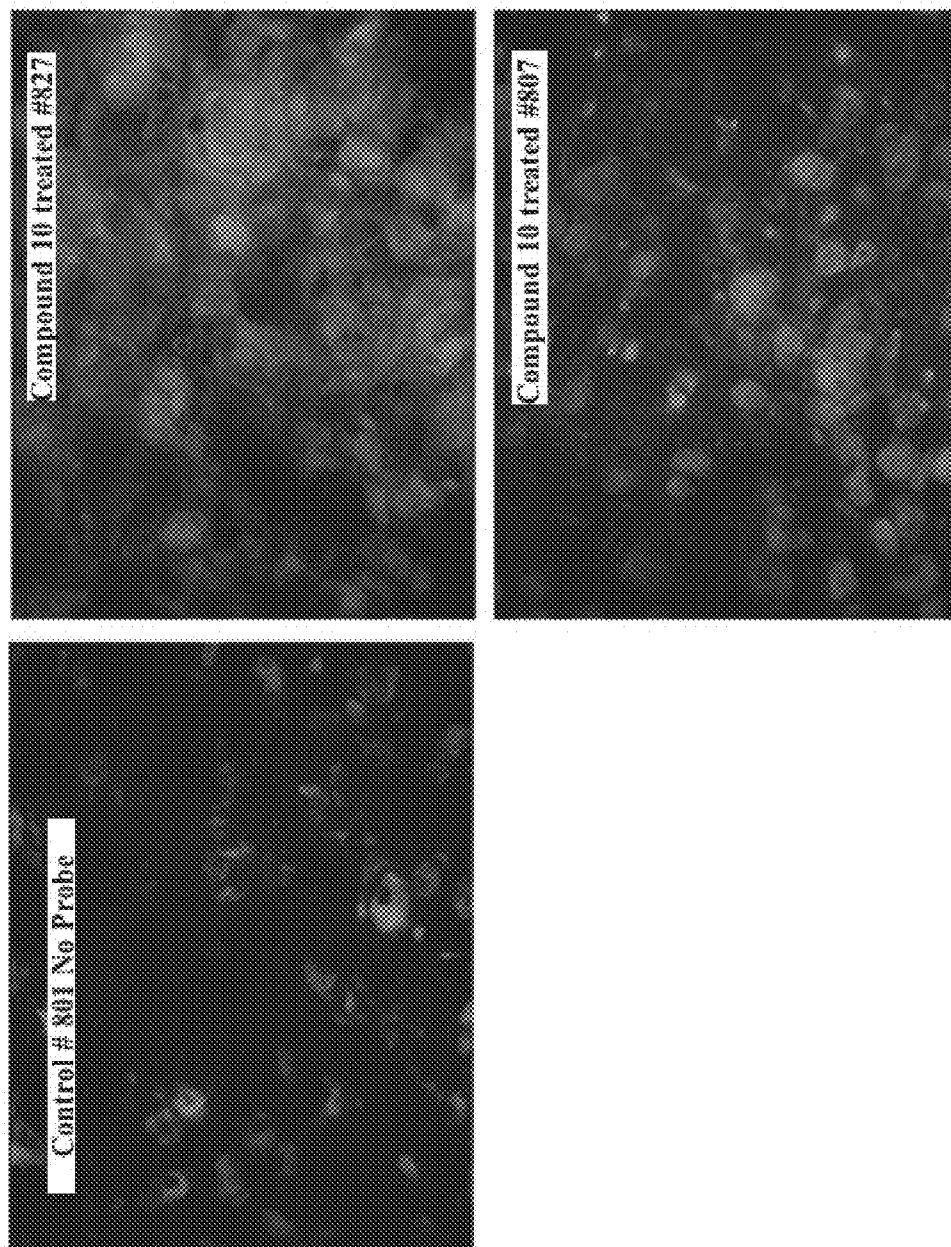


FIG. 8

Competition of Signal with Free Drug (5)

