

AD_____

Award Number: **W81XWH-12-1-0511**

TITLE: **Large Extremity Peripheral Nerve Repair**

PRINCIPAL INVESTIGATOR: **Robert W. Redmond Ph.D.**

CONTRACTING ORGANIZATION: **Massachusetts General Hospital
Boston, MA 02114**

REPORT DATE: October **2014**

TYPE OF REPORT: **Annual Report**

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

REPORT DOCUMENTATION PAGE			<i>Form Approved</i> <i>OMB No. 0704-0188</i>		
Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.					
1. REPORT DATE October 2014		2. REPORT TYPE Annual		3. DATES COVERED 30 Sep 2013 - 29 Sep 2014	
4. TITLE AND SUBTITLE Large Extremity Peripheral Nerve Repair			5a. CONTRACT NUMBER		
			5a. GRANT NUMBER W81XWH-12-1-0511		
			5c. PROGRAM ELEMENT NUMBER		
6. AUTHOR(S) Robert W. Redmond Ph.D. Jonathan M. Winograd M.D. CDR Mark E. Fleming M.D. MC, USN E-Mail: redmond@helix.mgh.harvard.edu			5d. PROJECT NUMBER		
			5e. TASK NUMBER		
			5f. WORK UNIT NUMBER		
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) The Massachusetts General Hospital, 55 Fruit Street, Boston, MA 02114. and Walter Reed National Military Medical Center 8901 Wisconsin Avenue, Bethesda, MD 20814			8. PERFORMING ORGANIZATION REPORT NUMBER		
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012			10. SPONSOR/MONITOR'S ACRONYM(S)		
			11. SPONSOR/MONITOR'S REPORT NUMBER(S)		
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT In current war trauma, 20-30% of all extremity injuries and >80% of penetrating injuries being associated with peripheral nerve damage, typically involve large segmental nerve deficits. Standard repair uses autologous nerve graft, secured by suture. Outcomes are unsatisfactory, affecting quality of life and return to active duty. We are investigating a sutureless, light-activated technology for sealing nerve grafts to produce an immediate water-tight seal that protects and optimizes the regenerating nerve environment. Our studies have shown that biocompatible chemical crosslinking of thin amnion and SIS sections considerably strengthens the materials and protects them from rapid biodegradation in vivo that would compromise their function as nerve wrap sealants during the regeneration process. Outcomes of rodent studies of segmental nerve deficit repair using isograft show the best performing wrap/ fixation method to be sutureless photochemical tissue bonding with the crosslinked amnion wrap. Often, autograft is unavailable due to extensive tissue damage and amputation. Importantly, We have shown nerve regeneration using our approach with an acellular nerve allograft to be equivalent to standard autograft repair in rodent models. An ongoing large animal validation study will pave the way for human studies of this technology.					
15. SUBJECT TERMS nothing listed					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON
a. REPORT	b. ABSTRACT	c. THIS PAGE			19b. TELEPHONE NUMBER (include area code)
U	U	U	UU	112	USAMRMC

Table of Contents

	<u>Page</u>
Introduction.....	4
Keywords	4
Overall Project Summary.....	4-13
Key Research Accomplishments.....	13
Reportable Outcomes.....	14-15
Conclusion.....	15
Supporting Data.....	16-37
Appendices.....	38-112

Introduction.

The goal of the research performed in this project is to develop a new technology for repair of peripheral nerve injuries involving significant neural deficit with improved functional outcomes for the wounded warrior. The research addresses drawbacks of current methods of suture attachment of nerve grafts and involves development of both a sutureless fixation method to place the nerve graft and an optimal wrap material to seal the endoneurial environment for regeneration. Reduction in needle trauma, reduced inflammation and scarring and sealing the endoneurial environment should all contribute to improved clinical outcomes.

Keywords: Nerve injury, nerve gap, nerve wrap, PTB, photosealing, Rose Bengal, amnion, nerve conduit, crosslinking, allograft, photochemistry.

Overall Project Summary:

Task 1– Determine mechanical properties, seal strength and resistance to biodegradation of candidate photochemical nerve wrap biomaterials. (Months 1-10)

Task 1a. Regulatory approval of use of human tissue by Partners (MGH) IRB and review and approval by USAMRMC Office of Research Protections (human amniotic membrane, HAM). (Months 1-4, MGH: Winograd/Redmond)

Regulatory approval for the use of discarded human tissue (Amniotic membrane) was obtained from both the MGH Institutional Review Board and the USAMRMC Office of Research Protections in August 2012.

Task 1b. Regulatory approval of rodent sciatic nerve for nerve wrap bond measurements by MGH IACUC and review and approval by USAMRMC Office of Research Protections (ACURO). (Months 1-4, MGH: Redmond)

Approvals for the rodent protocols to be used in Task 2 were obtained from the MGH IACUC (protocol #2012N000117) and ACURO approval on 11/19/2012.

Task 1c. Mechanical testing of AxoGuard→ nerve protector (Months 2-4, MGH: Redmond)

The AxoGuard nerve protector proved to be too thick for facile use in photochemical tissue bonding experiments in the rodent model. It was not possible to wrap this material around the small caliber rat sciatic nerve without undue mechanical tension on the wrap that tended to disrupt the contact between nerve and wrap. This required a search for a different source of commercial nerve wrap material, described below in Task 1i.

Task 1d. Processing of HAM and crosslinking with EDC to make xHAM. (Months 4-6, MGH: Redmond)

Task 1e. Mechanical testing (ultimate stress and Young's Modulus) of HAM and xHAM. (Months 4-6, MGH: Redmond)

We have completed processing of human amniotic membrane (HAM) and chemical crosslinking with EDC/NHS to make the crosslinked HAM that should resist biodegradation in vivo. A chemical crosslinking system (EDC (1-ethyl-3-(3-dimethylaminopropyl) carbodiimide), a water soluble agent used with N-hydroxysuccinimide (NHS) for coupling carboxyl groups with primary amines to form amide bonds in proteins) was used at different concentrations under conditions of one hour incubation at room temperature and the resultant mechanical properties measured using a microtensiometer. Figures 1A and 1B show the effect of chemical crosslinker on the maximum stress and Young's modulus

(stiffness) of HAM, measured using a microtensiometer. As expected, crosslinking imparts a greater strength and stiffness to the HAM, especially at the higher concentrations used.

Task 1f. Determine resistance of nerve wraps to collagenase digestion. (Months 4-6, MGH: Redmond).

Biodegradation of HAM as a function of EDC/NHS treatment was determined in the presence of 0.1% collagenase, a high concentration used in our laboratory for extraction of chondrocytes from cartilage. Two assays were used (a) time to complete dissolution and (b) rate of release of amine containing amino acids using the fluorescamine assay.

As can be seen in Figure 2, crosslinking of HAM with EDC/NHS has a large effect on the ability of HAM to resist biodegradation. Figure 2A shows that the uncrosslinked HAM is dissolved in the first hour of treatment whereas all treated samples remain intact even up to 24 h. A more detailed approach using the fluorescamine assay (Figure 2B) to detect amino-acid residues released on degradation shows that increasing EDC/NHS reduces rate and extent of degradation measured in this fashion. This is a highly positive result as a major limiting factor for use in nerve repair would be rapid degradation of the HAM wrap in vivo and this treatment affords considerable protection.

Task 1g. Rat sciatic nerve harvest from 20 Lewis rats. (Months 6-8, MGH: Randolph/Winograd)

Task 1h. Measure bonding strengths of wraps to ex vivo sciatic nerve (months 6-8, MGH: Redmond)

One of the strategies for sutureless graft fixation in this project involves photochemical bonding of a nerve wrap at the graft/nerve stump junction. Studies above show that chemical crosslinking with EDC/NHS strengthens the wrap material and increases its resistance to biodegradation. It is however, important to evaluate whether this chemical crosslinking could interfere with the ability to photochemically bond the wrap material around the epineurium. Thus, rat sciatic nerves were harvested from donor rats immediately post-euthanasia (Task 1g) and bonding of the wrap around the nerve ends performed following application of 0.1% Rose Bengal dye in saline to wrap and epineurium with illumination at 532 nm. The HAM wrap/nerve sample was then mounted in a microtensiometer, as shown in Figure 3 and the tensile load increased until bond failure.

The bond strength of the EDC/NHS treated HAM remains unchanged until the highest tested concentration of 8mM/2mM (EDC/NHS), when a statistically significant decrease is observed with respect to control ($p < 0.05$). At this higher concentration the xHAM becomes brittle and more difficult to handle. Figure 4 shows the data for bond strength as a function of treatment parameters.

In Task 1h we focused on determining the failure strength of the bond formed between ex vivo nerve segments as a function of the fixation procedure, in preparation for the corresponding rat experiments in Task 2. Figure 5 shows the results obtained with the three fixation methods under study (a) epineurial suture, (b) fibrin glue and (c) photochemical tissue bonding (PTB) with a wrap material. All methods induced bonding between the nerve segments with bond strength in the order of suture > PTB > fibrin glue. Conventional epineurial suturing using six 10.0 nylon sutures resulted in the strongest bond. This bond was significantly greater than any of the bonds created by PTB ($p < 0.05$). The strength of the bonds created by PTB were not significantly different from those created following 4-suture epineurial repair. The caliber of the rat sciatic nerve is comparable to a human digital nerve. The use of four epineurial sutures in this situation is clinically realistic and is therefore supportive of the bond strength imparted by PTB. Little difference was seen between uncrosslinked HAM or crosslinked xHAM, except at the highest crosslinker concentration used. In the rat repair model in Aim 2 we have chosen to use the crosslinked HAM due to its increased resistance to enzymatic degradation. Although bond strength is not really an issue in nerve repair, which should ideally be tension-free, these results show that the PTB method can provide fixation strengths

approaching that of conventional microsurgery and that the PTB repair is unlikely to be disturbed in vivo.

The results in Figure 5 were obtained with PTB parameters of 0.1% Rose Bengal with 532 nm light delivered at an irradiance of 0.5 W/cm² and a fluence of 60 J/cm². To further explore the optimal dosimetry conditions for the rat experiments in Aim 2, we performed a fluence dependence study using light delivered at an irradiance of 0.5 W/cm² for various durations. Figure 6 shows the fluence dependence of the bond strength formed using PTB/xHAM (4mM/1mM (EDC/NHS)) to reattach the nerve segments ex vivo. A fluence of 60 J/cm² resulted in superior bond strength in comparison to 30, 120 and 240 J/cm² (p<0.05). Interestingly, bond strength was significantly weaker with the highest fluence, presumably due to increasing friability of the amnion wrap. Those nerve/wrap preparations treated with no illumination predictably had negligible bond strength. An irradiance of 0.5W/cm² and a fluence of 60J/cm² equates to an illumination duration of 120 seconds (60 seconds per nerve/wrap side) and this was felt to be clinically acceptable for use in the animal studies in Aim 2.

The anatomy of HAM is shown pictorially in Figure 7. In vivo, the epithelial layer is in contact with the amniotic fluid while the spongy layer is in contact with the chorion. During preparation the amnion is de-epithelialized but there remains the question as to “which way up” the HAM should be bonded to the nerve. To that end we performed experiments where we were careful to note the surface in contact with the nerve and the resultant bond strength obtained after bonding with 60 J/cm² of 532 nm light delivered at an irradiance of 0.5 W/cm². Figure 8 shows that the bond strength between nerve and amnion was not significantly different to the epithelial and chorionic surfaces. This finding has helped simplify processing and storage of the amnion and also intra-operative handling.

The outcome of these experiments has helped confirm the following optimum conditions to apply to the in-vivo rodent survival operations in aim 2:

- Amnion cross-linked with 4mM 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC)/1mM H-hydroxysuccinimide (NHS)
- Laser irradiance of 0.5 W/cm² and fluence of 60 J/cm²
- Amniotic epithelial and chorionic surfaces bond equally well

Task 1i. Data analysis, conclusions and consideration of alternative wrap materials, if required (Months 6-10, MGH: Redmond)

In this task we focused on determining the best candidate commercial wrap to evaluate in PTB studies for nerve repair. As outlined in the original project proposal, our plan was to use a tubular swine intestinal submucosa (SIS) product called AxoGuard manufactured by AxoGen. Following initial trials this product was found to be unsuitable as it was too thick and possessed too much inherent shape memory to permit easy wrapping around the small diameter rat sciatic nerve. Intimate contact between tissue surfaces is an essential pre-requisite for photochemical tissue bonding. Following this discovery, we conducted a thorough search of alternative commercially available biomaterials that could satisfy our needs.

The following products were sampled and assessed for their conformability and bonding ability:

1. AxoGuard (multi-layer SIS – AxoGen)
2. NeuraGen (Collagen – Integra)
3. NeuraWrap (Collagen – Integra)
4. Tenoglide (Collagen – Integra)
5. NeuraMend (Collagen – Stryker)
6. NeuraMatrix (Collagen – Stryker)

7. Colafilm (Collagen – Innacol)
8. Amniofix (Collagen – MiMedx)
9. SurgiMend (Collagen – TEI Biosciences)
10. Oasis (Single layer SIS – HealthPoint)

Options 1-9 were also found to be unsuitable due to similar problems regarding excessive material thickness, stiffness, shape memory and inability of the material to conform satisfactorily around the rat sciatic nerve.

Option 10 is a single layer SIS product that met our requirements. Oasis is a product marketed and distributed as a wound dressing by HealthPoint. Although the material is approximately double the thickness of human amnion, it was sufficiently thin to allow circumferential nerve wrapping and close contact between wrap and epineurium. In fact, we discovered that both AxoGuard and Oasis SIS products are manufactured by Cook Medical. AxoGuard is simply a multi-layered SIS product. Given that the large animal studies with large caliber nerves in Aim 3 will use AxoGuard we feel that the single layer SIS material is totally appropriate for these small animal studies

Biomechanical testing of the single layer SIS material showed a Young's modulus (Fig. 9) and a maximum load to failure (Fig. 10) that were considerably greater than that of human amnion (See Figs. 1 and 2). Similar to earlier HAM studies the chemical crosslinking of SIS with increasing concentrations of EDC/NHS, gave an increase in Young's modulus and maximum load to failure. (Figs. 9 and 10). Digestion with 0.1% collagenase showed that increasing the concentration of EDC/NHS crosslinker reduced the rate of proteolytic degradation (Fig. 11) thus, extending the longevity of the material in vivo. This finding was consistent with those results observed with HAM and satisfied our goal to increase the in-vivo survival of nerve wrap biomaterial during long periods of recovery associated with large nerve deficit reconstruction and long nerve grafts.

As with the human amnion nerve wraps, it was important for us to confirm that, in addition to increasing the resistance to enzymatic degradation, EDC/NHS crosslinking of SIS did not interfere with photochemical tissue bonding. Figure 12 shows that there was no significant drop in bond strength measured by ex-vivo tensiometer testing. Interestingly, bond strength between SIS nerve wrap and sciatic epineurium was significantly greater than that found with untreated and crosslinked human amnion.

Milestones for Task 1 include the following, with decisions taken at the joint meetings of Partnering PI's, held at MGH, WRNMMC and mutual conferences.

- *Obtain MGH and DOD approvals for all protocols*
- *Determine mechanical properties of wrap materials and establish suitability for use in PTB. Modify processing of wraps and utilize alternate wraps if necessary*
- *Determine bond strength of wraps to ex-vivo nerve and synthetic graft. Modify processing of wraps and utilize alternate wraps if necessary.*
- *Collate results and determine best nerve wraps to use in Task 2*
- *Prepare publications and presentations based on Task 1 research results*

All the above milestones have been met so far, as described above.

Task 2 – Determine efficacy of nerve regeneration in a rodent model of segmental nerve deficit injury as a function of wrap and fixation procedure. (Months 6-22).

Task 2a. Regulatory approval for rodent study of segmental deficit repair by MGH IACUC and review and approval by USAMRMC Office of Research Protections (ACURO). (Months 1-4, MGH: Redmond/Winograd/Randolph)

This aim involves a large study of peripheral nerve repair in a rat sciatic nerve model using isogenic Lewis rats. The animal protocol for these experiments has been reviewed and received IACUC approval at the Massachusetts General Hospital (protocol #2012N000117) and was also granted ACURO approval on 11/19/2012.

Task 2b. Rodent surgeries for segmental deficit and repair using isograft with 110 Lewis rats. (Months 6-8, MGH: Winograd/Randolph)

A total of 110 rodents underwent survival surgery, commencing on 2/26/2013 and completed on 5/10/2013.

Task 2c. Biweekly functional recovery testing by gait analysis in isograft study. (Months 6-14, MGH: Redmond)

Functional recovery in each rodent during the 5-month follow-up period has been measured by monthly walking track analysis using the well-established Sciatic Function Index (SFI) as calculated from the paw-prints of the rodents as a function of time after surgery. As predicted, those animals in the negative control group (no repair following nerve deficit injury) experienced no functional recovery as illustrated by a complete lack of correction of the sciatic function index (-96.2+/-3.7). A value of -100 indicates zero functional recovery. Despite performing well in ex vivo experiments, those isografts wrapped with the commercially sourced SIS material performed worst out of all biological nerve wraps. In SIS+suture and SIS+PTB groups, this was statistically significant in comparison to standard repair (positive control). Isografts wrapped with cross-linked human amnion and secured with PTB (xHAM+PTB) exhibited the greatest functional recovery value although this did not reach statistical significance in comparison to the positive control group (SFI = -67.93+/-5.11 vs -71.69+/-4.80). There was no statistically significant difference between any of the remaining experimental groups compared to the positive control group. Table 1 provides a summary of the functional data.

It should be noted that although walking track analysis is a well-accepted outcome measure for functional assessment following rodent peripheral nerve surgery, the method has several recognised limitations that have been encountered. The inked footprints are rarely perfect and are open to considerable inter-observer variability. Walking track analysis is also limited by experimental hind-paw clawing. Although this was not a major problem, a small number of rodents at later time points in the majority of experimental groups were excluded from the calculation of mean SFIs due to unmeasurable footprints. Clawing results due to injury and incomplete recovery of the nerve supply to intrinsic musculature in the hind foot and may be exacerbated by the lack of physical rehabilitative measures that would ordinarily be introduced in human subjects. Although not thoroughly assessed, the presence of clawing did not seem to affect the reinnervation and retention of gastrocnemius muscle mass and therefore this problem may be limited to walking track analysis. Thus, SFI results are useful but a decision regarding which wrap and fixation method to proceed with in the next step could not be taken solely on this metric.

Task 2d. Gastrocnemius muscle harvest and muscle mass retentions in isograft study (Months 12-14, MGH: Redmond)

Following sacrifice, both experimental left sided gastrocnemius muscle and contralateral right-sided control muscle were harvested from each rodent. Wet muscle mass was recorded immediately following

harvest and percentage muscle mass retention calculated. Mean muscle mass retention in the negative control group was only 9.2% +/- 0.92. As expected, this was significantly less than that achieved in the standard repair group. Greatest muscle mass retention occurred in the xHAM+PTB group and the increase over standard repair was statistically significant (67.3% +/- 4.44 vs 60.0% +/- 5.16; p=0.02). There was no statistically significant difference between the positive control group and any of the remaining experimental groups. Wraps secured with fibrin glue performed consistently better than those secured with suture although these effects did not reach statistical significance. Table 2 provides a summary of muscle mass retention data.

Task 2e. Histomorphometric analysis of proximal and distal fibers in isograft study (Months 12-16, MGH: Redmond/Winograd)

Following sacrifice, left sciatic nerves were harvested and sent for histology. Nerves were harvested from a distance 5mm proximal to the proximal isograft neurotomy site to 5mm distal to the distal neurotomy. Following 24-hours of fixation, each nerve was cut into proximal, mid-graft and distal sections. Following dehydration and epoxy resin embedding, 1-micron slices of each specimen were cut and mounted for histomorphometric analysis. Axon counts and G-ratio (marker of myelination) will be measured from scanned images. All histologic specimens were collected and prepared for sectioning.

Histomorphometric analysis of all groups is now complete (Table 3; Fig 13). Axon counts at the distal section site did not differ significantly between treatment groups and positive controls. Those nerves that were repaired with crosslinked amnion and PTB (xHAM+PTB) recovered greatest fiber diameter, axon diameter and myelin thickness (Table 3; Fig 13). The corrected values (G-ratio) for myelination were not significantly different. These results support earlier functional data on sciatic function index (SFI) and muscle mass retention with xHAM+PTB performing best of all repair procedures.

Task 2f. Determination of axonal migration, endoneurial scarring in isograft study. (Months 12-16, MGH: Redmond/Randolph/Winograd)

Previous studies from our lab have suggested that, in addition to creating a watertight seal at the neurotomy site, photochemical tissue bonding also reduces the formation of fibrinous adhesions around the nerve. Although this is difficult to quantify, observations following rodent sacrifice in this study have supported this (Figure 14).

Task 2g. Decision on wrap/fixation method for Avance™ nerve graft studies in rodent model. (Month 16, All PI's)

This decision was made based on the composite results from gait analysis, muscle mass retention and histomorphometric studies that showed that of all approaches used the best outcomes were reproducibly observed when the PTB+xHAM procedure was used (see above). This was due to statistically significant increases in gastrocnemius muscle mass retention and histomorphometric outcomes including fiber diameter, axon diameter and myelin thickness along with equivalent SFI results in comparison to standard microsurgical repair of peripheral nerve injury involving significant nerve deficit using autograft (isograft) repair. Thus, for the allograft studies the wrap/fixation method chosen was PTB+xHAM in a model of rat sciatic nerve injury with deficit.

As detailed in the original project proposal, our intention was to purchase the Avance processed human allograft from AxoGen for use in the final phase of the rodent studies. After discussion with AxoGen and a review of the most recent literature on the use of human versus rat allograft in rodent models, we were advised that the human-sourced nerve may not be optimal for rodent studies (due to the potential

for cross-species immunoreactivity being greater than originally expected) and that for the purpose of these studies it would be better use processed rodent allograft. We have since harvested, frozen and stored donor rat sciatic nerve before shipping to AxoGen for processing. A Material Transfer Agreement was agreed between MGH and AxoGen and delivery of the processed nerve was concluded in December 2013. Allografts were of excellent quality. Microscopic examination revealed no evidence of epineurial fragmentation and fascicular architecture appeared intact. The grafts were pliable and resembled autogenous tissue.

Task 2h. Rodent surgeries for segmental deficit and repair using rat processed allograft (MGH: Winograd/Randolph)

Surgery has now been completed on the remaining 2 groups (n=10) of acellular nerve allograft (ANA) repairs in rodents. ANA secured using xHAM+PTB (using optimal crosslinking conditions of 4mM/1mM EDC/NHS and 532 nm illumination at an irradiance of 0.5 W/cm² and fluence of 60 J/cm²) were compared to a control group where ANA was secured using conventional 10-0 epineurial suture. The rat-processed nerve bore very close resemblance to freshly harvested autogenous nerve and was easy to handle intra-operatively. No issues were encountered when photochemically bonding the nerve. Walking track analysis, calculation of sciatic function index (SFI) (Fig 15, Table 4)) and measurement of gastrocnemius muscle mass retention is now complete (Fig 16, Table 5). Histology processing of excised nerve tissue has also now been completed and results of histomorphometric analysis are summarized in Table 6.

Task 2i. Biweekly functional recovery testing by gait analysis in Avance® nerve graft study. (Winograd/Randolph/Redmond - Months 16-20)

While photochemically sealed ANA showed an improvement in SFI in comparison to sutured ANA (-80.3+/-4.2 vs. -78.3+/-5.0; Table 4 and Fig.15), the difference was not statistically significant. When the analogous isograft studies from this Task were included for comparison, the isograft+xHAM/PTB group recovered greatest SFI after 5-months follow-up (Table 4). This was not statistically significant in comparison to isograft+suture. ANA+suture performed statistically worse than isograft+suture (-80.3+/-4.2 vs. -71.7+/-4.8; p=0.0019; Table 1 and 4). SFI was also statistically less for ANA+xHAM/PTB in comparison to isograft+xHAM/PTB (Table 4) but not significantly different from the standard of care isograft+suture repair.

Task 2j. Gastrocnemius muscle harvest and muscle mass retentions in Avance® nerve graft study. (Winograd/Randolph/Redmond - Months 20-22)

Although ANA+xHAM/PTB displayed a trend towards greater muscle mass recovery in comparison to ANA+suture, this result was not statistically significant (55.2+/-5.5% vs. 52.9+/-4.77%; Table 5, Fig 16). When the analogous isograft repairs are considered, the isograft+xHAM/PTB recovered greatest gastrocnemius muscle mass retention and this was statistically significant in comparison to all other groups (Table 5). Muscle mass recovery was statistically poorer in ANA+suture group in comparison to isograft+suture group. Likewise, recovery of ANA+xHAM/PTB was statistically poorer than isograft+xHAM/PTB. Muscle mass retention in the ANA+xHAM/PTB group was statistically comparable to that achieved using gold standard isograft+suture.

Task 2k. Histomorphometric analysis of proximal and distal fibers in Avance® nerve graft study. (Winograd/Randolph/Redmond - Months 20-22)

Although not statistically significant, fiber diameter, axon diameter and myelin thickness in the ANA+xHAM/PTB group displayed a trend towards greater recovery in comparison to ANA+suture. When considering analogous isograft repair groups, isograft+xHAM/PTB recovered the greatest fiber diameter, axon diameter and myelin thickness and this was statistically significant in comparison to all other groups (Table 6). Axon counts in the distal nerve stump were significantly greater for isograft+xHAM/PTB in comparison to ANA+suture. No other significant differences in axon counts or axon density existed between treatment groups. Histomorphometric recovery was poorest in the ANA+suture group and this was statistically significant in comparison to isograft+suture and isograft+xHAM/PTB. There was no significant difference between ANA+xHAM/PTB and gold standard isograft+suture (Table 6).

Task 2l. Determination of axonal migration, endoneurial scarring in Avance® nerve graft study. (Winograd/Randolph/Redmond - Months 20-22)

As with previous isograft studies, the formation of extraneural scar tissue was qualitatively less with photochemical repair in comparison to suture repair (Fig 17).

Task 2m. Decision on Avance® nerve graft procedure for large animal studies in Aim 3. (All PIs, Month 22)

This study shows that outcomes following light-activated sealing of ANA with PTB/xHAM are improved in comparison to sutured ANA, and are statistically equivalent to gold standard sutured isografts. Based on these findings, isografts that are photochemically sealed with crosslinked amnion nerve wraps have emerged as the superior reconstructive method for large gap nerve repair. However, the ability to elevate the performance of ANA to match the current gold standard is of major clinical interest when severe injuries, complicated with limb loss, preclude the use of autograft. The demonstrated success of light-activated sealing of isograft coaptation sites is likely related to the creation of a water-tight seal and the subsequent containment of the neurotrophic rich milieu. It is possible that removal of Schwann cells (SC), and the neurotrophic factors they liberate, during decellularization of ANAs, may partially abrogate this effect. However, the benefit observed suggests that the technique remains advantageous. The avoidance of suture-induced inflammation and fibrosis, the exclusion of infiltrating scar tissue and the prevention of axonal escape may be responsible.

Following decellularization, ANAs consist of basal lamina scaffolds. Components of the basal lamina such as fibronectin and laminin have pro-regenerative effects on neurite outgrowth and can support axonal regeneration in the absence of SCs. Whilst this may be sufficient over short lengths of ANA, successful regeneration across longer lengths is dependent on re-population of ANA by resident SCs. This study did not assess the extent of SC re-population but it is possible that photochemical sealing may augment SC migration, further improving regeneration through long ANAs. Investigating SC repopulation in ANAs of varying length may form the basis of future investigation. When combined with cell-based therapy and tissue engineering, sealing the regenerative milieu and maintaining high levels of growth promoting factors at the repair site, may amplify this effect.

With these acellular nerve allograft studies showing equivalent nerve regeneration outcomes for photochemical sealing of ANA or standard of care sutured autograft and promise in the use of ANA in this fashion for wounded warriors that lack sufficient autograft for standard repair, we are confident in this approach to progress to large animal studies.

Milestones for Task 2 include the following, with decisions taken at the quarterly meetings of Partnering PI's, held alternately at MGH and WRNMMC.

- *Obtain MGH and DoD approvals for all protocols.*
- *Complete all rodent surgeries and repair groups for isograft study.*
- *Determine lead wrap/fixation method for best functional recovery in isograft model and proceed with this method for Avance nerve graft study*
- *Determine method for rodent acellular nerve allograft (ANA) preparation*
- *Complete all rodent surgeries and repair groups for Avance nerve graft study*
- *Analyze recovery of function, muscle mass retention and histomorphometry data.*
- *Determine animal model and best photosealing procedure for Avance nerve graft vs autograft large animal study in Task 3*
- *Prepare publications and presentations based on Task 2 research results.*

All above milestones have now been completed

Task 3. Explore efficacy of lead wrap/fixation approach against standard of care autograft in large animal model of large segmental nerve deficit.

Task 3a. Regulatory approval for sheep study of segmental deficit repair. (Winograd, Randolph/Redmond, Months 18-22)

We have recently concluded a very long period of rodent studies to determine the best wrap/fixation method for use in repair of large segmental nerve deficits using autograft or allograft. The best performing repair method in the autograft study was clearly photosealing using a crosslinked human amnion wrap. In the allograft study, this method again performed well, reaching statistical equivalence to standard of care autograft repair using suture. Thus, the potential exists for application of this technology to wounded warriors where there is an insufficient supply of appropriate autograft.

Final validation of this technology for human application requires the large animal studies of segmental deficit repair in Phase 3. The original plan for these studies was repair of a 5 cm deficit in the median nerve of the sheep forelimb. The sheep model was chosen on the grounds of an established experimental model with negligible morbidity to the animal. However, in discussion with Axogen over the use of allograft in this model it was concluded that the sheep nerve structure was considerably different from human nerve and may not be suitable for the placement of the Avance product. We also considered the preparation of sheep allograft for use in this study but the structural differences caused our consultants at Axogen some trepidation as prior experience with sheep nerve using their patented detergent-based approach to human allograft preparation proved unsatisfactory. The decision was then taken to use a swine median nerve deficit model, which is equally well-tolerated by the animals, using the Avance acellular nerve allograft (from humans) in the swine model. Researchers at Axgen did not believe there would be a problem with immunogenicity with these studies but we proposed a small pilot study for immune reaction to the use of this material in the swine model before proceeding to the full study. The protocol for these studies was submitted to the IACUC at MGH in early October 2014 and is under review. On approval, the protocol will be forwarded to ACURO for approval before commencing experiments in December 2014.

The following tasks will be completed in Year 3.

- 3b. Surgical procedures to create segmental deficit in swine model. (Months 23-25)*
- 3c. Delayed repair of nerve deficit by Avance® and autograft in swine model. (Months 23-25)*
- 3d. Electrophysiology measurements in swine. (Months 23-30)*

- 3e. *Histomorphometric analysis of proximal and distal fibers in swine study. (Months 30-32)*
- 3f. *Determination of axonal migration, endoneurial scarring in swine study. (Months 30-32)*
- 3g. *Preparation of manuscript based on Task 3 studies and evaluation for recommendation for human studies.*

Key Research Accomplishments:

- Demonstrated that human amniotic membrane (HAM) can be strengthened by a biocompatible crosslinking process.
- Demonstrated that crosslinking dramatically increases resistance of HAM to biodegradation, thus, increasing its longevity as a wrap for nerve graft sealing in vivo.
- Demonstrated that chemical, crosslinking of HAM does not affect its ability as a wrap for photochemical sealing over nerve graft coaptation sites.
- Demonstrated that photochemical sealing of crosslinked HAM (xHAM) over nerve graft coaptation sites can be performed in a facile manner in a rat sciatic nerve model.
- Demonstrated that commercial single layer SIS (swine intestinal submucosa, Oasis) is a stronger material than HAM and can be further strengthened by chemical crosslinking with increased resistance to biodegradation.
- Demonstrated that both SIS and HAM and their crosslinked derivatives can be used as photochemical wraps in vivo over nerve graft coaptation sites in a facile manner in a rat sciatic nerve model.
- Demonstrated improved nerve regeneration in a functional recovery model (SFI, sciatic function index) using PTB/xHAM wrap compared to standard (suture) of care microsurgery.
- Demonstrated improved nerve regeneration in a muscle mass retention model (contralateral control (unoperated) vs treated sciatic nerve graft) using PTB/xHAM wrap compared to standard (suture) of care microsurgery.
- Demonstrated improved nerve regeneration in histomorphometric outcomes using PTB/xHAM wrap compared to standard (suture) of care microsurgery.
- Determined PTB/xHAM wrap to be the optimal fixation method for nerve repair involving large segmental deficit in a rodent model.
- Demonstrated that detergent-based processing of rat nerve can provide excellent acellular nerve allograft (ANA) for study of nerve regeneration in a rodent model.
- Demonstrated improved nerve regeneration outcomes of PTB/xHAM fixation of rat ANA compared to suture fixation.
- Demonstrated regeneration outcomes using ANA + PTB/xHAM repair of segmental nerve deficit in a rodent model equivalent to standard of care autograft/suture repair.
- Demonstrated that ANA + PTB/xHAM is a potential solution to providing optimal nerve regeneration outcomes in wounded warriors with insufficient autograft.
- Outlined a large animal protocol for comparative study of standard of care autograft vs ANA + PTB/xHAM in nerve regeneration using electrophysiological outcomes.

Reportable Outcomes:

Conference Presentations:

1. Fairbairn NG, Ng-Glazier J, Meppelink A, Randolph MA, Winograd JM, Fleming ME, Valerio IL, Kochevar IE, Redmond RW. Large extremity peripheral nerve repair. Military Health System Research Symposium (MHSRS) Fort Lauderdale, FL. August 12-15, 2013.
2. Fairbairn NG, Ng-Glazier J, Meppelink A, Randolph MA, Winograd JM, Fleming ME, Valerio IL, Kochevar IE, Redmond RW Treating Peripheral Nerve Injuries with Photochemical Tissue Bonding in Military and Civilians. 118th AMSUS Annual Continuing Education Meeting, Seattle, WA. November 3-8, 2013.
3. Fairbairn NG, Ng-Glazier J, Meppelink A, Randolph MA, Winograd JM, Fleming ME, Valerio IL, Kochevar IE, Redmond RW. Improved outcome following nerve graft reconstruction: The application of photochemical tissue bonding and human amnion nerve wraps in a rodent model of large deficit nerve injury. 41st Meeting of the New England Hand Society. Sturbridge MA, December 6, 2013.
4. Fairbairn NG, J Ng-Glazier, Meppelink A, Randolph MA, Winograd JM, Fleming ME, Valerio IL, Kochevar IE, Redmond RW. Annual Meeting of the American Society for Peripheral Nerve, Koloa, HI. January 10-12, 2014.
5. Fairbairn NG, Ng-Glazier J, Meppelink A, Randolph MA, Winograd JM, Fleming ME, Valerio IL, Redmond RW. Improving neuroregeneration following large deficit peripheral nerve injury: the application of human amniotic membrane scaffolds and photochemical tissue bonding (PTB). Poster presentation. 8th Symposium on Biologic Scaffolds for Regenerative Medicine, Silverado Resort, Napa, California, USA, 24-26th April, 2014.
6. Fairbairn NG, Ng-Glazier J, Meppelink A, Randolph MA, Winograd JW, Fleming ME, Valerio IL, Redmond RW. The application of Photochemical Tissue Bonding (PTB) for Large Deficit Peripheral Nerve Injury. Poster Presentation. 3rd Annual Harvard Research Day. 10th May 2014, Thomas Martin Conference Centre, Harvard Medical School, Boston, MA, USA.
7. Fairbairn NG, Ng-Glazier J, Meppelink A, Randolph MA, Winograd JM, Fleming ME, Valerio IL, Redmond RW. The application of photochemical tissue bonding for large deficit nerve repair. Oral presentation. 25th Annual Smith Day & Inaugural Jupiter International Forum. Royal Sonesta Hotel, Cambridge, MA, USA. 30th May 2014.
8. Fairbairn NG, Ng-Glazier J, Meppelink A, Randolph MA, Winograd JM, Fleming ME, Valerio IL, Redmond RW. The application of photochemical tissue bonding for large deficit nerve repair. Oral presentation. The 55th Annual meeting of the New England Society of Plastic and Reconstructive Surgeons (NESPRS), Sebasco Harbour Resort, Sebasco Harbour, Maine, USA, 6-8th June, 2014. *Joseph E Murray award for best presentation.
9. Fairbairn NG, Ng-Glazier J, Meppelink A, Randolph MA, Winograd JM, Fleming ME, Valerio IL, Redmond RW. Improving outcome following large deficit peripheral nerve injury: the application of a human amnion nerve wrap and photochemical tissue bonding (PTB). Oral presentation at the quadrennial congress of the European Society of Plastic Reconstructive and Aesthetic Surgeons (ESPRAS), Edinburgh, Scotland, UK, 6-11th July 2014.
10. Redmond RW. Clinical applications of photochemical crosslinking. Invited Talk at the Gordon Conference for Lasers in Medicine and Biology. Holderness, NH. July 13-18, 2014.

