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Regulation and Function of TIFAB in Myelodysplastic Syndrome

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INTRODUCTION

Myelodysplastic syndromes (MDS) are clonal bone marrow failure (BMF) disorders defined by blood cytopenias due to ineffective hematopoiesis, genomic instability, and a predisposition to acute myeloid leukemia (AML). The most commonly recurring genomic alteration in MDS is deletion of chromosome 5q (del(5q)). MDS patients with an isolated del(5q) presenting with anemia, neutropenia, and elevated platelets associated with dysplastic megakaryocytes are considered to have 5q- syndrome. The majority of MDS patients with del(5q) do not exhibit these particular symptoms and, instead, are referred to as “del(5q) MDS”. We have recently identified miR-146a, which target the TRAF6 arm of the innate immune pathway, a gene within the deleted region in del(5q) MDS. We posit that multiple genes on chr 5q coordinate TRAF6 activation in del(5q) MDS. A search of annotated genes within or near the CDRs revealed a known inhibitor of TRAF6, TIFAB, on band q31.1. We hypothesize that deletion of TIFAB promotes activation of the TRAF6 complex in human CD34⁺ cells resulting in hematopoietic defects resembling MDS with del(5q). The overall objectives of this proposal are to (1) determine whether loss of TIFAB in human CD34⁺ cells contributes to MDS in mice; (2) to investigate whether deletions of TIFAB activate TRAF6 in MDS; and (3) to determine the consequences of TIFAB deletion on signal transduction in human CD34⁺ cells, and whether these could explain features of MDS. In preliminary data from the first 2 year of the proposal, we have evidence that TIFAB is a regulator of human hematopoietic cells. Our key observations show that knockdown of TIFAB in human CD34⁺ hematopoietic stem/progenitor cells results in increased survival and proliferation, TIFAB inhibits TRAF6 protein expression and activation, resulting in lower NF-κB activation, and TIFAB expression impacts leukemic cell survival, growth and progenitor function. Given that TIFAB is deleted in many MDS patients, these findings could have major implications in MDS subtypes with deletions of chr 5q. The observation that del(5q) results in inappropriate activation of TRAF6 provides a strong rationale to study the contribution of TIFAB to deregulation of the TRAF6 pathway in MDS.

BODY

Task 1. Plasmid constructs and validation (months 1-4):

1a (Complete). For knockdown of TIFAB in human CD34⁺ or MDS/AML cell lines, we used RNAi-mediated gene silencing. Lentiviral vectors encoding two independent shRNAs targeting human TIFAB were purchased from OpenBiosystems. The bicistronic lentiviral vector contains a microRNA-adapted shRNA and a CMV-driven turboGFP (Figure 1A)¹.

1c (Complete). For qRT-PCR and immunoblot analysis to determine TIFAB knockdown, two independent shRNAs targeting TIFAB (#88 and #89) and a scrambled control vector (shCTL) were transduced into human HL60 and THP1 leukemia cell lines. Two days post transduction, cells were sorted for GFP expression and expanded for an additional 2 days in culture. RNA and protein were collected and examined for TIFAB knockdown by qRT-PCR (Figure 1B) and immunoblotting (Figure 1C), respectively. In addition, we have also confirmed efficient knockdown in primary human cord blood CD34⁺ cells (Figure 2D).

1d. The original plan involved knocking down both miR-145 and miR-146a in primary hematopoietic cells using a miRNA decoy retroviral vector. Given that more recent findings indicate that miR-146a specifically targets the TRAF6 pathway and its deletion results in many

MDS-like features in mice^{2,3}, we have opted to clone only the miR-146a decoy. We have generated a decoy that knocks down the expression of miR-146a only. To confirm knockdown, we have transduced human CD34+ cells and performed qRT-PCR analysis for miR-146a knockdown. Unfortunately, the miRNA decoy is not efficient at knocking down miR-146a (Figure 1E) and does not affect survival (Figure 1F) in primary human CD34+ cells (after repeated attempts). Given the technical difficulties of simultaneously knocking down miR-146a and TIFAB, we have focused our attention on the posttranscriptional regulation of TRAF6 (See below).

Task 2. Expression analysis of TIFAB and TIFA in hematopoietic cells (months 1-6):

2a (Complete). Normal expression patterns of TIFA and TIFAB were investigated in human marrow subpopulations. This task is complete.

Task 3. Isolation and infection of CD34+ cells (months 4-18):

3a/b (Complete). Human umbilical cord CD34+ cells were obtained and cultured in vitro. CD34+ cells were transduced with shTIFAB-GFP, sorted, and evaluated in vitro. Transduction efficiency was 4% for shTIFAB. As indicated above, knockdown of TIFAB resulted in a reproducible downregulation of TIFAB mRNA by ~50%, which is consistent with the expression in del(5q) MDS patient cells (Figure 1D).

3c. As described above, the miR-146a decoy is not efficient at knocking down miR-146a in CD34+ cells despite reasonable transduction efficiency. Given that we are not able to progress with the miR-146a knockdown experiments, we have extended our analysis of TIFAB knockdown to a preleukemic CD34+ cells expressing AML1-ETO fusion (AE)⁴. Since AE CD34+ cells are not fully immortalized, we will determine whether knockdown of TIFAB in AE CD34+ cells will make them malignant. The same assays will be performed as proposed for normal CD34+ experiments (Task 4 and 5).

Task 4. Application of *in vitro* hematopoietic assays (months 8-12):

4a (Complete). Transduced shTIFAB (and control) CD34+ cells from Task 3 were used to determine the (i) proliferation, (ii) survival, and (iii) clonogenic potential in methylcellulose containing differentiation cytokines (IL3, IL6, SCF, and Epo).

- (i) For proliferation assays, transduced and sorted CD34+ cells were cultured in vitro and evaluated for trypan blue exclusion at the indicated times. Knockdown of TIFAB resulted in increased proliferation of CD34+ cells (Figure 2A).
- (ii) For survival assays, transduced and sorted CD34+ cells were cultured for 4 days and then evaluated for AnnexinV staining. As show in in Figure 2B knockdown of TIFAB resulted in a significant reduction of apoptotic (AnnexinV+) cells (Figure 2B).
- (iii) For clonogenic potential, transduced and sorted CD34+ cells were plated into methylcellulose and then scored for colony formation after 10 days. The number of colonies formed following knockdown of TIFAB was similar as compared to control-transduced cells (Figure 2C). When the proportion of colony types was examined, knockdown of TIFAB resulted in slight expansion of CFU-GM colonies at the expense of CFU-G colonies (Figure 2C); however,

these differences are not significant. Therefore, we conclude that knockdown of TIFAB does not alter the differentiation of CD34+ cells.

- (iv) For in vitro differentiation, transduced CD34+ were cultured for ~ 7 days in StemSpan media containing 10ng/mL IL3, IL6, TPO, FLT3, and SCF. Immature (CD34+) and mature myeloid (CD33+) cell surface expression was determined by FACS. As show in Figure 2D, knockdown of TIFAB did not maintain cells in the immature CD34+ state. Conversely, CD34+ cells with knockdown of TIFAB exhibited an increase of mature CD33+CD34- cells.

These observations suggest that lower levels of TIFAB increase survival and proliferation of CD34+ cells but does not promote/maintain immature CD34+ cells.

Task 5. Application of bone marrow transplantation assays and analysis (months 8-36):

5a. (Partly Complete) We have transduced CD34+ cells with shCTL and shTIFAB and are ready for transplantation into NSGS mice (NOD/SCID mouse with IL2R γ knockout and transgenic expression of SCF, GM-CSF, and IL-3)⁵. The NSGS colony is maintained by our laboratory and ready for xenotransplantation. For the first experiment, we are prepared to engraft ~10 mice per group (shCTL and shTIFAB). Mice will be analyzed according to our original plan (Task 5b-5d). In addition, we will xenograft AE-CD34+ cells transduced with shCTL or shTIFAB. As for xenotransplantation of normal CD34+ cells, mice receiving AE-CD34+ will be monitored for MDS/AML-like features and hematopoietic defects.

Task 6. Identification of changes in TRAF6 activation and NF- κ B signaling by TIFAB (months 10-14):

It was previously reported that TIFAB functions by suppressing TRAF6-mediated NF- κ B activation^{6,7}. However, the mechanism by which TIFAB may inhibit TRAF6 and/or NF- κ B is not known. Based on our data from Year 1, we find that TIFAB reduces the TRAF6 protein and function. One of the key findings was that TIFAB suppressed LPS-induced NF- κ B activation but not TNF-induced NF- κ B activation. This observation is consistent with the finding that TIFAB inhibits TRAF6 function and stability. Based on this interesting finding, the experiments in Year 2 were focused on determining the effects of TIFAB on TRAF6-induced NF- κ B activation.

6a. (Complete) We have measured IL-6 and BFL1A1 (two NF- κ B target genes) in CD34+ cells transduced with shTIFAB (Figure 3A). We observed an ~1.5 increase in IL6 expression but no significant changes in BFL1A1 in cells with knockdown of TIFAB. This initial observation suggests that TIFAB does not regulate all functions of NF- κ B signaling.

6b. (Complete) We have devoted a significant effort to dissect the role of TIFAB on NF- κ B activation. Our initial efforts have been performed in HEK293 cells using NF- κ B luciferase reporter assays:

- i. To investigate the dosage effects of TIFAB on TNF or LPS-induced NF- κ B activation, we transfected increasing amounts of pcDNA-TIFAB into HEK293 cells and then stimulated with TNF (1 ng/ml) or LPS (1 ug/ml). As shown in Figure 3C, TIFAB (even at the lowest dose) efficiently inhibited LPS-induced kB-site luciferase activity, but only inhibited TNF-induced kB-luciferase at the highest dose (Figure 3B). Conversely, increasing the amount of LPS did not overcome the inhibitory effects of TIFAB on NF- κ B activation (Figure 3D).
- ii. To investigate the dosage effects of TRAF6 on TIFAB repression of NF- κ B, we transfected increasing amounts of pcDNA-TRAF6 and pcDNA-TIFAB into

- HEK293 cells. At all doses examined, TIFAB expression suppressed TRAF6-mediated kB-site activation (Figure 3E).
- iii. To further understand the specificity of TIFAB-mediated inhibition of NF-kB, we cotransfected various NF-kB activators and then assessed the effects of TIFAB (Figure 4A). As shown before, TIFAB effectively inhibits TRAF6-mediated activation of kB site-luciferase (Figure 4B). Since TRAF6 shares functional and sequence homology with TRAF2, we examined the effects of TIFAB on TRAF2-mediated activation of NF-kB (Figure 4C). Interestingly, TIFAB was not able to inhibit TRAF2-mediated kB site-luciferase activity. Since we hypothesize that TIFAB directly inhibits TRAF6, we wanted to determine whether transfecting in IKK (NF-kB kinase) or p65 (NF-kB transcription factor), which are both downstream of TRAF6, could still maintain kB site-luciferase activity in the presence of TIFAB (Figure 4A). As expected, IKK or p65 induced kB site-luciferase activity even in the presence of TIFAB (Figure 4D, 4E). These findings reinforce the hypothesis that TIFAB directly and specifically inhibits TRAF6-mediated NF-kB activation.
 - iv. To corroborate the NF-kB reporter assays, we investigated the consequences of TIFAB expression on TRAF6 by performing immunoblots (Figure 5A). As shown, we observed reduced levels of endogenous or co-transfected TRAF6 protein (Figure 5A). Moreover, expression of TIFAB reduced levels of co-transfected TRAF6, whereas the levels of TRAF2 were not affected (Figure 5B). The effects of TIFAB on TRAF6 protein are independent of *TRAF6* mRNA as overexpression of TIFAB did not reduce *TRAF6* mRNA in HEK293 cells (Figure 5C). To explore the possibility that TIFAB induces TRAF6 protein degradation, we used selective inhibitors of the proteasome (MG-132) or lysosome (3-MA). Treatment of HEK293 cells expressing TRAF6 and TIFAB with MG-132 did not restore TRAF6 protein levels (Figure 5D), indicating that TIFAB-mediated degradation of TRAF6 does not occur through the ubiquitin-proteasome pathway. However, treatment with 3-MA partially restored TRAF6 protein expression in TIFAB-expressing HEK293 cells (Figure 5D), suggesting that TIFAB-mediated degradation may occur through the lysosome/autophagy pathway.
 - v. TIFAB is 161 amino acids (aa) and contains a fork-head associated domain (FHA) spanning ~ aa 36-91 (Figure 6A). To identify the critical region for TIFAB-mediated inhibition of TRAF6, we created TIFAB deletion mutants (Figure 6A). Each mutant was fused to the FLAG epitope to conveniently detect expression by immunoblotting. Following transfection into HEK293 cells, the protein expression of the TIFAB deletion mutants was confirmed by FLAG immunoblots (Figure 6D). The consequences of TIFAB domain deletions on basal NF-kB activation were first evaluated without TRAF6 expression. As shown in Figure 6B, TIFAB D1-34, D92-161, and D35-161 did not inhibit basal kB-site luciferase activity; in contrast, the level of NF-kB appeared to increase when these mutants were expressed (Figure 6B). Mutant D1-91 inhibited basal NF-kB activation (Figure 6C).
 - vi. Next the TIFAB mutants were evaluated for their ability to repress TRAF6-mediated kB site-luciferase activity. As before, full-length TIFAB repressed TRAF6-mediated kB site activity (Figure 6C). Similar to the trend observed for

