

TECHNICAL REPORT  
NATICK/TP-16/001



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# **DEACTIVATING CHEMICAL AGENTS USING ENZYME-COATED NANOFIBERS FORMED BY ELECTROSPINNING**

by  
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January 2016

Reprint – Journal Article  
January 2011 – January 2012

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**U.S. Army Natick Soldier Research, Development and Engineering Center  
Natick, Massachusetts 01760-5000**

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<b>1. REPORT DATE (DD-MM-YYYY)</b> 11-01-2016		<b>2. REPORT TYPE</b> Reprint – Journal Article		<b>3. DATES COVERED (From - To)</b> January 2011-January 2012																	
<b>4. TITLE AND SUBTITLE</b>  DEACTIVATING CHEMICAL AGENTS USING ENZYME-COATED NANOFIBERS FORMED BY ELECTROSPINNING				<b>5a. CONTRACT NUMBER</b>																	
				<b>5b. GRANT NUMBER</b>																	
				<b>5c. PROGRAM ELEMENT NUMBER</b>																	
<b>6. AUTHOR(S)</b>  Daewoo Han*, Romy Kirby, Shaun F. Filocamo, and Andrew Steckl*				<b>5d. PROJECT NUMBER</b> 13-115a																	
				<b>5e. TASK NUMBER</b>																	
				<b>5f. WORK UNIT NUMBER</b>																	
<b>7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES)</b>  U.S. Army Natick Soldier Research, Development and Engineering Center ATTN: RDNS-SEW-TMM 10 General Greene Avenue, Natick, MA 01760-5000				<b>8. PERFORMING ORGANIZATION REPORT NUMBER</b>  NATICK/TP-16/001																	
<b>9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES)</b>				<b>10. SPONSOR/MONITOR'S ACRONYM(S)</b>																	
				<b>11. SPONSOR/MONITOR'S REPORT NUMBER(S)</b>																	
<b>12. DISTRIBUTION / AVAILABILITY STATEMENT</b>  Approved for public release; distribution is unlimited.																					
<b>13. SUPPLEMENTARY NOTES</b>  Reprint of article published in the peer-reviewed journal <i>ACS Applied Materials &amp; Interfaces</i> : 2011 December; 3(12):4633-9. doi: 10.1021/am201064b.  * University of Cincinnati, Nanoelectronics Laboratory, Cincinnati, Ohio 45221-0030																					
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<b>15. SUBJECT TERMS</b>  <table style="width: 100%; border: none;"> <tr> <td style="width: 25%;">MATS</td> <td style="width: 25%;">COATINGS</td> <td style="width: 25%;">ELECTROSPINNING</td> <td style="width: 25%;">CHEMICAL WARFARE</td> </tr> <tr> <td>FIBERS</td> <td>POLYMERS</td> <td>NANOSTRUCTURES</td> <td>DECONTAMINATION</td> </tr> <tr> <td>DFPASE</td> <td>NANOFIBERS</td> <td>CHEMICAL AGENTS</td> <td>COAXIAL ELECTROSPINNING</td> </tr> <tr> <td>ENZYMES</td> <td>NEUTRALIZATION</td> <td>SPINNING(INDUSTRIAL PROCESSES)</td> <td></td> </tr> </table>						MATS	COATINGS	ELECTROSPINNING	CHEMICAL WARFARE	FIBERS	POLYMERS	NANOSTRUCTURES	DECONTAMINATION	DFPASE	NANOFIBERS	CHEMICAL AGENTS	COAXIAL ELECTROSPINNING	ENZYMES	NEUTRALIZATION	SPINNING(INDUSTRIAL PROCESSES)	
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<b>16. SECURITY CLASSIFICATION OF:</b>			<b>17. LIMITATION OF ABSTRACT</b>	<b>18. NUMBER OF PAGES</b>	<b>19a. NAME OF RESPONSIBLE PERSON</b>																
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# Deactivating Chemical Agents Using Enzyme-Coated Nanofibers formed by Electrospinning

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## Abstract

The coaxial electrospinning technique was investigated as a novel method to create stabilized, enzyme-containing fibers that have the potential to provide enhanced protection from chemical agents. Electrospinning is a versatile technique for the fabrication of polymer fibers with large length (cm to km): diameter (nm to  $\mu\text{m}$ ) aspect ratios. The large surface to volume ratios, along with the bio-friendly nature of this technique, enables the fabrication of fiber mats with high enzyme concentrations, which amplify the catalytic activity per unit volume of membrane. Blended composite (single source) fibers incorporate enzyme throughout the fiber, which may limit substrate accessibility to the enzyme. In contrast, core/sheath fibers can be produced by coaxial electrospinning with very high enzyme loading (> 80%) in the sheath without noticeable loss of enzymatic activity. Several core-sheath combinations have been explored with the toxin-mitigating enzyme DFPase in order to achieve fibers with optimum properties. The concentration of fluoride released, normalized for the amount of protein incorporated into the sheath, was used

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as a measure of the enzyme activity versus time. The coaxial core/sheath combination of PEO and DFPase produced the highest activity (~7.3mM/mg).

### **Key words**

Coaxial electrospinning, DFPase, Enzyme, chemical warfare, nanofiber, decontamination.

### **Introduction**

Chemical warfare agents are a threat to both military and civilian populations. Many of these agents are part of a class of compounds known as organophosphates (OPs); they disrupt the central nervous system through inactivation of esterases such as acetylcholinesterase, and include such toxins as Sarin, Soman, Tabun and diisopropylfluorophosphate (DFP)<sup>1, 2</sup>. Current protocols for effective neutralization of these compounds involve the use of harsh chemicals (include using strong base, bleach, or strong oxidants), and in some cases can generate waste almost as harmful as the toxin. Naturally occurring substances specifically tailored to react with OPs would be an ideal choice for protection and decontamination.<sup>3,4</sup>

Organophosphatases are naturally occurring enzymes that can neutralize OPs. While these enzymes are able to effectively break down OPs, many of them are expensive and environmentally sensitive. Diisopropylfluorophosphatase (DFPase), a reasonably stable organophosphatase produced by squid, has shown broad based activity against OPs. Clones of

this enzyme have been produced from bacteria, providing a way to mass-produce this enzyme.<sup>5</sup>

Thus, DFPase is a logical choice to test stabilization methods and their effects on enzyme activity.