Publications

1. Fairbairn NG, Randolph MA, Redmond. The clinical applications of amnion in plastic surgery. *J Plast Reconstr Aesthet Surg*. 2014 May;67(5):662-675.

2. Fairbairn NG, Ng-Glazier J, Meppelink A, Randolph MA, Winograd JM, Fleming ME, Valerio IL, Redmond RW. Light-Activated Sealing of Nerve Graft Coaptation Sites Improves Outcome Following Large Gap Peripheral Nerve Injury. *Plast Reconstruct Surg.* 2014, in press.
3. Fairbairn NG, Ng-Glazier J, Meppelink A, Randolph MA, Winograd JM, Fleming ME, Valerio IL, Redmond RW. Large gap nerve reconstruction using acellular nerve allografts and photochemical tissue bonding. *Plast Reconstruct Surg.* 2014, submitted.

Conclusion:

At the end of the first two years of this project there have already been some notable discoveries that may impact military health care in the near future. There is a clear need in military medicine to improve outcomes in wounded warriors that undergo severe extremity injury involving large gap peripheral nerve injury. We have demonstrated that biocompatible chemical crosslinking can be used to strengthen thin nerve wraps and increase resistance to biodegradation such that the wrap retains its sealing ability throughout the time taken for the regenerating axons to traverse the nerve graft and pass the distal coaptation site. The light-activated sealing of the nerve wrap around the coaptation sites obviates the need for suture attachment of the graft and a host of advantages result from the lack of needle injury, inflammation and scarring, possible infection and axonal splay that can reduce functional recovery and cause neuroma formation. Processed crosslinked human amnion, a thin biological membrane (< 50 micron), has demonstrated the best potential as a nerve wrap for photochemical sealing in rodent models in vivo, with other commercial nerve wraps having proven less suitable due to greater thickness and inability to conform to the dimensions of the rat sciatic nerve. Using a photosealing approach with this material we have shown significant improvement in nerve regeneration outcomes in rodent models of segmental nerve defect bridged by autologous nerve graft

The severity of injury in wounded warriors exposed to IED blast can involve massive soft tissue damage and amputation. As such, donor autologous nerve may be unavailable for repair purposes and alternatives are sought. We focus on human allograft as a potential solution. Although allograft has proven to be less effective for nerve regeneration than autograft in clinical implementation using microsurgical attachment, we hypothesized that the photosealing benefit may improve outcomes. In the same rodent model we have demonstrated that photosealing allograft as a nerve bridge produces nerve regeneration that is not statistically different from standard of care sutured autograft. This represents major progress in the treatment of peripheral nerve injury associated with military trauma. If these improvements translate clinically, this could result in important improvements in peripheral nerve recovery in those cases of severe trauma and limb loss where the use of nerve autograft is not possible. With refinement, and with parallel advances in stem cell therapy and tissue engineering, this technique, when used in conjunction with acellular nerve allograft, has the potential to supplant the use of autografts following large gap injury. This approach is capable of rapid commercialization and translation into military medicine. The IP has been filed and the materials involved can be easily stored in a prolonged manner for rapid deployment.

Table 1. Monthly mean SFIs. After 5-months follow-up, those nerves repaired with photochemically sealed crosslinked amnion (xHAM+PTB) recovered greatest SFI, although this was not statistically significant. The negative control and those nerves repaired using xSIS+suture and xSIS+PTB performed statistically worse than graft+suture.

Experimental group	Mean SFI				
	1-month	2-month	3-month	4-month	5-month
No Repair	-89.0+/-3.9	-94.2+/-4.7*	-89.8+/-5.1*	-89.3+/-4.8*	-96.2+/-3.7*
Standard Graft + Suture	-87.6+/-5.0	-81.1+/-4.5	-71.8+/-7.3	-74.7+/-6.3	-71.7+/-4.8
HAM+suture	-90.3+/-5.2	-85.7+/-9.8	-80.6+/-3.5*	-79.7+/-5.7	-77.9+/-6.3
HAM+fibrin	-89.2+/-4.0	-81.6+/-4.1	-80.4+/-7.2*	-79.4+/-4.3	-75.2+/-4.6
HAM+PTB	-90.0+/-5.2	-81.2+/-2.4	-72.8+/-4.6	-75.6+/-3.3	-74.5+/-4.5
xHAM+suture	-96.6+/-7.5	-82.4+/-4.8	-80.0+/-4.0*	-81.4+/-4.8	-76.8+/-2.7
xHAM+fibrin	-90.9+/-3.2	-84.1+/-3.6	-79.8+/-3.3*	-81.2+/-3.1	-75.0+/-4.0
xHAM+PTB	-88.2+/-3.9	-80.3+/-3.5	-67.2+/-3.3	-71.6+/-5.5	-67.9+/-5.1
xSIS+suture	-94.7+/-3.9	-85.6+/-4.4	-82.5+/-4.4*	-81.4+/-4.4	-80.3+/-3.2*
xSIS+fibrin	-93.2+/-4.6	-84.7+/-4.9	-82.0+/-3.7*	-81.5+/-3.9	-78.8+/-3.9
xSIS+PTB	-92.5+/-2.0	-84.7+/-5.0	-84.3+/-4.6*	-85.3+/-6.3*	-85.0+/-6.0*

Table 2: Left gastrocnemius muscle mass retention at 150 days post-repair. Those nerves repaired using xHAM+PTB recovered greatest gastrocnemius muscle mass retention. This result was statistically significant. Those nerves repaired using xSIS+suture and xSIS+PTB recovered least gastrocnemius muscle mass. *Statistically significant improvement in comparison to standard graft+suture; p<0.05.

Experimental group	Mean left gastrocnemius muscle mass retention (%)	SD	P value*
No Repair	9.2	0.9	<0.0001
Standard Graft + Suture	60.0	5.2	1
HAM+suture	56.0	5.6	1
HAM+fibrin	59.8	5.4	1
HAM+PTB	62.5	4.0	1
xHAM+suture	57.7	5.1	1
xHAM+fibrin	62.7	4.3	1
xHAM+PTB	67.3*	4.4	0.02*
xSIS+suture	54.9	4.5	0.68
xSIS+fibrin	58.5	5.4	1
xSIS+PTB	54.1	3.2	0.37

Table 3: Histomorphometric analysis of nerve sections 5 mm distal to distal coaptation site in all groups. No difference in axon counts existed between experimental controls and treatment groups. Nerve fiber diameter, axon diameter and myelin thickness were all significantly larger in those nerves repaired using crosslinked amnion and PTB ($p < 0.05$).

Group	Total Axon Count $\times 10^{-3}$	Axon density ($\text{mm}^2 \times 10^{-3}$)	Nerve Fiber diameter (μm)	Axon diameter (μm)	Myelin thickness (μm)	G ratio
Negative Control	0.04+/-0.05*	0.48+/-0.49*	4.25+/-1.28*	2.64+/-1.01*	1.61+/-0.55*	0.61+/-0.09
Positive Control	7.61+/-3.42	29.36+/-18.10	5.47+/-1.70	3.50+/-1.44	1.96+/-0.47	0.62+/-0.08
HAM+suture	10.41+/-3.99	28.85+/-18.61	5.07+/-1.58*	3.44+/-1.39	1.63+/-0.50*	0.67+/-0.17
HAM+fibrin	10.42+/-1.54	29.95+/-14.00	5.22+/-1.67*	3.44+/-1.45	1.78+/-0.45*	0.64+/-0.09
HAM+PTB	9.31+/-4.19	30.70+/-8.94	5.19+/-1.76*	3.47+/-1.53	1.72+/-0.41*	0.65+/-0.09
xHAM+suture	9.79+/-3.35	27.12+/-9.22	5.14+/-1.66*	3.54+/-1.47	1.59+/-0.36*	0.67+/-0.08
xHAM+fibrin	10.87+/-4.32	32.12+/-20.28	5.24+/-1.68*	3.52+/-1.49	1.72+/-0.42*	0.65+/-0.09
xHAM+PTB	9.66+/-3.08	30.73+/-14.73	6.87+/-2.23*	4.51+/-1.83*	2.35+/-0.64*	0.64+/-0.08
xSIS+suture	9.36+/-2.41	30.30+/-16.46	4.83+/-1.42*	3.31+/-1.29*	1.52+/-0.38*	0.67+/-0.08
xSIS+fibrin	6.91+/-2.62	31.55+/-13.37	5.18+/-1.50*	3.58+/-1.30	1.59+/-0.52*	0.68+/-0.11
xSIS+PTB	7.84+/-2.04	30.06+/-13.38	4.81+/-1.49*	3.35+/-1.33*	1.45+/-0.34*	0.68+/-0.08

*Denotes statistical significance in comparison to standard repair group

Table 4: Mean SFI for analogous treatment groups over 5-month follow-up period. At each time point throughout recovery, no significant differences existed within each of the ANA groups.

Group	1 month	2 month	3 month	4 month	5 month
Isograft + Suture	-87.6+/-5.0	-81.1+/-4.5	-71.8+/-7.3	-74.7+/-6.3	-71.7+/-4.8
isograft+xHAM/PTB	-88.2+/-3.9	-80.3+/-3.5	-67.2+/-3.3	-71.6+/-5.5	-67.9+/-5.1
ANA+suture	-95.4+/-2.5	-90.3+/-10.6	-87.9+/-4.0	-84.1+/-3.2	-80.3+/-4.2
ANA+xHAM/PTB	-93.4+/-3.4	-91.1+/-5.4	-88.9+/-5.4	-83.4+/-4.8	-78.3+/-5.0

Table 5: Mean gastrocnemius muscle mass retention for analogous treatment groups at 150 days post-repair.

Experimental Group	Mean gastrocnemius muscle retention (%)	SD	P value*
Isograft+suture	60.0	5.2	----
Isograft+xHAM/PTB	67.3*	4.4	0.01
ANA+suture	52.9*	4.77	0.02
ANA+xHAM/PTB	55.2	5.5	0.22

*Denotes statistical significance in comparison to standard repair group

Table 6: Histomorphometric analysis of nerve sections 5 mm distal to distal coaptation site in all groups. (p<0.05).

Experimental Group	Total axon count (x10 ³)	Axon Density (mm ² x 10 ⁻³)	Nerve fiber diameter (μm)	Axon diameter (μm)	Myelin thickness (μm)	G-ratio
Isograft+suture	7.61+/-3.42	29.36+/-18.10	5.47+/-1.70	3.50+/-1.44	1.96+/-0.47	0.62+/-0.08
Isograft+xHAM/PTB	9.66+/-3.08	30.73+/-14.73	6.87+/-2.23*	4.51+/-1.83*	2.35+/-0.64*	0.64+/-0.08
ANA+suture	5.04+/-2.57	21.50+/-2.56	5.26+/-1.29	3.30+/-1.15	1.76+/-0.86	0.62+/-0.12
ANA+xHAM/PTB	6.04+/-3.20	22.03+/-5.15	5.38+/-1.22	3.41+/-0.99	1.97+/-0.69	0.63+/-0.11

*Denotes statistical significance in comparison to standard repair group

Figure 1A: Effect of EDC/NHS crosslinker concentration on maximum stress of HAM (n=5, * p<0.1, ** p<0.05).

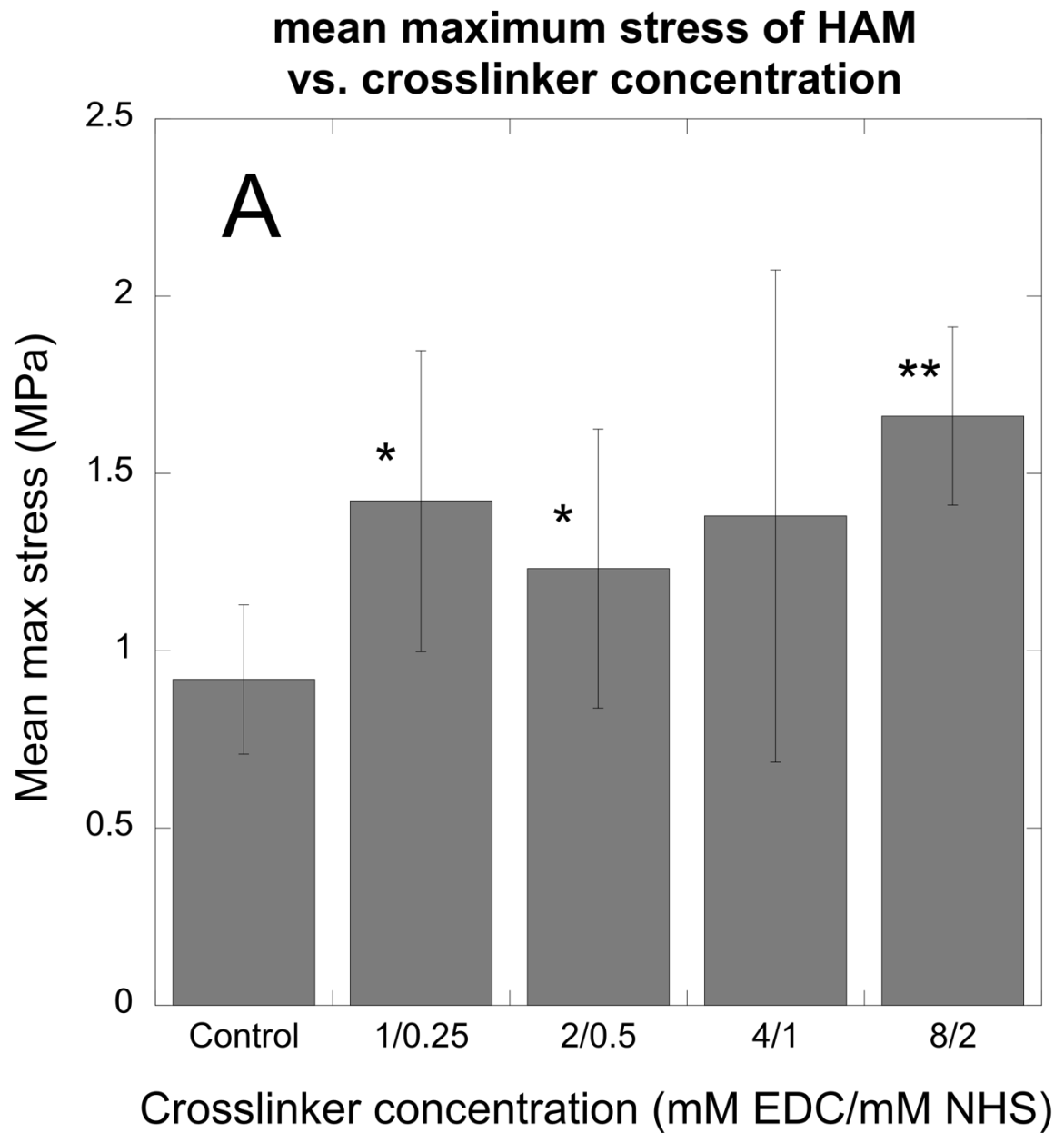


Figure 1B: Effect of EDC/NHS crosslinker concentration on Young's Modulus of HAM (n=5, * p<0.1, ** p<0.05).

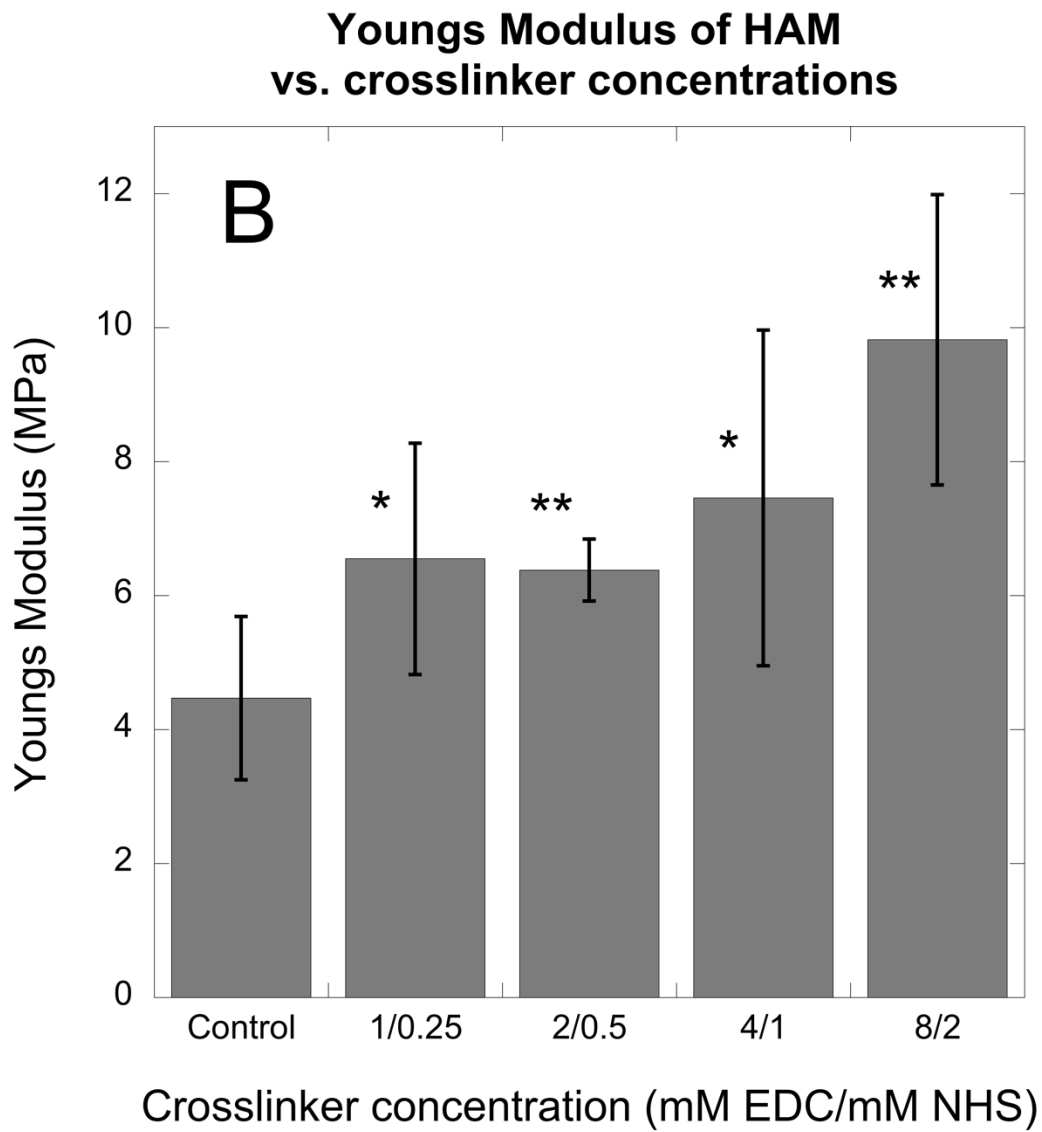


Figure 2A: Effect of EDC/NHS crosslinking on gross degradation time on incubation of control and crosslinked HAM samples with 0.1% collagenase in PBS at 37°C.

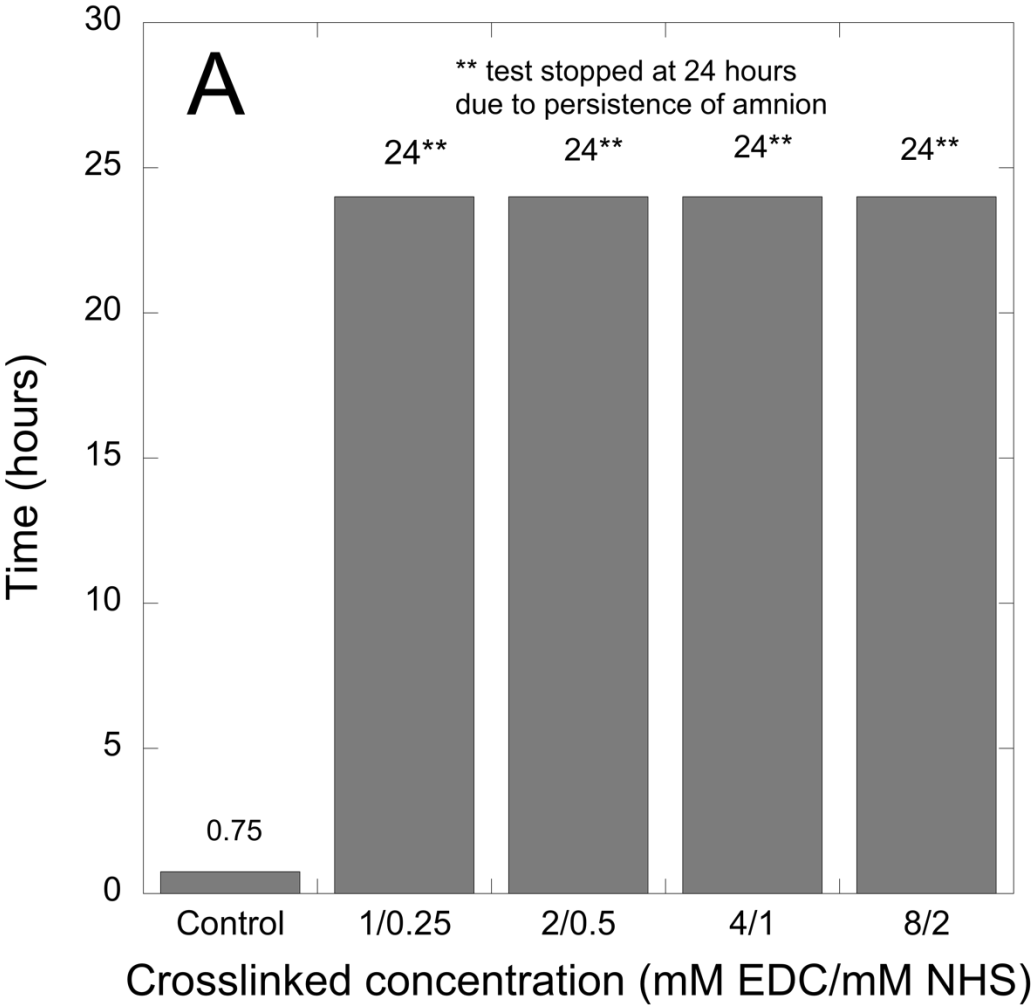


Figure 2B: Effect of EDC/NHS crosslinker concentration on amine containing amino-acid release detected by fluorescamine assay on incubation of control and crosslinked HAM samples with 0.1% collagenase in PBS at 37C.

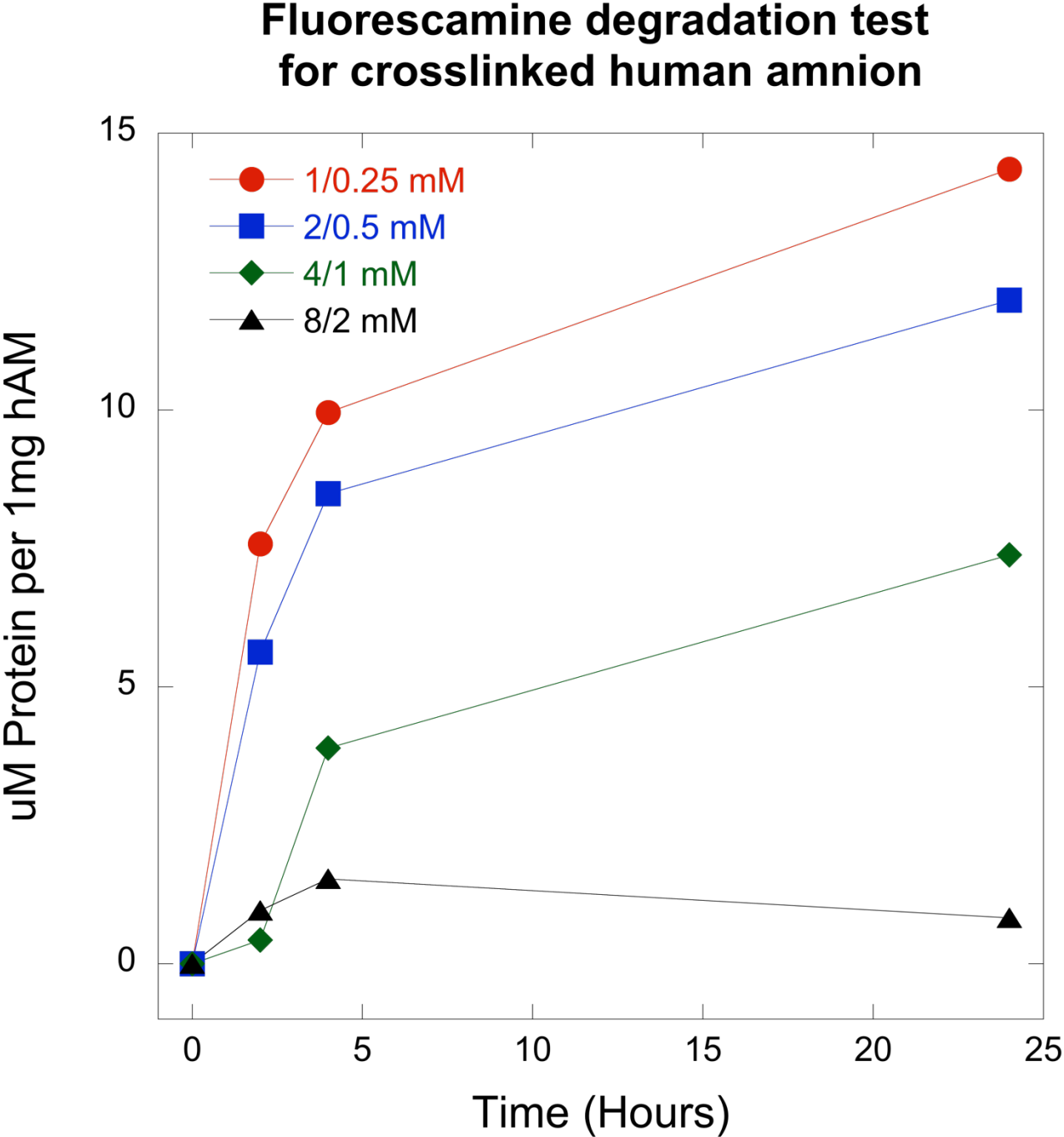


Figure 3: Schematic cartoon of wrap/nerve complex secured in the grips of the mechanical testing device.

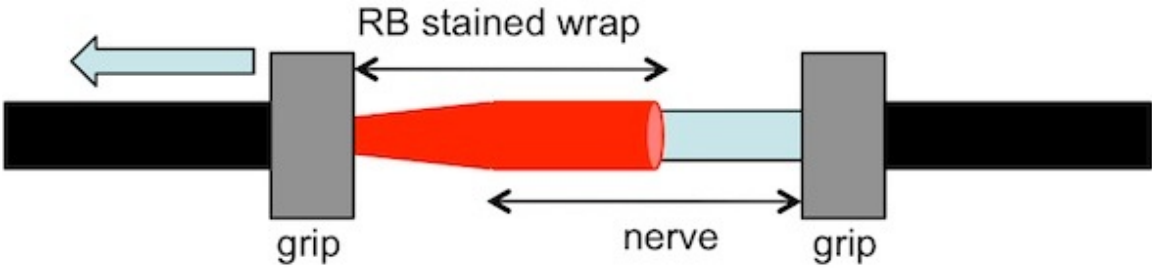


Figure 4: Bond strength between rat sciatic nerve ex vivo and HAM wrap as a function of crosslinking with photochemical bonding using 532 nm light delivered at 0.5 W/cm² and a total fluence of 60 J/cm² (n=5, ** p<0.5).

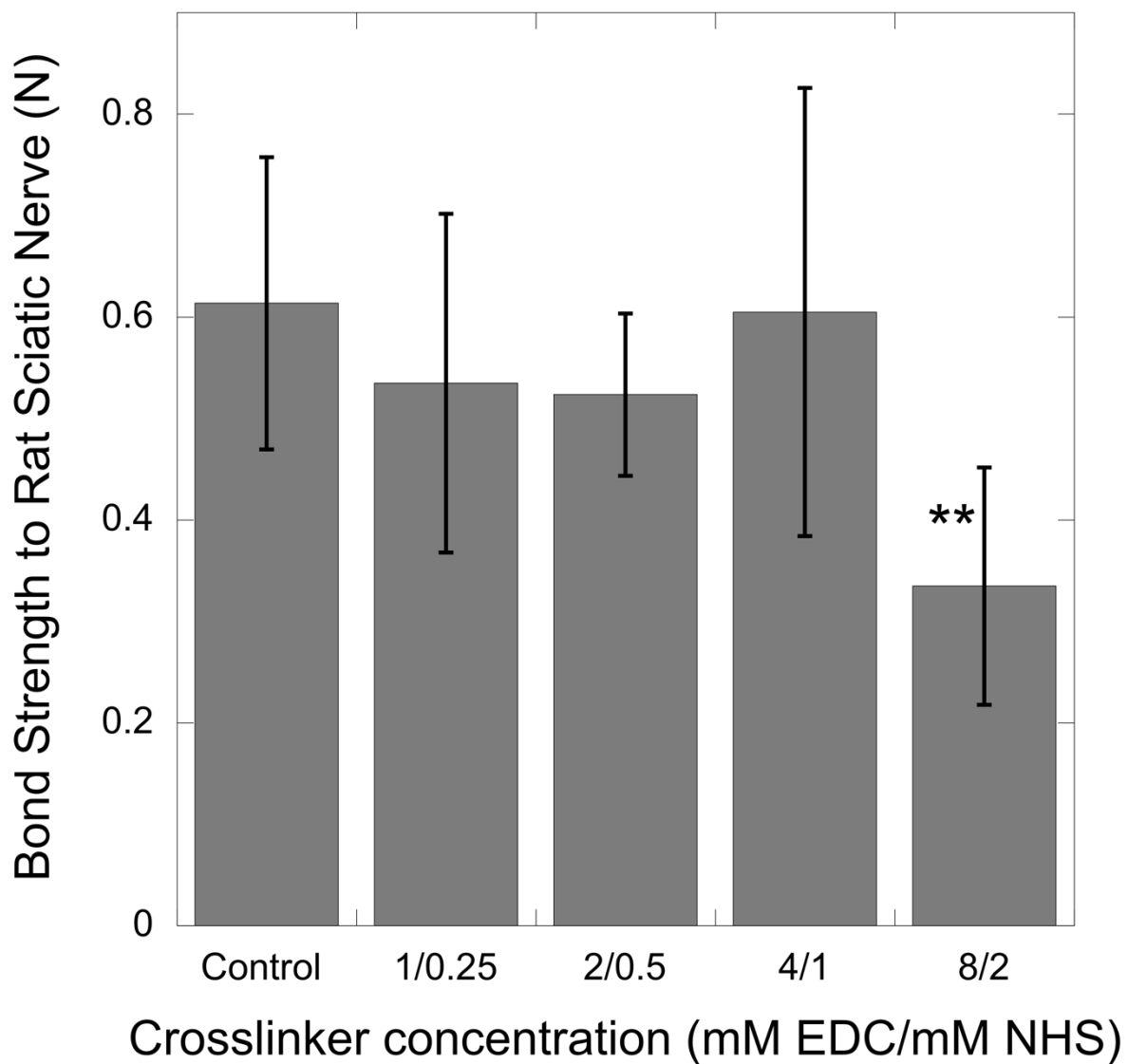


Figure 5: nerve-amnion bond strength as a function of fixation method using 532 nm light delivered at 0.5 W/cm² and a total fluence of 60 J/cm² (n=5).

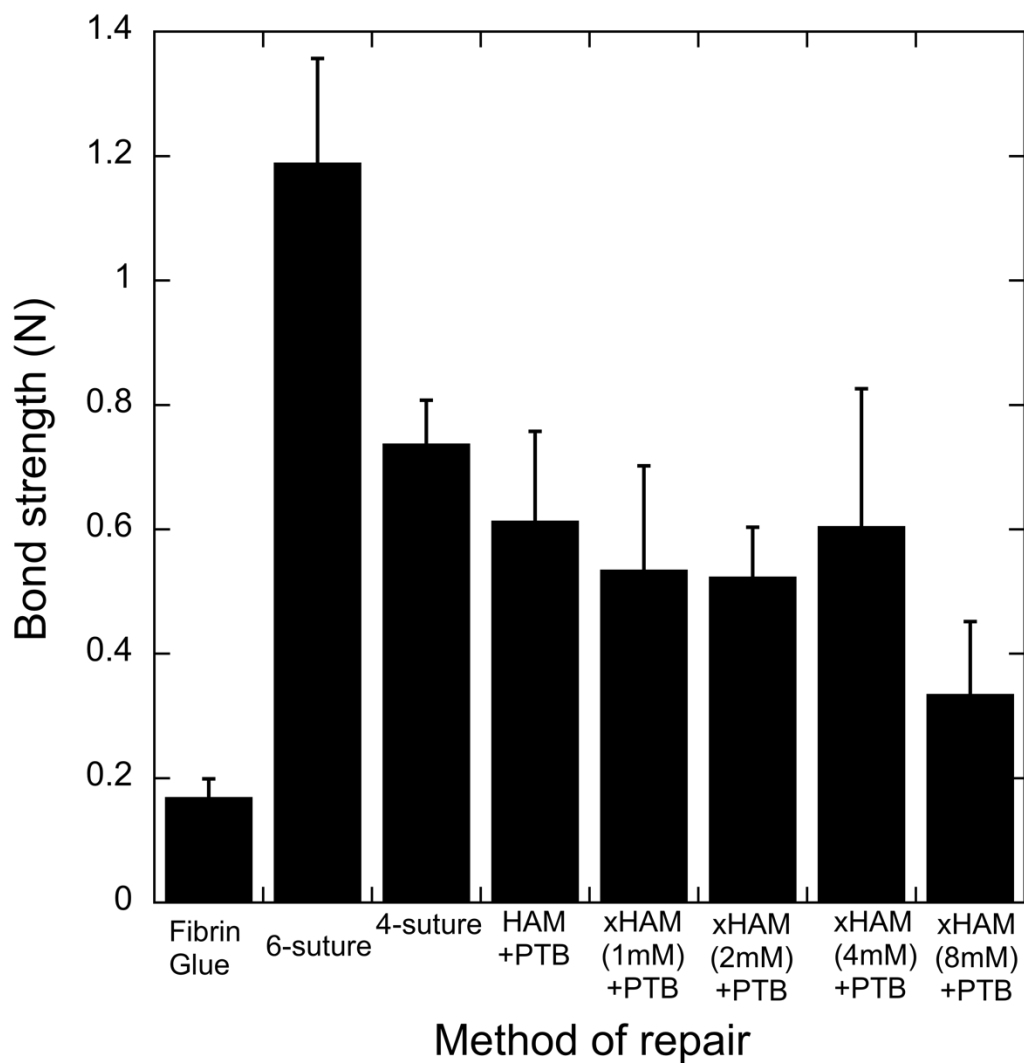


Figure 6: Nerve-amnion bond strength using PTB as a function of fluence (J/cm^2) using 532 nm light delivered at $0.5 W/cm^2$ ($n=5$, $** p<0.5$).