basal NF- κ B repression, TIFAB D1-34, D92-161, D36-91, and D35-161 did not efficiently repress TRAF6-mediated kB site activity (Figure 6C). However, D1-91 significantly repressed TRAF6-mediated kB site activity (Figure 6C). To extend these observations, we evaluated TRAF6 protein expression following transfection of the TIFAB mutants. Consistent with the kB-luciferase assays, D35-91 and D1-91 reduced endogenous TRAF6 expression (Figure 6D). These findings suggest that the inhibitory domain of TIFAB resides within aa 92-161. To further define the minimal sequence necessary to inhibit TRAF6, we have created additional TIFAB mutants: D1-125, D1-91+D126-161, D126-161, and D91-126 (Figure 6A).

- vii. To examine the cellular effects of restoring the minimal TIFAB domain that retains TRAF6 inhibition in disease-relevant cells, we transduced HL60 cells with TIFAB- and D1-91-expressing vectors. Although restoring full-length TIFAB had anti-leukemic effects in HL60 cells, restoring only the C-terminal domain (D1-91) in HL60 resulted in more pronounced anti-leukemic cell function. The C-terminal domain containing mutant (D1-91) inhibited proliferation ($P = 2.6 \times 10^{-6}$; Figure 7A), apoptosis (Figure. 7B), and impaired progenitor function ($P = 0.005$; Figure 7C) more dramatically than full-length TIFAB. Thus, restoring only the C-terminal domain of TIFAB is sufficient to destabilize TRAF6 protein, suppress NF- κ B activation, and impair leukemic progenitor function and survival.

Task 7. Gene expression analysis (months 16-24).

(Complete). Utilizing a novel TIFAB knockout mouse, we performed an expression analysis on sorted LSK isolated from 3 month old mice transplanted with WT or *Tifab*^{-/-} BM cells (n = 3 mice/group). We selected this time point to capture the gene expression profile of *Tifab*^{-/-} LSK after engraftment but prior to overt hematopoietic failure. Analysis of the gene expression data identified differentially expressed genes in *Tifab*^{-/-} LSK cells including 32 upregulated (> 1.7-fold) and 22 downregulated (< 1.7-fold). To elucidate the relevant signaling pathway regulated following *Tifab* loss in HSPC, we performed a gene set enrichment analysis (GSEA)⁸, which revealed overexpression of immune and infection response signatures, particularly related to interferon-regulated genes, and a TP53 response gene signature (Figure. 8A-B). Downregulated gene signatures in *Tifab*^{-/-} LSK also included immune response signatures, but these were related to enterotoxin and lipopolysaccharide (LPS) (Figure. 8A-B). Select genes were validated by qRT-PCR under basal and during LPS stimulation. TIFAB deficient cells exhibit hypersensitivity to LPS-induced gene expression.

Task 8. Validation of targets (months 24-36)

(Not complete). We were unable to perform this task within the specified time period.

Extra Tasks: Investigating the function of TIFAB by identifying interacting proteins:

(Initial Analysis Complete/Ongoing). The function of TIFAB is not well defined. To further identify the molecular role of TIFAB in MDS/AML cells, we performed a proteomics experiment to identify proteins that bind to TIFAB. For these experiments, we created a FLAG-tagged TIFAB cDNA in a retroviral vector. HL60 cells were transduced with vector or FLAG-TIFAB. HL60 cells expressing vector or TIFAB were immunoprecipitated with FLAG-containing beads. FLAG-TIFAB complexes were eluted off the beads and run on a

polyacrylamide gel for silver stain analysis. Bands that were unique to the lane containing FLAG-TIFAB (and the control bands) were cut from the gel and evaluated by mass spectrometry (Figure 9A). This was repeated in biological duplicates. TRAF6 was identified in the mass spectrometry analysis, indicating that TIFAB and TRAF6 form a complex. We further validated this by co-immunoprecipitation experiments in HL60 and HEK293 cells (Figure 9B-C). Based on this initial analysis, we have identified at least two unique TIFAB-interacting proteins. Future experiments will validate the interaction between TIFAB and USP15 or DDX21 and the consequences of this interaction on TRAF6 function. We will test the hypothesis that TIFAB-mediated inhibition of TRAF6 has consequences on USP15 and/or DDX21 function.

KEY RESEARCH ACCOMPLISHMENTS

The research generated during the funding period was extremely exciting and informative to the biology of del(5q) MDS. The new information is transformative in how we view immune regulation and the pathogenesis of del(5q) MDS. We have highlighted some of the key findings below.

- Identified an shRNA lentiviral vector containing a GFP reporter that knocks down the expression of human TIFAB by approximately 50% in human CD34+ cells.
- Successfully transduced normal CD34+ cells and human leukemic cell lines to evaluate TIFAB knockdown with the shRNA.
- Determined that knockdown of TIFAB in human CD34+ cells resulted in increased CD34+ survival and increased proliferation, without affecting differentiation potential.
- Discovered that TIFAB suppresses active TRAF6 and reduces TRAF6 protein expression in human cell lines.
- Determined that TIFAB suppresses NF- κ B activation following LPS/TRAF6-mediated stimulation, but not via TRAF2/TNFR.
- TIFAB mediates degradation of TRAF6 via lysosomes.
- Narrowed the region of TIFAB that possess the inhibitory function (between aa 91-161).
- Identified ~20 novel TIFAB interacting proteins by mass spectrometry.
- This work prompted us to generate a novel TIFAB knockout mouse.

REPORTABLE OUTCOMES

Accepted Research Article:

Fang J, G Rhyasen, L Bolanos, C Rasch, M Varney, M Wunderlich, S Goyama, G Jansen, J Cloos, C Rigolino, A Cortelezzi, JC Mulloy, EN Oliva, M Cuzzola, **DT Starczynowski**. (2012). Cytotoxic effects of Bortezomib in MDS/AML depend on autophagy-mediated lysosomal degradation of TRAF6 and repression of PSMA1. *Blood*, 120(4): 858-867. PMID: 22685174

Guo F, S Zhang, M Grogg, JA Cancelas, ME Varney, **DT Starczynowski**, W Du, J Yang, W Liu, G Thomas, S Kozma, Q Pang, Yi Zheng. Mouse gene targeting reveals an essential role of mTOR in hematopoietic stem cell engraftment and hematopoiesis. (2013). *Haematologica*, 98(9): 1353-1358. PMID: 23716557.

Rhyasen G, L Bolanos, J Fang, C Rasch, A Jerez, M Varney, M Wunderlich, C Rigolino, L Mathews, M Ferrer, N Southall, R Guha, J Keller, C Thomas, L Beverly, A Cortelezzi, EN Oliva, M Cuzzola, J Maciejewski, JC Mulloy, **DT Starczynowski**. Targeting IRAK1 as a novel therapeutic approach for Myelodysplastic Syndrome. (2013). *Cancer Cell*, 24(1): 90-104. PMID: 23845443.

Rhyasen G, L Bolanos and **DT Starczynowski***. Differential IRAK signaling in hematologic malignancies. (2013). *Experimental Hematology*, 41(12): 1005-1007. PMID: 24084080.

Rhyasen G, M Wunderlich, K Tohyama, G Garcia-Manero, JC Mulloy, **DT Starczynowski**. An MDS xenograft model utilizing a patient-derived cell line. (2014). *Leukemia*. 28(5): 1142-1145. PMID: 24326684.

Zhao JL, and **DT Starczynowski**. Role of microRNA-146a in hematopoietic stem cell function and myelodysplasia. (2014). *Frontiers Genetics*. 5:219. PMID: 25071842.

Fang J, B Barker, L Bolanos, X Liu, A Jerez, H Makishima, S Christie, X Chen, DS Rao, HL Grimes, Komurov, MT Weirauch, J Cancelas, JP Maciejewski, **DT Starczynowski**. Myeloid malignancies with chromosome 5q deletions acquire a dependency on an intrachromosomal NF- κ B gene network. (2014). *Cell Reports*. 11;8(5): 1328-1338. PMID: 25199827.

Research Articles Submitted:

Varney ME, M Niederkorn, K Konno, T Matsumura, J Gohda, N Yoshida, T Akiyama, S Christie, J Fang, D Miller, A Jerez, A Karsan, JP Maciejewski, J-I Inoue, **DT Starczynowski**. *Tifab* is required for restricting innate immune signaling and hematopoietic stem/progenitor function in del(5q) myelodysplasia. Under revision.

Abstracts at Conferences:

Melinda E. Varney, Andres Jerez, Jing Fang, David Miller, Lyndsey Bolanos, Aly Karsan, Jaroslaw P. Maciejewski, Daniel T. Starczynowski. American Society of Hematology, Atlanta, GA, December, 2012.

Melinda E. Varney, Andres Jerez, Jing Fang, David Miller, Lyndsey Bolanos, Aly Karsan, Jaroslaw P. Maciejewski, Daniel T. Starczynowski. Ohio State University Cancer Center Conference, April, 2013.

Awards:

ASH 2012 Abstract Achievement Award (for Melinda Varney)

CONCLUSIONS

The funding period for the project has yielded many interesting results and new research directions, most of which support our original hypothesis. Given that there are few publications on TIFAB, we had to develop novel assays and reagents. Overall, the experiments suggest that TIFAB, a novel and uncharacterized protein, exhibits tumor suppressor-like functions in human hematopoietic cells. Our key observations show that (1) TIFAB is primarily expressed in

hematopoietic progenitor cells (rather than in primitive hematopoietic stem cells or mature blood cells; (2) knockdown of TIFAB in human CD34+ hematopoietic stem/progenitor cells results in increased survival and altered hematopoietic progenitor function; (3) TIFAB inhibits TRAF6 protein expression and activation, resulting in lower NF- κ B activation; and (4) TIFAB expression impacts leukemic cell survival, growth and progenitor function. Given that TIFAB is deleted in many MDS patients, these findings could have major implications in MDS and AML subtypes with deletions of chr 5q.

As indicated above, majority of the goals in the Statement of Work have been accomplished within the 3 years of the proposal. In addition, our ongoing experiments support our hypothesis, permitting us to continue with our original outline of experiments. However, there have been a few minor alterations to the Statement of Work:

- We have limited our analysis of TIFA as it appears that TIFAB has a major effect on TRAF6 independent of TIFA levels. In addition, TIFA expression did not differ in hematopoietic subpopulations. As such, we propose to delay examining TIFA's potential role in linking TIFAB and TRAF6.
- Based on our preliminary findings, knockdown of miR-146a does not appear to be feasible in primary human CD34+ cells. As such we have opted to temporarily not focus on miR-146a until we find an alternative strategy. In place of these original experiments, we propose to focus on the direct inhibitory function of TIFAB on TRAF6 by performing detailed structure functional analysis and identification of novel TIFAB interacting proteins.
- Since we have optimized consistent knockdown of TIFAB in primary CD34+ cells, we will also investigate the consequences of knocking down TIFAB in preleukemic CD34+ that contain an AML1-ETO fusion gene. As with primary CD34+ cells, we will determine the effects of TIFAB knockdown on proliferation, progenitor function, survival, and xenotransplantation in NSGS mice.

