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CONTRACTING ORGANIZATION: University of Colorado, Denver, Colorado 80045

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For the last 12 months we have been working on data analysis and manuscript preparation. The first 6 papers from this study have been published, and another 6 are in preparation. The overview physiology paper was accepted last week at PLOS ONE, and the others have been accepted in major physiology journals. The next batch of papers will be from the OMICS portion of the study. We hired a new bioinformatics postdoc in October, and he has hit the ground running with comprehensive analyses of the OMICS dataset. The gene expression paper will be complete in another 3-4 weeks, and shortly after that the metabolomics and epigenetics data will be ready for publication. By the end of 2014 all major papers form the study will have been published. So far the project is from our perspective a complete success with identification of new physiological aspects of acclimatization, and first-ever insights into the underlying OMICS mechanisms. We are now looking for one additional year of funding to explore new analyses and integration that is possible because of the high quality of this dataset, and that could lead to even more comprehensive, "big picture" views of the process of human acclimatization to hypoxia at high altitude.						
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## **INTRODUCTION:**

The goal of this project is to advance high-altitude medical research by discovering the basic molecular mechanisms of acclimatization and de-acclimatization that protect soldiers from high-altitude illness.

## **BODY:**

All major milestones have been accomplished. Now we are working on papers integrating the findings from the extensive physiological studies and the OMICS studies. Since no one has done that work before, we are inventing the methods and approaches as we go along. A major breakthrough ahs been the application of an advanced clustering algorithm called WGCNA to our datasets. This will allow us to condense the enormous datasets generated by the gene expression and epigenetics chip studies into a manageable system that can easily be tested for relationships to physiological tests.

Accomplishments to date:

- IRB compliance and continuing review have been completed
- Analyses are completed for all subjects at all time points for epigenetics, gene expression, microRNA and metabolomics.
- All cytokine arrays are done, with follow-up and validation ELISAs completed. Writing of those manuscripts is underway.
- Six papers have been accepted for publication, three in Journal of Applied Physiology, one in Experimental Physiology, one in Acta Scandinavica and the overview paper at PLOS ONE
- A seventh paper is under review at NeuroReports.
- Nitric oxide analyses are done, adenosine and hydrogen sulfide analyses are done. Work has begun on a paper on NO and H2S with Drs. Roach, Kevil and Gladwin.
- Analysis of ADP, ATP and purigenic receptors is complete, writing that manuscript is underway. Another paper is in the works as well with Drs. Eltzschig, Blackburn, Xia, and Davis on adenosine in AltitudeOmics.
- The Lovering laboratory, home of our collaborators on AltitudeOmics, have two papers in preparation on AMS and intrapulmonary shunts, and one on gas exchange during AltitudeOmics.

## **KEY RESEARCH ACCOMPLISHMENTS:**

- 1. Completed the first ever measurements of acute mountain sickness, cognitive function and exercise capacity after 7 and 21 days of de-acclimatization. The results suggest near complete retention of acclimatization after 7 days de-acclimatization, and about 70% retention after 21 days. This key finding will be used in the OMICS analyses to help identify factors that occur with acclimatization, and are still present after de-acclimatization.
- 2. Six research papers have been completed and published on the physiology of human acclimatization to high altitude, and another is under review. Seven additional primary papers will be completed this year. Please see Appendices section for a table showing the "Status of Research Papers" and for a PDF of the published papers.

## **REPORTABLE OUTCOMES:**

- 1. Completed all regulatory steps to gain approval for this multi-site, multi-nation study.
- 2. Safely completed data collection on 23 young healthy student volunteers, and safely transported and cared for them and 40 scientists to/from Bolivia.
- 3. We are 100% in analysis and manuscript writing mode regarding all aspects of the study.

## **CONCLUSION:**

Humans retain acclimatization after 7 and 21 days of de-acclimatization. This was a key hypothesis of the study. Yet to be determined is what are the OMICS responses that can be linked to the process of gaining acclimatization, and its retention on descent to low altitude?

Status of manuscripts from the AltitudeOmics study, 2/15/2014

	#	Title	First/Last Authors	Status	PMID	Link to PDF
	1	Amann M, Goodall S, Twomey R, Subudhi AW, Lovering AT, Roach	Amman/Roach	Published, JAPPL	23813531	
		AltitudeOmics: on the consequences of high-altitude acclimatizati				
		the development of fatigue during locomotor exercise in humans.				
		Journal of applied physiology. 2013;115:634-42.				
	2	Goodall S, Twomey R, Amann M, Ross EZ, Lovering AT, Romer	Goodall/Roach	Accepted, Acta	24450855	
		LM, Subudhi AW, Roach RC. AltitudeOmics: Exercise-induced		Scandinavica		
		supraspinal fatigue is attenuated in healthy humans after				
		acclimatisation to high altitude. Acta Physiologica. 2014.				
	3	Subudhi AW, Fan JL, Evero O, Bourdillon N, Kayser B, Julian CG,	Subudhi/Roach	Accepted, Exp	24243839	
		Lovering AT, Roach RC. AltitudeOmics: Effect of ascent and		Physiol		
		acclimatization to 5260 m on regional cerebral oxygen delivery.				
		Experimental Physiology. 2013.				
	4	Fan JL, Subudhi AW, Evero O, Bourdillon N, Kayser B, Lovering	Fan/Roach	Accepted, JAPPL	24356520	
		AT, Roach RC. AltitudeOmics: Enhanced cerebrovascular				
		reactivity and ventilatory response to CO2 with high altitude				
		acclimatisation and re-exposure. Journal of Applied Physiology.				
_	-				04054040	
	5	Subudhi AW, Fan JL, Evero O, Bourdillon N, Kayser B, Julian CG,	Subudni/Roach	Accepted, JAPPL	243/1013	
		Lovering AI, Paneral RB, Roach RC. AltitudeOmics: Cerebral				
		autoregulation during ascent, acclimatization, and re-exposure				
		to high antitude and its relation with acute mountain sickness.				
-	6	Subudhi AW, Busher I, Bourdillon N, Davis C, Elliott I	Subudhi /Doach	Accorted DLOSOne		
	0	Subuulli Aw, bucher J, bour alloir N, bavis C, Elliott J, Eutormostor M, Evere O, Ean II, Jameson Van Houton S, Julian	Subuuiii/ Koacii	Accepted, PL050lle		
		CC Kark I Kark S Kawar B Karn ID Kim SE Lathan C Lauria				
		SS Lovering AT Paterson R Polaner D Ryan RI Spiral Teac IW				
		Wachsmuth NR Roach RC AltitudeOmics: The Integrative				
		Physiology of the Onset and Retention of Acclimatization to				
		Hypoxia in Humans, PLOS One (In Press) 2014.				

#	Title	First/Last Authors	Status	PMID	Link to PDF
7	AltitudeOmics: hemoglobin mass increases within 7 days of acclimatization to 5260m and is lost within 7 days of descent to	Ryan/Roach	Under revision		
	1525m				
8	AltitudeOmics: Detecting high altitude cognitive impairment	Roach/Roach	Submitted,		
			Neuroreport		
9	acclimatization, and re-exposure to 5,260m	Julian/Roach	In prep		
10	AltitudeOmics: Effect of ascent, acclimatization, and re-	Elliot/Roach	In prep		
	exposure to 5,260m on pulmonary shunt		-		
11	AltitudeOmics: Effect of ascent, acclimatization, and re-	Chicco/Roach	In prep		
	exposure to 5,260m on muscle mitochondrial respiration				
12	AltitudeOmics: Metabolomics during ascent, acclimatization,	Monte/Roach	In prep		
	and re-exposure to 5,260m				
13	AltitudeOmics: Effect of ascent, acclimatization, and re-	Kern/Roach	In prep		
	exposure to 5,260m on AMS and pulmonary shunt	1	1 1		
14	AltitudeOmics: Gene Expression and Acute Mountain Sickness	?/Roach	In prep		
15	AltitudeOmics: MicroRNA expression during ascent, acclimatization, and re-exposure to 5,260m	Kern/Roach	In prep		

# AltitudeOmics: on the consequences of high-altitude acclimatization for the development of fatigue during locomotor exercise in humans

Markus Amann,<sup>1</sup> Stuart Goodall,<sup>2</sup> Rosie Twomey,<sup>3</sup> Andrew W. Subudhi,<sup>4,5</sup> Andrew T. Lovering,<sup>6</sup> and Robert C. Roach<sup>4</sup>

<sup>1</sup>Department of Medicine, University of Utah, Salt Lake City, Utah; <sup>2</sup>Faculty of Health and Life Sciences, Northumbria University, Newcastle, United Kingdom; <sup>3</sup>School of Sport and Service Management, University of Brighton, Eastbourne, United Kingdom; <sup>4</sup>Altitude Research Center, Department of Emergency Medicine, University of Colorado Anschutz Medical Campus, Aurora, Colorado; <sup>5</sup>Department of Biology, University of Colorado, Colorado Springs, Colorado; and <sup>6</sup>Department of Human Physiology, University of Oregon, Eugene, Oregon

Submitted 20 May 2013; accepted in final form 24 June 2013

Amann M, Goodall S, Twomey R, Subudhi AW, Lovering AT, Roach RC. AltitudeOmics: on the consequences of high-altitude acclimatization for the development of fatigue during locomotor exercise in humans. J Appl Physiol 115: 634-642, 2013. First published June 27, 2013; doi:10.1152/japplphysiol.00606.2013.-The development of muscle fatigue is oxygen (O2)-delivery sensitive [arterial O<sub>2</sub> content (C<sub>a</sub>O<sub>2</sub>) × limb blood flow ( $Q_L$ )]. Locomotor exercise in acute hypoxia (AH) is, compared with sea level (SL), associated with reduced CaO2 and exaggerated inspiratory muscle work (Winsp), which impairs  $Q_{\rm L}$ , both of which exacerbate fatigue individually by compromising O2 delivery. Since chronic hypoxia (CH) normalizes CaO2 but exacerbates Winsp, we investigated the consequences of a 14-day exposure to high altitude on exercise-induced locomotor muscle fatigue. Eight subjects performed the identical constant-load cycling exercise (138  $\pm$  14 W; 11  $\pm$  1 min) at SL (partial pressure of inspired O<sub>2</sub>, 147.1  $\pm$  0.5 Torr), in AH (73.8  $\pm$  0.2 Torr), and in CH (75.7  $\pm$  0.1 Torr). Peripheral fatigue was expressed as pre- to postexercise percent reduction in electrically evoked potentiated quadriceps twitch force ( $\Delta Q_{tw,pot}$ ). Central fatigue was expressed as the exercise-induced percent decrease in voluntary muscle activation ( $\Delta$ VA). Resting C<sub>a</sub>O<sub>2</sub> at SL and CH was similar, but C<sub>a</sub>O<sub>2</sub> in AH was lower compared with SL and CH (17.3  $\pm$  0.5, 19.3  $\pm$  0.7,  $20.3 \pm 1.3$  ml O<sub>2</sub>/dl, respectively). W<sub>insp</sub> during exercise increased with acclimatization (SL:  $387 \pm 36$ , AH:  $503 \pm 53$ , CH:  $608 \pm 67$ cmH<sub>2</sub>O·s<sup>-1</sup>·min<sup>-1</sup>; P < 0.01). Exercise at SL did not induce central or peripheral fatigue.  $\Delta Q_{tw,pot}$  was significant but similar in AH and CH (21  $\pm$  2% and 19  $\pm$  3%; P = 0.24).  $\Delta$ VA was significant in both hypoxic conditions but smaller in CH vs. AH  $(4 \pm 1\% \text{ vs. } 8 \pm 2\%; P < 0.05)$ . In conclusion, acclimatization to severe altitude does not attenuate the substantial impact of hypoxia on the development of peripheral fatigue. In contrast, acclimatization attenuates, but does not eliminate, the exacerbation of central fatigue associated with exercise in severe AH.

altitude; respiratory muscle work; arterial  $O_2$  content; cerebral blood flow

THE DEVELOPMENT OF LOCOMOTOR muscle fatigue during wholebody endurance exercise is highly sensitive to the delivery of oxygen [O<sub>2</sub>; arterial O<sub>2</sub> content (C<sub>a</sub>O<sub>2</sub>) × leg blood flow ( $Q_L$ )]. Specifically, blunted O<sub>2</sub> delivery exaggerates, and augmented O<sub>2</sub> delivery attenuates the rate of development of locomotor muscle fatigue during exercise (1). Acute exposure to hypoxia (AH) has a substantial impact on the two determinants of leg muscle  $O_2$  delivery during strenuous locomotor exercise. First, despite a marked hyperventilatory response, arterial partial pressure of  $O_2$  [PO<sub>2</sub> (P<sub>a</sub>O<sub>2</sub>)] and arterial hemoglobin saturation (S<sub>a</sub>O<sub>2</sub>) fall below sea level (SL) values and cause a significant reduction in C<sub>a</sub>O<sub>2</sub>. In addition, inspiratory muscle work (W<sub>insp</sub>) is increased substantially at any given workload in hypoxia (2, 58), and these high levels of W<sub>insp</sub> compromise, in a dose-dependent manner,  $Q_L$  during exercise (34). Each of these two determinants of leg muscle O<sub>2</sub> delivery, namely C<sub>a</sub>O<sub>2</sub> and  $Q_L$ , accounts for, substantially and independently, the accelerated development of locomotor muscle fatigue in hypoxia (2).

During prolonged exposure to altitude, a progressive, timedependent hyperventilation, which increases alveolar PO<sub>2</sub>, occurs over the initial hours and days and advances more gradually over the ensuing 1-2 wk of acclimatization (56). This ventilatory acclimatization adds to an accompanying reduction in the alveolar-arterial O<sub>2</sub> gradient, which combined, substantially improves arterial oxygenation during exercise by increasing  $P_aO_2$  and  $S_aO_2$  (9, 13). Furthermore, chronic exposure to hypoxia (CH) is accompanied by erythropoiesis, and the combination of an increased hemoglobin concentration ([Hb]) plus improved oxygenation may serve to restore resting SL  $C_aO_2$  (8, 13). In contrast to this beneficial effect on  $O_2$  delivery,  $Q_L$ , during intense leg exercise at a given submaximal absolute workload, has been suggested to decline from SL to CH (8, 49, 64). The net effect of these acclimatization-induced, opposing consequences on leg O<sub>2</sub> delivery depends on the degree to which the increase in C<sub>a</sub>O<sub>2</sub> can counterbalance potential reductions in  $Q_{\rm L}$ . It has been documented previously that at a given absolute workload, locomotor muscle O2 delivery is reduced from SL to AH with no further changes following acclimatization (Pikes Peak, 4,300 m) (8, 64). Therefore, given the critical role of muscle  $O_2$  delivery in the development of fatigue, it could be argued that peripheral fatigue during constant-load endurance exercise is exacerbated in AH (vs. SL) and does not improve further during prolonged acclimatization. On the other hand, studies conducted at the same location as the present experiments [Mt. Chacaltaya (Bolivia), 5,260 m] document a reduction in locomotor muscle O<sub>2</sub> delivery from SL to AH and a full recovery following prolonged exposure, with the net effect of similar values in SL and CH (13). Based on these findings, it could be argued that the development of peripheral fatigue during constant-load endurance exercise is fastened in AH but recovers to SL values in CH.

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In this study, we sought to quantify exercise-induced locomotor muscle fatigue induced by the identical constant-load cycling trial performed at SL, in AH, and in CH (following 14 days at 5,260 m) to clarify the effects of acclimatization. We hypothesized that fatigue is, compared with SL, exacerbated significantly in AH and that altitude acclimatization would alleviate this impact.

## METHODS

This study was conducted as part of the AltitudeOmics project, examining the integrative physiology of human responses to hypoxia. All procedures conformed to the Declaration of Helsinki and were approved by the Universities of Colorado, Oregon, and Utah Institutional Review Boards and the U.S. Department of Defense Human Research Protection Program Office. All subjects were born and raised below 1,500 m and had not traveled to elevations >1,000 m for 3 mo before the experiments. Eight subjects (age  $21 \pm 1$  y, body weight  $69 \pm 11$  kg, height  $176 \pm 10$  cm) were studied at SL and following 14 days of altitude acclimatization at 5,260 m on Mt. Chacaltaya. At high altitude, subjects did not follow a systematic exercise-training program but were given the opportunity to participate, on a voluntary basis, in light hikes around the campsite (no significant change in altitude).

## Experimental Protocol

All participants were familiarized thoroughly with various experimental procedures involved in this investigation. The SL experiments of the present study were conducted  $\sim 130$  m above SL [Eugene, OR; barometric pressure (BP) 750.0  $\pm$  2.2 Torr]. The experiments in AH were conducted at the same altitude, while breathing a gas mixture containing 10.5% O<sub>2</sub> balance nitrogen, and experiments in CH were conducted on the 14th day of acclimatization at 5,260 m (BP 408.9  $\pm$ 0.7 Torr). Two participants were tested every morning. To assure that all subjects were tested exactly on day 14 after arrival on the mountain, the groups' transport to the mountain was staged, i.e., two new participants arrived every day. SL peak power output (W<sub>peak</sub>) was obtained from a maximal incremental exercise test (70, 100, 130, and 160 W for 3 min, each followed by 15 W/min increases thereafter) on a computer-controlled bicycle ergometer (Velotron, Dynafit; RacerMate, Seattle, WA). The experimental trial consisted of the identical constant-load cycling exercise (same absolute workload and duration) in each condition. Preliminary experiments (using different subjects), conducted to identify a workload that causes voluntary exhaustion between 8 and 12 min when acutely exposed to 5,260 m, revealed that a constant workload equal to 50% of SL W<sub>peak</sub> was required to reach this goal. Based on this, the workload during the experimental trials was set to equal 50% (138  $\pm$  14 W) of the subjects' SL W<sub>peak</sub> (275  $\pm$  14 W). Since an individual's endurance/aerobic capacity is lowest in AH (vs. SL and CH) (13), the first trial was performed to voluntary exhaustion in AH, and the achieved time (10.6  $\pm$  0.7 min) was then used for all subsequent trials. A 5-min warm-up at 10%  $W_{peak}$  (27  $\pm$ 8 W) preceded each trial. Throughout exercise, subjects were instructed to maintain their preferred pedal frequency, as determined during the practice sessions (88  $\pm$  3 rpm). Neuromuscular function was assessed before and within 2.5 min after exercise. During these procedures, subjects breathed ambient air at SL and in CH and a gas mixture (10.5% O<sub>2</sub>) in AH.