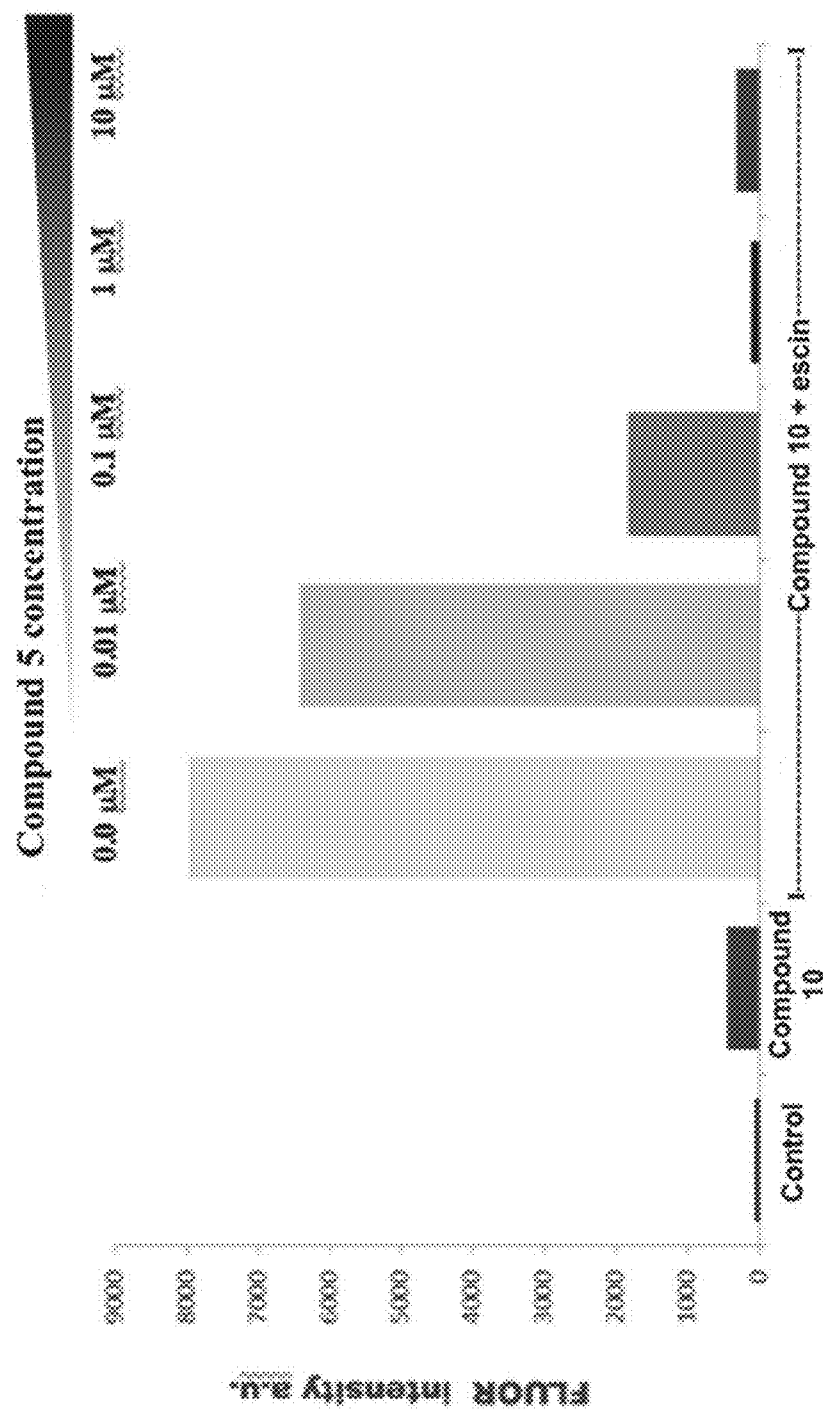


FIG. 9

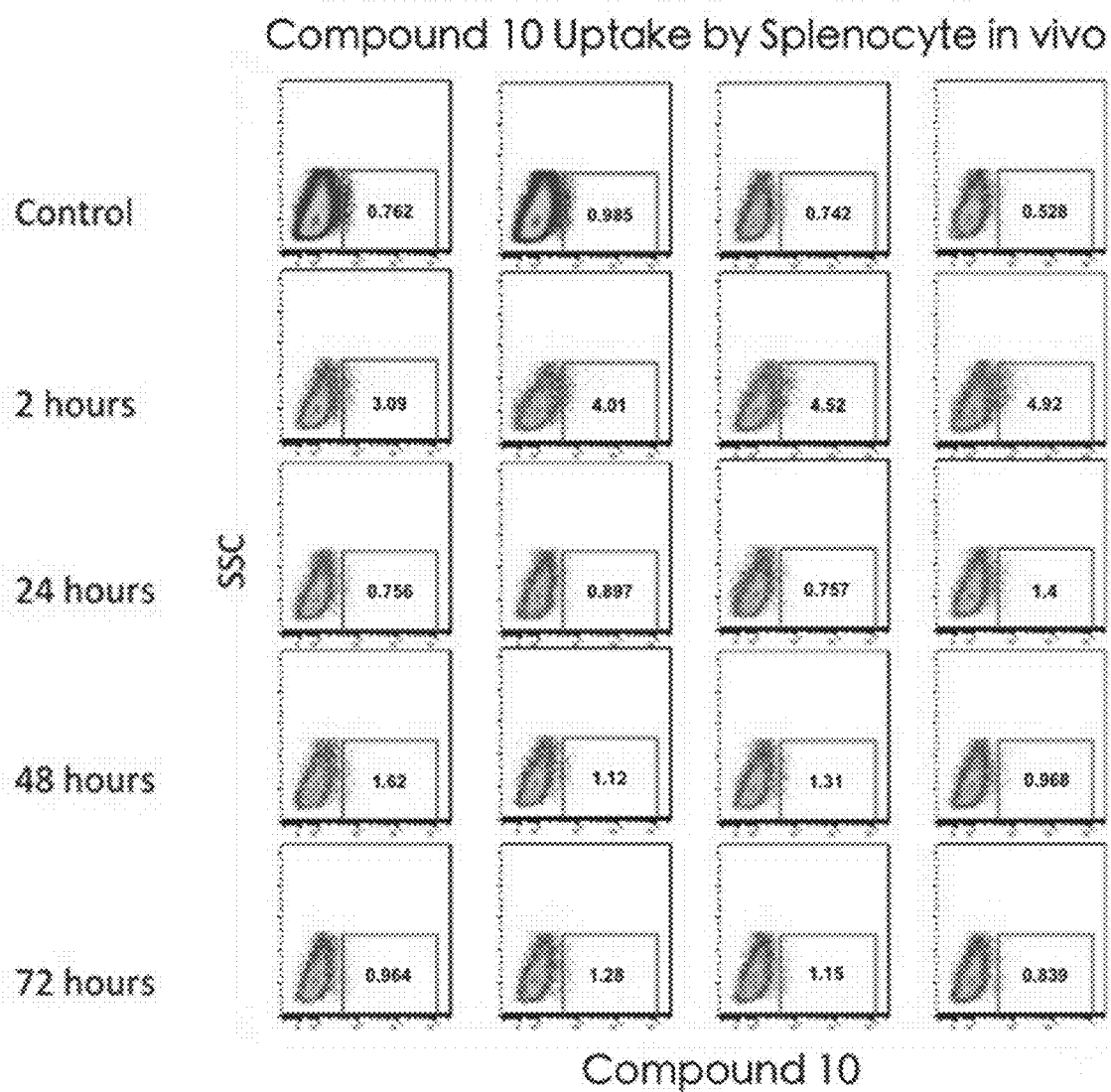


FIG. 10A

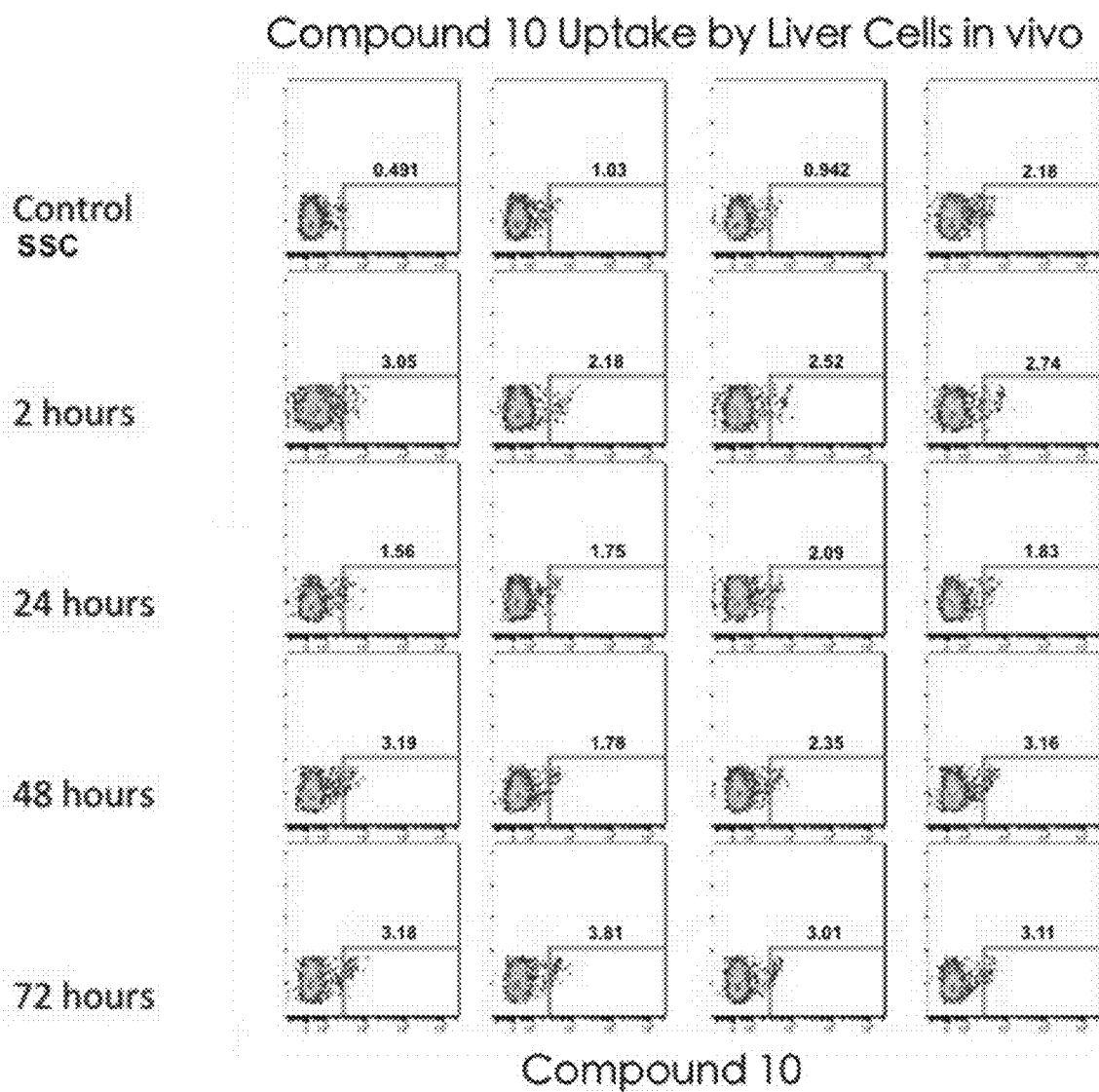


FIG. 10B

Compound 10 Uptake by Tumor Cells in vivo

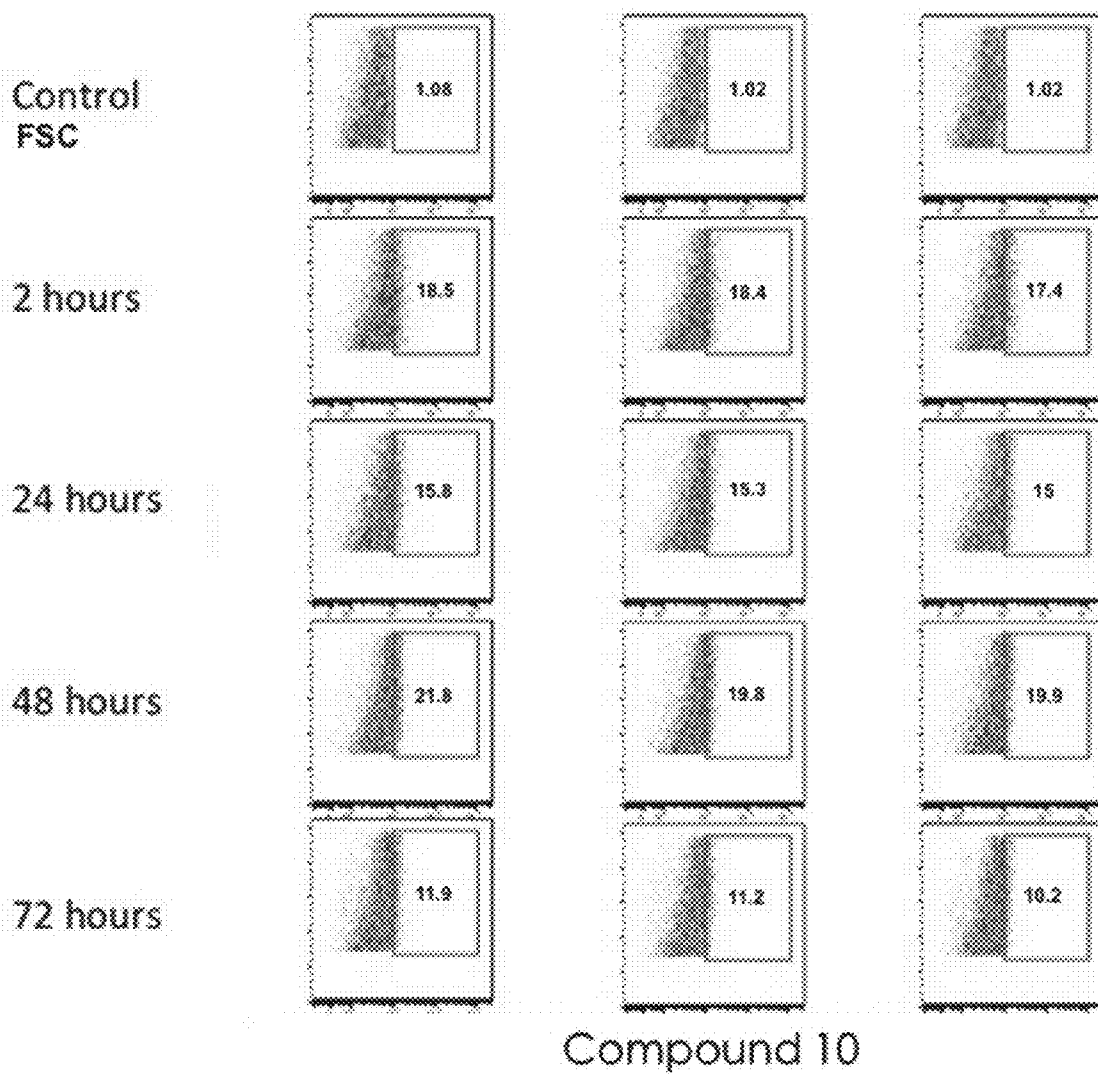


FIG. 10C

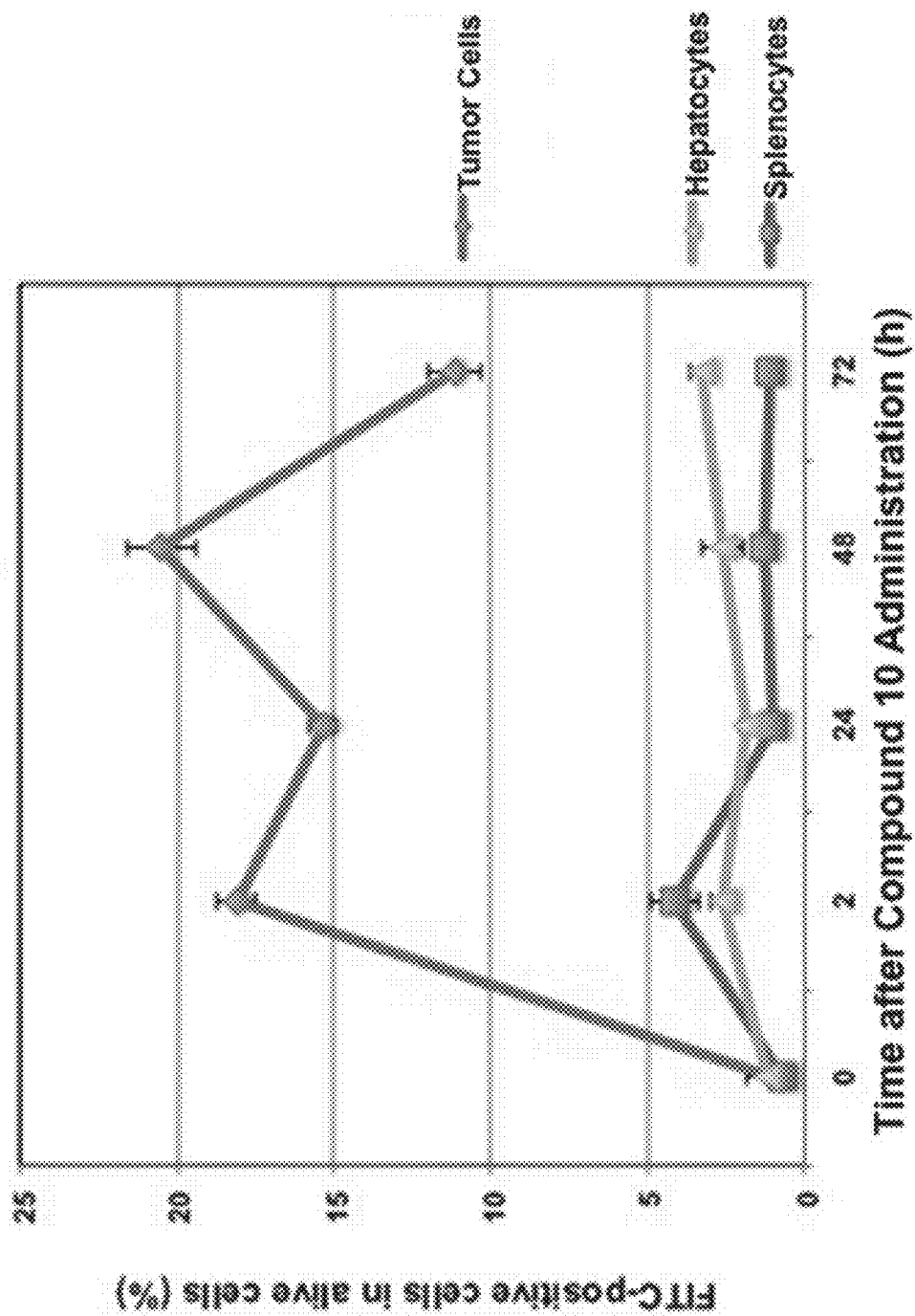


FIG. 11

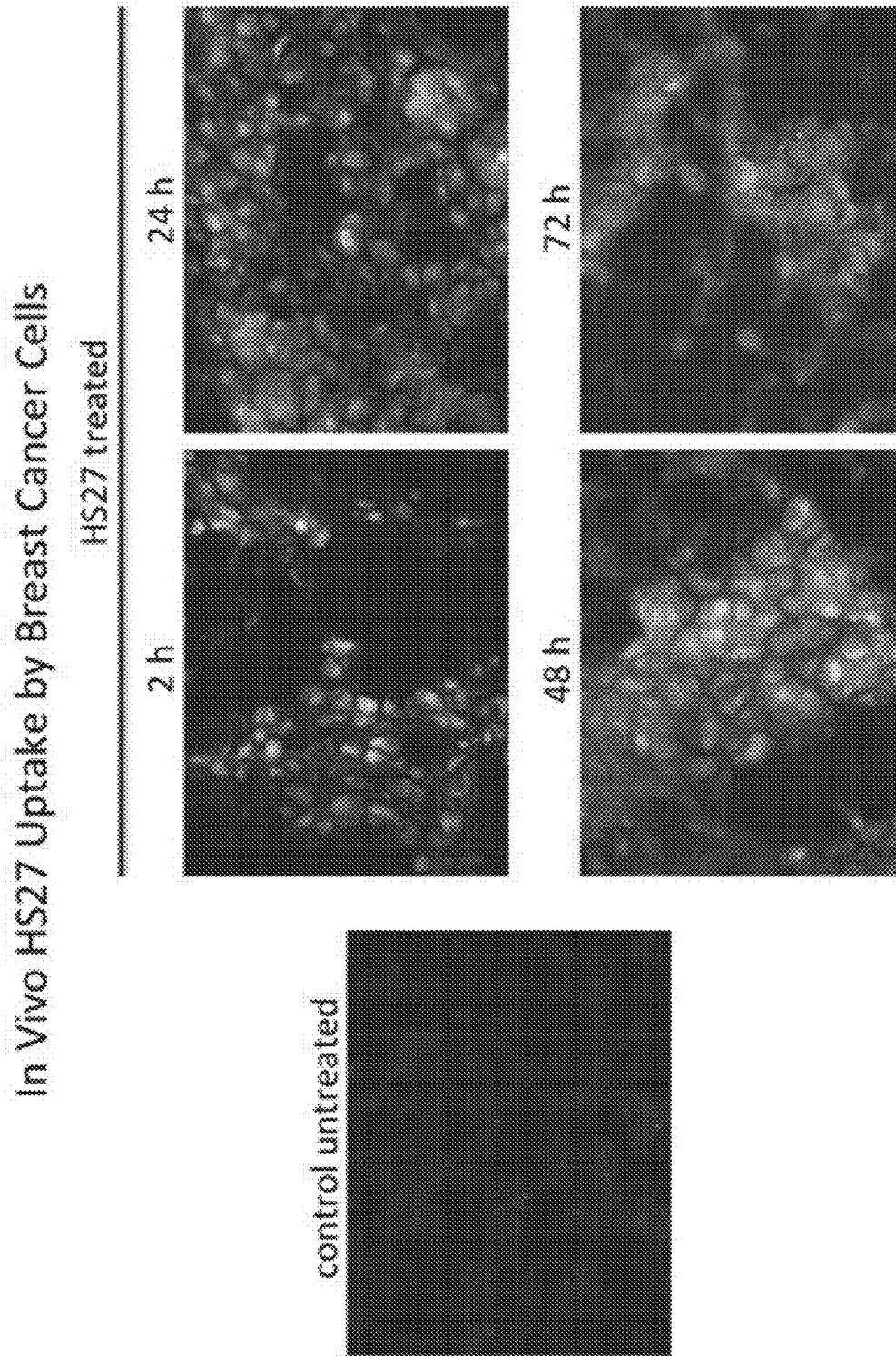


FIG. 12

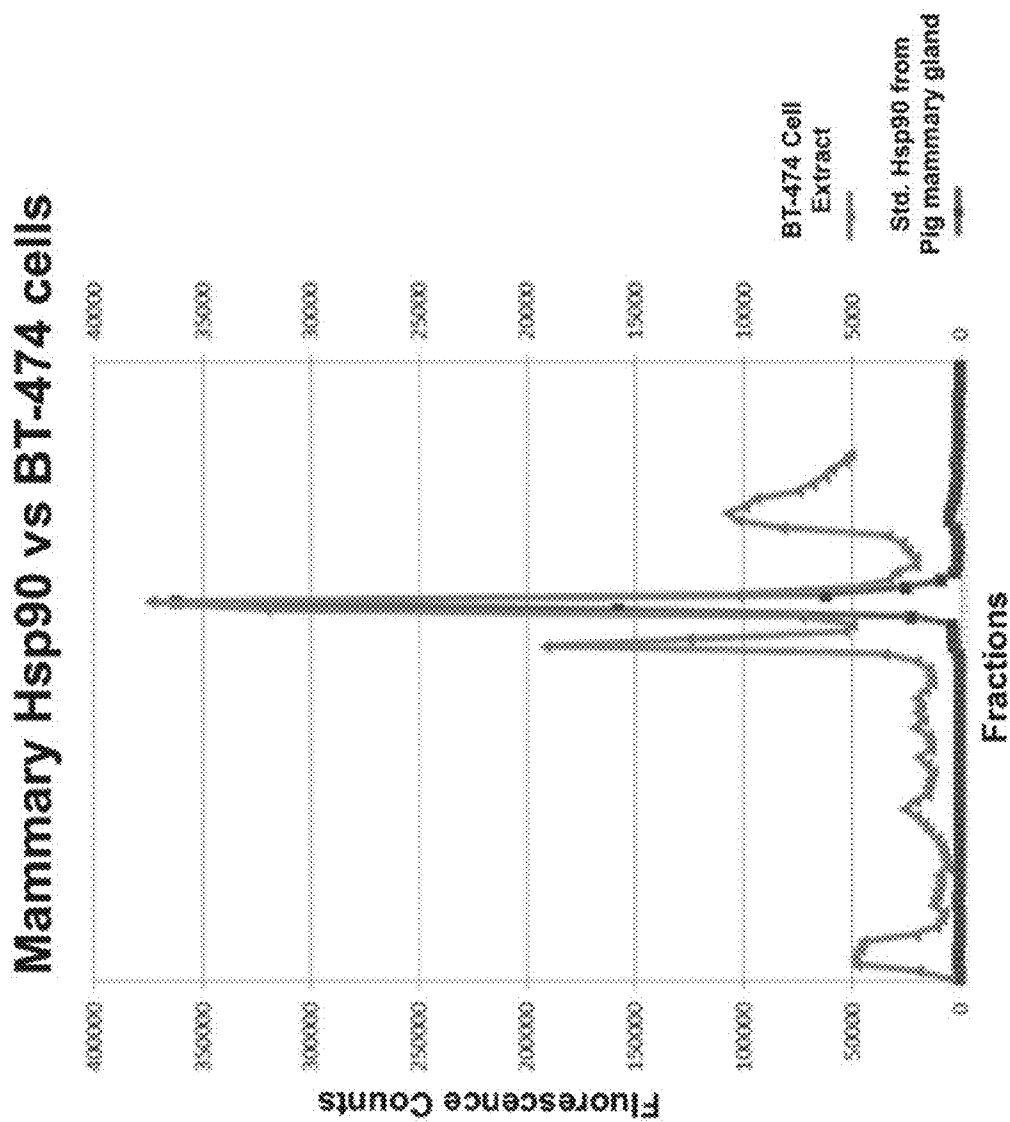


FIG. 13A

**MonoQ (anion exchange)
fractions of MCF-7 cells**

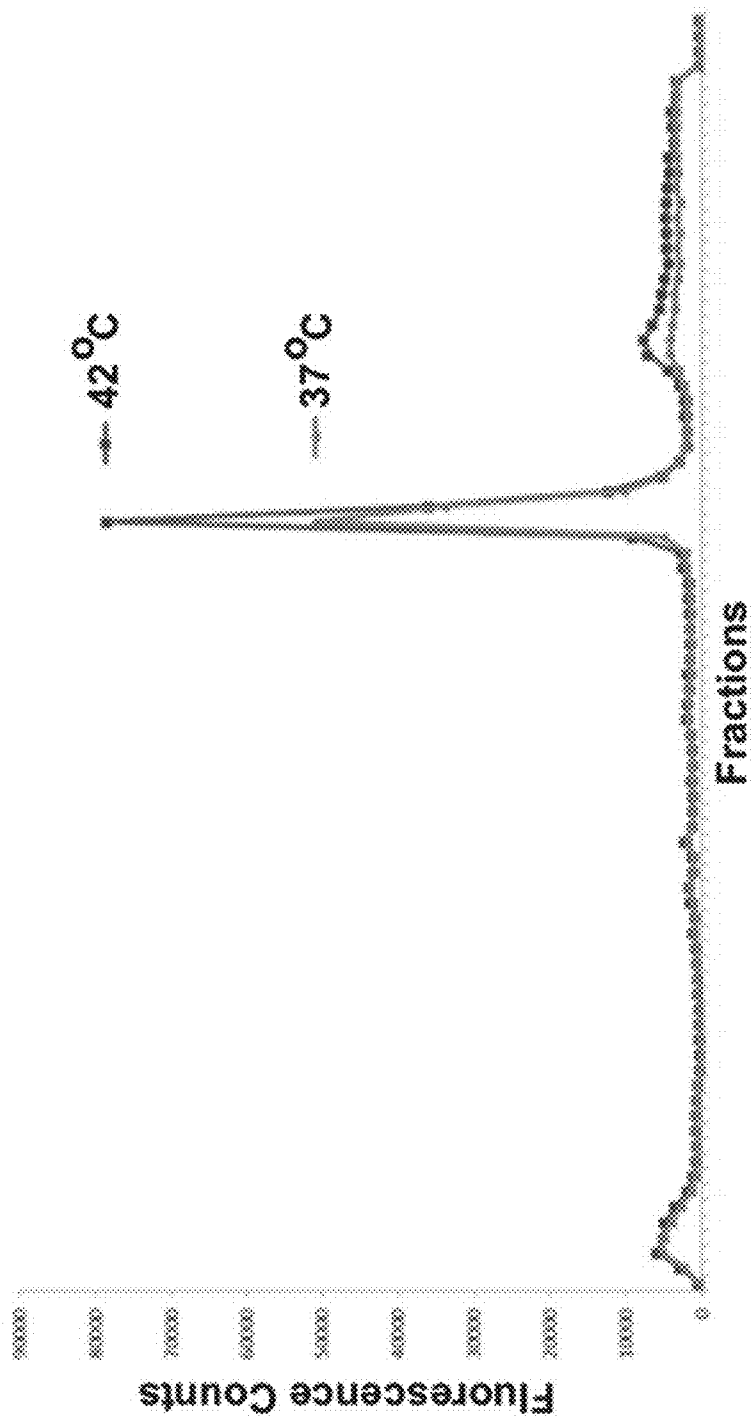


FIG. 13B

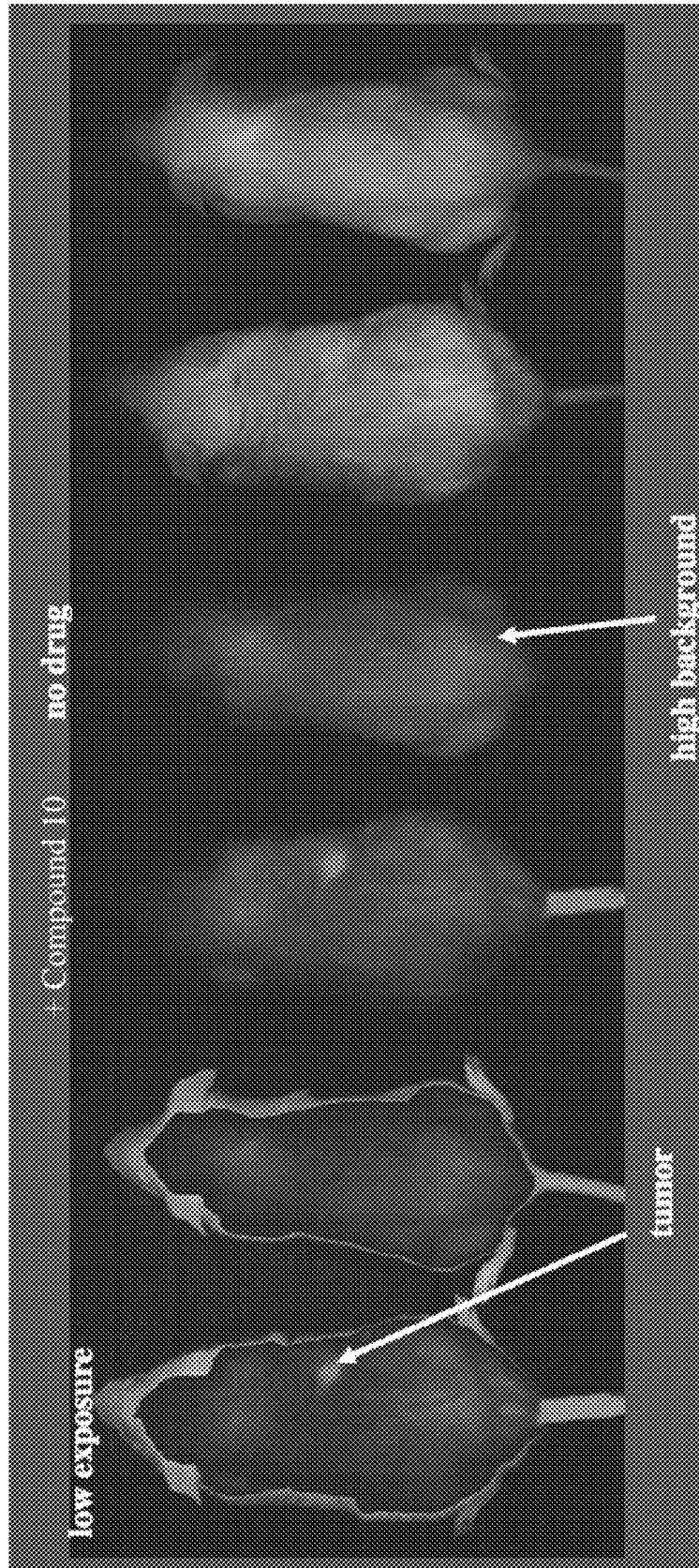


FIG. 14A

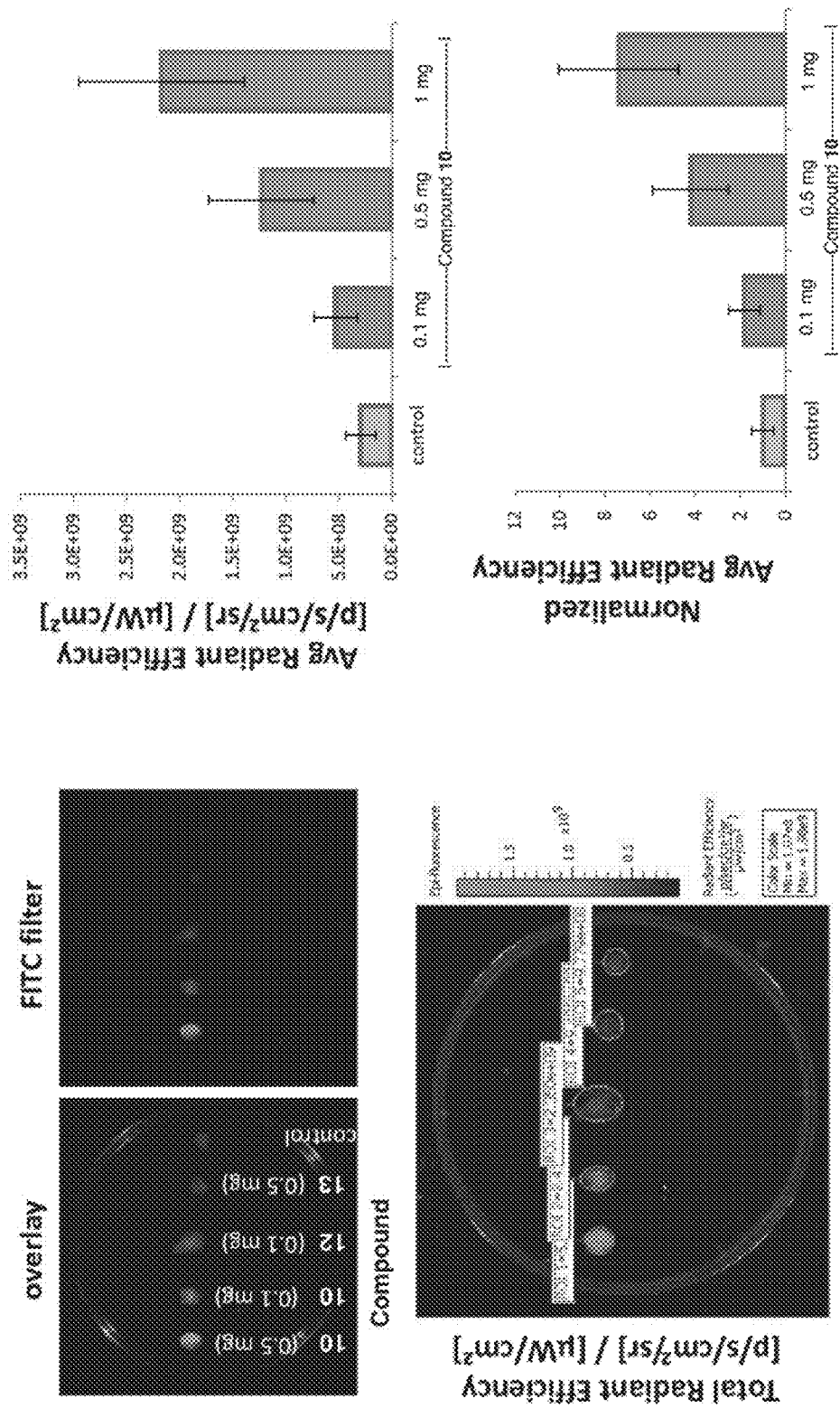


FIG. 14B

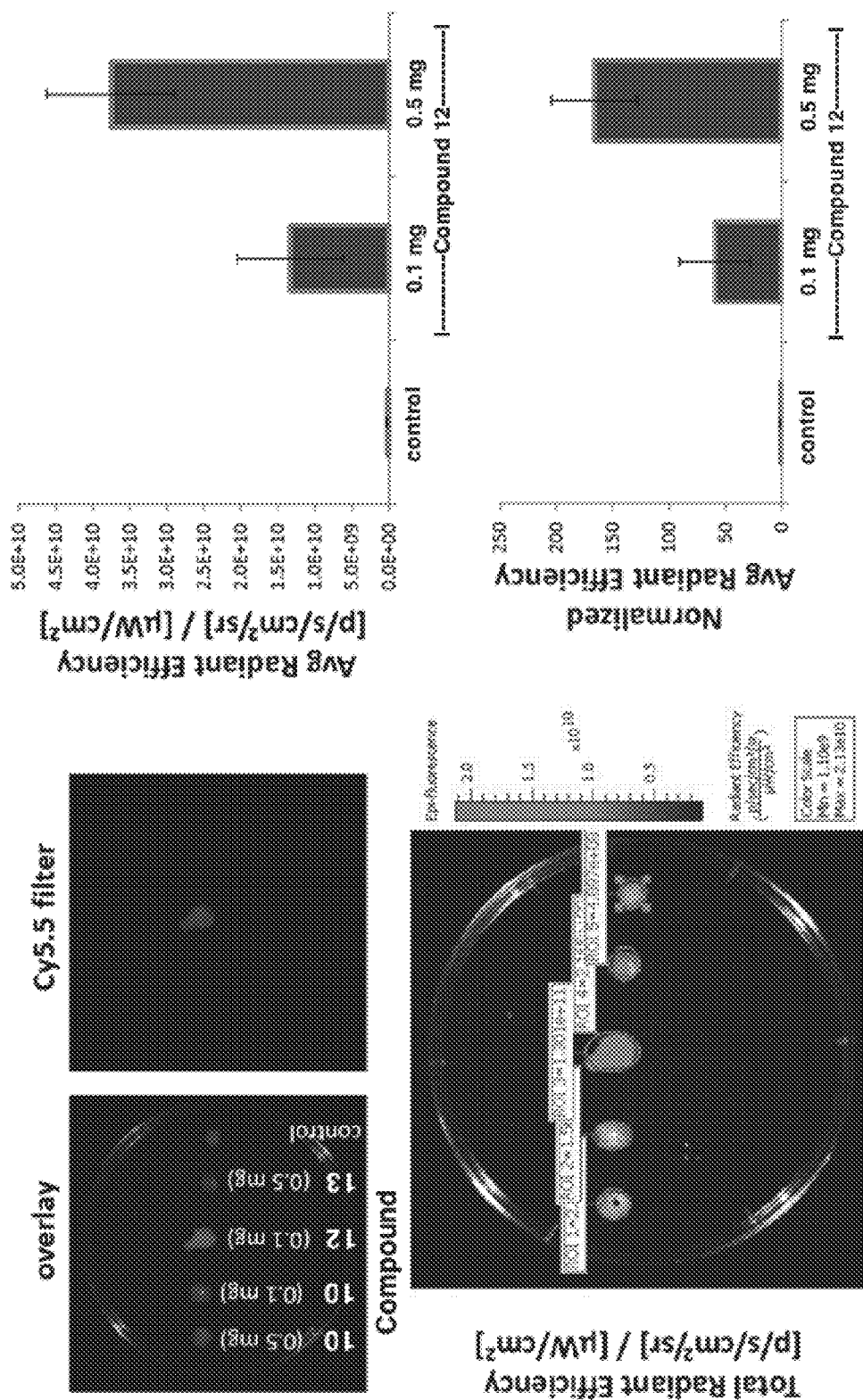


FIG. 14C

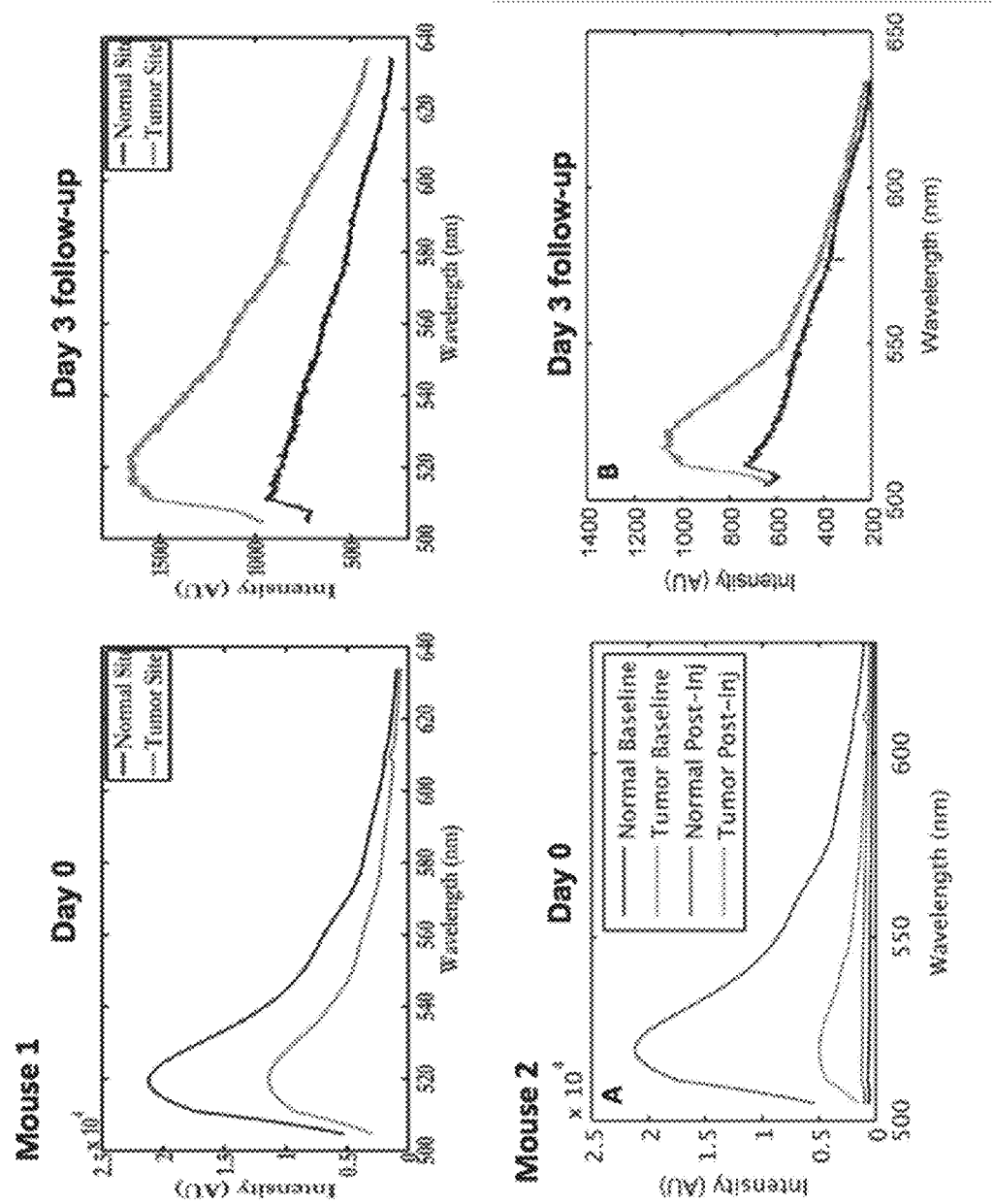


FIG. 15

FIG. 16A

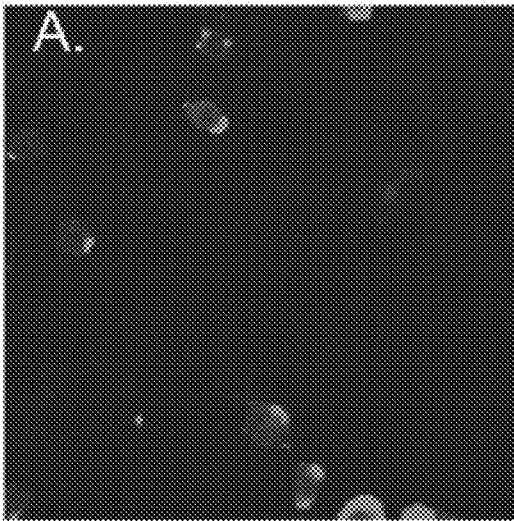


FIG. 16B

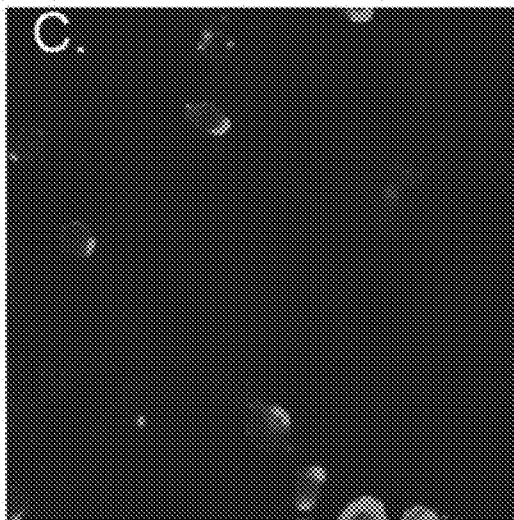
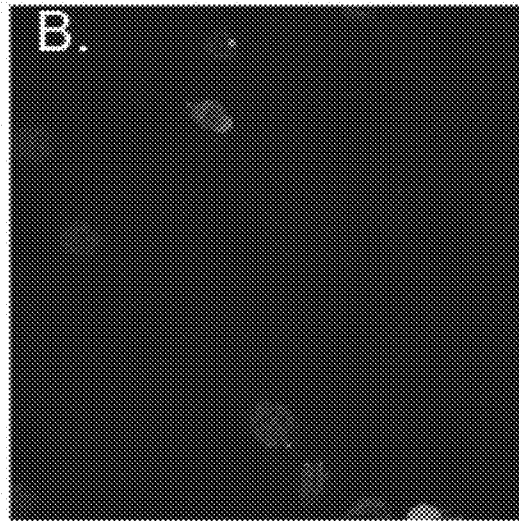


FIG. 16C

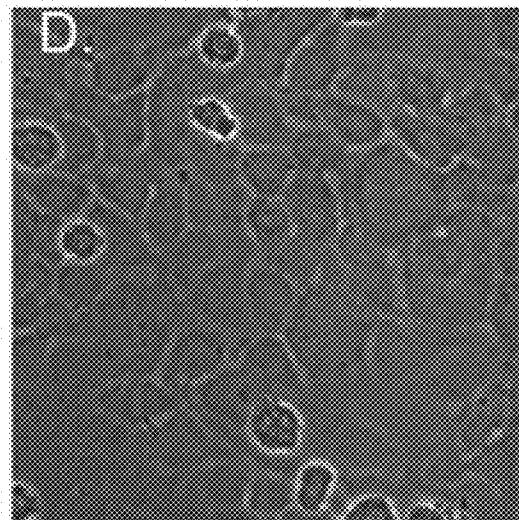


FIG. 16D

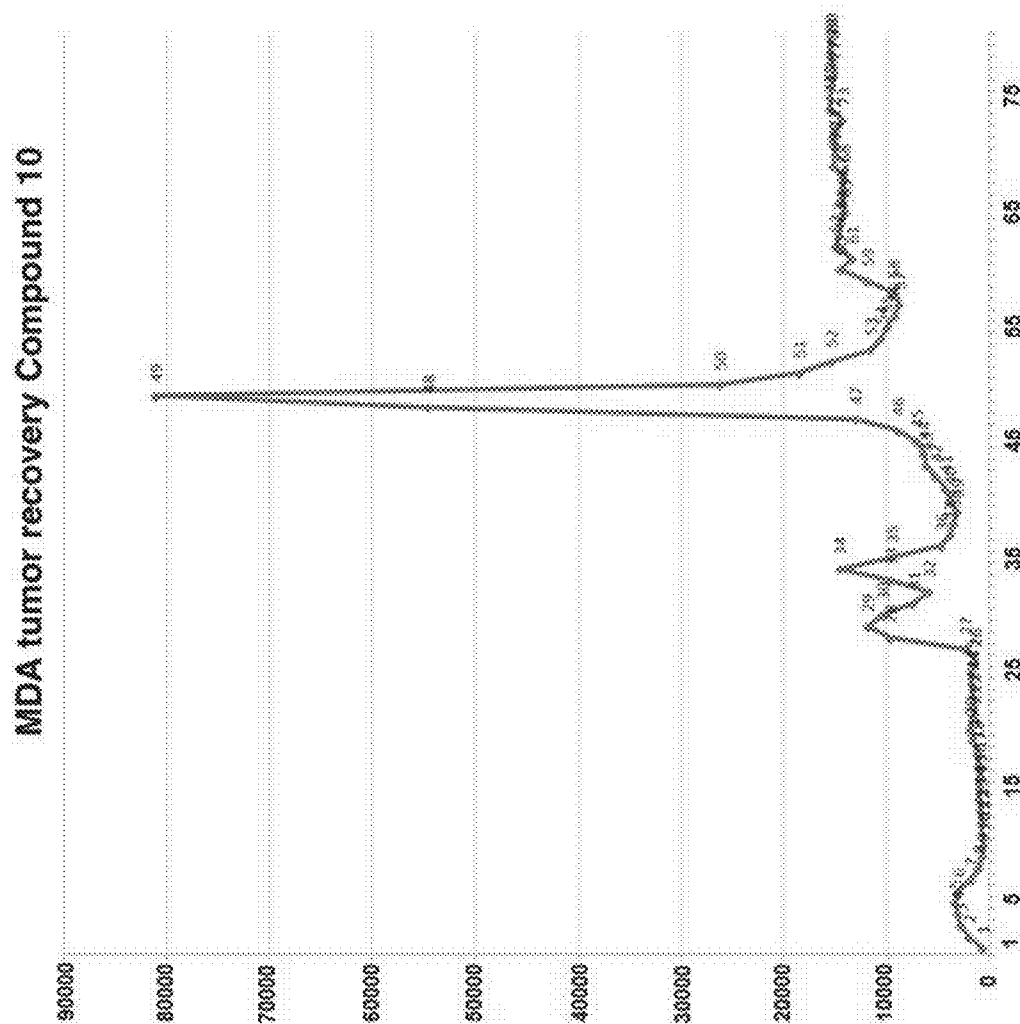
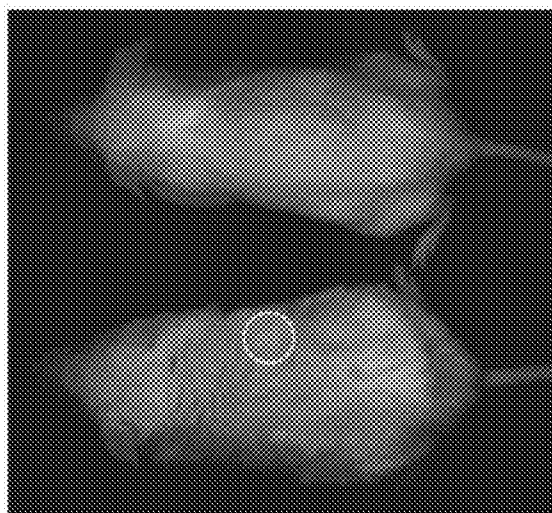


FIG. 17



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SUBSTITUTED 6-AMINOPURINES FOR TARGETING HSP90

CROSS-REFERENCE TO RELATED APPLICATIONS

This patent application is a divisional of U.S. patent application Ser. No. 14/419,965, filed Feb. 6, 2015, which is a national stage filing under 35 U.S.C. § 371 of International Application No. PCT/US2013/031614, filed on Mar. 14, 2013, which claims priority to U.S. Provisional Patent Application No. 61/680,107, filed on Aug. 6, 2012, the entire contents of which are hereby incorporated by reference in their entirety.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH

This invention was made with U.S. Government support awarded by the National Institutes of Health, Grant Nos. 1R01-A1089526-01 and 1R01-A1090644-01. The U.S. Government has certain rights in this invention.

BACKGROUND

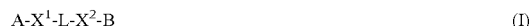
Heat shock protein 90 (Hsp90), one of the most abundant proteins expressed in cells, regulates cellular homeostasis by chaperoning protein folding and trafficking. Hsp90 is also highly upregulated in response to stress. The N-terminal domain of Hsp90 includes an ATP binding site, and ATPase activity is necessary for all of its cellular functions. To date, over 200 Hsp90 "client" proteins have been identified and many of these are involved in signal transduction.

Hsp90 has been implicated in diseases such as cancer and its expression is up-regulated during oncogenesis. High expression is associated with poor prognosis for cancers, such as breast cancer, prostate cancer, non-small cell lung cancer, colorectal cancer and melanoma.

Proteins such as glucose-regulated protein 94 (GRP94) and tumor necrosis factor receptor-associated protein (TRAP1) share homology with Hsp90 and both proteins also possess ATPase activity. While inhibitors of Hsp90 have been shown to have antiproliferative and antitumor activities, current Hsp90 inhibitors may bind nonspecifically to GRP94 and TRAP1. There is a need for compounds that selectively bind to Hsp90.

SUMMARY

In one aspect, the disclosure provides a compound of formula (1):



wherein:

A is an Hsp90 binding component;

X¹ is selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

X² is selected from the group consisting of —NR—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is selected from the group consisting of a detection moiety, an anti-cancer agent, and an Hsp90 binding component; and

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R is selected from the group consisting of hydrogen and a detection moiety.

In another aspect, the disclosure provides a method of detecting Hsp90 in a sample, comprising:

- a) contacting the sample with a compound having the following formula (Ia):



wherein:

A is an Hsp90 binding component;

X¹ is selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

X² is selected from the group consisting of —NR—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety; and

- b) detecting a signal from the detection moiety.

In another aspect, the disclosure provides a method of detecting cancer in a subject, comprising:

- a) contacting a biological sample from the subject with a compound having the following formula (Ia):



wherein:

A is an Hsp90 binding component;

X¹ is selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

X² is selected from the group consisting of —NR—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety; and

- b) detecting a signal from the detection moiety;

wherein cancer is detected in the sample when the signal from the detection moiety is higher relative to a signal from a reference sample.

In another aspect, the disclosure provides a method of treating cancer in a subject in need of treatment, comprising administering the subject a therapeutically effective amount of a compound having the following formula (Ib):



wherein:

A is an Hsp90 binding component;

X¹ and X² are each independently selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen; and

B is selected from the group consisting of an anticancer agent and an Hsp90 binding component.

In another aspect, the disclosure provides a method of detecting Human Immunodeficiency Virus (HIV) in a subject, the method comprising:

3

- a) contacting a biological sample from the subject with a compound having the following formula (Ia):



wherein:

A is an Hsp90 binding component;

X^1 is selected from the group consisting of $-\text{NH}-$, $-\text{O}-$, $-\text{S}-$, $-\text{C}(\text{O})-$ and $-\text{S}(\text{O})_2-$;

X^2 is selected from the group consisting of $-\text{NR}-$, $-\text{O}-$, $-\text{S}-$, $-\text{C}(\text{O})-$ and $-\text{S}(\text{O})_2-$;

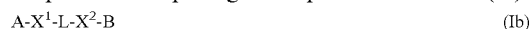
L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety; and

- b) detecting a signal from the detection moiety; wherein HIV is detected in the sample when the signal from the detection moiety is higher relative to a signal from a reference sample.

In another aspect, the disclosure provides a pharmaceutical composition comprising a compound of formula (Ib):



wherein:

A is an Hsp90 binding component;

X^1 and X^2 are each independently selected from the group consisting of $-\text{NH}-$, $-\text{O}-$, $-\text{S}-$, $-\text{C}(\text{O})-$ and $-\text{S}(\text{O})_2-$;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen; and

B is selected from the group consisting of an anticancer agent and an Hsp90 binding component; and a pharmaceutically acceptable carrier.

In another aspect, the disclosure provides a kit comprising a compound of formula (I) as described herein.

Other aspects and embodiments will become apparent in light of the following description and drawings.

BRIEF DESCRIPTION OF THE DRAWINGS

FIGS. 1A-1E is an SDS-PAGE silver stain showing the effect of different linker structures on Hsp90 recovery and recovery of non-specifically bound proteins. Selectivity towards Hsp90 was demonstrated by including 1 mM compound 5 in the tissue extract prior to mixing.

FIG. 2 is an SDS-PAGE silver stain showing elution of pig mammary gland proteins from ATP-Sepharose media with a compound described herein (8) and two known Hsp90 inhibitors.

FIG. 3 is an SDS-PAGE silver stain showing elution of pig mammary gland proteins from ATP-Sepharose media with various compounds.

FIG. 4 represents fluorescence microscopy images of cancer cell lines treated with a compound described herein (10).

FIGS. 5A and 5B, show fluorescence microscopy images of metastatic cancer cells and benign cells treated with: FIG. 5A an anti-Hsp90 antibody; and FIG. 5B compound 10.

FIGS. 6A-6C show fluorescence microscopy images of MCF7 cells treated with compound 10, in the presence or absence of an anti-Hsp90 antibody, and an Hsp90-binding compound.

FIGS. 7A and 7B show labeling of MDA-MB-468 tumor cells in NOD/SCID mice.

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FIG. 8 shows fluorescence microscopy images of xenografts harvested one hour after injection (IP) with compound 10.

- FIG. 9 shows the results of competitive binding experiments between compound 10 and the free parent drug compound 5.

FIGS. 10A-10C show the distribution of compound 10 in mice in FIG. 10A splenocytes; FIG. 10B liver cells; and FIG. 10C tumor cells.

- FIG. 11 shows quantitative analysis of the distribution of compound 10 in tumor cells, splenocytes and hepatocytes.

FIG. 12 shows fluorescence microscopy images of compound 10 uptake in breast cancer cells.

- FIGS. 13A and 13B show chromatograms of Hsp90 purified from: FIG. 13A pig mammary gland and BT474 cells, and FIG. 13B MCF7 cells, treated with compound 10.

- FIGS. 14A-14C show: FIG. 14A Fluorescence images of HSP90 expression in mouse xenografts; FIG. 14B quantitative analysis of compound 10 accumulation in tumors isolated from mice; and FIG. 14C accumulation of compound 12 within tumors isolated from mice.

FIG. 15 shows in vivo detection of compound 10 with optical spectroscopy.

- FIGS. 16A-16D show images of cells infected with human immunodeficiency virus, treated with compound 10 and an HIV-specific antibody.

FIG. 17 shows recovery of compound 10-bound protein from mice.

DETAILED DESCRIPTION

Described herein are compounds that may selectively bind to Hsp90. The compounds include an Hsp90 binding component, a linker, and an active moiety such as a detection group (e.g., a fluorophore or a radioisotope), an anti-cancer agent, or a second Hsp90 binding component. The compounds described herein may selectively bind to Hsp90 and may have significantly higher affinities for Hsp90 than for proteins with homology to Hsp90, such as GRP94 and TRAP1. The selective nature of the compounds may make them useful probes of Hsp90 in samples, and may allow for the selective targeting of Hsp90. For example, compounds described herein may be used to selectively detect Hsp90 in samples, and to selectively deliver anti-cancer agents to cells expressing high levels of Hsp90.

1. Definitions

The terminology used herein is for the purpose of describing particular embodiments only and is not intended to be limiting. As used in the specification and the appended claims, the singular forms "a," "and" and "the" include plural references unless the context clearly dictates otherwise.

Section headings as used in this section and the entire disclosure herein are not intended to be limiting.

- For the recitation of numeric ranges herein, each intervening number there between with the same degree of precision is explicitly contemplated. For example, for the range 6-9, the numbers 7 and 8 are contemplated in addition to 6 and 9, and for the range 6.0-7.0, the numbers 6.0, 6.1, 6.2, 6.3, 6.4, 6.5, 6.6, 6.7, 6.8, 6.9 and 7.0 are explicitly contemplated.

- As used herein, the term "about" is used synonymously with the term "approximately." Illustratively, the use of the term "about" indicates that values slightly outside the cited values, namely, plus or minus 10%. Such values are thus encompassed by the scope of the claims reciting the terms "about" and "approximately."

The terms “administer”, “administering”, “administered” or “administration” refer to any manner of providing a compound or a pharmaceutical composition (e.g., one described herein), to a subject or patient. Routes of administration can be accomplished through any means known by those skilled in the art. Such means include, but are not limited to, oral, buccal, intravenous, subcutaneous, intramuscular, transdermal, by inhalation and the like.