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4. Mulloy, J.C. *et al.* The AML1-ETO fusion protein promotes the expansion of human hematopoietic stem cells. *Blood* **99**, 15-23 (2002).
5. Wunderlich, M. *et al.* AML xenograft efficiency is significantly improved in NOD/SCID-IL2RG mice constitutively expressing human SCF, GM-CSF and IL-3. *Leukemia* **24**, 1785-8 (2010).
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8. Subramanian, A. *et al.* Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A* **102**, 15545-50 (2005).

APPENDICES

- Figures 1-9
- Curriculum Vitae

Figure 1.

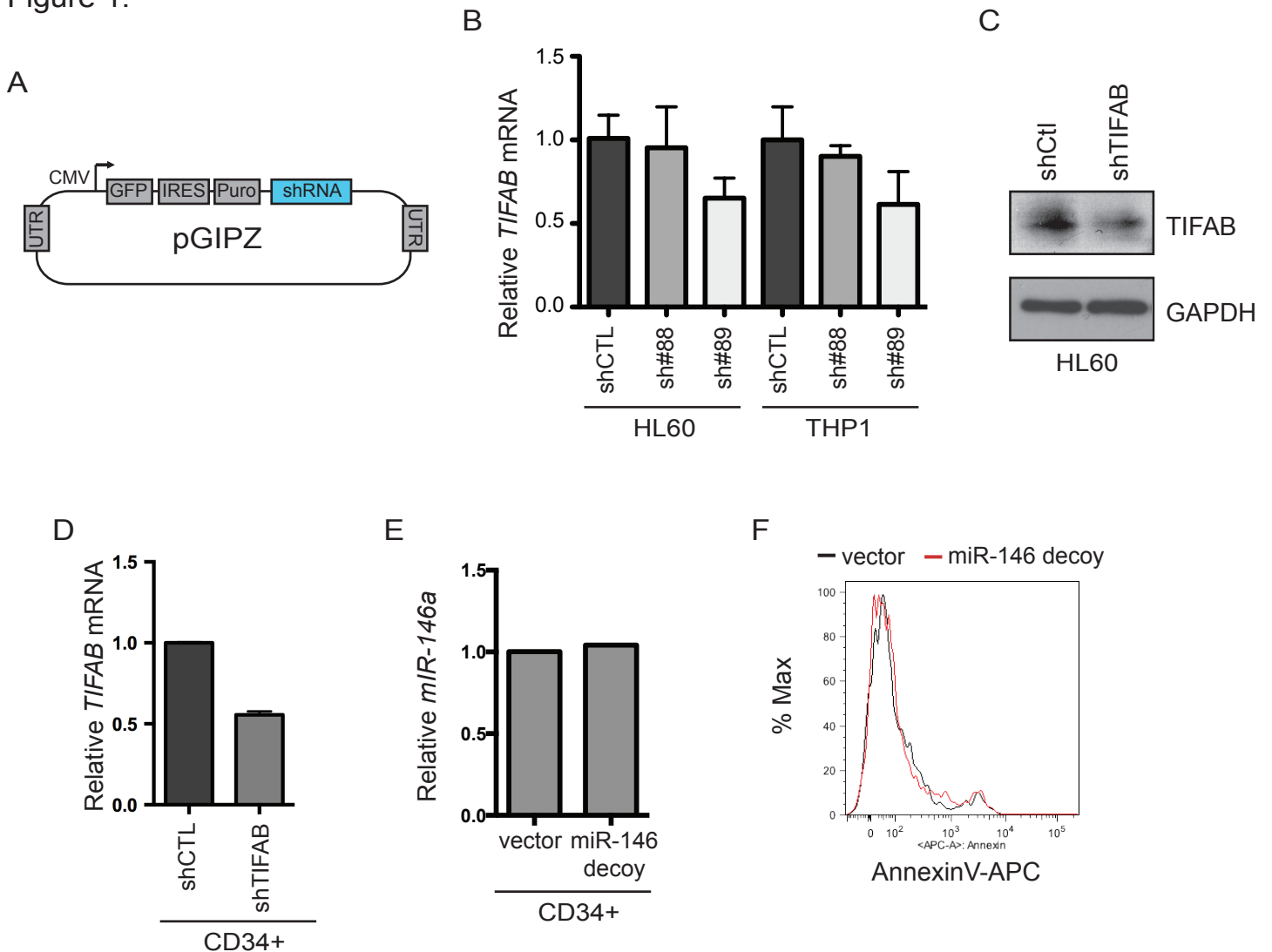


Figure 1: Task 1

A. Schematic representation of the pGIPZ lentiviral vector for knockdown of TIFAB is shown. On The bicistronic miRNA-based shRNA is driven by a CMV promoter and also expresses GFP for tracking expression in transduced cells. For simplicity, cPPT, WRE, Amp, pUC, and Ori are not shown.

B. qRT-PCR was performed on HL60 and THP1 cells transduced with the indicated shRNA-containing lentiviral vectors.

C. THP1 cells were transduced with control shRNA or shTIFAB and analyzed for TIFAB expression by immunoblotting with the indicated antibodies.

D. qRT-PCR was performed on primary CD34+ cord blood cells transduced with the indicated shRNA-containing lentiviral vectors.

E. pGK-miR-146a-GFP decoy was transduced into CD34+ cells. RNA was collected and analyzed by qRT-PCR for miR-146a expression.

F. CD34+ cells transduced with miR-146a decoy were evaluated for AnnexinV staining.

Figure 2.

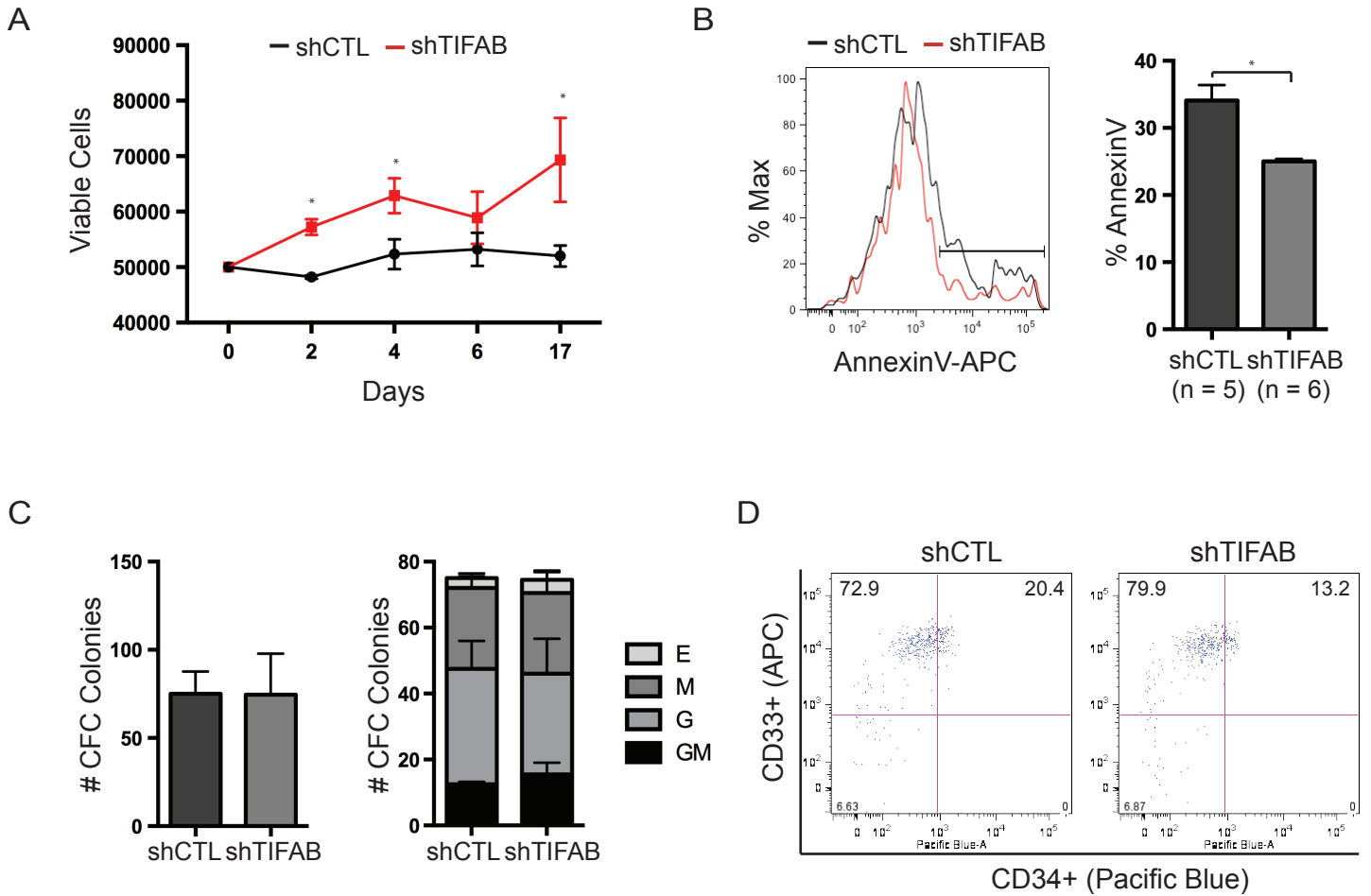


Figure 2: Task 4

A. 1×10^6 human CD34⁺ cells were transduced with control shRNA or shTIFAB (clone #89) and then sorted for GFP⁺ cells. 5×10^4 transduced cells were cultured in vitro and counted at the indicated days.

B Transduced CD34⁺ cells were evaluated for AnnexinV staining. The histogram is the summary of 2 independent transductions.

C. 5×10^4 transduced CD34⁺ were plated in methylcellulose and analyzed for colony formation. Colonies were scored after 14 days. E, erythroid; M, monocyte; G, granulocyte.

D. Transduced CD34⁺ cells were evaluated for CD34 and CD33 staining after 7 days in culture.

Figure 3.