### Exercise Responses

Pulmonary ventilation ( $V_E$ ) and gas exchange were measured at rest and throughout exercise using an open circuit system (Ultima PFX; Medical Graphics, St. Paul, MN, and O2cap; Oxigraf, Mountain View, CA). Arterial O<sub>2</sub> saturation (S<sub>p</sub>O<sub>2</sub>) was estimated continuously at rest and during exercise using a pulse oximeter (Nellcor N-200;

Pleasanton, CA) with adhesive forehead sensors. A correction factor based on arterial blood gases was used to adjust for the nonlinearity associated with the obtained pulse oximeter values (error between 60% and 80% saturation: 6%; error between >90% saturation: 3%). Heart rate was measured from the R-R interval of an ECG, using a three-lead arrangement. Ratings of perceived exertion were obtained using Borg's modified CR10 scale (10). [Hb] was measured (Radiometer OSM-3) in resting arterial blood samples collected at SL and on the 16th day at 5,260 m.  $C_aO_2$  was estimated as 1.39 [Hb]  $\times$  $(S_pO_2/100)$ . During all constant workload trials, esophageal pressure (Pes) was measured via a nasopharyngeal balloon (Cooper Surgical, Trumbull, CT), using standard procedures (7). To estimate W<sub>insp</sub>, P<sub>es</sub> was integrated over the period of inspiratory flow, and the results were multiplied by respiratory frequency  $(f_R)$  and labeled the inspiratory muscle pressure-time product. Vastus lateralis oxygenation was assessed using a multichannel near-infrared spectroscopy (NIRS) instrument (Oxymon Mk III; Artinis, Zetten, The Netherlands). As described previously (5), a NIR emitter and detector pair was affixed over the belly of the left vastus lateralis muscle (~15 cm proximal and 5 cm lateral to the midline of the superior border of the patella), using a spacer with an optode distance of 5.0 cm. Probes were secured to the skin using double-sided tape and shielded from light using elastic bandages. The Beer-Lambert Law was used to calculate micrometer changes in tissue oxygenation [oxyhemoglobin (O<sub>2</sub>Hb) and deoxyhemoglobin (HHb)] across time. using received optical densities from two continuous wavelengths of NIR light (780 and 850 nm) and a fixed differential path-length factor of 4.95 (26). Total hemoglobin (THb) was calculated as the sum of [O<sub>2</sub>Hb] and [HHb] changes to give an index of change in regional blood volume (59). Data were recorded continuously at 10 Hz and expressed relative to the resting baseline recorded in each experimental condition. Mean cerebral blood flow (CBF) was estimated from blood velocity (CBFv) in the left middle cerebral artery (MCA;  $50 \pm 4 \text{ mm deep}$ ), determined using a 2-MHz transcranial Doppler (Spencer Technologies, Seattle, WA). An index of cerebral O<sub>2</sub> delivery was calculated as the product of CBFv and C<sub>a</sub>O<sub>2</sub>. Changes in CBFv were assumed to reflect changes in CBF, based on evidence that the MCA changes minimally in response to hypoxia and hypocapnia (47, 54). The validity of this assumption at altitude has been challenged recently (62). Evidence of MCA dilation was demonstrated in subjects at altitudes above 6,400 m, but no changes in MCA diameter were observed at altitudes comparable with the present study (<5,300 m) (63). We acknowledge that these measurements must be interpreted with caution until definitive studies of MCA diameter at altitude are conducted.

## Expiratory Flow Limitations and Lung Volume Responses

*Expiratory flow limitations.* Subjects performed three maximal volitional flow-volume (FV) maneuvers before and after exercise (after assessment of neuromuscular function). Exercise tidal FV loops (FVLs) were plotted within the best of the six maximal loops (MFVLs), based on measured inspiratory capacity (IC) maneuvers (rest, 3 min of exercise, and immediately before the termination of exercise). Acceptable IC maneuvers during exercise required that peak inspiratory P<sub>es</sub> match that obtained at rest. The amount of expiratory flow limitation was defined as the percentage of the tidal volume (V<sub>T</sub>) that met the boundary of the expiratory portion of the MFVL (38).

*Lung volumes.* Functional residual capacity (FRC) was measured in a body plethysmograph (Platinum Elite Series; Medical Graphics), and total lung capacity (TLC) was calculated as the sum of FRC and IC. End-expiratory lung volume (EELV) was determined by subtracting the maximal IC, as measured during exercise from TLC, as measured at rest. End-inspiratory lung volume (EILV) was calculated as the sum of EELV and  $V_T$ . Inspiratory reserve volume, during exercise, was calculated by subtracting EILV from TLC, and expiratory reserve volume, during exercise, was determined by subtracting the residual volume from EELV.

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## Force and Compound Muscle Action Potentials

Knee-extensor force during voluntary and evoked contractions was measured using a calibrated load cell (Tedea, Basingstoke, UK). The load cell was fixed to a custom-built chair and connected to a noncompliant cuff, attached around the participant's right leg, just superior to the ankle malleoli. Participants sat upright in the chair with the hips and knees at 90° of flexion. Compound muscle action potentials (M-waves) were recorded from surface electrodes placed 2 cm apart over the vastus lateralis muscle belly. A reference electrode was placed over the patella. Evoked signals were amplified [gain: 1,000; force: custom-built bridge amplifier; electromyographic (EMG): PowerLab 26T; ADInstruments (Oxfordshire, UK)], bandpass filtered (EMG only: 20–2,000 Hz), digitized (4 kHz; PowerLab 26T, ADInstruments), acquired, and later analyzed (LabChart v7.0; ADInstruments) for peak-to-peak amplitude.

#### Neuromuscular Function

Force and EMG variables were assessed before and immediately (<2.5 min) after each trial. Before each trial, maximum voluntary contraction (MVC) force was determined from three control contractions. Femoral nerve stimulation was delivered during each 5-s MVC, and an additional stimulus was delivered after the MVC to determine the potentiated quadriceps twitch force ( $Q_{tw,pot}$ ) and voluntary muscle activation (VA) (42). Briefly, the force produced during the superimposed twitch (SIT), delivered within 0.5 s of attaining peak force during the MVC, was to be compared with the force produced by the single twitch, delivered during relaxation, ~2 s after the MVC: VA (%) = [1 - (SIT/Q\_{tw,pot})] × 100. The contraction sets were repeated three times, with 30 s between each set. Visual feedback of the target force was provided via a computer monitor.

Femoral nerve stimulation. Single electrical stimuli (200 µs pulse width) were delivered to the right femoral nerve via surface electrodes (32 mm diameter; CF3200; Nidd Valley Medical, North Yorkshire, UK) and a constant-current stimulator (DS7AH; Digitimer, Welwyn Garden City, Hertfordshire, UK). The cathode was positioned over the nerve, high in the femoral triangle; the anode was placed midway between the greater trochanter and the iliac crest (32). The site of stimulation that produced the largest resting twitch amplitude and M-wave was located. Single stimuli were delivered, beginning at 100 mA and increasing by 20 mA, until plateaus occurred in twitch amplitude and M-wave. Supramaximal stimulation was ensured by increasing the final intensity by 30% (mean current,  $250 \pm 55$  mA). Muscle contractility was assessed for each potentiated twitch as twitch amplitude (Qtw.pot: peak force - onset force), maximum rate of force development (MRFD), contraction time, maximum relaxation rate (MRR), and one-half relaxation time (RT<sub>0.5</sub>). Sarcolemmal membrane excitability was inferred from the peak-to-peak amplitude of the electrically evoked M-wave (27).

#### Reliability Measures

On a separate day, measures of neuromuscular function were repeated twice in all subjects at SL. The two assessment procedures were separated by a 2-min walk around the laboratory, followed by a 5-min rest period. Coefficient of variation (CV) and Pearson product-moment correlation coefficients (r) were calculated to evaluate test-retest error (precision) and test-retest reliability of the neuromuscular function-assessment procedure. All correlations were significant and indicated; in combination with the CVs, acceptable degrees of reproducibility include: MVC, CV = 3.1%, r = 0.97;  $Q_{tw,pot}$ , CV = 4.1%, r = 0.98; M-wave peak, CV = 4.8%, r = 0.98; VA, CV = 3.3%, r = 0.77.

#### Statistical Analysis

A one-way repeated-measures ANOVA was performed to evaluate differences among trials. A least-significance difference test identified the means that were significantly different with P < 0.05. Results are expressed as mean  $\pm$  SE.

#### RESULTS

## $C_aO_2$ and Cerebral $O_2$ Delivery

 $C_aO_2$  at rest was significantly lower in AH compared with SL and CH (17.3 ± 0.5, 19.3 ± 0.7, 20.3 ± 1.3 ml O<sub>2</sub>/dl, respectively). Acclimatization to altitude significantly increased [Hb] and S<sub>p</sub>O<sub>2</sub>, resulting in similar C<sub>a</sub>O<sub>2</sub> at SL and in CH (P = 0.16). Resting CBFv was similar among SL, AH, and CH (50.5 ± 3.7, 52.7 ± 2.3, and 55.7 ± 3.0 cm/s, respectively; P = 0.45). In all three conditions, CBFv increased significantly from rest to the final minute of exercise (22 ± 3%, 39 ± 6%, and 28 ± 5% for SL, AH, and CH, respectively; Table 1). The percent increase was significantly greater in AH compared with that observed at SL and in CH. The cerebral O<sub>2</sub> delivery index during the last minute of exercise was 18 ± 5% lower in AH vs. SL (Table 1) and 17 ± 8% greater in CH vs. SL (Table 1).

#### Ventilatory Effects

*Ventilatory response.* AH increased  $W_{insp}$  work by  $34 \pm 8\%$  above that at SL (P < 0.01) and dropped  $S_pO_2$  by  $36 \pm 3\%$  during the final minute of exercise. Following 14 days of acclimatization,  $W_{insp}$  was increased further by  $23 \pm 8\%$  from AH, and  $S_pO_2$ , during the final minute of exercise, was  $36 \pm 5\%$  higher in CH vs. AH. Breathing frequency and  $V_E$  rose

Table 1. Mean responses to the final minute of exercise (138  $\pm$  14 W, 10.6  $\pm$  0.7 min)

	Sea Level	Acute Hypoxia	Chronic Hypoxia
HR, beats/min	152 ± 5	$174 \pm 4*$	$166 \pm 4^{*}$
$V_{E}$ , 1 min <sup>-1</sup>	$64 \pm 4$	$113 \pm 8*$	$133 \pm 10*^{++}$
$f_{\rm R}$ , breaths min <sup>-1</sup>	$32 \pm 2$	$50 \pm 3*$	$54 \pm 3*$
V <sub>T</sub> , liter	$2.0 \pm 0.1$	$2.2 \pm 0.2$	$2.6 \pm 0.2*$ †
$\dot{V}O_2$ , 1 min <sup>-1</sup>	$2.58\pm0.19$	$2.44 \pm 0.19^{*}$	$2.39 \pm 0.16*$ †
$\dot{V}_{CO_2}$ , 1 min <sup>-1</sup>	$2.51\pm0.22$	$2.81 \pm 0.21*$	$2.40 \pm 0.15*$ †
$V_E / \dot{V}O_2$	$25 \pm 1$	$50 \pm 4*$	56 ± 3*†
$V_E/\dot{V}_{CO_2}$	$26 \pm 1$	$41 \pm 2^{*}$	58 ± 3*†
S <sub>p</sub> O <sub>2</sub> , %	$94.1 \pm 1.0$	$62.2 \pm 1.8^{*}$	$75.6 \pm 1.2*$ †
CBFv, cm/s	$59.1 \pm 4.8$	$74.2 \pm 3.8*$	$73.2 \pm 3.4*$
Cerebral O <sub>2</sub> delivery, a.u.	$1,105 \pm 62$	$895 \pm 40*$	1,289 ± 42*†
T <sub>i</sub> /T <sub>tot</sub>	$0.35 \pm 0.01$	$0.39 \pm 0.01*$	$0.39 \pm 0.01*$
T <sub>e</sub> , s	$1.30\pm0.08$	$0.74 \pm 0.05*$	$0.70 \pm 0.04*$
$W_{insp}$ , cmH <sub>2</sub> O · s <sup>-1</sup> · min <sup>-1</sup>	$387 \pm 36$	$503 \pm 53*$	$608 \pm 67^{*}$ †
IC, liter	$3.29\pm0.22$	$3.13 \pm 0.23$	$3.60 \pm 0.23^{*}$
V <sub>T</sub> /IC	$0.60\pm0.03$	$0.68 \pm 0.02*$	$0.72 \pm 0.02*$ †
IRV, liter	$1.30 \pm 0.14$	$0.99 \pm 0.13^{*}$	$0.96 \pm 0.05*$
ERV, liter	$1.98\pm0.25$	$2.14 \pm 0.29$	$1.67 \pm 0.25*$ †
EILV, %TLC	$80.5 \pm 1.6$	$85.4 \pm 1.7*$	$85.2 \pm 0.9*$
EELV, %TLC	$51.5 \pm 1.8$	$53.8 \pm 2.5$	$46.9 \pm 2.1*$ †
Expiratory flow limitation, <i>n</i>			
out of 8 subjects	0/8	2/8	4/8
RPE	$12.3 \pm 1.0$	$19.8 \pm 0.1*$	$17.9 \pm 0.6*$ †
Dyspnea	$11.5\pm0.7$	$19.5\pm0.2^*$	19.3 ± 0.2*

HR, heart rate; V<sub>E</sub>, minute ventilation;  $f_{\rm R}$ , breathing frequency; V<sub>T</sub>, tidal volume;  $\dot{\rm Vo}_2$ , maximum oxygen (O<sub>2</sub>) uptake;  $\dot{\rm VcO}_2$ , carbon dioxide production; S<sub>p</sub>O<sub>2</sub>, arterial O<sub>2</sub> saturation; CBFv, cerebral blood flow velocity; T<sub>i</sub>, duration of inspiration; T<sub>tot</sub>, duration of entire breath; T<sub>e</sub>, duration of expiration; W<sub>insp</sub>, inspiratory muscle work; IC, inspiratory capacity; IRV, inspiratory reserve volume; ERV, expiratory reserve volume; ELV, end-inspiratory lung volume; TLC, total lung capacity; EELV, end-expiratory lung volume; RPE, rating of perceived exertion. \*P < 0.05 vs. sea level;  $\dagger P < 0.05$  vs. acute hypoxia, n = 8.

substantially over the time of exercise in AH and CH, and V<sub>E</sub> was, during the final minute,  $79 \pm 13\%$  and  $110 \pm 12\%$ , respectively, higher compared with SL (P < 0.01). Pulmonary V<sub>E</sub> during the final minute of exercise was  $19 \pm 4\%$  higher in CH vs. AH (P < 0.01). Compared with SL, O<sub>2</sub> uptake, during the final minute of exercise, was  $5 \pm 2\%$  and  $7 \pm 2\%$  lower in AH and CH, respectively (both P < 0.05; Fig. 1).

*Expiratory flow limitation.* At SL, exercise flow rates during tidal breathing were well within the MFVL in all eight subjects. At end-exercise in AH, 6–51% of the  $V_T$  in two of the eight subjects reached flow limitation, as lung volume approached end-expiration. As  $V_E$  increased further in CH, expiratory flow rate became more limited, and 10–64% of the  $V_T$  in four of the eight subjects met the limit imposed by the MFVL.

## Membrane Excitability and Contractile Function

*M*-waves. As a measure of membrane excitability we examined pre- vs. postexercise vastus lateralis M-wave amplitudes in conjunction with the quadriceps muscle mechanical properties. Pre-exercise M-wave amplitudes were similar in all three conditions ( $10.2 \pm 1.0 \text{ mV}$ ,  $9.4 \pm 0.7 \text{ mV}$ , and  $12.9 \pm 1.8 \text{ mV}$  for SL, AH, and CH, respectively; P = 0.15). Postexercise M-wave amplitudes were unchanged from pre-exercise baseline values at SL and in AH ( $10.2 \pm 1.0 \text{ mV}$  and  $9.6 \pm 0.9 \text{ mV}$ , respectively; P > 0.3). However, following exercise in CH, M-wave amplitudes ( $7.8 \pm 2.1 \text{ mV}$ ) were reduced significantly from pre-exercise baseline levels (range: 1-18%; P < 0.01).

Quadriceps twitch force. Pre-exercise  $Q_{tw,pot}$  was similar in all three conditions (106 ± 4 N, 109 ± 4 N, and 110 ± 5 N for SL, AH, and CH, respectively; P = 0.18). Exercise in both hypoxic conditions caused a substantial (P < 0.01) but similar (P = 0.14) reduction in  $Q_{tw,pot}$  in all eight subjects. In contrast, exercise at SL did not induce measurable locomotor muscle fatigue; the postexercise  $Q_{tw,pot}$  was similar to pre-exercise baseline.

*MVC force.* Pre-exercise MVC was similar in all three conditions (391 ± 30 N, 394 ± 25 N, and 372 ± 30 N for SL, AH, and CH, respectively; P = 0.21). At SL, postexercise MVC was similar to pre-exercise baseline (P = 0.42). In



Fig. 1. Inspiratory muscle pressure-time product [esophageal pressure ( $P_{es}$ ) × respiratory frequency ( $f_R$ )] during the identical constant-load cycling exercise performed in all 3 conditions. \*P < 0.05 vs. acute hypoxia (AH), n = 8.

contrast, exercise in AH and CH caused a substantial reduction in MVC in all eight subjects. However, the exercise-induced reduction in MVC was  $30 \pm 9\%$  less in CH vs. AH (P < 0.05).

*Muscle activation.* Pre-exercise baseline values were similar in all three conditions (94  $\pm$  1%, 94  $\pm$  1%, and 93  $\pm$  1% for SL, AH, and CH, respectively; P = 0.19). Following the exercise at SL, muscle activation was unchanged from preexercise baseline (P = 0.88). In both AH and CH, postexercise muscle activation was significantly lower compared with preexercise baseline values. However, the pre- to postexercise decrease in muscle activation was 52  $\pm$  12% less in CH vs. AH (P < 0.01).

*Within-twitch measurements.* MRFD, MRR, and  $RT_{0.5}$  complement the findings reported for  $Q_{tw,pot}$ . The pre- to postexercise changes in within-twitch measurements of MRFD, MRR, and  $RT_{0.5}$  were similar in CH vs. AH.