“Contacting” as used herein, e.g., as in “contacting a sample” refers to contacting a sample directly or indirectly in vitro, ex vivo, or in vivo (i.e. within a subject as defined herein). Contacting a sample may include addition of a compound to a sample (e.g., a sample comprising cells that contain Hsp90), or administration to a subject. Contacting encompasses administration to a solution, cell, tissue, mammal, subject, patient, or human. Further, contacting a cell includes adding an agent to a cell culture.

“Detection moiety” as used herein includes one or more groups that are detectable, either directly or indirectly, by methods such as spectroscopic, photochemical, biochemical, chemical, or other methods. For example, useful detectable moieties or labels include chromophores, fluorophores, biotin, radioactive compounds, and the like. The detection moiety often generates a measurable signal, such as a radioactive, chromogenic, luminescent, or fluorescent signal, which can be used to quantitate the amount of the detection moiety in a sample. In some embodiments a detection moiety may include more than one detectable group, e.g., a fluorophore and a radioactive moiety.

“Effective amount,” as used herein, refers to a dosage or an amount of a compound or a composition effective for eliciting a desired effect. This term as used herein may also refer to an amount effective at bringing about a desired in vivo effect in an animal, e.g., a mammal, e.g., a human. For example, in methods of treating cancer, an effective amount may be an amount sufficient to treat the disorder.

“Member atom” as used herein refers to a polyvalent atom (e.g., a C, O, N, or S atom) in a chain or ring system that constitutes a part of the chain or ring. For example, in pyridine, five carbon atoms and one nitrogen atom are member atoms of the ring. In diethyl ether, four carbon atoms and one oxygen atom are member atoms of the chain. Member atoms will be substituted up to their normal valence. For example, in pyridine, the five carbon atoms will each be further substituted with a hydrogen or another substituent.

As used herein, the term “subject” is intended to include human and non-human animals. Exemplary human subjects include a human patient having a disorder, e.g., cancer, or a normal subject. The term “non-human animals” includes all vertebrates, e.g., non-mammals (such as chickens, amphibians, reptiles) and mammals, such as non-human primates, domesticated and/or agriculturally useful animals (such as sheep, dogs, cats, cows, pigs, etc.), and rodents (such as mice, rats, hamsters, guinea pigs, etc.).

As used herein, the term “treat” or “treating” a subject having a disorder refers to administering a compound or a composition described herein to the subject, such that at least one symptom of the disorder is cured, healed, alleviated, relieved, altered, remedied, ameliorated, or improved. Treating includes administering an amount effective to alleviate, relieve, alter, remedy, ameliorate, cure, improve or affect the disorder or the symptoms of the disorder. The treatment may inhibit deterioration or worsening of a symptom of a disorder.

Definitions of specific functional groups and chemical terms are described in more detail below. For purposes of

this disclosure, the chemical elements are identified in accordance with the Periodic Table of the Elements, CAS version, Handbook of Chemistry and Physics, 75th Ed., inside cover, and specific functional groups are generally defined as described therein. Additionally, general principles of organic chemistry, as well as specific functional moieties and reactivity, are described in *Organic Chemistry*, Thomas Sorrell, University Science Books, Sausalito, 1999; Smith and March *March's Advanced Organic Chemistry*, 5th Edition; John Wiley & Sons, Inc., New York, 2001; Larock, *Comprehensive Organic Transformations*, VCH Publishers, Inc., New York, 1989; Carruthers, *Some Modern Methods of Organic Synthesis*, 3rd Edition, Cambridge University Press, Cambridge, 1987; the entire contents of each of which are incorporated herein by reference.

The term “acyl” refers to an alkylcarbonyl, cycloalkylcarbonyl, heterocyclylcarbonyl, arylcarbonyl or heteroarylcarbonyl substituent, any of which may be further substituted (e.g., with one or more substituents).

The term “alkyl” refers to a straight or branched saturated hydrocarbon chain. Alkyl groups may include specified number of carbon atoms. For example, C₁-C₁₂ alkyl indicates that the alkyl group may have 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 or 12 carbon atoms. An alkyl group may be, e.g., a C₁-C₁₂ alkyl group, a C₁-C₁₀ alkyl group, a C₁-C₈ alkyl group, a C₁-C₆ alkyl group or a C₁-C₄ alkyl group. For example, exemplary C₁-C₄ alkyl groups include methyl, ethyl, n-propyl, isopropyl, n-butyl, sec-butyl, isobutyl and tert-butyl groups. An alkyl group may be optionally substituted with one or more substituents.

The term “alkylenyl” refers to a divalent alkyl group, examples of which include but are not limited to —CH₂—, —CH₂CH₂—, —CH₂CH₂CH₂— and —CH₂CH(CH₃)CH₂—. An alkylenyl group may be optionally substituted with one or more substituents.

The term “alkenyl” refers to a straight or branched hydrocarbon chain having one or more double bonds. Alkenyl groups may include a specified number of carbon atoms. For example, C₂-C₁₂ alkenyl indicates that the alkenyl group may have 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 or 12 carbon atoms. An alkenyl group may be, e.g., a C₂-C₁₂ alkenyl group, a C₂-C₁₀ alkenyl group, a C₂-C₈ alkenyl group, a C₂-C₆ alkenyl group or a C₂-C₄ alkenyl group. Examples of alkenyl groups include but are not limited to allyl, propenyl, 2-butenyl, 3-hexenyl and 3-octenyl groups. One of the double bond carbons may optionally be the point of attachment of the alkenyl substituent. An alkenyl group may be optionally substituted with one or more substituents.

The term “alkenylenyl” refers to a divalent alkenyl group, examples of which include but are not limited to —CH=CH—, —CH=CH—CH₂—, —CH=CH—CH₂—CH₂— and —CH₂—CH=CH—CH₂—. An alkenylenyl group may be optionally substituted with one or more substituents.

The term “alkynyl” refers to a straight or branched hydrocarbon chain having one or more triple bonds. Alkynyl groups may include a specified number of carbon atoms. For example, C₂-C₁₂ alkynyl indicates that the alkynyl group may have 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 or 12 carbon atoms. An alkynyl group may be, e.g., a C₂-C₁₂ alkynyl group, a C₂-C₁₀ alkynyl group, a C₂-C₈ alkynyl group, a C₂-C₆ alkynyl group or a C₂-C₄ alkynyl group. Examples of alkynyl groups include but are not limited to ethynyl, propargyl, and 3-hexynyl. One of the triple bond carbons may optionally be the point of attachment of the alkynyl substituent. An alkynyl group may be optionally substituted with one or more substituents.

The term “alkynylenyl” refers to a divalent alkynyl group, examples of which include but are not limited to $-\text{C}\equiv\text{C}-$, $-\text{C}\equiv\text{C}-\text{CH}_2-$, $-\text{C}\equiv\text{C}-\text{CH}_2-\text{CH}_2-$ and $-\text{CH}_2-\text{C}\equiv\text{C}-\text{CH}_2-$. An alkynylenyl group may be optionally substituted with one or more substituents.

The term “aryl” refers to an aromatic monocyclic, bicyclic, or tricyclic hydrocarbon ring system, wherein any ring atom capable of substitution can be substituted (e.g., with one or more substituents). Examples of aryl moieties include but are not limited to phenyl, naphthyl, and anthracenyl. Aryl groups may be optionally substituted with one or more substituents.

The term “arylalkyl” refers to an alkyl moiety in which at least one alkyl hydrogen atom is replaced with an aryl group. Arylalkyl includes groups in which more than one hydrogen atom has been replaced with an aryl group. Examples of arylalkyl groups include but are not limited to benzyl, 2-phenylethyl, 3-phenylpropyl, 9-fluorenyl, benzhydryl, and trityl groups. Arylalkyl groups may be optionally substituted with one or more substituents, on either the aryl moiety or the alkyl moiety.

The term “cycloalkyl” as used herein refers to non-aromatic, saturated or partially unsaturated cyclic, bicyclic, tricyclic or polycyclic hydrocarbon groups having 3 to 12 carbons. Any ring atom can be substituted (e.g., with one or more substituents). Cycloalkyl groups can contain fused rings. Fused rings are rings that share one or more common carbon atoms. Examples of cycloalkyl groups include but are not limited to cyclopropyl, cyclobutyl, cyclopentyl, cyclohexyl, cyclohexenyl, cyclohexadienyl, methylcyclohexyl, adamantyl, norbornyl, norbornenyl, tetrahydronaphthalenyl and dihydroindenyl. Cycloalkyl groups may be optionally substituted with one or more substituents.

The term “cycloalkylalkyl”, as used herein, refers to an alkyl group in which at least one hydrogen atom is replaced with a cycloalkyl group. Cycloalkylalkyl groups include those in which more than one hydrogen atom of the alkyl group is replaced with a cycloalkyl group. Examples of cycloalkylalkyl groups include but are not limited to cyclohexylmethyl, cyclopentylmethyl, cyclobutylmethyl and cyclopropylmethyl. Cycloalkylalkyl groups can be optionally substituted with one or more substituents, on either the cycloalkyl moiety or the alkyl moiety.

The term “halo” or “halogen” as used herein refers to any radical of fluorine, chlorine, bromine or iodine.

The term “haloalkyl” as used herein refers to an alkyl group as defined herein, in which one or more hydrogen atoms are replaced with halogen atoms, and includes alkyl moieties in which all hydrogens have been replaced with halogens (e.g., perfluoroalkyl such as CF_3).

“Heteroalkyl” refers to an alkyl, alkenyl or alkynyl group as defined herein, wherein at least one carbon atom of the alkyl group is replaced with a heteroatom. Heteroalkyl groups may contain from 1 to 18 non-hydrogen atoms (carbon and heteroatoms) in the chain, or 1 to 12 atoms, or 1 to 6 atoms, or 1 to 4 atoms. Heteroalkyl groups may be straight or branched, and saturated or unsaturated. Unsaturated heteroalkyl groups have one or more double bonds and/or one or more triple bonds. Heteroalkyl groups may be unsubstituted or substituted. Exemplary heteroalkyl groups include but are not limited to alkoxyalkyl (e.g., methoxymethyl), and aminoalkyl (e.g., alkylaminoalkyl and dialkylaminoalkyl). Heteroalkyl groups may be optionally substituted with one or more substituents.

The term “heteroalkylenyl” refers to a divalent heteroalkyl group, examples of which include but are not limited to $-\text{CH}_2\text{OCH}_2-$, $-\text{CH}_2\text{NHCH}_2-$, polyethyleneglycol

groups (e.g., $-(\text{CH}_2\text{CH}_2\text{O})_n-$), polyethyleneimine groups (e.g., $-(\text{CH}_2\text{CH}_2\text{NH})_n-$), and the like. A heteroalkylenyl group may be optionally substituted with one or more substituents.

The term “heteroaryl” as used herein refers to an aromatic 5-8 membered monocyclic, 8-12 membered bicyclic, or 11-14 membered tricyclic ring system having 1-3 heteroatoms if monocyclic, 1-6 heteroatoms if bicyclic, or 1-9 heteroatoms if tricyclic, said heteroatoms independently selected from O, N, S, P and Si (e.g., carbon atoms and 1-3, 1-6, or 1-9 heteroatoms independently selected from O, N, S, P and Si if monocyclic, bicyclic, or tricyclic, respectively). Any ring atom can be substituted (e.g., with one or more substituents). Heteroaryl groups can contain fused rings, which are rings that share one or more common atoms. Examples of heteroaryl groups include but are not limited to radicals of pyridine, pyrimidine, pyrazine, pyridazine, pyrrole, imidazole, pyrazole, oxazole, isoxazole, furan, thiazole, isothiazole, thiophene, quinoline, isoquinoline, quinoxaline, quinazoline, cinnoline, indole, isoindole, indolizine, indazole, benzimidazole, phthalazine, pteridine, carbazole, carboline, phenanthridine, acridine, phenanthroline, phenazine, naphthyridines and purines. Heteroaryl groups may be optionally substituted with one or more substituents.

The term “heteroarylalkyl” refers to an alkyl moiety in which at least one alkyl hydrogen atom is replaced with a heteroaryl group. Heteroarylalkyl includes groups in which more than one hydrogen atom has been replaced with a heteroaryl group. Examples of heteroarylalkyl groups include but are not limited to imidazolylmethyl (e.g., 1H-imidazol-2-ylmethyl and 1H-imidazol-4-ylmethyl), pyridinylmethyl (e.g., pyridin-3-ylmethyl and pyridin-4-ylmethyl), pyrimidinylmethyl (e.g., pyrimidin-5-ylmethyl), furylmethyl (e.g., fur-2-ylmethyl and fur-3-ylmethyl), and thienylmethyl (e.g., thien-2-ylmethyl and thien-3-ylmethyl) groups. Heteroarylalkyl groups may be optionally substituted with one or more substituents, on either the heteroaryl moiety or the alkyl moiety.

The term “heteroatom”, as used herein, refers to a non-carbon or hydrogen atom such as a nitrogen, sulfur, oxygen, silicon or phosphorus atom. Groups containing more than one heteroatom may contain different heteroatoms.

The term “heterocyclyl”, as used herein, refers to a nonaromatic, saturated or partially unsaturated 3-10 membered monocyclic, 8-12 membered bicyclic, or 11-14 membered tricyclic ring system having 1-3 heteroatoms if monocyclic, 1-6 heteroatoms if bicyclic, or 1-9 heteroatoms if tricyclic, said heteroatoms selected from O, N, S, Si and P (e.g., carbon atoms and 1-3, 1-6, or 1-9 heteroatoms of O, N, S, Si and P if monocyclic, bicyclic, or tricyclic, respectively). Any ring atom can be substituted (e.g., with one or more substituents). Heterocyclyl groups can contain fused rings, which are rings that share one or more common atoms. Examples of heterocyclyl groups include but are not limited to radicals of tetrahydrofuran, tetrahydrothiophene, tetrahydropyran, oxetane, piperidine, piperazine, morpholine, pyrrolidine, pyrimidine, pyrrolidine, indoline, tetrahydropyridine, dihydropyran, thianthrene, pyran, benzopyran, xanthene, phenoxathiin, phenothiazine, furazan, lactones, lactams such as azetidinones and pyrrolidinones, sultams, sultones, and the like. Heterocyclyl groups may be optionally substituted with one or more substituents.

The term “heterocyclylalkyl” refers to an alkyl moiety in which at least one alkyl hydrogen atom is replaced with a heterocyclyl group. Heterocyclylalkyl includes groups in which more than one hydrogen atom has been replaced with

a heterocyclyl group. Examples of heterocyclylalkyl groups include but are not limited to oxetanylmethyl, morpholinomethyl, and pyrrolidinylmethyl groups, and the like. Heterocyclylalkyl groups may be optionally substituted with one or more substituents, on either the heterocyclyl moiety or the alkyl moiety.

The term “hydroxy” refers to an —OH radical. The term “alkoxy” refers to an —O-alkyl radical. The term “aryloxy” refers to an —O-aryl radical.

The term “oxo” refers to an oxygen atom, which forms a carbonyl when attached to carbon, an N-oxide when attached to nitrogen, and a sulfoxide or sulfone when attached to sulfur (i.e. =O).

The term “mercapto” or “thiol” refers to an —SH radical. The term “thioalkoxy” or “thioether” refers to an —S-alkyl radical. The term “thioaryloxy” refers to an —S-aryl radical.

The term “substituents” refers to a group “substituted” on an alkyl, alkenyl, alkynyl, cycloalkyl, heterocyclyl, aryl, arylalkyl, heteroaryl or heteroarylalkyl group at any atom of that group. Any atom can be substituted. Suitable substituents include, without limitation: acyl, acylamido, acyloxy, alkoxy, alkyl, alkenyl, alkynyl, amido, amino, carboxy, cyano, ester, halo, hydroxy, imino, nitro, oxo (e.g., C=O), phosphonate, sulfinyl, sulfonyl, sulfonate, sulfonamino, sulfonamido, thioamido, thiol, thioxo (e.g., C=S), and ureido. In embodiments, substituents on a group are independently any one single, or any combination of the aforementioned substituents. In embodiments, a substituent may itself be substituted with any one of the above substituents.

The above substituents may be abbreviated herein. For example, the abbreviations Me, Et, Ph and Bn represent methyl, ethyl, phenyl and benzyl, respectively. A more comprehensive list of standard abbreviations used by organic chemists appears in a table entitled Standard List of Abbreviations of the Journal of Organic Chemistry. The abbreviations contained in said list are hereby incorporated by reference.

For compounds described herein, groups and substituents thereof may be selected in accordance with permitted valence of the atoms and the substituents, and such that the selections and substitutions result in a stable compound, e.g., a compound that does not spontaneously undergo transformation such as by rearrangement, cyclization, elimination, etc.

Where substituent groups are specified by their conventional chemical formulae, written from left to right, they optionally encompass substituents resulting from writing the structure from right to left, e.g., —CH₂O— optionally also recites —OCH₂—.

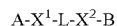
In accordance with a convention used in the art, the group:



is used in structural formulae herein to depict the bond that is the point of attachment of the moiety or substituent to the core or backbone structure.

2. Compounds

Compounds that may selectively bind to Hsp90 include compounds of formula (I):



(I)

wherein:

A is an Hsp90 binding component;

X¹ is selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

X² is selected from the group consisting of —NR—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is selected from the group consisting of a detection moiety, an anti-cancer agent, and an Hsp90 binding component; and

R is selected from the group consisting of hydrogen, and a detection moiety.

Compounds that may be used in methods described herein also include compounds of formula (Ia):



(Ia)

wherein:

A is an Hsp90 binding component;

X¹ is selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

X² is selected from the group consisting of —NR—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety.

Compounds that may be used in methods described herein also include compounds of formula (Ib):



(Ib)

wherein:

A is an Hsp90 binding component;

X¹ and X² are each independently selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen; and

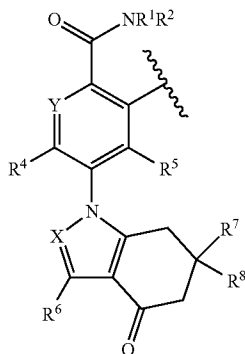
B is selected from the group consisting of an anti-cancer agent and an Hsp90 binding component.

a. Hsp90 Binding Components

In the compounds of formula (I), A is an Hsp90 binding component. The compounds of formula (I) include at least one Hsp90 binding component, and may include two Hsp90 binding components in embodiments in which B is also an Hsp90 component. Any molecule that binds to Hsp90 can be used as the basis for the Hsp90 binding component. Based on the complete structure of the compounds of formula (I), it will be understood that the Hsp90 binding component corresponds to an Hsp90 binding compound in which an atom or a group of atoms is removed, to provide a point of attachment to the —X¹-L-X²-B moiety of the remainder of the compound of formula (I) (or the —X²-L-X¹-A moiety in embodiments in which B is an Hsp90 binding component).

In embodiments, an Hsp90 binding component has the following formula (II):

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wherein:

R¹ and R² are independently selected from the group consisting of —H and —C₁₋₈-alkyl;

Y is CR³ or N;

R³ is —H, —F or —OCH₃;

R⁴ and R⁵ are independently selected from the group consisting of —H, —F and —OCH₃;

R⁶ is —C₁₋₈-alkyl, —C₂₋₈-alkenyl, —C₂₋₈-alkynyl, —C₃₋₈-cycloalkenyl, —C₃₋₈-cycloalkenyl-C₁₋₈-alkyl, —C₃₋₈-cycloalkyl, —C₃₋₈-cycloalkyl-C₁₋₈-alkyl, aryl, aryl-C₁₋₈-alkyl, halo-C₁₋₈-alkyl, heteroaryl, heteroaryl-C₁₋₈-alkyl, heterocyclyl, heterocyclyl-C₁₋₈-alkyl, or hydroxy-C₁₋₈-alkyl;

R⁷ and R⁸ are independently selected from the group consisting of —H and —C₁₋₈-alkyl; or R⁷ and R⁸ taken together with the carbon atom to which they are attached form a C₃₋₈-cycloalkyl group;

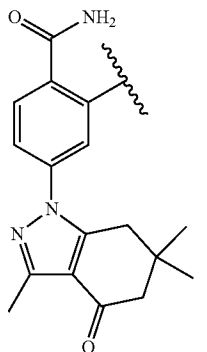
X is N or CR⁹; and

R⁹ is —H or —C₁₋₈-alkyl; and



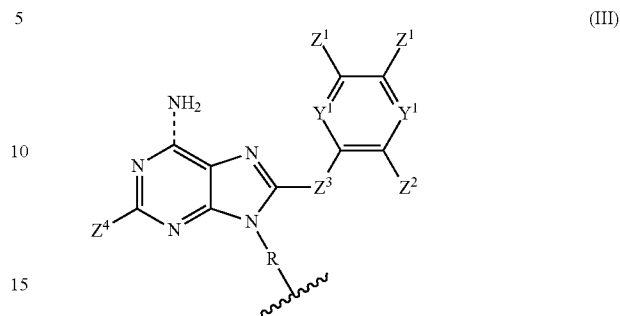
represents the point of attachment to the —X¹-L-X²-B moiety of the compound of formula (I).

In embodiments of formula (II), R¹ and R² are each —H, Y is CR³, R³ is —H, R⁴ and R⁵ are each —H, R⁶ is —C₁₋₈-alkyl (e.g., methyl) or halo-C₁₋₈-alkyl (e.g., trifluoromethyl), R⁷ and R⁸ are independently —C₁₋₈-alkyl (e.g., methyl), and X is N. In embodiments, formula (II) is the following:



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(II) In embodiments, an Hsp90 binding component has the following formula (III):



wherein:

each Y¹ is independently selected from the group consisting of CH and N;

R is alkylenyl or heteroalkylenyl;

each Z¹ is taken together with the carbon atoms to which they are attached to form a heterocyclic ring;

Z² is hydrogen or halo;

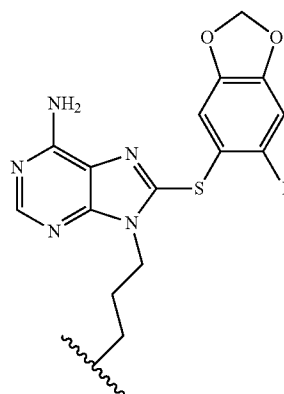
Z³ is CH₂, S, O, or NH;

Z⁴ is hydrogen or halo; and



represents the point of attachment to the remainder of the compound of formula (I), e.g., to the —X¹-L-X²-B moiety, or to the X²-L-X¹-A moiety when B is an Hsp90 binding component having the formula (III).

In embodiments, a group of formula (III) is the following:



The Hsp90 binding component may comprise any known Hsp90 binding compound or a portion thereof that retains affinity for Hsp90. Other compounds that bind to Hsp90 are known in the art, and include, for example, geldanamycin, herbimycin, radicicol, deguelin, derrubone and macebecin. Other compounds that bind to Hsp90 may include but are not limited to those described in U.S. Pat. Nos. 7,358,370, 7,678,803, 7,906,529 and 7,928,135, U.S. Patent Publication No. 2011/0183977, and International Patent Publication Nos. WO2008/130879 and WO2006/084030.

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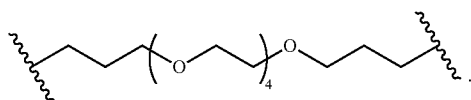
b. Linker

In the compounds of formula (I), L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen. In embodiments, L is a heteroalkylenyl group comprising at least twelve member atoms. In some embodiments, L comprises 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49 or 50 atoms. In some embodiments, the heteroalkylenyl group may be optionally substituted (e.g., a carbon may be substituted with an oxo group).

In embodiments, L is a straight-chain heteroalkylenyl group. In embodiments, L includes at least one monomer of a polyethylene glycol group (i.e. $-(CH_2CH_2O)-$), or it may include at least two, three, four, five, six or more such monomers.

In some embodiments, L has the formula $-(CH_2)_m-(OCH_2CH_2)_n-O-(CH_2)_p-$, wherein m is 2 or 3, n is 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20, and p is 2 or 3. In embodiments, m is 2. In embodiments, m is 3. In embodiments, n is 4, 5, 6, 7, 8, 9 or 10. In embodiments, n is 4. In embodiments, p is 2. In embodiments, p is 3.

A suitable linker has the following formula:

c. X^1 and X^2

The groups X^1 and X^2 in the compounds formula (I) are each independently selected from the group consisting of $-NH-$, $-O-$, $-S-$, $-C(O)-$ and $-S(O)_2-$. These groups serve as bridges between the linker and the A and B groups of the compound of formula (I). During the synthesis of these molecules (described in further detail below), the X^1 and X^2 groups may be derived from a linker precursor compound. For example, in some embodiments X^1 and X^2 may be $-NH-$. In such embodiments, a suitable linker precursor compound may have the formula $H_2N-Linker-NH_2$. In this case, the terminal amino groups of the linker precursor compound provide the source of the X^1 and X^2 groups.

In some embodiments, X^2 may be NR, wherein R is selected from the group consisting of hydrogen and a detection moiety. Such compounds may be synthesized, for example, by first synthesizing a group in which X^2 is NH and then further derivatizing the NH group with a compound comprising an additional detection moiety.

d. Detection Moieties

In embodiments of the compounds of formula (I), B is a detection moiety. The detection moiety comprises one or more groups that are, either directly or indirectly, detectable via a method such as a spectroscopic method. Exemplary detection moieties may comprise, for example, a chromophore, a fluorophore, a luminescent moiety or a radioisotope. In some embodiments, B comprises one or more detection moieties, such as, for example, a fluorophore and a radioisotope.

In embodiments, the detection moiety comprises a fluorophore. Suitable fluorophores include fluoresceins, rhodamines, coumarins, cyanines, and boron-dipyrromethenes (also known as BODIPYs). The fluorophores may be attached to the remainder of a compound of formula (I), for

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example, by using a reagent comprising a fluorophore and a reactive group such as a carboxylic acid, an isothiocyanate, a maleimide, or an ester such as a succinimidyl, pentafluorophenyl or tetrafluorophenyl ester. Such groups may react with a group present on a linker precursor compound, such as an amine, to attach the fluorophore to the remainder of the molecule of formula (I).

Suitable reagents comprising fluorophores, which may be used to prepare compounds of formula (I), are known in the art, and include but are not limited to fluoresceins, rhodamines, coumarins, cyanines and boron-dipyrromethenes. For example, reagents comprising fluorophores that are commercially available include but are not limited to: 5- and 6-carboxyfluoresceins and esters thereof; fluorescein-5-isothiocyanate and fluorescein-6-isothiocyanate; BODIPY® dyes commercially available from Molecular Probes; Alexa Fluor® dyes commercially available from Molecular Probes; CyDye fluors commercially available from GE Healthcare Biosciences, including but not limited to Cy3, Cy 5, Cy5.5 and Cy 7 esters; and VivoTag™ fluorophores available from PerkinElmer, including but not limited to VivoTag 645, VivoTag 645-MAL, VivoTag 680, VivoTag 680-MAL, VivoTag 680 XL, VivoTag-S 680, VivoTag 750, VivoTag-S 750, VivoTag 750-MAL, and VivoTag 800.

It will be understood by the skilled artisan that when B is a detection moiety, e.g., a detection moiety comprising a fluorophore, the detection moiety may include the detectable moiety as well as additional atoms or groups of atoms. For example, as will be further described in the synthesis section, a fluorophore may be attached to the remainder of a compound of formula (I), for example, by reacting a compound of the formula $A-X^1-L-NH_2$ with a reagent comprising a fluorophore and a reactive group, such as an isothiocyanate or N-succinimidyl ester. A reaction with an isothiocyanate will produce a compound $A-X^1-L-NH-C(S)-NH-fluorophore$. In such instances, it is understood that the $-NH-$ group attached to $-L-$ corresponds to the $-X_2-$ group of formula (I), while the group $-B$ corresponding to the "detection moiety" includes not only the fluorophore but also the $-C(S)-NH-$ linking atoms.

In other embodiments, the detection moiety may comprise a radioisotope. Such detection moieties may be useful for detection in samples by scintillation counting, or for radioimaging applications. Suitable radioisotopes include, but are not limited to, tritium (i.e. hydrogen-3), carbon-14, nitrogen-15, phosphorus-32, iodine-125 and iodine-131. For example, compounds comprising iodine-131 may be useful as radiopharmaceuticals.

e. Anti-cancer Agents

In embodiments of the compounds of formula (I), B comprises an anti-cancer agent. Exemplary anti-cancer/chemotherapeutic agents include, but are not limited to, the following:

alkylating agents (including, without limitation, nitrogen mustards, ethylenimine derivatives, alkyl sulfonates, nitrosoureas and triazines): uracil-mustard (Aminouracil Mustard®, Chlorethaminacil®, Demethylodopan®, Desmethyldopan®, Hacmanthamine®, Nordopan®, Uracil nitrogen Mustard®, Uracillost®, Uracilmotaza®, Uramustin®, Uramustine®), bendamustine (Treakisym®, Ribomustin®, Treanda®) chlormethine (Mustargen®), cyclophosphamide (Cytosan®, Neosar®, Clafen®, Endoxan®, Procytox®, Revimmune™), ifosfamide (Mitoxana®), melphalan (Alkeran®), Chlorambucil (Leukeran®), pipobroman (Amedel®, Vercyte®), triethyleriemelamine (Hemel®, Hexylen®, Hexastat®), triethylenethiophosphoramine, Temozolomide (Temodar®), thiotepa (Thioplex®), busulfan

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(Busilvex®), Myleran®), carmustine (BiCNU®), lomustine (CeeNU®), streptozocin (Zanosar®), estramustine (Emcyt®, Estracit®), fotemustine, irofulven, mannosulfan, mitobronitol, nimustine, procarbazine, ranimustine, semustine, triaziquone, treosulfan, and Dacarbazine (DTIC-Dome®).

anti-EGFR antibodies (e.g., cetuximab (Erbix®), panitumumab (Vectibx®), and gefitinib (Iressa®)).

anti-Her-2 antibodies (e.g., trastuzumab (Herceptin®) and other antibodies from Genentech).

antimetabolites (including, without limitation, folic acid antagonists (also referred to herein as antifolates), pyrimidine analogs, purine analogs and adenosine deaminase inhibitors): methotrexate (Rheumatrex®, Trexall®), 5-fluorouracil (Aducil®, Efudex®, Fluoroplex®), floxuridine (FUDF®), carmofur, cytarabine (Cytosar-U®, Tarabine PFS), 6-mercaptopurine (Puri-Nethol®), 6-thioguanine (Thioguanine Tabloid®), fludarabine phosphate (Fludara®), pentostatin (Nipent®), pemetrexed (Alimta®), raltitrexed (Tomudex®), cladribine (Leustatin®), clofarabine (Clofarex®, Clolar®), mercaptopurine (Puri-Nethol®), capecitabine (Xeloda®), nelarabine (Arranon®), azacitidine (Vidaza®), decitabine (Dacogen®), enocitabine (Sunrabin®), sapacitabine, tegafur-uracil, tiazofurine, tioguanine, trofosfamide, and gemcitabine (Gemzar®).

vinca alkaloids: vinblastine (Velban®, Velsar®), vincristine (Vincasar®, Oncovin®), vindesine (Eldisine®), vinorelbine (Navelbine®), vinflunine (Javlor®).

platinum-based agents: carboplatin (Paraplat®, Paraplatin®), cisplatin (Platinol®), oxaliplatin (Eloxatin®), nedaplatin, satraplatin, triplatin.

anthracyclines: daunorubicin (Cerubidine®, Rubidomycin®), doxorubicin (Adriamycin®), epirubicin (Ellence®), idarubicin (Idamycin®), mitoxantrone (Novantrone®), valrubicin (Valstar®), aclarubicin, amrubicin, liposomal doxorubicin, liposomal daunorubicin, pirarubicin, pixantrone, zorubicin.

topoisomerase inhibitors: topotecan (Hycamtin®), irinotecan (Camptosar®), etoposide (Toposar®, VePesid®), teniposide (Vumon®), lamellarin D, SN-38, camptothecin (e.g., IT-101), belotecan, rubitecan.

taxanes: paclitaxel (Taxol®), docetaxel (Taxotere®), larotaxel, cabazitaxel, ortataxel, tesetaxel.

antibiotics: actinomycin (Cosmegen®), bleomycin (Blenoxane®), hydroxyurea (Droxia®, Hydrea®), mitomycin (Mitozytrex®, Mutamycin®).

immunomodulators: lenalidomide (Revlimid®), thalidomide (Thalomid®).

immune cell antibodies: alemtuzumab (Campath®), gemtuzumab (Myelotarg®), rituximab (Rituxan®), tositumomab (Bexxar®).

interferons (e.g., IFN-alpha (Alferon®, Roferon-A®, Intron®-A) or IFN-gamma (Actimmune®)).

interleukins: IL-1, IL-2 (Proleukin®), IL-24, IL-6 (Sigosix®), IL-12.