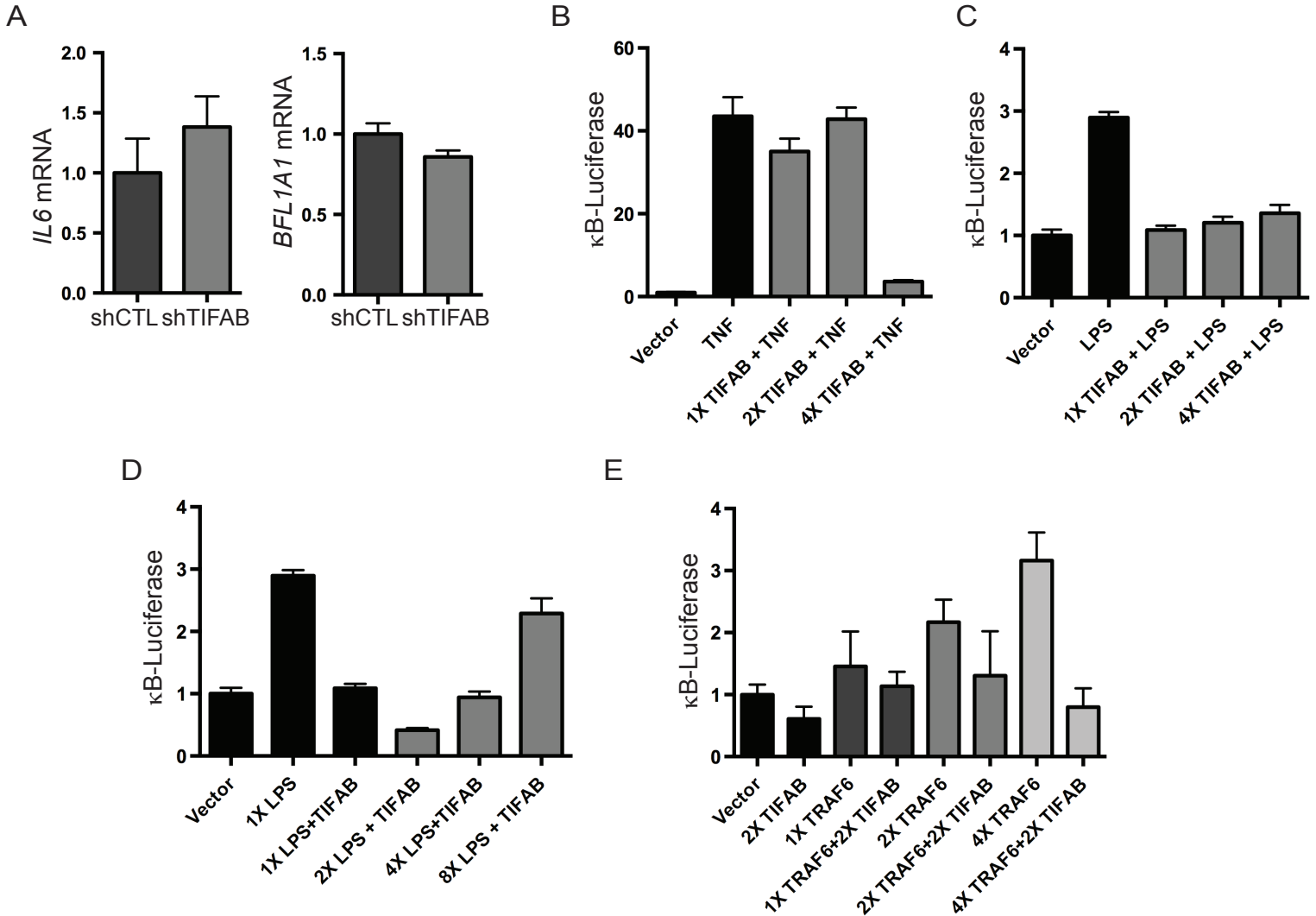


Figure 3: Task 5

A. qRT-PCR was performed for IL6 and BFL1A1 expression in human CD34+ transduced with the indicated vectors.

B. HEK293 cells were transfected with pcDNA3.1 (vector, 250 ng) or pcDNA3.1-FLAG-TIFAB (250, 500, or 1000 ng) and κB-luciferase. Following transfection, cells were simulated with TNFα (1 ng/ml) for 6 hours. Values represent relative luciferase.

C. HEK293 cells were transfected with pcDNA3.1 (vector, 250 ng) or pcDNA3.1-FLAG-TIFAB (250, 500, or 1000 ng) and κB-luciferase. Following transfection, cells were simulated with LPS (1 ug/ml) for 6 hours. Values represent relative luciferase.

D. HEK293 cells were transfected with pcDNA3.1 (vector, 250 ng) or pcDNA3.1-FLAG-TIFAB (250 ng) and κB-luciferase. Following transfection, cells were simulated with LPS (1, 2, 4, or 8 ug/ml) for 6 hours. Values represent relative luciferase.

E. HEK293 cells were transfected with pcDNA3.1 (vector, 250 ng), pcDNA3.1-FLAG-TIFAB (250, 500, or 1000 ng), and/or pcDNA3.1-FLAG-TRAF6 (250, 500, or 1000 ng) and κB-luciferase. Values represent relative luciferase.

Figure 4.

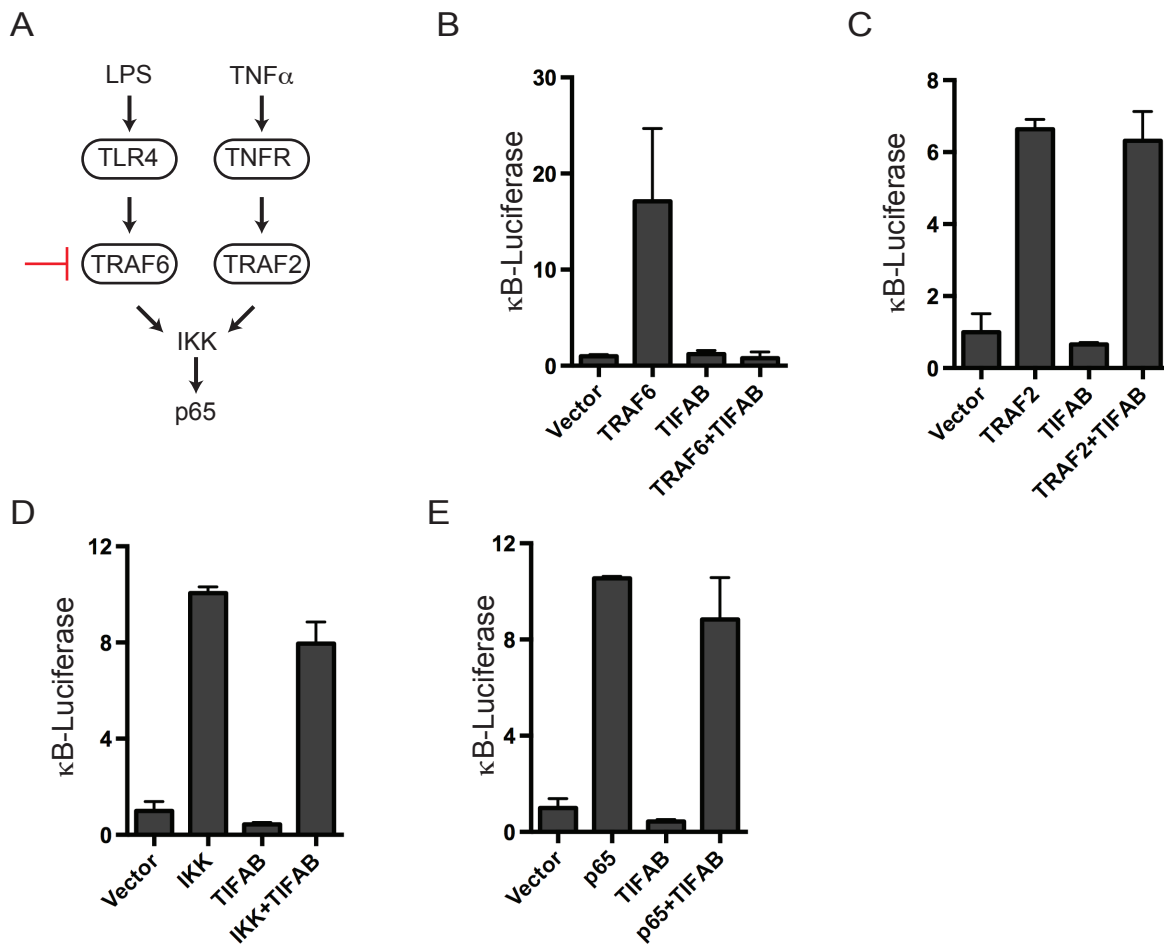


Figure 4: Task 6b

A. Model distinguishing LPS/TLR4 and TNF α /TNFR activation of NF- κ B. We propose that TIFAB selectively inhibits TRAF6.

B. HEK293 cells were transfected with pcDNA3.1 (vector, 250 ng), pcDNA3.1-FLAG-TRAF6 (250 ng) and/or pcDNA3.1-FLAG-TIFAB (250 ng) and κ B-luciferase. Following transfection, cells were simulated with TNF α (1 ng/ml) for 6 hours. Values represent relative luciferase.

C. HEK293 cells were transfected with pcDNA3.1 (vector, 250 ng), pcDNA3.1-FLAG-TRAF2 (250 ng) and/or pcDNA3.1-FLAG-TIFAB (250 ng) and κ B-luciferase. Following transfection, cells were simulated with TNF α (1 ng/ml) for 6 hours. Values represent relative luciferase.

D. HEK293 cells were transfected with pcDNA3.1 (vector, 250 ng), pcDNA3.1-FLAG-IKK β (250 ng) and/or pcDNA3.1-FLAG-TIFAB (250 ng) and κ B-luciferase. Following transfection, cells were simulated with TNF α (1 ng/ml) for 6 hours. Values represent relative luciferase.

E. HEK293 cells were transfected with pcDNA3.1 (vector, 250 ng), pcDNA3.1-FLAG-p65/RelA (250 ng) and/or pcDNA3.1-FLAG-TIFAB (250 ng) and κ B-luciferase. Following transfection, cells were simulated with TNF α (1 ng/ml) for 6 hours. Values represent relative luciferase.

Figure 5.

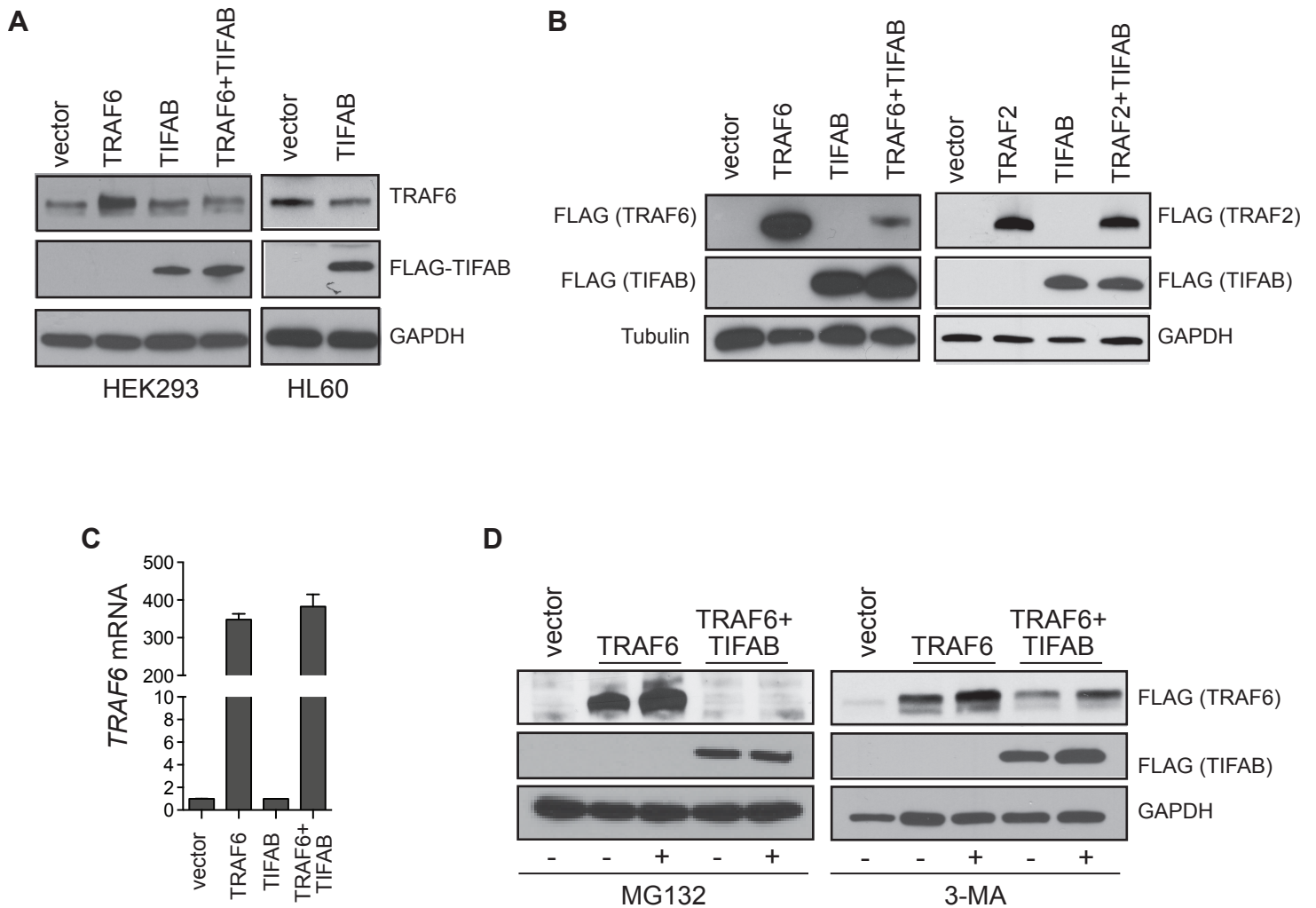


Figure 5: Task 6b

(A) Immunoblot analysis of HEK293 cells transfected with empty vector, TRAF6, or FLAG-TIFAB (left panel), and HL60 cells transduced with empty vector (pMSCV-pGK-GFP) or FLAG-TIFAB (right panel).