#### Vastus Lateralis Tissue Oxygenation

O<sub>2</sub>Hb was unchanged from baseline to warm-up at SL (P = 0.40) but decreased in AH (P < 0.05) and CH (P = 0.05). Compared with baseline, O<sub>2</sub>Hb was unchanged during the final minute of exercise at SL (P = 0.73) but was significantly lower in AH and CH (both P < 0.01). This decrease was significantly greater in AH vs. CH. HHb was unchanged from baseline to warm-up at SL (P = 0.80) but decreased significantly in AH and CH. Compared with baseline, HHb was unchanged during the final minute of exercise at SL (P = 0.24) but similarly increased in AH and CH (both P < 0.01). This was unchanged from baseline to warm-up at SL (P = 0.37) during the final minute of exercise in all three conditions.

## DISCUSSION

The purpose of this investigation was to evaluate the effect of altitude acclimatization on the development of fatigue during whole-body endurance exercise. Subjects repeated the identical constant-load cycling exercise at SL, in AH, and in CH. No measurable degree of fatigue was found following the exercise at SL. However, the identical exercise in AH, characterized by a reduced CaO2 and increased Winsp, resulted in a substantial degree of both peripheral and central fatigue. Two weeks of exposure to 5,260 m restored C<sub>a</sub>O<sub>2</sub> to SL values but increased W<sub>insp</sub> further over that observed in AH. The critical finding was that the rate of development of peripheral locomotor muscle fatigue failed to recover from AH to CH and was similar in both conditions. In contrast, the development of central fatigue was attenuated significantly in CH (vs. AH) but still greater compared with SL. Taken together, our findings suggest that acclimatization to high altitude attenuates the impact of AH on the development of central fatigue but fails to improve the exacerbated development of peripheral fatigue present during exercise in AH.

## Peripheral Fatigue

*Acute hypoxia.* The cycling bout in AH was, compared with SL, characterized by a substantially exaggerated rate of peripheral fatigue (Table 2 and Fig. 2). These observations confirm numerous earlier findings using whole-body (4, 31, 57) and single-muscle exercise (28, 39).

Table 2.	Effects of	constant-load	cycling	exercise	on
quadrice	ps muscle	function			

Percent Change from Pre- to Immediately Postexercise					
	Sea Level	Acute Hypoxia	Chronic Hypoxia		
Q <sub>tw.pot</sub> MRFD MRR RT <sub>0.5</sub> MVC Voluntary muscle activation M-wave amplitude	$\begin{array}{c} -3.1 \pm 1.8^{*} \\ -4.1 \pm 2.5^{*} \\ 2.7 \pm 2.8^{*} \\ 1.0 \pm 2.2^{*} \\ -1.3 \pm 1.2^{*} \\ -0.1 \pm 1.0^{*} \\ 0.7 \pm 2.7^{*} \end{array}$	$\begin{array}{r} -20.9 \pm 2.4 \\ -21.2 \pm 4.2 \\ -13.2 \pm 3.1 \\ 9.2 \pm 1.3 \\ -12.3 \pm 1.2 \\ -6.9 \pm 1.1 \\ 2.5 \pm 2.0 * \end{array}$	$\begin{array}{c} -18.8 \pm 3.4 \\ -17.9 \pm 3.5 \\ -9.0 \pm 2.2 \\ 8.2 \pm 1.4 \\ -8.9 \pm 1.3 \dagger \\ -3.7 \pm 1.2 \dagger \\ -7.8 \pm 2.1 \end{array}$		

Changes in muscle function are expressed as a percent change from pre-exercise baseline. All exercise trials were performed for the same duration  $(10.6 \pm 0.7 \text{ min})$  and at the same absolute workload  $(138 \pm 14 \text{ W})$ . Values are expressed as means  $\pm$  SE.  $Q_{tw,pot}$ , potentiated single twitch; MRFD, maximal rate of force development; MRR, maximal rate of relaxation; RT<sub>0.5</sub>, 1/2 relaxation time; MVC, maximal voluntary contraction force; M-wave, compound muscle action potential. Percent muscle activation is based on super-imposed twitch technique. Various variables in acute and chronic hypoxia were, compared with baseline, altered significantly, 2.5 min after exercise (P < 0.01). \*Not significantly different from pre-exercise baseline.  $\dagger P < 0.05$  vs. acute hypoxia, n = 8.

Compared with SL,  $C_aO_2$  was approximately one-third lower and  $W_{insp}$ , ~34% higher during exercise in AH. These substantial alterations are known to contribute about equally to the exacerbated development of peripheral fatigue in AH (2). The impact of an acutely lowered  $C_aO_2$  on muscle fatigability is mediated via the facilitating effects of the associated reduction in muscle  $O_2$  delivery on the intramuscular accumulation of metabolites known to cause peripheral fatigue, i.e., hydrogen ion and inorganic phosphate (37, 61). The W<sub>insp</sub>-induced exacerbation of peripheral fatigue results from the same intramuscular metabolic consequences associated with reductions in locomotor muscle  $O_2$  delivery. However, in the case of the W<sub>insp</sub>-related impairment in peripheral fatigue, the compromised  $O_2$  delivery is the consequence of a sympathetically mediated impact on  $Q_L$ , secondary to the activation of the respiratory muscle metaboreflex (34). Taken together, the combined effects of a significantly reduced  $C_aO_2$  and a higher W<sub>insp</sub> has a profound impact on leg  $O_2$  delivery and thus peripheral locomotor muscle fatigue (1).

*Chronic hypoxia.* Despite 2 wk of acclimatization to altitude, the rate of development of peripheral locomotor muscle fatigue was similar in AH and CH (Table 2 and Fig. 2). Somewhat conflicting data from earlier investigations suggest different mechanisms as a potential explanation of this finding. On the one hand, studies conducted by Reeves and colleagues (8, 64), following 2–3 wk at 4,300 m, report similar locomotor muscle  $O_2$  delivery during submaximal endurance exercise in AH and CH. Given the critical dependency of the development of peripheral fatigue on muscle  $O_2$  delivery, this similarity might explain the nearly identical levels of end-exercise locomotor muscle fatigue in AH and CH. On the other hand, experiments conducted at the same location as the present study (Mt. Chacaltaya, 5,260 m) have documented a significant improvement in leg muscle  $O_2$  delivery from AH to CH, with the net



Fig. 2. Individual data illustrating the effects of constant-load bike exercise ( $138 \pm 14$  W; 10.6  $\pm$  0.7 min) on potentiated quadriceps twitch force (Q<sub>tw,pot</sub>; *top*) and voluntary muscle activation (VA; *bottom*) at sea level [SL; resting arterial oxygen (O<sub>2</sub>) content: 19.3  $\pm$  0.7 ml O<sub>2</sub>/dl] and in AH (17.3  $\pm$  0.5 ml O<sub>2</sub>/dl) and chronic hypoxia (CH; 20.3  $\pm$  1.3 ml O<sub>2</sub>/dl).

effect of similar values during submaximal bike exercise at SL and in CH (13). It might be important to emphasize that these latter experiments involved a greater altitude (5,260 m vs. 4,300 m) and a 9–10 wk acclimatization period vs. only a 2–3 wk period, as in the experiments by Reeves and colleagues (8, 64), as well as the present study. Regardless, based on the findings from the earlier Chacaltaya experiments, it appears that the similar degrees of end-exercise fatigue in AH and CH in the present study (Fig. 2) might have occurred in the face of a significant difference in bulk muscle O<sub>2</sub> delivery, i.e., higher in CH vs. AH.

 $Q_{\rm L}$  was not measured directly in the present study. However, changes in THb, a NIRS-derived variable, are thought to reflect changes in regional blood volume and potentially  $Q_{\rm L}$  (24, 59). The previously documented similarity in resting  $Q_{\rm L}$  at SL, in AH, and in CH (11, 12, 36, 49, 50) is a critical prerequisite when using THb as an estimate of potential differences in  $Q_{\rm L}$ and O<sub>2</sub> delivery during exercise. Since C<sub>a</sub>O<sub>2</sub> was comparable at SL and CH (see RESULTS), the same exercise-induced increase in THb (Fig. 3) suggests a similar degree of  $O_2$  delivery in these conditions. Furthermore, the combination of a lower C<sub>a</sub>O<sub>2</sub> in AH vs. CH (and SL; see RESULTS) plus the similar increase in THb during exercise (Fig. 3) insinuates a lower locomotor muscle O<sub>2</sub> delivery in AH vs. CH (and by extension, SL). Both of these observations might support earlier blood flow studies conducted at the same location as the present experiments (13) but might contradict others performed at a lower altitude (8, 64). However, NIRS findings obtained from skeletal muscle need to be interpreted with caution. A significant limitation associated with NIRS is that this measurement is confined to a finite location, and changes in THb might not be representative of the whole muscle. Indeed, significant blood flow heterogeneity has been documented previously in skeletal muscle (35). Whereas heterogeneity diminishes with higher exercise intensities and is not affected by hypoxia (36), the exact location of NIRS probe placement from day to day is a potential source of error. To minimize this risk, we had strict criteria regarding probe placement (see METHODS), and at least two investigators independently assured correct probe positioning before each experiment.

Assuming that the similar degrees of peripheral fatigue in AH vs. CH occurred in the face of a greater O<sub>2</sub> delivery in CH, other, rather disadvantageous adaptations associated with acclimatization must have outweighed this benefit. A potential candidate is the documented impairment in the capacity of skeletal muscle to extract O<sub>2</sub> in CH, i.e., a decreased capillary muscle  $O_2$  conductance (41). This impact might, despite a similar O<sub>2</sub> delivery at SL and in CH, potentially lower extracellular PO<sub>2</sub> to or beyond a previously suggested critical value  $(\sim 30 \text{ Torr})$  associated with exacerbated development of peripheral fatigue (55). Alternatively, the higher  $O_2$  delivery in CH vs. AH (13), combined with the same degree of peripheral fatigue, might suggest that CaO2 and bulk O2 delivery, per se, might not depict key determinants of the exaggerated fatigability in hypoxia. Important here is the fact that despite the normalized C<sub>a</sub>O<sub>2</sub> and bulk O<sub>2</sub> delivery in CH, P<sub>a</sub>O<sub>2</sub> only partially recovers with acclimatization and remains fairly low in CH. This could hint toward a key role of PaO2 in exacerbating the development of peripheral fatigue at altitude.

In CH,  $V_E$  was ~20% higher compared with AH. Given the substantially lower air density at 5,260 m (0.64 kg/m<sup>3</sup> vs. 1.18



Fig. 3. Vastus lateralis oxygenation at resting baseline, during the final 30 s of a 3-min warm-up (28 W), and during the final 30 s of constant-load exercise (131 W) at SL (A), in AH (B), and in CH (C).  $\dagger P < 0.05$  vs. respective baseline; \*P < 0.05 vs. AH, n = 8. O<sub>2</sub>Hb, oxyhemoglobin; HHb, deoxyhemoglobin; THb, total hemoglobin.

kg/m<sup>3</sup> at 130 m, where AH experiments occurred), it could be argued that in terms of respiratory muscle work, the reduced density might balance the acclimatization-induced increase in  $V_E$ , with the net effect of a similar  $W_{insp}$  in CH and AH. However,  $W_{insp}$  was, similar to  $V_E$ , ~20% higher in CH vs. AH. This observation, per se, might suggest that the lower air density at altitude had no effect on the relationship between minute  $V_E$  and respiratory muscle work. However, it has been shown that bronchoconstriction, associated with severe hypoxia, increases the resistive component of respiratory work and offsets the theoretical benefit of a reduced air density (22). This results in a similar respiratory muscle work for a given  $V_E$  at altitude and at SL (18). Therefore, any increase in  $W_{insp}$  observed in hypobaric CH is attributable to the exaggerated ventilatory response associated with altitude acclimatization.

The increase in minute  $V_E$  in the present study was mainly due to the increase in  $V_T$ ;  $f_R$  was similar in both conditions. The higher  $V_T$  was achieved via reductions in EELV (Table 1), which is compared with increasing EILV to raise  $V_T$ , more economical, since higher lung volumes are associated with a reduced compliance (38). We therefore conclude that the 23% higher  $W_{insp}$  at the same workload in CH vs. AH resulted from the substantially higher  $V_E$  following acclimatization. Finally, this exaggerated  $W_{insp}$  likely aggravated the respiratory muscle metaboreflex and associated impact on leg vascular conductance (25) and presumably blunted exercise  $Q_L$  more in CH compared with AH.

In contrast to our findings, it was suggested previously that acclimatization to high altitude might eliminate the impact of AH on the rate of development of fatigue during single muscle exercise (adductor pollicis) and restore it to that observed at SL (28). However, submaximal, intermittent exercise, including a small muscle mass, does not maximally challenge  $O_2$  delivery and use. Therefore, the observed positive effect could, at least in part, be explained by the use of the available reserve capacity. Specifically, various compensatory mechanisms, including increases in cardiac output and muscle  $O_2$  delivery and extraction, could have reduced the hypoxia-induced impact on the development of fatigue. Such an effective compensation might not—or only to a much smaller degree—be possible during intense, whole-body exercise, performed close to a human's maximal circulatory and ventilatory capacity (14, 15).

CH had a significant impact on the effect of exercise on M-wave amplitude. Reductions in M-wave amplitude have been associated with decreases in sarcolemma excitability (19). The attenuated excitability results from reduced sarcolemma sodium (Na<sup>+</sup>)-potassium (K<sup>+</sup>)-ATPase activity (46) and can contribute to compromised muscle force output (21). Preexercise M-wave amplitudes (and Q<sub>tw,pot</sub>) in our experiments were similar in all three conditions. This suggests that neither severe AH nor CH impairs sarcolemma Na<sup>+</sup>-K<sup>+</sup>-ATPase activity and membrane excitability of resting locomotor muscle. This confirms earlier findings (40); however, it contrasts with others (16) who report decreased resting M-wave amplitudes following 10 days of exposure to severe hypoxia (>4,300 m). Regardless, although M-wave amplitudes did not change from pre- to postexercise at SL and in AH, we observed, in contrast to Garner et al. (30), a significant exercise-induced decrease in CH (Table 2). AH has recently been shown to have no effect on exercise-induced changes in Na<sup>+</sup>-K<sup>+</sup>-ATPase activity, which explains the similar M-wave behavior in SL and AH (51). However, altitude acclimatization causes a downregulation of Na<sup>+</sup>-K<sup>+</sup>-ATPase pump concentration, and although this does not alter resting M-wave characteristics, it likely explains the exerciseinduced decrease in M-wave amplitude observed in CH (20, 33).

The lower postexercise M-wave amplitude in CH indicates a failure of the motor nerve/sarcolemma to propagate evoked stimuli to the contractile apparatus and might have masked potential benefits of acclimatization on fatigue resistance. Put simply, postexercise twitch forces might have been larger in CH if M-waves had remained unchanged from pre-exercise. If so, this would have resulted in a smaller exercise-induced reduction in  $Q_{tw,pot}$  in CH. Regardless, failure of neuromuscular transmission/sarcolemmal excitability contributes to reduced force output in response to a given central nervous activation and can therefore be

considered a key determinant of the impaired fatigue resistance in CH.

#### Central Fatigue

Exercise in AH induced a substantial degree of central fatigue, which was attenuated by  $\sim$ 50% when the same trial was repeated in CH (Table 2). This significant improvement, associated with acclimatization, clearly contrasts with the absence of a beneficial effect of CH on peripheral fatigue, as described above. Since the development of central fatigue is highly sensitive to O<sub>2</sub> (1), we attribute this improvement to the effects of high-altitude acclimatization on O<sub>2</sub> availability within the brain. Specifically, the cerebral O<sub>2</sub> delivery index at the end of exercise in CH was improved from AH (Table 1) (65) and may explain the lower degree of central fatigue in CH vs. AH.

Despite the similar CBFv and a slightly higher brain O<sub>2</sub> delivery in CH vs. SL (Table 1), which agrees with earlier Chacaltaya studies using the Kety-Schmidt technique to measure CBF/O<sub>2</sub> delivery (44), exercise-induced central fatigue was greater in CH. Two considerations discussed previously might account for this observation. First, the significant degree of peripheral fatigue in CH (vs. no fatigue at SL) presumably facilitated central fatigue via increases in inhibitory neural feedback from locomotor muscle (mediated by group III/IV muscle afferents), which limit central motor drive (3, 6). Second, although CaO2 and brain O2 delivery were similar/ higher in CH vs. SL, the still substantially lower P<sub>a</sub>O<sub>2</sub> might have contributed to the greater degree of central fatigue during exercise in this condition. Indeed, a low  $P_aO_2$  was recently suggested to impair cerebral metabolism (48) and alterations in neurotransmitter turnover (23), and both of these factors have been linked to the development of central fatigue (17, 53).

Taken together, the current findings provide a global indication of the positive effects of altitude acclimatization on the development of central fatigue during exercise. However, we cannot comment on the specific sites of the central motor pathway involved or the relative contribution of  $C_aO_2$  and  $P_aO_2$ in mediating these beneficial adaptations.

## Implications of Findings for Performance-Related Questions in CH

AH generally impairs endurance exercise performance (60). Prolonged exposure to hypoxia is known to recover some of this impairment (29, 52); however, SL performance is never matched at altitude. Our current findings indicate that the acclimatization-induced partial recovery of endurance performance occurs independent of any improvement of peripheral locomotor muscle fatigue from AH to CH. This insinuates that peripheral locomotor muscle fatigability, per se, does not contribute to the improvement of endurance performance observed from AH to CH. We therefore propose that the significantly attenuated central fatigue during exercise in severe CH likely accounts, at least in part, for the improvement of endurance performance associated with altitude acclimatization.

Mechanisms underlying the hypoxia-induced curtailment of central motor drive (i.e., increase in central fatigue) and endurance exercise performance have been documented previously to differ depending on the severity of arterial hypoxemia. Specifically, peripheral fatigue might depict the dominant determinant of central motor drive and thus the limiting factor above 70–75%  $S_pO_2$ . At more severe degrees of hypoxemia (<70%  $S_pO_2$ ), central motor drive and endurance performance might primarily—but not exclusively—be determined/limited by central nervous system (CNS) hypoxia (5). Since peripheral fatigue did not change with acclimatization in the present study, but  $S_pO_2$  increased from below to above the "threshold" described previously (5), reductions in central fatigue might be mediated mainly by improved arterial oxygenation and associated smaller influence of CNS hypoxia on central motor drive.