### *Enzymatic detoxification*

The enzyme hydrolysis methods are highly effective in neutralizing DFP and other OP nerve agents. As shown in Fig. 1, the DFPase enzyme catalyzes the hydrolytic cleavage of the P-F bond in the DFP molecule, releasing diisopropylphosphate (DIP) and fluoride, which do not have the toxic effect of DFP. A calcium ion and a histidine residue (H287) play active roles in the detoxification mechanism; the calcium ion polarizes the fluorophosphate, leaving it susceptible to attack by hydroxyls formed through histidine-mediated proton abstraction from water.<sup>6</sup>

### *Electrospinning*

Organophosphatases have seen limited application due to their sensitivity to environmental conditions, such as pH, temperature and humidity. Encapsulation and covalent attachment techniques have been used to stabilize enzymes with some success.<sup>7-17</sup> However, encapsulation can cause a decrease in activity rate due to rate of diffusion effects, and surface attachment can decrease enzyme activity as well as limit the amount of enzyme loading on the material. Previous studies using DFPase have described decreases in enzyme activity of up to 70%.<sup>18-20</sup> Nanostructures, such as nanoparticles and nanofibers, have attracted much attention due to their extremely high surface to volume ratio (SVR) that can promote the catalytic activity. While the

use of nanoparticles has some difficulty with uniform dispersion and recycling<sup>21</sup>, nanofibers provide several very attractive aspects: (a) easy handling & recycling; (b) high SVR; (c) large variety of materials; (d) high porosity for easy access on fiber surfaces; (e) multi-layer composition.

Since electrospinning is a very attractive method for nanofiber production, electrospun nanofibers have been used as a supporting material for enzymes in different ways. The most common approach for enzyme loading on electrospun fibers is the dip-coating method. After a fibrous mat is formed by electrospinning, it is immersed into the enzyme solution for a certain period of time for enzyme adsorption. Cross-linkers<sup>22, 23</sup> can be utilized to add sequential layers of enzyme on the fiber mat. Modifications to the fiber surface<sup>7, 17, 24</sup> can be performed to improve the surface chemistry for enzyme attachment and subsequent increased enzyme activity. Another well-known method is to electrospin from a solution containing a blend of enzyme and electrospinnable polymer.<sup>13, 25</sup> While this method provides a one-step simple route for enzyme loading on fibers, the fraction of the polymer in the solution is quite large in order for the blend to be electrospinnable. Therefore, a relatively small amount of enzyme is present on the fiber surface, leading to lower catalytic activity.

### *Coaxial electrospinning*

In this report, coaxial electrospinning that produces fibers with a core-sheath structure in one



step is compared to single nozzle electrospinning of a polymer and enzyme blend (co-electrospinning). Using coaxial electrospinning, enzymes encapsulated by a biocompatible polymer sheath layer have been reported.<sup>26</sup> This approach can be used for controllable drug release systems without an initial burst release. Using electrospinning to form fiber mats with very high SVR allows the incorporated enzyme to interact most efficiently with the chemical agents. The versatility of the coaxial core/sheath electrospinning fiber formation is clearly a key asset in producing the optimum material for protection from chemical toxin exposure. To the best of the authors' knowledge, this article provides the first report of enzyme coated polymer fibers using coaxial electrospinning.

## **Experimental conditions**

### *Materials*

Crude recombinant diisopropylfluorophosphatase (DFPase) at 40% purity was purchased from Codexis and used without further purification. For the purpose of this paper, this crude material is collectively referred to as DFPase. Poly( $\epsilon$ -caprolactone) (PCL) (M.W. = 80 kDa), piperazine-N,N'-bis(2-ethanesulfonic acid) (PIPES) and diisopropylfluorophosphate (DFP) were purchased from Sigma-Aldrich (St. Louis, MO), and used as received. Fluoride concentration measurements were obtained with an Orion 9609 ionplus fluoride electrode

purchased from Thermo Fisher Scientific Inc. (Waltham, MA). Poly(ethylene oxide) (PEO) (M.W. = 900 kDa) and 2,2,2,-trifluoroethanol (TFE, 99.8% purity) solvent was purchased from Acros Organics (Geel, Belgium). These materials were used as received without any further modification.

### *Fabrication*

The coaxial electrospinning method is illustrated in [Fig. 2](#). Solutions for the core and sheath materials are separately fed into the coaxial nozzle from which they are ejected simultaneously. Upon application of a sufficiently high voltage, a compound Taylor cone is formed and a liquid jet is ejected consisting of the core material enveloped by the sheath material. The ejected compound liquid jet experiences whipping and bending instabilities within a sufficient distance for evaporating its solvent thoroughly and, consequently, becomes a solid micro/nanofiber with a core-sheath structure. Different characteristics from each material can thus be combined into a single fiber. Using the coaxial electrospinning method we have previously explored the formation of specialized fibers for several applications, including tissue engineering<sup>27</sup>, superhydrophobic fabrics<sup>28</sup>, photocatalytic materials<sup>29</sup>, etc.

To demonstrate the versatility of coaxially electrospun DFPase fibers, DFPase containing fiber mats were also formed by alternative methods (blend solution electrospinning and dip-coating).

### *Enzyme blended electrospinning*

Most enzymes are not directly electrospinnable because of low viscosity (even at very high concentrations). To facilitate the electrospinning of enzymes, the most convenient method is to form solutions with electrospinnable polymers. When the solution contains a large portion of electrospinnable polymer, the resulting fibers have a relatively small portion of enzyme exposed at the fiber surface since a significant amount of the enzyme is embedded within the fiber. To optimize the amount of viable enzyme, and therefore increase catalytic efficiency of the blend fiber approach, one needs to determine the minimum polymer concentration required for uniform fiber production.