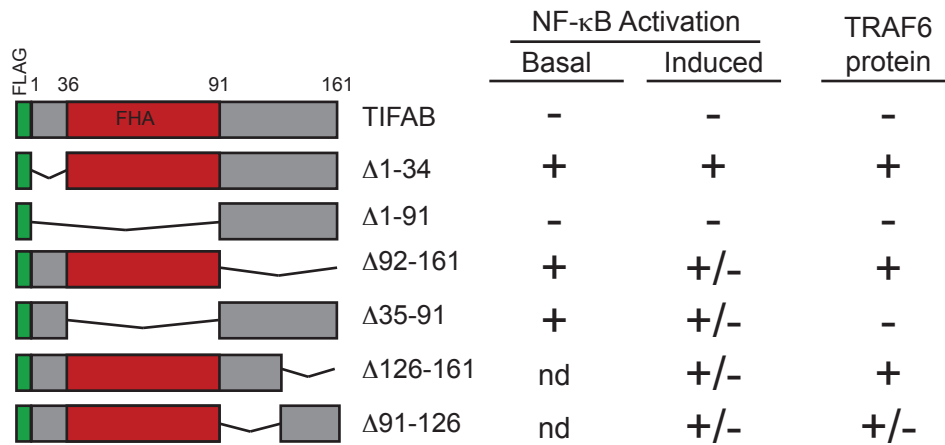
(B) Immunoblot analysis of HEK293 transfected with empty vector, TIFAB, and/or TRAF6 (left panel) or TRAF2 (right panel).

(C) Expression analysis of TRAF6 mRNA by qRT-PCR in HEK293 cells transfected with empty vector, TRAF6, and/or FLAG-TIFAB.

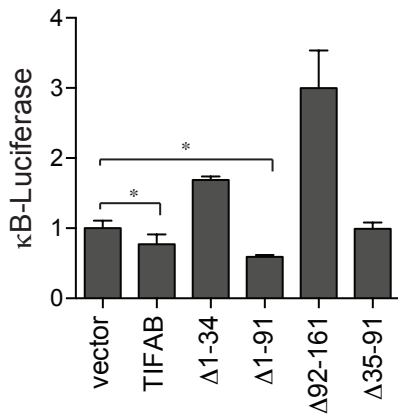
(D) Immunoblot analysis of HEK293 transfected with empty vector, TIFAB, and/or TRAF6 and then treated with a proteasome inhibitor (MG132, 10 μ M for 12 hrs) or a lysosome inhibitor (3-MA, 5 mM for 24 hrs).

Figure 6.

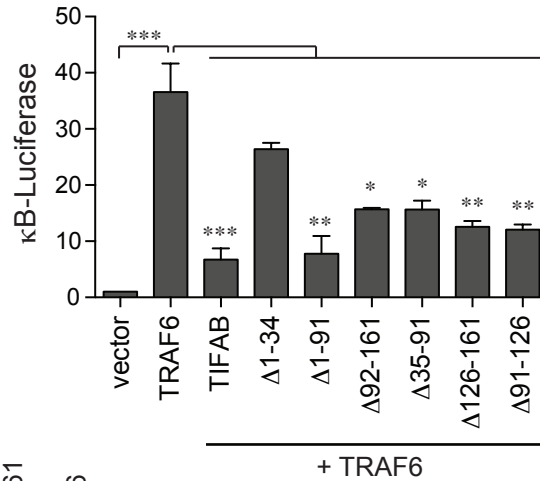
A



B



C



D

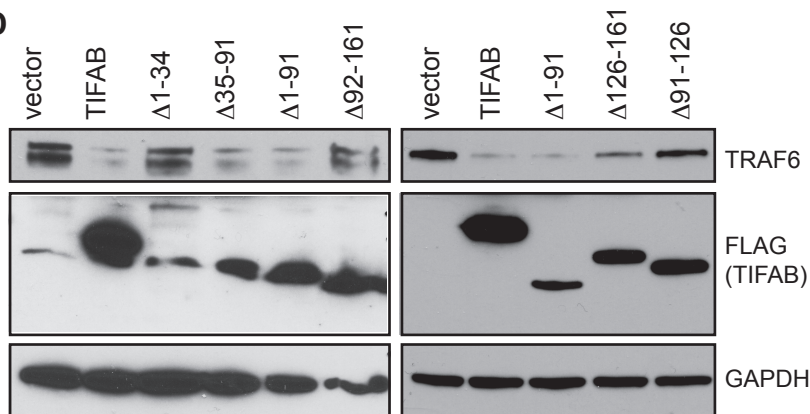


Figure 6: Task 6d

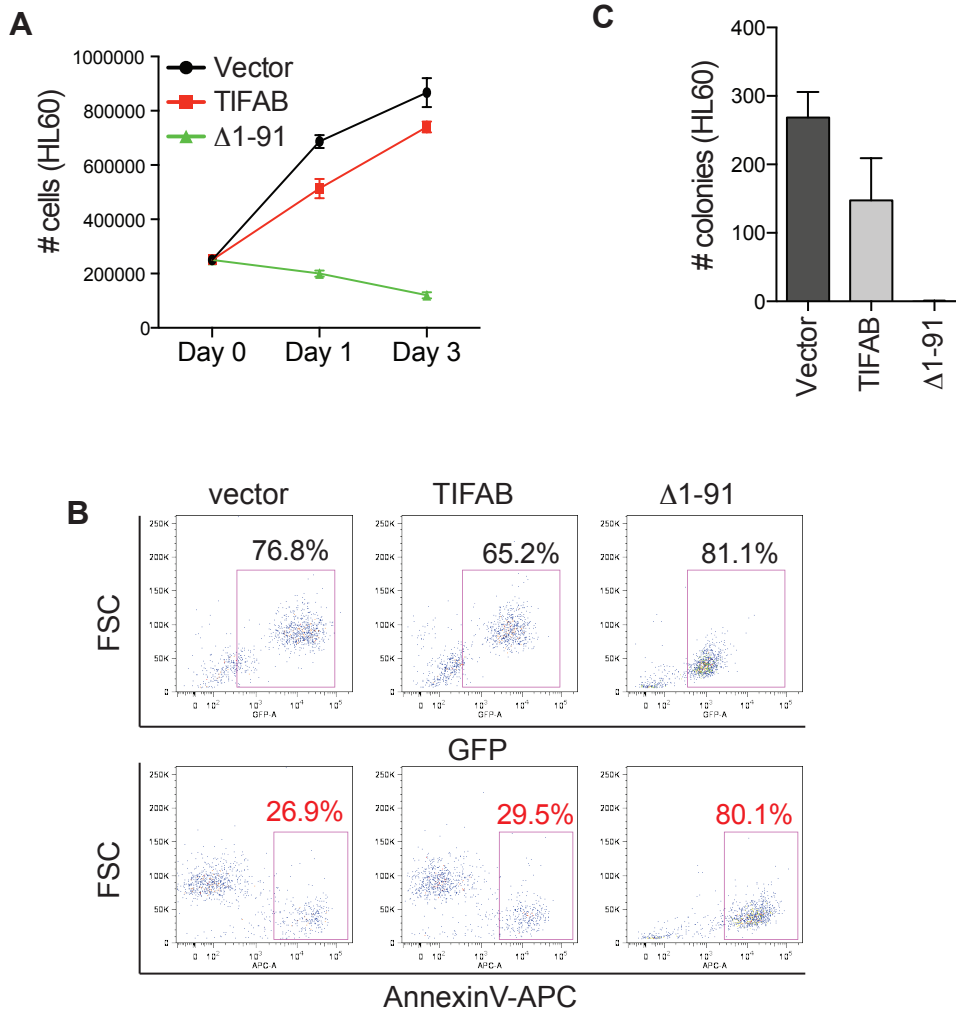
A. Structure of wild-type and TIFAB deletion mutants.

B. HEK293 cells were transfected with pcDNA3.1 (vector, 250 ng), pcDNA3.1-FLAG-TIFAB (250 ng), or the TIFAB deletion mutants, and κB-luciferase. Values represent relative luciferase.

D. HEK293 cells were transfected with pcDNA3.1 (vector, 250 ng), pcDNA3.1-FLAG-TRAF6 (250 ng) and pcDNA3.1-FLAG-TIFAB (250 ng) or TIFAB deletion mutants, and κB-luciferase. Values represent relative luciferase.

D. HEK293 cells were transfected with the indicated vectors and then evaluated by immunoblotting.

Figure 7



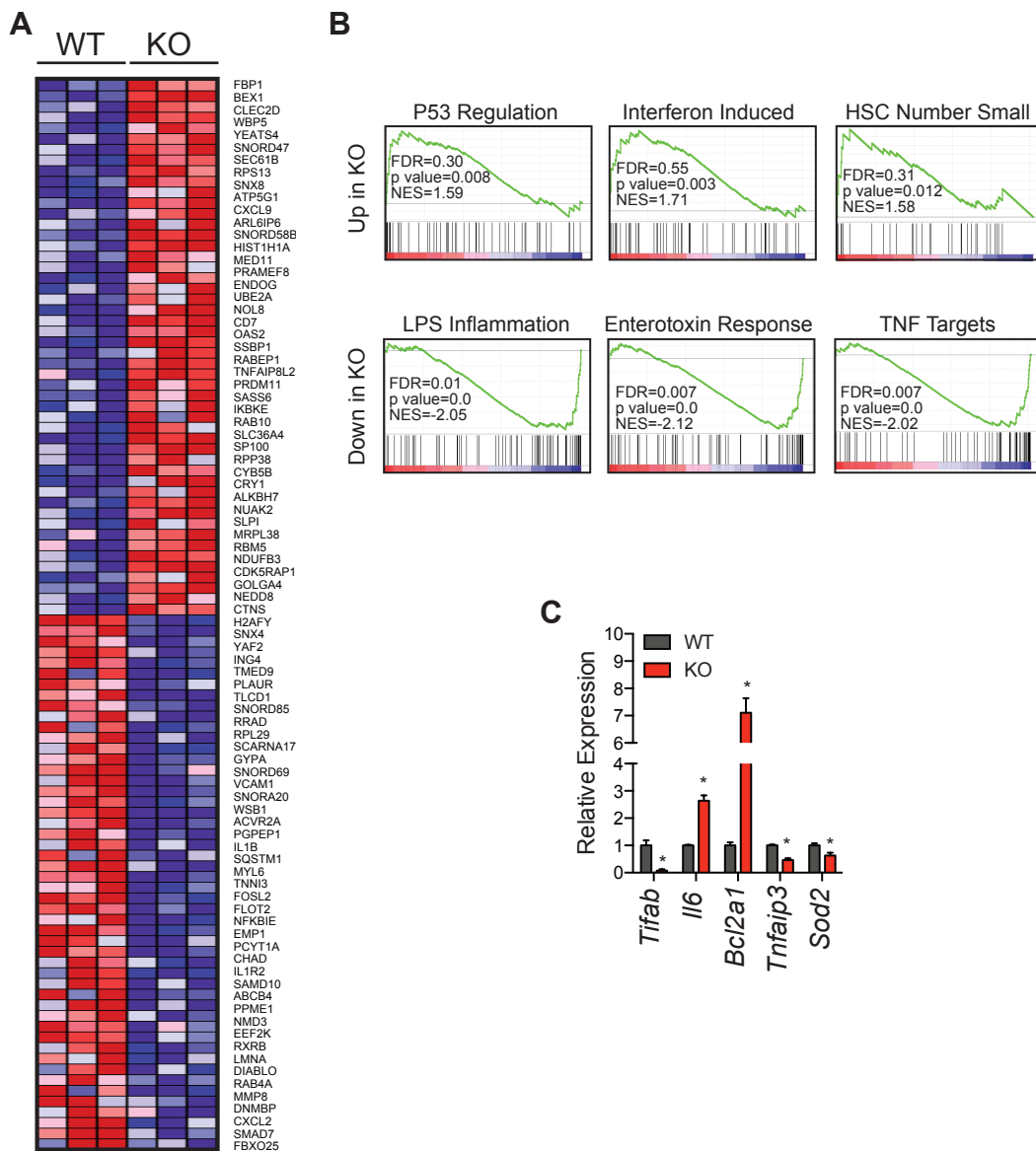
Task 6, Figure 7:

(A) Viable cell growth of HL60 transduced with vector, FLAG-TIFAB, or FLAG-TIFAB Δ 1-91 was assayed by trypan blue exclusion for the indicated number of days in culture.