A recent Point:Counterpoint debate in this journal has focused on the potential existence/relevance of differences in physiological responses to exercise performed in normobaric vs. hypobaric hypoxia (43, 45). Since the present AH and CH experiments were performed in normobaric and hypobaric hypoxia, respectively, these potential differences, if indeed existent, might have influenced our findings.

#### Conclusion

AH exacerbates central and peripheral fatigue during endurance exercise. Our experiments indicate that acclimatization to high altitude significantly attenuates the development of central fatigue but does not improve the development of peripheral fatigue observed during whole-body endurance exercise in AH.

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#### DISCLOSURES

The authors declare no conflicts of interest.

## AUTHOR CONTRIBUTIONS

Author contributions: M.A., S.G., and A.W.S. conception and design of research; M.A., S.G., R.T., and A.W.S. performed experiments; M.A., S.G., and R.T. analyzed data; M.A., S.G., R.T., and A.W.S. interpreted results of experiments; M.A. and S.G. prepared figures; M.A. and A.W.S. drafted manuscript; M.A., S.G., R.T., A.W.S., A.T.L., and R.C.R. edited and revised manuscript; M.A., S.G., R.T., A.W.S., A.T.L., and R.C.R. approved final version of manuscript.

#### REFERENCES

- Amann M, Calbet JA. Convective oxygen transport and fatigue. J Appl Physiol 104: 861–870, 2008.
- Amann M, Pegelow DF, Jacques AJ, Dempsey JA. Inspiratory muscle work in acute hypoxia influences locomotor muscle fatigue and exercise performance of healthy humans. *Am J Physiol Regul Integr Comp Physiol* 293: R2036–R2045, 2007.
- Amann M, Proctor LT, Sebranek JJ, Pegelow DF, Dempsey JA. Opioid-mediated muscle afferents inhibit central motor drive and limit peripheral muscle fatigue development in humans. *J Physiol* 587: 271– 283, 2009.
- Amann M, Romer LM, Pegelow DF, Jacques AJ, Hess CJ, Dempsey JA. Effects of arterial oxygen content on peripheral locomotor muscle fatigue. J Appl Physiol 101: 119–127, 2006.
- Amann M, Romer LM, Subudhi AW, Pegelow DF, Dempsey JA. Severity of arterial hypoxaemia affects the relative contributions of peripheral muscle fatigue to exercise performance in healthy humans. J Physiol 581: 389–403, 2007.
- Amann M, Venturelli M, Ives SJ, McDaniel J, Layec G, Rossman MJ, Richardson RS. Peripheral fatigue limits endurance exercise via a sensory feedback-mediated reduction in spinal motoneuronal output. J Appl Physiol; doi:10.1152/japplphysiol.00049.2013.
- Baydur A, Behrakis PK, Zin WA, Jaeger M, Milic-Emili J. A simple method for assessing the validity of the esophageal balloon technique. *Am Rev Respir Dis* 126: 788–791, 1982.
- Bender PR, Groves BM, McCullough RE, McCullough RG, Huang SY, Hamilton AJ, Wagner PD, Cymerman A, Reeves JT. Oxygen transport to exercising leg in chronic hypoxia. J Appl Physiol 65: 2592– 2597, 1988.
- Bisgard GE, Forster HV. Ventilatory response to acute and chronic hypoxia. In: *Handbook of Physiology, Section 4: Environmental Physiology*, edited by Fregly MJ and Blatteis CM. Bethesda, MD: Oxford University Press, 1996, p. 1207–1239.
- 10. **Borg G.** *Borg's Perceived Exertion and Pain Scales.* Champaign, IL: Human Kinetics, 1998.
- Brooks GA, Wolfel EE, Butterfield GE, Cymerman A, Roberts AC, Mazzeo RS, Reeves JT. Poor relationship between arterial [lactate] and leg net release during exercise at 4,300 m altitude. *Am J Physiol Regul Integr Comp Physiol* 275: R1192–R1201, 1998.
- Calbet JA. Chronic hypoxia increases blood pressure and noradrenaline spillover in healthy humans. J Physiol 551: 379–386, 2003.
- Calbet JA, Boushel R, Radegran G, Sondergaard H, Wagner PD, Saltin B. Why is VO2 max after altitude acclimatization still reduced despite normalization of arterial O2 content? *Am J Physiol Regul Integr Comp Physiol* 284: R304–R316, 2003.
- Calbet JA, Lundby C. Air to muscle O2 delivery during exercise at altitude. *High Alt Med Biol* 10: 123–134, 2009.
- Calbet JA, Radegran G, Boushel R, Saltin B. On the mechanisms that limit oxygen uptake during exercise in acute and chronic hypoxia: role of muscle mass. J Physiol 587: 477–490, 2009.
- Caquelard F, Burnet H, Tagliarini F, Cauchy E, Richalet JP, Jammes Y. Effects of prolonged hypobaric hypoxia on human skeletal muscle function and electromyographic events. *Clin Sci (Lond)* 98: 329–337, 2000.
- Chaudhuri A, Behan PO. Fatigue and basal ganglia. J Neurol Sci 179: 34–42, 2000.
- Cibella F, Cuttitta G, Romano S, Grassi B, Bonsignore G, Milic-Emili J. Respiratory energetics during exercise at high altitude. *J Appl Physiol* 86: 1785–1792, 1999.
- Clausen T. Na+-K+ pump regulation and skeletal muscle contractility. *Physiol Rev* 83: 1269–1324, 2003.
- Clausen T, Nielsen OB, Harrison AP, Flatman JA, Overgaard K. The Na+,K+ pump and muscle excitability. *Acta Physiol Scand* 162: 183– 190, 1998.
- Clausen T, Overgaard K, Nielsen OB. Evidence that the Na+-K+ leak/pump ratio contributes to the difference in endurance between fastand slow-twitch muscles. *Acta Physiol Scand* 180: 209–216, 2004.
- 22. Cruz JC. Mechanics of breathing in high altitude and sea level subjects. *Respir Physiol* 17: 146–161, 1973.
- Davis JN, Carlsson A, MacMillan V, Siesjo BK. Brain tryptophan hydroxylation: dependence on arterial oxygen tension. *Science* 182: 72– 74, 1973.

- De Blasi RA, Ferrari M, Natali A, Conti G, Mega A, Gasparetto A. Noninvasive measurement of forearm blood flow and oxygen consumption by near-infrared spectroscopy. J Appl Physiol 76: 1388–1393, 1994.
- Dempsey JA, Amann M, Romer LM, Miller JD. Respiratory system determinants of peripheral fatigue and endurance performance. *Med Sci Sports Exerc* 40: 457–461, 2008.
- Duncan A, Meek JH, Clemence M, Elwell CE, Tyszczuk L, Cope M, Delpy DT. Optical pathlength measurements on adult head, calf and forearm and the head of the newborn infant using phase resolved optical spectroscopy. *Phys Med Biol* 40: 295–304, 1995.
- Fowles JR, Green HJ, Tupling R, O'Brien S, Roy BD. Human neuromuscular fatigue is associated with altered Na+-K+-ATPase activity following isometric exercise. J Appl Physiol 92: 1585–1593, 2002.
- Fulco CS, Cymerman A, Muza SR, Rock PB, Pandolf KB, Lewis SF. Adductor pollicis muscle fatigue during acute and chronic altitude exposure and return to sea level. J Appl Physiol 77: 179–183, 1994.
- Fulco CS, Kambis KW, Friedlander AL, Rock PB, Muza SR, Cymerman A. Carbohydrate supplementation improves time-trial cycle performance during energy deficit at 4,300-m altitude. *J Appl Physiol* 99: 867–876, 2005.
- Garner SH, Sutton JR, Burse RL, McComas AJ, Cymerman A, Houston CS. Operation Everest II: neuromuscular performance under conditions of extreme simulated altitude. *J Appl Physiol* 68: 1167–1172, 1990.
- Goodall S, Gonzalez-Alonso J, Ali L, Ross EZ, Romer LM. Supraspinal fatigue after normoxic and hypoxic exercise in humans. *J Physiol* 590: 2767–2782, 2012.
- Goodall S, Ross EZ, Romer LM. Effect of graded hypoxia on supraspinal contributions to fatigue with unilateral knee-extensor contractions. J Appl Physiol 109: 1842–1851, 2010.
- Green H, Roy B, Grant S, Burnett M, Tupling R, Otto C, Pipe A, McKenzie D. Downregulation in muscle Na(+)-K(+)-ATPase following a 21-day expedition to 6,194 m. J Appl Physiol 88: 634–640, 2000.
- Harms CA, Babcock MA, McClaran SR, Pegelow DF, Nickele GA, Nelson WB, Dempsey JA. Respiratory muscle work compromises leg blood flow during maximal exercise. *J Appl Physiol* 82: 1573–1583, 1997.
- 35. Heinonen I, Nesterov SV, Kemppainen J, Nuutila P, Knuuti J, Laitio R, Kjaer M, Boushel R, Kalliokoski KK. Role of adenosine in regulating the heterogeneity of skeletal muscle blood flow during exercise in humans. *J Appl Physiol* 103: 2042–2048, 2007.
- Heinonen IH, Kemppainen J, Kaskinoro K, Peltonen JE, Borra R, Lindroos M, Oikonen V, Nuutila P, Knuuti J, Boushel R, Kalliokoski KK. Regulation of human skeletal muscle perfusion and its heterogeneity during exercise in moderate hypoxia. *Am J Physiol Regul Integr Comp Physiol* 299: R72–R79, 2010.
- Hogan MC, Richardson RS, Haseler LJ. Human muscle performance and PCr hydrolysis with varied inspired oxygen fractions: a 31P-MRS study. J Appl Physiol 86: 1367–1373, 1999.
- Johnson BD, Saupe KW, Dempsey JA. Mechanical constraints on exercise hyperpnea in endurance athletes. J Appl Physiol 73: 874–886, 1992.
- Katayama K, Amann M, Pegelow DF, Jacques AJ, Dempsey JA. Effect of arterial oxygenation on quadriceps fatigability during isolated muscle exercise. *Am J Physiol Regul Integr Comp Physiol* 292: R1279– R1286, 2007.
- Kayser B, Bokenkamp R, Binzoni T. Alpha-motoneuron excitability at high altitude. Eur J Appl Physiol 66: 1–4, 1993.
- Lundby C, Sander M, van Hall G, Saltin B, Calbet JA. Maximal exercise and muscle oxygen extraction in acclimatizing lowlanders and high altitude natives. J Physiol 573: 535–547, 2006.
- 42. Merton PA. Voluntary strength and fatigue. J Physiol 123: 553–564, 1954.
- Millet GP, Faiss R, Pialoux V. Point: hypotaic hypoxia induces different physiological responses from normobaric hypoxia. *J Appl Physiol* 112: 1783–1784, 2012.
- 44. Moller K, Paulson OB, Hornbein TF, Colier WN, Paulson AS, Roach RC, Holm S, Knudsen GM. Unchanged cerebral blood flow and oxidative metabolism after acclimatization to high altitude. J Cereb Blood Flow Metab 22: 118–126, 2002.
- Mounier R, Brugniaux JV. Counterpoint: hypobaric hypoxia does not induce different responses from normobaric hypoxia. J Appl Physiol 112: 1784–1786, 2012.

- Nielsen OB, Clausen T. The Na+/K(+)-pump protects muscle excitability and contractility during exercise. *Exerc Sport Sci Rev* 28: 159–164, 2000.
- 47. Poulin MJ, Fatemian M, Tansley JG, O'Connor DF, Robbins PA. Changes in cerebral blood flow during and after 48 h of both isocapnic and poikilocapnic hypoxia in humans. *Exp Physiol* 87: 633–642, 2002.
- Rasmussen P, Nielsen J, Overgaard M, Krogh-Madsen R, Gjedde A, Secher NH, Petersen NC. Reduced muscle activation during exercise related to brain oxygenation and metabolism in humans. *J Physiol* 588: 1985–1995, 2010.
- Roberts AC, Butterfield GE, Cymerman A, Reeves JT, Wolfel EE, Brooks GA. Acclimatization to 4,300-m altitude decreases reliance on fat as a substrate. *J Appl Physiol* 81: 1762–1771, 1996.
- Rowell LB, Saltin B, Kiens B, Christensen NJ. Is peak quadriceps blood flow in humans even higher during exercise with hypoxemia? *Am J Physiol Heart Circ Physiol* 251: H1038–H1044, 1986.
- Sandiford SD, Green HJ, Duhamel TA, Schertzer JD, Perco JD, Ouyang J. Muscle Na-K-pump and fatigue responses to progressive exercise in normoxia and hypoxia. *Am J Physiol Regul Integr Comp Physiol* 289: R441–R449, 2005.
- Schuler B, Thomsen JJ, Gassmann M, Lundby C. Timing the arrival at 2340 m altitude for aerobic performance. *Scand J Med Sci Sports* 17: 588–594, 2007.
- Secher NH, Seifert T, Van Lieshout JJ. Cerebral blood flow and metabolism during exercise: implications for fatigue. *J Appl Physiol* 104: 306–314, 2008.
- Serrador JM, Picot PA, Rutt BK, Shoemaker JK, Bondar RL. MRI measures of middle cerebral artery diameter in conscious humans during simulated orthostasis. *Stroke* 31: 1672–1678, 2000.
- Stary CM, Hogan MC. Effect of varied extracellular PO2 on muscle performance in Xenopus single skeletal muscle fibers. *J Appl Physiol* 86: 1812–1816, 1999.
- 56. Sutton JR, Reeves JT, Wagner PD, Groves BM, Cymerman A, Malconian MK, Rock PB, Young PM, Walter SD, Houston CS. Operation Everest II: oxygen transport during exercise at extreme simulated altitude. J Appl Physiol 64: 1309–1321, 1988.
- Taylor AD, Bronks R, Smith P, Humphries B. Myoelectric evidence of peripheral muscle fatigue during exercise in severe hypoxia: some references to m. vastus lateralis myosin heavy chain composition. *Eur J Appl Physiol Occup Physiol* 75: 151–159, 1997.
- Thoden JS, Dempsey JA, Reddan WG, Birnbaum ML, Forster HV, Grover RF, Rankin J. Ventilatory work during steady-state response to exercise. *Fed Proc* 28: 1316–1321, 1969.
- 59. Van Beekvelt MC, Colier WN, Wevers RA, Van Engelen BG. Performance of near-infrared spectroscopy in measuring local O(2) consumption and blood flow in skeletal muscle. J Appl Physiol 90: 511–519, 2001.
- Wehrlin JP, Hallen J. Linear decrease in VO2max and performance with increasing altitude in endurance athletes. *Eur J Appl Physiol* 96: 404–412, 2006.
- Westerblad H, Allen DG, Lannergren J. Muscle fatigue: lactic acid or inorganic phosphate the major cause? *News Physiol Sci* 17: 17–21, 2002.
- Willie CK, Macleod DB, Shaw AD, Smith KJ, Tzeng YC, Eves ND, Ikeda K, Graham J, Lewis NC, Day TA, Ainslie PN. Regional brain blood flow in man during acute changes in arterial blood gases. *J Physiol* 590: 3261–3275, 2012.
- 63. Wilson MH, Edsell ME, Davagnanam I, Hirani SP, Martin DS, Levett DZ, Thornton JS, Golay X, Strycharczuk L, Newman SP, Montgomery HE, Grocott MP, Imray CH, Caudwell Xtreme Everest Research Group. Cerebral artery dilatation maintains cerebral oxygenation at extreme altitude and in acute hypoxia—an ultrasound and MRI study. J Cereb Blood Flow Metab 31: 2019–2029, 2011.
- 64. Wolfel EE, Groves BM, Brooks GA, Butterfield GE, Mazzeo RS, Moore LG, Sutton JR, Bender PR, Dahms TE, McCullough RE, Huang SY, Sun SF, Grover RF, Hultgren HN, Reeves JT. Oxygen transport during steady-state submaximal exercise in chronic hypoxia. J Appl Physiol 70: 1129–1136, 1991.
- Xu K, Lamanna JC. Chronic hypoxia and the cerebral circulation. J Appl Physiol 100: 725–730, 2006.

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## AltitudeOmics: Exercise-induced supraspinal fatigue is attenuated in healthy humans after acclimatisation to high altitude

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## Abstract

Aims: We asked whether acclimatisation to chronic hypoxia (CH) attenuates the level of supraspinal fatigue that is observed after locomotor exercise in acute hypoxia (AH). Methods: Seven recreationally-active participants performed identical bouts of constant-load cycling (131±39W, 10.1±1.4min) on three occasions: 1) in normoxia (N, P<sub>1</sub>O<sub>2</sub>, 147.1mmHg); 2) in AH (F<sub>1</sub>O<sub>2</sub>, 0.105; P<sub>1</sub>O<sub>2</sub>, 73.8mmHg); 3) after 14 days in CH (5,260m; P<sub>I</sub>O<sub>2</sub>, 75.7mmHg). Throughout trials, prefrontal-cortex tissue oxygenation and middle cerebral artery blood velocity (MCAv) were assessed using nearinfrared-spectroscopy and transcranial Doppler sonography. Pre- and post-exercise twitch responses to femoral nerve stimulation and transcranial magnetic stimulation were obtained to assess neuromuscular and corticospinal function. Results: In AH, prefrontal oxygenation declined at rest ( $\Delta7\pm5\%$ ) and end-exercise ( $\Delta26\pm13$ ) (P<0.01); the degree of deoxygenation in AH was greater than N and CH (P<0.05). The cerebral  $O_2$  delivery index (MCA<sub>v</sub>×C<sub>a</sub>O<sub>2</sub>) was 19±14% lower during the final minute of exercise in AH compared to N (P=0.013) and 20±12% lower compared to CH (P=0.040). Maximum voluntary and potentiated twitch force were decreased below baseline after exercise in AH and CH, but not N. Cortical voluntary activation decreased below baseline after exercise in AH ( $\Delta$ 11%, P=0.014), but not CH ( $\Delta$ 6%, P=0.174) or N ( $\Delta$ 4%, P=0.298). A twofold greater increase in motor evoked potential amplitude was evident after exercise in CH compared to AH and N. Conclusion: These data indicate that exacerbated supraspinal fatigue after exercise in AH is attenuated after 14 days of acclimatisation to altitude. The reduced development of supraspinal fatigue in CH may have been attributable to increased corticospinal excitability, consequent to an increased cerebral O<sub>2</sub> delivery.