Blended solutions for the electrospinning were prepared by dissolving DFPase (3 wt.%) and PEO (3 wt.%) into DI water. Sodium dodecyl sulfate (SDS) surfactant was added at 0.05 wt.% not only to reduce the surface tension of the aqueous blended solution but also to provide a weakly basic environment for the enzyme solution. A high voltage of 20 kV was applied over a 20 cm gap distance and a solution flow rate of 0.4 mL/hr was used. The solution contained equal amounts by weight of DFPase enzyme and PEO polymer. A total solution amount of 670  $\mu$ L was dispensed.

### *Enzyme coaxial electrospinning*

Two polymers were utilized in coaxial electrospinning experiments: water-soluble PEO, and

PCL that is not water-soluble and requires select organic solvents. Coaxial electrospinning conditions are summarized in [Table 1](#). DI water was used as the solvent for PEO and DFPase and chloroform was used for the PCL solution.

The total dispensed amount of polymer in the various samples was maintained roughly constant. [Table 2](#) summarizes the compositions and weights of the 9 types of fiber mats fabricated: the weight of fiber mats, the weight ratio of polymer to DFPase and the amounts DFPase loaded for each sample type.

#### *Dip-coating method*

PCL-only fiber mats were dip-coated into a 3.3 wt.% DFPase aqueous solution. After 5 min immersion, the dip-coated fiber mats were lightly blotted and dried at room temperature overnight.

#### *Enzyme activity measurement*

The ability of electrospun DFPase to defluorinate DFP was measured based upon a previously described method with a fluoride probe.<sup>20, 30</sup> Briefly, samples were added to a glass Petri dish with 7.00 mL of 10-25 mM PIPES buffer, pH 7.2 and a stir bar. The samples were allowed to equilibrate for 3 minutes under the reference and fluoride electrodes. DFP solutions were prepared in Milli-Q deionized water at a concentration of 0.010 M. Once equilibrated, a

3.00 mL aliquot of the DFP solution was added, and fluoride ion measurements were recorded every 5 seconds for three minutes. Controls were also established for comparison: (1) a 3.00 mL aliquot of DFP solution was added to 7.00 mL of PIPES buffer and monitored for 3 minutes, to determine the rate of self-degradation; (2) DFPase was added to 7.00 mL of PIPES buffer, allowed to equilibrate; (3) then 3.00 mL of DFP added and monitored as described.

## Results and Discussion

The core-sheath structure of the coaxially electrospun fibers was demonstrated by TEM and fluorescence microscopy. TEM observation in [Fig. 3a](#) shows the core-sheath structure. The core and fiber diameters are 114 nm and 128 nm, respectively, and the wall thickness is  $\sim 7$  nm. This is well matched with our quantitative analysis considering material densities and flow rate ratio, which results in a calculated ratio of core radius to fiber radius of  $\sim 1 : 1.1$ . For fluorescence microscopy, we have added a Keystone Red XB dye to the sheath DFPase solution. Keystone Red dye is soluble in the aqueous sheath solution, but not in the PCL core solution. Fibers formed in this manner exhibited strong red fluorescence, as shown in [Fig. 3b](#). To confirm the location of the dye in the sheath, the fibers were immersed in water for 3 hr followed by rinsing with running water. Red fluorescence was not observed in the rinsed fiber mat ([Fig. 3c](#)). At the same time no noticeable change was observed in the overall morphology of the fiber mat as can

be seen in the bright-field photographs taken before and after rinsing (Figs. 3d and 3e). Therefore, it appears that the coaxially electrospun fibers have a core-sheath structure with a DFPase/dye sheath layer and a PCL polymer core.

Fiber morphologies have been observed in more detail using an EVEX mini-SEM, SX-3000. As shown in Fig. 4a-c, the coaxial and blended fibers provide good porosity, as there are no filled pores or inter-diffused fibers. In contrast, the porosity of the dip-coated PCL fiber mat shown in Fig. 4d was significantly decreased by loading of the DFPase. Additionally, blended composite fibers (Fig. 4a) and coaxially electrospun fibers (Fig. 4b-c) show uniform fiber morphology without beads.

For the dip-coated fiber mat, as shown in Fig. 4d, the total surface area was decreased by the reduced porosity and DFPase was not loaded uniformly over the entire PCL fiber mat, especially when the PCL fiber mat was thick. On the other hand, for fibers formed by coaxial or blend electrospinning, the amount of DFPase enzyme can be manipulated by adjusting concentration and/or flow rates without sacrificing fiber mat porosity and uniformity. Therefore, the dip-coating technique was not pursued in further studies.

Chemical composition of coaxial fiber mats has been investigated using energy dispersive X-ray analysis as shown in Fig. 5. In contrast to PEO fibers, PEO-DFPase coaxial fibers (Fig. 5a), show additional peaks of sodium, silicon, sulfur, chlorine, and calcium. DFPase includes

numerous sulfur-containing amino acids in its structure. Since sulfur is not present in any of the polymers used, we chose it for elemental mapping in order to show the dispersion of DFPase throughout the fiber mats. . As shown in [Fig. 5b-c](#), the coaxial fiber mat shows very strong sulfur content, while the non-fiber area exhibits only noise level detection. It is clear that the DFPase exists uniformly over the entire fiber mat without any localized large clusters of DFPase.