(B) AnnexinV staining of HL60 cells transduced with vector, FLAG-TIFAB, or FLAG-TIFAB Δ 1-91.

(C) Colony formation of HL60 cells transduced with vector, FLAG-TIFAB, or FLAG-TIFAB Δ 1-91 was measured after 14 days. Error bars are mean \pm SEM values.

Figure 8.



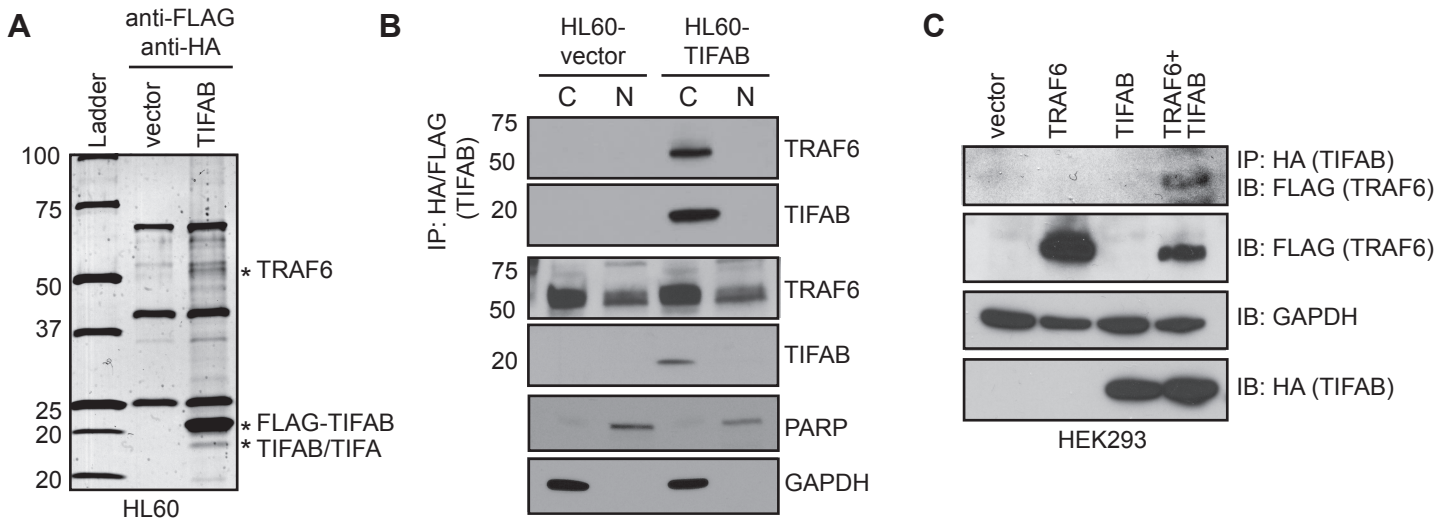
Task 7, Figure 8:

(A) Heat map generated from GSEA showing gene expression differences in LSK isolated from 3 month old mice transplanted with Tifab WT or KO BM cells (n = 3 mice/group)

(B) Gene set enrichment analysis (GSEA) of statistically significant gene sets enriched in the LSK cells of transplanted Tifab^{-/-} cells (up in KO) or WT cells (down in KO).

(C) Validation of gene expression of the indicated genes in BM MNC by qRT-PCR.

Figure 9



Extra Task, Figure 9:

(A) HL60 cells were transduced with vector or FLAG-TIFAB. HL60 cells expressing vector or TIFAB were immunoprecipitated with FLAG-containing beads. FLAG-TIFAB complexes were eluted off the beads and run on a polyacrylamide gel for silver stain analysis. Bands that were unique to the lane containing FLAG-TIFAB (and the control bands) were cut from the gel and evaluated by mass spectrometry (Figure 9A).

(B-C) Co-immunoprecipitation experiments were performed in HL60 and HEK293 cells.

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Personal Statement

Research in my laboratory focuses on the molecular, cellular, and genetic basis of hematologic malignancies, with a specific focus on Myelodysplastic Syndromes (MDS) and Acute Myeloid Leukemia (AML). We have uncovered aberrant activation of the innate immune pathway in MDS, and now find that this activation perturbs normal hematopoietic stem/progenitor (HSPC) function. Therefore, one of the main objectives of my research program is to evaluate the role of the innate immune pathway in normal HSPC and in MDS. Broader initiatives involve characterizing novel candidate genes relevant to MDS, dissecting molecular and cellular alterations in MDS (and related hematologic malignancies), and attempting to identify novel therapeutics.

Education and Training

2005-2010 Postdoctorate
BC Cancer Research Centre/University of British Columbia, Vancouver, Canada
2000-2005 Ph.D., Molecular Biology, Cell Biology and Biochemistry
Boston University, Boston, MA, USA.
1996-2000 B.Sc., Honors Biology
Concentrations in Chemistry and Biotechnology
Fairleigh Dickinson University, Teaneck, NJ, USA.

Research and Professional Experience

2010- present Assistant Professor, Division of Experimental Hematology, Cincinnati Children's Hospital Medical Center, Cincinnati, OH
2010-present Affiliate Assistant Professor, Department of Cancer and Cell Biology, University of Cincinnati, Cincinnati, OH
2010- 2011 Lecturer, Ulm University, Germany
2005-2010 Postdoctoral Fellow
Research Focus: *Identification and functional analysis of genetic and molecular determinants of hematological malignancies: deregulation of miR-146/TRAF6 signaling in Myelodysplastic syndromes*
Advisor: Dr. Aly Karsan, BC Cancer Research Centre/University of British Columbia
2001-2005 Graduate Research Assistant
Dissertation: *A mutational and functional analysis of C-terminal sequences of transcription factor REL: their role in cellular transformation and transcriptional activation*
Advisor: Dr. Thomas D. Gilmore, Boston University
2000 Summer Research Assistant
Research Focus: *Functional analysis of androgen receptor in prostate cancer*
Advisor: Dr. Marianne Sadar, BC Cancer Research Centre
1999-2000 Honors Research
Dissertation: *Inhibitory effects of somatostatin on the viability of a cell line*
Advisor: Dr. Anjali Saxena, Fairleigh Dickinson University
1997-1999 Research Assistant
Research Focus: *Development of enzyme-friendly biosensors for lactate detection*
Advisor: Dr. Mihaela Leonida, Fairleigh Dickinson University

Teaching Experience

- 2014 Individual Lecture
“Exploring Biomedical Research”, University of Cincinnati Honors Program.
- 2013 Individual Lecture
“Effective Oral Scientific Presentations”, Xavier University.
- 2013 Course
“Introduction to Grant Writing”, Department of Cancer and Cell Biology, University of Cincinnati.
Developed and co-taught a course on basic grant writing fundamentals, experience putting them in practice, and responding to critiques. The course is offered to graduate students and implements traditional didactic and web-based components.
- 2013 Lectures
“Critical data presentation III”, Department of Cancer and Cell Biology, University of Cincinnati.
Participated in primary research evaluation and interpretation with graduate students.
- 2012 Individual Lecture
“Effective Oral Scientific Presentations”, Xavier University.
- 2012 Course
“Introduction to Grant Writing”, Department of Cancer and Cell Biology, University of Cincinnati.
Developed and co-taught a course on basic grant writing fundamentals, experience putting them in practice, and responding to critiques. The course is offered to graduate students and implements traditional didactic and web-based components.
- 2012 Lectures
“Critical data presentation III”, Department of Cancer and Cell Biology, University of Cincinnati.
Participated in primary research evaluation and interpretation with graduate students.
- 2010 Lectures
“Innate Immunity and Cancer”, Master Online Program in Advanced Oncology. Ulm University.
Developed a video-based lecture on innate immunity and its role in human cancer for postgraduates of medicine.
- 2010 Individual Lecture
“MicroRNAs in cancer”, Medical Oncology Residency Training Program. BC Cancer Agency.
Organized and conducted a lecture for 1st and 2nd year residents of medical training on microRNAs and their role in clinical oncology.
- 2009 Individual Lecture
“MicroRNAs in hematological malignancies”, Hematology Fellows Series. Department of Medicine, University of British Columbia.
Organized and conducted a lecture for hematology and pathology fellows on the emerging role of microRNAs in hematological malignancies.
- 2009 Workshop
“MicroRNAs: small RNAs with big impact”. Terry Fox Laboratory, BC Cancer Research Centre.
Organized and co-lead a workshop on microRNAs, their biogenesis, diversity, and mechanism as related to gene regulation and to human disease.
- 2005 Teaching Assistant
“Molecular Biology Laboratory”, Boston University.
Prepared and delivered weekly labs for advanced (500-level) molecular biology students.
- 2000-2001 Teaching Fellow
“Life Sciences Chemistry I and II”, Boston University
Organized and delivered weekly labs for introductory (100-level) undergraduate science students to supplement course material. Prepared and evaluated exams and final grades. Held weekly office hours.
- 1998 Teaching Assistant
“Chemistry for Health Science”, Fairleigh Dickinson University.
Organized and delivered weekly labs for introductory (100-level) undergraduate science students.

Awards and Honors

2014	Research Achievement Award, Cincinnati Children's Hospital Medical Center
2013-2016	Gabrielle's Angel Foundation Medical Research Award
2011-14	American Society of Hematology Scholar Award, Basic Research Junior Faculty
2010	National Institute of Allergy and Infectious Disease, Keystone Symposium Scholarship
2008	Travel Award, American Society of Hematology
2008	Eugene Cronkite Award: New Investigator Award (1 st place). International Society of Experimental Hematology
2008	Travel Grant, International Society of Experimental Hematology Scientific Meeting
2007-2010	Postdoctoral Fellowship, Canadian Institute of Health Research
2006	Frank A. Belamarich Award; Outstanding Scholarship and Performance in Graduate Studies, Biology Department, Boston University
2006-2009	Postdoctoral Fellowship, Michael Smith Foundation for Health Research
2001-2005	Graduate Research Fellowship, Boston University
2002-2004	Postgraduate Research Scholarship, Natural Sciences and Engineering Research Council of Canada
2002	Travel Scholarship, NF- κ B: Bench to Bedside - Keystone Symposium
2000-01, 2005	Teaching Fellowships, Boston University
2000	J.M. Warren Summer Research Scholarship, British Columbia Cancer Agency
2000	ECAC Award: Graduating athlete with highest cumulative grade point average, Fairleigh Dickinson University, NJ
2000	Summa Cum Laude, Fairleigh Dickinson University
2000	Phi Omega Epsilon, Honor Society
1999	Phi Zeta Kappa, Honor Society
1999	Charter Day Scholarship, Fairleigh Dickinson University
1998	University-College Dean's Award, Fairleigh Dickinson University
1996-2000	University Honors List, Fairleigh Dickinson University
1996-2000	Tennis Scholarship, Fairleigh Dickinson University