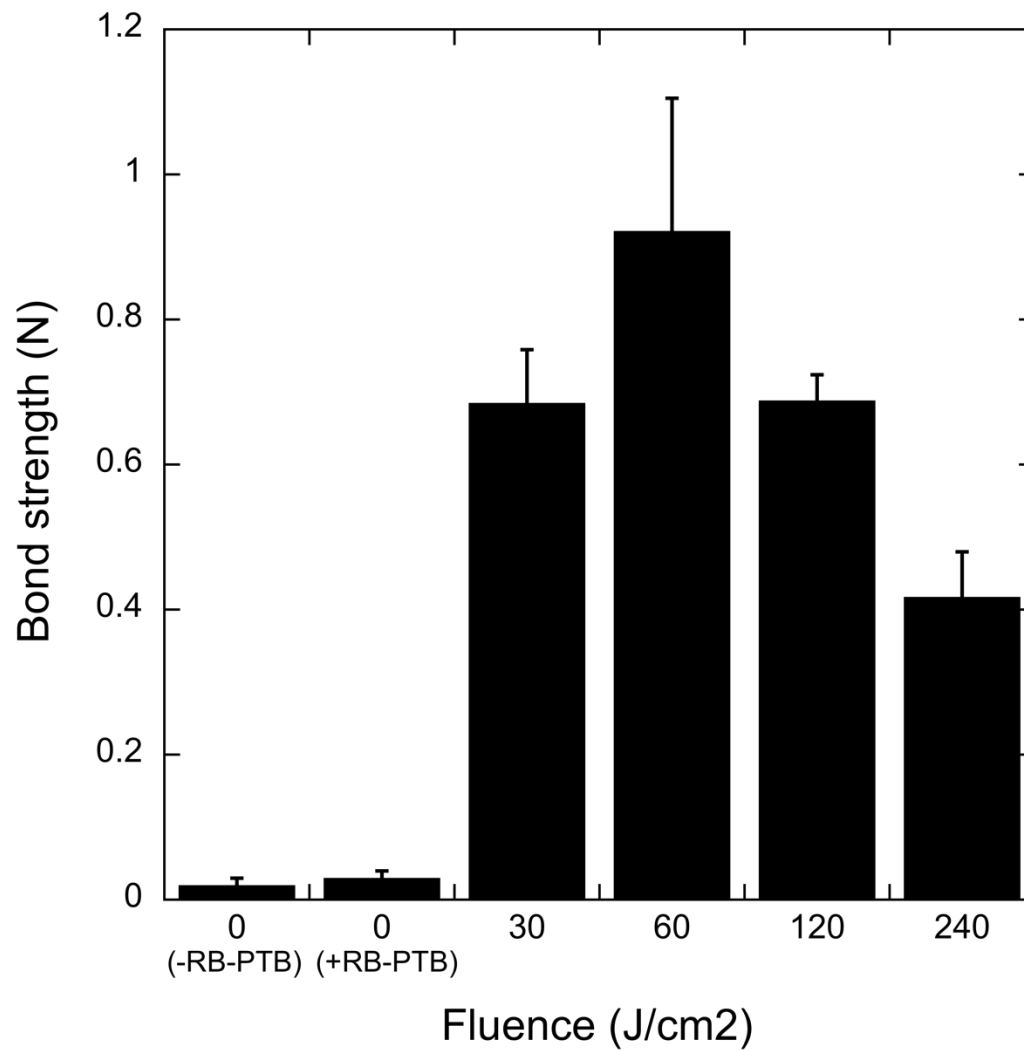


Figure 7: Schematic representation of the anatomy of human amniotic membrane (HAM)

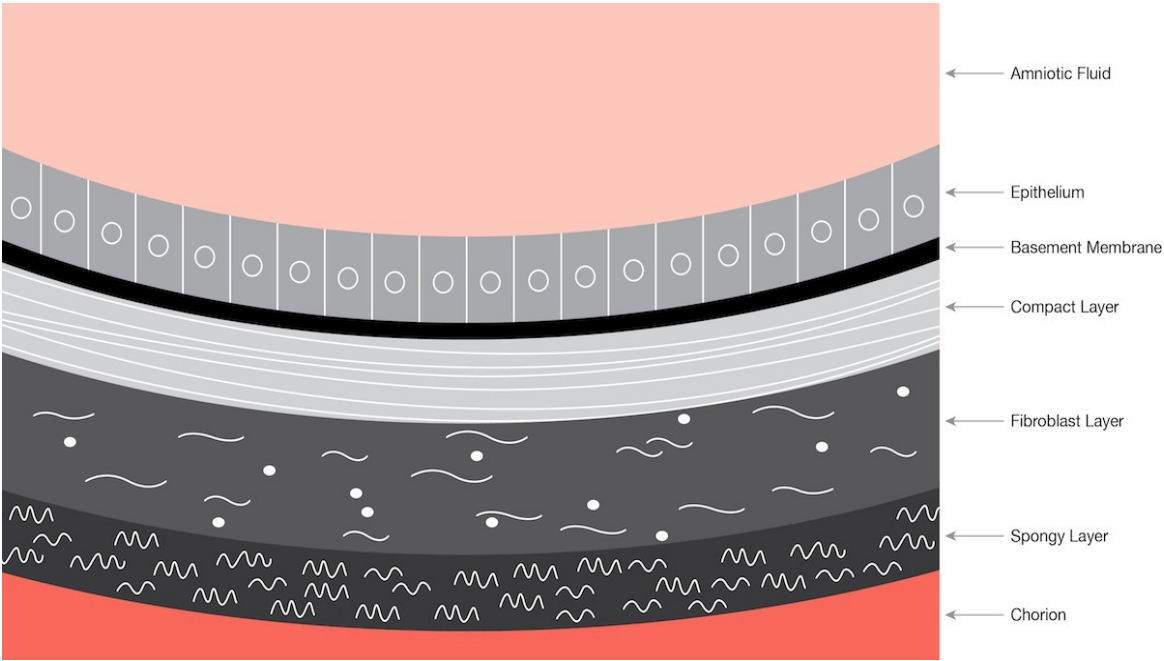


Figure 8: Amnion-nerve bond strength via PTB as a function of which surface was used as interface with nerve using 532 nm light delivered at 0.5 W/cm² and a total fluence of 60 J/cm².

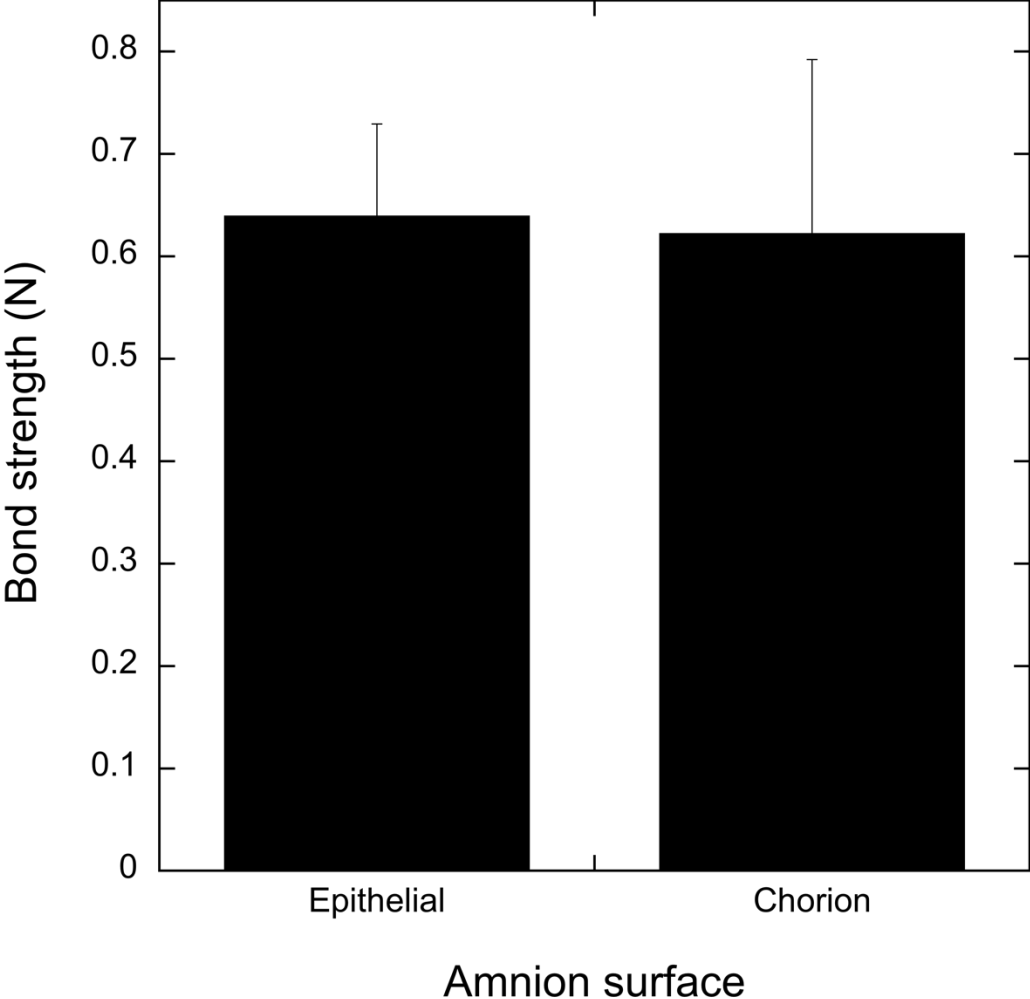


Figure 9: Effect of EDC/NHS crosslinker concentration on Young's Modulus of SIS (n=5, *p<0.5)

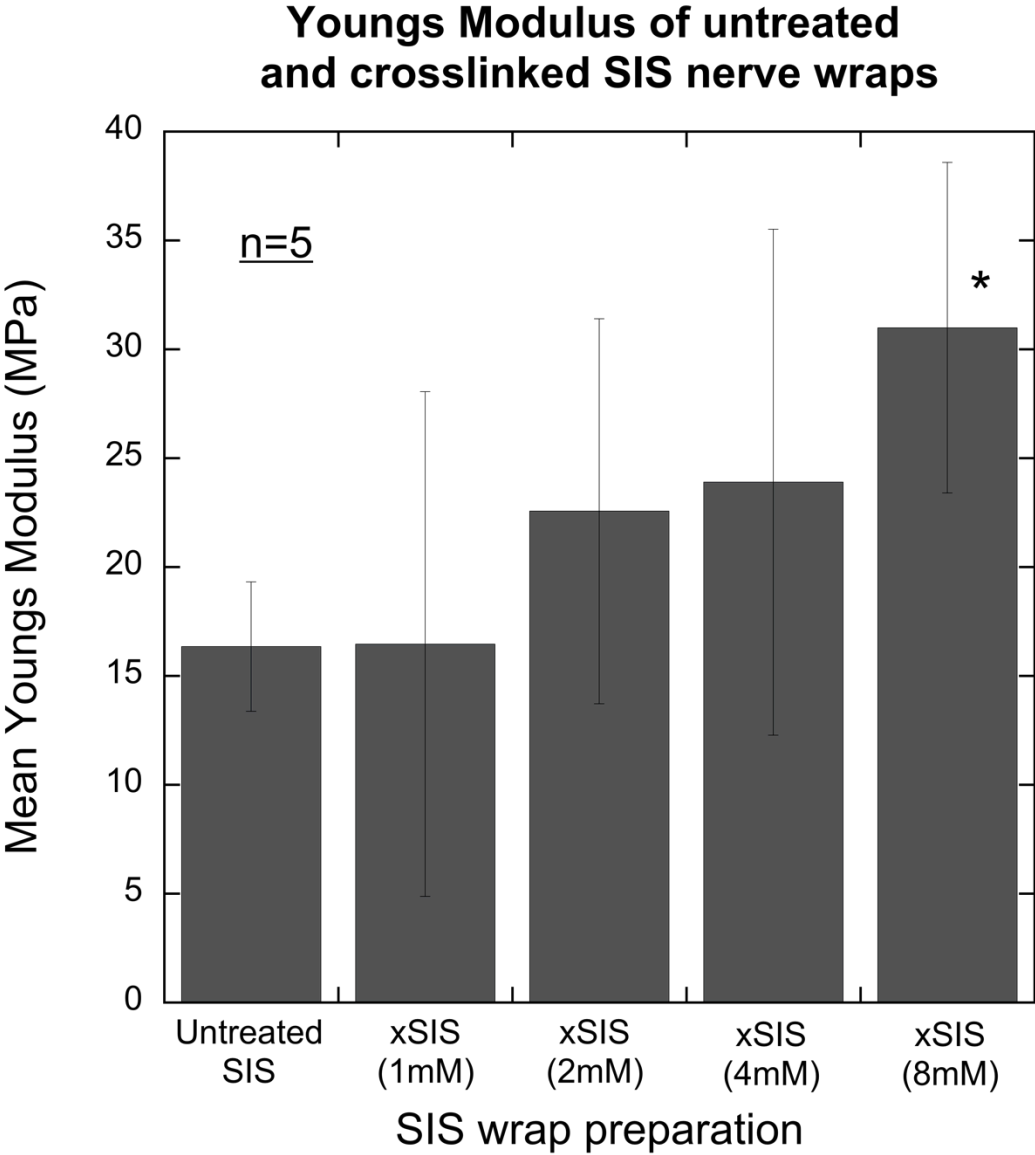


Figure 10: Effect of EDC/NHS crosslinker concentration on Young's Modulus of SIS (n=5, * p<0.5).

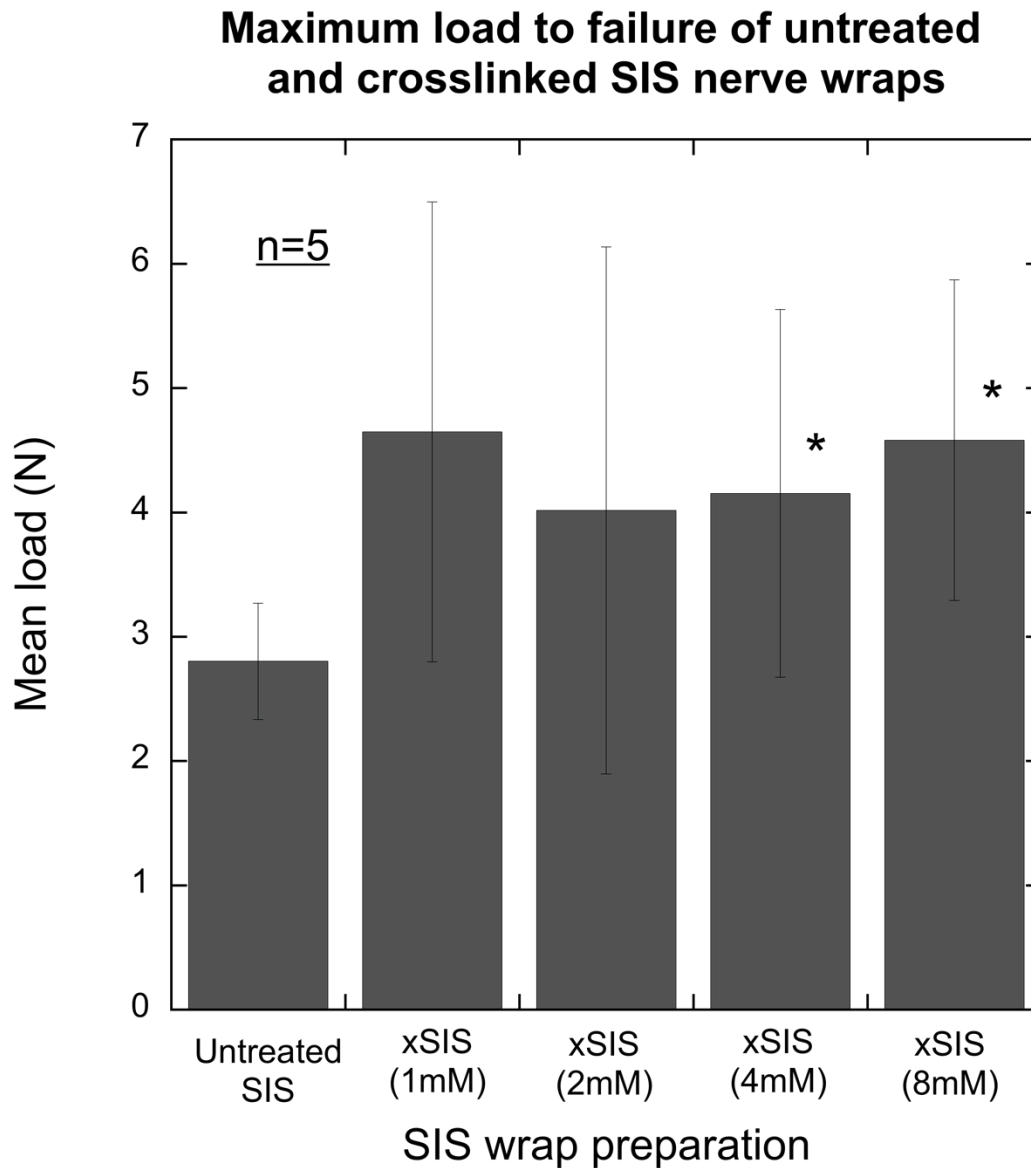


Figure 11: Digestion rates of crosslinked SIS as a function of crosslinker concentration in 0.1% collagenase solution

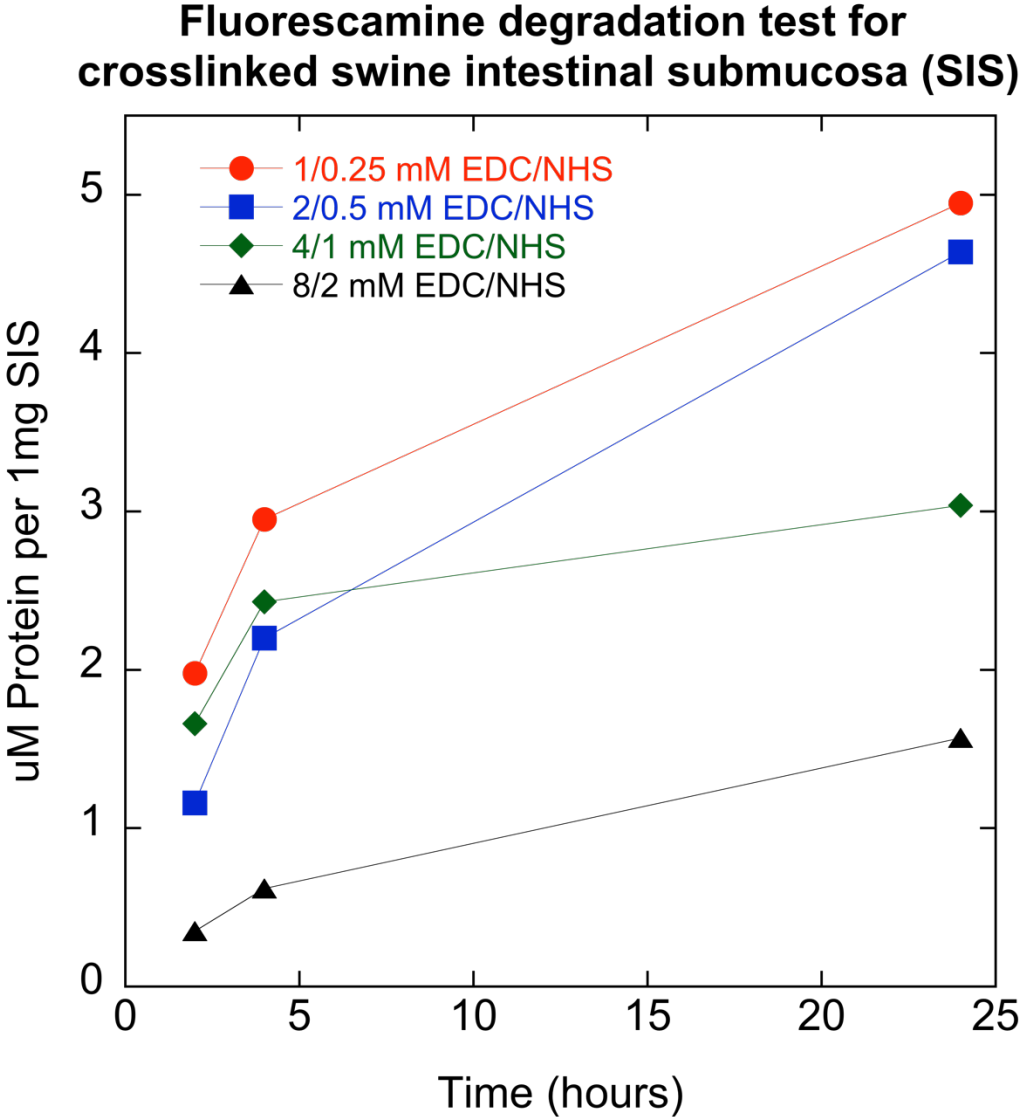


Figure 12:

Bond strength between sciatic nerve and candidate nerve wraps with varying crosslinker concentration

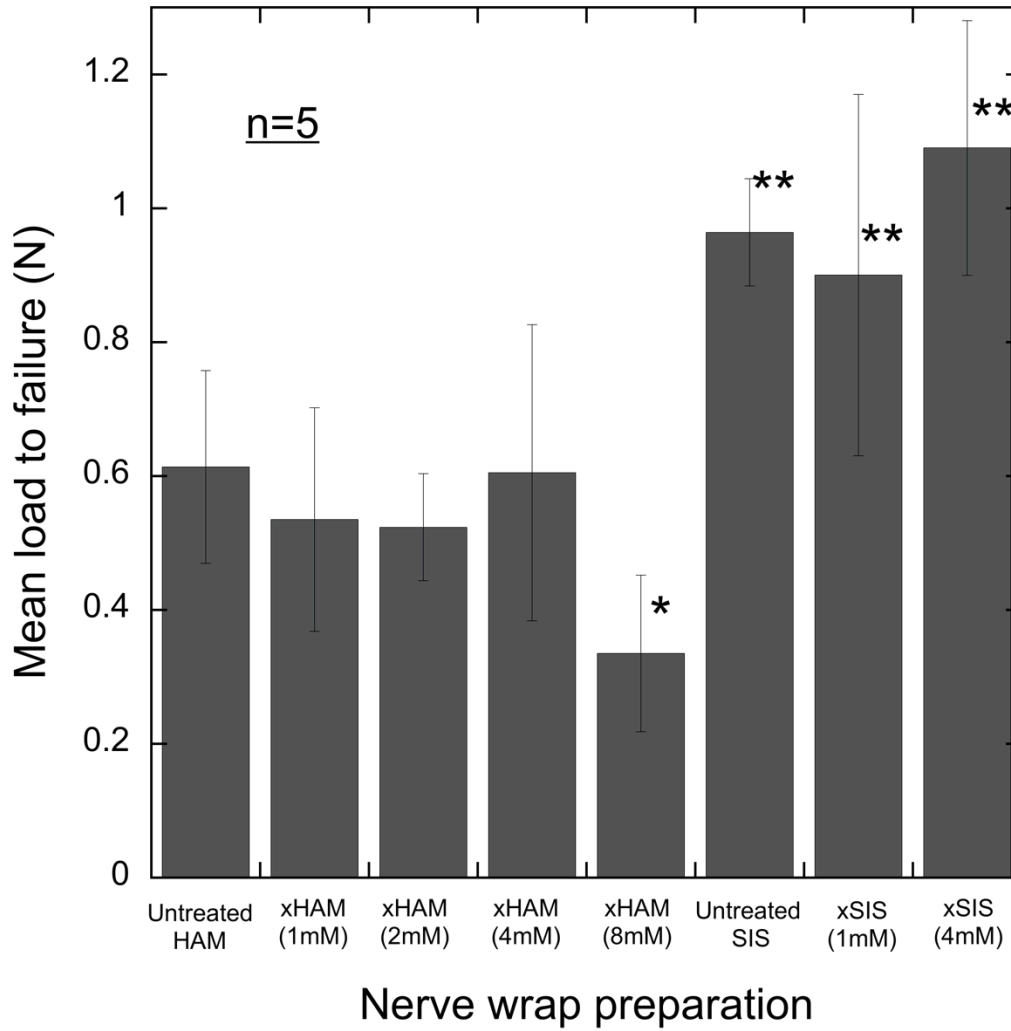


Figure 13: Histology slides from distal nerve sites in each of the 11 treatment groups. Note the almost complete absence of axons in the no repair group. Nerves in all remaining groups successfully regenerated axons distal to the isograft. As shown in Table 3, fiber and axon size in the xHAM+PTB group were significantly greater than standard repair and other treatment groups.

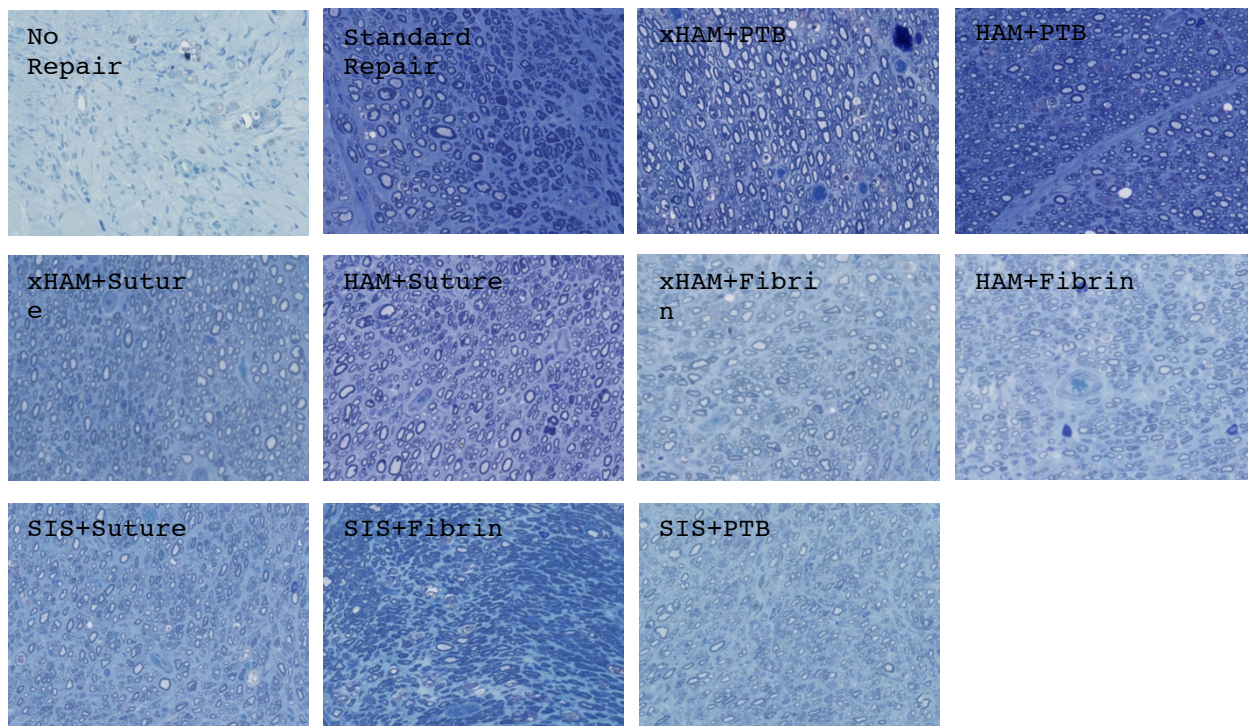


Figure 14: A. Findings following sacrifice in standard repair (positive control) group. Note the extent of fibrinous adhesions around proximal and distal neurorrhaphy sites (arrows). B. Findings following sacrifice in xHAM+PTB group. Note the relative absence of fibrinous adhesions around proximal and distal neurorrhaphy sites

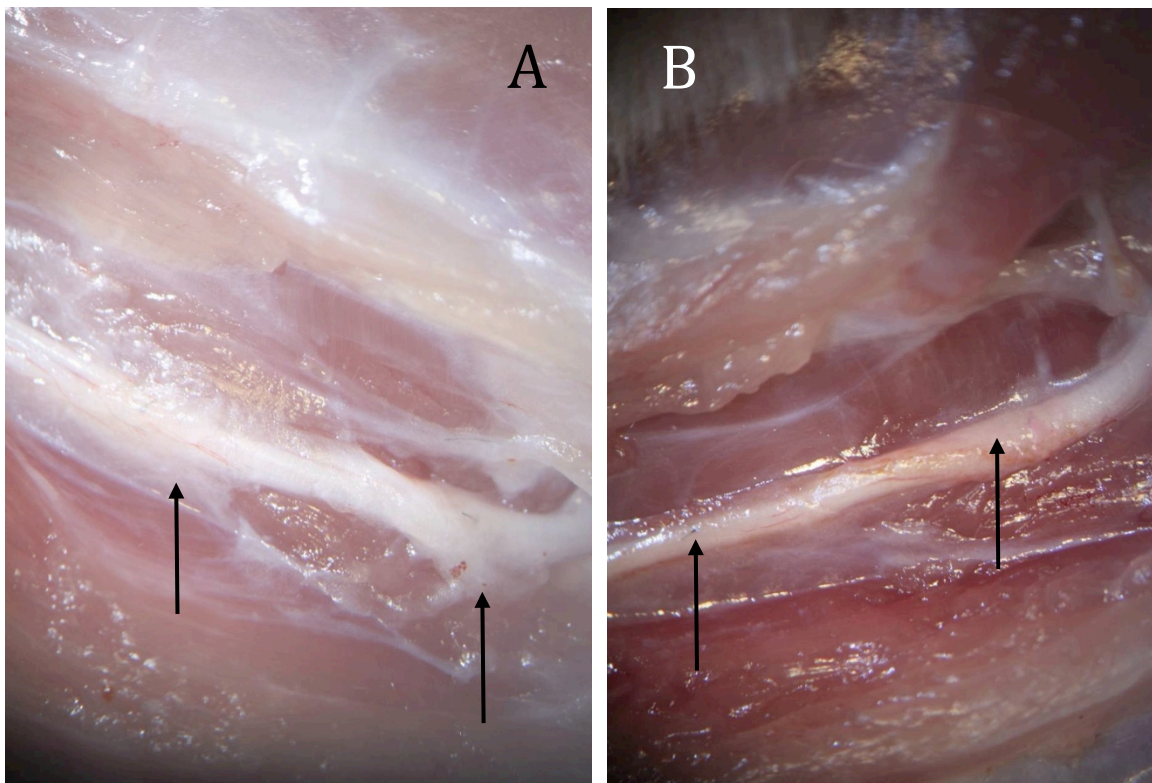


Figure 15: Sciatic Function Index for rat acellular nerve allograft secured with conventional suture vs. rat acellular nerve allograft secured with xHAM+PTB. No significant differences were found between the two groups.

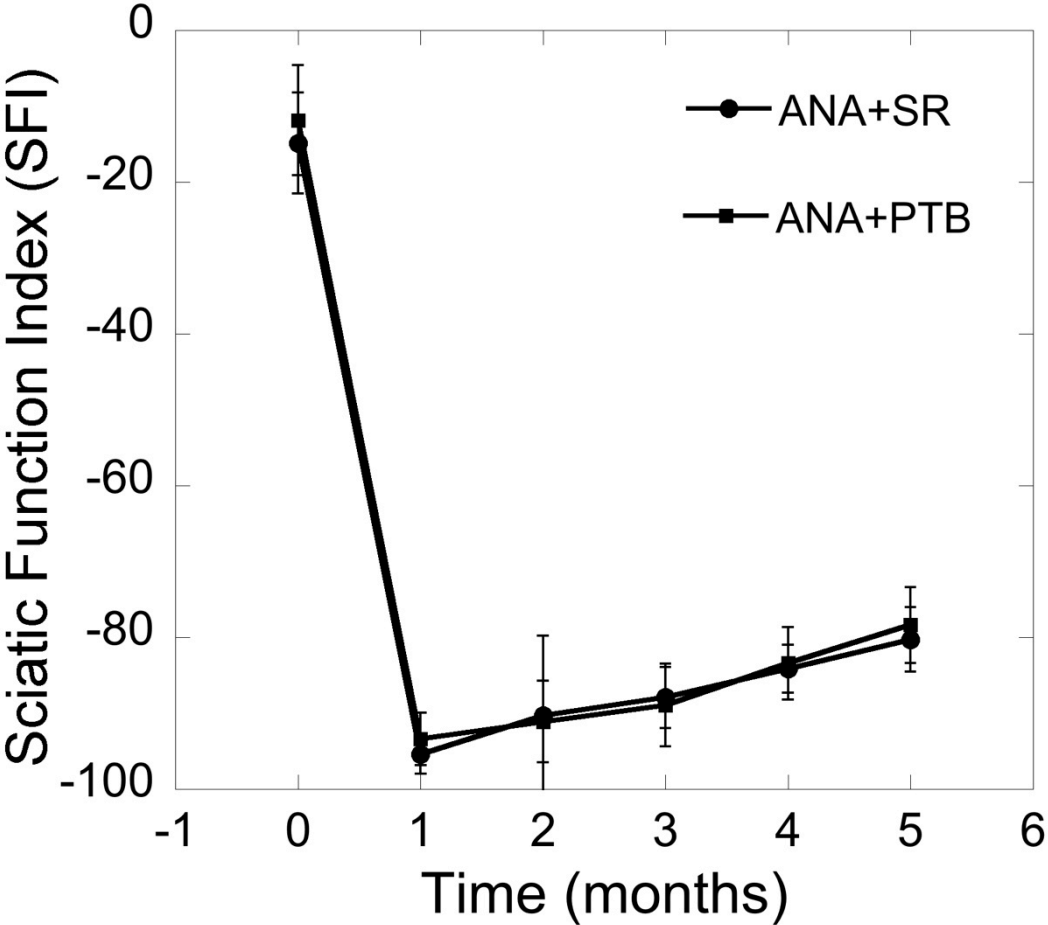


Figure 16: Mean gastrocnemius muscle mass retention. No significant difference in muscle mass retention existed between rat acellular nerve allograft secured with conventional suture vs. rat acellular nerve allograft secured with xHAM+PTB.