Catalytic activity of DFPase enzymes has been characterized by measuring the fluoride concentration ( $[F]$ ), which is a product of the hydrolysis reaction with DFP. Initial experiments were carried out to confirm the activity of the enzyme incorporated into electrospun fibers. For this purpose a relatively large amount of DFPase was introduced into the electrospun fiber mat. [Fig. 6a](#) shows  $[F]$  as a function of time for the 6 types of fiber mats described in [Table 1](#), as well as two reference samples: a DFPase-only (no fibers) sample and a fiber-only (PEO) sample.. The DFPase-only sample produced a similar amount of  $[F]$  to that generated by the DFPase-containing fibers. As expected, the PEO-only sample showed only a trace  $[F]$ . More insight into the differences between fiber types is obtained by studying the  $[F]$  normalized to the DFPase amount introduced into each sample type. As shown in [Fig. 6b](#), the coaxial fiber sample with PCL core and DFPase/PEO sheath has the fastest reaction rate and highest level of  $[F]$  released per weight of initial DFPase. Among the fiber mats, the sample with uniform fibers of a blend of DFPase and PEO (1:1 wt. ratio) shows the lowest level of normalized  $[F]$  release. Interestingly,

the DFPase-only sample exhibited an even lower sensitivity.

All samples show a similar [F] vs. time, with a relatively linear initial region followed by saturation to roughly the same [F] value after ~ 2 min. Given that the samples had varying DFPase amounts (from ~2.5 to 18 mg) it is clear that the saturation effect is caused by other factors. The most likely explanation is that at these levels of DFPase and DFP, an excess amount of acid is quickly generated. This overwhelms the buffering capacity of the solution, causing a drop in pH. In turn, this deactivates the enzyme by protonation of the active site during hydrolysis. A reduction in pH is known to reduce the DFPase-DFP reaction rate.<sup>6</sup> Fig. 7 shows both the [F] generated by DFP-DFPase reaction and the pH of the solution as a function of time. At first the [F] increases linearly with time while the pH is reduced. However, when the pH has dropped to the 5.5-6.0 range, a saturation in the [F] level begins to occur. This effect is more dramatic at a lower buffer concentration (Fig. 7a), with the pH dropping abruptly and enzymatic reaction saturating at a lower [F] level. As shown in Fig. 7b, increasing the buffer concentration leads to a slower pH drop and a higher reaction rate level.

Quantitative comparisons of enzyme efficiency were obtained from later experiments (see Fig. 7) because of pH decrease during these preliminary measurements. The decrease in pH is known to reduce enzyme activity<sup>6</sup>. Therefore, experiments on DFPase-DFP reaction kinetics were carried out to determine optimum reaction conditions and minimize pH effects. Fig. 8a



shows [F] vs. time for varying DFPase concentrations (0.1-1.0 mg/mL) at a fixed DFP concentration of 1mM. For the two higher DFPase concentrations the [F] signal was essentially the same, indicating that the DFP amount present was limiting the reaction. Reducing the DFPase concentration to 0.1 mg/mL resulted in a lower [F] and had a much reduced pH effect. Therefore, the next step was to investigate higher DFP concentrations while holding the DFPase concentration at 0.1 mg/mL. As shown in [Fig. 8b](#), improved linear [F] dependence was obtained for 10 and 100 mM DFP concentrations. Due to the toxicity of DFP, subsequent experiments were carried out with 10 mM concentration to minimize exposure. In addition, to minimize pH effects the buffer concentration was increased to 25 mM.

These conditions were utilized to produce and test optimized enzymatically active electrospun fiber mats. [Fig. 9a](#) shows normalized [F] with respect to time for these optimized coaxial fiber mats. These data were used to determine enzyme activity as a function of these constructs ([Table 3](#)). The coaxial core/sheath combination of PEO core and DFPase sheath produced the highest enzyme activity (~7.28 mM/mg), with slightly lower activity levels for PCL core - DFPase/PEO sheath and and PEO core - DFPase/PEO sheath constructs. It is important to note that the activity values for these optimized fibers have increased by a factor 2-3 from previous fibers (in [Fig. 6b](#)). To confirm that the enzyme was not affected by the electric field it experienced during electrospinning, a cast film sample was formed by coaxial dispensing with no

applied voltage. As can be seen in [Fig. 9a](#), this sample (dashed line) has a similar [F] as its fiber counterpart, indicating that there is no reduction in the enzymatic activity due to the electrospinning process, in contrast to other incorporation procedures, where activity can decrease by up to 70% in comparison to free enzyme in solution. Since in our case no reduction in enzymatic activity was observed, a more rigorous kinetic analysis (such as Michaelis Menten kinetics) was not pursued.

PEO fibers are water-soluble; therefore, they can only undergo a single exposure to the DFP solution. Since PCL is not water soluble, the PCL core/PEO sheath construction was chosen for further experimentation. Fibers were created with DFPase in the sheath, and then exposed to an aqueous DFP solution. The PEO sheath should dissolve after the first exposure, resulting in potential interactions between DFPase and the PCL core. [Fig. 9b](#) shows [F] vs. time for a single coaxial PCL core – DFPase/PEO sheath fiber mat sequentially exposed to fresh DFP solution for a total of three times. It appears from the resulting data that the sheath material is completely dissolved during the first exposure and subsequent DFP exposures only exhibited a background level of [F]. This is a similar result to that observed with PCL core – DFPase/dye experiments shown in [Fig. 3](#). This indicates that a different fiber construction is necessary to achieve a material that can undergo multiple exposures without degradation.

A different approach was pursued in the design of an enzyme containing fiber mat that is

also water resistant. A multi-layer scheme, shown in Fig. 10a, places the region containing the enzyme-coated PEO fibers between two layers of superhydrophobic Teflon-coated PCL fibers. A comparison between a conventional enzyme fiber mat and the multi-layer fiber mat is shown in Fig. 10b. The two mats, which also contained a dye in the enzyme solution for visualization purposes, were immersed in water for 12 hours. The conventional mat (Fig. 10b, left) released the dye immediately upon being immersed, whereas the multi-layer mat (Fig. 10b, right) completely contained the dye within the mat even after 12 hours. The resulting solutions were tested for the presence of DFPase, which has an absorption peak at ~ 267 nm. As shown in Fig. 10c, the release of DFPase is greatly suppressed by the multi-layer structure. This is one of many possible constructs that can be designed using this versatile technique. Studies are ongoing to determine the effects of different approaches to enzyme incorporation on enzyme activity.