Key words: adaptation, altitude, exercise, transcranial magnetic stimulation

#### Glossary

 $C_aO_2$ , arterial  $O_2$  content; CSP, cortical silent period; ERT, estimated resting twitch;  $F_1O_2$ , fraction of inspired  $O_2$ ;  $f_R$ , respiratory frequency; [Hb], haemoglobin concentration; MCA<sub>V</sub>, middle cerebral artery blood velocity; MEP, motor evoked potential;  $M_{max}$ , maximum M-wave; MVC, maximum voluntary contraction;  $P_aO_2$ , partial pressure of arterial  $O_2$ ;  $P_1O_2$ , partial pressure of inspired  $O_2$ ;  $Q_{tw,pot}$ , potentiated quadriceps twitch force; rMT, resting motor threshold; SIT, superimposed twitch;  $S_pO_2$ , arterial  $O_2$  saturation; TMS, transcranial magnetic stimulation;  $\dot{V}CO_2$ , carbon dioxide output;  $\dot{V}_E$ , minute ventilation;  $\dot{V}O_2$ , oxygen uptake;  $V_T$ , tidal volume.

## Introduction

The mechanisms underpinning impairments in exercise performance in hypoxia are not fully understood, but multiple peripheral and central mechanisms of fatigue have been proposed (Amann and Calbet, 2008, Nybo and Rasmussen, 2007, Perrey and Rupp, 2009). The rate of development of peripheral fatigue is increased during intense locomotor exercise in acute hypoxia (Amann et al., 2006b, Goodall et al., 2012). This has been documented in numerous human studies as an increased decline in the force response to motor nerve stimulation after exercise and an increased rate of rise in electromyogram (EMG) signals during exercise (Amann and Calbet, 2008). Amann et al. (2006a) suggested that the accelerated development of peripheral fatigue and associated intramuscular metabolic changes in acute moderate hypoxia restricts central motor drive preventing excessive end-exercise locomotor muscle fatigue under conditions of attenuated arterial oxygenation. It was subsequently demonstrated that in acute severe hypoxia, peripheral fatigue becomes the less important variable and the primary limitation to exercise transfers to a hypoxia-sensitive central component of fatigue (Amann et al., 2007). Less is known about the mechanism(s) of fatigue during locomotor exercise in chronic hypoxia. We recently reported the accelerated development of peripheral fatigue after locomotor exercise in acute hypoxia to be similar after a period of acclimatisation (14 days) to high altitude; conversely, the level of central fatigue was attenuated (Amann et al., 2013). The measure of central fatigue, however, was determined using peripheral stimulation and the responsiveness of the brain-to-muscle pathway after a period of chronic hypoxia remains unknown.

Transcranial magnetic stimulation (TMS) has been used to specify the site of fatigue within the central nervous system in acute severe hypoxia (Goodall *et al.*, 2012, Goodall *et al.*, 2010). When TMS is delivered over the motor cortex during a maximal voluntary contraction (MVC), it is possible to detect a twitch-like increment in force in the active muscle. That is, despite maximal effort, motor cortical output at the time of stimulation is insufficient to drive the motoneurons maximally. An increase in this increment in force after exercise provides evidence of a reduced cortical voluntary activation, indicative of supraspinal fatigue (Gandevia *et al.*, 1996, Todd *et al.*, 2003). Further, EMG recordings in response to cortical stimuli (motor evoked potential [MEP]) can be monitored to assess changes in excitability of the brain to muscle pathway. Descending volleys evoked from cortical stimulation depend on the stimulus intensity and excitability of corticospinal cells, whereas responses in the muscle depend on transmission through relevant excitatory and inhibitory interneurons and excitability of the motoneuron pool (Taylor and Gandevia, 2001). Hypoxia affects

neuronal function *in-vitro* (Nieber *et al.*, 1999), however, acute hypoxia appears to have negligible effects on resting MEPs elicited by TMS (Goodall *et al.*, 2010, Rupp *et al.*, 2012, Szubski *et al.*, 2006). A MEP evoked during muscular contraction is followed by an interval of EMG silence, the so-called cortical silent period (CSP). The initial phase of the CSP has been attributed to inhibitory spinal mechanisms (Inghilleri *et al.*, 1993), whereas the later period (>100 ms) represents increased cortical inhibition (Chen *et al.*, 1999, Inghilleri *et al.*, 1993, Taylor and Gandevia, 2001). Szubski *et al.* (2006) found a shorter CSP in acute hypoxia, suggestive of a reduced corticospinal inhibition during the exercise.

Responsiveness of the corticospinal pathway and the associated development of central fatigue after locomotor exercise during periods of prolonged hypoxia have not been studied. A recent investigation found an increase in corticospinal excitability (increased resting MEP) after a period of prolonged acute hypoxia (Rupp *et al.*, 2012); however, the mechanisms for this response and the associated effects upon the development of central fatigue during locomotor exercise have not been studied. We have recently related the development of supraspinal fatigue during exercise in severe acute hypoxia to a reduction in cerebral O<sub>2</sub> availability (Goodall *et al.*, 2012). Acclimatisation to altitude not only brings about improvements in arterial oxygenation, but also improvements in cerebrovascular function (Ainslie and Ogoh, 2009, Lucas *et al.*, 2011). It is unknown how haematologic (e.g., hemodynamic and cerebrovascular) adaptations might serve to impact corticospinal excitability and the development of supraspinal fatigue during locomotor exercise in chronic hypoxia. Accordingly, the aim of the present study was to assess corticospinal excitability and supraspinal fatigue after locomotor exercise in chronic hypoxia. We hypothesised that altered cerebrovascular and corticospinal responses after a period of acclimatisation to high altitude would reduce the severity of supraspinal fatigue compared to that observed in acute hypoxia.

## Methods

## **Ethical Approval**

All procedures conformed to the Declaration of Helsinki and were approved by the Universities of Colorado Denver, Oregon and Utah Institutional Review Boards and the US Department of Defense Human Research Protection Office.

## Participants

This study was conducted as part of the AltitudeOmics project examining the integrative physiology of human responses to hypoxia (Subudhi *et al.* under review at PLoSOne). After written informed consent, seven (five male) recreationally active sea level habitants participated in the study (mean ± SD age,  $21 \pm 1$  yr; stature,  $1.78 \pm 0.10$  m; body mass,  $69 \pm 11$  kg; maximum O<sub>2</sub> uptake [ $\dot{V}O_{2max}$ ],  $46.4 \pm$ 8.2 ml·kg<sup>-1</sup>·min<sup>-1</sup> [participant IDs: 1,2,3,5,6,7,10]). The participants were non-smokers, free from cardiorespiratory disease, born and raised at <1500 m, and had not travelled to elevations >1000 m in the 3 months prior to investigation. Participants arrived at the laboratory in a rested and fully hydrated state, at least 3 h postprandial, and avoided strenuous exercise in the 48 h preceding each trial. They also refrained from caffeine for 12 h before each test, while alcohol and prophylactic altitude medication were prohibited for the entire duration of the investigation. All of the subjects participated in a companion study investigating the acclimatisation-induced effects on peripheral measures of neuromuscular fatigue (Amann *et al.*, 2013); while the data were obtained from the same protocol described below, the primary TMS and cerebral oxygenation related outcome measures in the current study do no overlap with previous analyses.

## **Experimental design**

Participants completed a preliminary trial and three experimental trials. Each trial was conducted at the same time of day, and separated by at least 5 d during a 12 wk period. During the preliminary trial, participants were thoroughly familiarized with the methods used to assess neuromuscular function and performed a maximal incremental exercise test in normoxia for the determination of  $Vo_{2max}$  and peak workload ( $W_{peak}$ ); further maximal incremental tests were performed in AH and CH (Subudhi et al. under review at PLoSOne). During the experimental trials, participants performed constant-load exercise at a workload equal to 50% W<sub>peak</sub> obtained in the preliminary trial: 1) to the limit of tolerance in acute normobaric hypoxia (AH: F<sub>1</sub>O<sub>2</sub> = 0.105; Eugene, Oregon, barometric pressure [BP] =  $750 \pm 2$  mmHg; P<sub>1</sub>O<sub>2</sub> =  $73.8 \pm 0.2$  mmHg); 2) for the same absolute intensity and duration as in trial 1, but in normoxia (N: Eugene, Oregon, BP =  $750 \pm 2$  mmHg; P<sub>1</sub>O<sub>2</sub> = 147.1 ± 0.5 mmHg); and 3) for the same absolute intensity and duration as in trial 1, but after 14 d at 5,260 m above sea level (CH: Mt. Chacaltaya, Bolivia, BP =  $409 \pm 1$  mmHg; P<sub>1</sub>O<sub>2</sub> = 75.7 \pm 0.1 mmHg). Participants were flown to La Paz, Bolivia where they spent two nights at low altitude (Coroico, 1,525 m), before being driven to the Chacaltaya Research Station at 5,260 m. Before and within 2.5 min after each exercise trial, twitch responses to supramaximal femoral nerve stimulation and TMS were obtained to assess fatigue. During AH, the post-exercise measurements were made while participants continued to breathe the hypoxic gas. Cerebrovascular, cardiorespiratory and perceptual responses, as well as EMG activity of the vastus lateralis (VL), were assessed throughout each trial.

## Force and EMG recordings

Knee-extensor force during voluntary and evoked contractions was measured using a calibrated load cell (Tedea, Basingstoke, UK). The load cell was fixed to a custom-built chair and connected to a non-compliant cuff attached around the participant's right leg just superior to the right ankle. Participants sat upright in the chair with the hips and knees at 90° of flexion. EMG activity was recorded from the VL and biceps femoris (BF). Surface electrodes were placed 2 cm apart over the muscle bellies and a reference electrode was placed over the patella. The electrodes were used to record the compound muscle action potential (M-wave) elicited by electrical stimulation of the femoral nerve and the MEP elicited by TMS. Signals were amplified (gain 1000; Force: custom-built bridge amplifier; EMG: PowerLab 26T, ADInstruments Inc, Oxfordshire, UK), band-pass filtered (EMG only: 20-2000 Hz), digitised (4 kHz; PowerLab 26T, ADInstruments Inc), acquired and later analysed (LabChart v7.0, ADInstruments Inc).

## Neuromuscular function

Force and EMG variables were assessed before and immediately after each exercise trial. Prior to each trial, MVC force was determined from three, 3 s contractions. Femoral nerve stimulation was delivered at rest ~2 s after the MVC to determine the potentiated quadriceps twitch force ( $Q_{tw,pot}$ ). TMS was delivered during brief (~5 s) maximal and submaximal voluntary contractions for the determination of cortical voluntary activation. Each set of contractions comprised 100, 75, and 50% MVC efforts separated by ~5 s of rest. The contraction sets were repeated three times, with 15 s between each set. Visual feedback of the target force was provided via a computer monitor.

## Femoral nerve stimulation

Single electrical stimuli (200  $\mu$ s) were delivered to the right femoral nerve via surface electrodes (CF3200, Nidd Valley Medical Ltd, North Yorkshire, UK) and a constant-current stimulator (DS7AH, Digitimer Ltd, Welwyn Garden City, Hertfordshire, UK). The cathode was positioned over the nerve high in the femoral triangle; the anode was placed midway between the greater trochanter and the iliac crest. The site of stimulation that produced the largest resting twitch amplitude and M-wave ( $M_{max}$ ) was located. Single stimuli were delivered beginning at 100 mA and increasing by 20 mA until

plateaus occurred in twitch amplitude and  $M_{max}$ . Supramaximal stimulation was ensured by increasing the final intensity by 30% (mean current 253 ± 60 mA).

#### Transcranial magnetic stimulation

TMS was delivered via a concave double cone coil (110 mm diameter; maximum output 1.4 T) powered by a mono-pulse magnetic stimulator (Magstim 200, The Magstim Company Ltd, Whitland, UK). The coil was held over the vertex to preferentially stimulate the left hemisphere (posteroanterior intracranial current flow), and was placed in an optimal position to elicit a large MEP in the VL and a small MEP in the antagonist (BF). The optimal coil position was marked on the scalp with indelible ink to ensure reproducibility of the stimulation. Resting motor threshold (rMT) was determined at the beginning of each experimental trial. Briefly, TMS was first delivered with the coil placed over the optimal site of stimulation at a sub-threshold intensity of 35% maximum stimulator output. Stimulus intensity was then increased in 5% steps until consistent motor evoked potentials (MEPs) with peak-to-peak amplitudes of more than 50  $\mu$ V were evoked. Thereafter, stimulus intensity was reduced in 1% steps until an intensity was reached that elicited an MEP of at least 50  $\mu$ V in 5 out of 10 trials (Groppa *et al.*, 2012). The stimulation intensity that elicited rMT was increased by 30%; thus, the experimental stimulation intensity was 130% of rMT. This stimulation intensity elicited a large MEP in the VL (area between 60 and 100% of M<sub>max</sub> during knee-extensor contractions ≥50% MVC; Figure 1); indicating the TMS stimulus activated a high proportion of knee extensor motor units, while causing only a small MEP in the BF (amplitude <20% of MEP during kneeextensor contractions).

#### Constant-load exercise

Participants sat on an electromagnetically-braked cycle ergometer (Velotron Dynafit Pro, Racermate, Seattle, WA) while baseline cardiorespiratory and cerebrovascular data were collected for 3 min. The participants warmed-up for 5 min at 10%  $W_{peak}$  (26 ± 8 W) before the workload was increased to 50% normoxic  $W_{peak}$  (131 ± 39 W). This intensity was chosen to maximise the tolerable duration of exercise in the hypoxic conditions. The participants remained seated throughout exercise and maintained a target pedal cadence equivalent to that chosen during the incremental exercise test (88 ± 3 rpm). Task-failure was reached when cadence dropped below 60% of the target rpm for >5 s. Constant load exercise was performed firstly in AH; the achieved time (10.1 ± 1.4 min) was then replicated in N and CH.

## Tissue oxygenation and cerebrovascular responses

Cerebral oxygenation was assessed using a multi-channel NIRS instrument (Oxymon III, Artinis) (Subudhi et al., 2009, Subudhi et al., 2011). Changes in oxygenated , deoxygenated and total cerebral haeme concentrations ( $\mu$ M) were expressed relative to the resting baseline recorded in each experimental condition. Arterial oxygen saturation was estimated using forehead pulse oximetry (S<sub>0</sub>O<sub>2</sub>; Model N-595, Nellcor, Pleasonton, CA). Excellent agreement between the pulse oximeter and arterial  $O_2$  saturation across the range of values in the present study has been published (Romer et al., 2007). Hemoglobin concentration [Hb] was measured (OSM-3, Radiometer, Copenhagen, Denmark) in resting arterial blood samples. Samples were collected during the primary physiological protocols at sea level (2-4 d prior to the first exercise trial in the present study) and on the 16<sup>th</sup> day at 5,260 m (2 d following the constant load exercise trial in the present study) (Subudhi et al. under review at PLoSOne). Arterial  $O_2$  content ( $C_aO_2$ ) was estimated using the equation: ([Hb]  $\times$  1.39  $\times$  S<sub>p</sub>O<sub>2</sub> / 100). Resting [Hb] in combination with the measured S<sub>p</sub>O<sub>2</sub> during the exercise protocol were used to obtain C<sub>a</sub>o<sub>2</sub> throughout exercise in all conditions. Blood velocity in the left middle cerebral artery (MCA<sub>v</sub>) was determined using transcranial Doppler (Spencer Technologies, Seattle, WA). The custom-made NIRS headset was modified to hold a 2 MHz probe positioned over the left temporal window. Measurements were optimised at an average penetration depth of  $50 \pm 3$ mm. An index of cerebral  $O_2$  delivery was calculated as the product of MCA<sub>v</sub> and  $C_aO_2$ . It was assumed that changes in MCA<sub>v</sub> would reflect changes in cerebral blood flow based on evidence that the middle cerebral artery diameter changes minimally in response to hypoxia and hypocapnia (Poulin and Robbins, 1996).

## Cardiorespiratory and perceptual responses

Ventilatory and pulmonary gas exchange indices were assessed using an online system (in AH & N Medical Graphics PFX, St. Paul, MN, USA; & in CH Oxigraf O<sub>2</sub>cap, Mountain View, CA, USA). Heart rate was identified from the peak MCA<sub>v</sub> envelopes. Ratings of perceived exertion for dyspnea and limb discomfort were obtained using the CR10 scale at baseline and every minute throughout exercise (Borg, 1982). In CH, symptoms of acute mountain sickness were assessed on the day of a trial using the Lake Louise Score (Roach *et al.*, 1993).

## Data analysis

Cortical voluntary activation was assessed by measuring the force responses to motor-cortex stimulation during submaximal and maximal contractions. Corticospinal excitability increases during

voluntary contraction (Rothwell *et al.*, 1991); thus, we estimated the amplitude of the resting twitch evoked by TMS (ERT; Goodall *et al.*, 2009, Sidhu *et al.*, 2009a). Cortical voluntary activation (%) was subsequently quantified using the equation:  $(1 - [SIT / ERT] \times 100)$ .

The peak-to-peak amplitude and area of evoked MEPs and  $M_{max}$  were measured offline. To ensure the motor cortex stimulus activated a high proportion of the knee-extensor motor units, the area of vastus lateralis MEP was normalised to that of  $M_{max}$  elicited during the MVC at the beginning of each trial (Taylor *et al.*, 1999) (Figure 1). The duration of the CSP evoked by TMS during MVC was quantified as the duration from stimulation to the continuous resumption of post-stimulus EMG exceeding  $\pm$  2 SD of pre-stimulus EMG (>50 ms prior to stimulus). VL EMG signals during exercise were rectified and smoothed (15 ms), then quantified as the mean integrated area during each cycle revolution and averaged over each minute of exercise. A computer algorithm identified the onset and offset of activity where the rectified EMG signals deviated >2 SD from baseline for >100 ms.

## **Reliability coefficients**

On a separate day, the responses to TMS, femoral nerve stimulation and MVC were repeated twice in all participants. The two assessment procedures were separated by a 2 min walk followed by 5 min of rest. Coefficient of variation (CV) and intraclass correlation coefficient (ICC) were calculated to evaluate test-retest reliability. All correlations were statistically significant and indicated, in combination with the CVs, a high level of reproducibility: cortical voluntary activation, CV = 1.4%, ICC = 0.82; CSP, CV = 7.1%, ICC = 0.93; ERT, CV = 10.2%, ICC = 0.84; MEP/M<sub>max</sub>, CV = 9.6%, ICC = 0.66; M<sub>max</sub>, CV = 11.4%, ICC = 0.98; 100% MVC MEP, CV = 14.1%, ICC = 0.96; 75% MVC MEP, CV = 10.2%, ICC = 0.98; 50% MVC MEP, CV = 7.2%, ICC = 0.99; MVC, CV = 4.7%, ICC = 0.94; Q<sub>tw,pot</sub>, CV = 4.8%, ICC = 0.97.