HSP90 inhibitors (e.g., geldanamycin or any of its derivatives). In certain embodiments, the HSP90 inhibitor is selected from geldanamycin, 17-alkylamino-17-desmethoxygeldanamycin ("17-AAG") or 17-(2-dimethylaminoethyl)amino-17-desmethoxygeldanamycin ("17-DMAG").

anti-androgens which include, without limitation nilutamide (Nilandron®) and bicalutamide (Caxodex®).

antiestrogens which include, without limitation tamoxifen (Nolvadex®), toremifene (Fareston®), letrozole (Femara®), testolactone (Teslac®), anastrozole (Arimidex®), bicalutamide (Casodex®), exemestane (Aromasin®), flutamide (Eu-

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lexin®), fulvestrant (Faslodex®), raloxifene (Evista®, Keoxifene®) and raloxifene hydrochloride.

anti-hypercalcaemia agents which include without limitation gallium (III) nitrate hydrate (Ganite®) and pamidronate disodium (Aredia®).

apoptosis inducers which include without limitation ethanol, 2-[[3-(2,3-dichlorophenoxy)propyl]amino]-(9CI), gambogic acid, elesclomol, embelin and arsenic trioxide (Trisenox®).

Aurora kinase inhibitors which include without limitation binucleine 2.

Bruton's tyrosine kinase inhibitors which include without limitation terreic acid.

calcineurin inhibitors which include without limitation cypermethrin, deltamethrin, fenvalerate and tyrphostin 8.

CaM kinase II inhibitors which include without limitation 5-Isoquinolinesulfonic acid, 4-[(2S)-2-[(5-isoquinoliny)sulfonyl)methylamino]-3-oxo-3-[4-phenyl-1-piperazinyl]propyl]phenyl ester and benzenesulfonamide.

CD45 tyrosine phosphatase inhibitors which include without limitation phosphonic acid.

CDC25 phosphatase inhibitors which include without limitation 1,4-naphthalene dione, 2,3-bis[(2-hydroxyethyl)thio]-(9CI).

CHK kinase inhibitors which include without limitation debromohymenialdisine.

cyclooxygenase inhibitors which include without limitation 1H-indole-3-acetamide, 1-(4-chlorobenzoyl)-5-methoxy-2-methyl-N-(2-phenylethyl)-(9CI), 5-alkyl substituted 2-arylamino-phenylacetic acid and its derivatives (e.g., celecoxib (Celebrex®), rofecoxib (Vioxx®), etoricoxib (Arcoxia®), lumiracoxib (Prexige®), valdecoxib (Bextra®) or 5-alkyl-2-arylamino-phenylacetic acid).

cRAF kinase inhibitors which include without limitation 3-(3,5-dibromo-4-hydroxybenzylidene)-5-iodo-1,3-dihydroindol-2-one and benzamide, 3-(dimethylamino)-N-[3-[(4-hydroxybenzoyl)amino]-4-methylphenyl]-(9CI).

cyclin dependent kinase inhibitors which include without limitation olomoucine and its derivatives, purvalanol B, roscovitine (Seliciclib®), indirubin, kenpaullone, purvalanol A and indirubin-3'-monooxime.

cysteine protease inhibitors which include without limitation 4-morpholinecarboxamide, N-[(1S)-3-fluoro-2-oxo-1-(2-phenylethyl)propyl]amino]-2-oxo-1-(phenylmethyl)ethyl]-(9CI).

DNA intercalators which include without limitation plamycin (Mithracin®) and daptomycin (Cubicin®).

DNA strand breakers which include without limitation bleomycin (Blenoxane®).

E3 ligase inhibitors which include without limitation N-((3,3,3-trifluoro-2-trifluoromethyl)propionyl)sulfanilamide.

EGF Pathway Inhibitors which include, without limitation tyrphostin 46, EKB-569, erlotinib (Tarceva®), gefitinib (Iressa®), lapatinib (Tykerb®) and those compounds that are generically and specifically disclosed in WO 97/02266, EP 0 564 409, WO 99/03854, EP 0 520 722, EP 0 566 226, EP 0 787 722, EP 0 837 063, U.S. Pat. No. 5,747,498, WO 98/10767, WO 97/30034, WO 97/49688, WO 97/38983 and WO 96/33980.

farnesyltransferase inhibitors which include without limitation a-hydroxyfarnesylphosphonic acid, butanoic acid, 2-[(2S)-2-[(2S,3S)-2-[[[(2R)-2-amino-3-mercaptopropyl]amino]-3-methylpent-yl]oxy]-1-oxo-3-phenylpropyl]amino]-4-(methylsulfonyl)-1-methylethylester (2S)-(9CI), tipifarnib (Zarnestra®), and manumycin A.

Flk-1 kinase inhibitors which include without limitation 2-propenamide, 2-cyano-3-[4-hydroxy-3,5-bis(1-methyl-ethyl)phenyl]-N-(3-phenylpropyl)-(2E)-(9CI).

glycogen synthase kinase-3 (GSK3) inhibitors which include without limitation indirubin-3'-monooxime.

histone deacetylase (HDAC) inhibitors which include without limitation suberoylanilide hydroxamic acid (SAHA), [4-(2-amino-phenylcarbamoyl)-benzyl]-carbamic acid pyridine-3-ylmethyl ester and its derivatives, butyric acid, pyroxamide, trichostatin A, oxamflatin, apicidin, depeptide, depudecin, trapoxin, vorinostat (Zolinza®), and compounds disclosed in WO 02/22577.

I-kappa B-alpha kinase inhibitors (IKK) which include without limitation 2-propenenitrile, 3-[(4-methylphenyl)sulfonyl]-(2E)-(9CI).

imidazotetrazinones which include without limitation temozolomide (Methazolastone®, Temodar® and its derivatives (e.g., as disclosed generically and specifically in U.S. Pat. No. 5,260,291) and Mitozolomide.

insulin tyrosine kinase inhibitors which include without limitation hydroxyl-2-naphthalenylmethylphosphonic acid.

c-Jun-N-terminal kinase (JNK) inhibitors which include without limitation pyrazoleanthrone and epigallocatechin gallate.

mitogen-activated protein kinase (MAP) inhibitors which include without limitation benzenesulfonamide, N-[2-[[[3-(4-chlorophenyl)-2-propenyl]methyl]amino]methyl]phenyl]-N-(2-hydroxyethyl)-4-methoxy-(9CI).

MDM2 inhibitors which include without limitation trans-4-iodo, 4'-boranyl-chalcone.

MEK inhibitors which include without limitation butanedinitrile, bis[amino[2-aminophenyl]thio]methylene]-(9CI).

MMP inhibitors which include without limitation Actinonin, epigallocatechin gallate, collagen peptidomimetic and non-peptidomimetic inhibitors, tetracycline derivatives marimastat (Marimastat®), prinomastat, incyclinide (Metastat®), shark cartilage extract AE-941 (Neovastat®), Tanomastat, TAA211, MMI270B or AAJ996.

mTor inhibitors which include without limitation rapamycin (Rapamune®), and analogs and derivatives thereof, AP23573 (also known as ridaforolimus, deforolimus, or MK-8669), CCI-779 (also known as temsirolimus) (Torisel®) and SDZ-RAD.

NGFR tyrosine kinase inhibitors which include without limitation tyrphostin AG 879.

p38 MAP kinase inhibitors which include without limitation Phenol, 4-[4-(4-fluorophenyl)-5-(4-pyridinyl)-1H-imidazol-2-yl]-(9CI), and benzamide, 3-(dimethylamino)-N-[3-[(4-hydroxybenzoyl)amino]-4-methylphenyl]-(9CI).

p56 tyrosine kinase inhibitors which include without limitation damnacanthal and tyrphostin 46.

PDGF pathway inhibitors which include without limitation tyrphostin AG 1296, tyrphostin 9, 1,3-butadiene-1,1,3-tricarbonitrile, 2-amino-4-(1H-indol-5-yl)-(9CI), imatinib (Gleevec®) and gefitinib (Iressa®) and those compounds generically and specifically disclosed in European Patent No.: 0 564 409 and PCT Publication No.: WO 99/03854.

phosphatidyl inositol 3-kinase inhibitors which include without limitation wortmannin, and quercetin dihydrate.

phosphatase inhibitors which include without limitation cantharidic acid, cantharidin, and L-leucinamide.

protein phosphatase inhibitors which include without limitation cantharidic acid, cantharidin, L-P-bromotetramisole oxalate, 2(5H)-furanone, 4-hydroxy-5-(hydroxymethyl)-3-(1-oxohexadecyl)-(5R)-(9CI) and benzylphosphonic acid.

PKC inhibitors which include without limitation 1-H-pyrrolo-2,5-dione, 3-[1-3-(dimethylamino)propyl]-1H-indol-3-yl]-4-(1H-indol-3-yl)-(9CI), Bisindolylmaleimide IX, Sphingosine, staurosporine, and Hypericin.

PKC delta kinase inhibitors which include without limitation rottlerin.

polyamine synthesis inhibitors which include without limitation DMFO.

PTP1B inhibitors which include without limitation L-leucinamide.

protein tyrosine kinase inhibitors which include, without limitation tyrphostin Ag 216, tyrphostin Ag 1288, tyrphostin Ag 1295, geldanamycin, genistein and 7H-pyrrolo[2,3-d]pyrimidine derivatives as generically and specifically described in PCT Publication No.: WO 03/013541 and U.S. Publication No.: 2008/0139587.

SRC family tyrosine kinase inhibitors which include without limitation PP1 and PP2.

Syk tyrosine kinase inhibitors which include without limitation piccatanol.

Janus (JAK-2 and/or JAK-3) tyrosine kinase inhibitors which include without limitation tyrphostin AG 490 and 2-naphthyl vinyl ketone.

retinoids which include without limitation isotretinoin (Accutane®, Amnesteem®, Cistane®, Claravis®, Sotret®) and tretinoin (Aberel®, Aknoten®, Avita®, Renova®, Retin-A®, Retin-A MICRO®, Vesanoid®).

RNA polymerase II elongation inhibitors which include without limitation 5,6-dichloro-1-beta-D-ribofuranosylbenzimidazole.

serine/Threonine kinase inhibitors which include without limitation 2-aminopurine.

sterol biosynthesis inhibitors which include without limitation squalene epoxidase and CYP2D6.

VEGF pathway inhibitors, which include without limitation anti-VEGF antibodies, e.g., bevacizumab, and small molecules, e.g., sunitinib (Sutent®), sorafenib (Nexavar®), ZD6474 (also known as vandetanib) (Zactima™), SU6668, CP-547632 and AZD2171 (also known as cediranib) (Recentin™).

Examples of chemotherapeutic agents are also described in the scientific and patent literature, see, e.g., Bulinski (1997) J. Cell Sci. 110:3055-3064; Panda (1997) Proc. Natl. Acad. Sci. USA 94:10560-10564; Muhlradt (1997) Cancer Res. 57:3344-3346; Nicolaou (1997) Nature 387:268-272; Vasquez (1997) Mol. Biol. Cell. 8:973-985; Panda (1996) J. Biol. Chem. 271:29807-29812.

Other exemplary anti-cancer agents include alitretinon, altretamine, aminopterin, aminolevulinic acid, amsacrine (Amsidine®), asparaginase (crisantaspase, Erwinase®), atrasentan, bexarotene (Targretin®), carboquone, demecolcine, efaproxiral, elsamitucin, etoglucid, ferrocene, Gliadel implants, hydroxycarbamide, leucovorin, lonidamine, lucanthone, masoprocil, methyl aminolevulinate, mitoguanzone, mitotane (Lysodren®), oblimersen, omacetaxine (Genasense®), pegaspargase (Oncaspar®), porfimer sodium (Photofrin®), prednimustine, sitimagene ceradenovec (Cerepro®), talaporfin, temoporfin, trabectedin (Yondelis®), and verteporfin.

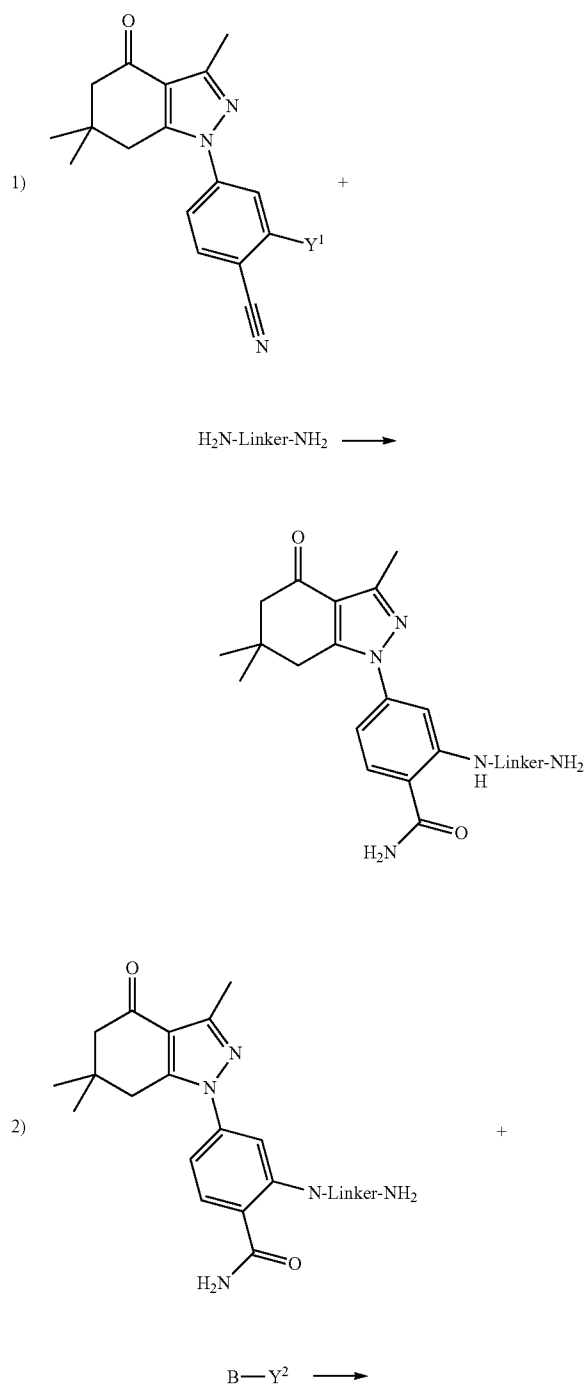
It will be understood by the skilled artisan that when B comprises an anti-cancer agent, the anti-cancer agent may be attached to the linker group of the compound of formula (I) by any suitable linkage. For example, if the anti-cancer agent has a reactive moiety such as a hydroxyl group, an amine group, or the like, such groups may be used to attach the anti-cancer agent to the remainder of the compound of formula (I). Alternatively, an anti-cancer agent may be

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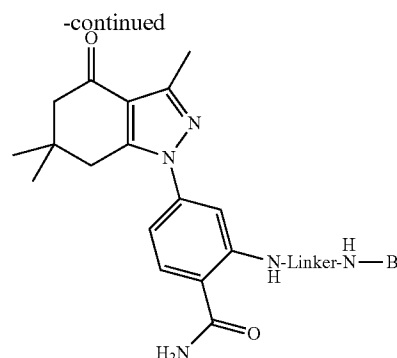
further derivatized with an atom or a group of atoms in order to provide a reactive group. In such instances, it is understood that the group -B corresponding to the anti-cancer agent includes not only the anti-cancer agent itself but also the linking atoms.

f. Preparation of Compounds

Compounds described herein may be prepared according to a variety of methods. A representative synthesis of exemplary compounds of formula (I) is illustrated in Scheme 1. Scheme 1 Exemplary Synthesis



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In reaction 1 of Scheme 1, the starting material includes an exemplary Hsp90-binding moiety precursor, and the group Y^1 is a leaving group or a reactive group. For example and as illustrated in Scheme 1, Y^1 may be leaving group such as a halogen, such that the compound $H_2N-Linker-NH_2$ may react with the compound via a reaction such as nucleophilic aromatic substitution. The nitrile can then be subsequently hydrolyzed to produce the Hsp90-binding moiety.

In reaction 2 of Scheme 1, the product of the initial reaction is further reacted with a compound $B-Y^2$, where B comprises a detection moiety, an anti-cancer agent or an Hsp90 binding component, and Y^2 is a leaving group or a reactive group. For example, Y^2 may be reactive group such as an isothiocyanate or a maleimide.

As can be appreciated by the skilled artisan, further methods of synthesizing the compounds of the formulae herein will be evident to those of ordinary skill in the art. Additionally, the various synthetic steps may be performed in an alternate sequence or order to give the desired compounds. Synthetic chemistry transformations and protecting group methodologies (protection and deprotection) useful in synthesizing the compounds described herein are known in the art and include, for example, those such as described in R. Larock, Comprehensive Organic Transformations, VCH Publishers (1989); T. W. Greene and P. G. M. Wuts, Protective Groups in Organic Synthesis, 2d. Ed., John Wiley and Sons (1991); L. Fieser and M. Fieser, Fieser and Fieser's Reagents for Organic Synthesis, John Wiley and Sons (1994); and L. Paquette, ed., Encyclopedia of Reagents or Organic Synthesis, John Wiley and Sons (1995), and subsequent editions thereof.

g. Evaluation of Compounds

Compounds can be evaluated by determining their ability to bind to Hsp90. The selectivity for Hsp90 over other proteins, such as GRP94 and TRAP1, may also be determined.

For example, prior to attachment of a detection moiety, an A-Linker- NH_2 compound can be bound to Sepharose resin. Such a resin can be exposed to a pig mammary gland extract, which is a tissue known to be high in ATP binding proteins including native forms of Hsp90, GRP94 and TRAP1. Media can be incubated with extract, washed stringently, and bound proteins removed with SDS and subsequently characterized. Preincubation of the resin with a known Hsp90 inhibitor should selectively block binding of Hsp90 and associated proteins, while those recovered in the presence of such a quenching agent are likely to be non-specifically bound.

Compounds of formula (I) may have increased selectivity for Hsp90 compared to proteins that share sequence homology with Hsp90. For example, the compounds may bind to

Hsp90 with an affinity that is at least about 100 to 1000 fold higher than the affinity for GRP94, a molecular chaperone that functions in the processing and transport of secreted proteins. The Hsp90 affinity probe may bind to Hsp90 with an affinity that is at least about 100 fold to 1000-fold higher than the affinity for TRAP1, (TNF receptor-associated protein 1), a heat shock protein of 75 kDa. This increased selectivity may lead to more accurate detection and targeting of Hsp90.

3. Methods of Use

Compounds of formula (I) may be used in a variety of methods, such as methods of detecting Hsp90 in a sample, methods of detecting cancer in a subject, and methods of treating cancer in a subject.

a. Methods of Detecting Hsp90

In some embodiments, compounds described herein can be used in methods of detecting Hsp90 in a sample, the methods comprising:

- a) contacting the sample with a compound having the following formula (Ia):



wherein:

A is an Hsp90 binding component;

X¹ is selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

X² is selected from the group consisting of —NR—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety; and

- b) detecting a signal from the detection moiety.

In some embodiments, the sample is an in vitro sample, such as a cell or tissue extract. In some embodiments, the sample is a cell culture. For example, the sample may be a culture of cells such as cancer cells.

In some embodiments, the sample is a biological sample from a subject, such as a human. In some embodiments, the biological sample is selected from the group consisting of a tissue sample, bodily fluid, whole blood, plasma, serum, urine, bronchoalveolar lavage fluid, and a cell culture suspension or fraction thereof. In embodiments in which Hsp90 is detected in a biological sample from a subject, the methods may further involve providing or obtaining a biological sample from the subject, which can be obtained by any known means including needle stick, needle biopsy, swab, and the like. In an embodiment of such methods, the biological sample is a blood sample, such as a blood plasma or serum sample; which may be obtained by any standard technique such as, for example, by venipuncture. Biological samples used in the methods may be stored or banked under suitable tissue storage conditions, or can be accessed from samples that have been previously stored or banked under suitable conditions.

Following contacting the sample with a compound, the method further includes detecting a signal from the detection moiety. A signal may be detected by any suitable means appropriate for the particular detection moiety being used. For example, when the detection moiety comprises a fluorophore, a signal may be detected using a fluorometer or a fluorescence plate reader, or by using fluorescence techniques such as fluorescence microscopy, fluorescence resonance energy transfer, flow cytometry and fluorescence-

activated cell sorting. In embodiments in which the detection moiety comprises a radioisotope, a signal may be detected using scintillation counting or radioimaging techniques.

A signal from the detection moiety may be quantitated, for example, by comparing the quantity of the signal to that of a reference sample.

b. Methods of Detecting Cancer

In some embodiments, compounds described herein can be used in a method of detecting cancer in a subject, the method comprising:

- a) contacting a biological sample from the subject with a compound having the following formula (Ia):



wherein:

A is an Hsp90 binding component;

X¹ is selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

X² is selected from the group consisting of —NR—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety; and

- b) detecting a signal from the detection moiety;

wherein cancer is detected in the sample when the signal from the detection moiety is higher relative to a signal from a reference sample.

In some embodiments, the method further comprises obtaining the biological sample from the subject, such as a biological sample described herein, according to methods described herein. In some embodiments, the subject is a human.

A reference sample may be a sample from a healthy subject, i.e. a subject having no clinical signs or symptoms of cancer. Suitably, the healthy subject may be clinically evaluated for otherwise undetected signs or symptoms of cancer, which evaluation may include routine physical examination and/or laboratory testing.

In embodiments, the cancer may be any type of cancer, such as a cancer recognized by the National Cancer Institute. In embodiments, the cancer may be a type of cancer associated with elevated levels of Hsp90. Exemplary types of cancers include the following:

Digestive/gastrointestinal cancers such as anal cancer; bile duct cancer; extrahepatic bile duct cancer; appendix cancer; carcinoid tumor, gastrointestinal cancer; colon cancer, colorectal cancer including childhood colorectal cancer; esophageal cancer including childhood esophageal cancer; gallbladder cancer; gastric (stomach) cancer including childhood gastric (stomach) cancer, hepatocellular (liver) cancer including adult (primary) hepatocellular (liver) cancer and childhood (primary) hepatocellular (liver) cancer; pancreatic cancer including childhood pancreatic cancer; sarcoma, rhabdomyosarcoma; islet cell pancreatic cancer; rectal cancer; and small intestine cancer;

Breast cancer, including childhood breast cancer, male breast cancer and breast cancer during pregnancy;

Genitourinary cancers such as bladder cancer including childhood bladder cancer, renal cell (kidney) cancer; ovarian cancer including childhood ovarian cancer; ovarian epithelial cancer; ovarian low malignant potential tumor; penile cancer; prostate cancer; renal cell cancer including child-

hood renal cell cancer; renal pelvis and ureter, transitional cell cancer; testicular cancer; urethral cancer; vaginal cancer; vulvar cancer, cervical cancer; Wilms tumor and other childhood kidney tumors; endometrial cancer, and gestational trophoblastic tumor;

Lung cancer such as non-small cell lung cancer; and small cell lung cancer;

Respiratory cancers such as malignant mesothelioma, adult; malignant mesothelioma, childhood; malignant thymoma; childhood thymoma; thymic carcinoma; bronchial adenomas/carcinoids including childhood bronchial adenomas/carcinoids; pleuropulmonary blastoma; non-small cell lung cancer; and small cell lung cancer; and

Skin cancers such as Kaposi's sarcoma; Merkel cell carcinoma; melanoma; and childhood skin cancer.

In suitable embodiments, the cancer may be a cancer that is associated with increased levels of Hsp90, including but not limited to breast cancer, prostate cancer, non-small cell lung cancer, colorectal cancer, pancreatic cancer, ovarian cancer and melanoma. In particular embodiments, the cancer is breast cancer or prostate cancer.

c. Methods of Treating Cancer

In some embodiments, compounds described herein can be used in a method of treating cancer in a subject in need of treatment. Such methods comprise administering the subject a therapeutically effective amount of a compound having the following formula (Ib):



wherein:

A is an Hsp90 binding component;

X¹ and X² are each independently selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen; and

B is selected from the group consisting of an anti-cancer agent and an Hsp90 binding component.

In embodiments, the cancer is a cancer described herein. In embodiments, the cancer is selected from the group consisting of breast cancer, prostate cancer, non-small cell lung cancer, colorectal cancer, pancreatic cancer, ovarian cancer and melanoma. In particular embodiments, the cancer is breast cancer or prostate cancer.

In embodiments, the anti-cancer agent is an anti-cancer agent described herein. In embodiments, B is an anti-cancer agent selected from the group consisting of a protein kinase inhibitor, a protein phosphatase inhibitor and a histone deacetylase inhibitor.

In the methods of treating cancer, a compound, or a pharmaceutical composition comprising the compound, may be administered to the subject by any convenient route of administration, whether systemically/peripherally or at the site of desired action, including but not limited to, oral (e.g. by ingestion); topical (including e.g. transdermal, intranasal, ocular, buccal, and sublingual); pulmonary (e.g. by inhalation or insufflation therapy using, e.g. an aerosol, e.g. through mouth or nose); rectal; vaginal; parenteral, for example, by injection, including subcutaneous, intradermal, intramuscular, intravenous, intraarterial, intracardiac, intrathecal, intraspinal, intracapsular, subcapsular, intraorbital, intraperitoneal, intratracheal, subcuticular, intraarticular, subarachnoid, and intrasternal; by implant of a depot, for example, subcutaneously or intramuscularly. Additional modes of administration may include adding the compound

and/or a composition comprising the compound to a food or beverage, including a water supply for an animal, to supply the compound as part of the animal's diet.

While it is possible for the compound to be administered alone, in some embodiments the compound may be presented as a pharmaceutical composition (e.g., formulation) comprising at least one compound, as defined above, together with one or more pharmaceutically-acceptable carriers, adjuvants, excipients, diluents, fillers, buffers, stabilizers, preservatives, lubricants, or other materials well known to those skilled in the art and optionally other therapeutic or prophylactic agents.

Thus, the disclosure further provides pharmaceutical compositions, as defined above, and methods of making a pharmaceutical composition comprising admixing; at least one compound, as defined above, together with one or more pharmaceutically acceptable carriers, excipients, buffers, adjuvants, stabilizers, or other materials, as described herein.

Suitable carriers, excipients, etc. can be found in standard pharmaceutical texts, for example Remington's Pharmaceutical Sciences, 18th edition, Mack Publishing Company, Easton, Pa., 1990.

The formulations may conveniently be presented in unit dosage form and may be prepared by any method well known in the art of pharmacy. Such methods include the step of bringing into association the compound with the carrier which constitutes one or more accessory ingredients. In general, the formulations may be prepared by uniformly and intimately bringing into association the compound with liquid carriers or finely divided solid carriers or both, and then if necessary shaping the product.

Formulations may be in the form of liquids, solutions, suspensions, emulsions, elixirs, syrups, tablets, lozenges, granules, powders, capsules, cachets, pills, ampoules, suppositories, pessaries, ointments, gels, pastes, creams, sprays, mists, foams, lotions, oils, boluses, electuaries, or aerosols.

Formulations suitable for oral administration (e.g., by ingestion) may be presented as discrete units such as capsules, cachets or tablets, each containing a predetermined amount of the compound; as a powder or granules; as a solution or suspension in an aqueous or non-aqueous liquid; or as an oil-in-water liquid emulsion or a water-in-oil liquid emulsion; as a bolus; as an electuary; or as a paste.

A tablet may be made by conventional means, e.g., compression or molding, optionally with one or more accessory ingredients. Compressed tablets may be prepared by compressing in a suitable machine the compound in a free-flowing form such as a powder or granules, optionally mixed with one or more binders (e.g., povidone, gelatin, acacia, sorbitol, tragacanth, hydroxypropylmethyl cellulose); fillers or diluents (e.g., lactose, microcrystalline cellulose, calcium hydrogen phosphate); lubricants (e.g., magnesium stearate, talc, silica); disintegrants (e.g., sodium starch glycolate, cross-linked povidone, cross-linked sodium carboxymethyl cellulose); surface-active or dispersing or wetting agents (e.g., sodium lauryl sulfate); and preservatives (e.g., methyl p-hydroxybenzoate, propyl p-hydroxybenzoate, sorbic acid). Molded tablets may be made by molding in a suitable machine a mixture of the powdered compound moistened with an inert liquid diluent. The tablets may optionally be coated or scored and may be formulated so as to provide slow or controlled release of the compound therein using, for example, hydroxypropylmethyl cellulose in varying proportions to provide the desired release profile. Tablets may optionally be provided with an enteric coating, to provide release in parts of the gut other than the stomach.

Formulations suitable for topical administration (e.g., transdermal, intranasal, ocular, buccal, and sublingual) may be formulated as an ointment, cream, suspension, lotion, powder, solution, past, gel, spray, aerosol, or oil. Alternatively, a formulation may comprise a patch or a dressing such as a bandage or adhesive plaster impregnated with compounds and optionally one or more excipients or diluents. In addition, a formulation may be added to a conventional bandage, e.g. to a gauze portion that contacts a wound, as an antimicrobial agent.

Formulations suitable for topical administration in the mouth include lozenges comprising the compound in a flavored basis, usually sucrose and acacia or tragacanth; pastilles comprising the compound in an inert basis such as gelatin and glycerin, or sucrose and acacia; and mouthwashes comprising the compound in a suitable liquid carrier.

Formulations suitable for topical administration to the eye also include eye drops wherein the compound is dissolved or suspended in a suitable carrier, especially an aqueous solvent for the compound.

Formulations suitable for nasal administration, wherein the carrier is a solid, include a coarse powder having a particle size, for example, in the range of about 20 to about 500 microns which is administered in the manner in which snuff is taken, i.e., by rapid inhalation through the nasal passage from a container of the powder held close up to the nose. Suitable formulations wherein the carrier is a liquid for administration as, for example, nasal spray, nasal drops, or by aerosol administration by nebulizer, include aqueous or oily solutions of the compound.

Formulations suitable for administration by inhalation include those presented as an aerosol spray from a pressurized pack, with the use of a suitable propellant, such as dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide, or other suitable gases.

Formulations suitable for topical administration via the skin include ointments, creams, and emulsions. When formulated in an ointment, the compound may optionally be employed with either a paraffinic or a water-miscible ointment base. Alternatively, the compounds may be formulated in a cream with an oil-in-water cream base. If desired, the aqueous phase of the cream base may include, for example, at least about 30% w/w of a polyhydric alcohol, i.e., an alcohol having two or more hydroxyl groups such as propylene glycol, butane-1,3-diol, mannitol, sorbitol, glycerol and polyethylene glycol and mixtures thereof. The topical formulations may desirably include a compound which enhances absorption or penetration of the compound through the skin or other affected areas. Examples of such dermal penetration enhancers include dimethylsulfoxide and related analogues.

When formulated as a topical emulsion, the oily phase may optionally comprise merely an emulsifier (otherwise known as an emulgent), or it may comprises a mixture of at least one emulsifier with a fat or an oil or with both a fat and an oil. Preferably, a hydrophilic emulsifier is included together with a lipophilic emulsifier which acts as a stabilizer. It is also preferred to include both an oil and a fat. Together, the emulsifier(s) with or without stabilizer(s) make up the so-called emulsifying wax, and the wax together with the oil and/or fat make up the so-called emulsifying ointment base which forms the oily dispersed phase of the cream formulations.

Suitable emulgents and emulsion stabilizers include Tween 60, Span 80, cetostearyl alcohol, myristyl alcohol, glyceryl monostearate and sodium lauryl sulfate. The choice of suitable oils or fats for the formulation is based on

achieving the desired cosmetic properties, since the solubility of the compound in most oils likely to be used in pharmaceutical emulsion formulations may be very low. Thus the cream should preferably be a non-greasy, non-staining and washable product with suitable consistency to avoid leakage from tubes or other containers. Straight or branched chain, mono- or dibasic alkyl esters such as diisoadipate, isocetyl stearate, propylene glycol diester of coconut fatty acids, isopropyl myristate, decyl oleate, isopropyl palmitate, butyl stearate, 2-ethylhexyl palmitate or a blend of branched chain esters known as Crodamol CAP may be used, the last three being preferred esters. These may be used alone or in combination depending on the properties required. Alternatively, high melting point lipids such as white soft paraffin and/or liquid paraffin or other mineral oils can be used.

Formulations suitable for rectal administration may be presented as a suppository with a suitable base comprising, for example, cocoa butter or a salicylate.