### **Summary & Conclusions**

We have demonstrated a novel fiber mat construct that incorporates a biological material that is active against harmful chemicals. Coaxially electrospun fibers containing DFPase in the sheath and polymer in the core have shown very promising catalytic activity. This process has been optimized to maximize the amount of enzyme (>80%) loaded into the blended sheath layer, while retaining the enzymatic activity. The process is amenable to changes in the polymer make-up of the construct, which provides a means to form reactive fiber mats that would be

active over multiple uses, while protecting the sensitive biological material from environmental damage, leading to inactivation. Future efforts will investigate different layering constructs, as exemplified by the enzyme sandwich described above. This would provide increased environmental protection, as well as potential reusability, and therefore can be studied in conditions that simulate those for extended storage. The coaxial DFPase/polymer fiber formation appears to be a very promising approach for the fabrication of mats or fabrics for decontaminating hazardous chemicals and for protecting people and equipment.

### **Acknowledgement**

The work at the University of Cincinnati is partially supported by a contract with the Natick Soldier Research Development and Engineering Center.

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Table 3 Summary of electrospun fiber mats with linear DFPase-DFP kinetics. (\*Estimated based on experimental conditions such as solute concentration, solution density and flow rates).

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Fig. 10 Multi-layer fiber mat with outer superhydrophobic fibers and inner enzyme containing fibers: (a) schematic of multi-layer fiber mat; (b) demonstration of multi-layer protection of enzyme containing fibers; (c) absorption spectrum (peak wavelength  $\sim 267$  nm) comparison between enzyme/PEO fiber mat and multi-layer fiber mat.

Fig #	Description (wt.% in solution)	Distance (cm)	Applied voltage (kV)	Flow rates (mL/hr) core & sheath
5	PEO(4) core & DFPase(10) sheath	20	13-13.5	0.9 & 0.2
	PEO(4) core & DFPase(4)/PEO(1)/SDS(1) sheath	20	12	0.6 & 0.2
	PCL(10) core & DFPase(4)/PEO(1)/SDS(1) sheath	20	11-12.6	0.5 & 0.2
7	PEO(4) core & DFPase(1)/SDS(0.1) sheath	20	12.5	1.2 & 0.1
	PEO(4) core & DFPase(1)/PEO(2)/SDS(1) sheath	20	13-13.5	0.8 & 0.1
	PCL(10) core & DFPase(1)/PEO(2)/SDS(1) sheath	20	13-15	0.5 & 0.25

Table 1 Coaxial electrospinning conditions for coaxial fiber mats: high DFPase loading (Fig. 5) and low DFPase loading (Fig. 7).

Fig #	Description	Wt. ratio (polymer: DFPase)	Fiber mat weight * (mg)	DFPase amount * (mg)
5	DFPase dip coated PCL fibers	13 : 1	44.19	3.02
	PEO/DFPase blended composite fibers	1 : 1	34.91	17.46
	Coaxial fibers (PEO core & DFPase sheath)	1.8 : 1	21.83	7.80
	Coaxial fibers (PEO core & DFPase/PEO sheath)	3.25 : 1	26.43	5.87
	Coaxial fibers (PCL core & DFPase/PEO sheath)	9.31 : 1	23.95	2.57
7	Coaxial fibers (PEO core & DFPase sheath)	48.1 : 1	24.86	0.52
	Coaxial fibers-R (PEO core & DFPase sheath)	48.1 : 1	24.4	0.52
	Coaxial fibers (PEO core & DFPase/PEO sheath)	34 : 1	20.59	0.57
	Coaxial fibers (PCL core & DFPase /PEO sheath)	30.9 : 1	12.63	0.38

\* Apparent area: 4 inch<sup>2</sup>

Table 2 Summary of material compositions for electrospun fiber mats: high DFPase loading (Fig. 5) and low DFPase loading (Fig. 7). (\*estimated based on experimental conditions such as solute concentration, solution density and flow rates).

#	Description	Fiber diameter (nm)	Surface area * (cm <sup>2</sup> )	Maximum activity**
				[F]/DFPase (mM/mg)
1	Coaxial fibers (PEO core & DFPase sheath)	120 ± 27	8.3 × 10 <sup>3</sup>	7.28
2	Coaxial fibers-R (PEO core & DFPase sheath)	-	-	6.26
3	Coaxial fibers (PEO core & DFPase/PEO sheath)	126 ± 33	6.2 × 10 <sup>3</sup>	3.86
4	Coaxial fibers (PCL core & DFPase /PEO sheath)	218 ± 41	1.3 × 10 <sup>3</sup>	4.51

\* Apparent area: 4 inch<sup>2</sup>, \*\* Apparent area: 1 inch<sup>2</sup>

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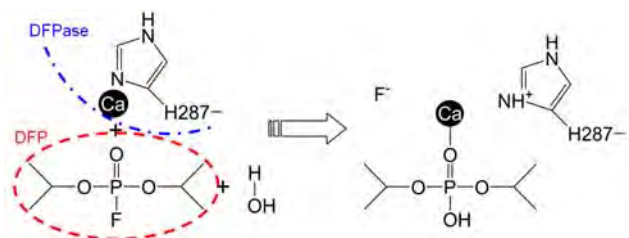