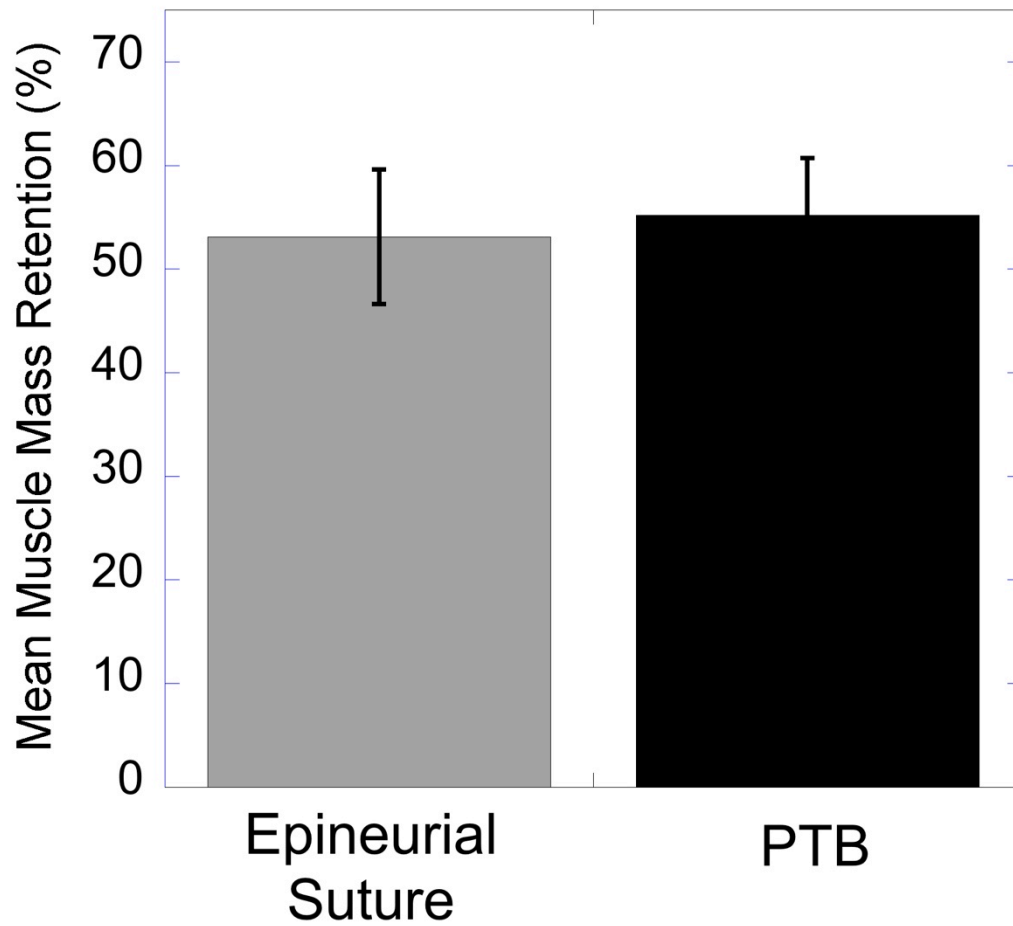
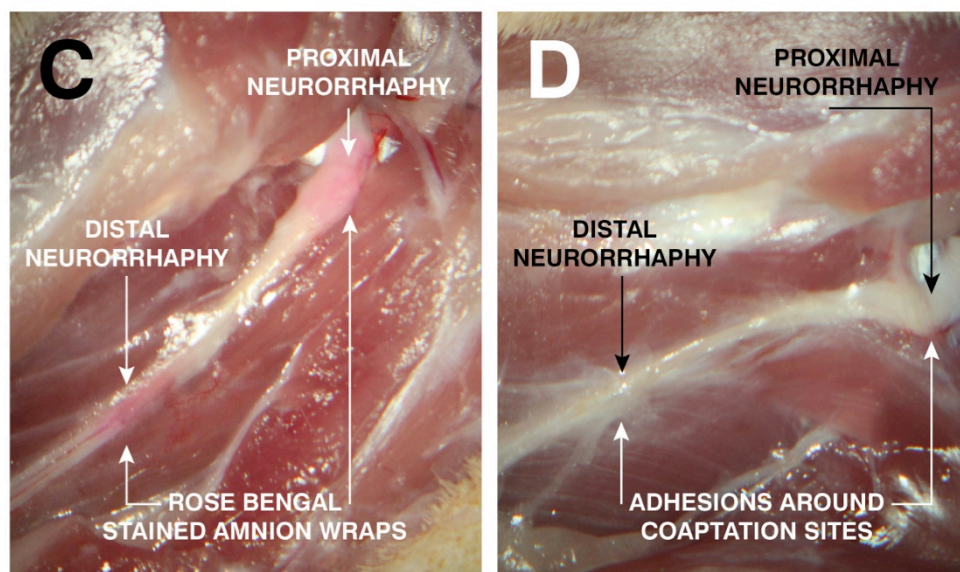


Figure 17. C. Findings following sacrifice in ANA +xHAM+PTB group. Note the relative absence of fibrinous adhesions around proximal and distal neurorrhaphy sites. Photochemically sealed ANA showed remnants of RB stained amnion wraps. D. Findings following sacrifice in ANA/standard repair (positive control) group. Note the extent of fibrinous adhesions around proximal and distal neurorrhaphy sites (arrows).





ELSEVIER



REVIEW

The clinical applications of human amnion in plastic surgery



N.G. Fairbairn*, M.A. Randolph, R.W. Redmond

Division of Plastic Surgery, Massachusetts General Hospital, Harvard Medical School, 15 Parkman Street, WAC 453, Boston, MA 02114, USA

Received 29 May 2013; accepted 23 January 2014

KEYWORDS

Human amnion;
Plastic surgery;
Biological dressing

Summary Since the early 1900s, human amnion has been applied to a wide variety of clinical scenarios including burns, chronic ulcers, dural defects, intra-abdominal adhesions, peritoneal reconstruction, genital reconstruction, hip arthroplasty, tendon repair, nerve repair, microvascular reconstruction, corneal repair, intra-oral reconstruction and reconstruction of the nasal lining and tympanic membrane. Amnion epithelial and mesenchymal cells have been shown to contain a variety of regulatory mediators that result in the promotion of cellular proliferation, differentiation and epithelialisation and the inhibition of fibrosis, immune rejection, inflammation and bacterial invasion. The full repertoire of biological factors that these cells synthesise, store and release and the mechanisms by which these factors exert their beneficial effects are only now being fully appreciated. Although many commercially available biological and synthetic alternatives to amnion exist, ethical, religious, and financial constraints may limit the widespread utilisation of these products. Amnion is widely available, economical and is easy to manipulate, process and store. Although many clinical applications are of historical interest only, amnion offers an alternative source of multi-potent or pluripotent stem cells and therefore may yet have a great deal to offer the plastic surgery and regenerative medicine community. It is the purpose of this article to review the clinical applications of human amnion relevant to plastic surgery.

© 2014 British Association of Plastic, Reconstructive and Aesthetic Surgeons. Published by Elsevier Ltd. All rights reserved.

Introduction and history

Prior to the realisation of its medical and surgical applications, human amnion was the focus of myth and superstition. Being born with the fetal membranes or “caul” intact was considered extremely lucky. Children were gifted with life-long happiness, the ability to see spirits, and protection from death by arms and drowning. The magical powers of

* Corresponding author. Division of Plastic Surgery, Massachusetts General Hospital, Harvard Medical School, 78 Winter Street, Cambridge, Boston, MA 02141, USA. Tel.: +1 617 515 2701.

E-mail address: NGF174@hotmail.com (N.G. Fairbairn).

the caul were not confined to the original bearer and could be transferred by inheritance or legitimate sale. As a result, the trade of caul amulets became extremely popular, particularly between seafaring men during the 1800s at the time of the Napoleonic War.¹

In 1910, Davis reported on early experience with fetal membranes in skin transplantation.² Over the last century, the beneficial effects of amnion have been applied to burns, chronic vascular and diabetic ulcers, dural defects, intra-abdominal adhesions, peritoneal reconstruction, genital reconstruction, hip arthroplasty, tendon repair, nerve repair, microvascular grafts, corneal repair, intra-oral reconstruction and reconstruction of the nasal lining and tympanic membrane. More recently amnion has been shown to be a viable source of stem cells with a potentially exciting future in tissue engineering and regenerative medicine. Although many of these roles are of historical interest only, an awareness of this history is an important pre-requisite for future development and innovation. It is the purpose of this article to review past and present applications of human amnion relevant to plastic surgery and how it may contribute to our future.

Anatomy and physiology

Amnion forms during the transition of the morula into the blastocyst at approximately 7-days following fertilisation.³ Amnion is between 0.02 and 0.05 mm thick and consists of five distinct layers: (1) epithelium, (2) basement membrane, (3) compact layer, (4) fibroblast layer, (5) spongy layer (see Figure 1). The innermost epithelium consists of a single layer of cells in direct contact with amniotic fluid. Microvilli at the apical surface of these cells play an important role in amniotic fluid homeostasis.

The basement membrane border of the cells contains blunt projections that inter-digitate with similar processes in the basement membrane, forming a densely adherent bond. The basement membrane is a thin layer composed of reticular fibers. The compact, fibroblast and spongy layers are referred to as the amniotic mesenchyme and originate

from the primary extra-embryonic mesoderm of the blastocyst. The mesenchyme contains collagen I–VII and non-collagenous proteins such as elastin, laminin, fibronectin and vitronectin. The compact layer is composed of a dense network of fibers and is almost entirely free from cells. Abundant type I, II and III collagen and elastin within this layer endow amnion with tensile strength and elasticity.⁴ These properties help protect the fetus from mechanical stress and desiccation. The fibroblast layer is the thickest layer and is composed of a loose fibroblast network within a matrix of reticulin. The outermost spongy layer represents the transitional layer between amnion and chorion and is composed of bundles of reticulin within a background of mucin. The two layers are loosely adherent, allowing a degree of gliding during gestation and easy separation by blunt dissection during harvest.⁵

In spite of being devoid of vascularity, nerves, muscles and lymphatics, amnion is highly metabolically active.⁵ Oxygen and nutrients are obtained by diffusion from amniotic fluid and chorionic vasculature. The epithelial layer is a source of prostaglandins, particularly prostaglandin-E₂, and is thought to play an important role in the initiation and maintenance of uterine contractions.⁶ The epithelium also contains human chorionic gonadotrophin receptors that regulate prostaglandin production and activity. Epithelial cells manufacture multiple vasoactive peptides, growth factors, cytokines and extracellular matrix (ECM) proteins.⁵ These biological factors may reside in the epithelium or may be transported and accumulated in the mesenchyme where they act as a reservoir from which the amnion exerts its therapeutic effects following transplantation.

Mechanism of therapeutic effect

As a barrier and analgesic

The application of amnion to a wound bed prevents desiccation and excessive fluid loss and provides an analgesic effect by protecting exposed nerve ends from the environment.

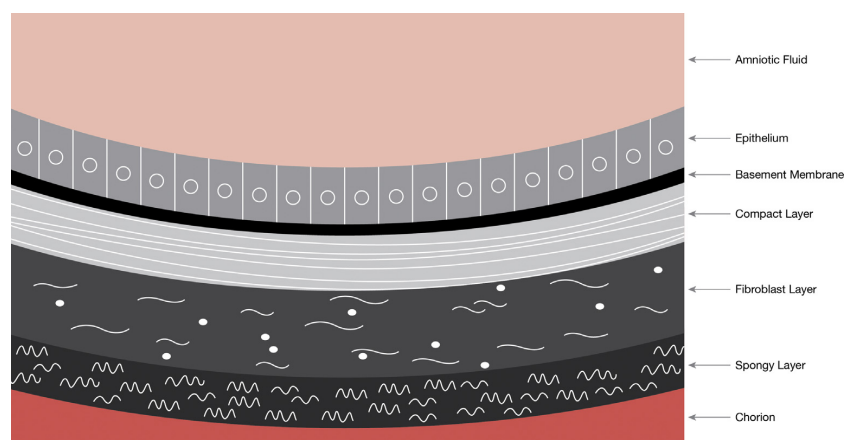


Figure 1 Schematic of amnion structure.

As a non-immunogenic material

Several investigators have concluded that amniotic epithelial and mesenchymal cells lack HLA class A, B, DR and co-stimulatory molecules CD-40, CD-80 and CD-86.⁷ In contrast, others have shown the presence of class-1 and class-1b antigens in epithelial cells, mesenchymal cells and fibroblasts.⁸ Radiobiological studies suggest that although amnion cells retain the ability to synthesise HLA, they do not express HLA-A, B, C or DR antigens of β -2 microglobulin on the cell surface.⁹ Mesenchymal stromal cells may inhibit the maturation of peripheral blood monocytes into antigen-presenting dendritic cells.

As a promoter of epithelialisation and an inhibitor of fibrosis and scar

Amniotic epithelial and mesenchymal cells contain epidermal growth factor (EGF), keratinocyte growth factor (KGF), keratinocyte growth factor receptor (KGFR), hepatocyte growth factor (HGF), and hepatocyte growth factor receptor (HGFR). These growth factors are responsible for proliferation, migration and differentiation of epithelial cells and the promotion of epithelialisation.¹⁰ Basic fibroblast growth factor (bFGF), and transforming growth factor (TGF) β -1, β 2, β 3 have also been demonstrated in amnion cells. bFGF is a pro-angiogenic factor and plays a role in the formation of granulation tissue through the proliferation of fibroblasts. The TGF- β family is responsible for the synthesis and deposition of ECM proteins and the regulation and transformation of fibroblasts into myofibroblasts.¹¹ Mesenchymal hyaluronic acid may inhibit TGF- β and the generation of excessive fibrosis and scar.¹⁰ This may explain the beneficial effect amnion has on scar formation and why fetal wound healing is essentially scarless.

As an anti-inflammatory and anti-bacterial

Amniotic epithelial cells contain interleukin 10 (IL-10) that down-regulates the expression of Th1 cytokines, major histocompatibility complex (MHC) class II antigens and co-stimulatory molecules on macrophages.¹² IL-10 also enhances B-cell survival, proliferation and antibody production and has been shown to inhibit the production of pro-inflammatory cytokines such as interferon- γ , IL-2, IL-3, tumour necrosis factor- α (TNF- α), and granulocyte macrophage colony stimulating factor (GM-CSF). Other anti-inflammatory mediators such as IL-1 receptor antagonist and tissue inhibitors of metalloproteinase-1, 2, 3, 4 (TIMPs) have also been found in amniotic cells.

Amniotic fluid contains lysozymes and immunoglobulins.¹³ In vitro experiments confirm reduced viability of group-A and group-B *Streptococcus*, *Staphylococcus aureus* and *Staphylococcus saprophyticus* in the presence of amnion.¹⁴ Amnion has also been shown to produce human-beta-3-defensin. These antimicrobial peptides are implicated in the resistance of epithelial surfaces to microbial colonisation and have been shown to be upregulated in inflamed amnion.¹⁵ Amnion epithelial cells can be induced to express intercellular adhesion molecule-1 (ICAM-1) by pro-inflammatory cytokines such as tumour necrosis factor- α (TNF- α) and IL-1 β .¹⁶ ICAM-1 has a

role in the attraction and adhesion of leukocytes and may also have a role in signal transduction in pro-inflammatory pathways resulting in the recruitment of inflammatory mediators such as macrophages and granulocytes.¹⁷

As a regulator of angiogenesis

The angiogenic influence of amnion is uncertain. The presence of platelet derived growth factor (PDGF) and vascular endothelial derived growth factor (VEGF) are suggestive of a pro-angiogenic role.¹⁸ bFGF may have an even greater pro-angiogenic influence than PDGF and VEGF. However, a large amount of ophthalmological research contends that it is the ability of amnion to suppress angiogenesis that renders it useful in corneal healing. The expression of tissue inhibitors of metalloproteinase (TIMP-1, 2, 3, 4), thromboplastin-1 and endostatin in amniotic cells supports these claims.¹²

Amnion collection and processing

Elective cesarean section donors undergo rigorous serological screening for human immunodeficiency virus-1/2, Hepatitis B, Hepatitis C, human T-cell lymphotropic virus, syphilis, cytomegalovirus, and tuberculosis.¹⁹ Following delivery, amnion is separated from the placenta by blunt dissection (see Figure 2). Once gross contaminants are removed, amnion is usually de-epithelialised to limit immunogenicity, sterilised to reduce risks of disease transmission, and preserved to improve longevity and convenience for storage. Improvements in processing have focused on preserving membrane architecture and growth factor content in order to optimise therapeutic effect.

De-epithelialisation can be performed by mechanical scraping or exposure to chemicals.¹⁹ It is uncertain how these protocols affect the levels of growth factors and ECM proteins. Koizumi et al. showed that, although amnion denuded of its epithelium contained EGF, TGF- α , KGF, HGF, bFGF, TGF- β 1, and TGF- β 2, protein levels were reduced in comparison to samples with intact epithelium.¹⁰ Whether this is clinically significant is uncertain. Neurotransmitters, neurotrophic factors and neuropeptides are concentrated in the epithelium and therefore amnion with intact epithelium may be

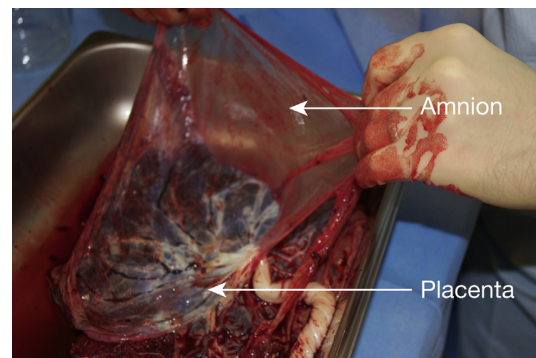


Figure 2 Amnion being bluntly dissected from human placenta.

superior when applied to neural injury.²⁰ In contrast, denuded amnion results in superior cell adhesion, migration and proliferation and therefore may be preferable when applied to acute and chronic wounds.²¹ As the majority of clinical applications concern wound healing, the use of denuded amnion has greater representation in the literature.

Developed in the late 1980s, cryopreservation in glycerol is the most widely used preservation technique. Antibacterials and anti-fungals are often added before freezing at -80°C . Cells are devitalised although not sterilised.²³ Viable bacteria and viruses can be present following several months of storage.²⁴ The effect on biological properties is uncertain. Thomasen et al. reported no detrimental impact on sterility, histological integrity or the availability of biological mediators. Amnion cryopreserved in 50% glycerol/DMEM at -80° for 1-month contained EGF, TGF- α , KGF, HGF, bFGF, TGF- β 1, - β 2, β 3, KGFR and HGFR.²² Cryopreservation requires expensive equipment that may be unavailable for some institutions, particularly in developing nations.

Lyophilisation is an alternative technique allowing storage of amnion at room temperature, obviating the requirement for deep freeze facilities and increasing surgeon convenience. Lyophilised membranes are commonly sterilised with gamma irradiation. Concerns exist regarding detrimental changes to membrane architecture and growth factor levels. Nakamura et al. reported no significant difference in tensile properties, tissue structure or ECM composition between lyophilised, gamma-irradiated and cryopreserved membrane.²⁵ Lim et al. showed that lyophilisation reduced the levels of

several growth factors and ECM proteins although there was no appreciable difference in clinical performance when compared with cryopreserved samples.²⁶ Other methods of preservation and sterilisation exist although these are less well accepted.

The variation in processing within the literature makes it difficult to draw definitive conclusions on the optimal method. Variation also exists amongst commercially available products (Table 1). Independent of processing technique, several donor specific factors can influence the biochemical composition of amniotic membrane. Lopez-Valladares et al. showed that in fresh, cryopreserved and lyophilised amnion, levels of bFGF, HGF, KGF and TGF- β 1 were significantly lower in those membranes of greater chronological and gestational age.²⁷ Velez et al. found significant differences in cytokine profiles between African Americans and Caucasians.²⁸ Membrane architecture and growth factor profile can also vary depending on what area of amnion a specimen originates from.²⁹ As a result, standardisation of collection and processing may be essential if consistent therapeutic results are to be achieved. If consistent relationships between donor variables and biochemical profile exist, it may become possible to select certain varieties of amnion for specific clinical situations.

Applications relevant to plastic surgery

Broadly speaking, amnion has been applied as an alternative biological dressing or has in some way augmented reconstruction. Table 2 provides examples of

Table 1 Commercially available human amnion products.

Manufacturer	Product	Membrane thickness	Indications for use	Processing technique
MiMedx (Marietta, Georgia)	AmnioFix membrane	50–100 microns	Dural reconstruction, spinal surgical barrier	Proprietary Purion process (dehydration and sterilization)
	AmnioFix Wrap	50–100 microns	Tendon and soft tissue inflammatory conditions	As above
	AmnioFix Injectable	N/A	Nerve and tendon repair	As above
	EpiFix	50–100 microns	Chronic and acute partial and full thickness wounds	As above
Bio-Tissue Inc (Miami, Florida)	Prokera corneal bandage	50–100 microns	Corneal erosion, infectious and inflammatory keratitis, herpes, superficial epithelial defects	Proprietary CryoTek process (cryopreservation)
	AmnioGraft	50–100 microns	Chemical burns, Pterygium, corneal defects, leaking glaucoma blebs, Stevens-Johnson syndrome, Strabismus	As above
	AmnioGuard	300–400 microns	Coverage of glaucoma drainage devices	As above
AcelaGraft Cellular Therapeutics (Cedar Knolls, New Jersey)	AcelaGraft	50–100 microns	General wound dressing and ophthalmic wounds	Deoxycholic acid, gel drying, electron beam irradiation

Table 2 Summary of evidence on the applications of human amnion relevant to plastic surgery.

Clinical Scenario	Author	Study design	Application	Amnion Prep	Summary of outcomes
Biological dressing Burns	Lin et al., 1985 ³⁰	OCC (n = 11)	Overlay on autograft	Fresh	Amnion compared to conventional dressings. Good adherence, not rejected by patient, reduced pain, infection, bleeding, number of dressing changes and time to healing in amnion group
	Subrahmanyam, 1995 ³¹	CS (n = 22)	Overlay on micro-skin grafts	Fresh	Epithelialisation within 7–10 days in 16 patients. Superior wound healing due to occlusive, growth promoting effect of amnion
	Sawhney, 1989 ³²	OCC (n = 90)	PT burns	Processed	Amnion vs conventional silver dressings. Superficial, mid-dermal and full thickness burns. Amnion reduced wound exudate, expedited epithelialisation and reduced granulation tissue and scar formation in all groups. In mid-dermal burns, amnion degraded and required regular replacement. Amnion applied to FT burns once eschar separated
	Ramakrishnan et al., 1997 ²³	OCC (n = 350)	PT burns	Processed	Amnion compared to conventional dressings. Amnion had superior adherence, porosity allowing egress of exudate, transparency allowing wound monitoring, reduction in pain, healing times demands on nursing staff and cost
	Branski et al., 2007 ³⁴	P-RCT (n = 102)	PT burns	Processed	Amnion vs topical antimicrobials. Significantly less dressing changes with amnion. Time to healing, length of stay and incidence of hypertrophic scarring were not significantly different between groups
	Singh et al., 2007 ³⁵	OCC (n = 50)	PT burns	Processed	Gamma irradiated amnion compared with glycerol preserved amnion. Radiation sterilised amnion easier to apply than glycerol sterilized. No significant difference in time to healing, infection, scarring
	Fraser et al., 2009 ³⁶	Animal (n = 21)	PT burns	Processed	Symmetrical lower limb deep dermal burns. Amnion vs paraffin gauze. Histopathological and immunohistochemical analysis showed significantly reduced scar tissue formation in amnion group.
	Mostaque et al., 2011 ³⁷	P-RCT (n = 102)	PT burns	Processed	Paediatric burns. Amnion vs silver sulfadiazine dressings. Amnion resulted in: significantly reduced mean hospital stay, dressing changes, mean time to epithelialisation, reduced pain, increased mobility. Patient and surgeon

	Adly et al., 2010 ³⁸	P-RCT (n = 46)	PT + FT burns	Processed	satisfaction high. Amniotic membrane group compared to Tegaderm dressings. Amnion resulted in significantly faster healing, lower rates of infection, lower pain scores and lower levels of electrolyte and albumin loss.
	Mohammadi et al., 2013 ³⁹	P-RCT (n = 38)	Symmetric upper and lower limb burns	Fresh	Right limb autograft + amnion overlay vs left limb autograft + conventional dressing. Graft success assessed after 21 days. Mean graft take in right limbs was significantly higher than left (90% vs 67%)
<i>Acute wounds</i>	Seashore et al., 1975 ⁴⁰	CS (n = 16)	Omphalocele and gastroschisis	Fresh	Fresh amnion compared to porcine xenograft and silastic sheeting; mean time to healing 55 days; amnion superior due to ready availability, reduced bacterial counts, rapid epithelialisation
	Tekin et al., 2007 ⁴¹	CS (n = UNK)	Coverage of exposed viscera	Fresh	Amnion applied as a cover in place of Bogota bag every 48 h; reduction in serosal erosions and adhesions between bowel loops; visceral and abdominal wall oedema reduced
<i>Chronic wounds</i>	Troensegaard-Hansen et al., 1950 ⁴²	CS (n = 7)	Chronic leg ulcers	Processed	Amnion used in case patients compared with 1 control treated with conventional dressings. Chronic ulceration duration 4–15 years. All amnion patients healed within 10 weeks. No healing in control patient. No wound breakdown during follow-up
	Faulk et al., 1980 ¹⁸	CS (n = 15)	Chronic leg ulcers	Fresh	Amnion vs regular dressings. Samples for histology + immunohistochemistry before and after 5-days amnion. New vessel formation + granulation tissue superior in amnion group.
	Ward et al., 1984 ⁴³	CS (n = 28)	Chronic leg ulcers	Fresh	Amnion applied for 5-days after which ulcer was autografted. 50% recurrence at 1-year (defined as ulceration >1 cm)
	Ward et al. 1989 ⁴⁴	CS (n = 27)	Chronic leg ulcers	Various	Healing compared amongst groups treated with fresh, frozen, tissue cultured maintained or lyophilized amnion. No statistically significant difference between groups. Lyophilised judged to be easiest to use and store
	Singh et al., 2004 ⁴⁵	CS (n = 50)	Chronic leg ulcers	Processed	Successful pain relief and healing of ulcers of varying aetiologies
	Gajiwala et al., 2003 ⁴⁶	CS (n = 8)	Pressure sore	Processed	Superficial sores treated with lyophilized, irradiated human amnion. Easy to handle and apply, analgesic, reduction in exudate, accelerated epithelialisation. Complete healing with single application

(continued on next page)

Table 2 (continued)

Clinical Scenario	Author	Study design	Application	Amnion Prep	Summary of outcomes
Reconstruction <i>Dura</i>	Insausti et al., 2010 ⁴⁷	CS (n = 2)	Large post-traumatic wounds	Processed	Amnion applied to large, deep wounds. Accelerated epithelialisation. Up-regulation of c-Jun expression and modification of keratinocyte migration
	Tomita et al., 2012 ⁴⁸	CS (n = 10)	Skull base	Processed	No CSF leakage or adverse outcome directly related to amnion were observed
	Hasegawa et al., 2004 ⁴⁹	CR (n = 1)	Myelomeningocele	Fresh autograft	Autologous onlay graft; no rejection; prevention of infection following wound dehiscence; graft epithelialised; absence of excessive scar tissue formation; rapid, water-tight solution; neurotrophic factors from amnion promoted neural healing
<i>Oral cavity</i>	De Weerd et al., 2013 ⁵⁰	CR (n = 1)	Myelomeningocele	Fresh, autograft	No rejection; absence of excessive scar tissue formation; graft epithelialised; rapid, water-tight solution; neurotrophic factors from amnion promoted neural healing
	Lawson, 1985 ⁵²	CS (n = 12)	Pectoralis major flap oral mucosal lining	Fresh	Amnion provided scaffolding function. Flaps formed granulation tissue and epithelialised twice as fast untreated flaps. Wound contracture reduced.
	Samandari et al., 2004 ⁵⁵	CS (n = 7)	Mandibular vestibuloplasty	Fresh	No infection or graft rejection. Amnion present for 3 weeks and led to rapid granulation tissue formation, mucosalisation and maintenance of post-operative buccal vestibular height. Prosthodontic surgery possible at 1-month
<i>Genitalia</i>	Kothari et al., 2012 ⁵⁶	CS (n = 10)	Mandibular vestibuloplasty	Processed	No infection or graft rejection. Amnion present for 3 weeks and led to rapid granulation tissue formation, mucosalisation and maintenance of post-operative buccal vestibular height. Prosthodontic surgery possible at 1-month
	Tancer et al., 1979 ⁵³	CS (n = 4)	Vaginal reconstruction	Fresh	Amnion applied over obturator; no rejection or infective complications; epithelialisation complete in all 4 cases by 8 weeks
	Ashworth et al., 1986 ⁵⁴	CS (n = 15)	Vaginal reconstruction	Fresh	No rejection; purulent discharge between obturator changes although no overt infection; excellent results in partial or complete vaginal agenesis reconstruction; improvement in vaginal strictures; epithelialisation by 4 weeks
<i>Flap and microvascular</i>	Ozkaya et al., 2012 ⁵⁷	Animal (n = 32)	Random pattern skin flaps	Fresh	Amnion applied to undersurface of flaps; greater survival of treated flaps. Significant reduction in polymorphonuclear leukocyte number; significant increase in capillary

	Gray et al., 1987 ⁵⁸	Animal (n = 120)	Vascular interpositional grafts	Processed	proliferation and density Grafts soft, pliable; no collapse of graft walls; transparent wall helped to prevent back wall suturing; easier to suture than alternative synthetic (PTFE) grafts; No rejection. Patency after 3 months comparable to other synthetic grafts but inferior to control autogenous vein grafts
<i>Tendon and nerve</i>	Ozbuluk et al., 2010 ⁵⁹	Animal (n = 42)	Flexor tendon repair	Fresh	Adhesion formation reduced in amnion treated group compared with untreated control group after 6 weeks follow-up
	Meng et al. ⁶⁰	Animal (n = 36)	Nerve wrap	Processed	Significant improvements in functional recovery and nerve histomorphometric outcomes at early time points; no significant difference after 12 weeks; significantly less perineural scar tissue formation in amnion group
	Henry et al., 2009 ⁶¹	Animal (n = 24)	Nerve wrap	Processed	Photochemical sealing of amnion wrap to neurorrhaphy site resulted in significant improvement of electrophysiological and histomorphometric outcomes and reduction in axonal escape
	O'Neill et al., 2009 ⁶²	Animal (n = 48)	Nerve wrap	Processed	Photochemical sealing of amnion wrap to neurorrhaphy site showed significantly improved functional and histomorphometric outcomes and reduction in extraneural adhesions
	Mohammad et al., 2000 ⁶³	Animal (n = 66)	Nerve conduit	Processed	Amnion conduits vs silicone conduits vs standard autograft over 1 cm defect. Regeneration through amnion conduit comparable to autograft and superior to silicone conduit. Functional recovery statistically better at early time points in amnion group. Amnion degraded by 4-months.
	O'Neill et al., 2009 ⁶⁴	Animal (n = 24)	Nerve conduit	Processed	Amnion conduits secured with either photochemical tissue bonding (PTB) or suture and compared to autograft. Functional outcomes, muscle mass retention and histomorphometry in amnion conduit + PTB comparable to autograft

PT = partial thickness; FT = full thickness; OCC = observational case controlled trial; CS = case series; CR = case report; P-RCT = prospective randomised controlled trial; UNK = unknown.

experimental and clinical evidence supporting therapeutic benefit according to these categories. In spite of the large evidence base, there is a paucity of well-designed, randomised controlled trials testing amnion performance against gold standard alternatives. A list of ongoing human clinical trials, as listed by ClinicalTrials.gov, is provided in [Table 3](#). The following discussion makes reference to this evidence and aims to provide a more cohesive understanding. A summary of the advantages and disadvantages of amnion according to application are listed in [Table 4](#). The applications amnion has for tissue engineering and regenerative medicine is also discussed.

Biological wound dressing

Burns

The history of amnion use for the management of burns is extensive. The use of amnion for corneal burns and other ophthalmological epithelial defects is commonplace and has led to the development of several commercially available products ([Table 1](#)). Membranes have been used as overlay following standard autografting and microskin grafting and also in place of conventional dressings following superficial and mid-dermal burns, including cadaveric allograft and porcine xenograft.^{30–39}

Acute wounds

Amnion has been used as an alternative temporary biological dressing to protect exposed viscera in cases of congenital abdominal wall defects such as omphalocele and gastroschisis and also full thickness defects secondary to

major trauma, infection or oncological resection.⁴⁰ Amnion provides an alternative to the Bogota bag and can form part of a staged abdominal wall reconstruction with or without negative pressure therapy (NPT).⁴¹

Chronic wounds

Chronic wounds represent a major financial burden on healthcare services worldwide. Multiple studies have reported superior wound healing following the application of amnion to chronic leg ulcers of varying aetiology.^{18,42–45} Areas of pressure necrosis have also been treated although the suitability of amnion in these complex and often extensive wounds is most likely limited to only the most early and superficial cases.⁴⁶ Amnion has also been applied to areas of stalled healing following large traumatic soft tissue loss in patients unfit for complex reconstruction.⁴⁷ In each of these situations, amnion can be applied in conjunction with NPT.

Reconstruction

Dural repair

Amnion has been used to reconstruct dural defects in the skull base and in cases of myelomeningocele.^{48–50} Water tight closure in these situations is essential in order to prevent CSF leak and potentially life threatening infection. Although synthetic materials are available in these situations, autologous solutions are preferred. In congenital anomalies such as myelomeningocele, amnion can be applied as an autograft immediately or as a delayed procedure following storage. When soft tissue defects are large, amnion can form part of a layered closure under loco-regional or free tissue transfer.^{49,50} Amnion may

Table 3 Summary of registered clinical trials using human amnion.

Description	Design	Trial identifier	Institution	Status
The treatment of partial thickness burns: treated amnion versus currently in use topical medication	Phase 2/3 RCT	NCT00674999	University of Texas, Galveston	Recruiting
Evaluation of the cryopreserved amniotic membranes in the care of resistant vascular ulcers	Phase 2 single group interventional	NCT00820274	University Hospital, Limoges Etablissement Français du Sang	Recruiting
An Evaluation of the Effect of the AmnioFix™ Amniotic Membrane Allograft on Scar Tissue and Adhesions in Patients Undergoing Posterior Instrumentation Removal	Phase 2/3 Observational case controlled	NCT01357187	UNKNOWN	Not yet recruiting
The role of AMT (amniotic membrane transplantation) in treating epithelial defects and spherophoron, preventing corneal opacification, decreasing pain, improving visual acuity and treating acute chemical burns	Phase 2/3 RCT	NCT00370812	Shaheed Beheshti Medical University	Recruiting

RCT = Randomised controlled trial.

Table 4 Advantages and disadvantages of human amnion for different clinical application.