## Statistical analysis

Data are presented as means  $\pm$  SD in the text and means  $\pm$  SE in the figures. A 3 × 2 repeated measures ANOVA on condition (3 [AH, N, CH]) and time (2 [pre, post]) was used to test for withingroup differences. When ANOVA revealed significant interactions, post-hoc comparisons were made using the least significant differences test. Statistical significance was set at P < 0.05. All analyses were conducted using SPSS (v19, IBM Corporation, New York, USA).

## Results

## **Exercise responses**

The exercise workload was  $131 \pm 39$  W (50% N W<sub>peak</sub>), which equated to 83% W<sub>peak</sub> in AH and 74% W<sub>peak</sub> in CH. Cerebral oxygenation data are shown in Figure 2. During N, oxyhaemoglobin was unchanged from baseline to warm up and total haemoglobin was increased during the final minute of exercise (P = 0.658 and 0.007, respectively). During AH, deoxygenated haemoglobin increased from baseline to warm up (P = 0.006); this response was exaggerated towards end exercise (P < 0.001). During CH, deoxygenated haemoglobin increased at end exercise (P = 0.015) in line with increased total haemoglobin (P = 0.043). Overall, these results demonstrate that the degree of cerebral deoxygenation ( $\Delta$  deoxygenated haemoglobin) in AH was greater than that observed in N and CH (P < 0.05).

 $S_pO_2$  and MCA<sub>v</sub> data are shown in Figure 3. Acute exposure to hypoxia decreased  $S_pO_2$  at rest ( $\Delta 7 \pm 4\%$ ; P = 0.009) and during the final minute of exercise ( $\Delta 34 \pm 10\%$ ; P < 0.001). Resting  $S_pO_2$  in CH was 85 ± 2% (P < 0.001 vs. N; P = 0.330 vs. AH), and in the final minute of exercise had fallen to 78 ± 5% (P < 0.001 vs. N; P = 0.002 vs. AH). No changes in  $S_pO_2$  were apparent in N (P > 0.702). Resting MCA<sub>v</sub> did not differ between conditions at baseline (pooled average, 54 ± 9 cm·s<sup>-1</sup>; P = 0.544). MCA<sub>v</sub> did not increase from rest at any time point in N (P > 0.108). MCA<sub>v</sub> increased from rest to the final minute of exercise in AH (40 ± 15%; P < 0.001) and CH (25 ± 14%; P = 0.016), but did not differ between conditions (Figure 3).

Hemoglobin concentration was  $1.42 \pm 0.03 \text{ g}\cdot\text{L}^{-1}$  in N and  $1.63 \pm 0.31 \text{ g}\cdot\text{L}^{-1}$  in CH (P = 0.005). Resting P<sub>a</sub>O<sub>2</sub> was reduced in AH compared to N (39.1 ± 4.8 vs. 103.3 ± 8.7 mmHg, P < 0.001), was increased in CH relative to AH (58.8 ± 3.2 mmHg, P < 0.001), but was still lower than N (P < 0.001). C<sub>a</sub>O<sub>2</sub> was lower at rest in AH vs. N (19.8 ± 1.9 vs. 21.5 ± 2.9 ml·dl<sup>-1</sup>; P = 0.013); during the final minute of exercise C<sub>a</sub>O<sub>2</sub> in AH was 36 ± 8% lower than N (P < 0.001) and 22 ± 9% lower than in CH (P = 0.001). C<sub>a</sub>O<sub>2</sub> was lower at rest in CH vs. N (19.4 ± 2.6 vs. 21.5 ± 2.9 ml·dl<sup>-1</sup>; P < 0.001) and during the final minute of exercise (17.6 ± 2.9 vs. 21.2 ± 2.9 ml·dl<sup>-1</sup>; P = 0.725). Consequently, cerebral O<sub>2</sub> delivery index (MCA<sub>v</sub> × C<sub>a</sub>O<sub>2</sub>) was 19 ± 14% lower during the final minute of exercise in AH compared to N (P = 0.013) and 20 ± 12% lower compared to CH (P = 0.040). No differences were evident between N and CH at rest (P = 0.783) or during the final minute of exercise (P = 0.797) (Figure 3).

Cardiorespiratory data are shown in Table 1. Respiratory frequency and minute ventilation (V<sub>E</sub>) rose substantially over time in all conditions.  $\dot{V}_E/\dot{V}CO_2$  during the final minute of exercise in AH and CH was approximately twofold greater than in N (P < 0.001);  $\dot{V}_E/\dot{V}CO_2$  during the final minute of exercise was 28% higher in CH compared to AH (P < 0.001). During the final minute of exercise, whole body  $\dot{V}O_2$ was not different across the three conditions (P = 0.411). Dyspnea and limb discomfort at endexercise were higher in AH compared to N (P < 0.001 and P = 0.048, respectively), but were not different compared to CH (P = 0.714 and 0.549, respectively). Integrated EMG activity at end exercise was higher in AH compared to N (32%; P = 0.029), but not CH (16%; P = 0.303). There were no reported symptoms of acute mountain sickness during CH.

## Pre- and post-exercise responses

Peripheral and central measures of excitability are shown in Table 2.

#### Neuromuscular responses

MVC did not differ between conditions at baseline (AH,  $392 \pm 77$  N; N,  $386 \pm 90$  N; CH,  $376 \pm 39$  N; P = 0.942). MVC was reduced post-exercise in AH ( $339 \pm 77$  N, P = 0.011) and CH ( $346 \pm 93$  N, P = 0.032), but not N ( $387 \pm 87$  N, P = 0.684). The reductions in MVC were not different between conditions (P  $\ge$  0.119). Q<sub>tw,pot</sub> did not differ between conditions at baseline (AH,  $107 \pm 13$  N; N,  $105 \pm 12$  N; CH,  $110 \pm 16$  N; P = 0.752). Q<sub>tw,pot</sub> was reduced post-exercise in AH ( $84 \pm 14$  N, P = 0.005) and CH ( $90 \pm 18$  N, P = 0.011), but not N ( $102 \pm 12$  N, P = 0.692). On average, resting M<sub>max</sub> in CH displayed a twofold increase compared to AH and N (P < 0.019); however, the change in M<sub>max</sub> during MVC was not statistically significant (P > 0.058). Neither measure of M<sub>max</sub> changed pre- to post-exercise in any condition (P  $\ge$  0.610). Pooled across conditions, pre-exercise ERT (mean  $r^2 = 0.95$ ) was 70% of the pre-exercise Q<sub>tw,pot</sub> and did not differ between conditions (mean ERT 75  $\pm 25$  N; P = 0.811). Post-exercise ERT was reduced in AH ( $52 \pm 27$  N, P = 0.049), but was unchanged in N and CH (P  $\ge 0.107$ ).

## Corticomotor responses

rMT in AH, N and CH was  $54 \pm 5$ ,  $53 \pm 3$  and  $51 \pm 6\%$  maximum stimulator output (P = 0.276), respectively. During CH, resting MEP amplitude was twofold greater compared to AH (P = 0.014) and N (P = 0.014). Exercise elicited a reduction in resting MEP amplitude in CH (P = 0.022), but not AH (P = 0.346) or N (P = 0.369). MEPs evoked during brief knee extensor contractions at 100, 75 and 50% MVC pre-exercise were higher in CH compared to AH (P < 0.020) and N (P < 0.030) (see also Figure

4). MEPs evoked during the brief knee-extensor contractions (50-100% MVC) post-exercise were not significantly different from pre-exercise values in any condition. MEP amplitude, however, was higher post-exercise during CH compared to AH (50% MVC, P = 0.018; 75% MVC, P = 0.030) and N (50% MVC, P = 0.034). The MEP/M<sub>max</sub> ratio increased for within contraction responses during CH (vs. AH 50 and 75% MVC; P  $\leq$  0.014 and N 50% MVC; P = 0.019) (Table 2). The CSP did not differ between conditions pre-exercise (pooled average, 186 ± 47 ms; P = 0.880) or post-exercise (pooled average, 185 ± 50 ms; P = 0.760). Baseline cortical voluntary activation did not differ between conditions (AH, 93 ± 5%; N, 97 ± 3%; CH, 93 ± 6%; P = 0.310) (Figure 5). Cortical voluntary activation was reduced post-exercise in AH ( $\Delta$ 11%, P = 0.014), but not in N ( $\Delta$ 4%, P = 0.298) or CH ( $\Delta$ 6%, P = 0.174); the decrease in AH was greater compared to N (P = 0.022) (Figure 5).

## Discussion

The aim of the present study was to assess corticospinal excitability and supraspinal fatigue after locomotor exercise in chronic hypoxia. The main finding was that exercise-induced supraspinal fatigue, as quantified via changes in cortical voluntary activation, was attenuated after two weeks of acclimatisation to high altitude whereas it was exacerbated in AH vs. N. Importantly, the diminished level of central fatigue in CH occurred in parallel with improvements in cerebral haemodynamics and arterial oxygenation (increased  $C_aO_2$  and  $S_pO_2$ ) brought about by the two weeks at altitude. Moreover, the attenuated development of central fatigue occurred in line with a substantial increase in corticospinal excitability. This latter finding suggests that a period of acclimatisation to altitude reduces the level of exercise-induced central fatigue and that this is attributable, at least in part, to an increased overall excitability of the brain to muscle pathway.

## **Supraspinal Fatigue**

A key aim of the present study was to determine the effect of acclimatisation on the development of central fatigue assessed after exercise. We hypothesised that improvements in cerebral oxygenation known to occur after a prolonged stay at altitude would bring about positive modifications on the development of central fatigue. We show that the development of supraspinal fatigue during locomotor exercise is recovered after 2 weeks at high altitude and similar to that observed in normoxia. Thus, the adaptive processes that take place during acclimatisation to high altitude seemingly protect healthy humans against the development of supraspinal fatigue.

## **Corticomotor responses**

The present study found no change in corticospinal excitability ( $\Delta$  resting MEP) in AH, a finding which is in line with literature utilising varying severities of hypoxia ( $F_1O_2 = 0.14 - 0.10$ ; resting  $S_pO_2 = 93 -$ 74%) for as little as 10 min to 1 h (Goodall *et al.*, 2010, Rupp *et al.*, 2012, Millet *et al.*, 2012). However, Szubski *et al.* (2006) reported increased corticospinal excitability, expressed as a reduced rMT (not  $\Delta$ MEP), after ~30 min of breathing hypoxic air ( $F_1O_2 = 0.12$ ; resting  $S_pO_2 = 75\%$ ). Moreover, the present study found a twofold increase in corticospinal excitability after 14 d acclimatisation to severe altitude (5,260 m, equivalent to  $F_1O_2$  0.105; resting  $S_pO_2 = 91 \pm 2\%$ ) with accompanying increases in the MEP/M<sub>max</sub> ratio, suggesting that the increases in MEP size were due to adaptive mechanisms within spinal and/or supraspinal sites. Similarly, Rupp *et al.* (2012) found a 26% increase in corticospinal excitability ( $\Delta$ MEP amplitude) after 3 h of exposure to normobaric hypoxia ( $F_1O_2 = 0.12$ ; resting  $S_pO_2 = 86\%$ ), demonstrating a time-dependent, hypoxia-induced modification in the brain-to-muscle pathway. Thus, a prolonged stay at altitude modifies the integrity of the corticospinal pathway which may contribute to reduce the level of central fatigue; however, a duration-dependent adaptation cannot yet be established with certainty.

TMS over the motor cortex preferentially activates corticospinal neurons trans-synaptically through excitatory interneurons and corticocortical axons (Di Lazzaro et al., 1998). The response to TMS critically depends on membrane excitability of motor cortical neurons and ion-channel function (Boroojerdi et al., 2001, Rothwell et al., 1991). In vitro investigations using isolated cerebral neurons from rats demonstrate that ion-channel function is affected by O<sub>2</sub> availability and that neuronal hyper-excitability is the consequence of chronic hypoxia (Donnelly et al., 1992). A heightened neural response is necessary to maintain membrane integrity and ionic homeostasis that occur from a period of insufficient metabolic activity (Nieber et al., 1999). Thus, the twofold increase in MEP observed in the present study might be due to facilitated cortical neurons acting to restore the loss of neuronal activity associated with a prolonged exposure to altitude. Additionally, an increased level of muscle sympathetic nerve activity (peroneal microneurography) has been reported during a prolonged stay at the same altitude as in the present study (Hansen and Sander, 2003). That study showed a significant increase in muscle sympathetic nerve activity just 3 days after exposure to high altitude, suggesting that the prolonged stay induced a striking and long-lasting sympathetic overactivity. More recently, Buharin et al. (2013) found that a transient increase in sympathetic nerve activity (induced via lower body negative pressure) enhances corticospinal excitability as identified using TMS. The mechanism responsible for the increase in corticospinal excitability was postulated

to be due to an elevated concentration of noradrenaline, a monoamine that is known to increase exponentially during sustained periods at altitudes exceeding 4,000 m (Cunningham *et al.*, 1965, Mazzeo *et al.*, 1994). Thus, the increased corticospinal excitability observed following 2 weeks of acclimatisation in the present study might be attributable, at least in part, to a heightened sympathetic nerve activity and associated increases in corticospinal excitability as well as hyper-excitable cerebral neurons. The increased corticospinal excitability in this investigation occurred in line with no symptoms of mountain sickness, a finding that opposes that of Miscio *et al.* (2009). Miscio *et al.* (2009) found that exposure to high altitude changes cortical excitability by affecting both inhibitory and excitatory circuits and that this is reflected in acute mountain sickness symptoms. This conclusion was based on a group of participants who resided at 4,554 m for only 3-5 days, a time frame in which acute mountain sickness is said to be most prominent (Hackett and Roach, 2001) and much shorter than the present study.

Despite substantial differences in end-exercise peripheral fatigue, CSP duration immediately after exercise (i.e., pre-to post-exercise change) was similar in all conditions. This suggests that locomotor exercise in N, AH and CH does not influence intracortical inhibition. These findings are in agreement with investigations using locomotor exercise in N and AH (Goodall *et al.*, 2012, Sidhu *et al.*, 2009b). However, Oliviero *et al.* (2002) reported decreased intracortical inhibition and CSP duration in chronic hypoxemic patients with COPD. These changes, mediated by cerebral GABA receptors, were reversed after 3-4 months of O<sub>2</sub> therapy, demonstrating that the changes were O<sub>2</sub> sensitive. However, factors other than chronic hypoxaemia might influence intracortical inhibition in patients with COPD making it difficult to quantify the influence that chronic hypoxaemia has on cortical inhibition.

On balance, we judge the increased corticospinal excitability in CH noted in the present study to be the result of adaptations in ion-channel function and elevations in circulating catecholamines serving to facilitate neurotransmission rather than mechanisms related to intracortical inhibition (Buharin *et al.*, 2013, Nieber *et al.*, 1999, Palange, 1998).

## Hematological and cerebrovascular responses

Upon initial exposure to high altitude, acute hypoxia dilates cerebral arterioles thereby overriding the vasoconstrictive effect of hyperventilation-associated hypocapnia (Iwasaki *et al.*, 2011). During a prolonged stay at altitude, hypocapnia further develops and arterial hypoxaemia is ameliorated, as

reflected by increases in arterial [Hb],  $PO_2$  and  $O_2$  saturation (Figure 3). Furthermore, the increase in P<sub>a</sub>O<sub>2</sub> and further decrease in P<sub>a</sub>CO<sub>2</sub> with acclimatisation causes relative vasoconstriction reducing CBF down to SL values (Subudhi et al. 2013). We estimated an index of cerebral O<sub>2</sub> delivery using the product of MCA<sub>v</sub> and C<sub>a</sub>O<sub>2</sub>. Our data demonstrate a reduced cerebral O<sub>2</sub> delivery index during exercise in AH compared to N; however, an improved cerebral O2 delivery index was evident after two weeks of acclimatisation (Figure 3). The data in AH support a relationship between cerebral  $O_2$ delivery and supraspinal fatigue (Goodall et al., 2012). The calculation of C<sub>a</sub>O<sub>2</sub> during exercise from resting [Hb] should be interpreted with caution as a hemoconcentration could have impacted this measure. At sea level, the hemoconcentration accompanying maximal exercise for approximately 10 min is counterbalanced by the concomitant exercise-induced arterial hypoxemia with the net effect of similar C<sub>a</sub>O<sub>2</sub> at rest and during exercise (Amman et al., 2006a). At altitude, despite significant hemoconcentration, C<sub>a</sub>O<sub>2</sub> actually falls from rest to submaximal/maximal exercise by 10-25% (Calbet et al., 2003). This would suggest that exercise C<sub>a</sub>O<sub>2</sub> calculations, based on a resting C<sub>a</sub>O<sub>2</sub> measure, might actually overestimate  $C_aO_2$  measured during exercise at altitude. Furthermore, we assumed that MCA diameter would remain constant in hypoxia (Poulin and Robbins, 1996, Serrador et al., 2000). While there is evidence of MCA dilatation at rest in hypoxia (Willie et al., 2012, Wilson et al., 2011), there is currently no evidence of MCA dilatation during intense exercise accompanied with substantial exercise-induced hyperventilation and associated hypocapnia. We acknowledge, however, that our measurements of blood velocity (rather than flow) must be interpreted with caution.

We found acclimatisation-induced increases in  $O_2$  saturation and content (Figure 3). Furthermore, arterial  $O_2$  tension increased from AH to CH (~39 mmHg to ~59 mmHg). Subudhi *et al.* (2013) has shown resting cerebral  $O_2$  delivery to be maintained at levels observed in N during AH and CH, although it is presumed that the delivery of  $O_2$  to the mitochondria within the parenchyma will be reduced because the driving gradient for diffusion from capillary to tissue is the PO<sub>2</sub> difference between capillary and tissue (Xu and Lamanna, 2006). The tissue PO<sub>2</sub> would be close to zero; thus, the driving force is essentially the  $P_aO_2$ . In the present study the  $P_aO_2$  increased in line with acclimatisation, thereby improving the gradient for diffusion and perhaps restoring brain tissue  $O_2$ tension to pre-hypoxic levels (Dunn *et al.*, 2000). Thus, we postulate that the lack of central fatigue in chronic hypoxia may be related to increases in brain tissue  $O_2$  tension. However, the link between increases in  $P_aO_2$  and  $C_aO_2$  and the reduction in central fatigue that occurs after a period of acclimatisation warrants further investigation.