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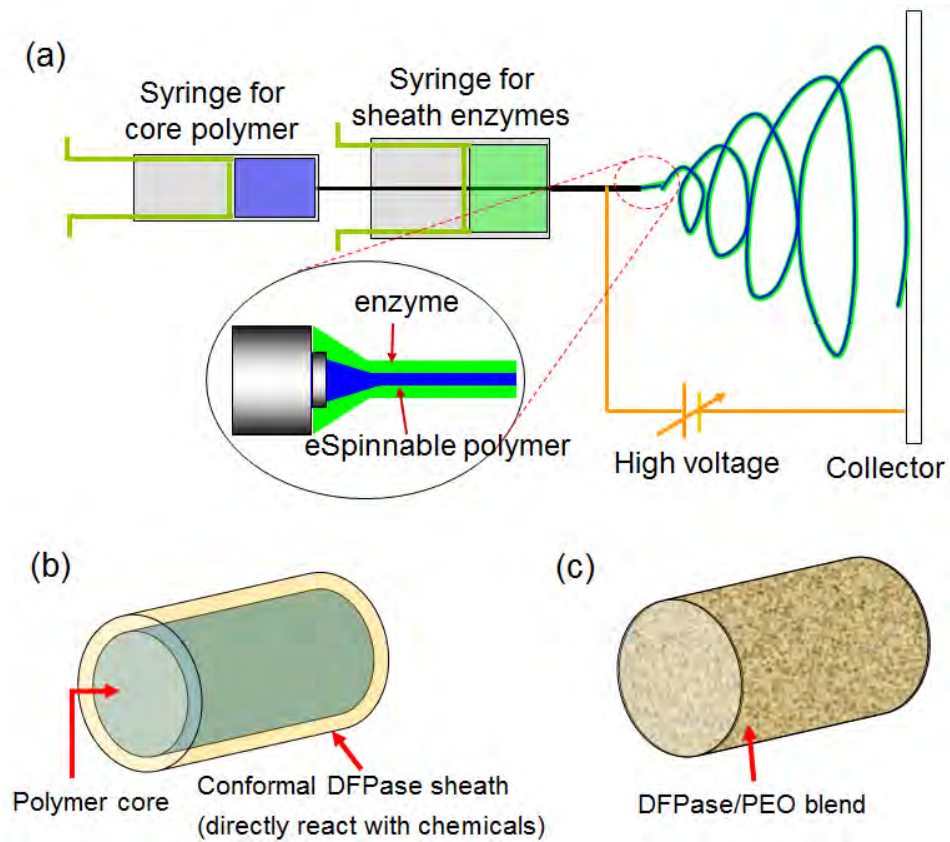


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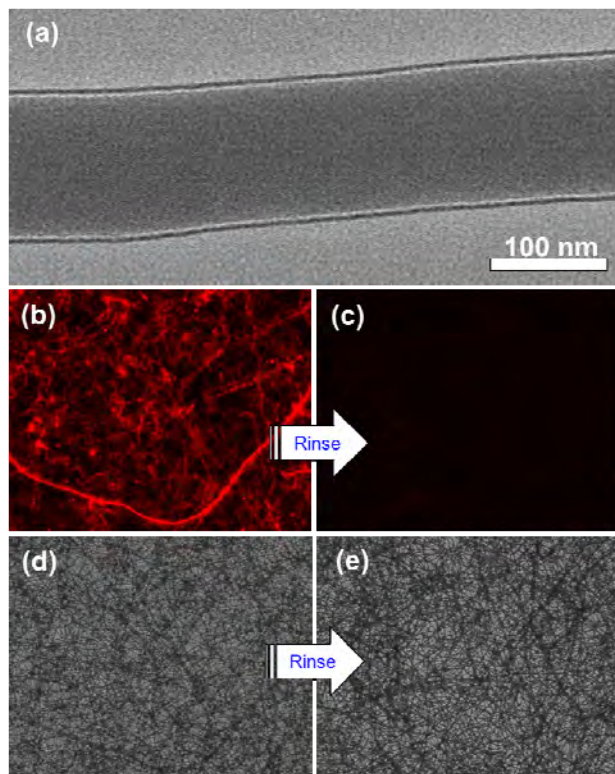


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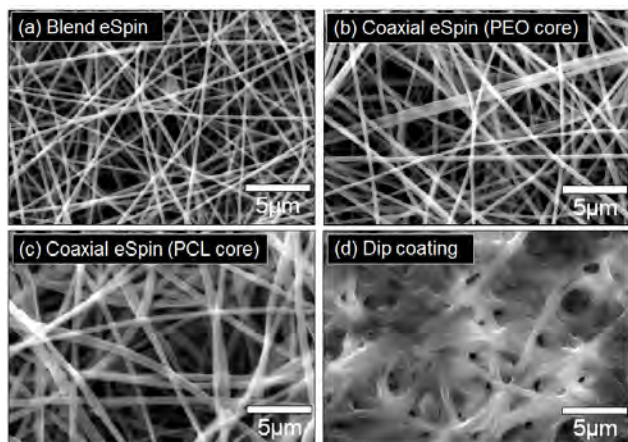


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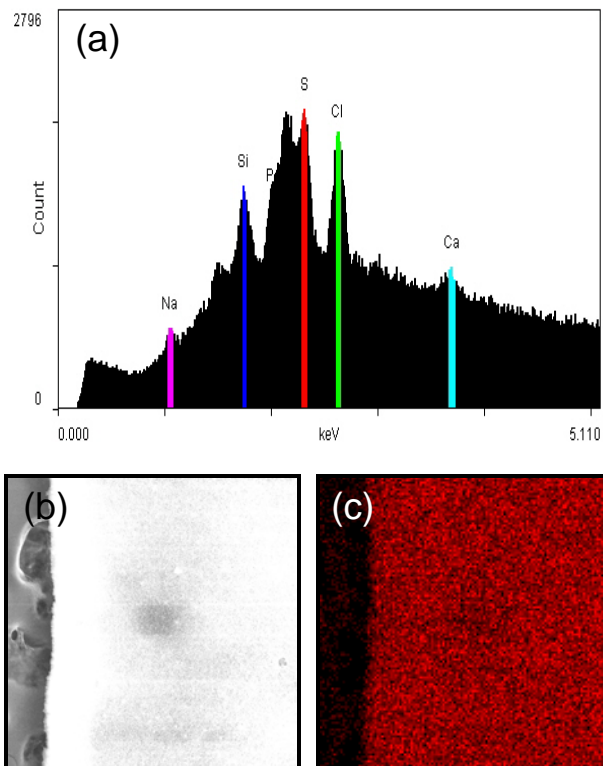