Amniotic membrane for clinical application		
Application	Advantages	Disadvantages
General		
	Abundant supply; no donor morbidity; inexpensive; easy processing and storage; off-the-shelf availability; high tensile strength; non-immunogenic; anti-bacterial, anti-inflammatory, anti-fibrotic, regulator of angiogenesis; reduced social, cultural, religious obstacles compared with allograft and xenograft products	Donor screening; risk of disease transmission; optimal processing method uncertain; problematic handling and suturing; variable biological properties depending on sample location, donor and gestational age and race; proteolytic degradation
Biological dressing		
Burns	Readily adherent; transparent allowing wound monitoring; reduction in exudate and infection; accelerated epithelialisation; analgesic; reduction in dressing changes, analgesia requirements, demand on nursing staff; reduced cost; reduced scarring	Handling and suturing may be difficult; Membrane architecture and growth factor content varies with location, gestational age, donor age and race; Degradation may require reapplication; no adherence in full thickness burns
Acute wounds	As above; allows temporary coverage of exposed abdominal viscera; autograft possible in omphalocele and gastroschiasis;	As above
Chronic wounds	Conformable to deep, irregular wounds; permeable allowing egress of exudates; reduced requirement for autografting, time to autografting and graft failure; reduced scarring; use with NPT	As above
Reconstructive		
Dural repair	Water tight barrier; neurotrophic factors support neural tissue; autografting possible (myelomeningocele)	As above
Oral cavity and vaginal vault	Rapid epithelialisation removing need for autograft; bone coverage possible	As above
Flap and microvascular	Angiogenic effect; inhibits neutrophils and free radicals; allows manufacture of vascular grafts; amnion scaffold permits growth factor and stem cell seeding	As above; degradation may interfere with vascular graft success
Nerve and tendon	Wrap reduces scar tissue/adhesions; prevents leakage of growth factors; provides neurotrophic support; allows manufacture of conduits; amnion scaffold permits growth factor and stem cell seeding	As above; empty nerve conduits limited to short distances; degradation may interfere with conduit support

support underlying neurological tissue through the production of neurotrophic factors such as nerve growth factor (NGF), brain derived neurotrophic factor (BDNF) and brain natriuretic peptide (BNP).^{14,51}

Mucosal lining

Amnion has been used to expedite epithelialisation of muscle flaps following intra-oral and vaginal vault reconstruction.^{52–54} In the absence of a cutaneous paddle, these surfaces can be reconstructed with split and full thickness skin grafts, buccal mucosa grafts, intestinal mucosa, peritoneum or commercially available collagen based products. Skin grafts are perhaps the most widely practiced technique although they are associated with complications such as donor site wound, colour and texture mismatches, dryness, desquamation, hair growth, poor mobility and contracture. Commercial products are expensive and require complex processing that can reduce their clinical efficacy. Amnion has also been used to successfully cover bone following excision of gingival leukoplakia and vestibuloplasty.^{55,56}

Flap and microvascular

Partial or complete flap necrosis is a dreaded complication when performing tissue transfer. Amnion applied to the undersurface of random pattern skin flaps has been shown to significantly increase capillary proliferation, reduce infiltrating neutrophils and improve flap survival.⁵⁷ Local factors liberated by amnion may reduce leukocyte activation and free radical formation, limiting endothelial injury, thrombosis and flap necrosis.

The survival of free tissue transfer relies on successful microvascular anastomosis. In major trauma or complex elective reconstruction, this can require interpositional grafting. Autogenous vein remains the gold standard graft material although the associated donor morbidity is unsavoury. Alternative biological or synthetic materials with equivalent patency and functional outcomes are desirable. Amnion has been rolled into interpositional grafts and, in a rat model, resulted in re-endothelialisation at equal time points compared with vein autografts.⁵⁸

Tendon and nerve

Successful functional recovery, particularly in the hand and upper limb, is dependent on accurate reconstruction of tendons and nerves. Wrapping tenorrhaphy and neuroorrhaphy sites with amnion can reduce adhesions and improve functional recovery.^{59,60} Reducing suture burden at the repair site can also reduce scar tissue formation. Amnion wraps have been sealed around neuroorrhaphy sites with a novel, sutureless, photochemical tissue bonding technique, resulting in improved functional and histological outcomes.^{61,62} Amnion scaffolds may act as a reservoir of neurotrophic factors. Sealing the regenerative milieu may prevent the elution of these and other endogenous neuroregenerative factors into the surrounding tissue.

As with vessel injury, loss of nerve tissue may require bridging techniques. Autogenous nerve grafts remain the gold standard technique although in situations of major trauma, demand for autogenous material can exceed supply. Processed allografts and biological and synthetic nerve conduits are options although all are associated with limitations. Amnion provides an alternative material for the construction of nerve conduits.^{63,64} As with any hollow conduit, applications remain limited to short deficits.

Tissue engineering and regenerative medicine

Amnion as a scaffold

Biological scaffolds require the presence of extracellular matrix proteins such as collagen, laminin and fibronectin. Adhesion molecules specific to these proteins facilitate cell adhesion, transmembrane receptor activation and intracellular signalling cascades that regulate cell migration, proliferation, differentiation and apoptosis.⁶⁵ Ideal scaffolds are biocompatible, mechanically stable, flexible, resorbable at a rate consistent with repair and allow the incorporation of growth factors and genetic materials.⁶⁶ Amnion basement membrane contains collagen III, IV and other glycoproteins such as laminin and fibronectin. Amnion scaffolds have been used to cultivate epithelial cells *in vitro* before *in vivo* transplantation. This has been used to reconstruct corneal surfaces following chemical burns, limbal stem cell deficiency and other related pathology.⁶⁷ Amnion scaffolds seeded with human keratinocytes have generated living skin equivalents and have been successfully transplanted into an animal model.⁶⁸ Denuded amnion has been used as a carrier matrix for chondrocytes and cartilage regeneration.⁶⁹ Amnion seeded with human umbilical vein endothelial cells and human vascular smooth muscle cells has been rolled into a cell dense, mechanically stable, multi-layered blood vessel conduit.⁷⁰ Although growth factor levels in denuded amnion may be reduced, several studies have suggested scaffolding function is more effective in the absence of epithelium. Due to the interference of hemidesmosome formation, amniotic epithelium may hinder uniform cell expansion.⁷¹

Amnion as an alternative source of stem cells

The use of pluripotent embryonic stem cells (ESCs) is hindered by ethical controversy. Mesenchymal stem cells (MSCs) are a less controversial, non-embryonic source of multipotent cells. Bone marrow mesenchymal stem cells

(BM-MSCs) are perhaps the gold standard adult multipotent cell. However, due to the invasive and painful nature of harvest, alternatives such as adipose derived mesenchymal stem cells (AD-MSCs) have become popular. Adipose tissue is abundant, readily accessible with low morbidity, provides cell numbers and stem cell fractions that greatly exceed that of BM-MSCs, and have superior proliferation capacity and differentiation potential *in vitro*.⁷² Adipose derived stem cells can also be induced into pluripotent cells. These cells are reprogrammed into pluripotency by inducing the expression of transcription factors characteristic of undifferentiated embryonic stem cells.⁷³

Several limitations of AD-MSCs exist. Cell populations are not homogenous. Considerable variations in phenotype, proliferative capacity and differentiation potential exist between and within individuals. Proliferative capacity and differentiation potential may decrease with donor age, a characteristic shared by all adult derived MSCs.⁷⁴ The secretion of tumour promoting factors such as IL-6 and the pro-angiogenic effect of these cells have also raised concerns regarding malignant transformation.⁷⁵ With regards to induced pluripotency, the persistence of source cell epigenetic memory may render these cells unstable and unpredictable.⁷⁶

Amnion has advantages over all adult derived MSCs. Amnion supply is unlimited and is arguably more convenient to obtain than adipose tissue. Total cell number and stem cell fraction from amnion is thought to greatly exceed both BM-MSCs and AD-MSCs.⁷² In addition to amnion, placental tissue provides chorionic membrane, chorionic villi, maternal decidua, umbilical cord, umbilical cord blood and Whartons jelly. These provide additional MSCs and also embryonic populations such as endothelial and haematopoietic stem cells.⁷⁷ Proliferative capacity and differentiation potential of amnion derived cells is thought to exceed that of adipose tissue. Derivatives from all three germ layers such as adipogenic, osteogenic, chondrogenic, hepatic, pancreatic, cardiac, vascular and neural cells have been cultured and shown to possess reparative and functional capabilities. Placental cells of fetal origin (amnion, chorion, chorionic villi) may have greater differentiation potential than those of maternal origin (decidua).⁷⁸ Fetal origins may also prevent age related reductions in proliferative and differentiation potential characteristic of adult cells. Due to a maximum gestational age of 9–10 months, it is also likely that amnion provides a population of cells that have accumulated less genetic damage than adult sources.

It is currently uncertain whether amnion cells are truly pluripotent or whether multiple sub-populations of multipotent stem cells exist. The existence of multiple sub-populations is potentially problematic. Not unlike growth factor level, the proliferative and differentiation characteristics of these cells may vary according to membrane location, gestational and donor age, race and processing technique. In addition, different methods of culture, isolation and expansion may artificially select certain sub-populations and obscure true biological activity. Pluripotency is supported by the identification in amniotic cells of multiple molecular markers typically found on embryonic stem cells, such as octamer-4 (OCT-4), NANOG, sex-determining Y-box-2 (SOX-2), Lefty-A, FGF-4, REX-1 and

Table 5 Comparison between amnion, adipose tissue and bone marrow as alternative sources of mesenchymal stem cells.

Variable	Tissue		
	Amnion	Adipose	Bone marrow
Invasiveness of procurement	–	+	++
Ethical issues	–	–	–
Availability of tissue	+++	++	+
Stem cell fraction (% of total cells)	5–50	1–5	0.01–0.05
Proliferation capacity in vitro	+++	++	+
Senescence with passage	+	++	+++
Differentiation potential	+++	++	+
Reduction in differentiation potential with donor age	–	+	+
Age and environmental acquired DNA damage	–	+	+
Cryogenic storage of cells for future use following birth	+	–	–

teratocarcinoma derived growth factor-1 (TDGF-1).⁷⁹ OCT-4 is responsible for the maintenance of pluripotency and it has been shown that the level of this marker decreases with increasing cellular differentiation. Embryonic stem cells are derived from the inner cell mass of the blastocyst, which in turn gives rise to the epiblast. The epiblast, from which the amnion is derived, gives rise to all 3 germ cell layers. It is therefore possible that amniotic cells retain epiblastic pluripotency. In addition, gastrulation plays an important role in the differentiation and determination of cell fate. Amnion forms prior to this phase and it is therefore possible that these cells are pluripotent.⁸⁰ Table 5 provides a comparison of the salient characteristics of bone marrow, adipose tissue and amnion as sources of stem cells.

Conclusion

Human amnion provides the plastic surgeon with an incredibly versatile material. It is economical, widely available, easy to harvest and store and has no ethical constraints. Amnion contains a plethora of biological mediators and is a well-established alternative wound dressing. It is biocompatible, highly conformable, thin, and yet retains considerable tensile strength. Amnion can mechanically support and improve survival of transferred tissue and, through the manufacture of vessel and nerve conduits, may also directly contribute to neurovascular reconstruction. Amnion has provided a vehicle for the development of a novel photochemical tissue bonding technique that has proved efficacious for the sutureless repair of skin, tendon, nerve and vessel. Amnion may prove useful as a biological scaffold for tissue engineering and is emerging as an alternative source of multipotent and even pluripotent stem cells. After more than a century of clinical

use, the application of human amnion in plastic and reconstructive surgery continues to evolve.

Conflict of interest

None.

Funding

None.

References

- Richardson R. Copperfields caul. *Lancet* 2002;**359**:2209.
- Davis JW. Skin transplantation with a review of 550 cases at the Johns Hopkins Hospital. *Johns Hopkins Med J* 1910;**15**:307–96.
- Trelford JD, Anderson DG, Hanson FW, Mendel V, Sawyer RH. Amnion autografts and allografts as a cover for skin defects in sheep. A preliminary report. *J Med* 1972;**3**:81–7.
- Mamaede AC, Carvalho MJ, Abrantes AM, Laranjo M, Maia CJ, Botelho MF. Amniotic membrane: from structure and functions to clinical applications. *Cell Tissue Res* 2012;**349**:447–58.
- Bourne GL. The microscopic anatomy of the human amnion and chorion. *Am J Obstet Gynecol* 1960;**79**:1070–3.
- Okazaki T, Casey ML, Okita JR, MacDonald PC, Johnston JM. Initiation of human parturition. XII. Biosynthesis and metabolism of prostaglandins in human fetal membranes and uterine decidua. *Am J Obstet Gynecol* 1981;**139**:373–81.
- Bilic G, Zeisberger SM, Mallik AS, Zimmermann R, Zisch AH. Comparative characterization of cultured human term amnion epithelial and mesenchymal stromal cells for application in cell therapy. *Cell Transpl* 2008;**17**:955–68.
- Hammer A, Hutter H, Blaschitz A, et al. Amnion epithelial cells, in contrast to trophoblast cells, express all classical HLA class I molecules together with HLA-G. *Am J Reprod Immunol* 1997;**37**:161–71.
- Adinolfi M, Akle CA, McColl I, et al. Expression of HLA antigens, beta 2-microglobulin and enzymes by human amniotic epithelial cells. *Nature* 1982;**295**:325–7.
- Koizumi NJ, Inatomi TJ, Sotozono CJ, Fullwood NJ, Quantock AJ, Kinoshita S. Growth factor mRNA and protein in preserved human amniotic membrane. *Curr Eye Res* 2000;**20**:173–7.
- Sporn MB, Roberts AB, Wakersfield LM, de Crombrughe B. Some recent advances in the chemistry and biology of transforming growth factor-beta. *J Cell Biol* 1987;**105**:1039–45.
- Hao Y, Ma DH, Hwang DG, Hwang DG, Kim WS, Zhang F. Identification of antiangiogenic and antiinflammatory proteins in human amniotic membrane. *Cornea* 2000;**19**:348–52.
- Thadepalli H, Bach VT, Davidson EC. Antimicrobial effect of amniotic fluid. *Obstet Gynecol* 1978;**52**:198–204.
- Kjaergaard N, Hein M, Hyttel L, et al. Antibacterial properties of human amnion and chorion in vitro. *Eur J Obstet Gynecol Reprod Biol* 2001;**94**:224–9.
- Buhimschi IA, Jabr M, Buhimschi CS, et al. The novel antimicrobial peptide beta3-defensin is produced by the amnion: a possible role of the fetal membranes in innate immunity of the amniotic cavity. *Am J Obstet Gynecol* 2004;**191**:1678–87.
- Marvin KW, Hansen WR, Miller HC, Eykholt RL, Mitchell MD. Amnion-derived cell express intercellular adhesion molecule-1: regulation by cytokines. *J Mol Endocrinol* 1999;**22**:193–205.

17. Etienne-Manneville S, Chaverot N, Strosberg AD, Couraud PO. ICAM-1-coupled signaling pathways in astrocytes converge to cyclic AMP response element-binding protein phosphorylation and TNF-alpha secretion. *J Immunol* 1999;163:668–74.
18. Faulk WP, Matthews R, Stevens PJ, Bennett JP, Burgos H, Hsi BL. Human amnion as an adjunct in wound healing. *Lancet* 1980;1:1156–8.
19. Riau AK, Beurman RW, Lim LS, Mehta JS. Preservation, sterilization and de-epithelialisation of human amniotic membrane for use in ocular surface reconstruction. *Biomaterials* 2010;31:216–25.
20. Sakuragawa N, Elwan MA, Uchida S, Fujii T, Kawashima K. Non-neuronal neurotransmitters and neurotrophic factors in amniotic epithelial cells: expression and function in humans and monkey. *Jpn J Pharmacol* 2001;85:20–3.
21. Koizumi N, Rigby H, Fullwood NJ, et al. Comparison of intact and denuded amniotic membrane as a substrate for cell-suspension culture of human limbal epithelial cells. *Graefes Arch Clin Exp Ophthalmol* 2007;245:123–34.
22. Hennerbichler S, Reichl B, Pleiner D, Gabriel C, Eibl J, Redi H. The influence of various storage conditions on cell viability in amniotic membrane. *Cell Tissue Bank* 2007;81–8.
23. Van Baare J, Cameron PU, Vardaxis N, et al. Comparison of glycerol preservation with cryopreservation methods on HIV-1 inactivation. *J Burn Care Rehabil* 1998;19:494–500.
24. Thomasen H, Pauklin M, Noelle B, et al. The effect of long term storage on the biological and histological properties of cryopreserved amniotic membrane. *Curr Eye Res* 2001;36:247–55.
25. Nakamura T, Yoshitani M, Rigby H, et al. Sterilized, freeze-dried amniotic membrane: a useful substrate for ocular surface reconstruction. *Invest Ophthalmol Vis Sci* 2004;45:93–9.
26. Lim LS, Poh RW, Riau AK, Beuerman RW, Tan D, Mehta JS. Biological and ultrastructural properties of acelagraft, a freeze dried γ -irradiated human amniotic membrane. *Arch Ophthalmol* 2010;128:1303–10.
27. Lopez-Valladares MJ, Teresa Rodriguez-Ares M, Tourino R, Gude F, Teresa Silva M, Couceiro J. Donor age and gestational age influence on growth factor levels in human amniotic membrane. *Acta Ophthalmol* 2010;88:e211–6.
28. Velez DR, Fortunato SJ, Morgan N, et al. Patterns of cytokine profiles differ with pregnancy outcome and ethnicity. *Hum Reprod* 2008;23:1902–9.
29. Gicquel JJ, Dua HS, Brodie A, et al. Epidermal growth factor variations in human amniotic membrane used for ex vivo tissue constructs. *Tissue Eng Part A* 2009;15:1919–27.
30. Lin SD, Lai CS, Hou MF, Yang CC. Amnion overlay meshed autograft. *Burns Incl Therm Inj* 1985;11:374–8.
31. Subrahmanyam M. Amniotic membrane as a cover for micro-skin grafts. *Br J Plast Surg* 1995;48:477–8.
32. Sawhney CP. Amniotic membrane as a biological dressing in the management of burns. *Burns* 1989;15:339–42.
33. Ramakrishnan KM, Jayaraman V. Management of partial thickness burn wounds by amniotic membrane: a cost-effective treatment in developing countries. *Burns* 1997;23:533–6.
34. Branski LK, Herndon DN, Celis MM, et al. Amnion in the treatment of pediatric partial-thickness facial burns. *Burns* 2008;34:393–9.
35. Singh R, Purohit S, Chacharkar, et al. Microbiological safety and clinical efficacy of radiation sterilized amniotic membranes for treatment of second degree burns. *Burns* 2007;33:505–10.
36. Fraser JF, Cuttle L, Kempf M, et al. A randomised controlled trial of amniotic membrane in the treatment of standardised burn injury in the merino lamb. *Burns* 2009;35:998–1003.
37. Mostaque AK, Rahman KB. Comparisons of the effects of biological membrane (amnion) and silver sulfadiazine in the management of burn wounds in children. *J Burn Care Res* 2011;32:200–9.
38. Adly OA, Moghazy AM, Abbas AH, et al. Assessment of amniotic and polyurethane membrane dressings in the treatment of burns. *Burns* 2010;36:703–10.
39. Mohammadi AA, Seyed Jafari SM, Kiasat M, et al. Effect of fresh human amniotic membrane dressing on graft take in patients with chronic burn wounds compared with conventional methods. *Burns* 2013;39:349–53.
40. Seashore JH, MacNaughton RJ, Talbert JL. Treatment of gastroschisis and omphalocele with biological dressings. *J Pediatr Surg* 1975;10:9–17.
41. Tekin S, Tekin A, Kucukkartallar T, Cakir M, Kartal A. Use of chorioamniotic membrane instead of bogota bag in open abdomen: how i do it? *World J Gastroenterol* 2008;14:815–6.
42. Troensegaard-Hansen E. Amniotic grafts in chronic skin ulceration. *Lancet* 1950;1:859–60.
43. Ward DJ, Bennett JP. The long-term results of the use of human amnion in the treatment of leg ulcers. *Br J Plast Surg* 1984;37:191–3.
44. Ward DJ, Bennett JP, Burgos H, et al. The healing of chronic venous leg ulcers with prepared human amnion. *Br J Plast Surg* 1989;42:463–7.
45. Singh R, Chouhan US, Purohit S, et al. Radiation processed amniotic membranes in the treatment of non-healing ulcers of different aetiologies. *Cell Tissue Bank* 2004;5:129–34.
46. Gajiwala K, Lobo Gajiwala A. Use of banked tissue in plastic surgery. *Cell Tissue Bank* 2003;4:141–6.
47. Insausti CL, Alcaraz A, Garcia-Vizcaino, et al. Amniotic membrane induces epithelialization in massive posttraumatic wounds. *Wound Repair Regen* 2010;18:368–77.
48. Tomita T, Hayashi N, Okabe M, et al. New dried human amniotic membrane is useful as a substitute for dural repair after skull base surgery. *J Neurol Surg* 2012;73:302–7.
49. Hasegawa M, Fujisawa H, Hayashi Y, Yamashita J. Autologous amnion graft for repair of myelomeningocele: technical note and clinical implication. *J Clin Neurosci* 2004;11:408–11.
50. de Weerd L, Weum S, Sjavik K, Acharya G, Hennig RO. A new approach in the repair of a myelomeningocele using amnion and a sensate perforator flap. *J Plast Reconstr Aesthet Surg* 2013;66:860–3.
51. Itoh H, Sagawa N, Hasegawa M, et al. Brain natriuretic peptide is present in the human amniotic fluid and is secreted from amnion cells. *J Clin Endocrinol Metab* 1993;76:907–11.
52. Lawson VG. Oral cavity reconstruction using pectoralis major muscle and amnion. *Arch Otolaryngol* 1985;111:230–3.
53. Tancer ML, Katz M, Veridiano NP. Vaginal epithelialization with human amnion. *Obstet Gynecol* 1979;54:345–9.
54. Ashworth MF, Morton KE, Dewhurst J, Lilford RJ, Bates RG. Vaginoplasty using amnion. *Obstet Gynecol* 1986;67:443–6.
55. Samandari MH, Yaghmaei M, Ejllali M, Moshref M, Saffar AS. Use of amnion as a graft material in vestibuloplasty: a preliminary report. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2004;97:574–8.
56. Kothari CR, Goudar G, Hallur N, Sikkerimath B, Gudi S, Kothari MC. Use of amnion as a graft material in vestibuloplasty: a clinical study. *Br J Oral Maxillofac Surg* 2012;50:545–9.
57. Ozkaya O, Egemen O, Yesilada A, Sakiz D, Ugurlu K. The effect of non-preserved human amniotic membrane on the survival of ischaemic skin flaps in rats. *J Plast Reconstr Aesthet Surg* 2012;65:1700–5.
58. Gray KJ, Shenaq SM, Engelmann UH, Fishman IJ, Jeraj K, Spira M. Use of human amnion for microvascular interpositional grafts. *Plast Reconstr Surg* 1987;79:778–85.
59. Ozboluk S, Ozkan Y, Ozturk A, Gul N, Ozdemir RM, Yanik K. The effects of human amnion membrane and periosteal autograft on tendon healing: experimental study in rabbits. *J Hand Surg Eur Vol* 2010;35:262–8.

60. Meng H, Li M, You F, Du J, Luo Z. Assessment of processed human amniotic membrane as a protective barrier in rat model of sciatic nerve injury. *Neurosci Lett* 2011;**496**:48–53.
61. Henry FP, Goyal NA, David WS, et al. Improving electrophysiologic and histologic outcomes by photochemically sealing amnion to the peripheral nerve repair site. *Surgery* 2009;**145**:313–21.
62. O'Neill AC, Randolph MA, Bujold KE, Kochevar IE, Redmond RW, Winograd JM. Photochemical sealing improves outcome following peripheral neurotomy. *J Surg Res* 2009;**151**:33–9.
63. Mohammad J, Shenaq J, Rabinovsky E, Shenaq S. Modulation of peripheral nerve regeneration: a tissue engineering approach. The role of amnion tube nerve conduit across a 1 cm nerve gap. *Plast Reconstr Surg* 2000;**105**:660–6.
64. O'Neill AC, Randolph MA, Bujold KE, Kochevar IE, Redmond RW, Winograd JM. Preparation and integration of human amnion nerve conduits using a light activated technique. *Plast Reconstr Surg* 2009;**124**:428–37.
65. Alpin AE. Cell adhesion molecule regulation of nucleocytoplasmic trafficking. *FEBS Lett* 2003;**534**:11–4.
66. Yang S, Leong KF, Du Z, Chua CK. The design of scaffolds for use in tissue engineering. Part 1. Traditional factors. *Tissue Eng* 2001;**7**:679–89.
67. Koizumi N, Inatomi T, Suzuki T, Sotozono C, Kinoshita S. Cultivated corneal epithelial stem cell transplantation in ocular surface disorders. *Ophthalmology* 2001;**108**:1569–74.
68. Yang L, Shirikata Y, Shudou M, et al. New skin equivalent model from de-epithelialised amnion membrane. *Cell Tissue Res* 2006;**326**:69–77.
69. Jin CZ, Park SR, Choi BH, Lee KY, Kang CK, Min BH. Human amniotic membrane as a delivery matrix for articular cartilage repair. *Tissue Eng* 2007;**13**:693–702.
70. Amensag S, McFetridge PS. Rolling the human amnion to engineer laminated vascular tissues. *Tiss Eng Part C Methods* 2012;**18**:903–12.
71. Burman S, Tejwani S, Vemuganti GK. Ophthalmic applications of preserved human amniotic membrane: a review of current indications. *Cell Tissue Bank* 2004;**5**:161–75.
72. Miki T, Lehmann T, Cai H, Stolz DB, Strom SC. Stem cell characteristics of amniotic epithelial cells. *Stem Cells* 2005;**23**:1549–59.
73. Park IH, Zhao R, West JA. Reprogramming of human somatic cells to pluripotency with defined factors. *Nature* 2008;**451**:141–6.
74. Witkowska-Zimny M, Walenko K. Stem cells from adipose tissue. *Cell Mol Biol Lett* 2011;**16**:236–57.
75. Walter M, Liang S, Ghosh S, Hornsby PJ, Li R. Interleukin 6 secreted from adipose stromal cells promotes migration and invasion of breast cancer cells. *Oncogene* 2009;**28**:2745–55.
76. Blasco MA, Serrano M, Fernandez-Capetillo O. Genomic instability in iPS: time for a break. *EMBO J* 2011;**30**:991–3.
77. Hass R, Kasper C, Bohm S, Jacobs R. Different populations and sources of human mesenchymal stem cells (MSC): a comparison of adult and neonatal tissue derived MSC. *Cell Commun Signal* 2011;**9**:12.
78. De Coppi P, Bartsch G, Siddiqui MM, et al. Isolation of amniotic stem cell lines with potential for therapy. *Nat Biotechnol* 2007;**25**:100–6.
79. Miki T. Amnion-derived stem cells: in quest of clinical applications. *Stem Cell Res Ther* 2011;**2**:25.
80. Enders AC, King BF. Formation and differentiation of extra-embryonic mesoderm in the rhesus monkey. *Am J Anat* 1988;**181**:327–40.

Manuscript 2

Light-Activated Sealing of Nerve Graft Coaptation Sites Improves Outcome Following Large Gap Peripheral Nerve Injury.

Neil G. Fairbairn^{1,2}, Joanna Ng-Glazier^{1,2}, Amanda Meppelink², Mark A. Randolph²,
John M. Winograd², Mark E. Fleming³, Ian L. Valerio³ and Robert W. Redmond^{1*}.

¹ Wellman Center for Photomedicine, Harvard Medical School, Massachusetts General Hospital, Boston, MA 02115.

² Division of Plastic Surgery, Department of Surgery, Harvard Medical School, Massachusetts General Hospital, Boston, MA 02115.

³ Walter Reed National Military Medical Center, 8901 Wisconsin Avenue, Bethesda, MD 20814.

To whom correspondence should be addressed at:

Wellman Center for Photomedicine
Harvard Medical School
Massachusetts General Hospital
55 Fruit Street
Boston, MA 02115
Tel: 617 726-8857
E-mail: Redmond@helx.mgh.harvard.edu

INTRODUCTION

Outcomes following peripheral nerve repair remain frustratingly poor. This is particularly true when tissue loss results in large gaps between nerve ends. The gold standard method of reconstruction involves autografting and suture fixation. However, even with meticulous tissue handling and microsurgical technique, needle trauma and the presence of suture material at the repair site result in extra- and intraneural scar tissue formation¹. This causes a direct obstacle for regenerating axons and can result in external tethering, compression and traction neuritis. These effects are amplified in the context of nerve grafting when axons must traverse two coaptation sites. Reducing manipulation of injured nerve ends and suture burden at the repair site is a logical solution to this problem.

Several sutureless methods of nerve repair have been thoroughly investigated. Fibrin sealants and laser-assisted techniques represent two of the most extensively studied. Fibrin sealants rely on the combination of fibrinogen and thrombin to form a fibrin clot. Although originally designed for hemostasis, “off-label” application as an adhesive for peripheral nerve repair dates back to the 1940s^{2,3}. Proponents claim that fibrin glue can be rapidly applied, results in less scar tissue formation, and can improve outcomes^{4,5,6,7,8,9}. In contrast, others have reported excessive fibrosis, inferior bond strength and high rates of dehiscence^{10,11,12,13}. Introduced in the 1980s, laser welding of peripheral nerves has generally been performed using carbon dioxide and argon lasers¹⁴. A photothermal reaction at the repair site creates a bond consisting of a coagulum of denatured protein. Proposed advantages include shorter repair times, reduced incidence of inflammation, scarring and neuroma. In one rodent model, laser welding was found to be at least as effective as fibrin glue and epineurial suture^{14,15}. However, due to thermal damage, poor bond strength and high rates of dehiscence, the technique has not been clinically adopted^{16,17,18,19,20,21}. The introduction of biological solder such as fascia or bovine albumin promised to enhance bond strength and limit thermal damage but interest in its potential has largely been abandoned.

Photochemical tissue bonding (PTB) differs from laser welding as it uses visible light to create covalent bonds between proteins of opposed tissues that have been pre-stained with a non-toxic, photoactive dye, resulting in a water-tight seal with no thermal damage. Dye photoactivation results in the creation of reactive species and subsequent crosslinking between amino acid residues^{22,23,24,25}. The absence of collateral thermal damage is a distinct advantage over laser welding. In recent years our group has consistently showed that photochemical sealing of neurorrhaphy sites with a human amnion nerve wrap (Fig 1.) is a viable alternative to suture and can result in superior functional and histological outcomes in comparison with standard suture repair^{26,27,28,29}. These observations are likely due to a reduction in intra- and extra-neural scar tissue, the prevention of axonal escape, and the enclosure and prevention of leakage of beneficial neuro-regenerative factors. It is also possible that growth promoting and anti-fibrotic factors within amniotic membrane may contribute to this effect.

As with any biological material, amnion is susceptible to proteolytic degradation. When applied to corneal ulcers, untreated amnion degrades within 7 days³⁰. In unpublished work, we observed that, when applied to long nerve grafts, photochemically bonded amnion wraps at the distal neurorrhaphy site failed prior to the arrival of regenerating axons, resulting in nerve dehiscence. Cross-linking of biomaterials reduces enzymatic degradation and can be readily applied to collagen matrix of amnion. Chemicals such as glutaraldehyde and physical methods such as gamma and electron beam irradiation are common crosslinking methods but are limited in this application by toxicity issues and inconsistent cross-linking density respectively^{31,32,33,34,35}. The water soluble carbodiimide 1-ethyl-3-(3-dimethylamionopropyl) carbodiimide hydrochloride (EDC) is an alternative non-toxic agent resulting in carboxyl-to-amine cross-linking between proteins. Often used in combination with N-hydroxysuccinimide (NHS), EDC has been successfully used to improve biomechanical strength and resistance to degradation of several collagen-based biomaterials, including amnion^{36,37,38,39,40}.

The mechanism of chemical crosslinking is distinct from that of photocrosslinking and we hypothesized that pre-treatment of nerve wraps with EDC/NHS prior to PTB should result in a stronger, more durable nerve wrap that retains the ability to undergo photochemical bonding. Recent *ex vivo* work assessing the biomechanical properties of human amnion and a commercially available SIS material has confirmed this hypothesis and has demonstrated that 4mM EDC/1mM NHS represents the optimum crosslinker concentration to achieve these goals (manuscript in preparation). By applying various different methods of fixation (suture, fibrin glue, PTB) to these nerve wraps, this study has conducted a comprehensive and clinically applicable *in vivo* assessment of the optimal repair strategy following large gap injury.

METHODS

Nerve wrap biomaterials

Human amnion (HAM) harvest and processing

Human amniotic membrane was obtained from elective caesarean section patients who had been screened serologically for human immunodeficiency virus-1/2, Hepatitis B, Hepatitis C, human T-cell lymphotropic virus, syphilis, cytomegalovirus, and tuberculosis. Following delivery, amnion was bluntly removed from the placenta and washed liberally with phosphate buffered saline (PBS; Sigma-Aldrich, Co., St Louis, Mo). Membranes were mechanically de-epithelialized using a cell scraper, cut into strips, wrapped around nitrocellulose paper and placed in a storage solution containing a 1:1 mix of 100% sterile glycerol and Dulbeccos modified Eagles medium (DMEM; Gibco, Grand Island, NY), penicillin-streptomycin-neomycin (PSN; Gibco, Grand Island, NY) and amphotericin B. HAM was stored at -80°C until required. Following thawing, HAM was mounted onto nitrocellulose paper before being dried and cut into 1cm x 1cm sections.

Swine intestinal submucosa (SIS)

Single layer SIS material was obtained from HealthPoint Biotherapeutics Ltd. (Fort Worth, TX). This dehydrated material was removed from packaging and cut into 1x1cm sections prior to use.

Nerve wrap crosslinking

Crosslinking solution was made by adding EDC/NHS (Sigma-Aldrich, Co., St Louis, Mo) to 2-(N-morpholino)ethanesulfonic (MES) acid buffer (Sigma-Aldrich, Co., St Louis, Mo). Nerve wraps were immersed in EDC/NHS for 1-hour on a platform shaker. Based on *ex vivo* experiments, optimal crosslinker concentration was 4mM EDC/1mM NHS.

Sciatic nerve injury and reconstruction

The Institutional Animal Care and Use Committee (IACUC) at the Massachusetts General Hospital approved all procedures. One hundred and ten male, inbred Lewis rats weighing 250-300g were randomized to one of eleven experimental groups. This breed was selected in order to permit immunotolerant isograft exchange between rodents for nerve gap reconstruction. Induction and maintenance anesthesia was achieved using isoflurane (Baxter Healthcare Corp. Deerfield IL; 5% induction/2-3% maintenance). A dorsolateral, muscle-splitting incision was made on the left hindquarter of each animal and, under the operating microscope, 1.5cm sections of nerve were excised and exchanged between two simultaneously anesthetized animals. Two groups (n=10) served as positive (6-epineurial sutures) and negative (no repair) controls. The remaining nine groups (n=10) had nerves reconstructed using different combinations of the three different nerve wraps and three different fixation methods (Table 1; Fig.2). Wounds were closed in three layers with 4.0 vicryl (muscle and deep dermal) and 4.0 monocril (subcuticular). Topical antibacterial ointment and bitter apple were applied liberally to wounds. Rodents were housed in the Massachusetts General Hospital small animal facility and had access to food and water as required.

Group 1: Negative control (n=10)

Following the creation of nerve defects, a small incision was made in adjacent muscle. Proximal nerve ends were sutured into muscle pockets using two 10.0 Ethilon sutures (Ethicon, Sommerville, NJ).

Distal nerve ends were left free.

Group 2: Positive control (n=10)

Following nerve exchange, grafts were secured with six 10.0 epineurial sutures at each neurorrhaphy site. Following repair, any axons protruding from the repair site were trimmed.

Suture fixation (n=10) – groups 3, 4, 5

On the morning of surgery, candidate nerve wraps were prepared as described above. Following nerve graft exchange, each graft was secured into the defect using two 10.0 nylon sutures at each end. Nerve wraps were rehydrated for 60 seconds in PBS before being transferred into the surgical field. Once applied circumferentially at each repair site, wraps were secured with one proximal and one distal 10.0 suture. Great care was taken to include only the wrap and the underlying epineurium in each bite.