Formulations suitable for vaginal administration may be presented as pessaries, tampons, creams, gels, pastes, foams or spray formulations containing in addition to the compound, such carriers as are known in the art to be appropriate.

Formulations suitable for parenteral administration (e.g., by injection, including cutaneous, subcutaneous, intramuscular, intravenous and intradermal), include aqueous and non-aqueous isotonic, pyrogen-free, sterile injection solutions which may contain anti-oxidants, buffers, preservatives, stabilizers, bacteriostats in addition to the compound, and solutes which render the formulation isotonic with the blood of the intended recipient; and aqueous and non-aqueous sterile suspensions which may include suspending agents and thickening agents, and liposomes or other microparticulate systems which are designed to target the compound to blood components or one or more organs. Examples of suitable isotonic vehicles for use in such formulations include Sodium Chloride Injection, Ringer's Solution, or Lactated Ringer's Injection. Typically, the concentration of the compound in the solution is from about 1 ng/ml to about 1 µg/ml, although other concentrations are possible and are encompassed within the invention. The formulations may be presented in unit-dose or multi-dose scaled containers, for example, ampoules and vials, and may be stored in a freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid carrier, for example water for injections, immediately prior to use. Extemporaneous injection solutions and suspensions may be prepared from sterile powders, granules, and tablets; Formulations may be in the form of liposomes or other microparticulate systems which are designed to target the compound to blood components or one or more organs.

It will be appreciated that appropriate dosages of the compounds, and compositions comprising the compounds, can vary from patient to patient. Determining the optimal dosage will generally involve the balancing of the level of therapeutic benefit against any risk or deleterious side effects of the treatments of the present invention. The selected dosage level will depend on a variety of factors including, but not limited to, the activity of the particular compound, the route of administration, the time of administration, the rate of excretion of the compound, the duration of the treatment, other drugs, compounds, and/or materials used in combination, and the age, sex, weight, condition, general health, and prior medical history of the patient. The amount of compound and route of administration will ultimately be at the discretion of the physician, although generally the

dosage will be to achieve local concentrations at the site of action which achieve the desired effect without causing substantial harmful or deleterious side-effects.

Administration in vivo can be effected in one dose, continuously or intermittently (e.g., in divided doses at appropriate intervals) throughout the course of treatment. Methods of determining the most effective means and dosage of administration are well known to those of skill in the art and will vary with the formulation used for therapy, the purpose of the therapy, the target cell being treated, and the subject being treated. Single or multiple administrations can be carried out with the dose level and pattern being selected by the treating physician. In general, a suitable dose of the compound is in the range of about 100 µg to about 250 mg per kilogram body weight of the subject per day.

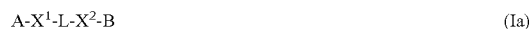
The composition may be administered once, on a continuous basis (e.g. by an intravenous drip), or on a periodic/intermittent basis, including about once per hour, about once per two hours, about once per four hours, about once per eight hours, about once per twelve hours, about once per day, about once per two days, about once per three days, about twice per week, about once per week, and about once per month. The composition may be administered until a desired reduction of symptoms is achieved.

The present compounds, compositions, and methods may be administered as part of a therapeutic regimen along with other treatments appropriate for the particular injury or disease being treated.

d. Methods of Detecting HIV

In some embodiments, compounds described herein can be used in a method of detecting Human Immunodeficiency Virus (HIV) in a subject, the method comprising:

- a) contacting a biological, sample from the subject with a compound having the following formula (Ia):



wherein:

A is an Hsp90 binding component;

X¹ is selected from the group consisting of —NH—, —O—, —S—, —C(O)— and —S(O)₂—;

X² is selected from the group consisting of —NR—, —O—, —S—, —C(O)— and —S(O)₂—;

L is a divalent linker comprising at least twelve member atoms independently selected from carbon, nitrogen and oxygen, wherein at least one member atom is nitrogen or oxygen;

B is a detection moiety; and

R is selected from the group consisting of hydrogen and a detection moiety; and

- b) detecting a signal from the detection moiety; wherein HIV is detected in the sample when the signal from the detection moiety is higher relative to a signal from a reference sample.

4. Kits

In another aspect, the disclosure provides a kit, which may be used for detecting Hsp90 in a sample, for detecting cancer in a sample, or for treating cancer in a subject.

A kit will include a compound of formula (I) as described herein. A kit may also include instructions for use of the compound of formula (I). Instructions included in kits can be affixed to packaging material or can be included as a package insert. While the instructions are typically written or printed materials they are not limited to such. Any medium capable of storing such instructions and communicating them to an end user is contemplated by this disclosure. Such media include, but are not limited to, electronic storage media (e.g., magnetic discs, tapes, cartridges, chips),

optical media (e.g., CD, DVD), and the like. As used herein, the term “instructions” can include the address of an internet site that provides the instructions.

In one embodiment, the disclosure provides a kit for detecting Hsp90 in a sample. The kit comprises at least one compound of formula (I), and instructions for assaying the test sample for Hsp90. For example, the kit can comprise instructions for assaying the test sample for Hsp90 by fluorescence detection. The kit may further comprise a calibrator or control, e.g., purified, and optionally lyophilized, (e.g., Hsp90), and/or at least one container (e.g., tube, microtiter plates or strips) for conducting the assay, and/or a buffer, such as an assay buffer or a wash buffer, either one of which can be provided as a concentrated solution. Preferably, the kit comprises all components, i.e., reagents, standards, buffers, diluents, etc., which are necessary to perform the assay. The instructions also can include instructions for generating a standard curve or a reference standard for purposes of quantifying Hsp90.

The kit can also optionally include other reagents required to conduct a diagnostic assay or facilitate quality control evaluations, such as buffers, salts, enzymes, enzyme cofactors, substrates, detection reagents, and the like. Other components, such as buffers and solutions for the isolation and/or treatment of a test sample (e.g., pretreatment reagents), also can be included in the kit. The kit can additionally include one or more other controls. One or more of the components of the kit can be lyophilized, in which case the kit can further comprise reagents suitable for the reconstitution of the lyophilized components.

The various components of the kit optionally are provided in suitable containers as necessary, e.g., a microtiter plate. The kit can further include containers for holding or storing a sample (e.g., a container or cartridge for a blood sample). Where appropriate, the kit optionally also can contain reaction vessels, mixing vessels, and other components that facilitate the preparation of reagents or the test sample. The kit can also include one or more instrument for assisting with obtaining a test sample, such as a syringe, pipette, forceps, measured spoon, or the like.

It will be readily apparent to those skilled in the art that other suitable modifications and adaptations of the compounds and methods of the present disclosure described herein are readily applicable and appreciable, and may be made using suitable equivalents without departing from the scope of the present disclosure or the aspects and embodiments disclosed herein. Having now described the present disclosure in detail, the same will be more clearly understood by reference to the following examples which are merely intended only to illustrate some aspects and embodiments of the disclosure, and should not be viewed as limiting to the scope of the disclosure; The disclosures of all journal references, U.S. patents and publications referred to herein are hereby incorporated by reference in their entireties.

EXAMPLES

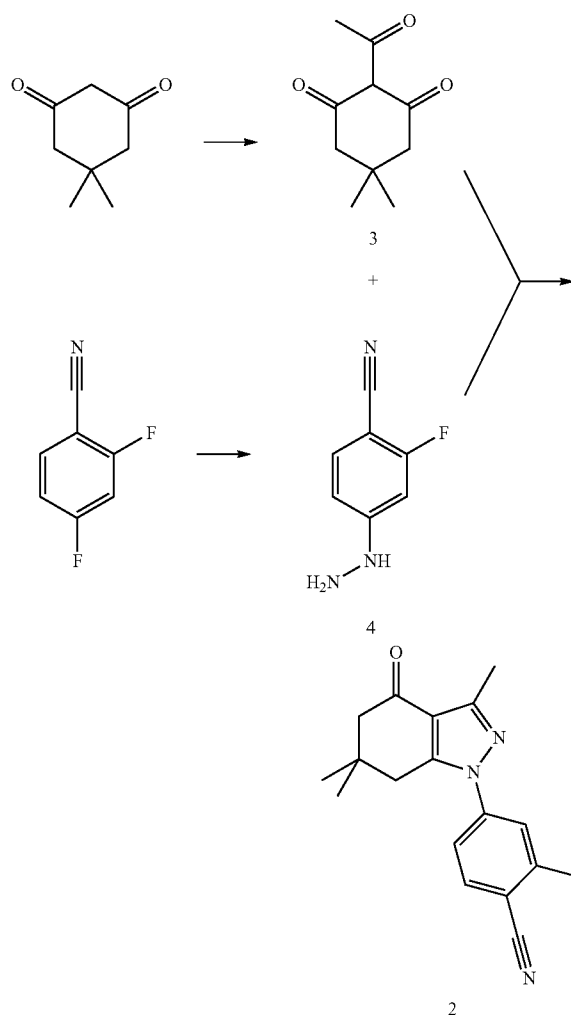
General Experimental and Analytical Details

Reagents were obtained from commercial sources and used without further purification. 1,19-Diamino-4,7,10,13,16-pentaaxanonadecane was obtained from Berry and Associates. Proton NMR spectra were obtained on Varian 400 and 500 MHz spectrometers. LC/MS were obtained on an Agilent ion-trap LC/MS system. HRMS results were obtained on an Agilent 6224 LCMS-TOF and are reported as an average of four runs.

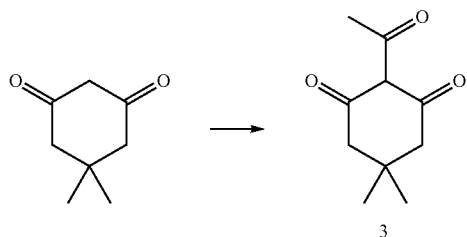
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Example 1

Synthesis of 2-Fluoro-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzonitrile (2)



2-Acetyl-5,5-dimethylcyclohexane-1,3-dione (3)

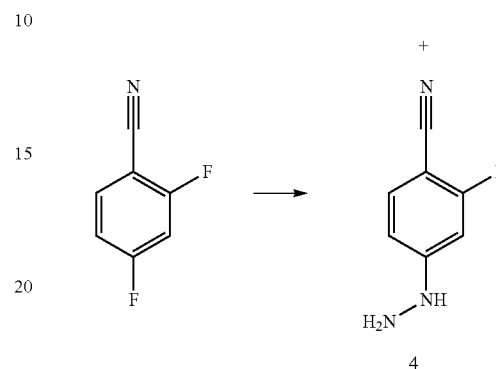


Dimedone (10 g, 71.3 mmol) was dissolved in methylene chloride (200 mL) and treated with Hunig's base (9.7 g, 74.9 mmol) and DMAP (440 mg, 3.6 mmol) followed by slow addition of acetic anhydride (7.65 g, 74.9 mmol). After 24 h, the mixture was concentrated, partitioned between hexanes

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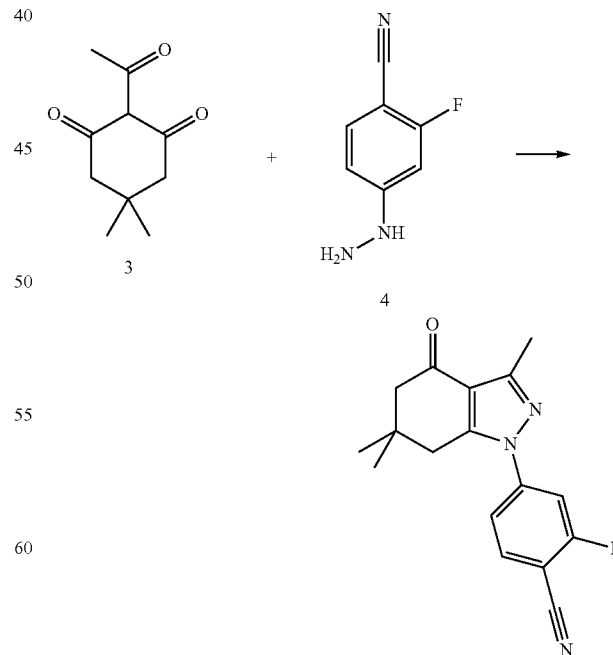
(150 mL) and 1 N HCl (70 mL), then washed with brine (50 mL), treated with Norit A and then dried (MgSO_4), filtered and concentrated to give 2-acetyl-5,5-dimethylcyclohexane-1,3-dione 3 (~11 g) as a yellow oil. The entire product was used in the next step.

2-Fluoro-4-hydrazinylbenzonitrile (4)



2,4-difluorobenzonitrile (10 g, 72 mmol) was dissolved in methanol (100 mL) and treated drop wise with hydrazine hydrate (18 g, 0.36 mol) and stirred at RT. After 16 h, the reaction mixture, containing a mixture of isomers, was concentrated then partitioned between ethyl acetate (100 mL), water (70 mL) and 1 N NaOH (30 mL). The organic layer was washed with brine (40 mL) then concentrated to give 2,4-difluorobenzonitrile (4) in a mixture of isomers as a white solid.

2-Fluoro-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzonitrile (2)

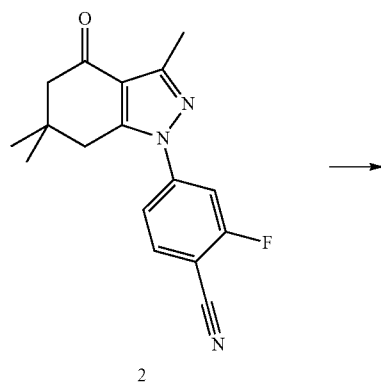


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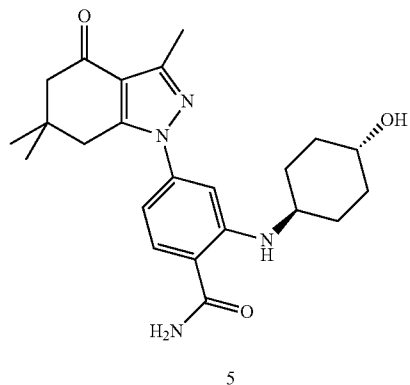
All of 3 (assume 13 g, 71.8 mmol) and 4 (assume 10.9 g, 71.8 mmol) were combined, then dissolved in methanol (40 mL) and treated with acetic acid (1 mL) and stirred at RT for 3 days. The mixture was concentrated and then dissolved in methylene chloride (20 ml). The mixture was chromatographed in a number of batches, with the best approach to load a DCE solution onto a dry column, wait a bit (maybe 15 to 20 min) then elute with 10% ethyl acetate in hexanes to remove the yellow (isomeric product) followed by elution with 20% to get the product. The chromatographed products were recrystallized from ethyl acetate/hexanes to give 2 (10.4 g, 49%). TLC (hexane/EtOAc: 60/40) R_f =0.47; ^1H NMR (CDCl_3) δ 7.73 (dd, J =7, 8.5 Hz, 1H), 7.49 (dd, J =2, 9.8 Hz, 1H), 7.45 (dd, J =2, 8.5 Hz, 1H), 2.90 (s, 2H) 2.54 (s, 3H) 2.43 (s, 2H), 1.14 (s, 6H); MS (ESI): m/z 298.2 $[\text{M}+\text{H}]^+$.

Example 2

Synthesis of 2-(((1r,4r)-4-Hydroxycyclohexyl) amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (5)



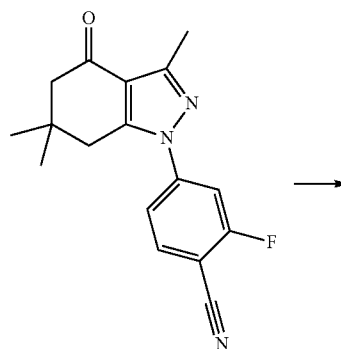
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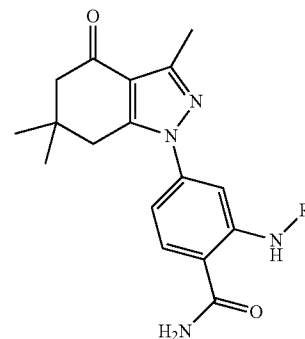
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A mixture of nitrile 2 (200 mg, 0.67 mmol) and trans-4-aminocyclohexanol (232 g, 2.0 mmol), Hunig's base (117 μL) and DMSO (300 μL) were heated to 90° C. for 30 m. The mixture diluted with ethanol (2 mL) and treated with 50% NaOH (5 drops) and then, very slowly, a drop at a time, with hydrogen peroxide. After each drop, the reaction foamed up a bit. After adding 5 drops over 10 m, the mixture was diluted with water (18 mL) and allowed to cool slowly with rapid stirring. After stirring overnight, the solid was filtered off to give the product 5 (251 mg, 91%) as a white powder. TLC (EtOAc) R_f =0.25; ^1H NMR (CDCl_3) δ 8.07 (d, J =7.2 Hz), 7.44 (d, J =8.6 Hz, 1H), 6.75 (d, J =2 Hz, 1H), 6.60 (dd, J =2, 8.6 Hz, 1H), 5.6 (brs, 2H), 3.71 (m, 1H), 3.35 (m, 1H), 2.80 (s, 2H) 2.53 (s, 3H) 2.38 (s, 2H), 2.13 (m, 2H), 2.02 (m, 2H), 1.64 (br s, 1H), 1.41 (m, 4H), 1.09 (s, 6H); MS (ESI): m/z 411.3 $[\text{M}+\text{H}]^+$.

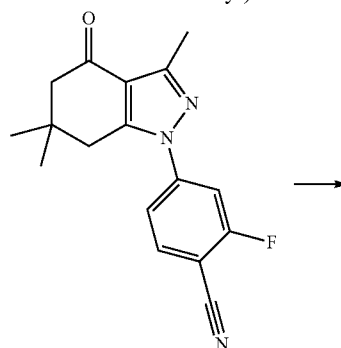
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Example 3
Attachment of Linkers

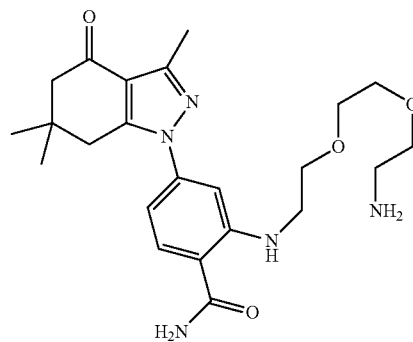
2

6 R = $-(\text{CH}_2)_2-\text{O}-\text{CH}_2\text{CH}_2\text{O}-(\text{CH}_2)_2-\text{NH}_2$ 7 R = $-(\text{CH}_2)_{10}-\text{NH}_2$ 8 R = $-(\text{CH}_2)_3-\text{O}-(\text{CH}_2\text{CH}_2\text{O})_4-(\text{CH}_2)_3-\text{NH}_2$ 9 R = $-(\text{CH}_2)_3-\text{O}-(\text{CH}_2\text{CH}_2\text{O})_2-(\text{CH}_2)_3-\text{NH}_2$

2-((2-(2-(2-aminoethoxy)ethoxy)ethyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (6)



2

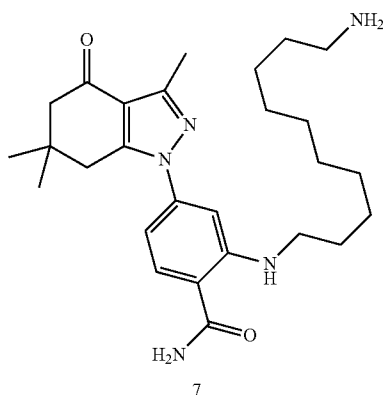


6

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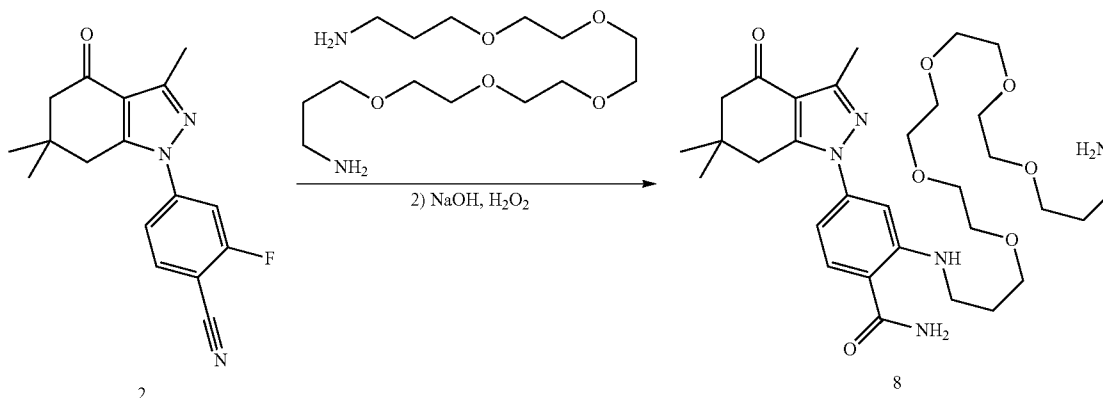
A mixture of 2 (500 mg, 1.7 mmol) and 2,2'-(ethylene-dioxy) bis(ethylamine) (1.2 g, 8.4 mmol) in DMSO (1 mL) were and heated to 90° C. for 30 minutes. The mixture was diluted with ethanol (2 mL) and, still at 90° C., treated with 50% NaOH (20 drops) and very slowly with 30% hydrogen peroxide (40 drops). The reaction mixture was diluted with methylene chloride and methanol and adsorbed onto silica gel. The mixture was chromatographed (silica gel, 1.5 cm×20 cm) and eluted with 9/1 CH₂Cl₂/MeOH, followed by 9/1/0.1 CH₂Cl₂/MeOH/NH₃ to give 6 (670 mg, 89%) as a clear glass. TLC (4/1/0.1 CH₂Cl₂/MeOH/NH₃) R_f=0.14; ¹H NMR (CDCl₃) δ 8.17 (t, J=5 Hz), 7.47 (d, J=8.4 Hz, 1H), 6.77 (d, J=2 Hz, 1H), 6.60 (dd, J=2, 8.4 Hz, 1H), 6.1 (br s, 2H), 3.73 (t, 3H), 3.65 (m, 4H), 3.50 (t, 2H), 3.36 (m, 2H), 2.85 (t, 2H), 2.77 (s, 2H) 2.51 (s, 3H) 2.36 (s, 2H), 1.07 (s, 6H); HRMS (ESI) [M+H]⁺ calcd for C₂₃H₃₄N₅O₄, 444.2605; found 444.2607.

2-((10-aminodecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (7)



Compound 7 was made in an analogous manner to 6. HRMS (ESI) [M+H]⁺ calcd for C₂₇H₄₂N₅O₂, 468.3333; found 468.3342.

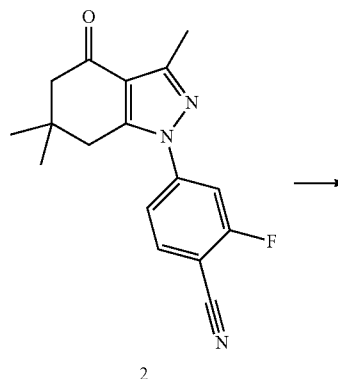
2-((19-Amino-4,7,10,13,16-pentaoxonadecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (8)



34

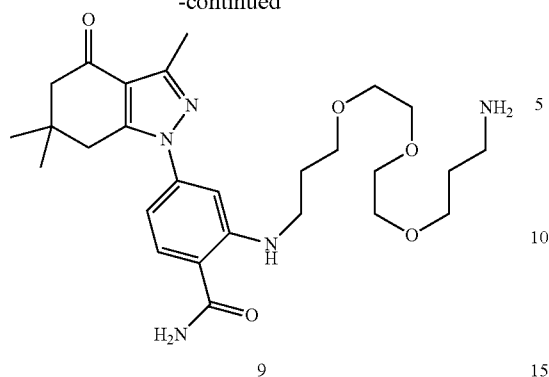
A mixture of 2 (482 mg, 1.62 mmol) and 1,19-diamino-4,7,10,13,16-pentaoxonadecane (1 g, 3.24 mol), diisopropylethylamine (628 mg, 4.8 mmol) and DMSO (1 mL) were heated to 90° C. for 20 m. Still at 90° C., the mixture was diluted with ethanol (2 mL) and treated with 50% NaOH (10 drops) and then, very slowly, a drop at a time, with hydrogen peroxide. After each drop, the reaction foamed up substantially. After about 10 drops over 10 m, the reaction mixture was diluted with ethanol and added to silica gel (6 g) and left overnight. The next day, the slurry was concentrated to a powder, added to a silica gel column (2.5×20 cm) and chromatographed with CH₂Cl₂ (300 mL), CH₂Cl₂/MeOH/NH₃ 19/0.9/0.1 (300 mL), 9/0.9/0.1 (300 mL) and 4/0.9/0.1 (500 mL). Fractions containing two by-products were set aside (see below). The cleanest fractions were combined to give 8 (600 mg, 61%) as a lightly yellow glass. TLC (4/1/0.1 CH₂Cl₂/MeOH/NH₃) R_f=0.30; ¹H NMR (CDCl₃) δ 7.98 (t, J=4 Hz), 7.47 (d, J=8.4 Hz, 1H), 6.77 (d, J=2 Hz, 1H), 6.60 (dd, J=2, 8.4 Hz, 1H), 6.0 (brs, 2H), 3.61 (mf 16H), 3.28 (m, 2H), 2.85 (t, 2H), 2.79 (s, 2H), 2.52 (s, 3H) 2.37 (s, 2H), 2.28 (brs, 2H), 1.94 (m, 2H), 1.76 (m, 2H), 1.07 (s, 6H); HRMS (ESI) [M+H]⁺ calcd for C₃₁H₅₀N₅O₇, 604.3705; found 604.3715.

2-((3-(2-(2-(3-aminopropoxy)ethoxy)ethoxy)propyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (9)



35

-continued



Compound 9 was prepared in the same way as compound 6. TLC (4/1/0.1 CH₂Cl₂/MeOH/NH₃) R_f=0.39; ¹H NMR

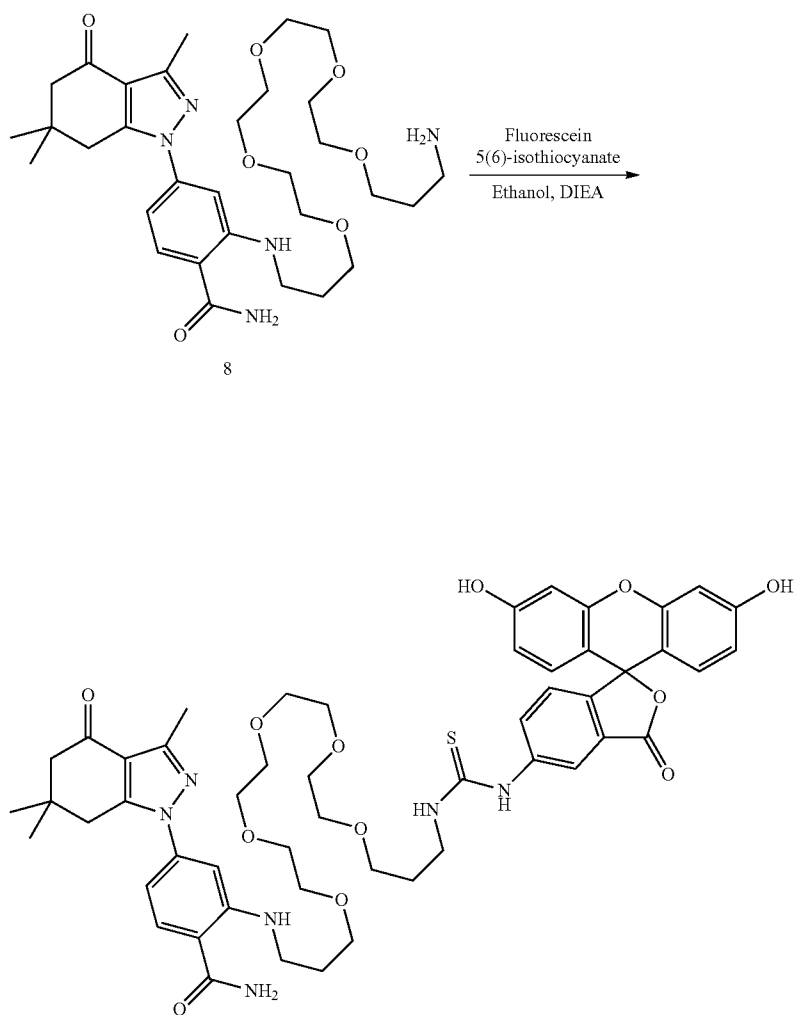
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(CDCl₃) δ 8.00 (t, J=5 Hz, 1H), 7.46 (d, J=8.2 Hz, 1H), 6.77 (s, 1H), 6.60 (d, J=8.4 Hz, 1H), 6.1 (brs, 2H), 3.59 (m, 8H), 3.29 (m, 2H), 2.82 (t, 2H), 2.79 (s, 2H), 2.59 (brs, 2H), 2.51 (s, 3H), 2.37 (s, 2H), 1.93 (M, 2H), 1.73 (m, 2H), 1.07 (s, 6H); HRMS (ESI) [M+H]⁺ calcd for C₂₇H₄₂N₅O₅, 516.3180; found 516.3191.

Example 4

Synthesis of Probe Compounds

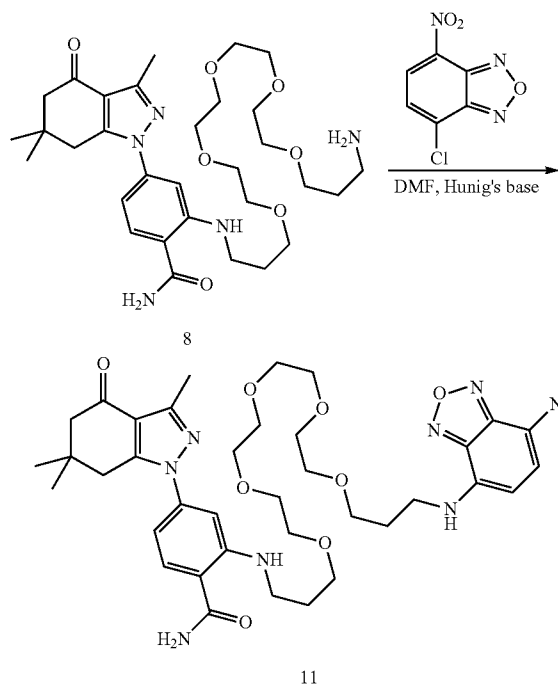
2-((1-((3',6'-dihydroxy-3-oxo-3H-spiro[isobenzofuran-1,9'-xanthen]-5-yl)amino)-1-thioxo-6,9,12,15,18-pentaoxa-2-azahenicosan-21-yl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (10)



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2-((1-((3',6'-dihydroxy-3-oxo-3H-spiro[isobenzofuran-1,9'-xanthen]-5-yl)amino)-1-thioxo-6,9,12,15,18-pentaoxa-2-azahenicosan-21-yl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (10). Fluorescein 5(6)-isothiocyanate (35 mg, 90 μ mol) was dissolved in methanol (10 mL) and treated with amine 8 (54.2 mg, 90 μ mol) followed by Hunig's base (35 mg, 270 μ mol) and stored in a drawer overnight. TLC ($\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{AcOH}$: 4/1/0.1) showed formation of a new product. The reaction mixture was concentrated to a glass and dissolved in DMSO/water (4/1, 2.5 mL). About half of the product was purified in two injections by chromatography (5 to 100% MeOH, Agilent C-18 21.2x250 mm) to give the product (45 mg) as a yellow solid. TLC (4/0.9/0.1 $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$) R_f =0.3; ^1H NMR ($\text{DMSO}-d_6$) δ 10.2 (br s, 2H), 8.40 (br t, 1H), 8.34 (br s, 1H), 8.23 (s, 1H), 7.92 (b s, 1H), 7.73 (d, J =8.1 Hz, 2H), 7.26 (brs, 1H), 7.15 (d, J =8.1 Hz, 1H), 6.76 (s, 1H), 6.67 (s, 1H), 6.65 (br s, 2H), 6.59 (d, J =8.5 Hz, 2H), 6.54 (d, J =8.5 Hz, 2H), 3.47 (m, 20H), 3.19 (m, 2H), 2.91 (s, 2H), 2.38 (s, 3H), 2.31 (s, 2H), 1.79 (m, 4H), 1.00 (s, 6H); MS (ESI): m/z 993.5 $[\text{M}+\text{H}]^+$.