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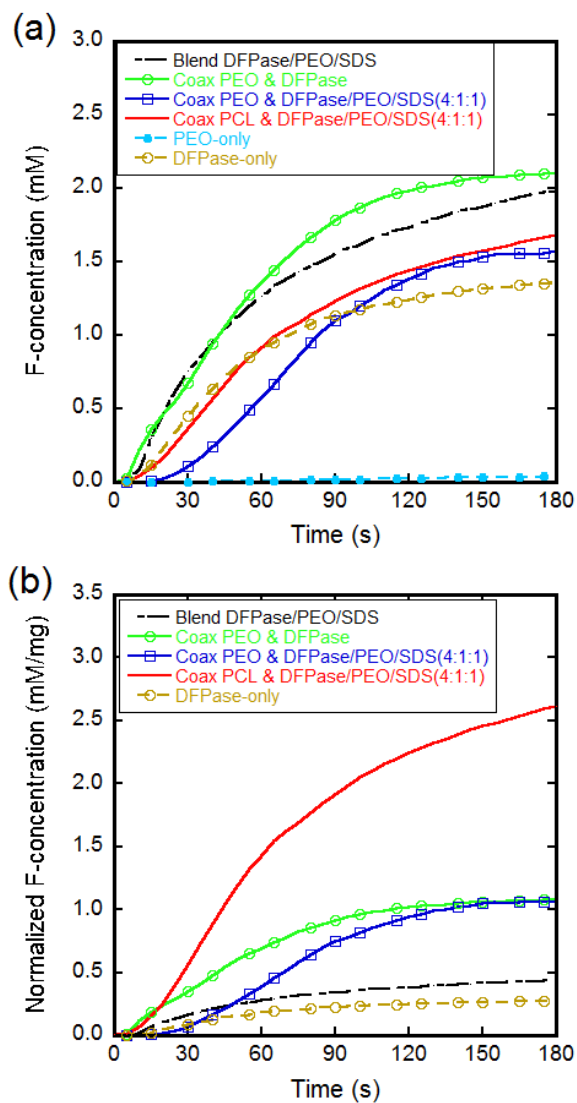


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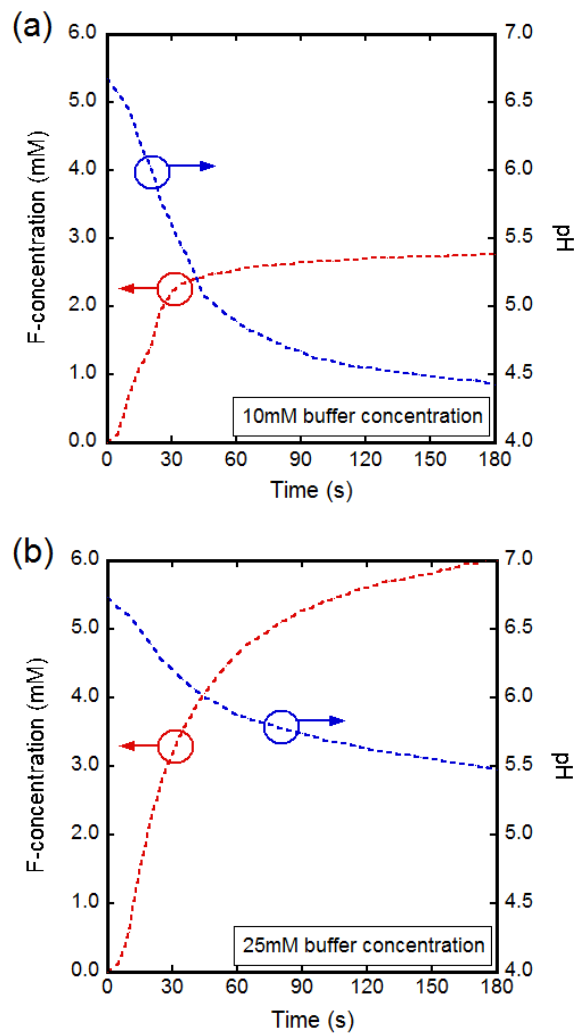


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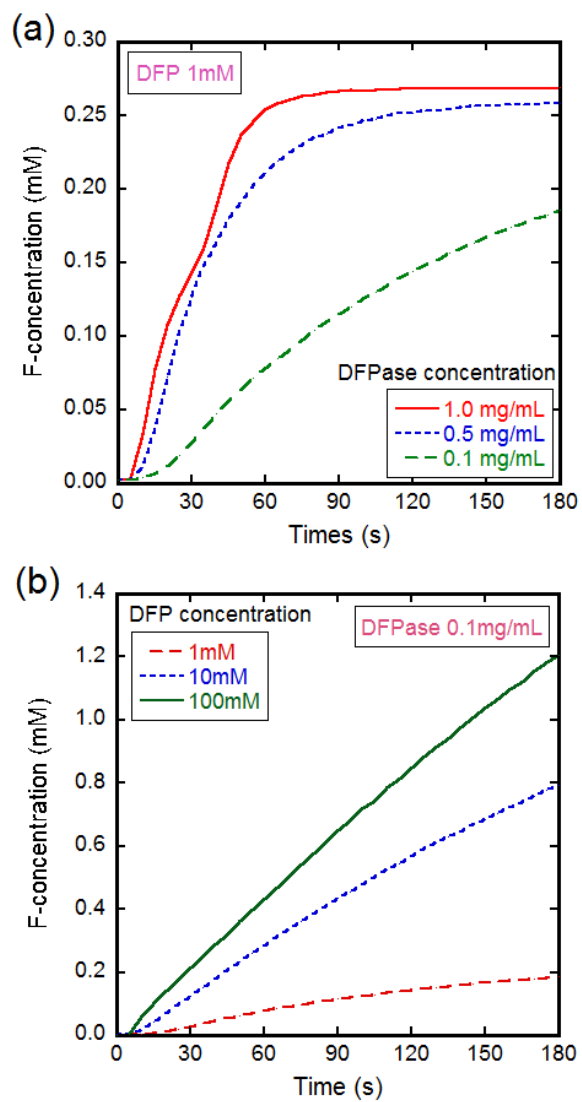