## **Technical Considerations**

Exercising in a hypobaric environment was not feasible for the trials in AH. Thus, the two modes of hypoxia (normobaric [AH] vs. hypobaric [CH]) differed. The literature concerning the responses in normobaric and hypobaric hypoxia is equivocal and readers are directed elsewhere to a point:counterpoint debate (Girard *et al.*, 2012). Briefly, it was proposed that evidence is growing, suggestive that hypobaric hypoxia affects responses (ventilation, fluid balance, acute mountain sickness and performance) to a greater extent than normobaric hypoxia (Girard *et al.*, 2012). However, this argument was opposed by the fact that in terms of O<sub>2</sub> sensing, hypobaric hypoxia does not induce different responses compared to normobaric hypoxia (Mounier and Brugniaux, 2012). Moreover, it is unknown how any such differences which might exist between hypobaric and normobaric hypoxia may affect indices of exercise-induced fatigue. We set the  $F_1O_2$  (0.105) at sea level to obtain the same  $P_1O_2$  (~74 mmHg) that was expected at the subsequent altitude in Bolivia (5,260 m).

In line with other investigations that have measured exercise-induced fatigue of the knee extensors (Goodall *et al.*, 2012, Goodall *et al.*, 2010, Sidhu *et al.*, 2009b, Rossman *et al.*, 2013), measurements were made within 2.5 min after exercise termination. Corticospinal excitability associated with maximal single muscle contractions recovers within 1 min post-exercise (Taylor *et al.*, 1999). Thus, the present experimental design, utilising whole body exercise, might not have captured all elements of central fatigue. However, the methods and time to assess fatigue after exercise in all three conditions were identical and even though our measurements were made more than 1 min post-exercise, significant differences were observed, testifying to the strength of our data.

## Conclusion

The novel finding was that supraspinal fatigue, present after exercise in acute hypoxia, was attenuated after a period of acclimatisation to high altitude. Importantly, the reduced development of central fatigue in chronic hypoxia occurred in parallel with an increase in the excitability of the brain to muscle pathway consequent to an increased cerebral O<sub>2</sub> delivery. The attenuated rate of development of central fatigue in chronic hypoxia might explain, at least in part, the improvements in locomotor exercise performance that are commonly observed after acclimatisation to high altitude.

## **Author Contributions**

SG, RT, and MA contributed to conception and design of the experiments, data collection, data analysis, data interpretation, manuscript drafting and editorial process. ER contributed to conception and design of the experiments, data interpretation and manuscript revision. AL contributed to data collection. LR contributed to conception and design of the experiments, data interpretation, manuscript drafting and revision. AL, AS and RR conceived, designed and executed the AltitudeOmics study of which the present study was a part, and contributed to manuscript revision. AS also contributed to data collection and data interpretation. All authors approved the final version of the manuscript.

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## **Conflict of Interest**

Nothing to declare

## References

- Ainslie, P. N. & Ogoh, S. 2009. Regulation of cerebral blood flow during chronic hypoxia: a matter of balance. *Exp Physiol* 95, 251-262.
- Amann, M. & Calbet, J. A. 2008. Convective oxygen transport and fatigue. J Appl Physiol 104, 861-70.
- Amann, M., Eldridge, M. W., Lovering, A. T., Stickland, M. K., Pegelow, D. F. & Dempsey, J. A. 2006a. Arterial oxygenation influences central motor output and exercise performance via effects on peripheral locomotor muscle fatigue in humans. *J Physiol* 575, 937-952.
- Amann, M., Goodall, S., Twomey, R., Subudhi, A. W., Lovering, A. T. & Roach, R. C. 2013. AltitudeOmics: on the consequences of high-altitude acclimatization for the development of fatigue during locomotor exercise in humans. J Appl Physiol 115, 634-42.
- Amann, M., Romer, L. M., Pegelow, D. F., Jacques, A. J., Hess, C. J. & Dempsey, J. A. 2006b. Effects of arterial oxygen content on peripheral locomotor muscle fatigue. J Appl Physiol 101, 119-127.
- Amann, M., Romer, L. M., Subudhi, A. W., Pegelow, D. F. & Dempsey, J. A. 2007. Severity of arterial hypoxaemia affects the relative contributions of peripheral muscle fatigue to exercise performance in healthy humans. J Physiol 581, 389-403.
- Borg, G. A. 1982. Psychophysical bases of perceived exertion. Med Sci Sports Exerc 14, 377-81.
- Boroojerdi, B., Battaglia, F., Muellbacher, W. & Cohen, L. G. 2001. Mechanisms influencing stimulusresponse properties of the human corticospinal system. *Clin Neurophysiol* 112, 931-7.
- Buharin, V. E., Butler, A. J., Rajendra, J. K. & Shinohara, M. 2013. Enhanced corticospinal excitability with physiologically heightened sympathetic nerve activity. *J Appl Physiol* 114, 429-35.
- Calbet J.A., Boushel, R., Radegran, G., Sondergaard, H., Wagner, P.D. & Saltin, B. 2003. Why is VO<sub>2</sub> max after altitude acclimatization still reduced despite normalization of arterial O<sub>2</sub> content? *Am J Phys: Reg Integr Comp Phys* 284, R304-316.
- Chen, R., Lozano, A. M. & Ashby, P. 1999. Mechanism of the silent period following transcranial magnetic stimulation. Evidence from epidural recordings. *Exp Brain Res* 128, 539-42.
- Cunningham, W. L., Becker, E. J. & Kreuzer, F. 1965. Catecholamines in plasma and urine at high altitude. J Appl Physiol 20, 607-10.
- Di Lazzaro, V., Restuccia, D., Oliviero, A., Profice, P., Ferrara, L., Insola, A., Mazzone, P., Tonali, P. & Rothwell, J. C. 1998. Effects of voluntary contraction on descending volleys evoked by transcranial stimulation in conscious humans. *J Physiol* 508, 625-33.
- Donnelly, D. F., Jiang, C. & Haddad, G. G. 1992. Comparative responses of brain stem and hippocampal neurons to O2 deprivation: in vitro intracellular studies. *Am J Physiol* 262, L549-54.

- Dunn, J. F., Grinberg, O., Roche, M., Nwaigwe, C. I., Hou, H. G. & Swartz, H. M. 2000. Noninvasive assessment of cerebral oxygenation during acclimation to hypobaric hypoxia. J Cereb Blood Flow Metab 20, 1632-5.
- Gandevia, S. C., Allen, G. M., Butler, J. E. & Taylor, J. L. 1996. Supraspinal factors in human muscle fatigue: evidence for suboptimal output from the motor cortex. *J Physiol* 490, 529-536.
- Girard, O., Koehle, M. S., MacInnis, M. J., Guenette, J. A., Verges, S., Rupp, T., Jubeau, M., Perrey, S., Millet, G. Y. & Chapman, R. F. 2012. Comments on Point: Counterpoint: Hypobaric hypoxia induces/does not induce different responses from normobaric hypoxia. J Appl Physiol 112, 1788-1794.
- Goodall, S., Gonzalez-Alonso, J., Ali, L., Ross, E. Z. & Romer, L. M. 2012. Supraspinal fatigue after normoxic and hypoxic exercise in humans. *J Physiol* 590, 2767-82.
- Goodall, S., Romer, L. M. & Ross, E. Z. 2009. Voluntary activation of human knee extensors measured using transcranial magnetic stimulation. *Exp Physiol* 94, 995-1004.
- Goodall, S., Ross, E. Z. & Romer, L. M. 2010. Effect of graded hypoxia on supraspinal contributions to fatigue with unilateral knee-extensor contractions. *J Appl Physiol* 109, 1842-1851.
- Groppa, S., Oliviero, A., Eisen, A., Quartarone, A., Cohen, L. G., Mall, V., Kaelin-Lang, A., Mima, T., Rossi, S., Thickbroom, G. W., Rossini, P. M., Ziemann, U., Valls-Sole, J. & Siebner, H. R. 2012.
  A practical guide to diagnostic transcranial magnetic stimulation: report of an IFCN committee. *Clin Neurophysiol* 123, 858-82.
- Hackett, P. H. & Roach, R. C. 2001. High-altitude illness. N Engl J Med 345, 107-14.
- Hansen, J. & Sander, M. 2003. Sympathetic neural overactivity in healthy humans after prolonged exposure to hypobaric hypoxia. *J Physiol* 546, 921-9.
- Inghilleri, M., Berardelli, A., Cruccu, G. & Manfredi, M. 1993. Silent period evoked by transcranial stimulation of the human cortex and cervicomedullary junction. *J Physiol* 466, 521-534.
- Iwasaki, K., Zhang, R., Zuckerman, J. H., Ogawa, Y., Hansen, L. H. & Levine, B. D. 2011. Impaired dynamic cerebral autoregulation at extreme high altitude even after acclimatization. J Cereb Blood Flow Metab 31, 283-92.
- Lucas, S. J., Burgess, K. R., Thomas, K. N., Donnelly, J., Peebles, K. C., Lucas, R. A., Fan, J. L., Cotter, J. D., Basnyat, R. & Ainslie, P. N. 2011. Alterations in cerebral blood flow and cerebrovascular reactivity during 14 days at 5050 m. J Physiol 589, 741-53.
- Mazzeo, R. S., Wolfel, E. E., Butterfield, G. E. & Reeves, J. T. 1994. Sympathetic response during 21 days at high altitude (4,300 m) as determined by urinary and arterial catecholamines. *Metabolism* 43, 1226-32.
- Millet, G. Y., Muthalib, M., Jubeau, M., Laursen, P. B. & Nosaka, K. 2012. Severe hypoxia affects exercise performance independently of afferent feedback and peripheral fatigue. *J Appl Physiol* 112, 1335-44.
- Miscio, G., Milano, E., Aguilar, J., Savia, G., Foffani, G., Mauro, A., Mordillo-Mateos, L., Romero-Ganuza, J. & Oliviero, A. 2009. Functional involvement of central nervous system at high altitude. *Exp Brain Res* 194, 157-62.

- Mounier, R. & Brugniaux, J. V. 2012. Last Word on Counterpoint: Hypobaric hypoxia does not induce different physiological responses from normobaric hypoxia. *J Appl Physiol* 112, 1796-1796.
- Nieber, K., Eschke, D. & Brand, A. 1999. Brain hypoxia: effects of ATP and adenosine. *Prog Brain Res* 120, 287-97.
- Nybo, L. & Rasmussen, P. 2007. Inadequate cerebral oxygen delivery and central fatigue during strenuous exercise. *Ex Sport Sci Rev* 35, 110-118.
- Oliviero, A., Corbo, G., Tonali, P. A., Pilato, F., Saturno, E., Dileone, M., Versace, V., Valente, S. & Di Lazzaro, V. 2002. Functional involvement of central nervous system in acute exacerbation of chronic obstructive pulmonary disease A preliminary transcranial magnetic stimulation study. J Neurol 249, 1232-6.
- Palange, P. 1998. Renal and hormonal abnormalities in chronic obstructive pulmonary disease (COPD). *Thorax* 53, 989-91.
- Perrey, S. & Rupp, T. 2009. Altitude-induced changes in muscle contractile properties. *High Alt Med Biol* 10, 175-82.
- Poulin, M. J. & Robbins, P. A. 1996. Indexes of flow and cross-sectional area of the middle cerebral artery using doppler ultrasound during hypoxia and hypercapnia in humans. *Stroke* 27, 2244-50.
- Roach, R. C., Bartsch, P., Olez, O. & Hackett, P. H. 1993. The Lake Louise acute mountain sickness scoring system. In: SUTTON, J. R., HOUSTON, C. S. & COATES, G. (Eds.) *Hypoxia and Mountain Medicine*. Burlinton, VT, Queen City Printers Inc.
- Romer, L. M., Haverkamp, H. C., Amann, M., Lovering, A. T., Pegelow, D. F. & Dempsey, J. A. 2007. Effect of acute severe hypoxia on peripheral fatigue and endurance capacity in healthy humans. *Am J Physiol. Reg, Int Com Physiol* 292, R598-606.
- Rossman, M. J., Venturelli, M., McDaniel, J., Amann, M. & Richardson, R. S. 2012. Muscle mass and peripheral fatigue: a potential role for afferent feedback? *Acta Physiol (Oxf)* 206, 242-50.
- Rothwell, J. C., Thompson, P. D., Day, B. L., Boyd, S. & Marsden, C. D. 1991. Stimulation of the human motor cortex through the scalp. *Exp Physiol* 76, 159-200.
- Rupp, T., Jubeau, M., Wuyam, B., Perrey, S., Levy, P., Millet, G. Y. & Verges, S. 2012. Time-dependent effect of acute hypoxia on corticospinal excitability in healthy humans. *J Neurophysiol* 108, 1270-7.
- Serrador, J. M., Picot, P. A., Rutt, B. K., Shoemaker, J. K. & Bondar, R. L. 2000. MRI measures of middle cerebral artery diameter in conscious humans during simulated orthostasis. *Stroke* 31, 1672-8.
- Sidhu, S. K., Bentley, D. J. & Carroll, T. J. 2009a. Cortical voluntary activation of the human knee extensors can be reliably estimated using transcranial magnetic stimulation. *Muscle Nerve* 39, 186-96.
- Sidhu, S. K., Bentley, D. J. & Carroll, T. J. 2009b. Locomotor exercise induces long-lasting impairments in the capacity of the human motor cortex to voluntarily activate knee extensor muscles. *J Appl Physiol* 106, 556-65.
- Subudhi, A. W., Fan, J. L., Evero, O., Bourdillon, N., Kayser, B., Julian, C. G., Lovering, A.T. & Roach, R. 2013. AltitudeOmics: Effect of ascent and acclimatization to 5260 m on regional cerebral oxygen delivery. *Exp Physiol*, DOI: 10.1113/expphysiol.2013.075184.
- Subudhi, A. W., Miramon, B. R., Granger, M. E. & Roach, R. C. 2009. Frontal and motor cortex oxygenation during maximal exercise in normoxia and hypoxia. *J Appl Physiol* 106, 1153-8.
- Subudhi, A. W., Olin, J. T., Dimmen, A. C., Polaner, D. M., Kayser, B. & Roach, R. C. 2011. Does cerebral oxygen delivery limit incremental exercise performance? J Appl Physiol 111, 1727-34.
- Szubski, C., Burtscher, M. & Loscher, W. N. 2006. The effects of short-term hypoxia on motor cortex excitability and neuromuscular activation. *J Appl Physiol* 101, 1673-1677.
- Taylor, J. L., Butler, J. E. & Gandevia, S. C. 1999. Altered responses of human elbow flexors to peripheral-nerve and cortical stimulation during a sustained maximal voluntary contraction. *Exp Brain Res* 127, 108-15.
- Taylor, J. L. & Gandevia, S. C. 2001. Transcranial magnetic stimulation and human muscle fatigue. *Muscle & nerve* 24, 18-29.
- Todd, G., Taylor, J. L. & Gandevia, S. C. 2003. Measurement of voluntary activation of fresh and fatigued human muscles using transcranial magnetic stimulation. *J Physiol* 551, 661-671.
- Willie, C. K., Macleod, D. B., Shaw, A. D., Smith, K. J., Tzeng, Y. C., Eves, N. D., Ikeda, K., Graham, J., Lewis, N. C., Day, T. A. & Ainslie, P. N. 2012. Regional brain blood flow in man during acute changes in arterial blood gases. *J Physiol* 590, 3261-75.
- Wilson, M. H., Edsell, M. E., Davagnanam, I., Hirani, S. P., Martin, D. S., Levett, D. Z., Thornton, J. S., Golay, X., Strycharczuk, L., Newman, S. P., Montgomery, H. E., Grocott, M. P., Imray, C. H. & Caudwell Xtreme Everest Research, G. 2011. Cerebral artery dilatation maintains cerebral oxygenation at extreme altitude and in acute hypoxia--an ultrasound and MRI study. J Cereb Blood Flow Metab 31, 2019-29.
- Xu, K. & Lamanna, J. C. 2006. Chronic hypoxia and the cerebral circulation. *J Appl Physiol* 100, 725-30.

### **Figure Legends**

**Figure 1.** Mean area of motor evoked potentials (MEP) recorded from the vastus lateralis (VL) in response to stimulation over the motor cortex during varying contraction intensities pre- ( $\circ$ ) and post-exercise ( $\bullet$ ) (mean for all conditions). The TMS responses were compared to the area of the maximal M-wave ( $M_{max}$ ) evoked by peripheral stimulation of the femoral nerve. Data are means  $\pm$  SE for 7 participants.

**Figure 2.** Cerebral oxygenation at resting baseline, during the final 30 s of a 3 min warm up (28 W) and during the final 30 s of constant-load exercise (131 W) in normoxia (N; panel a), acute hypoxia (AH; panel b) and chronic hypoxia (CH; panel c). Data are means  $\pm$  SE for 7 participants.  $\pm$  P < 0.05 vs. respective baseline;  $\pm$  P < 0.05 vs. respective warm up; \* P < 0.05 vs. AH; # P < 0.05 vs. CH.

Resting baseline in AH denotes the value after 10 min wash in of the hypoxic gas.  $O_2Hb$ , oxygenated haemoglobin; HHb, deoxygenated haemoglobin and THb, total haemoglobin.

**Figure 3.** Arterial oxygen saturation ( $S_pO_2$ ) (a), cerebral blood flow velocity (MCA<sub>v</sub>) (b) and middle cerebral artery  $O_2$  delivery index (MCA<sub>v</sub> ×  $C_aO_2$ ) during constant-load exercise (131 W) in normoxia (N), acute hypoxia (AH), and chronic hypoxia (CH). Values are plotted for the duration of the shortest trial (8 min) and extrapolated to the group mean exercise time (10.1 min). Data are means  $\pm$  SE for 7 participants.  $\pm P < 0.05$  vs. rest;  $\pm P < 0.05$  vs. N;  $\pm P < 0.05$  vs. CH.

**Figure 4.** Representative MEPs evoked during knee extensor contractions at 50% MVC before exercise in each condition. Traces are shown from a representative participant in each condition; 8 stimuli were delivered from which an average value was obtained. Note the increase in MEP amplitude (corticospinal excitability) after acclimatisation.