Memberships and Committees

2014	Scholar Awards Study Section Reviewer, American Society of Hematology (Reviewer)
2014	NIH MONC Study Section (Ad hoc reviewer)
2013	Aplastic Anemia & MDS International Foundation/Edward P. Evans Foundation: MDS Basic Research Summit (Invited discussant)
2013	Grant Reviewer, Canadian Cancer Society (Ad hoc)
2013	NIDDK Workshop: Role of ncRNAs in Hematopoiesis Workshop, Bethesda, MD. (Invited talk/discussant)
2013	Medical Scientist Training Program, University of Cincinnati College of Medicine (Training Faculty)
2013	Scholar Awards Study Section Reviewer, American Society of Hematology
2013	Grant Reviewer, Swiss Cancer League (Ad hoc reviewer)
2012	Scholar Awards Study Section Reviewer, American Society of Hematology
2012-present	Early Career Reviewer (ECR) at the Center for Scientific Review, National Institute of Health
2011 (Sept)	NIH State of the Science Symposium: Myelodysplastic Syndrome Working Group, Bethesda MD (Invited member)
2011-present	Graduate student recruitment committee, Cancer and Cell Biology Graduate Program, University of Cincinnati
2011-present	Immunobiology Graduate Program, Cincinnati Children's Hospital (Training Faculty)
2010-present	Cancer and Cell Biology Graduate Program, University of Cincinnati (Training Faculty)
2008	Campus Provost Search Committee, Fairleigh Dickinson University-Vancouver
2008-present	International Society of Experimental Hematology (Member)

2005-present American Society of Hematology (Associate Member)
 2006-2011 Board of Directors, Fairleigh Dickinson University-Vancouver (Member)
 1999-2000 *TriBeta* Biological Society, Fairleigh Dickinson University Chapter (Vice President)
 1997-2000 University Honors Program, Fairleigh Dickinson University

University Service

2014 Straus Fellows Program (Reviewer)
 2014-present CCB Graduate Program (Committee Member)
 2014 Cincinnati Cancer Center, Pilot Grant Program (Reviewer)
 2013 Postdoctoral Recruitment Symposium, CCHMC (Organizing Committee)
 2013 Midwest Blood Club Annual Meeting (Co-Organizer)
 2013 ASBMB Career Symposium (Planning Committee Member)
 2013-14 Office of Postdoctoral Affairs, CCHMC (Assistant Director)
 2012 Qualifying Exam Committee, Graduate Program in Cancer and Cell Biology, University of Cincinnati (Standing Member)
 2011 (Sept) CCB Graduate Program Student Symposium, University of Cincinnati (Faculty co-organizer)
 2011 (Feb) Research Ethics, University of Cincinnati (Discussion leader)
 2011-14 Cancer and Blood Diseases Institute Seminar Series (Co-coordinator)
 2011 Scholarship Oversight Committee, Hematology/Oncology Clinical Fellowship (Committee Member)
 2011 Immunohematology Club Seminar Series (Speaker)

Patents

Daniel Starczynowski and Garrett Rhyasen, "Combination therapy for MDS", U.S patent, pending (2014)

Aly Karsan and Daniel Starczynowski, "TRAF6 as a therapeutic target and predictive biomarker for lung and colorectal cancer," U.S. provisional patent (2010)

Aly Karsan and Daniel Starczynowski, "Novel methods to predict therapeutic response to Lenalidomide and related drugs", U.S. provisional patent (2008)

Peer-Reviewed Publications

1. Gilmore TD, M-E Gapuzan, D Kalaitzidis, and **D Starczynowski**. (2002) Rel/NF-kB/IkB signal transduction in the generation and treatment of human cancer. Cancer Letters, 181: 1-9. PMID:12430173. (Review)
 *Top 25 Hottest Article" on ScienceDirect within Cancer Letters (April-June 2005)
2. Leonida MD, **DT Starczynowski**, R Waldman, and B Aurian-Blajeni. (2003) Polymeric FAD used as enzyme-friendly mediator in lactate detection. Analytical and Bioanalytical Chemistry, 376: 832-837. PMID: 12811450.
3. **Starczynowski DT**, JG Reynolds, and TD Gilmore. (2003). Deletion of either C-terminal transactivation subdomain enhances the *in vitro* transforming activity of human transcription factor REL. Oncogene, 22: 6929-6936. PMID: 14534540.
4. Gilmore TD, D Kalaitzidis, M-C Liang, and **DT Starczynowski**. (2004). The c-Rel transcription factor and B-cell proliferation: a deal with the devil. Oncogene, 23: 2275-2286. PMID: 14755244. (Review)
5. Gilmore TD, D Kalaitzidis, and **DT Starczynowski**. (2004). *RE*levant gene amplification in B-cell lymphomas. Blood. 103: 3243. PMID: 15070712. (Letter)

6. Kalaitzidis D, J Ok, L Sulak, **DT Starczynowski**, and TD Gilmore. (2004). Characterization of a human REL-estrogen receptor fusion protein with a reverse conditional transforming activity in chicken spleen cells. Oncogene, 23: 7580-7587. PMID:15326488.
7. **Starczynowski DT**, JG Reynolds, and TD Gilmore. (2005). Mutations of tumor necrosis factor α -responsive serine residues 460 and 471 within the C-terminal transactivation domain of human transcription factor REL can enhance its *in vitro* transforming ability. Oncogene, 24: 7355-7368. PMID: 16027730.
8. **Starczynowski DT**, H Trautmann, C Pott, L Harder, N Arnold, R Siebert, and TD Gilmore. (2007). Mutation of an IKK phosphorylation site within the transactivation domain of REL in two patients with B-cell lymphoma enhances REL's *in vitro* transforming activity. Oncogene, 26: 2685-2694. PMID: 17072339.
*Oncogene's Featured Article (April 2007)
9. **Starczynowski DT**, S Vercauteren, S Sung, A Brooks-Wilson, J Spinelli, C Eaves, A Eaves, D Horsman, W Lam, and A Karsan. (2008). High-resolution array comparative genomic hybridization of CD34⁺ cells from patients with low-risk myelodysplastic syndromes predicts overall and leukemia-free survival. Blood, 112(8): 3412-3424. PMID: 18663149.
10. **Starczynowski DT**, F Kuchenbauer, B Argiropoulos, S Sung, R Morin, A Muranyi, D Hogue, R Wells, M Marra, WL Lam, K Humphries, and A Karsan. (2010). Identification of miR-145 and miR-146a as microRNAs involved in the pathogenesis of 5q- syndrome. Nature Medicine, 16(1): 49-58. PMID: 19898489.
*Comment in Nature Medicine. 2010. Myelodysplasia: Battle in the Bone Marrow. 16(1): 30-32.
*Featured in The Hematologist. 2010. Clarifying the genetic underpinnings of the 5q- syndrome. July/Aug:7(4).
11. **Starczynowski DT** and A Karsan. (2010). Deregulation of innate immune signaling in myelodysplastic syndromes is associated with deletion of chromosome arm 5q. Cell Cycle, 11; 9(5). PMID: 20160505. (Review)
12. **Starczynowski DT** and A Karsan. (2010). Innate immune signaling in Myelodysplastic Syndromes. Hematology/Oncology Clinics of North America, 24(2):343-359. PMID: 20359630. (Review)
13. Vercauteren SM, S Sung, **DT Starczynowski**, WL Lam, H Bruyere, DE Horsman, P Tsang, H Leitch, and A Karsan. (2010). Array comparative genomic hybridization of peripheral blood granulocytes of patients with myelodysplastic syndrome detects karyotypic abnormalities. American Journal of Clinical Pathology, 134(1):119-126. PMID: 20551276.
14. **Starczynowski DT**, S Vercauteren, S Sung, A Brooks-Wilson, J Spinelli, C Eaves, A Eaves, D Horsman, W Lam, and A Karsan. (2011). Copy number alterations at polymorphic loci may be acquired somatically in patients with myelodysplastic syndromes. Leukemia Research, 35(4): 444-447. PMID: 20801506.
15. **Starczynowski DT**, RD Morin, A McPherson, J Lam, R Chari, J Wegrzyn, A Delaney, AL Prabhu, Y Zhao, M Hirst, W Lam, MA Marra, and A Karsan. (2011). Genome-wide identification of human microRNAs located in leukemia-associated genomic alterations. Blood, 117(2): 595-607. PMID: 20962326.
16. **Starczynowski DT**, F Kuchenbauer, J Wegrzyn, K Humphries, and A Karsan. (2011). MicroRNA-146a disrupts hematopoietic differentiation and survival. Experimental Hematology, 39(2): 167-178. PMID: 20933052.
17. Kuchenbauer F, SM Mah, M Heuser, A McPherson, J Ruschmann, A Rouhi, T Berg, L Bullinger, B Argiropoulos, RD Morin, D. Lai, **DT Starczynowski**, A Karsan, CJ Eaves, A Watahiki, Y Wang, SA Aparicio, A Ganser, J Krauter, H Doehner, K Doehner, MA Marra, FD Carmargo L Palmquist, C Buske, and RK Humphries. (2011). Comprehensive analysis of mammalian miRNA* species and their role in myeloid cells. Blood, 118(12) 3350-3358. PMID: 21628414.

18. Rhyasen G and **DT Starczynowski***. (2011). Deregulation of microRNAs in Myelodysplastic Syndromes. Leukemia, 26(1): 13-22. PMID: 21852786.
19. **Starczynowski DT**, WL Lockwood, S Delehouzee, R Chari, J Wegrzyn, M Fuller, MS Tsao, S Lam, AF Gazdar, WL Lam, and A Karsan. TRAF6 is an amplified oncogene bridging the Ras and nuclear factor- κ B cascade in lung cancer. (2011). Journal of Clinical Investigation, 121(10):4095-4105. PMID: 21911935.
20. Vercauteren S, **DT Starczynowski**, S Sung, K McNeil, C Salski, C-L Jensen, W Lam, A Karsan. (2012). T cells of patients with Myelodysplastic syndrome are frequently derived from the malignant clone. British Journal of Haematology, 156(3): 409-412.
21. Fang J, M Varney, and **DT Starczynowski***. Implication of miRNAs in the pathogenesis of MDS. (2012) Current Pharmaceutical Design, 18(22): 3170-3779. PMID: 22685174
22. Fang J, G Rhyasen, L Bolanos, C Rasch, M Varney, M Wunderlich, S Goyama, G Jansen, J Cloos, C Rigolino, A Cortelezzi, JC Mulloy, EN Oliva, M Cuzzola, **DT Starczynowski***. (2012). Cytotoxic effects of Bortezomib in MDS/AML depend on autophagy-mediated lysosomal degradation of TRAF6 and repression of PSMA1. Blood. 120(4): 858-867. PMID: 22685174.
23. Guo F, S Zhang, M Grogg, JA Cancelas, ME Varney, **DT Starczynowski**, W Du, J Yang, W Liu, G Thomas, S Kozma, Q Pang, Yi Zheng. Mouse gene targeting reveals an essential role of mTOR in hematopoietic stem cell engraftment and hematopoiesis. (2013). Haematologica. 98(9): 1353-1358. PMID: 23716557.
24. Rhyasen G, L Bolanos, J Fang, C Rasch, A Jerez, M Varney, M Wunderlich, C Rigolino, L Mathews, M Ferrer, N Southall, R Guha, J Keller, C Thomas, L Beverly, A Cortelezzi, EN Oliva, M Cuzzola, J Maciejewski, JC Mulloy, **DT Starczynowski***. Targeting IRAK1 as a novel therapeutic approach for Myelodysplastic Syndrome. (2013). Cancer Cell, 24(1): 90-104. PMID: 23845443.
 * Comment in Cancer Discovery (2013)
 * Comment in Nature Reviews Drug Discovery (2013)
25. Rhyasen G, L Bolanos, and **DT Starczynowski***. Differential IRAK signaling in hematologic malignancies. (2013). Experimental Hematology, 41(12): 1005-1007. PMID: 24084080.
26. Wilson IM, EA Vucic, KS Enfield, KL Thu, YA Zhang, R Chari, WW Lockwood, N Radulovich, **DT Starczynowski**, JP Banáth, M Zhang, A Pusic, M Fuller, KM Lonergan, D Rowbotham, J Yee, JC English, TP Buys, SA Selamat, IA Laird-Offringa, P Liu, M Anderson, M You, MS Tsao, CJ Brown, KL Bennewith, CE Macaulay, A Karsan, AF Gazdar, S Lam, WL Lam. EYA4 is inactivated biallelically at a high frequency in sporadic lung cancer and is associated with familiar lung cancer risk. (2013). Oncogene. 4;36(36): 4464-4473. PMID: 24096489.
27. Rhyasen G, M Wunderlich, K Tohyama, G Garcia-Manero, JC Mulloy, **DT Starczynowski***. An MDS xenograft model utilizing a patient-derived cell line. (2014). Leukemia. 28(5): 1142-1145. PMID: 24326684.
28. Beverly LJ, and **DT Starczynowski***. IRAK1: oncotarget in MDS and AML. (2014). OncoTarget. 5(7): 1699-1700. PMID: 2488061.
29. **Starczynowski DT***. Errant innate immune signaling in del(5q) MDS. (2014). Blood. 124(5): 669-671. PMID: 25082861.
30. Zhao JL, and **DT Starczynowski***. Role of microRNA-146a in hematopoietic stem cell function and myelodysplasia. (2014). Frontiers Genetics. 5;219. PMID: 25071842.
31. Rhyasen G, and **DT Starczynowski***. IRAK signaling in cancer. (2014). British Journal of Cancer. In press.