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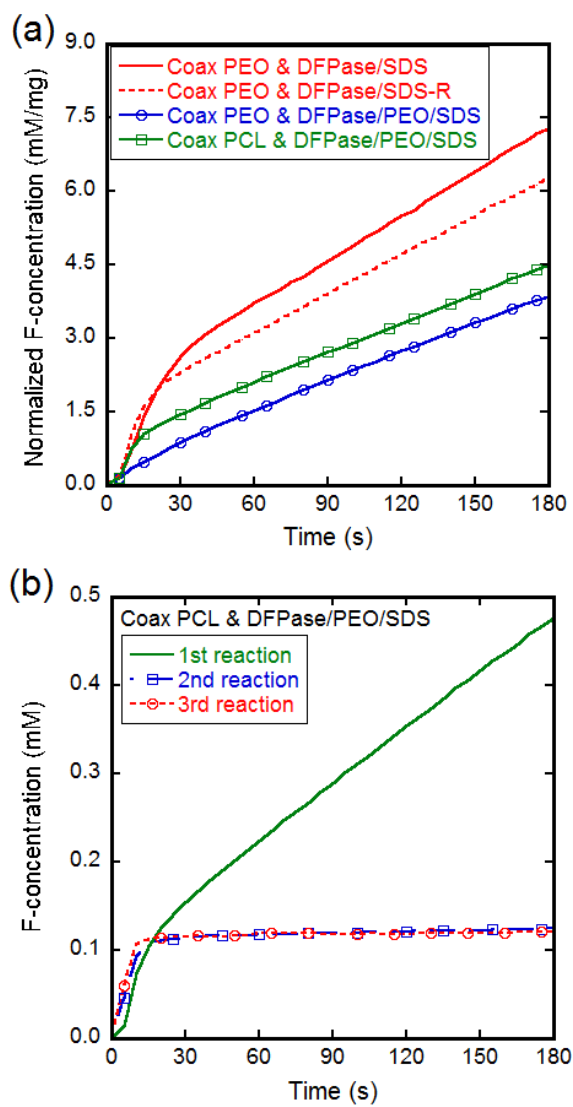


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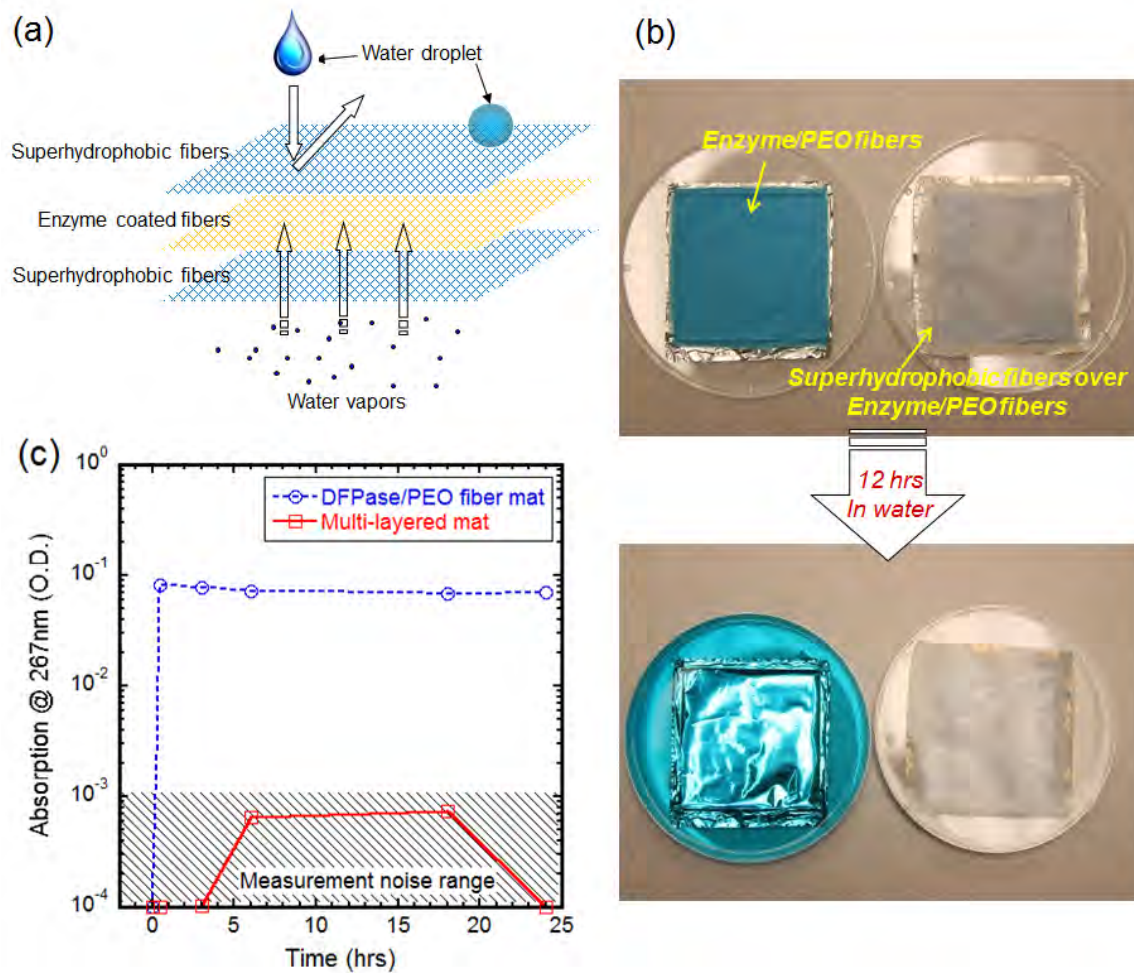


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