Fibrin glue fixation (n=10) – groups 6, 7, 8

Wraps were prepared and nerve grafts tacked into place as described above. Following wrap application, Tisseel fibrin glue (Baxter Healthcare Corp. Deerfield IL) was applied to each nerve wrap interface, ensuring that the entire wrap was covered in glue.

PTB fixation (n=10) – groups 9, 10, 11

Wraps were prepared and nerve grafts tacked into place as described above. Prior to transfer into the surgical field, wraps and neurorrhaphy sites were stained with 0.1% (w/v) Rose Bengal (RB) (Sigma-Aldrich, Co., St Louis, Mo) for 60 seconds. After 60 seconds, excess dye was removed. RB-stained wraps were wrapped circumferentially around sciatic nerves ensuring that a minimum of 5mm overlap existed. The area of overlap was irradiated for 60-seconds using a 532nm KTP laser (Laserscope, San Jose, Ca) at an irradiance of $0.5\text{W}/\text{cm}^2$. The nerve/wrap was then rotated 180° in order to irradiate the back wall in the same manner for an additional 60-seconds (Fig 1).

Outcome assessment

Walking track analysis

Walking track analysis was performed immediately prior to surgery for baseline sciatic function index (SFI). Following surgery, walking track analysis was performed at 30-day intervals. After dipping both hind paws in water soluble ink, rats were encouraged to walk up a 10 x 60 cm, partially enclosed ramp lined with white paper and set at an incline of 30° to horizontal. Measurements of print length, toe spread and intermediary toe spread were measured from the resulting prints using digital calipers. Mean values from three normal and experimental prints were entered into the SFI formula described by Bain and colleagues⁴¹.

Muscle weight retention

All rodents were sacrificed 150-days post-operatively by carbon dioxide inhalation. Left and right gastrocnemius muscles were harvested. Wet weights were recorded immediately and percentage muscle mass retention calculated.

Histology and histomorphometric analysis

Following sacrifice, nerves were harvested 5mm proximal and distal to the graft and immediately fixed in a mixture of 2% glutaraldehyde/2% paraformaldehyde (Electron Microscopy Sciences, Hatfield, PA). After 48 hours, fixed nerves were washed in sodium cacodylate buffer (0.1M; pH=7.4) and post-fixed in 2% osmium tetroxide (Electron Microscopy Sciences, Hatfield, PA) for 2 hours. Following further washing in sodium cacodylate buffer, specimens were dehydrated in increasing concentrations of ethanol (25%, 50%, 75%, 95%, 100%). Following dehydration, all specimens were washed with propylene oxide (Electron Microscopy Sciences, Hatfield, PA). Specimens were then placed in increasing concentrations of Epoxy resin (DDSA (dodecyl succinic anhydrides 98+%; Free Acid 2%)/tEPON-812 (Epoxy Resin)/NMA Ultrapure (methyl-5-norbornene-2,3-dicarboxylic anhydride)/DMP-30 (2,4,6-tri (dimethylaminomethyl) phenol (Tousimis Research Corporation, Rockville, MD) before being baked overnight in an oven at 60°C. Using a diamond blade, 1µm sections were cut 5mm proximal and 5mm distal to the graft.

Histology slides were scanned using a Hamamatsu NanoZoomer 2.0-HT slide scanner (Meyer Instruments, Houston TX) and read using NDP.com software (Hamamatsu Corp. Bridgewater, NJ). Nerve area was calculated at 40x magnification. From these images, five 400x images were randomly selected and imported into Adobe Photoshop. Axon counts were measured and from fifty randomly (randomizer.org) selected fibers (250 nerve fibers per location), fiber diameter, axon diameter, myelin thickness and G-ratio were calculated.

Statistical analysis

Statistical analysis was performed using KaleidaGraph for Windows v4.1 (Synergy Software, Reading, PA). Testing between experimental groups was achieved using analysis of variance (ANOVA) and the post-hoc Bonferroni test. Statistical significance was set at <0.05 .

RESULTS

Sciatic nerve graft reconstruction

Gross observations

Following sacrifice, two rodents from the xSIS+PTB group were found to have dehisced repairs. All other nerve repairs were intact. Crosslinked amnion and SIS nerve wraps that had been stained with RB, although not fully intact, were identifiable (Fig. 3C). No evidence of untreated nerve wraps could be found in any of the fixation groups. Although not quantitatively assessed, those nerves repaired photochemically were found to have considerably less extraneural scar tissue formation in comparison to standard graft+suture (Fig. 3A-3C).

Sciatic function Index

All repairs experienced some degree of functional recovery. After 5-months follow-up, nerves repaired with xHAM+PTB showed greatest functional recovery although this was not statistically significant in comparison to graft+suture (-67.9 ± 5.1 vs -71.7 ± 4.8 ; Table 2, Fig. 4). SIS+suture and SIS+PTB groups experienced significantly poorer functional recovery in comparison to positive controls ($-80.3\pm$

3.2 and -85.0 ± 6.0 vs -71.7 ± 4.8 respectively; $p < 0.01$). There were no other significant differences between any of the remaining groups and positive controls.

Gastrocnemius muscle mass retention

Nerves repaired with xHAM+PTB experienced a statistically significant improvement in muscle mass retention compared with graft+suture ($67.3\% \pm 4.4$ vs $60.0\% \pm 5.2$; $p = 0.02$). There were no other significant differences between remaining treatment groups and graft+suture controls (Table 3, Fig. 5). Although not statistically significant, those wraps secured with fibrin glue and PTB displayed a trend towards superior recovery in comparison to suture fixation.

Histomorphometry

All distal nerve sections were populated with axons (Fig. 6). As expected, axon counts in the no repair group were significantly lower than all other repair groups. Distal axon counts in the remaining treatment groups did not differ significantly in comparison to graft+suture (Table 4). However, nerve fiber diameter, axon diameter and myelin thickness were all significantly increased in the xHAM+PTB group in comparison to graft+suture repair (Table 4).

DISCUSSION

This study shows that photochemical sealing of nerve graft coaptation sites using crosslinked human amnion results in statistically significant improvements in muscle mass retention and histomorphometric outcomes compared with standard graft+suture. With the exception of xSIS, all crosslinked nerve wraps displayed a trend towards greater functional and histological recovery. Although *in vivo* nerve wrap degradation was not formally assessed throughout the regenerative period, there was visible evidence of persisting crosslinked RB-stained wraps following sacrifice (Fig. 3C). In contrast, untreated wraps were completely degraded (Fig. 3B). Improved wrap durability may maintain a protective seal at nerve graft coaptation sites for longer, preventing wrap and bond degradation and subsequent dehiscence prior to the arrival of regenerating axons. Partial degradation of crosslinked material did occur. Although EDC

successfully protects against collagenase degradation, it may be unable to offer protection against other proteolytic enzymes.

Fixation methods associated with reduced suture burden (fibrin glue and PTB) also trended towards greater functional and histomorphometric recovery. These observations are consistent with widely accepted tenets regarding the detrimental, pro-inflammatory effect of suture. Outcomes following photochemical sealing surpassed those achieved using fibrin glue and only those nerves repaired with photochemically-sealed crosslinked amnion resulted in statistically significant improvements in muscle mass retention and histomorphometry in comparison to standard graft+suture. This suggests that the effects of reduced suture burden and photochemical sealing are additive. Substantial volumes of axoplasmic fluid, rich in neuroregenerative factors, are released from nerve stumps following injury^{45,46,47}. The containment of this fluid at the site of repair by water-tight sealing may explain these observations. The lack of significant improvement in those repairs photochemically sealed with untreated amnion supports the claim that water-tight sealing of nerve grafts in large deficit injuries is only efficacious if amnion can resist proteolytic degradation over extended periods of recovery.

As a stand-alone technique, it is generally accepted that fibrin glue provides insufficient bond strength to hold nerve ends in close apposition. Indeed, most favorable outcomes have been reported when fibrin glue is used to augment suture repair^{4,5,6,7,8,9}. Fibrin bonds have also been shown to degrade within three weeks⁴⁴. As a result, the suitability of fibrin glue for wrap fixation was questionable. In spite of these concerns, no dehiscence occurred in this group and recovery was comparable to standard graft+suture. This verifies that fibrin glue, in combination with epineurial tacking sutures, provides sufficient support and is a viable alternative for repair.

Those nerves repaired photochemically had considerably less extraneural adhesions in comparison to standard graft+suture (Fig. 3A-3C), a finding consistent with previous studies²⁷. The scar-reducing properties of this technique may be related, not only to reduced suture burden, but also to an inhibitory

photochemical effect on tissue fibroblasts and also to the widely documented anti-fibrotic effects of transplanted amniotic membrane^{42,43}.

Two cases of nerve dehiscence occurred in the xSIS+PTB group. Despite superior mechanical strength and durability, *in vivo* use of SIS was problematic. Single layer SIS is considerably thicker than amniotic membrane (100µm vs. 20-50µm). This impaired circumferential wrapping and adherence to the epineurium of the small diameter rodent sciatic nerve. As a result, photochemical bonding was sub-optimal and visibly not water-tight. Although ineffective for very small nerves, we are optimistic that this material may still be suitable for larger caliber nerves.

The limitations of current functional and histological outcome assessment for peripheral nerve research in rodent models have been well reported^{48, 49, 50, 51}. Several additional limitations are inherent to our animal model. During suture fixation of nerve wraps, it was difficult to ensure that underlying axons were not included within the suture bite. This was particularly relevant when attempting fixation of relatively thick SIS. Inadequate fixation due to excessively superficial bites and inadvertent axonal strangulation due to excessively deep bites may have compromised recovery in these groups.

Photochemical bonding required clear access 5mm proximal and distal to the coaptation site. As a result, the maximum achievable nerve gap before sciatic trifurcation was 1.5-2cm. The limited “large” gap, coupled with the large regenerative capacity of rodents may have interfered with the detection of differences between treatment groups.

We exchanged isografts between two simultaneously anaesthetized rodents. Slight discrepancies in nerve graft length made tensionless repair impossible. The insertion of two tacking sutures at each coaptation site was necessary to permit photochemical sealing. As a result, we were unable to achieve truly “sutureless” repairs. However, in the clinical arena, nerve autografts and allografts can be

oversized to eliminate tension and obviate this requirement for sutures. Whilst tacking sutures may still be necessary for group fascicular repair and cable grafting, subsequent wrapping of the entire gap and photochemical sealing of coaptation sites still offers the benefit of water-tight sealing and containment of neurotrophic-rich fluid.

CONCLUSIONS

Photochemical sealing of nerve graft coaptation sites using crosslinked human amnion nerve wraps results in a statistically significant improvement in muscle mass retention and nerve histomorphometry in comparison to conventional graft+suture. This observation is likely related to the creation of a water tight seal at nerve graft coaptation sites, the improved longevity of this seal as a result of nerve wrap crosslinking, and a reduction in inflammation and scar tissue formation secondary to reduced suture burden and exposure to low power laser. Although unsuitable for small caliber nerves, we are optimistic that SIS may represent a viable, commercially available nerve wrap that may facilitate the rapid clinical translation of photochemical sealing of larger caliber nerves.

ACKNOWLEDGEMENTS

Financial support for this study was provided by the Peer-Reviewed Orthopedic Research Program (PRORP) through USAMRAA award W81XWH-12-1-0511.

REFERENCES

1. DeLee JC, Smith MT, Green DP. The reaction of nerve tissue to various suture materials: A study in rabbits. *J Hand Surg (Am)*. 1977;2:38-43.
2. Tarlov IM, Denslow C, Swarz S, Pineless D. Plasma clot suture of nerves. *Arch Surg*. 1943;47:44-58
3. Young JZ and Medawar PB. Fibrin suture of peripheral nerves. *Lancet*. 1940;239:126-128
4. Narakas A. The use of fibrin glue in repair of peripheral nerves. *Orthop Clin North Am*. 1988;19:187
5. Bento RF and Miniti A. Anastomosis of the intratemporal facial nerve using fibrin tissue adhesive. *Ear Nose Throat J*. 1993;72:663
6. Martins RS, Siqueira MG, Da Silva CF, Plese JP. Overall assessment of regeneration in peripheral nerve lesion repair using fibrin glue, suture or a combination of the 2 techniques in a rat model. Which is the ideal choice? *Surg Neurol*. 2005;64(Supp 1):10-16; discussion 16.
7. Menovsky T and Beek JF. Laser, fibrin glue, or suture repair of peripheral nerves: a comparative functional, histological, and morphometric study in the rat sciatic nerve. *J Neurosurg*. 2001;95:694-699.
8. Feldman MD, Sataloff RT, Epstein G, Ballas SK. Autologous fibrin tissue adhesive for peripheral nerve anastomosis. *Arch Otolaryngol Head Neck Surg*. 1987;113:963-967.
9. Smahel J, Meyer VE, Bachem U. Gluing of peripheral nerves with fibrin: experimental studies. *J Reconstr Microsurg*. 1987;3:211-220.
10. Cruz NI, Debs N, Fiol RE. Evaluation of fibrin glue in rat sciatic nerve repairs. *Plas Reconstr Surg*. 1986;178:369-73.
11. Maragh H, Meyer BS, Davenport D, Gould JD, Terzis JK. Morphofunctional evaluation of fibrin glue versus microsuture nerve repairs. *J Reconstr Microsurg*. 1990;6:331-337.
12. Sames M, Blahos J, Rokyta R, Benes V. Comparison of microsurgical suture with fibrin glue connection of the sciatic nerve in rabbits. *Physiol Res*. 1997;46:303-306.

13. Herter T. Problems with the use of fibrin adhesion of the nerves. *Neurosurg Rev.* 1988;11:249-258
14. Menovsky T, Beek JF, Thomsen SL. Laser (-assisted) nerve repair. A review. *Neurosurg Rev.* 1995;18:225-235
15. Fischer DW, Beggs JL, Kenshalo DL, Jr., et al. Comparative study of microepineurial anastomoses with the use of CO₂ laser and suture techniques in rat sciatic nerves. Part 1. Surgical technique, nerve action potentials, and morphological studies. *Neurosurgery.* 1985;17:300-308.
16. Eppley BL, Kalenderian E, Winkelmann T, et al. Facial nerve graft repair: Suture versus laser-assisted anastomosis. *Int J Oral Maxillofac Surg.* 1989;18:50-54.
17. Bass LS, Moazami N, Pocsidio J, et al. Changes in type I collagen following laser welding. *Lasers Surg Med.* 1992;12:500-505
18. Dort JC, Wolfensberger M, Felix H. CO₂ laser repair of the facial nerve: An experimental study in the rat. *J Laryngol Otol.* 1994;108:466-469.
19. Korff M, Bent SW, Havig MT, et al. An investigation of the potential for laser nerve welding. *Otolaryngol Head Neck Surg.* 1992;106:345-350.
20. Huang TC, Blanks RH, Berns MW, et al. Laser versus suture nerve anastomosis. *Otolaryngol Head Neck Surg.* 1992;107:14-20.
21. Menovsky T, Beek JF. Carbon dioxide laser-assisted nerve repair: Effect of solder and suture material on nerve regeneration in rat sciatic nerve. *Microsurgery.* 2003;23:109-116.
22. Chan BP, Hui TY, Chan OCM, et al. Photochemical cross-linking for collagen based scaffolds: A study on optical properties, mechanical properties, stability and hematocompatibility. *Tissue Eng.* 2007;13:73-85
23. Kochevar IE, Redmond RW. Photosensitized production of singlet oxygen. *Methods Enzymol.* 2000;319:20-28

24. Balasubramanian D, Du X, Zigler JS Jr. The reaction of singlet oxygen with proteins, with special reference to crystallins. *Photochem Photobiol.* 1990;52:761-768
25. Webster A, Britton D, Apap-Bologna A, Kemp G. A dye-photosensitized reaction that generates stable protein-protein crosslinks. *Anal Biochem.* 1989;179:154-157
26. Johnson TS, O'Neill AC, Motarjem PM, et al. Photochemical tissue bonding: A promising technique for peripheral nerve repair. *J Surg Res.* 2007;143:224-9
27. O'Neill AC, Randolph MA, Bujold KE, Kochevar IE, Redmond RW, Winograd JM. Photochemical sealing improves outcome following peripheral neurorrhaphy. *J Surg Res.* 2009;151:33-9
28. Henry FP, Goyal NA, David WS, et al. Improving electrophysiologic and histologic outcomes by photochemically sealing amnion to the peripheral nerve repair site. *Surgery.* 2009;145:313-21
29. O'Neill AC, Randolph MA, Bujold KE, Kochevar IE, Redmond RW, Winograd JM. Preparation and integration of human amnion nerve conduits using a light activated technique. *Plast Reconstr Surg.* 2009;12:428-37
30. Spoerl E, Wollensak G, Reber F, Pillunat L. Cross-linking of human amniotic membrane by glutaraldehyde. *Ophthalmic Res.* 2004;36:71-77
31. Fujisato T, Tomihata K, Tabata Y, Iwamoto Y, Burczak K, Ikada Y. Cross-linking of amniotic membranes. *J Biomater Sci Polym Ed.* 1999;10:1171-1181
32. Spira M, Liu B, Xu Z, Harrell R, Chahadeh H. Human amnion collagen for soft tissue augmentation-biochemical characterizations and animal observations. *J Biomed Mater Res.* 1994;28:91-96.
33. Bigi A, Cojazzi G, Panzavolta S, Rubini K, Roveri N. Mechanical and thermal properties of gelatin films at different degrees of glutaraldehyde crosslinking. *Biomaterials.* 2001;22:763-768.
34. Isenburg JC, Simionescu DT, Vyavahare NR. Tannic acid treatment enhances biostability and reduces calcification of glutaraldehyde fixed aortic wall. *Biomaterials.* 2005;26:1237-1245.
35. Guldner NW, Jasmund I, Zimmermann H, et al. Detoxification and endothelialization of

- glutaraldehyde-fixed bovine pericardium with titanium coating: A new technology for cardiovascular tissue engineering. *Circulation*. 2009;119:1653-1660.
36. Liu Y, Griffith M, Watsky MA, et al. Properties of porcine and recombinant human collagen matrices for optically clear tissue engineering applications. *Biomacromolecules*. 2006;7:1819-1828.
 37. Duan X and Sheardown H. Dendrimer crosslinked collagen as a corneal tissue engineering scaffold: Mechanical properties and corneal epithelial cell interactions. *Biomaterials*. 2006;27:4608-4617.
 38. Rafat M, Li F, Fagerholm P, et al. PEG-stabilized carbodiimide crosslinked collagen-chitosan hydrogels for corneal tissue engineering. *Biomaterials*. 2008;29:3960-3972.
 39. Liu W, Deng C, McLaughlin CR, et al. Collagen-phosphorylcholine interpenetrating network hydrogels as corneal substitutes. *Biomaterials*. 2009;30:1551-1559.
 40. Ma DH, Lai JY, Cheng HY, Tsai CC, Yeh LK. Carbodiimide cross-linked amniotic membranes for cultivation of limbal epithelial cells. *Biomaterials*. 2010;31:6647-6658
 41. Bain JR, Mackinnon SE, Hunter DA. Functional evaluation of complete sciatic, peroneal, and posterior tibial nerve lesions in the rat. *Plast Reconstr Surg* 1989;83:129-38
 42. Fairbairn N, Randolph MA, Redmond RW. The clinical applications of human amnion in plastic surgery. *J Plast Reconstr Aesthet Surg*. 2014;67:662-675
 43. Mamede AC, Carvalho MJ, Abrantes AM, Laranjo M, Maia CJ, Botelho MF. Amniotic membrane: From structure and functions to clinical applications. *Cell Tissue Res*. 2012;349:447-58
 44. Dunn CJ, Goa KL. Fibrin sealant: A review of its use in surgery and endoscopy. *Drugs*. 1999;58:863-886.
 45. Demirkan F, Snyder CC, Latifoglu O, et al. A method of enhancing regeneration of conventionally repaired peripheral nerves. *Ann Plast Surg*. 1995;34:67-72
 46. Lundborg G, Longo FM, Varon S. Nerve regeneration model and trophic factors in vivo. *Brain*

Res. 1982;232:157-161

47. Longo FM, Manthorpe M, Skaper SD, et al. Neuronotropic activities accumulate in vivo within silicone nerve regeneration chambers. *Brain Res.* 1983;261:109-117
48. Reddet R, Jari R, Crawford T, Chen YG, Rohde C, Brushart TM. Peripheral nerve pathways regulate motoneuron collateral dynamics. *J Neuroscience.* 2005;25:9406-9412
49. Streppel M, Azzolin N, Dohm S, et al. Focal application of neutralizing antibodies to soluble neurotrophic factors reduces collateral axonal branching after peripheral nerve lesion. *Eur J Neurosci.* 2002;15:1327-1342.
50. Boyd JG and Gordon T. Glial cell line-derived neurotrophic factor and brain-derived neurotrophic factor sustain the axonal regeneration of chronically axotomized motoneurons in vivo. *Exp Neurol.* 2003;183:610-619.
51. Guntinas-Lichius O, Irintchev A, Streppel M, et al. Factors limiting motor recovery after facial nerve transection in the rat: Combined structural and functional analyses. *Eur J Neurosci.* 2005;21:391-402.

FIGURE LEGENDS

Figure 1. Technique of nerve repair using PTB. Divided nerve ends brought into close apposition. If retraction of nerve ends prevents this, two epineurial tacking sutures may be placed 180° apart to facilitate apposition. Nerve ends and nerve wrap stained with Rose Bengal for 60 seconds. Nerve wrap is wrapped circumferentially around nerve ends and is illuminated with 532nm laser for 60 seconds. Nerve is rotated 180° and back wall illuminated for further 60 seconds.

Figure 2. Methods of nerve repair for control and experimental groups. Positive controls had nerves repaired using isografts secured with standard epineurial suture. Negative controls had 15mm sections of nerve excised and no repair. Proximal nerve ends were buried into adjacent muscle and secured with 2x10.0 nylon suture. Nine experimental groups were composed of different combinations of the three different nerve wraps and three different fixation methods illustrated.

Figure 3. Gross observations following sacrifice. (A) Graft+suture (B) HAM+PTB (C) xHAM+PTB. Crosslinked nerve wraps (xHAM+PTB) were still identifiable after 5-months, as evidence by the pink staining from Rose Bengal (C). In contrast, no evidence of non-crosslinked wraps, that had also been stained with Rose Bengal (HAM+PTB), could be found (B). Although not formally assessed, those nerves repaired photochemically developed considerably less scar tissue formation in comparison to those repairs performed using standard epineurial suture (B and C vs. A).

Figure 4. SFI for select groups. Those nerve repaired using xHAM+PTB recovered greatest SFI although this was not statistically significant compared with standard graft+suture (67.9±5.1 vs - 71.7±4.8). Photochemical repairs using un-crosslinked amnion (HAM+PTB) performed less well than

xHAM+PTB and standard graft+suture although this was not statistically significant (see table 2).

xSIS+PTB performed least well out of all treatment groups.

Figure 5. Gastrocnemius Muscle Mass Retention. Those nerves repaired using xHAM+PTB recovered greatest gastrocnemius muscle mass and this was statistically significant in comparison to graft+suture (67.3+/-4.4 vs 60.0+/-5.2; p=0.02). Photochemical sealing of amnion wraps tended to outperform fibrin and suture fixation. This trend was not observed with xSIS, which performed least well out of all treatment groups.

Figure 6. Histology slides from distal section site. Those nerves that were not repaired lacked axons. All treatment groups regenerated axons distal to isografts with the exception of those nerves that dehisced in the xSIS+PTB group. Axons in the xHAM+PTB group had statistically larger fiber diameter, axon diameter and myelin thickness in comparison to graft+suture (see table 4).

Tables

Table 1. Experimental groups. Nine treatment groups composed of different combinations of 3 different nerve wraps and three different fixation methods. Two control groups consisted of no repair (negative control) and nerve graft and standard epineurial suture (positive control)

	Nerve wrap material		
	HAM	xHAM	xSIS
Suture	Group 3 (HAM+suture)	Group 4 (xHAM+suture)	Group 5 (xSIS+suture)
Fibrin glue	Group 6 (HAM+Fibrin)	Group 7 (xHAM+Fibrin)	Group 8 (xSIS+Fibrin)
PTB	Group 9 (HAM+PTB)	Group 10 (xHAM+PTB)	Group 11 (xSIS+PTB)

Table 2. Monthly mean SFIs. After 5-months follow-up, those nerves repaired with photochemically sealed crosslinked amnion (xHAM+PTB) recovered greatest SFI, although this was not statistically significant. No repairs and those nerves repaired using xSIS+suture and xSIS+PTB performed statistically worse than graft+suture.

No Repair	-89.0+/-3.9	-94.2+/-4.7*	-89.8+/-5.1*	-89.3+/-4.8*	-96.2+/-3.7*
Standard Graft + Suture	-87.6+/-5.0	-81.1+/-4.5	-71.8+/-7.3	-74.7+/-6.3	-71.7+/-4.8
HAM+suture	-90.3+/-5.2	-85.7+/-9.8	-80.6+/-3.5*	-79.7+/-5.7	-77.9+/-6.3
HAM+fibrin	-89.2+/-4.0	-81.6+/-4.1	-80.4+/-7.2*	-79.4+/-4.3	-75.2+/-4.6
HAM+PTB	-90.0+/-5.2	-81.2+/-2.4	-72.8+/-4.6	-75.6+/-3.3	-74.5+/-4.5
xHAM+suture	-96.6+/-7.5	-82.4+/-4.8	-80.0+/-4.0*	-81.4+/-4.8	-76.8+/-2.7
xHAM+fibrin	-90.9+/-3.2	-84.1+/-3.6	-79.8+/-3.3*	-81.2+/-3.1	-75.0+/-4.0
xHAM+PTB	-88.2+/-3.9	-80.3+/-3.5	-67.2+/-3.3	-71.6+/-5.5	-67.9+/-5.1
xSIS+suture	-94.7+/-3.9	-85.6+/-4.4	-82.5+/-4.4*	-81.4+/-4.4	-80.3+/-3.2*
xSIS+fibrin	-93.2+/-4.6	-84.7+/-4.9	-82.0+/-3.7*	-81.5+/-3.9	-78.8+/-3.9
xSIS+PTB	-92.5+/-2.0	-84.7+/-5.0	-84.3+/-4.6*	-85.3+/-6.3*	-85.0+/-6.0*
*Statistically significantly worse in comparison to standard graft+suture; p<0.05					

Table 3. Left gastrocnemius muscle mass retention. Those nerves repaired using xHAM+PTB recovered greatest gastrocnemius muscle mass retention. This result was statistically significant. Those nerves repaired using xSIS+suture and xSIS+PTB recovered least gastrocnemius muscle mass.

No Repair	9.2	0.9	<0.0001
Standard Graft + Suture	60.0	5.2	1
HAM+suture	56.0	5.6	1
HAM+fibrin	59.8	5.4	1
HAM+PTB	62.5	4.0	1
xHAM+suture	57.7	5.1	1
xHAM+fibrin	62.7	4.3	1
xHAM+PTB	67.3*	4.4	0.02
xSIS+suture	54.9	4.5	0.68
xSIS+fibrin	58.5	5.4	1
xSIS+PTB	54.1	3.2	0.37
*Statistically significant improvement in comparison to standard graft+suture; p<0.05			

Table 4: Histomorphometric analysis of distal nerve sections. Axon number did not differ significantly between treatment groups. Fiber diameter, axon diameter and myelin thickness were all significantly greater in those nerves repaired using xHAM+PTB in comparison to graft+suture.

Experimental Group	Total axon count (x0.001)	Axon Density (mm ² x 0.001)	Nerve fiber diameter (μm)	Axon diameter (μm)	Myelin thickness (μm)	G-ratio
No repair	0.04+/-0.05	0.48+/-0.49	4.14+/-1.15	3.13+/-1.17	1.01+/-0.48	0.74+/-0.12
Standard Graft+ Suture	7.61+/-3.42	29.36+/-18.10	5.47+/-1.70	3.50+/-1.44	1.96+/-0.47	0.62+/-0.08
HAM+suture	10.41+/-3.99	28.85+/-18.61	5.07+/-1.58	3.44+/-1.39	1.63+/-0.50	0.67+/-0.17
HAM+fibrin	10.42+/-1.54	29.95+/-14.00	5.22+/-1.67	3.44+/-1.45	1.78+/-0.45	0.64+/-0.09
HAM+PTB	9.31+/-4.19	30.70+/-8.94	5.19+/-1.76	3.47+/-1.53	1.72+/-0.41	0.65+/-0.09
xHAM+suture	9.79+/-3.35	27.12+/-9.22	5.14+/-1.66	3.54+/-1.47	1.59+/-0.36	0.67+/-0.08
xHAM+fibrin	10.87+/-4.32	32.12+/-20.28	5.24+/-1.68	3.52+/-1.49	1.72+/-0.42	0.65+/-0.09
xHAM+PTB	9.66+/-3.08	30.73+/-14.73	6.87+/-2.23*	4.51+/-1.83*	2.35+/-0.64*	0.64+/-0.08
xSIS+suture	9.36+/-2.41	30.30+/-16.46	4.83+/-1.42	3.31+/-1.29	1.52+/-0.38	0.67+/-0.08
xSIS+fibrin	6.91+/-2.62	31.55+/-13.37	5.18+/-1.50	3.58+/-1.30	1.59+/-0.52	0.68+/-0.11
xSIS+PTB	7.84+/-2.04	30.06+/-13.38	4.81+/-1.49	3.35+/-1.33	1.45+/-0.34	0.68+/-0.08
*Statistically significant improvement in comparison to standard graft+suture; p<0.05						

Figure 1. Technique of nerve repair using PTB. Divided nerve ends brought into close apposition. If retraction of nerve ends prevents this, two epineurial tacking sutures may be placed 180° apart to facilitate apposition. Nerve ends and nerve wrap stained with Rose Bengal for 60 seconds. Nerve wrap is wrapped circumferentially around nerve ends and is illuminated with 532nm laser for 60 seconds. Nerve is rotated 180° and back wall illuminated for further 60 seconds.

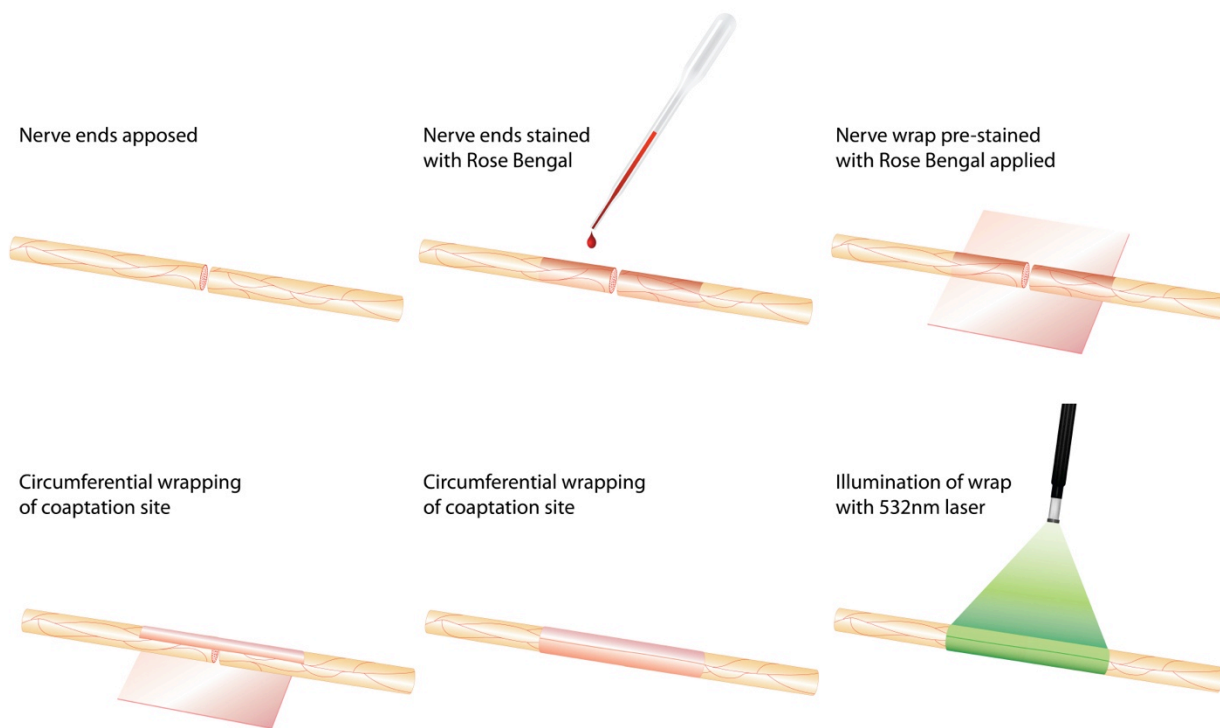
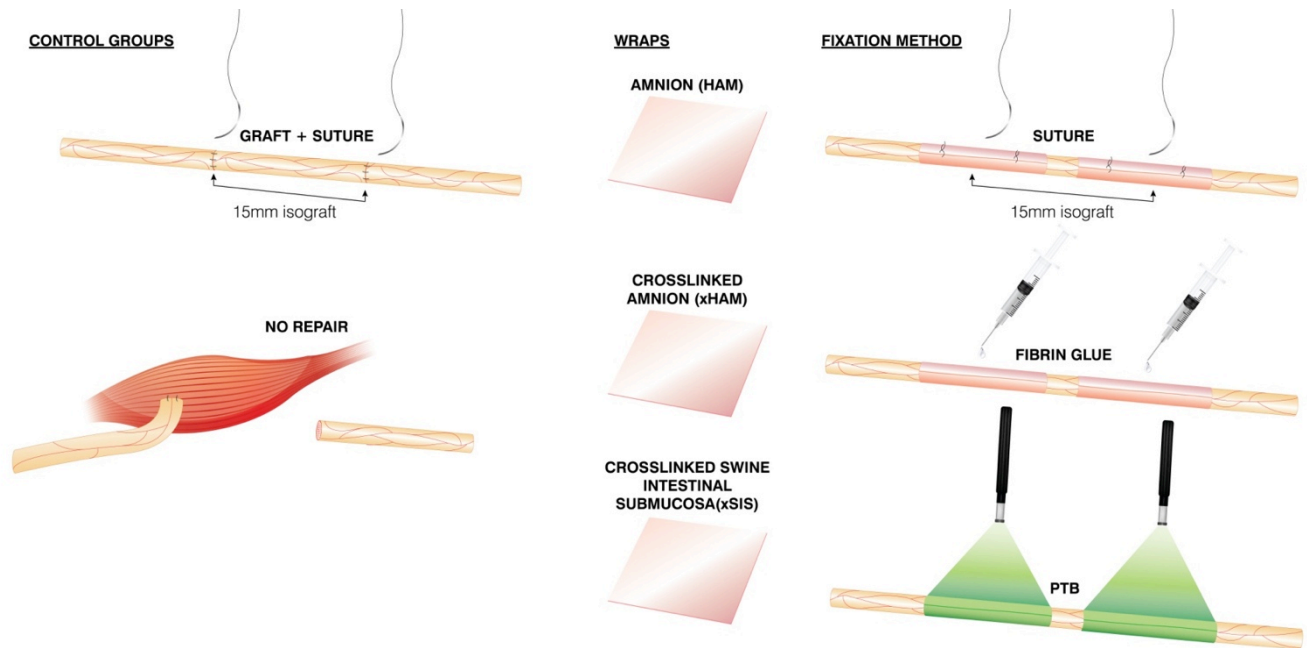


Figure 2. Methods of nerve repair for control and experimental groups. Positive controls had nerves repaired using isografts secured with standard epineurial suture. Negative controls had 15mm sections of nerve excised and no repair. Proximal nerve ends were buried into adjacent muscle and secured with 2x10.0 nylon suture. Nine experimental groups were composed of different combinations of the three



different nerve wraps and three different fixation methods illustrated.