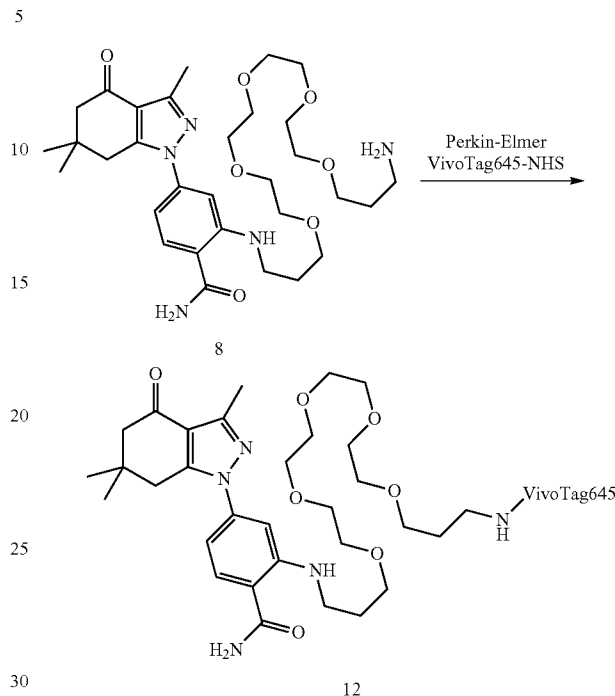
2-((19-((7-nitrobenzo[c][1,2,5]oxadiazol-4-yl)amino)-4,7,10,13,16-pentaoxonadecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (11)



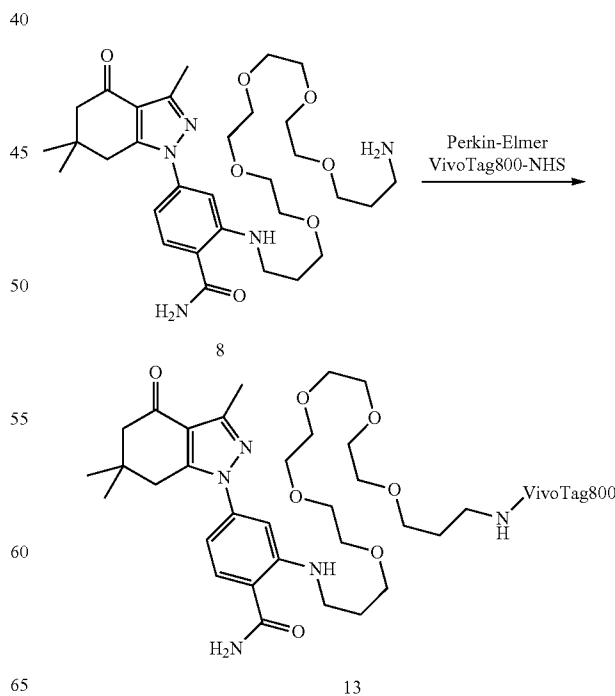
2-((19-((7-nitrobenzo[c][1,2,5]oxadiazol-4-yl)amino)-4,7,10,13,16-pentaoxonadecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (11). A mixture of Amine 8 (100 mg, 165 μ mol) and 4-chloro-7-nitro-1,2,3-benzoxadiazole (40 mg, 200 μ mol), Hunig's base (87 μ L) and DMF (1 mL) were mixed. The solution turned immediately much darker. TLC (9/1: $\text{CH}_2\text{Cl}_2/\text{MeOH}$) showed no starting materials and a major new yellow product, which changed color under UV light. The mixture was concentrated and then chromatographed with CH_2Cl_2 (200 mL), $\text{CH}_2\text{Cl}_2/\text{MeOH}$ 19/1 (300 mL), 9/1 (300 mL). The cleanest fractions were combined and then rechromato-

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graphed with CH_2Cl_2 (200 mL), $\text{CH}_2\text{Cl}_2/\text{MeOH}$ 19/1 (500 mL), 9/1 (300 mL), to give the product (11) (52 mg, 41%) as a dark brown oil. MS (ESI): m/z 767.4 $[\text{M}+\text{H}]^+$. VivoTag 645 Compound (12)



Amine 8 (4.5 mg, 7.45 μ mol) was dissolved in DMSO (200 μ L) and added to VivoTag 645 (Perkin Elmer, 2.5 mg, 1.8 μ mol) in its original container and placed back into the freezer overnight. The sample was purified in 3 portions by reverse-phase chromatography and concentrated to give VivoTag 645 compound (12) (~3 mg) as a blue solid. MS (ESI): m/z 1577.5 $[\text{M}]^+$. VivoTag 800 Compound (13)

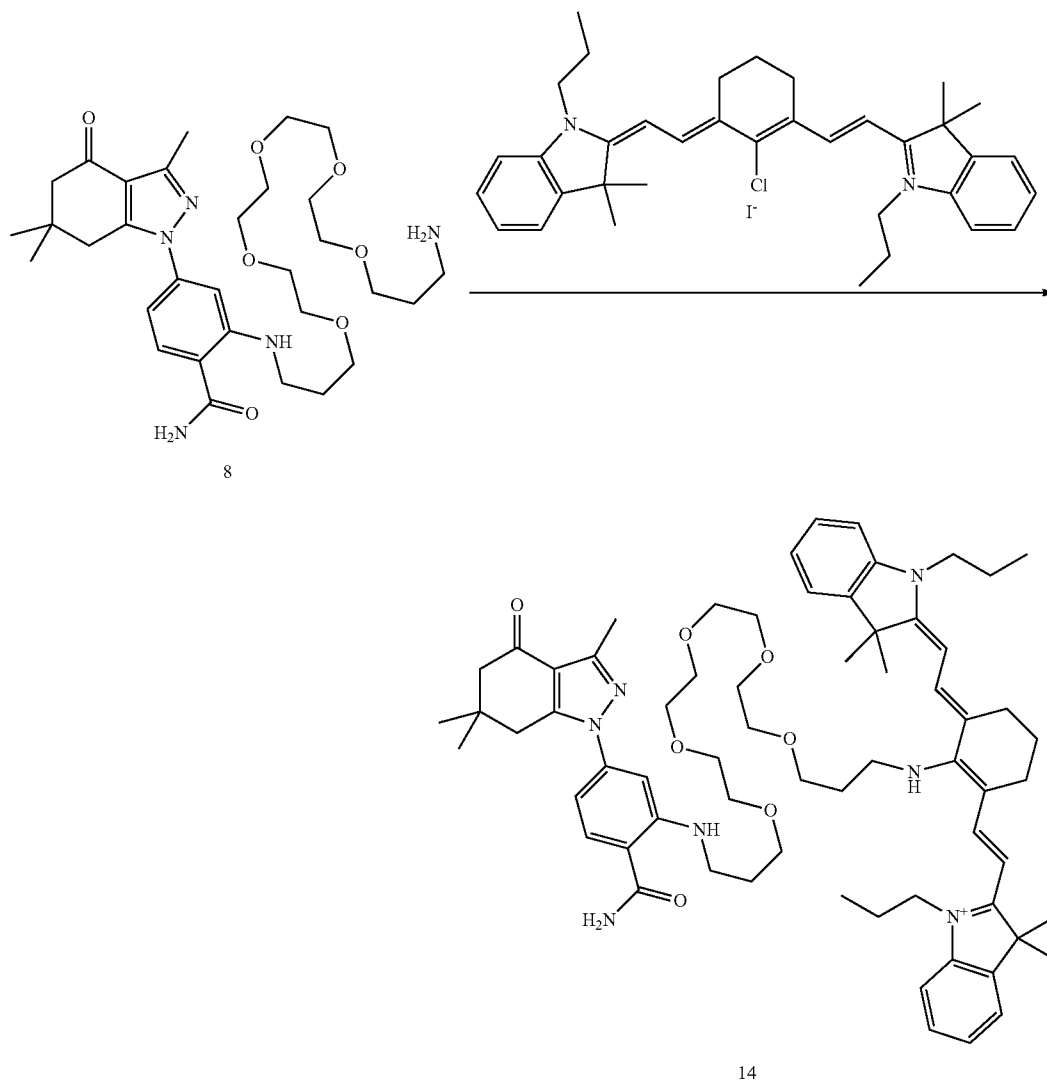


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VivoTag 800 compound (13). Amine 8 (4.3 mg, 7.2 μmol) was dissolved in DMSO (100 μL) and treated with Hunig's base (10 μL) and added to VivoTag 800 (Perkin Elmer, 5 mg, 3.42 μmol) in its original container. The starting material vial was washed twice with DMSO (100 μL), which was added to the reaction mixture. The mixture was left in an aluminum bag at RT for 30 m, then placed back into the freezer for the 3 d. The sample was purified in 3 portions by reverse-phase chromatography and concentrated to give VivoTag 800 compound (13) (~9 mg) as a blue solid. MS (ESI): m/z 823.7 $[\text{M}-2]^{2-}$.

2-((E)-2-((E)-2-((19-((2-carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-4,7,10,13,16-pentaoxononadecyl)amino)-3-((E)-2-(3,3-dimethyl-1-propylindolin-2-ylidene)ethylidene)cyclohex-1-en-1-yl)vinyl)-3,3-dimethyl-1-propyl-3H-indol-1-ium (14)



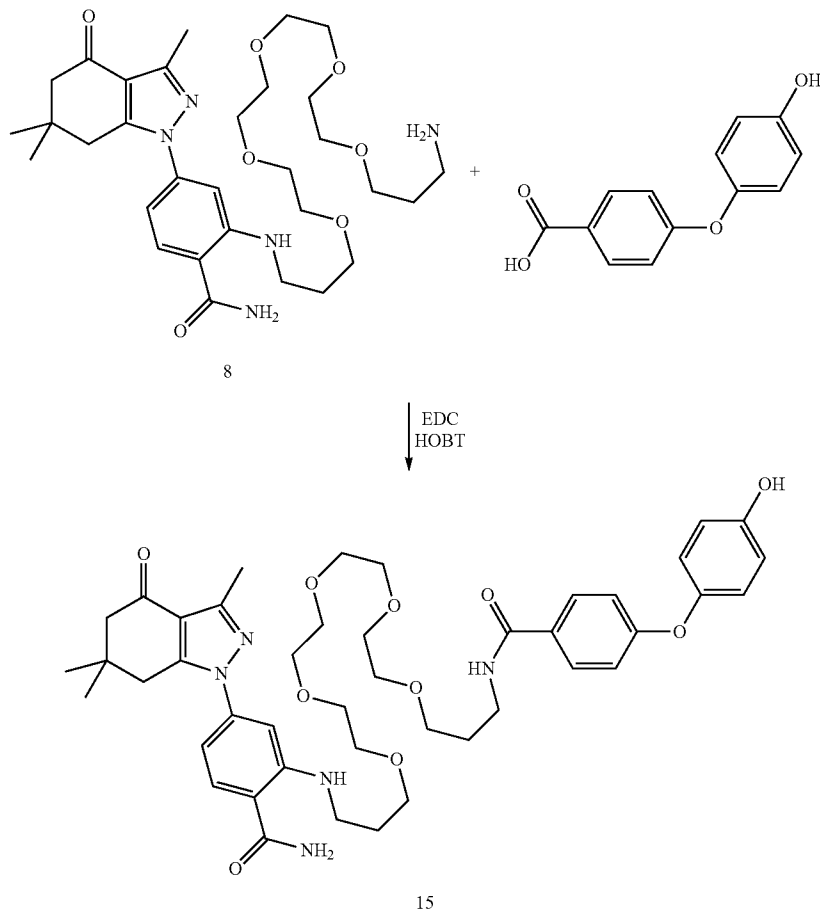
Amine 8 (12.1 mg, 20 μmol) was dissolved in DMSO (200 μL) and added to IR-780 iodide (7 mg, 10 μmol) and treated with Hunig's base (15 μL) followed by ethanol (100 μL). The sample was heated for 3 h at 70° C. The entire sample was added to a silica gel column (2.5x20 cm) and

chromatographed with CH_2Cl_2 (200 mL), $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$ 39/0/9/0.1 (240 mL), 19/0.9/0.1 (240 mL) and 12.3/0.9/0.1 (240 mL). The blue band was concentrated to give product 14 (2.4 mg, 22%) as a blue solid. MS (ESI): m/z 554.1 $[\text{M}+\text{H}]^{2+}$.

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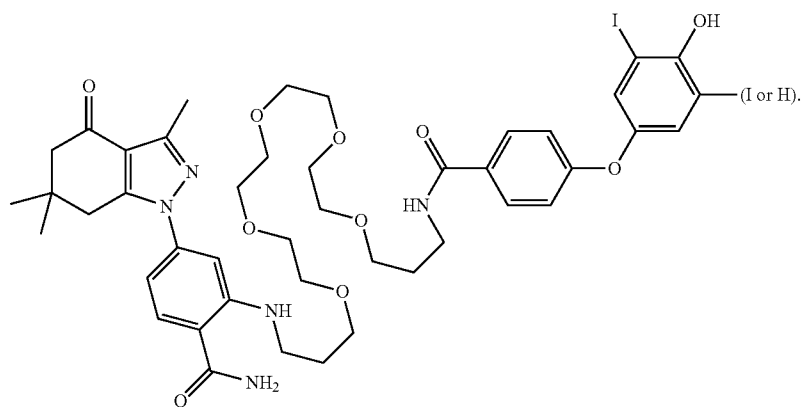
N-((19-((2-Carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-4,7,10,13,16-pentaoxonadecyl)-4-(4-hydroxyphenoxy)benzamide (15)



Amine 8 (90 mg, 149 μmol), 4-(4-hydroxyphenoxy) benzoic acid (38 mg, 164 μmol), EDC (43 mg, 224 μmol), HOBt (21 mg, 156 μmol) and 1 chip of DMAP were dissolved in DMF (1 mL) and stirred at RT. Additional acid and EDC were added until starting amine disappeared (by TLC analysis). The mixture was then treated with 1N HCl (5 mL) and extracted with ethyl acetate (5 \times 5 mL). The organic

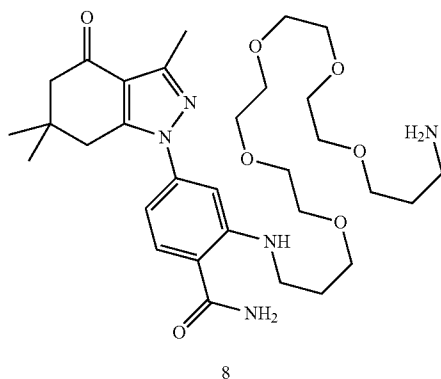
layer was concentrated, dissolved in DMSO and injected on the prep HPLC (50 to 100% methanol, Agilent C-18, 21.1 \times 25 cm). The product peak was concentrated to give 15 (44 mg, 36%) as a clear glass. (ESI): m/z 816.6 $[\text{M}+\text{H}]^+$.

The phenol moiety of compound 15 will be iodinated using standard methods to produce the following compounds:

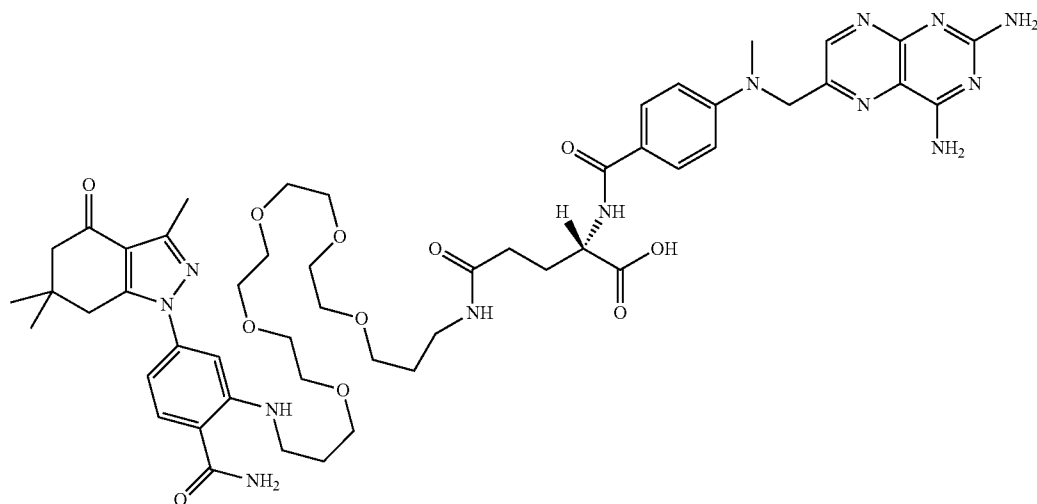
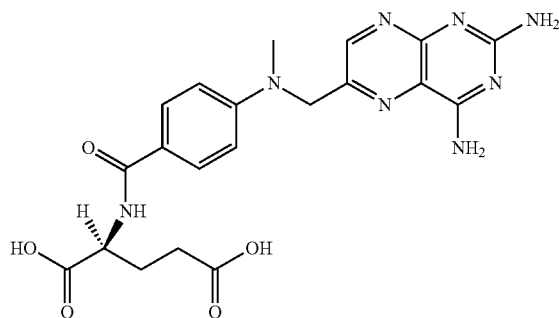


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(S)-1-((2-Carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-24-(4-(((2,4-diaminopteridin-6-yl)methyl)(methyl)amino)benzamido)-21-oxo-4,7,10,13,16-pentaoxa-20-azapentacosan-25-oic acid (16)



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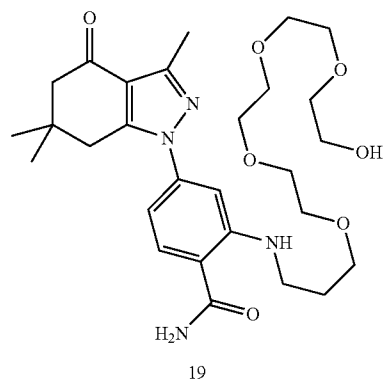
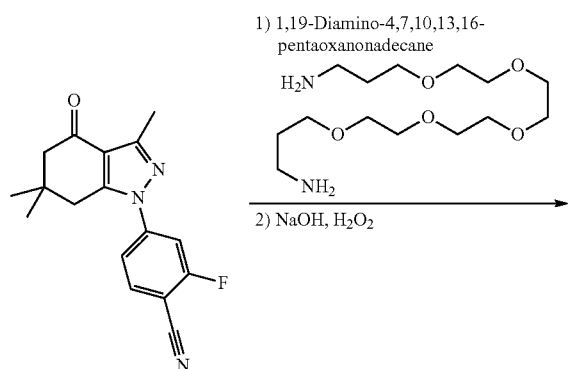
16

Amine 8 (36 mg, 60 μ mol) and methotrexate (MTX, 30 mg, 65 μ mol) were dissolved in DMF (0.5 mL) and then treated with EDC (12 mg, 62 μ mol) in DMF (150 μ L) quickly and stirred at room temperature. After one day, more MTX (6 mg) was added to the mixture followed by more EDC (1.2 mg). After another 2 h the reaction mixture was shot directly onto a prep HPLC (Agilent Prep C-18, 2.5 \times 25

cm, 5 to 100 % MeOH w/2% formic acid) and the product collected and concentrated to give product 16 (40 mg, 64%) as a yellow glass.

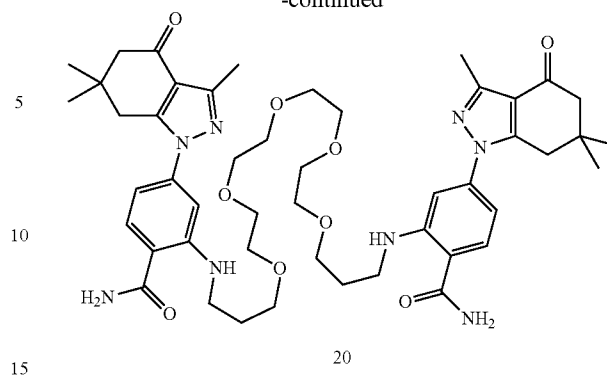
2-((1-hydroxy-3,6,9,12-tetraoxapentadecan-15-yl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (19) and 2,2'-(4,7,10,13,16-pentaoxaanonadecane-1,19-diylbis(azanediyl))bis(4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide) (20)

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-continued



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Using the synthesis of Amine 8, two by-products resulting from bis-coupling and a diamine impurity were recovered. The mixture (~100 mg) was dissolved in DMSO (1 mL) and injected on a prep HPLC (PFH-002_078Prep, 5 to 100% methanol, Agilent C-18, 21.1×25 cm). The early eluting peaks was concentrated to give alcohol 19 (48 mg). (ESI): m/z 547.5 [M+H]⁺. The later eluting peak was concentrated to give the symmetrical compound 20 (34 mg). (ESI): m/z 899.7 [M+H]⁺.

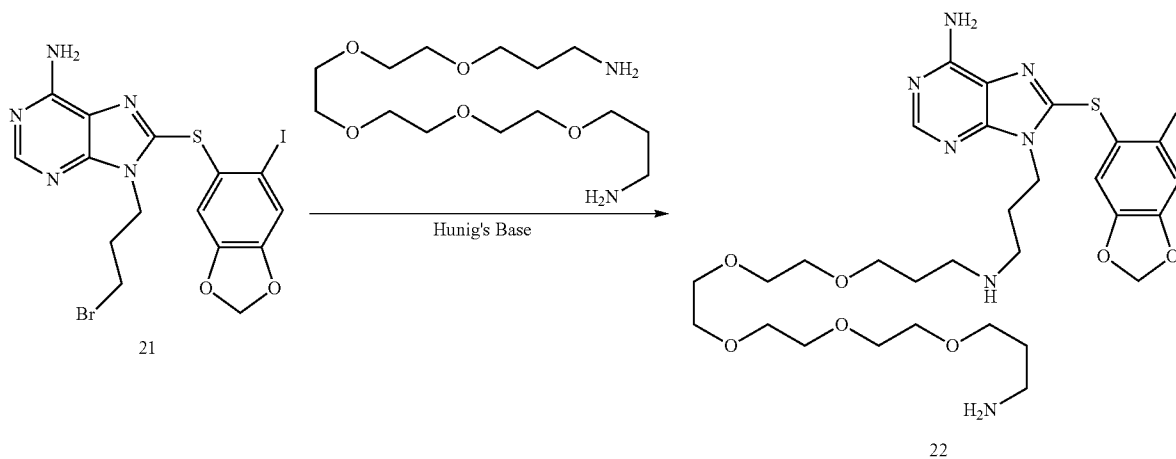
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Example 5

Synthesis of PU-H71 Derivatives

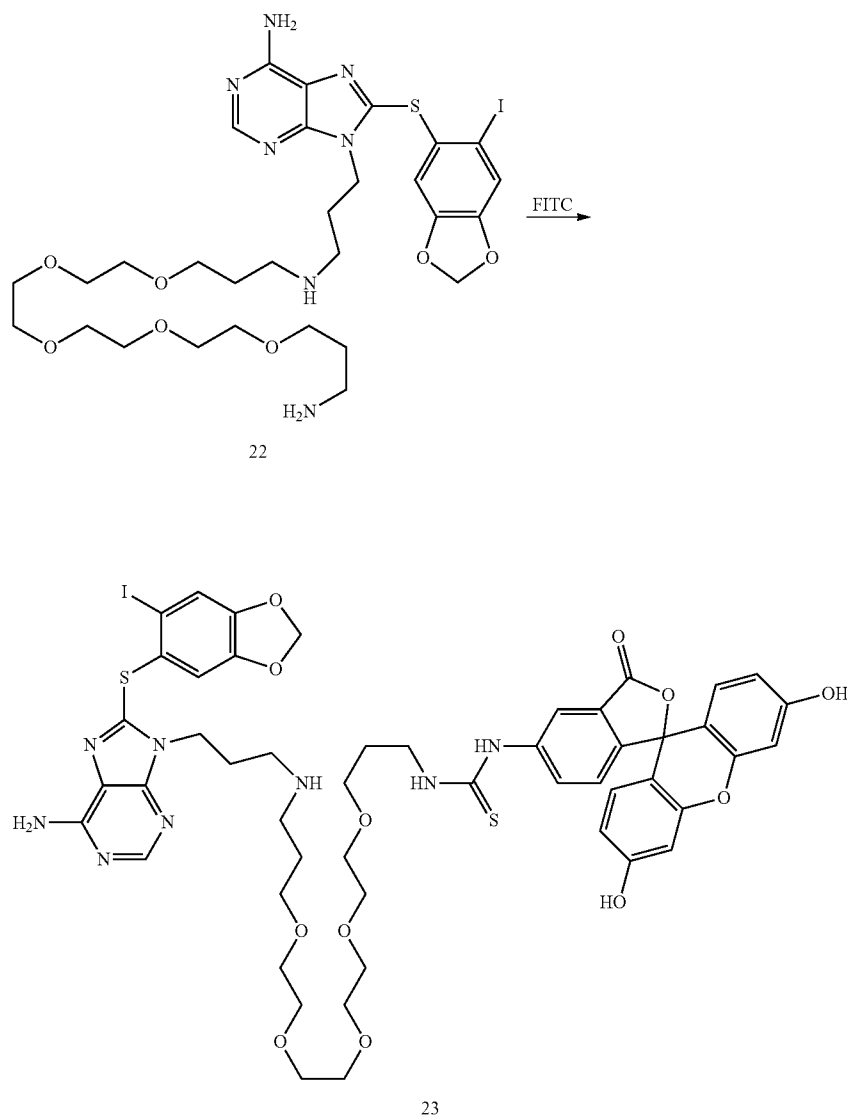
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N1-(3-(6-amino-8-((6-iodobenzo[d][1,3]dioxol-5-yl)thio)-9H-purin-9-yl)propyl)-4,7,10,13,16-pentaoxonadecane-1,19-diamine (22)



9-(3-Bromopropyl)-8-((6-iodobenzo[d][1,3]dioxol-5-yl)thio)-9H-purin-6-amine (21, prepared according to *Bioorg. Med. Chem.* 19, 2603-2614 (2011)) (14 mg, 26 μ mol) was treated with 1,19-diamino-4,7,10,13,16-pentaoxanonadecane (24 mg, 79 μ mol) in ethanol (1 mL) and DMSO (400 μ L). The ethanol was removed under vacuum and the mixture stirred at RT for 1 day. The mixture was purified by preparative HPLC (5 to 100% methanol, 20 mL/m, Agilent C-18, 21.1 \times 25 cm) to give the product 22 (14 mg, 70%) as a clear oil. (ESI): m/z 762.2 [M+H]⁺.

1-(23-(6-amino-8-((6-iodobenzo[d][1,3]dioxol-5-yl)thio)-9H-purin-9-yl)-4,7,10,13,16-pentaoxa-20-azatricosyl)-3-(3',6'-dihydroxy-3-oxo-3H-spiro[isobenzofuran-1,9'-xanthen]-5-yl)thiourea (23)



Amine (22) (all 14 mg, 18 μ mol) was dissolved in ethanol (1 mL) and added to FITC (10 mg, 25.7 μ mol). Additional solid persisted so more ethanol (250 mL) was added, followed by DMSO (250 μ L) and Hunig's base (25 μ L). The

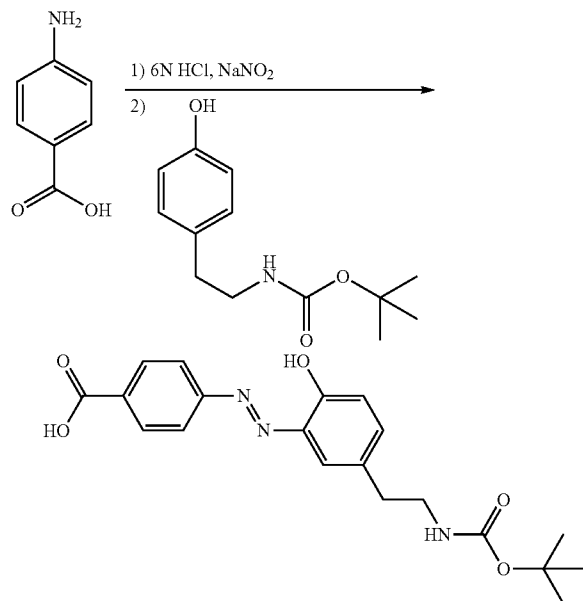
mixture was concentrated and purified by preparative HPLC (PFH-003-027Prep, 5 to 100% methanol, 20 mL/m, Agilent C-18, 21.1 \times 25 cm) to give the fluorescent product (23) (16 mg, 75 %) as a yellow glass. (ESI): m/z 1151.3 [M+H]⁺.

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Example 6

Synthesis of Compound with a Cleavable Linker

(E)-4-((5-(2-((tert-butoxycarbonyl)amino)ethyl)-2-hydroxyphenyl)diazenyl)benzoic acid (17)

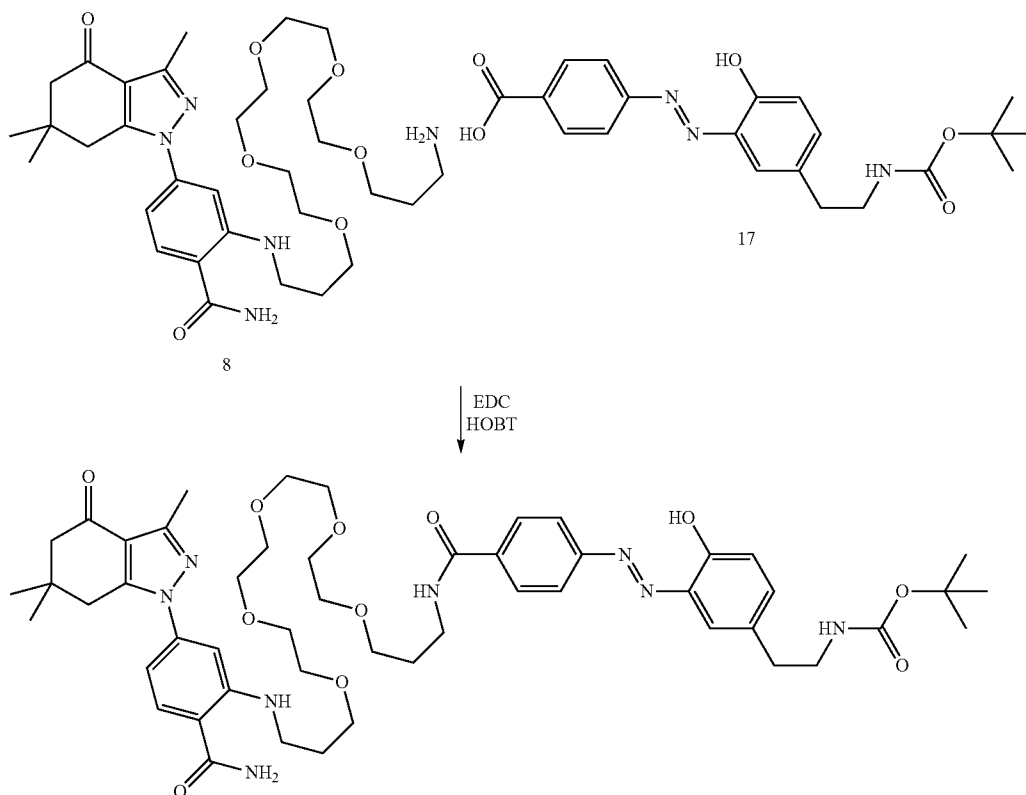


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4-Aminobenzoate (500 mg, 3.6 mmol) was slurried in 6N HCl (10 mL), cooled to 0° C. and treated slowly with sodium nitrite (629 mg, 9.11 mmol). After stirring for 20 m, the mixture was added slowly to an ice-cooled solution of N-Boc tyramine (865 mg, 3.6 mmol) in saturated sodium bicarbonate solution (40 mL) with added sodium bicarbonate (4 g) and a bit of acetone (~5 mL). The orange reaction slurry was left to stir overnight. The reaction mixture was treated 1N HCl (100 mL) until acidic and then stirred an additional 2 h. The solids were filtered off, washed with water and air dried, then dried under vacuum to give 17 (1.03 g, 73%) as a reddish-orange solid. The product was used as is though it contained a minor impurity by NMR. TLC (9/1 CH₂Cl₂/MeOH) R_f=0.20; ¹H NMR (DMSO-d₆) δ 13.19 (br s, 1H), 10.82 (s, 1H), 8.11 (d, J=8.5 Hz, 2H), 8.05 (d, J=8.5 Hz, 2H), 7.55 (s, 1H), 7.28 (d, J=8.3 Hz, 1H), 7.00 (d, J=8.3 Hz, 1H), 6.88 (t, 1H), 3.12 (m, 2H), 2.67 (t, J=6.8 Hz, 2H), 1.33 (s, 9H); MS (ESI): m/z 384.2 [M]⁻, 791.4 [2M+Na]⁻; HRMS (ESI) [M+Na]⁺ calcd for C₂₆H₂₃N₃O₅Na, 408.1536; found 408.1518.