PMID: 25290089.

32. Fang J, B Barker, L Bolanos, X Liu, A Jerez, H Makishima, S Christie, X Chen, DS Rao, HL Grimes, K Komurov, MT Weirauch, J Cancelas, JP Maciejewski, **DT Starczynowski***. Myeloid malignancies with chromosome 5q deletions acquire a dependency on an intrachromosomal NF- κ B gene network. (2014). Cell Reports. 11;8(5): 1328-1338. PMID: 25199827.
33. Lai CK, B Moon, F Kuchenbauer, **DT Starczynowski**, B Argiropoulos, E Yung, P Beer, A Schwarzer, A Sharma, G Park, M Leung, G Lin, S Vollett, S Fung, CJ Eaves, A Karsan, AP Weng, RK Humphries, M Heuser. Cell fate decisions in malignant hematopoiesis: leukemia phenotype is determined by distinct functional domains of the MN1 oncogene. (2014). PLoS One. 17; 9(11): epub. PMID: 25401736.
34. Chang KH, A Sengupta, RC Nayek, A Duran, SJ Lee, RG Pratt, AM Wellendorf, SE Hill, M Watkins, D Gonzalez-Nieto, BJ Aronow, **DT Starczynowski**, R Civitelli, MT Diaz-Meco, J Moscat, JA Cancelas. p62 is required to retain short-term repopulating and myeloid progenitor cells through inhibition of IKK/NF- κ B/Ccl4 signaling at the bone marrow macrophage-osteoblast niche. (2014). Cell Reports. Accepted.
35. Fang J, L Bolanos, KM Choi, X Liu, S Christie, ME. Figueroa, S Akunuru, R Kumar, KD Greis, P Stoilov, MD Filippi, JP Maciejewski, N Salamonis, G Garcia-Manero, H Geiger, Y Zheng, **DT Starczynowski***. Ubiquitin editing of a spliceosome auxiliary protein by TRAF6 induces hematopoietic stem cell defects and Myelodysplasia. Submitted.
36. Varney ME, M Niederkorn, K Konno, T Matsumura, J Gohda, N Yoshida, T Akiyama, S Christie, J Fang, D Miller, A Jerez, A Karsan, JP Maciejewski, J-I Inoue, **DT Starczynowski***. *Tifab* is required for restricting innate immune signaling and hematopoietic stem/progenitor function in del(5q) myelodysplasia. Submitted.

*Corresponding author

Current Funding

1. National Heart, Lung, and Blood Institute
Type: RO1 (RO1HL111103)
Role: Principal Investigator
Title: Identification and characterization of genes in del(5q) myelodysplastic syndrome
Period: 12/1/11 – 12/1/16
\$1,250,000
2. Gabrielle's Angel Foundation Medical Research Award
Type: Faculty Scholar Award
Role: Principal Investigator
Period: 1/13-1/16
Title: Defining the role and therapeutic potential of TNF receptor-associated factor 6 in MDS
\$225,000
3. Celgene Corporation
Type: Sponsored research agreement
Role: Principal Investigator
Period: 4/13-
\$ 34,000
5. Cincinnati Children's Basic Science Research and the Clinical Translational, Outcomes and Health Services Redesign award
Type: Pilot grant
Role: Principal Investigator
Title: Immune signaling and altered spliceosome function in MDS.
Period: 12/13 – 12/14
\$75,000

6. National Heart, Lung, and Blood Institute
 Type: RO1 (RO1HL114582)
 Role: Principal Investigator
 Title: Role of TRAF6 in Myelodysplastic Syndromes
 Period: 7/14-6/18
 \$1,250,000
7. Edwards P. Evans Foundation
 Type: Research Grant
 Role: Principal Investigator
 Title: Chronic innate immune signaling in the pathogenesis of MDS
 Period: 7/14-7/16
 \$400,000
8. National Institute of Diabetes and Digestive and Kidney Diseases
 Type: RO1 (RO1DK102759)
 Role: Principal Investigator
 Title: Molecular Pathogenesis of MDS
 Period: 9/14-9/17
 \$450,000

Completed Funding

1. Trustee Grant Award, Cincinnati Children's Hospital Medical Center
 Type: Junior Faculty Award
 Role: Principal Investigator
 Period: 1/11-12/13
 Title: Deregulation of TIFAB in myelodysplastic syndromes
 \$120,000
2. BSRC/CTOHSC Research Innovation/Pilot Grant Cincinnati Children's Hospital Medical Center
 Type: Pilot grant
 Role: Principal Investigator
 Title: Mechanism and treatment of Lenalidomide Resistance
 Period: 8/12 – 8/13
 \$66,000
3. American Society of Hematology (ASH) Scholar Awards Program
 Type: Basic Junior Faculty
 Role: Principal Investigator
 Title: Deregulation of TIFAB in myelodysplastic syndromes
 Period: 7/1/11 – 7/1/14
 \$150,000
4. Department of Defense (DOD) Bone Marrow Failure Research Program
 Type: New Investigator Award
 Role: Principal Investigator
 Period: 6/1/11 – 6/1/14
 Title: Regulation and function of TIFAB in myelodysplastic syndrome
 \$380,000 total

Invited Talks

Myeloid Workshop, American Society of Hematology, San Francisco, CA. (Dec, 2014). Invited talk.

Gabrielle's Angel Foundation for Cancer Research Annual Medical Symposium, New York City, NY (October, 2014). Invited talk

Cincinnati Children's Hospital Medical Center, Annual Immunology Retreat, Cincinnati, OH (October, 2014). Invited talk.

Sanford Burnham Medical Research Institute, San Diego, CA (September, 2014). Seminar Series/Invited talk.

Northwestern University, Chicago, IL (September, 2014). Seminar Series/Invited talk.

Edward P. Evans Foundation MDS Research Summit, Philadelphia, PA (September, 2014). Seminar/Invited talk.

MD Anderson Cancer Center, Houston, TX. (May, 2014). Grand Rounds.

Myeloid Workshop, Cincinnati, OH. (May, 2014). Invited talk

Indiana University, Indianapolis, IN. (April, 2014). Seminar Series/Invited talk.

University of West Virginia. (March, 2014). Seminar Series/Invited talk.

City of Hope Cancer Research Center. (March 2014). Seminar Series/Invited talk.

University of Cincinnati. (September, 2013). Grand Rounds/Invited talk.

FASEB Science Research Conference: Hematologic Malignancies, Saxton River Falls, VT. (July, 2013). Invited talk.

UCLA, Stem Cell Program. Los Angeles, CA. (April, 2013). Seminar series/Invited talk.

Myeloid Workshop, American Society of Hematology, Atlanta, GA. (Dec, 2012). Invited talk.

James Brown Cancer Center, University of Louisville, Louisville, KY. (Oct, 2012). Seminar series/Invited talk.
Hematologic Malignancies, Houston, TX (October, 2012). Invited talk.

Midwest Blood Club Symposium, Indianapolis, IN (March, 2012). Oral Abstract.

American Society of Hematology, San Diego, CA (December, 2011): New Therapies in Myelodysplastic Syndromes. Oral Abstract Session.

MD Anderson Cancer Center, Houston, TX. Leukemia Group (November, 2011)

Japanese Society of Hematology, Nagoya, Japan (October, 2011). Plenary Session.

Baltic Stem Cell Meeting, Szczecin, Poland (May, 2011). Plenary Session.

Midwest Blood Club Symposium, Cincinnati, OH (April, 2011). Plenary Session.

Cepheid Inc., Sunnyvale, CA (March, 2011).

Taussig Cancer Institute, Cleveland Clinic, Cleveland, OH (March, 2011). Translational Hematology and Oncology Research Lecture Series.

Ulm University, Ulm, Germany (February, 2011). Workshop on Translational Research: Cellular and Molecular Biology of Cancer.

James Graham Brown Cancer Center, University of Louisville, Louisville, KY (February, 2011).

Keystone Symposia, NF- κ B in Inflammation and Disease, Santa Fe, NM (January 2010). Workshop: NF- κ B in Disease Pathogenesis.

Department of Cancer and Cell Biology, University of Cincinnati, OH (November, 2009). Department Seminar Series.

Department of Cytogenetics, Vancouver Hospital and Health Sciences Centre, Vancouver General Hospital, Vancouver, BC (March 2009). Cytogenetics Department Seminar Series.

Center for Advanced Biotechnology and Medicine, Rutgers University, New Brunswick, NJ (February 2009). Seminar Series.

Centre for Blood Research, University of British Columbia, Vancouver, BC (February 2009). Centre for Blood Research Seminar Series.

American Society of Hematology, San Francisco, CA (December 2008). Myelodysplastic Syndromes: Basic Biology: Ribosomal Proteins, MicroRNAs and Animal Models. Oral Abstract Session.

Stem Cell Network, Annual General Meeting. Vancouver, BC (November 2008). Plenary Session.

BC Cancer Agency Annual Conference, Vancouver, BC (November 2008). Knowledge Translation - Basic vs. Clinical Research: Hematological Malignancies.

International Society of Experimental Hematology. Boston, MA. (July 2008). New Investigator Session.

Editorial Board

2012- Leukemia

2012- PLoS One

Ad Hoc Reviewer

PLoS One, Genes and Cancer, Current Pharmaceutical Review, Experimental Hematology, Leukemia, Blood, International Journal of Cancer, Haematologica, Journal of Hematology & Oncology, Annals of Hematology, Leukemia & Lymphoma, Nature Communication, Epigenetics.

Current Trainees

1. Jing Fang - Postdoctoral fellow; Cincinnati Children's Hospital Medical Center (March, 2011 – current)
2. Melinda Varney – Postdoctoral fellow; Cincinnati Children's Hospital Medical Center (February, 2011 – current)
3. Lyndsey Bolanos – lab manager/technician: Cincinnati Children's Hospital Medical Center (February, 2011 – current)
4. Xiaona Liu – technician: Cincinnati Children's Hospital Medical Center (July, 2013 – current)
5. Madeline Niederkorn – PhD student; Department of Cancer Biology: University of Cincinnati (June, 2013 – current)
6. Molly Smith - PhD student; Department of Cancer Biology: University of Cincinnati

(June, 2014 – current)

7. Katelyn Melgar – MD/PhD student; Department of Immunobiology; Cincinnati Children’s Medical Center
(Aug, 2014 – current)

Past Trainees

1. Garrett Rhyasen – PhD student; Department of Cancer and Cell Biology, University of Cincinnati
(June, 2011 – July, 2014). Postdoctoral Fellow at AstraZeneca.
2. David Miller (BSc candidate): Undergraduate; Xavier University
(June, 2012 – December, 2012). Medical student at Indiana University Medical School.
3. Brenden Barker – technician: Cincinnati Children’s Hospital Medical Center
(July, 2012 – July, 2013). Medical student at Indiana University Medical School.