Figure 3. Gross observations following sacrifice. (A) Graft+suture (B) HAM+PTB (C) xHAM+PTB. Crosslinked nerve wraps (xHAM+PTB) were still identifiable after 5-months, as evidence by the pink staining from Rose Bengal (C). In contrast, no evidence of non-crosslinked wraps, that had also been stained with Rose Bengal (HAM+PTB), could be found (B). Although not formally assessed, those nerves repaired photochemically developed considerably less scar tissue formation in comparison to those repairs performed using standard epineurial suture (B and C vs. A).

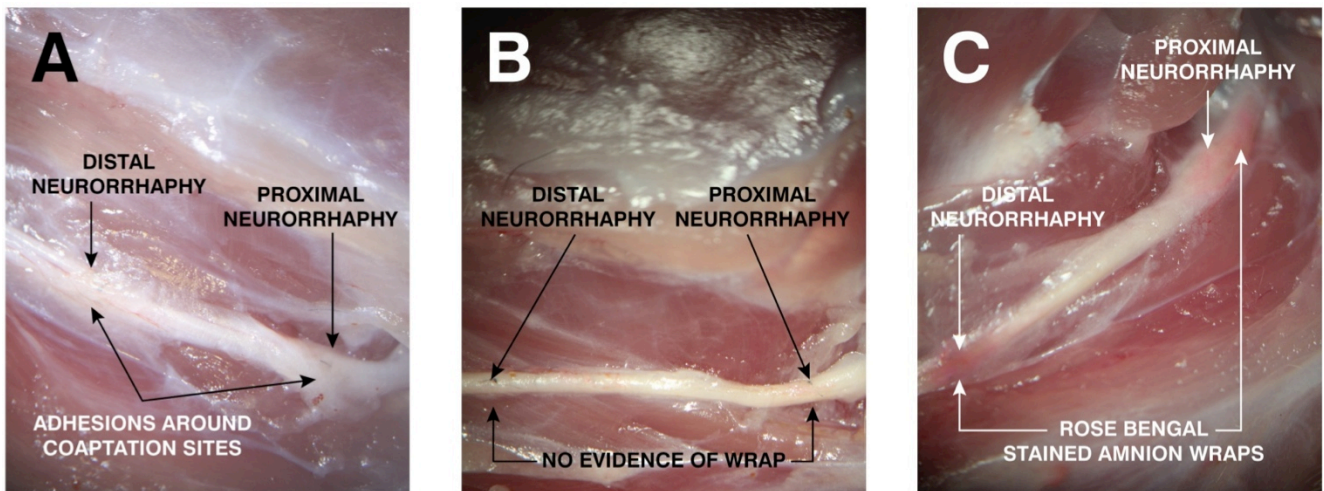


Figure 4. SFI for select groups. Those nerve repaired using xHAM+PTB recovered greatest SFI although this was not statistically significant compared with standard graft+suture (67.9+/-5.1 vs -71.7+/-4.8). Photochemical repairs using un-crosslinked amnion (HAM+PTB) performed less well than xHAM+PTB and standard graft+suture although this was not statistically significant (see table 2). xSIS+PTB performed least well out of all treatment groups.

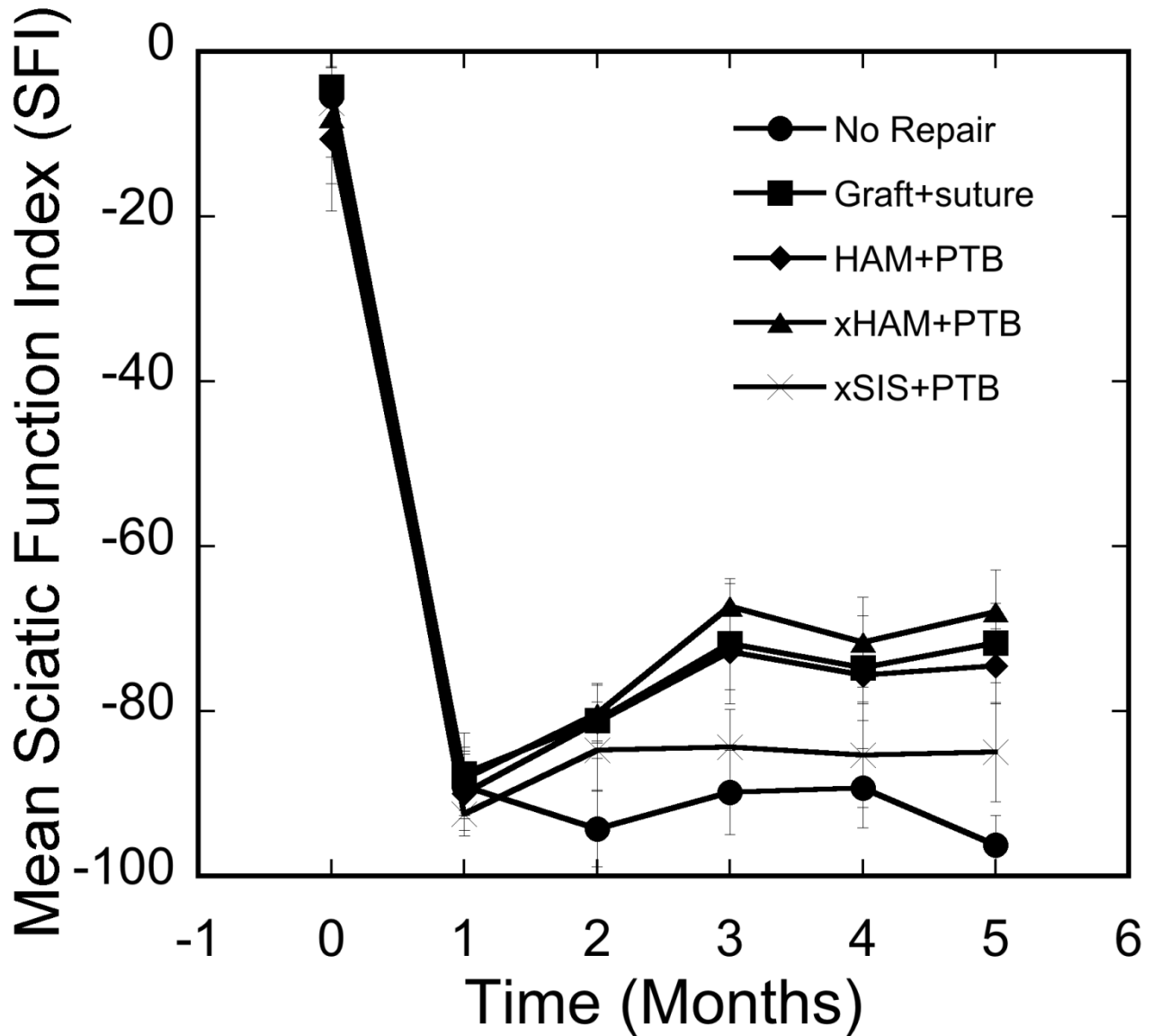


Figure 5. Gastrocnemius Muscle Mass Retention. Those nerves repaired using xHAM+PTB

recovered greatest gastrocnemius muscle mass and this was statistically significant in comparison to graft+suture (67.3+/-4.4 vs 60.0+/-5.2; p=0.02). Photochemical sealing of amnion wraps tended to outperform fibrin and suture fixation. This trend was not observed with xSIS, which performed least well out of all treatment groups.

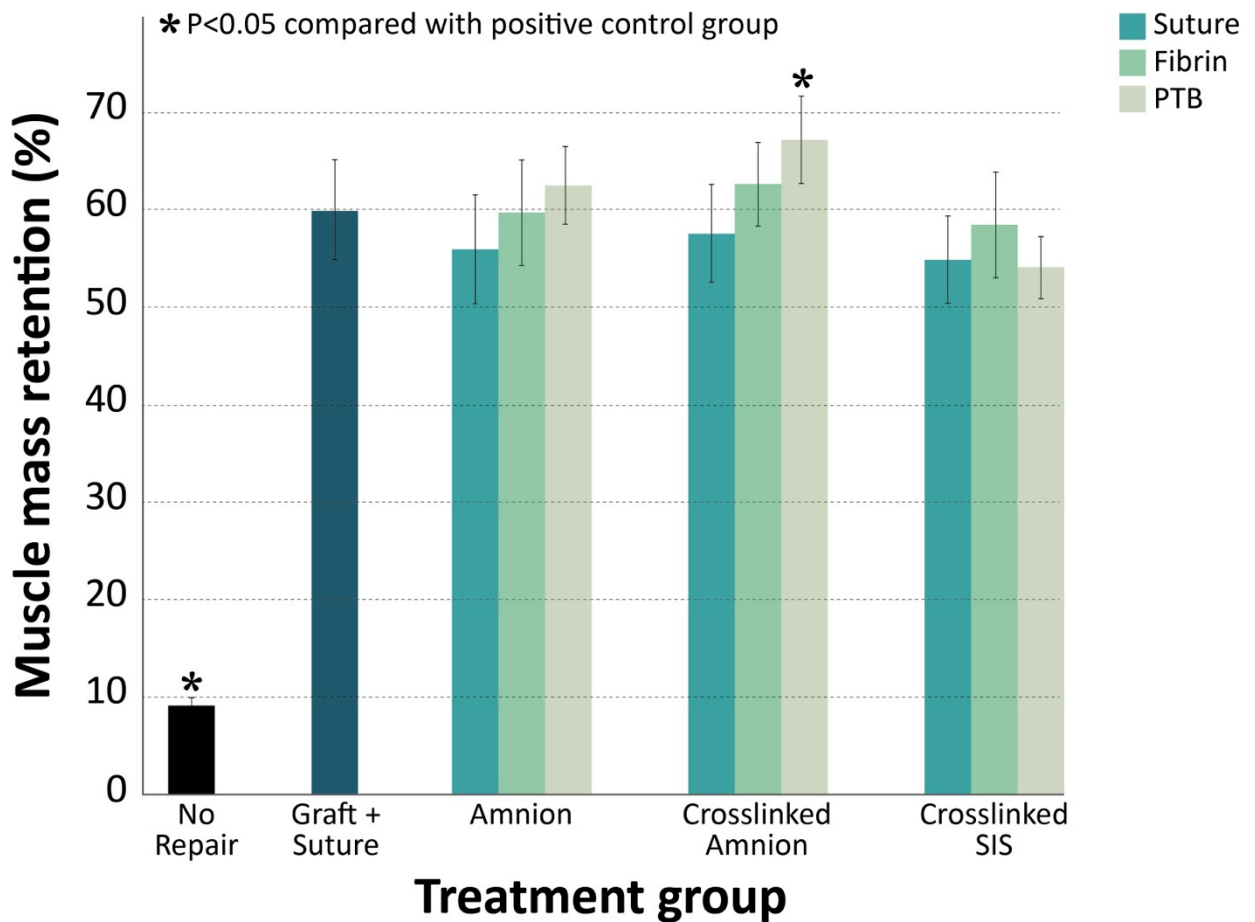
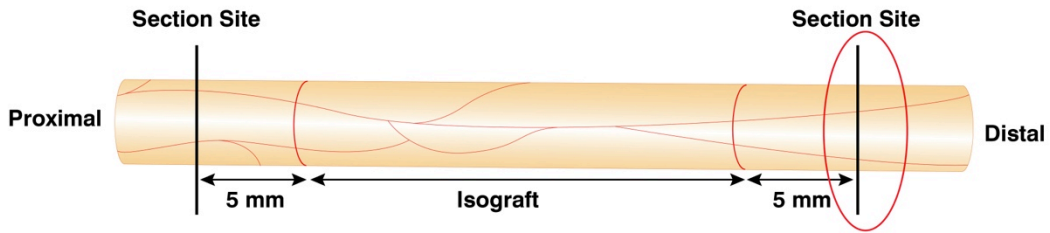
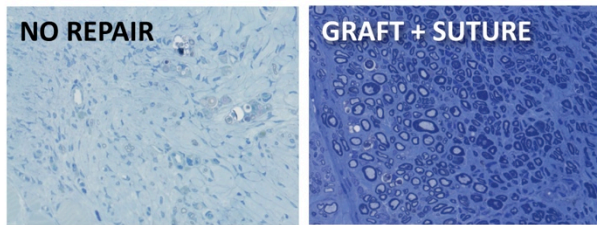


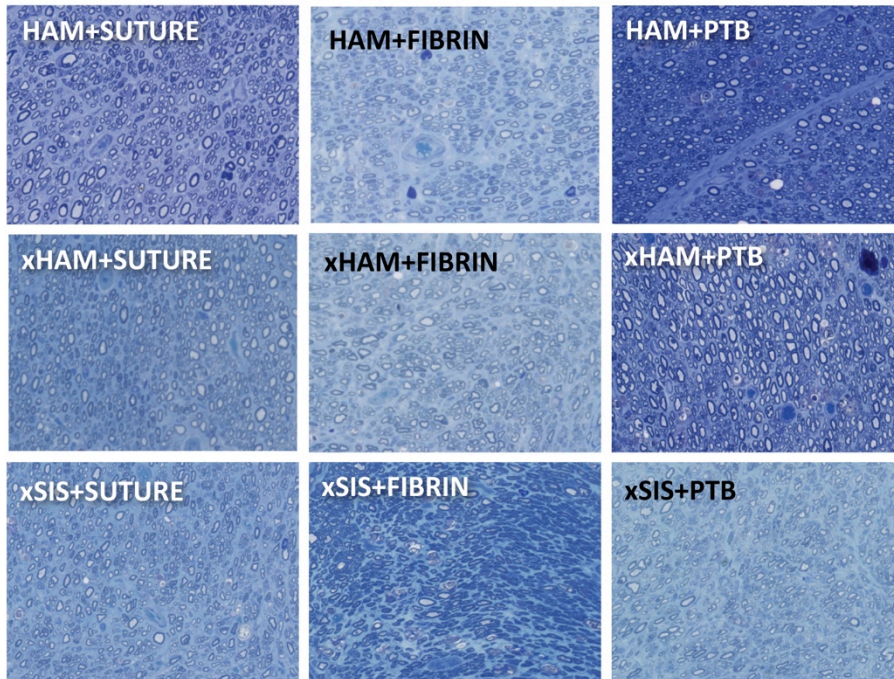
Figure 6. Histology slides from distal section site. Those nerves that were not repaired lacked axons. All treatment groups regenerated axons distal to isografts with the exception of those nerves that dehisced in the xSIS+PTB group. Axons in the xHAM+PTB group had statistically larger fiber diameter, axon diameter and myelin thickness in comparison to graft+suture (see table 4).



CONTROL GROUPS



TREATMENT GROUPS



Manuscript 3:

Large Gap Nerve Reconstruction Using Acellular Nerve Allografts And Photochemical
Tissue Bonding

Neil G. Fairbairn^{1,2}, Joanna Ng-Glazier^{1,2}, Amanda Meppelink², Mark A. Randolph²,
John M. Winograd², Mark E. Fleming³, Ian L. Valerio³ and Robert W. Redmond^{1*}.

¹ Wellman Center for Photomedicine, Harvard Medical School, Massachusetts General Hospital,
Boston, MA 02115.

² Division of Plastic Surgery, Department of Surgery, Harvard Medical School, Massachusetts General
Hospital, Boston, MA 02115.

³ Walter Reed National Military Medical Center, 8901 Wisconsin Avenue, Bethesda, MD 20814.

To whom correspondence should be addressed at:

Wellman Center for Photomedicine
Harvard Medical School
Massachusetts General Hospital
55 Fruit Street
Boston, MA 02115
Tel: 617 726-8857
E-mail: Redmond@helx.mgh.harvard.edu

INTRODUCTION:

The current gold standard method of repairing peripheral nerve injuries involving large gaps is the nerve autograft. However, the use of autografts necessitates the harvest of an “expendable” donor nerve and is associated with prolonged operating time, additional scarring, sensory loss and increased risk of neuroma formation. In cases of major trauma associated with substantial tissue loss, the demand for autograft may exceed that which can be harvested from the patient. Although rare in civilian practice, this scenario has become more frequent in recent years following the return of wounded warfighters from Iraq and Afghanistan who have sustained major nerve injury, often in addition to limb amputation. In these challenging patients, alternative strategies must be employed for nerve reconstruction.

Excluding nerve and tendon transfers, alternatives to nerve autograft for bridging gaps include conduits and nerve allografts. Conduits can be broadly categorized according to their biological or artificial composition and whether they are degradable or non-degradable. In the past, autogenous structures such as vein, artery and muscle have served as guidance channels but these techniques have equivalent limitations to that of autogenous nerve harvest. Alternative biological conduits are typically manufactured from components of the ECM such as collagen and fibrin and have theoretical advantages over synthetic varieties based on superior cell adhesion and predictable, non-toxic degradation. In spite of this, artificial conduits composed of biodegradable polymers and other novel materials have shown clinical efficacy and safety. However, the efficacy of inert, hollow conduits is limited to short gaps. Textured luminal surfaces, intraluminal fibers and guidance channels and the addition of schwann cells, neurotrophic factors and stem cells have attempted to improve efficacy but, despite increasingly elaborate strategies, outcomes are currently unable to match the nerve autograft.

Nerve allografts provide the most accurate representation of intraneural architecture and axonal guidance. Initially, allografts were fresh and cellularized, requiring immunosuppression to overcome rejection and preserve donor schwann cell viability over the course of recovery. Early experiments in immunosuppressed rodents and non-human primates produced outcomes that were comparable to

autograft⁽¹⁻⁶⁾. These observations were partly related to the pro-regenerative effect of immunosuppressant therapy. However, due to the risk of infective and neoplastic disease, this approach has been disfavored and has driven the development of decellularization protocols and the use of acellular nerve allograft (ANA). Initial attempts at decellularization involved thermal and chemical processing and, although technically successful, produced poor regenerative outcomes as a result of inadequate preservation of extracellular matrix (ECM) components and insufficient removal of cellular debris^(7, 8). In 2004, Hudson et al described an optimized detergent-based protocol that resulted in superior histological outcomes in comparison to those previously described methods^(9, 10). In 2007, human ANA (Avance, AxoGen Inc), processed by a modified detergent-based method, was approved for clinical use⁽¹¹⁾.

Although obviating the requirement and risks of immunosuppression, decellularization removes schwann cells (SCs) and other pro-regenerative components. The detrimental impact this has on the extent and rate of regeneration in comparison to conventional autograft has been consistently demonstrated in animal models of large gap injury^(7, 12). This long-standing impasse has prevented widespread clinical acceptance of ANA. Leaders in the field reserve ANA for small diameter, non-critical sensory nerve defects of less than 4cm, for restoring autograft donor site sensation, for nerve supercharging and for end-to-side nerve transfers in the hand. The use of ANA for the reconstruction of motor nerves, large diameter nerves, critical sensory nerves and sensory nerves greater than 4cm in length has been discouraged⁽¹³⁾.

This practice has been based on the scarcity of good quality clinical evidence and the fact that this evidence is often compounded by small sample sizes, the inclusion of sensory nerves only, small nerve gaps and the lack of autograft control groups⁽¹⁴⁻¹⁶⁾. However, in 2012, Brooks et al published results from the first comprehensive multi-centre trial and showed that Avance was safe and resulted in meaningful recovery in 87% of those cases recording quantitative outcomes⁽¹¹⁾. Cho et al have also

reported on results from the RANGER study and conclude that, for gaps between 5-50mm, ANAs can produce motor and sensory outcomes that are comparable to autograft and superior to hollow conduits (17).

Broadening the clinical application of ANAs and ultimately supplanting autograft provides strong impetus to improve regeneration through ANAs. Recent efforts have focused on the supplementation of ANA with neurotrophic factors, SCs and stem cells. Although conceptually exciting, preclinical experience has, up to now, been disappointing. Although a tissue engineered or cell-based solution is far from being realized, addressing the technical limitations of suture coaptation may offer a more simplistic and rapidly translatable solution.

The use of suture for neurorrhaphy has several well-known limitations. Suture material is inflammatory, results in fibrosis and ultimately leads to intra and extra-neural scar tissue formation. Scar tissue not only presents a direct obstacle to regenerating axons but can also lead to tethering and external compression, all of which can compromise outcome following injury. Even under high magnification and with meticulous surgical technique, coaptation sites are imperfect. Leakage of growth promoting factors and mis-guided axons into adjacent tissues further compromises outcome and likely contributes to neuroma formation. These effects are exacerbated in the context of gap injury and nerve grafting when axons must traverse two coaptation sites.

Sutureless repair can be achieved by a novel light-activated technique known as photochemical tissue bonding (PTB). Opposed nerve ends are wrapped circumferentially with human amnion that has been stained with a non-toxic, photoactive dye. Illumination of the nerve-wrap interface with a visible light source results in dye photoactivation, the formation of reactive species, subsequent crosslinking between amino acid residues and the creation of non-thermal, water-tight bonds⁽¹⁸⁻²¹⁾. In rodent models of simple end-to-end repair, this technique produced superior functional and histological outcomes in comparison

to conventional suture repair⁽²²⁻²⁵⁾. These observations are likely due to a reduction in intra- and extra-neural scar tissue, the prevention of axonal escape, and the enclosure and prevention of leakage of beneficial neuro-regenerative factors. Subsequent application of this technique in a rabbit model of large gap injury led to high rates of dehiscence, believed to be due to proteolytic degradation of amnion wraps and light-activated bonds during long periods of recovery and prior to the arrival of regenerating axons (unpublished work). Chemical crosslinking of amnion prior to light-activated sealing improves *ex-vivo* and *in vivo* nerve wrap durability. Crosslinked amnion and light-activated bonds were preserved for at least 5-months following repair and translated into superior muscle weight retention and histomorphometric outcomes in comparison to conventional graft and epineurial suture (manuscript submitted for publication). It is the aim of this study to test the efficacy of this strategy when used in conjunction with ANA. Our overarching goal is to optimize regeneration through ANAs and therefore improve recovery following large gap nerve injury, particularly those injuries complicated by severe multi-limb injury and amputation.

METHODS:

Allograft preparation

Sprague Dawley rats provided donor sciatic nerves. Following harvest, nerves were placed in PBS and stored at -80°C before being shipped to AxoGen Laboratories for decellularization. Once processed, nerves were returned and stored at -80°C until the day of surgery.

Human amnion (HAM) harvest and processing

Amniotic membrane was obtained from elective caesarean section patients who had been screened serologically for human immunodeficiency virus-1/2, Hepatitis B, Hepatitis C, human T-cell lymphotropic virus, syphilis, cytomegalovirus, and tuberculosis. Following delivery, amnion was bluntly removed from the placenta and washed liberally with phosphate buffered saline (PBS; Sigma-Aldrich, Co., St Louis, Mo). Membranes were mechanically de-epithelialized using a cell scraper, cut

into strips, wrapped around nitrocellulose paper and placed in a storage solution containing a 1:1 mix of 100% sterile glycerol and Dulbeccos modified Eagles medium (DMEM; Gibco, Grand Island, NY), penicillin-streptomycin-neomycin (PSN; Gibco, Grand Island, NY) and amphotericin B. HAM was stored at -80°C until required. Following thawing, HAM was mounted onto nitrocellulose paper before being dried and cut into 1cm x 1cm sections.

Nerve wrap crosslinking

Crosslinking solution was made by adding EDC/NHS (Sigma-Aldrich, Co., St Louis, Mo) to 2-(N-morpholino)ethanesulfonic (MES) acid buffer (Sigma-Aldrich, Co., St Louis, Mo). Nerve wraps were immersed in EDC/NHS for 1-hour on a platform shaker. Based on *ex vivo* experiments, optimal crosslinker concentration was 4mM EDC/1mM NHS (manuscript in progress).

Sciatic nerve injury and reconstruction

The Institutional Animal Care and Use Committee (IACUC) at the Massachusetts General Hospital approved all procedures. Twenty male inbred Lewis rats weighing 250-300g were randomized into two groups. In 1 group (n=10), nerve gaps were repaired using allografts secured with conventional epineurial suture. In the remaining group (n=10), nerves were repaired using allografts secured with photochemically sealed xHAM. Induction and maintenance anesthesia was achieved using isoflurane (Baxter Healthcare Corp. Deerfield IL; 5% induction/2-3% maintenance). A dorsolateral, muscle-splitting incision was made on the left hindquarter of each animal and, under the operating microscope, 15mm sections of sciatic nerve were excised and repaired by one of the following techniques. Wounds were closed in three layers with 4.0 vicryl (muscle and deep dermal) and 4.0 monocryl (subcuticular). Topical antibacterial ointment was applied liberally to wounds and the foot sprayed with bitter apple to discourage automutilation. Rodents were housed in the Massachusetts General Hospital small animal facility and had access to food and water as required.

Group 1- ANA+epineurial suture

Prior to anesthesia, allografts were thawed in a 37°C water bath. Following the excision of 15mm of sciatic nerve, allografts were placed into the field and cut to length, ensuring that no tension existed at either coaptation site. Grafts were secured with six 10.0 epineurial sutures at each site. Following repair, any protruding axons were trimmed and allowed to retract inside the repair (Fig 1).

Group 2 - ANA+PTB

Crosslinked amnion and nerve allografts were prepared as described above. Nerve grafts were tacked into place using two 10.0 epineurial sutures at each coaptation site. Prior to transfer into the surgical field, wraps and coaptation sites were stained with 0.1% Rose Bengal ((RB); Sigma-Aldrich, Co., St Louis, Mo) for 60 seconds. After 60 seconds, excess dye was removed. RB-stained wraps were wrapped circumferentially around sciatic nerves, creating a 5mm overlap either side of the coaptation. The area of overlap was irradiated for 60-seconds using a 532nm KTP laser (Laserscope, San Jose, Ca) at an irradiance of 0.5W/cm². The nerve/wrap was then rotated 180° in order to irradiate the back wall in the same manner for an additional 60-seconds (Fig 1).

Outcome assessment

Walking track analysis and calculation of sciatic function index

Walking track analysis was performed immediately prior to surgery for baseline sciatic function index (SFI). Following surgery, walking track analysis was performed at 30-day intervals. After dipping both hind paws in water soluble ink, rats were encouraged to walk up a 10 x 60 cm, partially enclosed ramp lined with white paper and set at an incline of 30° to horizontal. Measurements of print length, toe spread and intermediary toe spread were measured from the resulting prints using digital calipers. Mean values from three normal and experimental prints were entered into the SFI formula described by Bain and colleagues⁽²⁶⁾.

Gastrocnemius muscle mass retention

All rodents were sacrificed 150-days post-operatively by carbon dioxide inhalation. Left and right gastrocnemius muscles were harvested. Wet weights were recorded immediately and percentage muscle mass retention calculated.

Nerve histomorphometry

Following sacrifice, nerves were harvested 5mm proximal and distal to the graft and immediately fixed in a mixture of 2% glutaraldehyde/2% paraformaldehyde (Electron Microscopy Sciences, Hatfield, PA). After 48 hours, fixed nerves were washed in sodium cacodylate buffer (0.1M; pH=7.4) and post-fixed in 2% osmium tetroxide (Electron Microscopy Sciences, Hatfield, PA) for 2 hours. Following further washing in sodium cacodylate buffer, specimens were dehydrated in increasing concentrations of ethanol (25%, 50%, 75%, 95%, 100%). Following dehydration, all specimens were washed with propylene oxide (Electron Microscopy Sciences, Hatfield, PA). Specimens were then placed in increasing concentrations of Epoxy resin (DDSA (dodecyl succinic anhydrides 98+%; Free Acid 2%)/tEPON-812 (Epoxy Resin)/NMA Ultrapure (methyl-5-norbornene-2,3-dicarboxylic anhydride)/DMP-30 (2,4,6-tri (dimethylaminomethyl) phenol (Tousimis Research Corporation, Rockville, MD) before being baked overnight in an oven at 60°C. Using a diamond blade, 1µm sections were cut 5mm proximal and 5mm distal to the graft. Sections were stained with toluidine blue and were mounted and cover-slipped in preparation for histomorphometric analysis.

Histology slides were scanned using a Hamamatsu NanoZoomer 2.0-HT slide scanner (Meyer Instruments, Houston TX) and read using NDP.com software (Hamamatsu Corp. Bridgewater, NJ). Nerve area was calculated at 40x magnification. From these images, five 400x images were randomly selected and imported into Adobe Photoshop. Axon counts were measured and from fifty randomly (randomizer.org) selected fibers (250 nerve fibers per location), fiber diameter, axon diameter, myelin thickness and G-ratio were calculated.

Comparative analysis of ANA vs Isograft

This current work represents phase 2 of a large, three-phase study and builds on the recently successful application of photochemically sealed isografts for large gap nerve repair (phase 1 manuscript submitted for publication). Phase 1 isograft experiments used exactly the same animal model as found in this current study, with isografts taking the place of ANA. In order to avoid repetition of experimental groups and unnecessary animal morbidity, and to extend the clinical relevance of this current study, we have elected to include outcomes from two groups of animals from the phase 1 isograft study. One group (n=10) consisting of isografts secured with conventional suture (gold standard isograft+suture) and the optimal repair group (n=10) consisting of isografts secured with photochemically sealed crosslinked amnion wraps (isograft+PTB) are compared to the two ANA groups described above.

Statistical analysis

Statistical analysis was performed using KaleidaGraph for Windows v4.1 (Synergy Software, Reading, PA). Testing between experimental groups was achieved using ANOVA and the post hoc Bonferroni test. Statistical significance was set at <0.05 .

RESULTS

Sciatic nerve reconstruction

Gross observation following sacrifice

There were no cases of nerve dehiscence in either ANA group and no animals suffered from foot ulceration or automutilation. In those nerves repaired photochemically, remnants of the RB-stained amnion nerve wraps were evident. As with previous studies, extra-neural scar tissue formation was obvious in those nerves repaired using conventional suture and was qualitatively less in photochemically bonded nerves (Fig 2). Qualitatively, allografts were of smaller diameter than isografts.

Sciatic function index

Isograft+PTB from phase 1 recovered greatest SFI after 5-months follow-up (Table 1; Fig 3). This was not statistically significant in comparison to isograft+suture. Likewise, photochemically sealed ANA showed a non-statistically significant improvement in comparison to sutured ANA (-80.3+/-4.2 vs. -78.3+/-5.0; Table 1 and 4; Fig 3). ANA+suture performed statistically worse than isograft+suture (-80.3+/-4.2 vs. -71.7+/-4.8; p=0.0019; Table 1 and 4). SFI was also statistically less for ANA+PTB in comparison to isograft+suture and isograft+PTB (Table 1 and 4).

Gastrocnemius muscle mass retention

Isograft+PTB recovered greatest gastrocnemius muscle mass retention and this was statistically significant in comparison to all other groups (Table 2). Although ANA+PTB displayed a trend towards greater muscle mass recovery in comparison to ANA+suture, this result was not statistically significant (55.2+/-5.5% vs. 52.9+/-4.77%; Table 2, Fig 4). Muscle mass recovery was statistically poorer in ANA+suture group in comparison to isograft+suture group. Likewise, recovery of ANA+PTB was statistically poorer than isograft+PTB. Muscle mass retention in the ANA+PTB group was statistically comparable to that achieved using gold standard isograft+suture.

Nerve Histomorphometry

Axon counts in the distal nerve stump were significantly greater for isograft+PTB in comparison to ANA+suture. No other significant differences in axon counts or axon density existed between treatment groups. Isograft+PTB recovered the greatest fiber diameter, axon diameter and myelin thickness and this was statistically significant in comparison to all other groups (Table 3 and 4; Fig 5). Histomorphometric recovery was poorest in the ANA+suture group and this was statistically significant in comparison to isograft+suture and isograft+PTB. Although not statistically significant, fiber diameter, axon diameter and myelin thickness in the ANA+PTB group displayed a trend towards greater recovery in comparison to ANA+suture. There was no significant difference between ANA+PTB and gold standard isograft+suture (Table 3 and 4; Fig 5)

DISCUSSION

This study shows that outcomes following light-activated sealing of ANA are improved in comparison to sutured ANA, making them statistically equivalent to gold standard sutured isografts. Although the difference between photochemically bonded and sutured ANA was not statistically significant, the trend towards improvement is consistent with recent experience applying this technique to isograft reconstruction. Based on these findings, isografts that are photochemically sealed with crosslinked amnion nerve wraps have emerged as the superior reconstructive method for large gap nerve repair. However, the ability to elevate the performance of ANA to match the current gold standard is of major clinical interest when severe injuries, complicated with limb loss, preclude the use of autograft. The demonstrated success of light-activated sealing of isograft coaptation sites is likely related to the creation of a water-tight seal and the subsequent containment of the neurotrophic rich milieu. It is possible that removal of SCs, and the neurotrophic factors they liberate, during decellularization of ANAs, may partially abrogate this effect. However, the non-significant benefit observed suggests that the technique remains advantageous. The avoidance of suture-induced inflammation and fibrosis, the exclusion of infiltrating scar tissue and the prevention of axonal escape may be responsible.

Allografts were of excellent quality. Microscopic examination revealed no evidence of epineurial fragmentation and fascicular architecture appeared intact. The grafts were pliable and resembled autogenous tissue. Although the use of Avance® processed human cadaveric nerve is desirable in order to maximize clinical relevance, recent findings from Wood et al showed that, in rodent models of nerve injury, regeneration through rat ANA was statistically superior to Avance®⁽²⁷⁾. By definition, ANAs should avoid immunorejection. However, Gulati et al showed that acellular allografts were more immunogenic in comparison to acellular isografts. Although both successfully supported regeneration, outcomes were inferior to that of cellular isografts⁽²⁸⁾. Kvist et al performed comparative analysis between allografts from various species when used to reconstruct rodent nerve gaps and detected significant differences in outcome⁽²⁹⁾. It is possible that cross-species xenografts incite a minor,

subclinical inflammatory response that is sufficient to compromise recovery. In light of this possibility, rat-processed ANA was used in this study in favour of Avance®. ANAs from Sprague Dawley rats were used to repair nerve gaps in Lewis rats. It is uncertain whether cross-strain transplantation had any detrimental impact on regeneration. As with previous studies, the formation of extraneural scar tissue was qualitatively less with photochemical repair in comparison to suture repair (Fig 2).

SFI, muscle mass retention, fiber diameter and axon diameter, following the use of sutured ANA, were statistically poorer than sutured isografts after 5-months follow-up. These results are discordant with Whitlock et al who showed that, in a rodent model, there was no significant difference in outcome between isograft and allograft reconstruction of a 14mm sciatic nerve gap after 12 weeks, although results were significant after 6 weeks⁽³⁰⁾. A significant difference was detected after 16 weeks when nerve gaps were extended to 28mm. Interestingly, a comparative analysis by Moore et al showed that, although outcomes using a detergent processed allograft were equivalent to isograft, AxoGen processed rat allograft were inferior to isograft⁽³¹⁾. The 15mm gap used in this study was sufficient to detect significant differences between isograft and ANAs for the aforementioned outcomes although axon count, myelin thickness and G-ratio were not significantly different. It is possible that if a greater than 15mm gap had been used in this study, the observed differences between isograft and ANA, and between sutured and light activated sealing of ANA, may have been more apparent.