(E)-tert-butyl 3-((4-((19-((2-carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-4,7,10,13,16-pentaoxononadecyl)carbamoyl)phenyl)diazenyl)-4-hydroxyphenethylcarbamate (18)



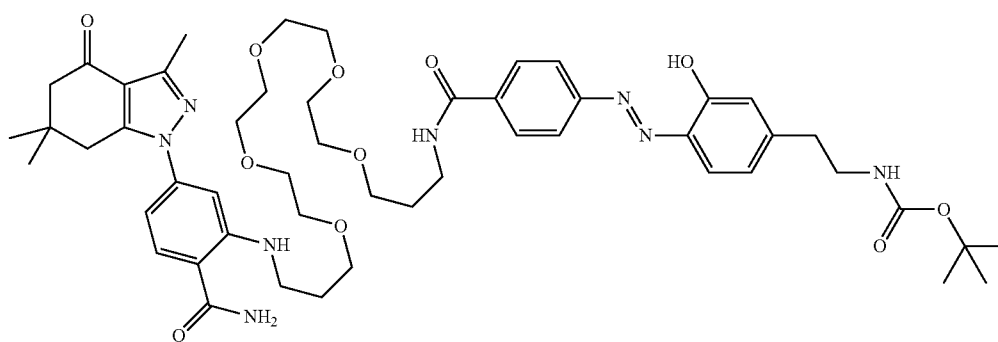
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Amine 8 (483 mg, 800 μmol), acid 17 (308 mg, 800 μmol), EDC (161 mg, 840 μmol) and HOBT (113 mg, 840 μmol) and 2 chips of DMAP and were dissolved in CH_2Cl_2 (10 mL) and stirred at RT for 2 h. The reaction mixture was added to a column and chromatographed (2.5 \times 20 cm, silica gel, CH_2Cl_2 (300 mL), 9/1 $\text{CH}_2\text{Cl}_2/\text{MeOH}$ (300 mL), 4/1 $\text{CH}_2\text{Cl}_2/\text{MeOH}$ (300 mL). The active fractions were combined and concentrated to a frothy glass to give 18 (560 mg, 72%). The hard glass was scraped out to give an orange powder (460 mg). TLC (9/0.9/0.1 $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$) R_f =0.44; ^1H NMR ($\text{DMSO}-d_6$) δ 10.85 (s, 1H), 8.60 (br t, 1H), 8.40 (br t, 1H), 8.02 (br s, 4H), 7.92 (b s, 1H), 7.73 (d, J =8.1 Hz, 1H), 7.55 (s, 1H), 7.27 (d, J =8.1 Hz, 1H), 7.00 (d, J =8.1 Hz, 1H), 6.88 (br t, 1H), 6.76 (s, 1H), 6.66 (d, J =8.1 Hz, 1H), 3.47 (m, 20H), 3.19 (m, 1H), 3.12 (m, 1H), 2.90 (s, 2H), 2.67 (t, 2H), 2.52 (m under DMSO, 2H), 2.38 (s, 3H), 2.31 (s, 2H), 1.77 (m, 4H), 1.33 (s, 9H), 0.99 (s, 6H); HRMS (ESI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{31}\text{H}_{71}\text{N}_8\text{O}_{11}$, 971.5237; found 971.523575.

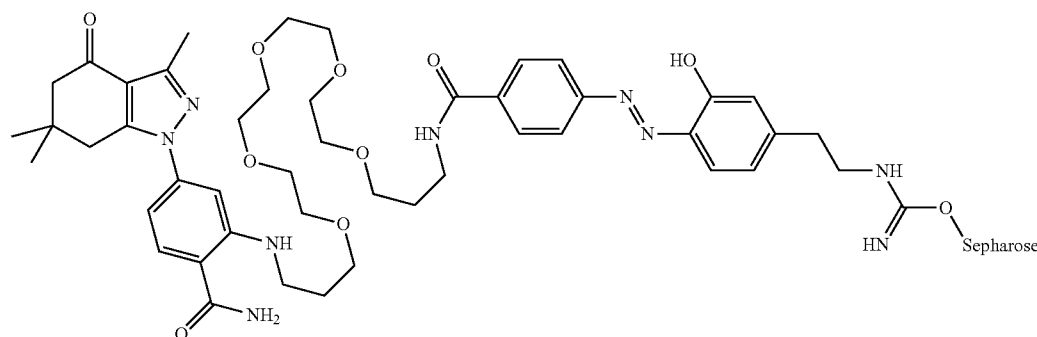
Example 7

Preparation of Affinity Chromatography Resin with a Cleavable Linker (E)



18

1) TFA
2) CNBr-activated Sepharose™ 4B



E

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CNBr-activated Sepharose™ 4B was obtained from GE Healthcare Life Sciences. The following procedure generally followed GE Healthcare Instructions 71-7086-00 AFA.

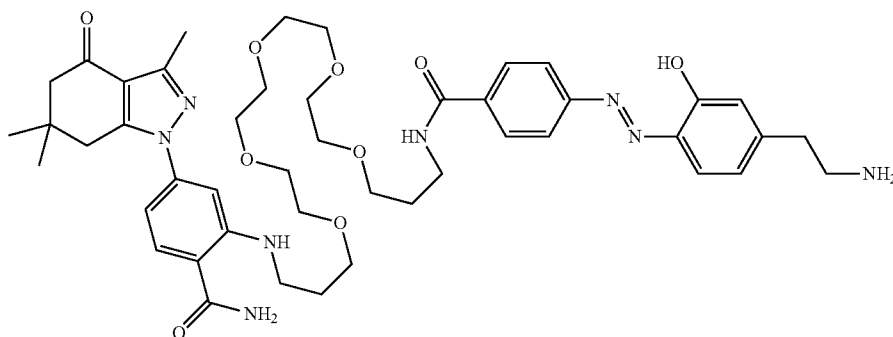
Buffers and Solutions:

Swelling solution	1 mM HCl
Coupling buffer	0.1M NaHCO_3 , 0.5M NaCl, pH = 8.3
Capping solution	1M ethanolamine
Low buffer	0.1M AcOH/NaAcOH, 0.5M NaCl pH = 4
High buffer	0.1M TRIS-HCl, 0.5M NaCl pH = 8
Storage buffer	0.1M KH_2PO_4 , pH = 7.4 w/ 200 mg NaN_3/L

Ligand 18 (25 mg 25.74 μmol) was dissolved in trifluoroacetic acid (1 mL). TLC (9/1/0.1: $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$) showed loss of starting material and formation of a lower product, corresponding to compound 24 (HRMS (ESI) $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{46}\text{H}_{63}\text{N}_8\text{O}_9$, 871.4713; found 871.4696). The mixture was concentrated, then dissolved in ethanol (5 mL) and concentrated again. The residue was then dissolved in ethanol (5 mL) for addition to the resin.

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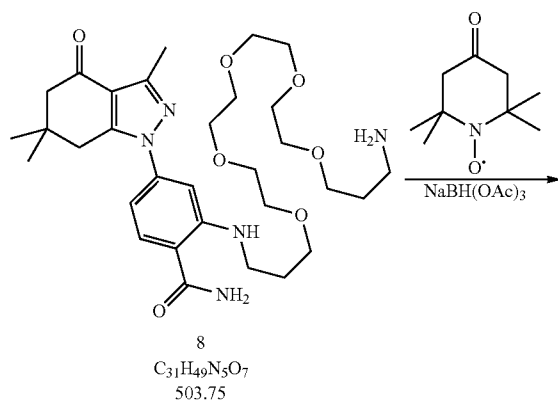
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In a big 275 mL column, CNBr-activated Sepharose™ 4B (25 g) was swelled in 1 mM HCl (450 ml) and then washed with 1 mM HCl (5 L). The resin was washed with coupling buffer (125 mL) and then slurried with coupling buffer (125 mL). The mixture was then treated with the compound described above. The mixture was tumbled at RT for 4 h. The resin was then drained and washed with coupling buffer (5 × 125 mL), diluted with more coupling buffer (~125 mL) and treated with capping solution (2 mL) and rotated at RT for 2 h. The solution was drained and washed with 3 rounds of high buffer/low buffer (250 mL ca.) and finally washed with water (250 mL) and transferred in storage buffer (125 mL) to a bottle and stored at 4° C.

Example 8

Synthesis of Other Compounds for Hsp90 Binding and Cell Penetration Comparisons

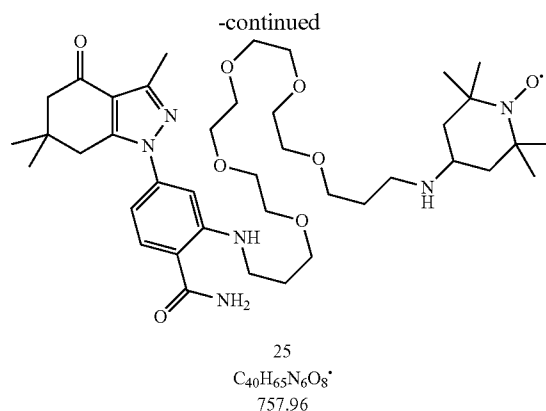
2-((19-((1-oxo-2,2,6,6-tetramethylpiperidin-4-yl)amino)-4,7,10,13,16-pentaoxonadecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (25)



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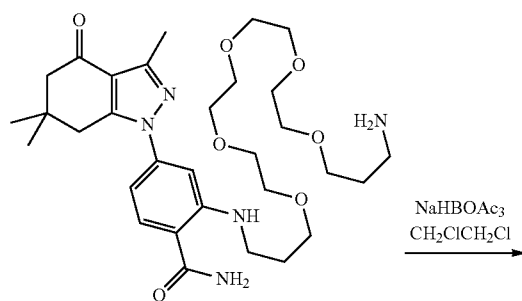
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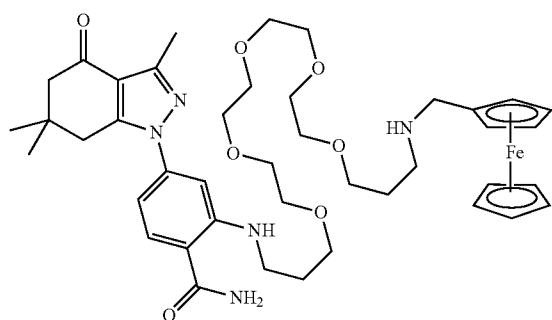
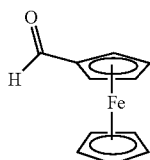
Amine 8 (50 mg, 83 μmol) was dissolved in dichloroethane (2 mL) and treated with 4-oxo-TEMPO (14 mg, 83 μmol) followed by sodium triacetoxyborohydride (23 mg, 107 μmol) and stirred at RT overnight. TLC (CH₂Cl₂/MeOH/AcOH: 4/1/0.1) showed primarily a new product. The mixture was concentrated then dissolved in DMSO/water and chromatographed (5 to 100% MeOH, Agilent C-18 21.2 × 250 mm) to give product 25 (39 mg, 63%) as a glass. TLC (9/0.9/0.1 CH₂Cl₂/MeOH/NH₃) R_f=0.55; ¹H NMR (DMSO-d₆) δ spectra not assigned due to broadening by TEMPO; MS (ESI): m/z 758.5 [M+H]⁺.

2-((19-(ferrocenemethylamino)-4,7,10,13,16-pentaoxonadecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (26), 2-((19-di(ferrocenemethylamino)-4,7,10,13,16-pentaoxonadecyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (27)

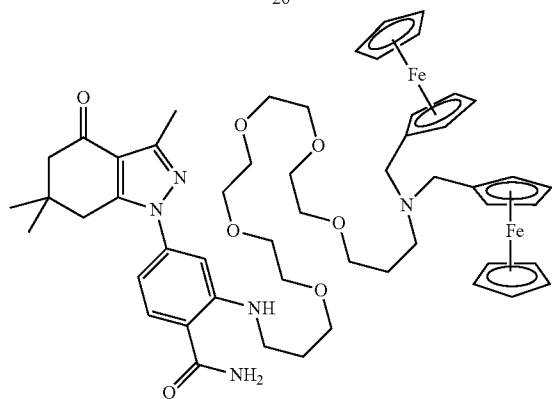
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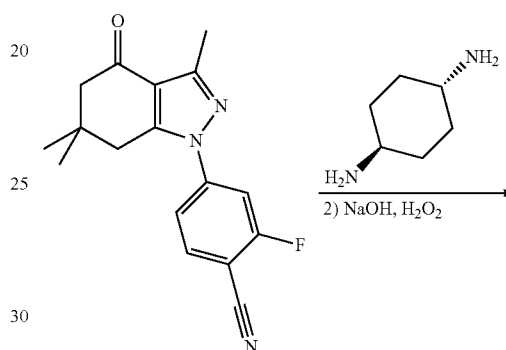
Amine 8 (673 mg, 1.11 mmol) and ferrocene carboxaldehyde (239 mg, 1.11 mmol) were dissolved in dichloroethane (5 mL) and treated with sodium triacetoxy-borohydride (283.5 mg, 1.34 mmol) followed by acetic acid (77 μ L). The reaction mixture was stirred over the weekend. The mixture was adsorbed onto silica (3 g), added to a dry column (1.5x12 cm silica gel) and eluted with 100%, 19/1/0.1, then 9/1/0.1: $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_4\text{OH}$ (300 mL ea.) to give the less polar product, 18 (~180 mg) and the more polar product (~650 mg), both as yellow glasses. The products were separately dissolved in DMSO and chromatographed by HPLC (Agilent Prep C-18, 2.5x25 cm, 5 to 100% MeOH

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w/2% formic acid) to give 26 (490 mg, 54%) and 27 (200 mg, 17%) as viscous yellow glasses. 26. TLC (9/0.9/0.1 $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$) $R_f=0.23$; ^1H NMR (CDCl_3) δ 8.52 (s, 1H), 7.98 (brs, 1H), 7.51 (d, $J=8.1$ Hz, 1H), 6.78 (d, $J=1.6$ Hz, 1H), 6.59 (dd, $J=1.6, 8.1$ Hz, 1H), 6.44 (brs, 1H), 4.3 (brs, 2H), 4.16 (br s, 2H), 4.11 (s, 5H), 3.91 (br s, 2H), 3.57 (br m, 20H), 3.28 (br s, 2H), 2.97 (br t, 2H), 2.79 (s, 2H), 5.52 (s, 3H), 2.37 (s, 2H), 1.91 (br m, 4H), 1.08 (s, 6H); MS (ESI): m/z 802.4 $[\text{M}+\text{H}]^+$. 27 TLC (9/0.9/0.1 $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$) $R_f=0.40$; MS (ESI): m/z 1000.5 $[\text{M}+\text{H}]^+$.

2-(((1*r*,4*r*)-4-aminocyclohexyl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1*H*-indazol-1-yl)benzamide (28)

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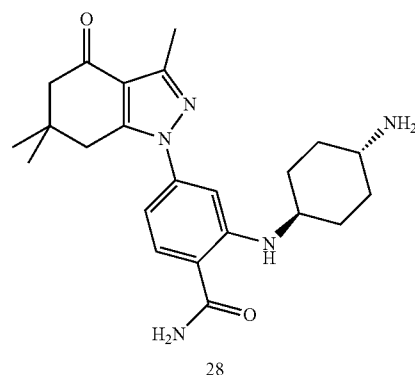
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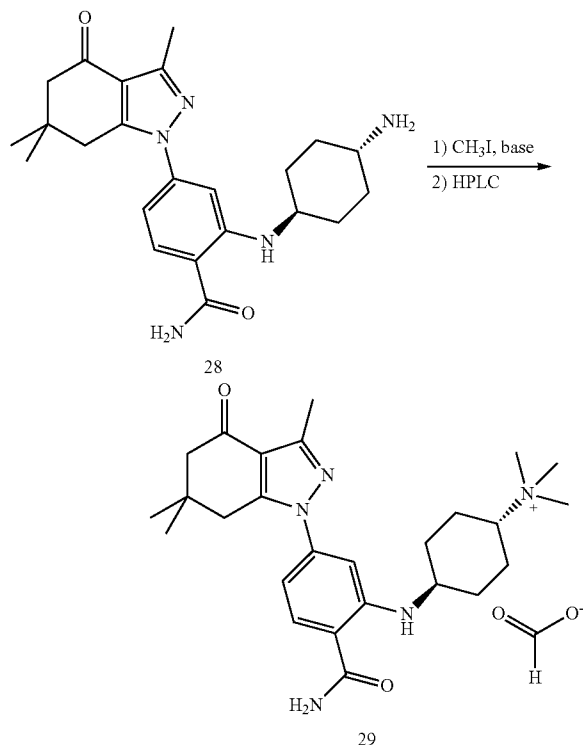
A mixture of 2 (500 mg, 1.68 mmol) and trans-1,4-diaminocyclohexane (1.15 g, 10 mmol) and DMSO (1.5 mL) were stirred at RT for 3 days. The mixture was then diluted with ethanol (3 mL), heated to 90° C. and treated with 50% NaOH (12 drops) and then treated, very slowly, a drop at a time, with 30% hydrogen peroxide. After each drop, the reaction foamed up a bit. After 12 drops the reaction mixture was diluted with water (20 mL) and stirred rapidly overnight. A lot of powder fell out and was filtered off, washed with water and air dried to give 28 (689 mg, 100%) as an off-white powder. MS (ESI): m/z 410.3 $[\text{M}+\text{H}]^+$.

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(1*r*,4*r*)-4-((2-carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1*H*-indazol-1-yl)phenyl)amino)-*N*,*N*,*N*-trimethylcyclohexanaminium (29)



A mixture of 28 (100 mg, 244 μ mol) and methyl iodide (208 mg, 1.5 mmol) were dissolved in methylene chloride (2 mL) containing solid sodium bicarbonate (123 mg, 1.5 mmol) and stirred at RT. With monitoring by LC/MS, additional methyl iodide (total 1.6 mL), sodium bicarbonate (123 mg) and DMF (500 μ L) were added over 4 days until some tetramethylation was detected. The sample was concentrated then diluted with DMSO and water to give a milky solution which was injected on the Prep HPLC (PFH-003_002Prep, 5 to 100% methanol (0.2% formic acid), 20 mL/m, Agilent C-18, 21.1x25 cm). The active fractions were combined and stirred in ethanol to yield a powder which was filtered off to give 29 (28 mg, 23%) as a white powder. MS (ESI): m/z 452.5 $[M+H]^+$.

Example 9

Selectivity Studies

Several media were prepared as described in Example 7, using compounds 6, 7, 8 and 9. These resins were termed A, B, C and D respectively. To prepare the resin in each case, the appropriate compound was added at 1-10 μ mol/gram of resin in minimal methanol.

The ability of each resin to selectively capture Hsp90 from pig mammary gland extract, a tissue shown to be high in ATP binding proteins including native forms of Hsp90, GRP94 and TRAP1, was evaluated. The resin was incubated in the protein solution then washed with a high salt buffer. The bound proteins were removed via an SDS boil procedure, separated by SDS-PAGE electrophoresis, located by

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silver staining and identified using mass spectrometry (MS) sequencing. Selectivity towards Hsp90 was demonstrated by inclusion of 1 mM compound 5 in the tissue extract prior to mixing with the affinity resin. In the case of resin E (Example 7), bound proteins were eluted with 25 mM sodium dithionite in phosphate-buffered saline.

Results are illustrated in FIG. 1. Numbers indicate bands that were sequenced by MS: (1) Fatty acid synthase. (2) Hsp90 (3) Hsp90 (4) Hsp90 proteolytic fragments.

A large number of proteins, including Hsp90, were retained on some resins. The competition experiment, performed by pre-incubating the protein solution with known ligand compound 5, showed clean exclusion of Hsp90 but not other proteins, demonstrating that proteins other than Hsp90 are not client proteins.

Resins A and B showed significant non-specific binding as illustrated in FIG. 1. Resin C showed an intense band of Hsp90, and quenching with compound showed complete blocking of Hsp90 binding as well as several N-terminal fragments. MS analysis demonstrated that all of the recovered proteins were Hsp90 or proteolytic fragments of the protein, suggesting that this resin is selective for Hsp90 over GRP94 and TRAP-1. Resin D was more selective than A and B, although some non-specific binding was observed.

Following exposure of resin E to pig mammary gland extract and treatment with 25 mM sodium dithionite, the eluted proteins were identified by MS. The retained protein-profile (FIG. 1, lane E) was essentially identical to resin C, except that the proteins were recovered using non-denaturing conditions.

Example 10

Additional Selectivity Studies

To further explore the selectivity, the elution of pig mammary gland proteins from gamma phosphate linked ATP Sepharose was studied with compound 8, as well as known potent Hsp90 inhibitors PU-H71 (Caldas-Lopes et al. *Proc. Natl. Acad. Sci. USA* 106 (20), 8368 (2009)) and SNX-2112 (Chandarlapaty et al. *Clin. Cancer Res.* 14 (1) 240 (2008)). This ATP resin has been used previously to study purine-binding proteins (Graves et al. *Mol. Pharmacol.* 2002, 62, 1364) and to discover novel Hsp90 inhibitors (Huang et al. *J. Med. Chem.* 2009, 52, 4288.)

The ATP resin was charged with the pig mammary gland extract and aliquots were distributed into individual wells. Proteins eluted from the wells with increasing amounts of indicated compound were analyzed as described above; results are illustrated in FIG. 2. Compound PU-H71 demonstrates strong potency towards both Hsp90 and GRP94, whereas SNX-2112 shows weaker affinity for GRP94. Compound 8 shows essentially no elution of GRP94.

Example 11

Additional Selectivity Studies

The purinome was captured from pig mammary gland extract as described (Hughes et al., *Bioorg Med Chem.* 2012, 20(10):3298-3305) on gamma phosphate linked ATP Sepharose. Following extensive washing with physiological buffers to remove non-specifically associated proteins, the resin was divided equally and interrogated with the indicated compounds in parallel as described (Hughes et al 2012). All eluted proteins were characterized by SDS-PAGE and silver staining, and then identified by mass spectrometry. Results

are illustrated in FIG. 3. The experiment shows that compounds 5, 17-AAG (Schulte et al. *Cancer Chemoth. Pharm.* 42 (4), 273 (1998)) and PU-H71 have specificity for both Hsp90 and GRP94, whereas compounds 8, 27, 11, 28, 29, 19, 20 only recognize Hsp90. Key: GRP94, glucose regulated protein 94; Hsp90, heat shock protein 90; LAPT, lapatinib; Hsp90 STD, purified Hsp90; 17-AAG, derivatized form of geldanamycin, BLK, vehicle control consisting of buffer containing 10% DMSO used to solubilize all compounds included in the test.

Example 12

Evaluation of Compound 10 in Cancer Cells

In one experiment, the fluorescein compound 10 was applied at micromolar concentrations to four types of tumor cells. As illustrated in FIG. 4, the cell lines MCF-7, BT474 and MDA-MB 468, which are metastatic forms of human breast cancer, take up the probe readily. The Huh7 cell line, a human liver non-metastatic cell line, does not absorb the probe. The probe (10) fluoresces to give green staining in images obtained; in FIG. 4, the green staining appears white. DAPI staining shows the nucleus in blue in images obtained; in FIG. 4, the blue staining appears gray.

In another experiment, MCF7 (metastatic) and MCF10 (benign) cells were treated with an Hsp90 antibody± membrane permeabilization with Triton X 100. As shown in FIG. 5A, in non-permeabilized cells the antibody recognizes surface Hsp90 in MCF7 cells but not in MCF10 cells. After permeabilization the antibody recognizes intracellular Hsp90. Significantly more Hsp90 staining is observed in the MCF7 line. The experiment was also performed using fluorescein compound 10. As shown in FIG. 5B, the probe enters and detects Hsp90 in MCF7 cells, but not in MCF10 cells. The probe also detects more intracellular Hsp90 in MCF7s following permeabilization compared with MCF 10s consistent with increased expression of Hsp90. These results suggest up-regulation of Hsp90 in metastatic cells only. In FIG. 5 the green staining from compound 10 fluorescence appears white, while blue from the DAPI-stained nuclei appears gray.

As shown in FIG. 6A, compound 10 was added to MCF7 cells alone. In FIG. 6B, the compound was added in the presence of an antibody to Hsp90. The antibody prevents internalization of surface 90 causing the probe to accumulate at the membrane. In FIG. 6C, compound 5 competes away the fluorescence signal. DAPI (blue) highlights the nucleus of the cell. In FIG. 6 the green staining from compound 10 fluorescence appears white, while blue from the DAPI-stained nuclei appears gray.

Example 13

Labeling of MDA-MB-468 Tumor Cells in NOD/SCID Mice

MDA-MB-468 tumor-bearing NOD-SCID mice were ip injected with compound 10 (1 mg/mouse, in 100 µl of DMSO). After 1 and 2 hours, mice were sacrificed and tumors and spleens were harvested. Tumors were digested with collagenase/hyaluronidase/Dnase for 1 hour, and ficoll, and washed, then stained with anti-mouse CD45+ anti-H2-kD (APC). Tumor cells were analyzed by flow cytometry, acquired by LSRII for FITC color compound 10 and mouse lineage expression. Lineage negative cells were

analyzed for compound 10 labeling (% positivity is shown in each dot plot). Results are illustrated in FIG. 7A.

Spleen were minced and passed through the strainer. Splenocytes were acquired as described above to analyze compound 10 labeling. Results are illustrated in FIG. 7B.

Example 14

Xenografts

FIG. 8 shows images of xenografts (MDA-MB 468) harvested one hour after injection (IP) with compound 10. Tissue slices were prepared from the harvested xenografts and prepared for analysis by confocal analysis. The presence of compound 10 was detected within the tumor cells confirming uptake of the probe in vivo. A tissue slice from the control experiment is shown in which the animal was injected with vehicle alone (DMSO). Only background fluorescence is detected. In FIG. 8 the green staining from compound 10 fluorescence appears white or light gray, while blue from the DAPI-stained nuclei appears dark gray.

Example 15

Competition Experiments

BT474 cells were incubated with varying doses of compound 5 (0 µM, 0.01 µM, 0.1 µM, 1 µM and 10 µM) in the presence of a fixed concentration of compound 10 (10 µM) and a cell permeabilizing agent (escin) for 30 minutes. The cells were then washed to remove the free probe and cell extracts prepared from the homogenized cells. The amount of drug uptake was then measured by fluorescence (Ex488/Em522 nm) in a micro plate fluorescence reader. FIG. 9 shows that the signal from compound 10 can be effectively competed in vivo with the free parent drug, compound 5, in a dose-dependent manner. This data demonstrates that the fluorescence signal measured in cell and xenograft studies is derived from the probe bound to Hsp90.

Example 16

Measuring the Biodistribution of Compound 10 In Vivo in Mouse Xenografts

MDA-MB-468 cells (5M cells/mouse) were injected to the flank of NOD/SCID mice. When tumors reached 5-10 mm in diameter, tumor-bearing mice were intraperitoneally injected with compound 10 (1 mg/mouse, in 100 µL of DMSO). After 2, 24, 48 or 72 hours, mice were sacrificed and tumors, livers and spleens were harvested. Tumors and livers were digested with collagenase/hyaluronidase/DNase for 1 hour at 37° C., and ficoll to eliminate debris and dead cells, and then washed. Cells in tumor digest were labeled with anti-mouse CD45-biotin+anti-H2-kD-biotin for 15 min on ice, and then 15 min with streptavidin-APC. Tumor cells were analyzed by flow cytometry to detect fluorescence from compound 10. Mouse lineage negative cells were gated for the analysis. Spleen were minced and passed through the strainer. Splenocytes, liver cells and tumor cells were analyzed by flow cytometry to detect fluorescence from compound 10.

Spleen were minced and passed through a strainer to get rid of aggregations/membranous tissue. Cells were stained with violet viability dye. Viable cells were analyzed for their compound 10 uptake. FIG. 10A shows little accumulation of compound 10 in splenocytes.

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Liver tissues were minced with blades and digested with triple enzyme buffer (collagenase III, hyaluronidase, DNase) for 1 h at 37° C., and cells were passed through a strainer to get rid of aggregations. Cells were stained with violet viability dye. Viable cells were analyzed for their compound 10 uptake. FIG. 10B shows little accumulation of compound 10 in liver cells.

Tumor tissues were minced with blades and digested with triple enzyme buffer (collagenase III, hyaluronidase, DNase) for 1 h at 37° C., and cells were passed through a strainer to get rid of aggregations. Cells were stained with anti-mouse CD45-biotin/anti-H2-kd-biotin and then streptavidin-APC and violet viability dye. Viable and CD45/H2-kd negative cells were analyzed for their compound 10 uptake. FIG. 10C shows a high degree of accumulation of compound 10 in tumor cells.

FIG. 11 shows quantitation of compound 10 accumulation in vivo following harvest of the indicated tissues from mice injected with the probe at the indicated time points (5 mice/time point).

Example 17

Tumor Labeling Experiment

MDA-MB-468 tumor cells (5 M cells/mouse) were injected to the flank of NOD/SCID mice. After 3 weeks, tumor volumes reached 400-450 mm³. Mice were treated/untreated with intraperitoneal injection of compound 10 (1 mg/mouse). After 2, 24, 48 and 72 hours, mice were sacrificed, and tumor tissues were frozen in OCT compound. FITC(+) cells in tumor tissues (frozen sections) were analyzed under fluorescent microscope (Axio Observer). From 2 to 48 h, tumor cells showed intense FITC signal, as shown in FIG. 12. In FIG. 12, the green staining from compound 10 fluorescence appears white or light gray.

Example 18

Recovery of Compound 10 Bound to Hsp90 In Vitro and In Vivo

Hsp90 was purified from pig mammary gland as described in Hughes et al., *Bioorg Med Chem.* 2012, 20 (10):3298-3305. The purified protein (0.3 µg) was mixed with an excess of compound 10. The unbound probe was removed by dialysis and the labeled protein subjected to microanion exchange chromatography using a Pharmacia SMART system. Column fractions (100 µL) were collected and analyzed for fluorescence in a plate reader. Chromatographs are shown in FIG. 13A, showing a single peak was recovered by fluorescence that correlated with elution of Hsp90 as determined by Western blot and mass spectrometry. The chromatography experiment was then repeated on cell extracts prepared from MCF7 and BT474 cells treated with compound 10. In both cases a peak of fluorescence was recovered at 49 minutes that co-elutes with Hsp90.

MCF cells were heated to 42° C. or maintained at 37° C. for 60 minutes. Compound 10 was added (10 µM) and after 60 minutes the cells harvested. Extracts were prepared and separated by micro anion-exchange chromatography. Column fractions were analyzed for the presence of compound 10. FIG. 13B shows that the major peak of fluorescence correlates with elution of Hsp90 and that heat treatment induces activation of the protein as reflected by increased uptake of compound 10.

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Collectively these data demonstrate that compound 10 binds to Hsp90 in tumor cells and is not associated with any other cellular protein, and that the probe can be recovered bound to the protein from cell extracts.

Example 19

Imaging of HSP90 Expression in Mouse Xenografts by Fluorescence Imaging

Mice with MDA MB 468 xenografts were injected with either 0.1, 0.5 or 1 mg of compound 10 intravenously. The animals were then immediately imaged in real time in an TVIS Kinetic imager (Caliper Life Science) for the presence of fluorescence at 465ex/520em. After two hours the tumors were excised and examined for the presence of compound 10.

FIG. 14A shows rapid accumulation of compound 10 in the xenograft following tail vein injection. Red image (false, 1 vs. 2) clearly demarks the tumor over background signal (appearing very dark gray in FIG. 14A). Green image (actual, 3 vs. 4) shows clear demarcation of the tumor over and above the natural background fluorescence of the animal (with the tumor appearing bright white and background fluorescence appearing gray).

FIG. 14B shows quantitative analysis of compound 10 accumulation in tumors isolated from mice treated with the indicated doses of the compound. FIG. 14B shows the probe accumulates selectively within the tumor mass in a dose dependent manner. Measurements were made in the IVIS imager or by preparing cells extracts from the tumors and measuring their fluorescence in a microtitre plate reader.

FIG. 14C shows accumulation of compound 12 within tumors isolated from mice injected with the indicated concentrations of the compound. FIG. 14C shows the probe accumulates selectively within the tumor mass in a dose dependent manner. Measurements were made in the IVIS imager or by preparing cells extracts from the tumors and measuring their fluorescence in a microtitre plate reader.

Example 20

In Vivo Detection of Compound 10 with Optical Spectroscopy

MDA-MB-468 tumor cells (1 M cells/mouse) were injected to the flank of SCID mice. Three weeks later, when tumor size reached 5-8 mm in diameter, the experiment was performed. Baseline autofluorescence levels were measured for tumor and normal tissues (skin adjacent to tumor), 1 mg or 0.5 mg of compound 10 was injected via tail vein. After drug injection, fluorescence signals were measured with the optical spectroscopy (~30 min). Two days after injection, measurement of fluorescence signal was repeated.

FIG. 15 shows results from two separate mice injected with compound 10 (1 mg) and the presence of the compound detected tumor by optical spectroscopy at time 0 and then 3 days later. Normal: Normal skin adjacent to tumor. All data are normalized to peak fluorescence intensity to compare fluorescence line shapes. In both mice at Day 0 Normal shows higher signal than tumor after compound 10 injection (due to poor vasculature in tumor). Measurements were made within a few minutes of injection; On Day 3, normal site has declined to pre-injection baseline level whereas the tumor still retains FITC signal in both cases. These data again demonstrate accumulation of compound 10 in tumors only and not other tissues.

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Example 21

Compound 10 Recognizes Cells Infected with HIV-1

TZM-b1 (Hela derived) cells were added on glass cover-slips in 24-well plates (1 × 10⁵ cells/well) and were allowed to attach to cover slips overnight. For infection NL4.3 virus (HIV-1) was added to medium to achieve 10 ng/mL P24 protein concentration along with DEAE dextran at a final concentration 15 µg/mL. After 48 hours of infection, compound 10 was added to the medium and incubated for 1 hour at 37° C. Later the infected cells were washed twice with phosphate-buffered saline (PBS) and fixed for 20 min in 4% paraformaldehyde. They were washed twice with PBS and permeabilized for 30 min in 50% ice cold methanol on ice. Blocking was carried out using 1% FBS for 30 minutes at room temperature after two washes with PBS. Cells were then incubated for 2 hours at room temperature with antibody against HIV-1 capsid protein P24 at 1:100 dilutions in PBS. The antibody was obtained from the AIDS Reference and Reagent Program (HIV-1 p24 Monoclonal Antibody-183-H12-5C, Catalogue number 3537). The cells were washed three times in PBS before and after 1 h of incubation with anti-mouse Texas Red secondary antibody (1:250). (Texas Red® Goat Anti-Mouse IgG (H+L), catalogue number: T682, Life technologies). Cover slips were mounted on the slides using Prolong gold with DAPI (ProLong® Gold Antifade Reagent with DAPI Catalog Number P36935, life technologies) and observed by confocal microscopy (Fluoview FV10i Olympus).

As shown in FIG. 16, panel A is an overlay of B and C; panel B shows the fluorescence from compound 10 (where green staining appears white or light gray); panel C shows fluorescence from the antibody to SP4 viral capsid protein (where red staining appears white or light gray); D shows an optical image of the entire field, showing that the probe and antibody do not recognize uninfected cells. These data demonstrate that compound 10 can be used to detect cells infected with HIV.

Example 22

Recovery of Drug-Bound Protein From Mice

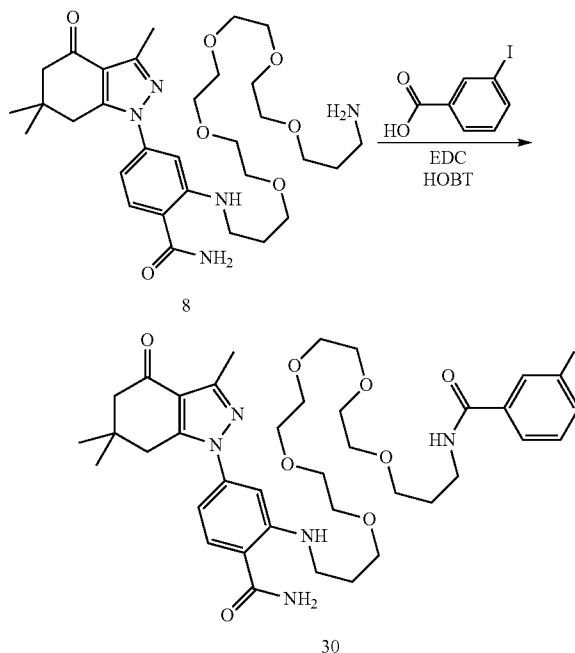
Mice were injected (IV) with 1 mg of compound 10 and the tumors were excised from the animal after 2 hours. Tissue extracts were prepared and characterized by micro anion-exchange chromatography. Column fractions were analyzed for the presence of compound 10 by fluorescence and mass spectrometry. Results are shown in FIG. 17. The major peak of fluorescence (fractions 49-53) co-elutes with elution of full length Hsp90. The two minor peaks (fractions 27-35) were attributable to binding to degraded forms of the N terminal domain of Hsp90.

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Example 23

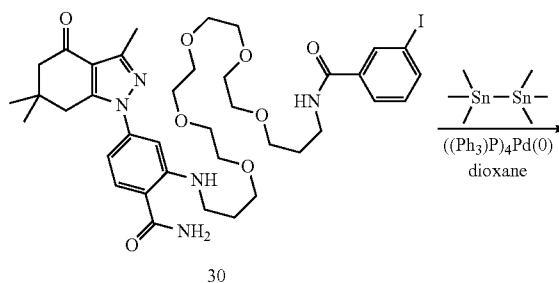
Synthesis of Iodine-Containing Compounds

N-(19-((2-carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-4,7,10,13,16-pentaoxonadecyl)-3-iodobenzamide (30)



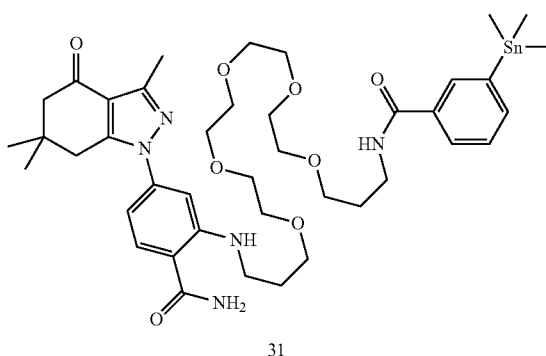
Amine (8) (165 mg, 273 µmol), 3-iodobenzoic acid (68 mg, 273 µmol), EDC (79 mg, 410 µmol) and HOBT (37 mg, 227 µmol) and were dissolved in methylene chloride (4 mL) and stirred at room temperature for 1 day. The reaction mixture was concentrated then dissolved in DMSO (1.5 mL) and purified by prep HPLC (30 to 100% methanol, 20 mL/min, Agilent C-18, 21.1×25 cm) to give 30 (155.7 mg, 68%) as a clear glass. MS (ESI): m/z 834.3 [M+H]⁺.

N-(19-((2-carbamoyl-5-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)phenyl)amino)-4,7,10,13,16-pentaoxonadecyl)-3-(trimethylstannyl)benzamide (31)



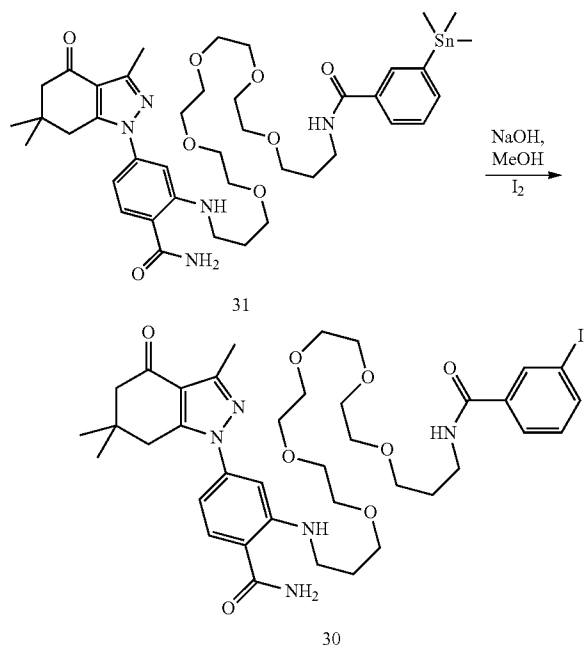
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From *J. Org. Chem.* 2012, 77, 1931-1938 and *Appl. Radiat. Isot.* Vol. 49, No. 8, pp. 955-959, 1998. Iodide (30) (92.7 mg, 111 μmol), hexamethylditin (40 mg, 122 μmol) and tetrakis triphenylphosphine palladium(0) (2.6 mg 2.22 μmol) were slurried in dioxane (2 mL), purged with nitrogen and heated to 100° C. for 1 h then 80° C. for 16 h. The reaction mixture was concentrated and chromatographed (silica gel, 2.5x25, CH_2Cl_2 (100 mL), $\text{CH}_2\text{Cl}_2/\text{MeOH}$: 19/1 (250 mL), $\text{CH}_2\text{Cl}_2/\text{MeOH}$: 9/1 (250 mL)) to give 31 (92 mg, 95%) as a slightly yellowish oil. The product was further purified by prep HPLC (30 to 100% methanol, 20 mL/m, Agilent C-18, 21.1x25 cm) to give a clear glass. MS (ESI): base peak m/z 872.4 $[\text{M}+\text{H}]^+$.

Development of Method for Iodine Incorporation from I_2



Tin compound 31 (15.4 mg, 17.7 μmol) was dissolved in methanol (1 mL) and treated with 1N NaOH (53 μL of 1N solution) followed by Iodine (194 μL of 0.1 M solution). The iodine color went away almost immediately. After stirring for 20 m, an aliquot was removed for LC/MS analysis. LC/MS shows clean conversion to the iodide, about 60% there. Additional iodine (194 μL) was added to test for over

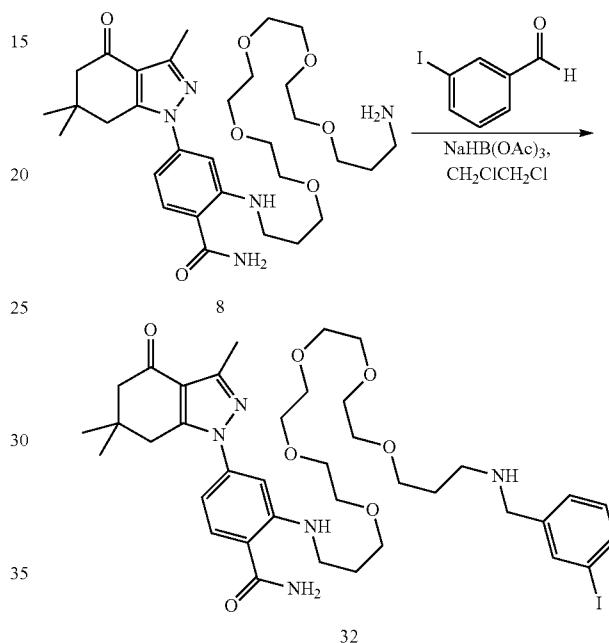
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iodination. After a day, LC/MS showed complete and clean conversion to the mono-iodo compound 30. MS (ESI): m/z 834.3 $[\text{M}+\text{H}]^+$.

Example 24

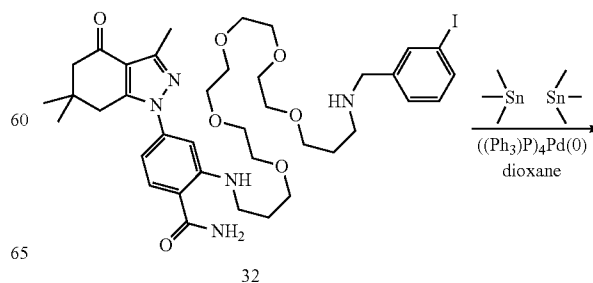
Synthesis of Dual-Function Compounds

2-((1-(3-iodophenyl)-6,9,12,15,18-pentaoxa-2-azahenicosan-21-yl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (32)



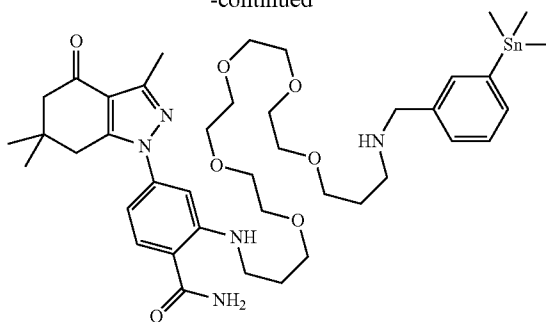
Amine 8 (100 mg, 166 μmol) and 3-iodobenzaldehyde (38 mg, 166 μmol) were dissolved in dichloroethane (2 mL) and treated with sodium triacetoxyborohydride (60 mg, 282 μmol) and stirred at room temperature for 16 h. The sample was then loaded onto a column and chromatographed (silica gel, 2.5x15 cm, CH_2Cl_2 (100 mL), then 19/0.9/0.1: $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$ (200 mL), then 9/0.9/0.1: $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{NH}_3$ (450 mL)) to give 32 (108 mg, 63%) as a clear oil. MS (ESI): m/z 820.3 $[\text{M}+\text{H}]^+$.

4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)-2-((1-(3-(trimethylstannyl)phenyl)-6,9,12,15,18-pentaoxa-2-azahenicosan-21-yl)amino)benzamide (33)



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Iodide 32 (230 mg, 280 umol), hexamethylditin (101 mg, 308 umol) and tetrakis triphenylphosphine palladium(0)

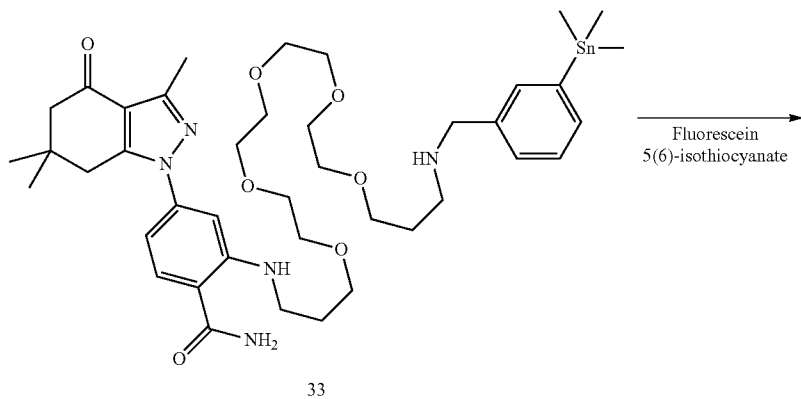
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(6.48 mg 5.61 umol) were slurried in dioxane (5 mL), purged with nitrogen and heated to 100° C. for 45 m. The reaction mixture was concentrated then added to a column and chromatographed (silica gel, 2.5x25, CH₂Cl₂ (100 mL), CH₂Cl₂/MeOH/NH₃: 19/9/1 (250 mL), CH₂Cl₂/MeOH/NH₃: 9/0.9/0.1 (500 mL)) to give 33 (213 mg, 89%) as a dark glass. MS (ESI): base peak m/z 858.4 [M+H]⁺.

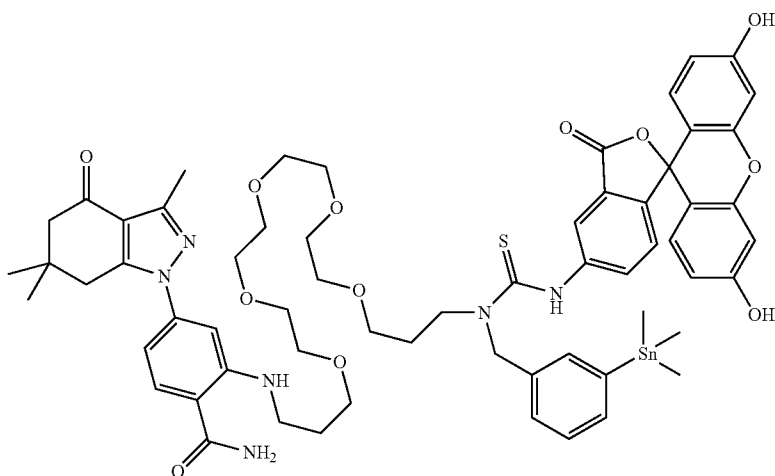
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2-((1-((3',6'-dihydroxy-3-oxo-3H-spiro[isobenzofuran-1,9'-xanthen]-5-yl)amino)-1-thioxo-2-(3-(trimethylstannyl)benzyl)-6,9,12,15,18-pentaoxa-2-azahenicosan-21-yl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (34)



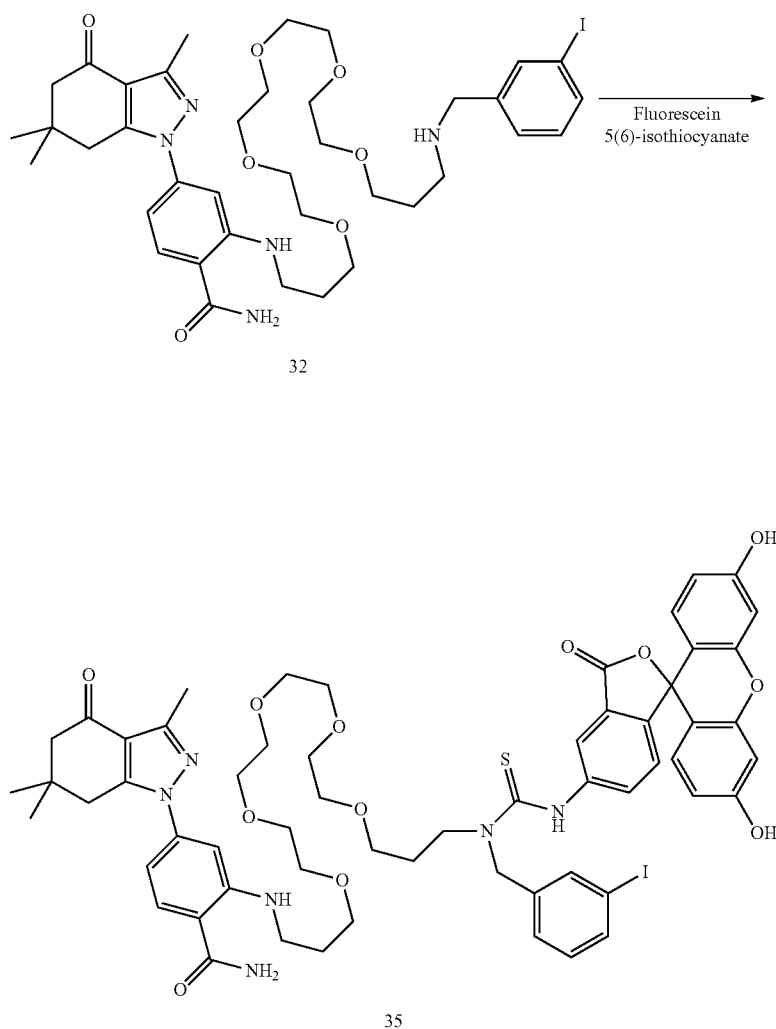
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Tin compound 33 (116 mg, 135 μmol) was dissolved in DMSO (200 μL) and treated with FITC (53 mg, 135 μmol) dissolved in DMSO (200 μL) followed by Hunig's base (around 35 mg, 270 μmol). The mixture was purified by prep HPLC (30 to 100% methanol, 20 mL/min, Agilent C-18, 21.1 \times 25 cm) to give 34 (68 mg, 40%) as a yellow solid. MS (ESI): base peak m/z 1247.4 $[\text{M}]^+$.

Synthesis of Iodide Standard: 2-((1-((3',6'-dihydroxy-3-oxo-3H-spiro[isobenzofuran-1,9'-xanthen]-5-yl)amino)-2-(3-iodobenzyl)-1-thioxo-6,9,12,15,18-pentaoxa-2-azahenicosan-21-yl)amino)-4-(3,6,6-trimethyl-4-oxo-4,5,6,7-tetrahydro-1H-indazol-1-yl)benzamide (35)



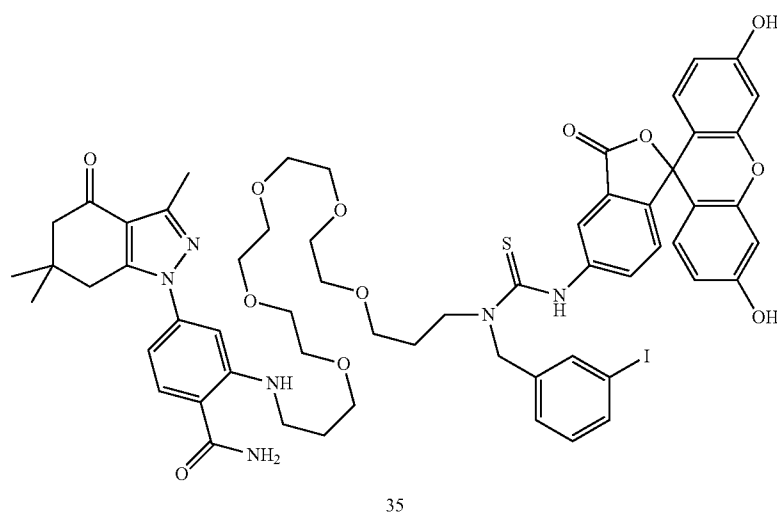
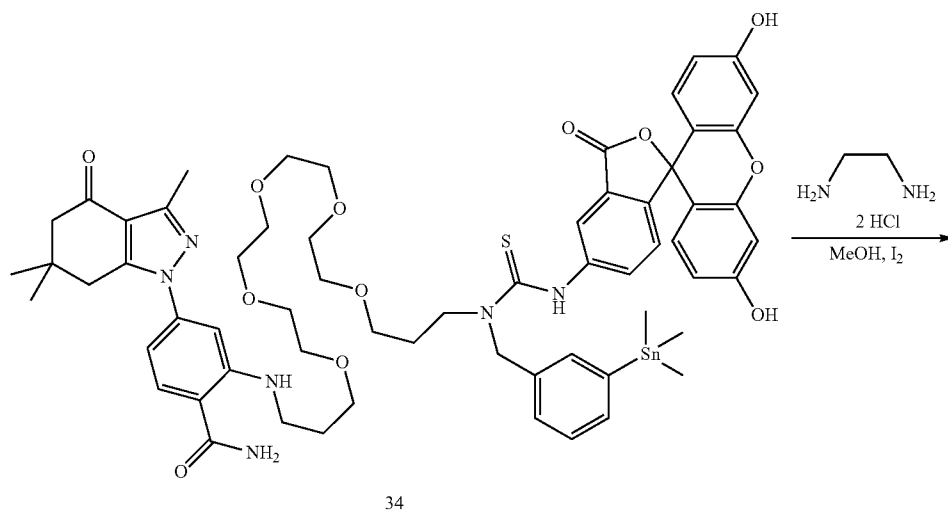
Iodide 32 (31 mg, 38 μmol) was dissolved in DMSO (200 μL) and treated with FITC (15 mg, 38 μmol) dissolved in DMSO (200 μL) and Hunig's base (10 mg, 76 μmol) and stirred for 1 h. The sample was then loaded onto a column

and chromatographed (2.5 \times 15 cm, CH_2Cl_2 (100 mL), then 19/1/0.1: $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{AcOH}$ (1500 mL), then 9/1/0.1: $\text{CH}_2\text{Cl}_2/\text{MeOH}/\text{AcOH}$ (1500 mL)) to give 35 (28 mg, 62%) as a yellow glass. MS (ESI): m/z 1207.4 $[\text{M}]^+$.

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Iodination Test

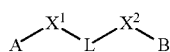


Tin compound 34 (1 mg, 0.8 μmol) was dissolved in methanol (1 mL) and treated with ethylenediamine diHCl (1 mg in 10 μL of water) followed by iodine (8 μL of 0.1 M solution). The color dissipated instantly. After stirring for 1 day, LC/MS showed clean formation of the iodide. MS (ESI): m/z 1207.4 $[\text{M}]^+$.

The iodination can be repeated using a radiolabeled iodine source to provide a dual function compound.

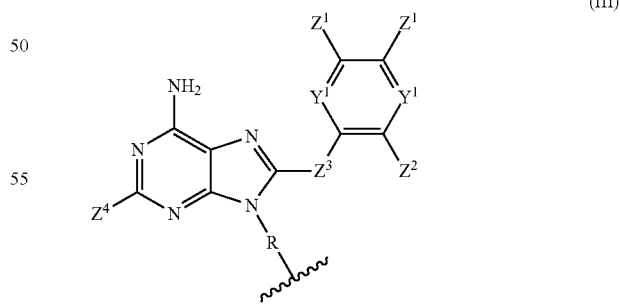
The invention claimed is:

1. A compound of formula (I):



wherein:

A is a heat shock protein 90 binding component of formula (III):



wherein:

R is alkylenyl or heteroalkylenyl;

each Y^1 is independently $-\text{CH}$ or $-\text{N}$;

each Z^1 is taken together with the carbon atoms to which they are attached to form a heterocyclic ring;

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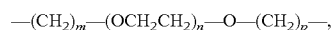
Z^2 is —H or halo;
 Z^3 is —CH₂—, —S—, —O— or —NH—;
 Z^4 is —H or halo; and



is the point of attachment in formula (I);

X^1 is —NH—, —O—, —S—, —C(O)— or —S(O)₂—;

L is a divalent linker of the following formula:



wherein:

m is 2 or 3;

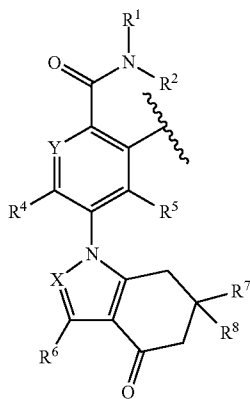
n is 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20; and

p is 2 or 3;

X^2 is —NR—, —O—, —S—, —C(O)— or —S(O)₂—;

R is —H or a detection moiety; and

B is a detection moiety, an anti-cancer agent, or a heat shock protein 90 binding component of formula (II):



wherein:

R^1 is —H or —C₁₋₈-alkyl;

R^2 is —H or —C₁₋₈-alkyl;

Y is —CR³ or —N;

R^3 is —H, —F or —OCH₃;

R^4 is —H, —F or —OCH₃;

R^5 is —H, —F or —OCH₃;

R^6 is —C₁₋₈-alkyl, —C₂₋₈-alkenyl, —C₂₋₈-alkynyl, —C₃₋₈-cycloalkenyl, —C₃₋₈-cycloalkenyl-C₁₋₈-alkyl, —C₃₋₈-cycloalkyl, —C₃₋₈-cycloalkyl-C₁₋₈-alkyl, aryl, aryl-C₁₋₈-alkyl, halo-C₁₋₈-alkyl, heteroaryl, heteroaryl-C₁₋₈-alkyl, heterocyclyl, heterocyclyl-C₁₋₈-alkyl, or hydroxy-C₁₋₈-alkyl;

R^7 is —H or —C₁₋₈-alkyl;

R^8 is —H or —C₁₋₈-alkyl; or

R^7 and R^8 , taken together with the carbon atom to which they are attached, form a —C₃₋₈-cycloalkyl;

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X is —CR⁹ or —N;

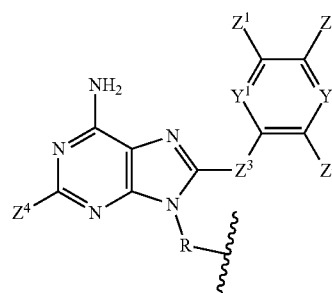
R^9 is —H or —C₁₋₈-alkyl; and



10 is the point of attachment in formula (I); or

B is a detection moiety, an anti-cancer agent, or a heat shock protein 90 binding component of formula (III):

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(III)

wherein:

R is alkylenyl or heteroalkylenyl;

each Y^1 is independently —CH or —N;

each Z^1 is taken together with the carbon atoms to which they are attached to form a heterocyclic ring;

Z^2 is —H or halo;

Z^3 is —CH₂—, —S—, —O— or —NH—;

Z^4 is —H or halo; and



is the point of attachment in formula (I);

wherein each detection moiety independently comprises a fluorophore or a radioactive compound;

wherein the fluorophore is a fluorescein, a rhodamine, a coumarin, a cyanine or a boron-dipyrromethene;

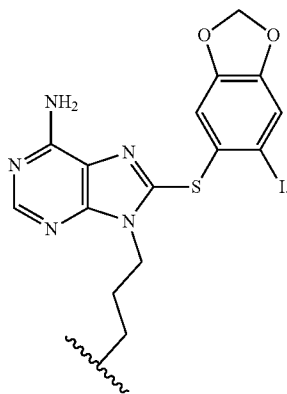
wherein the radioactive compound is a radioisotope; and

wherein the anti-cancer agent is an alkylating agent, an anti-epidermal growth factor receptor antibody, an anti-Her-2 antibody, an antimetabolite, a vinca alkaloid, an anthracycline, a platinum-based agent, a topoisomerase inhibitor, a taxane, an anti-cancer antibiotic, an immune cell antibody, an interferon, an interleukin, a heat shock protein 90 inhibitor, an anti-androgen, an anti-estrogen, an antihypercalcemia agent, an apoptosis inducer, an aurora kinase inhibitor, a Bruton's tyrosine kinase inhibitor, a calcineurin inhibitor, a Ca²⁺-calmodulin-dependent protein kinase II inhibitor, a CD45 tyrosine phosphatase inhibitor, a cell division cycle 25 phosphatase inhibitor, a checkpoint kinase inhibitor, a cyclooxygenase inhibitor, a cRAF kinase inhibitor, a cyclin dependent kinase inhibitor, a cysteine protease inhibitor, a deoxyribonucleic acid intercalator, a deoxyribonucleic acid strand breaker, an E3 ligase inhibitor, an epidermal growth factor pathway inhibitor, a farnesyltransferase inhibitor, a fetal liver kinase-1 inhibitor, a glycogen synthase kinase-3 inhibitor, a histone deacetylase inhibitor, an I-kappa B-alpha kinase inhibi-

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tor, an imidazotetrazinone, an insulin tyrosine kinase inhibitor, a c-Jun N-terminal kinase inhibitor, a mitogen-activated protein kinase inhibitor, a mouse double minute 2 inhibitor, an MEK inhibitor, a matrix metalloproteinase inhibitor, a mammalian target of rapamycin inhibitor, a nerve growth factor receptor tyrosine kinase inhibitor, a p38 mitogen-activated protein kinase inhibitor, a p56 tyrosine kinase inhibitor, a platelet-derived growth factor pathway inhibitor, a phosphatidylinositol 3-kinase inhibitor, a phosphatase inhibitor, a protein phosphatase inhibitor, a protein kinase C inhibitor, a protein kinase C delta kinase inhibitor, a polyamine synthesis inhibitor, a protein tyrosine phosphatase 1B inhibitor, a protein tyrosine kinase inhibitor, an SRC family tyrosine kinase inhibitor, a spleen tyrosine kinase inhibitor, a Janus tyrosine kinase inhibitor, a retinoid, a ribonucleic acid polymerase II elongation inhibitor, a serine/threonine kinase inhibitor, a sterol biosynthesis inhibitor, a vascular endothelial growth factor pathway inhibitor, alitretinon, altretamine, aminopterin, aminolevulinic acid, amsacrine, asparaginase, atrasentan, bexarotene, carboquone, demecolcine, efaproxiral, elsamitrucin, etoglucid, a Gliadel implant, hydroxycarbamide, leucovorin, lonidamine, lucanthone, masoprocol, methyl aminolevulinate, mitoguazone, mitotane, oblimersen, omacetaxine, pegaspargase, porfimer sodium, prednimustine, sitimagene ceradenovec, talaporfin, temoporfin, trabectedin or verteporfin.

2. The compound of claim 1, wherein A has the following formula:

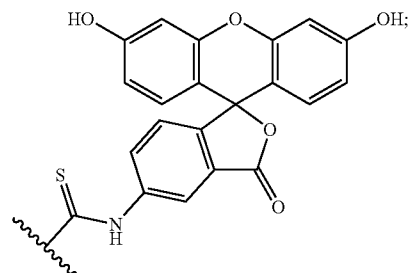


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3. The compound of claim 1, wherein B is the detection moiety.

4. The compound of claim 1, wherein B is the detection moiety comprising the fluorophore.

5. The compound of claim 4, wherein B has the following formula:



and



is the point of attachment to the $-X^2-L-X^1-A$ moiety of the compound of formula (I).

6. The compound of claim 4, wherein the fluorophore is a fluorescein.

7. The compound of claim 1, wherein B is the anti-cancer agent.

8. The compound of claim 7, wherein the anti-cancer agent is methotrexate, topotecan, irinotecan, etoposide, teniposide, lamellarin D, SN-38, camptothecin, belotecan, rubitecan, thalidomide, or verteporfin.

9. The compound of claim 1, wherein B is the heat shock protein 90 binding component of formula (II) or formula (III).

10. The compound of claim 9, wherein B is the heat shock protein 90 binding component of formula (II).

11. The compound of claim 9, wherein B is the heat shock protein 90 binding component of formula (III).

12. The compound of claim 1, wherein m is 3 and p is 3.

13. The compound of claim 1, wherein n is 4.

14. A kit comprising a compound of claim 1.

* * * * *