Following decellularization, ANAs consist of basal lamina scaffolds. Components of the basal lamina such as fibronectin and laminin have pro-regenerative effects on neurite outgrowth^(32, 33) and can support axonal regeneration in the absence of schwann cells⁽³³⁻³⁹⁾. Whilst this may be sufficient over short lengths of ANA, successful regeneration across longer lengths is dependent on re-population of ANA by resident schwann cells. Early evidence of the diminishing regenerative return with increasing graft length was reported by Gulati⁽⁷⁾. Contemporary studies by Saheb-Al-Zamani et al emphasized the finite migratory and proliferative capacity of SCs and showed that repopulation of increasingly long

ANAs by recipient SCs induces a state of proliferative duress that can lead to SC senescence. Axonal regeneration and functional recovery declined with increasing graft length and isografts outperformed ANAs at all lengths⁽⁴⁰⁾. This study did not assess the extent of SC re-population but it is possible that photochemical sealing may augment SC migration, further improving regeneration through long ANAs. Investigating SC repopulation in ANAs of varying length may form the basis of future investigation. When combined with cell-based therapy and tissue engineering, sealing the regenerative milieu and maintaining high levels of growth promoting factors at the repair site, may amplify this effect.

Ideally, both isograft groups included in this study would have been repeated with the ANA groups. By including data from phase 1 in the comparative analysis, particularly the isograft+PTB group as the “optimal” repair method, an element of selection bias has been introduced. An assumption has been made in this case that the outcomes in these particular groups would have been replicated if they had been repeated. The decision not to repeat these groups was rationalized by the desire to avoid unnecessary animal experimentation and in light of the growing body of evidence supporting this light-activated technique.

CONCLUSION

Photochemical sealing of isografts has emerged as the optimal method of reconstruction for a 15mm rodent sciatic nerve gap. SFI, muscle mass retention, fiber diameter and axon diameter are statistically poorer following the use of sutured ANA in comparison to sutured isografts. Photochemical sealing of ANAs led to non-significant improvements of SFI, gastrocnemius muscle mass retention and nerve histomorphometry, in comparison to sutured ANA, that was statistically comparable to gold standard sutured isografts. These results build on the recent successful application of this technique for isograft reconstruction. Although subtle, if these improvements translate clinically, this could result in important improvements in peripheral nerve recovery in those cases of severe trauma and limb loss where the use of nerve autograft is not possible. With refinement, and with parallel advances in stem cell therapy and

tissue engineering, this technique, when used in conjunction with ANA, has the potential to completely supplant the use of autografts following large gap injury.

ACKNOWLEDGEMENTS

Financial support for this study was provided by the Peer-Reviewed Orthopedic Research Program (PRORP) through USAMRAA award W81XWH-12-1-0511.

REFERENCES

1. Bain JR, Mackinnon SE, Hudson AR, Falk RE, Falk JA, Hunter DA. The peripheral nerve allograft: an assessment of regeneration across nerve allografts in rats immunosuppressed with cyclosporin A. *Plastic and reconstructive surgery*. 1988 Dec;82(6):1052-66. PubMed PMID: 3264409. Epub 1988/12/01. eng.
2. Bain JR, Mackinnon SE, Hudson AR, Wade J, Evans P, Makino A, et al. The peripheral nerve allograft in the primate immunosuppressed with Cyclosporin A: I. Histologic and electrophysiologic assessment. *Plastic and reconstructive surgery*. 1992 Dec;90(6):1036-46. PubMed PMID: 1448498. Epub 1992/12/01. eng.
3. Fish JS, Bain JR, McKee N, Mackinnon SE. The peripheral nerve allograft in the primate immunosuppressed with Cyclosporin A: II. Functional evaluation of reinnervated muscle. *Plastic and reconstructive surgery*. 1992 Dec;90(6):1047-52. PubMed PMID: 1448499. Epub 1992/12/01. eng.
4. Weinzweig N, Grindel S, Gonzalez M, Kuy D, Fang J, Shahani B. Peripheral-nerve allotransplantation in rats immunosuppressed with transient or long-term FK-506. *Journal of reconstructive microsurgery*. 1996 Oct;12(7):451-9. PubMed PMID: 8905545. Epub 1996/10/01. eng.
5. Mackinnon SE, Midha R, Bain J, Hunter D, Wade J. An assessment of regeneration across peripheral nerve allografts in rats receiving short courses of cyclosporin A immunosuppression. *Neuroscience*. 1992;46(3):585-93. PubMed PMID: 1545911. Epub 1992/01/01. eng.
6. Midha R, Mackinnon SE, Evans PJ, Best TJ, Hare GM, Hunter DA, et al. Comparison of regeneration across nerve allografts with temporary or continuous cyclosporin A immunosuppression. *Journal of neurosurgery*. 1993 Jan;78(1):90-100. PubMed PMID: 8416248. Epub 1993/01/01. eng.

7. Gulati AK. Evaluation of acellular and cellular nerve grafts in repair of rat peripheral nerve. *Journal of neurosurgery*. 1988 Jan;68(1):117-23. PubMed PMID: 3335896. Epub 1988/01/01. eng.
8. Sondell M, Lundborg G, Kanje M. Regeneration of the rat sciatic nerve into allografts made acellular through chemical extraction. *Brain research*. 1998 Jun 8;795(1-2):44-54. PubMed PMID: 9622591. Epub 1998/06/12. eng.
9. Hudson TW, Liu SY, Schmidt CE. Engineering an improved acellular nerve graft via optimized chemical processing. *Tissue engineering*. 2004 Sep-Oct;10(9-10):1346-58. PubMed PMID: 15588395. Epub 2004/12/14. eng.
10. Hudson TW, Zawko S, Deister C, Lundy S, Hu CY, Lee K, et al. Optimized acellular nerve graft is immunologically tolerated and supports regeneration. *Tissue engineering*. 2004 Nov-Dec;10(11-12):1641-51. PubMed PMID: 15684673. Epub 2005/02/03. eng.
11. Brooks DN, Weber RV, Chao JD, Rinker BD, Zoldos J, Robichaux MR, et al. Processed nerve allografts for peripheral nerve reconstruction: a multicenter study of utilization and outcomes in sensory, mixed, and motor nerve reconstructions. *Microsurgery*. 2012 Jan;32(1):1-14. PubMed PMID: 22121093. Epub 2011/11/29. eng.
12. Giusti G, Willems WF, Kremer T, Friedrich PF, Bishop AT, Shin AY. Return of motor function after segmental nerve loss in a rat model: comparison of autogenous nerve graft, collagen conduit, and processed allograft (AxoGen). *The Journal of bone and joint surgery American volume*. 2012 Mar 7;94(5):410-7. PubMed PMID: 22398734. Epub 2012/03/09. eng.
13. Boyd KU, Nimigan AS, Mackinnon SE. Nerve reconstruction in the hand and upper extremity. *Clinics in plastic surgery*. 2011 Oct;38(4):643-60. PubMed PMID: 22032591. Epub 2011/10/29. eng.

14. Karabekmez FE, Duymaz A, Moran SL. Early clinical outcomes with the use of decellularized nerve allograft for repair of sensory defects within the hand. *Hand (New York, NY)*. 2009 Sep;4(3):245-9. PubMed PMID: 19412640. Pubmed Central PMCID: PMC2724628. Epub 2009/05/05. eng.
15. Shanti RM, Ziccardi VB. Use of decellularized nerve allograft for inferior alveolar nerve reconstruction: a case report. *Journal of oral and maxillofacial surgery : official journal of the American Association of Oral and Maxillofacial Surgeons*. 2011 Feb;69(2):550-3. PubMed PMID: 21145638. Epub 2010/12/15. eng.
16. Gunn S, Cosetti M, Roland JT, Jr. Processed allograft: novel use in facial nerve repair after resection of a rare racial nerve paraganglioma. *The Laryngoscope*. 2010;120 Suppl 4:S206. PubMed PMID: 21225804. Epub 2011/01/13. eng.
17. Cho MS, Rinker BD, Weber RV, Chao JD, Ingari JV, Brooks D, et al. Functional outcome following nerve repair in the upper extremity using processed nerve allograft. *The Journal of hand surgery*. 2012 Nov;37(11):2340-9. PubMed PMID: 23101532. Epub 2012/10/30. eng.
18. Chan BP, Hui TY, Chan OC, So KF, Lu W, Cheung KM, et al. Photochemical cross-linking for collagen-based scaffolds: a study on optical properties, mechanical properties, stability, and hemocompatibility. *Tissue engineering*. 2007 Jan;13(1):73-85. PubMed PMID: 17518582. Epub 2007/05/24. eng.
19. Kochevar IE, Redmond RW. Photosensitized production of singlet oxygen. *Methods in enzymology*. 2000;319:20-8. PubMed PMID: 10907495. Epub 2000/07/25. eng.

20. Balasubramanian D, Du X, Zigler JS, Jr. The reaction of singlet oxygen with proteins, with special reference to crystallins. *Photochemistry and photobiology*. 1990 Oct;52(4):761-8. PubMed PMID: 2089424. Epub 1990/10/01. eng.
21. Webster A, Britton D, Apap-Bologna A, Kemp G. A dye-photosensitized reaction that generates stable protein-protein crosslinks. *Analytical biochemistry*. 1989 May 15;179(1):154-7. PubMed PMID: 2757189. Epub 1989/05/15. eng.
22. Henry FP, Goyal NA, David WS, Wes D, Bujold KE, Randolph MA, et al. Improving electrophysiologic and histologic outcomes by photochemically sealing amnion to the peripheral nerve repair site. *Surgery*. 2009 Mar;145(3):313-21. PubMed PMID: 19231584. Epub 2009/02/24. eng.
23. Johnson TS, O'Neill AC, Motarjem PM, Amann C, Nguyen T, Randolph MA, et al. Photochemical tissue bonding: a promising technique for peripheral nerve repair. *The Journal of surgical research*. 2007 Dec;143(2):224-9. PubMed PMID: 17543988. Epub 2007/06/05. eng.
24. O'Neill AC, Randolph MA, Bujold KE, Kochevar IE, Redmond RW, Winograd JM. Preparation and integration of human amnion nerve conduits using a light-activated technique. *Plastic and reconstructive surgery*. 2009 Aug;124(2):428-37. PubMed PMID: 19644257. Epub 2009/08/01. eng.
25. O'Neill AC, Randolph MA, Bujold KE, Kochevar IE, Redmond RW, Winograd JM. Photochemical sealing improves outcome following peripheral neurorrhaphy. *The Journal of surgical research*. 2009 Jan;151(1):33-9. PubMed PMID: 18599081. Epub 2008/07/05. eng.
26. Bain JR, Mackinnon SE, Hunter DA. Functional evaluation of complete sciatic, peroneal, and posterior tibial nerve lesions in the rat. *Plastic and reconstructive surgery*. 1989 Jan;83(1):129-38. PubMed PMID: 2909054. Epub 1989/01/01. eng.

27. Wood MD, Kemp SW, Liu EH, Szykaruk M, Gordon T, Borschel GH. Rat-derived processed nerve allografts support more axon regeneration in rat than human-derived processed nerve xenografts. *Journal of biomedical materials research Part A*. 2014 Apr;102(4):1085-91. PubMed PMID: 23630071. Epub 2013/05/01. eng.
28. Gulati AK, Cole GP. Nerve graft immunogenicity as a factor determining axonal regeneration in the rat. *Journal of neurosurgery*. 1990 Jan;72(1):114-22. PubMed PMID: 2294170. Epub 1990/01/01. eng.
29. Kvist M, Sondell M, Kanje M, Dahlin LB. Regeneration in, and properties of, extracted peripheral nerve allografts and xenografts. *Journal of plastic surgery and hand surgery*. 2011 Jun;45(3):122-8. PubMed PMID: 21682608. Epub 2011/06/21. eng.
30. Whitlock EL, Tuffaha SH, Luciano JP, Yan Y, Hunter DA, Magill CK, et al. Processed allografts and type I collagen conduits for repair of peripheral nerve gaps. *Muscle & nerve*. 2009 Jun;39(6):787-99. PubMed PMID: 19291791. Epub 2009/03/18. eng.
31. Moore AM, MacEwan M, Santosa KB, Chenard KE, Ray WZ, Hunter DA, et al. Acellular nerve allografts in peripheral nerve regeneration: a comparative study. *Muscle & nerve*. 2011 Aug;44(2):221-34. PubMed PMID: 21660979. Pubmed Central PMCID: PMC3136642. Epub 2011/06/11. eng.
32. Akers RM, Mosher DF, Lilien JE. Promotion of retinal neurite outgrowth by substratum-bound fibronectin. *Developmental biology*. 1981 Aug;86(1):179-88. PubMed PMID: 7286392. Epub 1981/08/01. eng.

33. Ide C, Tohyama K, Yokota R, Nitatori T, Onodera S. Schwann cell basal lamina and nerve regeneration. *Brain research*. 1983 Dec 12;288(1-2):61-75. PubMed PMID: 6661636. Epub 1983/12/12. eng.
34. Hall SM. Regeneration in cellular and acellular autografts in the peripheral nervous system. *Neuropathology and applied neurobiology*. 1986 Jan-Feb;12(1):27-46. PubMed PMID: 3703154. Epub 1986/01/01. eng.
35. Ide C. Nerve regeneration and Schwann cell basal lamina: observations of the long-term regeneration. *Archivum histologicum Japonicum = Nihon soshikigaku kiroku*. 1983 Apr;46(2):243-57. PubMed PMID: 6882155. Epub 1983/04/01. eng.
36. Fawcett JW, Keynes RJ. Muscle basal lamina: a new graft material for peripheral nerve repair. *Journal of neurosurgery*. 1986 Sep;65(3):354-63. PubMed PMID: 3734886. Epub 1986/09/01. eng.
37. Keynes RJ, Hopkins WG, Huang LH. Regeneration of mouse peripheral nerves in degenerating skeletal muscle: guidance by residual muscle fibre basement membrane. *Brain research*. 1984 Mar 19;295(2):275-81. PubMed PMID: 6713188. Epub 1984/03/19. eng.
38. Bryan DJ, Miller RA, Costas PD, Wang KK, Seckel BR. Immunocytochemistry of skeletal muscle basal lamina grafts in nerve regeneration. *Plastic and reconstructive surgery*. 1993 Oct;92(5):927-40. PubMed PMID: 8415975. Epub 1993/10/01. eng.
39. Glasby MA, Gilmour JA, Gschmeissner SE, Hems TE, Myles LM. The repair of large peripheral nerves using skeletal muscle autografts: a comparison with cable grafts in the sheep femoral nerve. *British journal of plastic surgery*. 1990 Mar;43(2):169-78. PubMed PMID: 2328378. Epub 1990/03/01. eng.

40. Saheb-Al-Zamani M, Yan Y, Farber SJ, Hunter DA, Newton P, Wood MD, et al. Limited regeneration in long acellular nerve allografts is associated with increased Schwann cell senescence. *Experimental neurology*. 2013 Sep;247:165-77. PubMed PMID: 23644284. Pubmed Central PMCID: PMC3863361. Epub 2013/05/07. eng.

TABLES

Table 1. Mean SFI for all treatment groups over 5-month follow-up period. At each time point throughout recovery, no significant differences existed within each of the isograft and ANA groups. Recovery of SFI was statistically poorer for both sutured and photochemically sealed ANA in comparison to sutured and photochemically sealed isograft at each time point.

Experimental group	Mean SFI				
	1-month	2-month	3-month	4-month	5-month
Isograft+suture	-87.6+/-5.0	-81.1+/-4.5	-71.8+/-7.3	-74.7+/-6.3	-71.7+/-4.8
Isograft+PTB	-88.2+/-3.9	-80.3+/-3.5	-67.2+/-3.3	-71.6+/-5.5	-67.9+/-5.1
ANA+suture	-95.4+/-2.5	-90.3+/-10.6	-87.9+/-4.0	-84.1+/-3.2	-80.3+/-4.2
ANA+PTB	-93.4+/-3.4	-91.1+/-5.4	-88.9+/-5.4	-83.4+/-4.8	-78.3+/-5.0

Table 2. Gastrocnemius muscle mass retention for all groups. Isograft+PTB recovered significantly greater muscle mass in comparison to isograft+suture. Muscle mass retention was significantly poorer in the ANA+suture group in comparison to isograft+suture. Retention in the ANA+PTB group was comparable to isograft+suture.

Experimental group	Mean left gastrocnemius muscle mass retention (%)	SD	P value*
Isograft+suture	60.0	5.2	----
Isograft+PTB	67.3	4.4	0.01
ANA+suture	52.9	4.77	0.02
ANA+PTB	55.2	5.5	0.22

Table 3. Histomorphometric analysis for all treatment groups. Isograft+PTB recovered significantly greater fiber diameter, axon diameter and myelin thickness than all other groups. With the exception of G-ratio, no significant differences existed between ANA+suture and ANA+PTB. ANA+suture recovered significantly less fiber diameter and axon diameter than isograft+suture. There was no difference in axon count, myelin thickness and G-ratio between these groups. ANA+PTB was statistically comparable to isograft+suture for all histomorphometric parameters (see Bonferroni all-pairs comparison (Table 4)).

Histomorphometric parameters 5mm distal from distal graft coaptation site (Mean+/-SD)						
Experimental Group	Total axon count (x0.001)	Axon Density (mm ² x 0.001)	Nerve fiber diameter (µm)	Axon diameter (µm)	Myelin thickness (µm)	G-ratio
Isograft+Suture	7.61+/-3.42	29.36+/-18.10	5.47+/-1.70	3.50+/-1.44	1.96+/-0.47	0.62+/-0.08
Isograft+PTB	9.66+/-3.08	30.73+/-14.73	6.87+/-2.23*	4.51+/-1.83*	2.35+/-0.64*	0.64+/-0.08
ANA+suture	5.04+/-2.57	21.50+/-2.56	5.26+/-1.29	3.30+/-1.15	1.76+/-0.86	0.62+/-0.12
ANA+PTB	6.04+/-3.20	22.03+/-5.15	5.38+/-1.22	3.41+/-0.99	1.97+/-0.69	0.63+/-0.11

Table 4. Bonferroni all-pairs comparison for all treatment groups. In comparison to isograft

+suture, isograft+PTB results in statistically significant improvements in muscle mass retention, fiber diameter, axon diameter and myelin thickness. ANA+suture had significantly poorer recovery of SFI, muscle mass retention, fiber diameter and axon diameter. With the exception of G-ratio, there was no significant difference between ANA+suture and ANA+PTB for any of the outcomes measured. There was no significant difference in muscle mass retention, axon count, fiber diameter, axon diameter, myelin thickness and G-ratio between ANA+PTB and isograft+suture.

Group Comparison	SFI (5-month)	Muscle Mass	Axon Count	Fiber Diameter	Axon Diameter	Myelin Thickness	G-ratio
Isograft+suture vs Isograft+PTB	0.59	0.01	0.90	< 0.0001	< 0.0001	< 0.0001	< 0.0001
Isograft+suture vs ANA+suture	0.002	0.02	0.47	< 0.0001	< 0.0001	1	1
Isograft+suture vs ANA+PTB	0.03	0.22	1	0.27	0.08	1	0.21
Isograft+PTB vs ANA+suture	< 0.0001	< 0.0001	0.02	< 0.0001	< 0.0001	< 0.0001	< 0.0001
Isograft+PTB vs ANA+PTB	0.0002	< 0.0001	0.08	< 0.0001	< 0.0001	< 0.0001	0.0005
ANA+suture vs ANA+PTB	1	1	1	0.07	0.06	1	0.0062

FIGURE LEGENDS

Figure 1. Methods of rodent sciatic nerve injury and repair. All left sciatic nerves had 15mm sections excised. These gaps were reconstructed with sections of processed rat ANA. In one group (n=10), ANAs were secured with conventional 10.0 ethilon suture. In the remaining group (n=10), ANAs were secured using photochemically sealed crosslinked amnion nerve wraps.

Figure 2. Gross observations following sacrifice. All nerves were found to be intact and showed evidence of regeneration across both isografts and ANA. Qualitatively, sutured isografts (A) and sutured ANA (D) had greater adhesion formation around coaptation sites. Photochemically sealed isografts (B) and photochemically sealed ANA (C) showed remnants of RB stained amnion wraps. Qualitatively, sutured ANA diameter was noticeably reduced in comparison to isograft, an observation consistent with their statistically inferior outcome.

Figure 3. Sciatic Function Index of ANA and isograft treated groups. After 5-months follow-up, no significant difference existed within either isograft or ANA groups. ANA+suture and ANA+PTB groups recovered statistically less SFI in comparison to both isograft groups.

Figure 4. Gastrocnemius Muscle Mass Retention for ANA and isograft treated groups. Isografts sealed photochemically with crosslinked amnion were statistically superior to all other groups. No significant difference existed between sutured and photochemically sealed ANA. ANA+suture was recovered statistically less muscle mass than isograft+suture but there was no significant difference between isograft+suture and ANA+PTB.

Figure 5. Nerve histomorphometry of ANA and isograft groups.

Figure 1. Methods of rodent sciatic nerve injury and repair. All left sciatic nerves had 15mm sections excised. These gaps were reconstructed with sections of processed rat ANA. In one group (n=10), ANAs were secured with conventional 10.0 ethilon suture. In the remaining group (n=10), ANAs were secured using photochemically sealed crosslinked amnion nerve wraps.

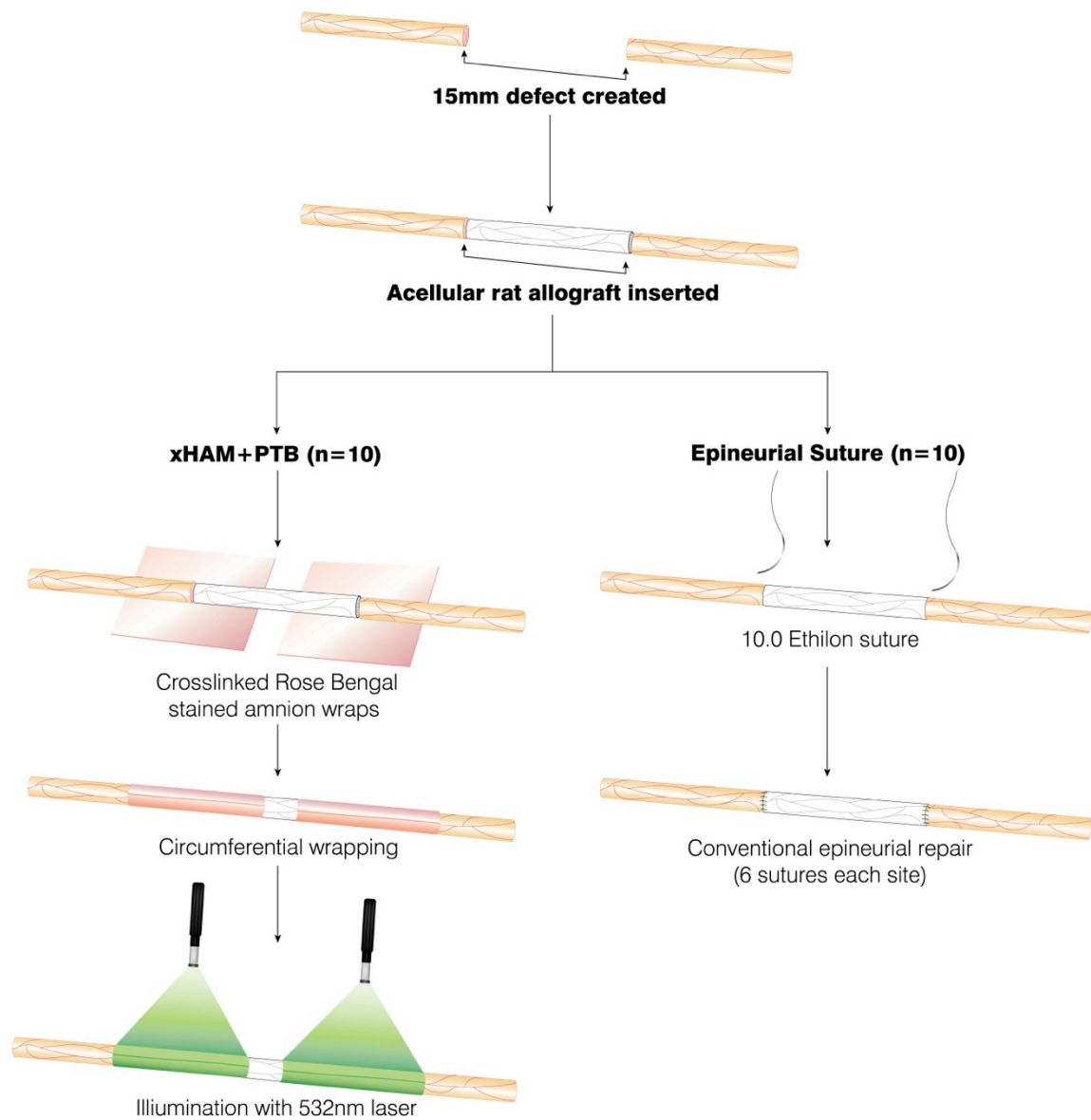


Figure 2. Gross observations following sacrifice. All nerves were found to be intact and showed evidence of regeneration across both isografts and ANA. Qualitatively, sutured isografts (A) and sutured ANA (D) had greater adhesion formation around coaptation sites. Photochemically sealed isografts (B) and photochemically sealed ANA (C) showed remnants of RB stained amnion wraps. Qualitatively, sutured ANA diameter was noticeably reduced in comparison to isograft, an observation consistent with their statistically inferior outcome.

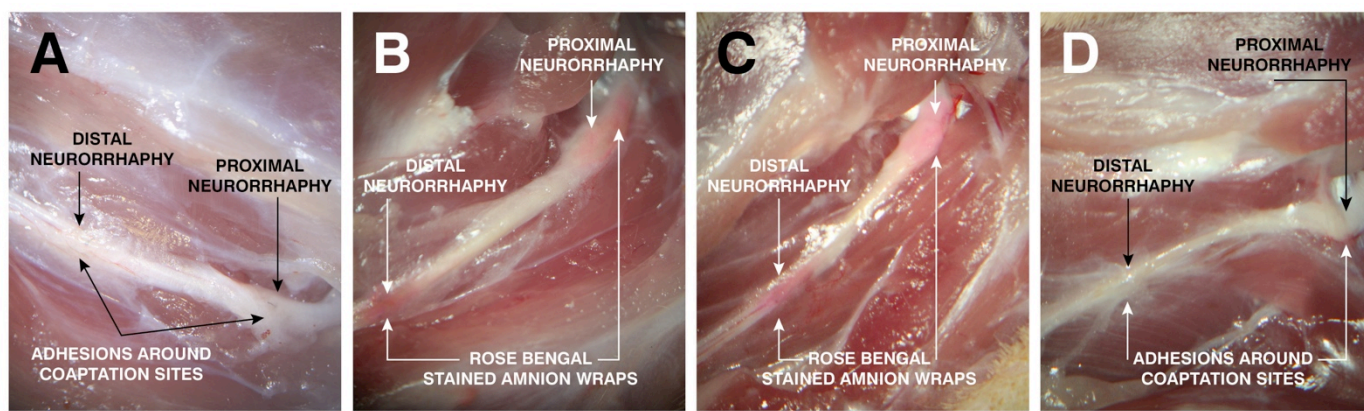


Figure 3. Sciatic Function Index of ANA and isograft treated groups. After 5-months follow-up, no significant difference existed within either isograft or ANA groups. ANA+suture and ANA+PTB groups recovered statistically less SFI in comparison to both isograft groups.

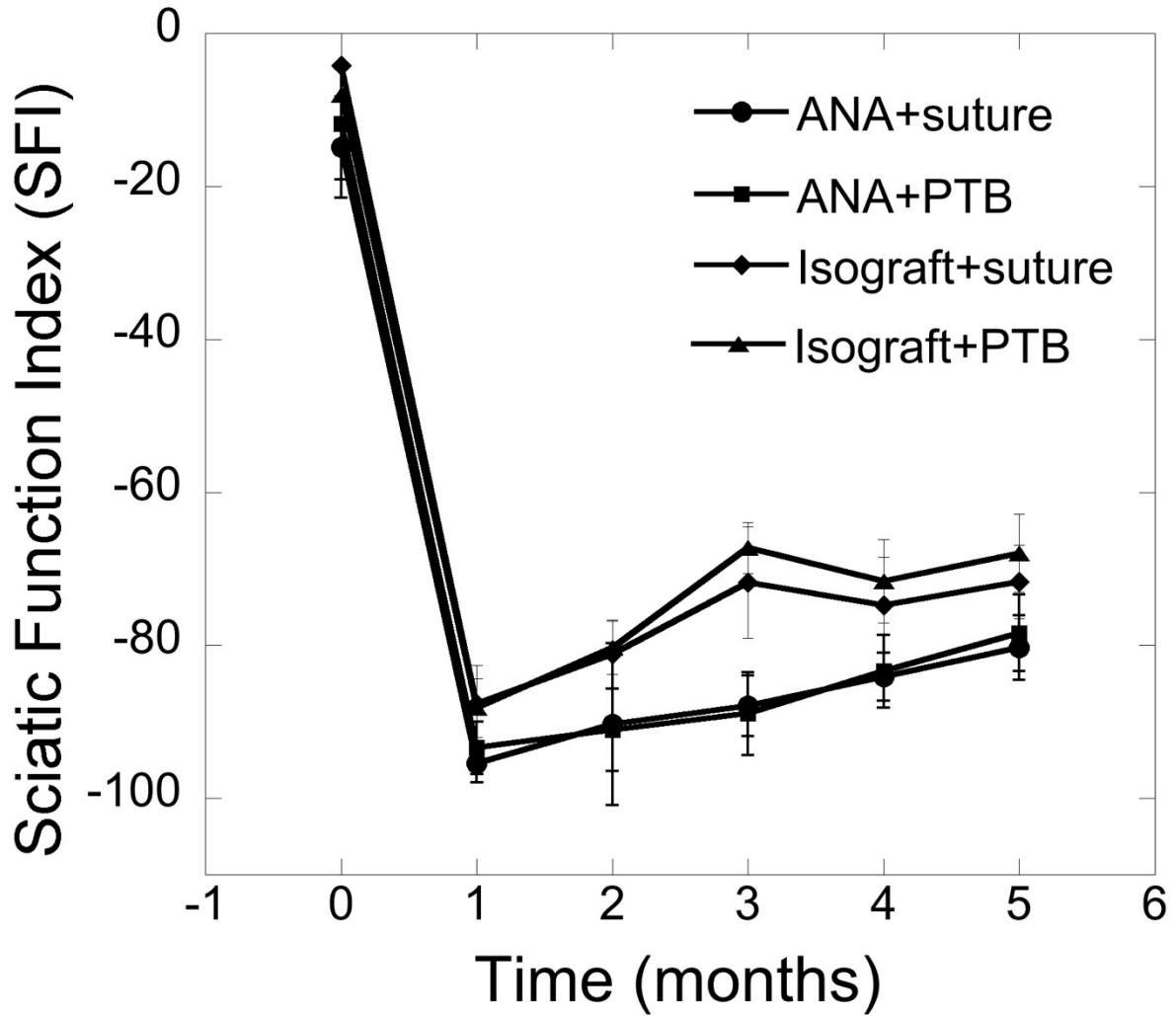


Figure 4. Gastrocnemius Muscle Mass Retention for ANA and isograft treated groups. Isografts sealed photochemically with crosslinked amnion were statistically superior to all other groups. No significant difference existed between sutured and photochemically sealed ANA. ANA+suture was recovered statistically less muscle mass than isograft+suture but there was no significant difference between isograft+suture and ANA+PTB.

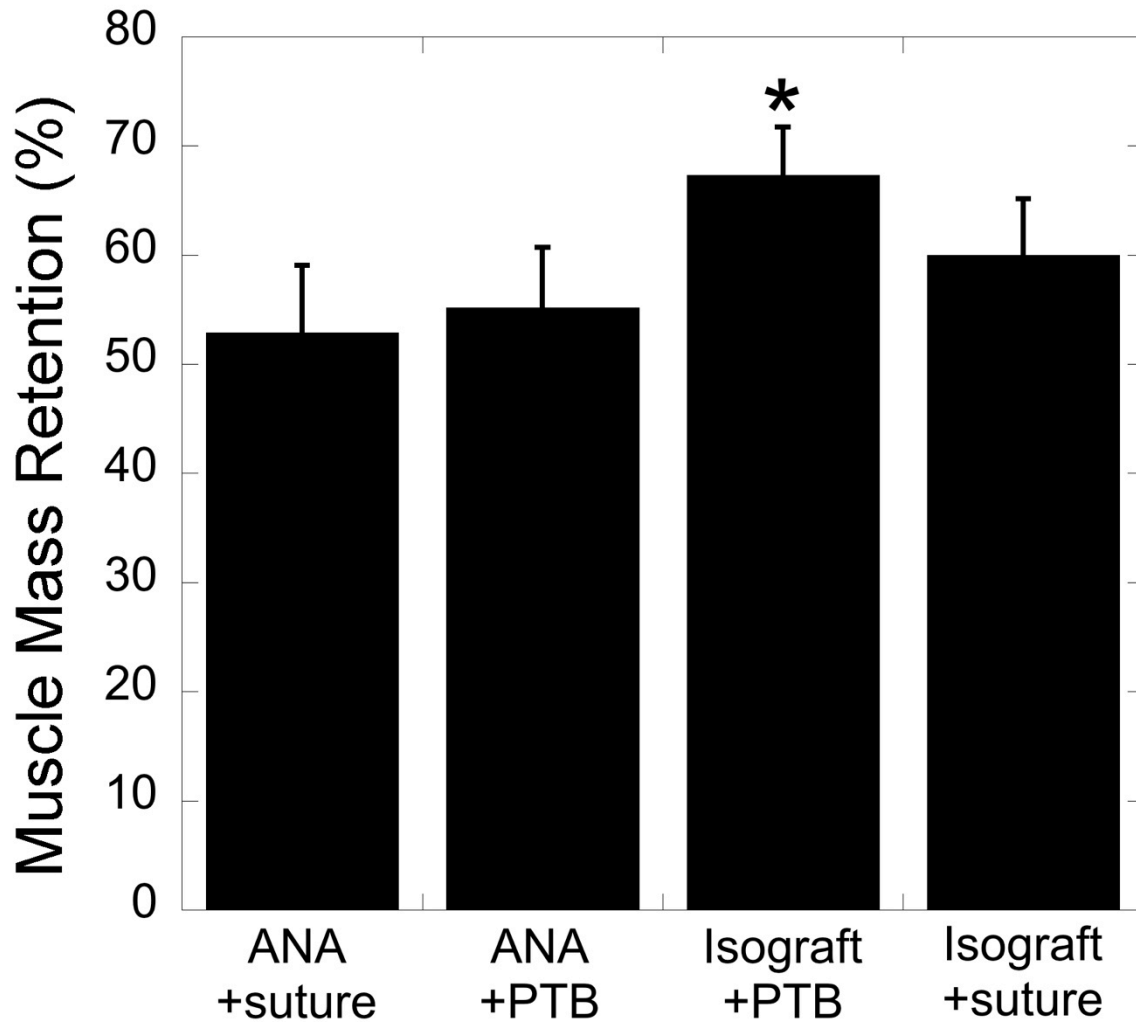


Figure 5. Nerve histomorphometry of ANA and isograft groups.