**Figure 5.** Cortical voluntary activation measured before (open bars) and immediately after (<2.5 min; closed bars) constant-load exercise (131 W) in normoxia (N), acute hypoxia (AH), and chronic hypoxia (CH). \* P < 0.05 pre- vs. post-exercise.

·		Normoxia	Acute Hypoxia	Chronic Hypoxia
HR (beats min <sup>-1</sup> )	Rest	81 ± 7†	90 ± 9	104 ± 16
	Final Min	150 ± 16*	173 ± 14	167 ± 16
$\dot{V}_{\rm E}$ (I min <sup>-1</sup> )	Rest	$14.3 \pm 2.4$	20.0 ± 2.6	24.5 ± 5.4
	Final Min	$60.0 \pm 9.6^{*+}$	108.8 ± 24.7†	128.5 ± 30.0
$f_{\rm R}$ (breaths min <sup>-1</sup> )	Rest	15.6 ± 3.6	17.5 ± 4.5	13.0 ± 3.4
	Final Min	31.4 ± 4.9*†	51.6 ± 8.7†	54.8. ± 9.9
V <sub>T</sub> (I)	Rest	$1.07 \pm 0.37$	$1.30 \pm 0.34$	$1.47 \pm 0.63$
	Final Min	$2.00 \pm 0.45$	2.07 \pm 0.43	$2.41 \pm 0.58$
VO <sub>2</sub> (I min <sup>-1</sup> )	Rest Final Min	$\begin{array}{rrrr} 0.49 & \pm & 0.10 \\ 2.45 & \pm & 0.51 \end{array}$	$0.45 \pm 0.08$ 2.34 ± 0.58	$\begin{array}{rrrr} 0.45 & \pm & 0.12 \\ 2.07 & \pm & 0.50 \end{array}$
$\dot{V}$ CO <sub>2</sub> (I min <sup>-1</sup> )	Rest	$0.44 \pm 0.09$	$0.55 \pm 0.09$	$0.39 \pm 0.08$
	Final Min	$2.32 \pm 0.51$	2.69 $\pm 0.62^+$	$1.94 \pm 0.50$
$\dot{V}_{\rm E}$ / $\dot{V}_{\rm O_2}$	Rest	30.7 ± 2.7*†	47.4 ± 6.5†	$55.9 \pm 14.9$
	Final Min	25.2 ± 2.4*†	51.2 ± 15.0†	$62.9 \pm 9.2$
$\dot{V}_{\rm E}$ / $\dot{V}_{\rm CO_2}$	Rest	33.9 ± 2.7†	$37.9 \pm 6.5^+$	$63.4 \pm 6.8$
	Final Min	26.2 ± 2.6*†	$41.7 \pm 6.9^+$	$67.1 \pm 9.1$
RPE, dyspnoea	Rest	$7.0 \pm 0.0$	$7.3 \pm 0.5$	$7.1 \pm 0.4$
	Final Min	11.4 ± 2.4*†	19.4 ± 0.8	19.1 ± 0.7
RPE, limb	Rest	$7.1 \pm 0.4$	$7.1 \pm 0.4$	7.0 ± 0.0
	Final Min	12.3 ± 3.3*	19.9 ± 0.4	17.6 ± 1.7

**Table 1.** Cardiorespiratory and perceptual responses at rest and during the final minute of constant-load cycling (131 W) in normoxia, acute hypoxia and chronic hypoxia.

Values are means ± SD for 7 participants. Resting values were measured during the 5<sup>th</sup> minute of breathing the test gas mixture. HR, heart rate;  $V_E$ , minute ventilation;  $f_R$ , respiratory frequency;  $V_T$ , tidal volume;  $VO_2$ , oxygen uptake;  $VCO_2$ , carbon dioxide output; RPE, ratings of perceived exertion. \* P < 0.05 vs. acute hypoxia; † P < 0.05 vs. chronic hypoxia.

		Norm	oxia	Acute	Hy	poxia	Chronic H	łypoxia
Rest								
	Pre	6.9 ±	2.0†	8.6	±	3.7†	14.9 ±	8.3
M <sub>max</sub> amplitude (mV)	Post	6.7 ±	1.7	9.0	±	4.1	14.0 ±	8.2
	Pre	0.19 ±	0.12†	0.19	±	0.11†	0.41 ±	0.28
MEP amplitude (mV)	Post	0.11 ±	0.06	0.11	±	0.10	0.21 ±	0.18 <sup>#</sup>
	Pre	2.6 ±	1.3	2.7	±	1.9	4.1 ±	4.2
IVIEP/IVI <sub>max</sub> (%)	Post	1.8 ±	1.2	1.5	±	1.3	2.6 ±	3.4
Within contraction								
	Pre	8.9 ±	1.7	9.9	±	3.2	13.0 ±	6.1
M <sub>max</sub> amplitude 100% (mv)	Post	9.0 ±	1.9	10.0	±	3.3	11.9 ±	5.4
	Pre	3.8 ±	1.5	3.1	±	1.0†	7.1 ±	4.7
WEP amplitude 100% (mv)	Post	4.0 ±	2.7	3.2	±	1.0	6.5 ±	4.4
MED amplitude 75% (m)()	Pre	3.9 ±	1.5†	2.9	±	1.4†	7.6 ±	4.9
WEP amplitude 75% (IIIV)	Post	4.3 ±	2.6	3.3	±	1.2†	6.9 ±	3.9
MEP amplitude 50% (mV)	Pre	2.54 ±	0.87†	2.16	±	0.52†	6.5 ±	4.8
	Post	2.99 ±	2.01†	2.56	±	0.95†	6.4 ±	4.5
MEP/M <sub>max</sub> (%) 100% MVC	Pre	35 ±	17	33	±	14	52 ±	17
····-· / ······························	Post	39 ±	20	37	±	15	52 ±	19
MEP/M <sub>max</sub> (%) 75% MVC	Pre	40 ±	15	34	±	19†	58 ±	18
	Post	42 ±	17	38	±	18†	57 ±	13
MEP/M (%) 50% MVC	Pre	28 ±	14†	26	±	10†	50 ±	21
	Post	30 ±	15†	31	±	17†	54 ±	23
(CSP (ms)	Pre	198 ±	58	174	±	46	186 ±	36
CSP (IIIS)	Post	188 +	64	171	+	25	106 +	51

**Table 2.** Peripheral and central measures of excitability assessed before and after constant-load cycling (131 W) in normoxia, acute hypoxia and chronic hypoxia.

Values are means ± SD for 7 participants.  $M_{max}$ , maximal motor response; MEP, motor evoked potential; CSP, cortical silent period. + P < 0.05 vs. chronic hypoxia; <sup>#</sup> P < 0.05 vs. Pre.















# AltitudeOmics: Effect of ascent and acclimatization to 5260 m on regional cerebral oxygen delivery

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### What is the central question of this study?

Hypoxia associated with ascent to high altitude may threaten cerebral oxygen delivery. We sought to determine if there are regional changes in the distribution of cerebral blood flow that might favor oxygen delivery to areas associated with basic homeostatic functions to promote survival in this extreme environment.

## What is the main finding and its importance?

We show evidence of a "brain sparing" effect during acute exposure to high altitude, in which there is a slight increase in relative oxygen delivery to the posterior cerebral circulation. This may serve to support basic regulatory functions associated with the brain stem and hypothalamus.

# Abstract

Accepted Articl

Cerebral hypoxemia associated with rapid ascent to high altitude can be life threatening; yet, with proper acclimatization, cerebral function can be maintained well enough for humans to thrive. METHODS: We investigated adjustments in global and regional cerebral oxygen delivery  $(DO_2)$  as 21 healthy volunteers rapidly ascended and acclimatized to 5260m. Ultrasound indices of cerebral blood flow (CBF) in internal carotid and vertebral arteries were measured at sea level (SL), upon arrival at 5260m (ALT1; Pbar = 409mmHg), and after 16 days of acclimatization (ALT16). Cerebral DO<sub>2</sub> was calculated as the product of arterial oxygen content ( $CaO_2$ ) and flow in each respective artery and summed to estimate global CBF. Vascular resistances were calculated as the quotient of mean arterial pressure and respective flows. RESULTS: Global CBF increased ~70% upon arrival at ALT1 (P<0.001) and returned to SL values at ALT16 as a result of changes in cerebral vascular resistance. A reciprocal pattern in CaO<sub>2</sub> maintained global cerebral  $DO_2$  across acclimatization, although  $DO_2$  to the posterior cerebral circulation was increased by  $\sim 25\%$  at ALT1 (P=0.032). CONCLUSIONS: Cerebral DO<sub>2</sub> is well maintained upon acute exposure and acclimatization to hypoxia, particularly in the posterior and inferior regions of the brain associated with vital homeostatic functions. This tight regulation of cerebral DO<sub>2</sub> was achieved through integrated adjustments in local vascular resistances to alter cerebral perfusion during both acute and chronic exposure to hypoxia.

# Introduction

Although the brain represents only about 2% of body weight, it is a highly metabolic tissue that receives ~15% of cardiac output and accounts for ~20% of total body oxygen consumption at rest (Wade & Bishop, 1962). Maintenance of cerebral oxygen delivery (DO<sub>2</sub>) is essential for vital cerebral functions associated with homeostasis. In the face of severe hypoxemia, such as experienced during rapid ascent to extreme altitudes (> 8,000 m), reduction in cerebral DO<sub>2</sub> results in loss of consciousness within seconds (Luft *et al.*, 1951; Luft & Noell, 1956) and death within minutes (Bert, 1943). However, with staged acclimatization to progressively higher elevations, cerebral DO<sub>2</sub> can be maintained well enough for humans to reach the summit of Mount Everest (8,848 m) without supplemental oxygen. The mechanisms responsible for this remarkable plasticity in cerebral DO<sub>2</sub> are complex and not completely understood.

Cerebral DO<sub>2</sub> is the product of cerebral blood flow (CBF) and arterial oxygen content (CaO<sub>2</sub>). It is well established that CBF rises upon acute exposure to high altitude and returns to near sea-level values with acclimatization (Severinghaus *et al.*, 1966; Huang *et al.*, 1987; Jensen *et al.*, 1990), while CaO<sub>2</sub> decreases in acute hypoxia and returns to sea-level values with acclimatization. These opposing CBF and CaO<sub>2</sub> responses to altitude appear to offset one another and maintain cerebral DO<sub>2</sub> across acclimatization (Severinghaus *et al.*, 1966; Wolff *et al.*, 2002). The pattern of CBF

change in response to hypoxia has been attributed to the relative balance of hypoxic vasodilation and hypocapnic vasoconstriction in the brain (Xu & Lamanna, 2006; Brugniaux *et al.*, 2007). During acute, severe hypoxia, vasodilation typically exceeds vasoconstriction, resulting in greater CBF (Mardimae *et al.*, 2012; Willie *et al.*, 2012). With acclimatization, increased ventilatory drive reduces PaCO<sub>2</sub> and improves PaO<sub>2</sub>, tipping the balance in favor of vasoconstriction and restoring CBF to pre-exposure values. Changes in the PaO<sub>2</sub>/PaCO<sub>2</sub> ratio have been shown to account for ~40% of the variation in global CBF over acclimatization (Lucas *et al.*, 2011), with other biochemical (e.g. pH, HCO<sub>3</sub><sup>-</sup>, nitric oxide) and hematological (e.g. hemoglobin, hematocrit, blood viscosity) factors presumably accounting for the rest of the response (Todd *et al.*, 1994; Tomiyama *et al.*, 1999; Severinghaus, 2001) to maintain global cerebral DO<sub>2</sub>.

Recent data demonstrate that acute normobaric hypoxia (i.e. breathing hypoxic gas) affects the regional distribution of CBF within the brain. Data from positron emission tomography (PET) studies show greater perfusion of the brain stem, hypothalamus, thalamus and cerebellum during acute hypoxia, with (Binks *et al.*, 2008) or without (Buck *et al.*, 1998) controlled levels of PaCO<sub>2</sub>. Regional differences in cerebrovascular reactivity to O<sub>2</sub> and CO<sub>2</sub> have been postulated to control the distribution of CBF. Vascular Doppler studies of the major tributary vessels of the brain suggest that a greater percentage of blood flow may be directed towards the posterior cerebral circulation, including the brain stem, in response to controlled levels of hypoxia and hypocapnia (Sato *et al.*, 2012). From a teleological perspective,

this could help preserve vital homeostatic functions at the expense of higher cognitive processing; however, it is unclear whether regional distribution of CBF is similarly affected in hypobaric hypoxia (i.e. high altitude) or if it changes with acclimatization, as not all studies report significant regional differences (Huang *et al.*, 1987; Willie *et al.*, 2012; Willie *et al.*, 2013).

Despite the importance of O<sub>2</sub> supply for cerebral function, longitudinal studies of cerebral DO<sub>2</sub> at high altitude are sparse. In a secondary analysis of data from Severinghaus et al.'s original study of CBF at high altitude, global cerebral DO<sub>2</sub> in four subjects appeared stable and in excess of oxygen demand after 6-12 hours and 3-5 days of exposure to 3,810m (Severinghaus, 2001; Wolff et al., 2002). Using similar methodology (Kety-Schmidt technique), no differences were found in global cerebral DO<sub>2</sub> measured after 5 weeks at 5,260 m and return to sea level (Moller et al., 2002). Unfortunately, these two studies were based on a limited number of observations, which makes it difficult to detect small differences if they existed (type II error), and utilized methodology that can only measure global cerebral  $DO_2$ . A more recent MRI study with a larger sample size reported a tendency towards elevation of cerebral DO<sub>2</sub> after subjects returned from 2 days at 3,800 m (Smith et al., 2013), but no measurements of regional cerebral  $DO_2$  were made. Based the limited data to date, it is uncertain if global or regional cerebral DO<sub>2</sub> varies over time at high altitude.

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In this study we used vascular Doppler technology in conjunction with arterial blood sampling to allow us to quantify global and regional changes in CBF and cerebral DO<sub>2</sub> in the field as healthy people rapidly ascended and acclimatized to high altitude (5,260 m). We tested the hypothesis that upon acute exposure cerebral DO<sub>2</sub> would be maintained to regions of the brain associated with homeostasis at the expense of other tissues, but that these changes would normalize with acclimatization.

# **Methods**

### Subject recruitment and screening

This study was conducted as part of the AltitudeOmics project, for which a detailed description of the protocol is published elsewhere (Subudhi *et al.*, In Review). Briefly, following institutional ethics approval from the Universities of Colorado and Oregon and the US Department of Defense Human Research Protection Office, young, healthy sea-level residents were recruited from the greater Eugene, Oregon area (elevation 128 m). Potential subjects were screened to exclude anyone who was born or had lived at altitudes >1,500 m for more than one year or had traveled to altitudes > 1,000 m in the past 3 months. After obtaining written consent, physical exams and the Army Physical Fitness Test (push ups, sit ups and 3.2 km run) were performed to verify health and fitness status.

To evaluate effects of altitude acclimatization on cerebrovascular hemodynamics, subjects were studied on 3 occasions: 1) at sea level (SL, 130 m), 2) upon acute exposure to 5,260 m (ALT1), and 3) after 16 days of acclimatization (ALT16). Specifically, ~4 weeks following SL measurements in Eugene, Oregon, subjects were flown to La Paz, Bolivia. They spent two nights at low altitude (Coroico, Bolivia, 1,525 m) before being driven to the Chacaltaya Research Station at 5,260 m while breathing supplemental oxygen. Acute responses to high altitude were assessed 2 to 4 hours after arrival and cessation of supplemental oxygen (ALT1). Subjects acclimatized to altitudes ranging from 3,800 to 5,260 m over the next 15 days, with a majority of the time (75%) spent at 5,250 m. Measurements were repeated on ALT16.

### Instrumentation

Subjects were studied in an upright, seated position with feet on the floor. Arterial blood pressure (ABP) was monitored via a fluid filled pressure transducer (Utah Medical, Salt Lake City, UT, USA) positioned at heart level and attached to a 22-gauge catheter in a radial artery. Blood flow velocity in the left middle cerebral artery was measured by transcranial Doppler (MCA<sub>velocity</sub>: 2MHz probe, Spencer Technologies, Seattle, WA, USA, affixed to a custom-made headset) at depths ranging from 43 to 54 mm. Signal quality was optimized and an M-mode screen shot was recorded to facilitate subsequent probe placements. Arterial saturation was measured on the right side of the forehead by pulse oximetry (Nellcor N-200,

Mansfield, MA, USA). Limb lead electrodes were used to measure ECG (ADInstruments BioAmp, Colorado Springs, CO, USA and Sonosite Micromaxx, Bothell, WA, USA). Metabolic variables, including expired ventilation and gas concentrations were assessed via breath-by-breath (Medgraphics PFX, St. Paul, MN, USA and Vacumed UVM, Ventura, CA, USA) and mixing chamber (Oxigraf O<sub>2</sub>cap, Mountain View, CA, USA) systems, calibrated with the same 3-L syringe and known concentrations of O<sub>2</sub> and CO<sub>2</sub> prior to each test. Additionally, core temperature was monitored by telemetry pill (CorTemp HQInc., Palmetto, FL, USA) Analog data were sampled and recorded at 200Hz (ADinstruments Powerlab 16/30, Colorado Springs, CO, USA).

### **Cerebral Blood Flow**

After verification of signal quality, resting data were recorded for 10 min while subjects breathed room air. At 6 min, 2 ml of arterial blood was drawn anaerobically for blood gas analysis (described below). During the last 4 min of the resting period, diameter and blood flow velocity in the left internal carotid (ICA: 1.5 cm distal to the carotid bifurcation) and vertebral arteries (VA: between spinous processes of C4 and C5) were recorded over a minimum of 5 cardiac cycles by a registered diagnostic sonographer (SonoSite Micromaxx L25 probe, Bothell, WA, USA). Briefly, vessel diameter from a longitudinal view was identified and measured with digital calipers in synchronization with the ECG tracing to identify systole and diastole. Velocity was measured in the center of the vessel with an insonation angle < 60 degrees and a sample volume maximized for vessel diameter. The peak velocity tracing across cardiac cycles was used for calculation of mean velocity (time averaged peak) and volumetric flow. This procedure was used to verify accurate tracing of the spectral envelop during data collection and results in higher values than the time averaged mean method (Schoning *et al.*, 1994). All data were downloaded in DICOM format for verification of measurements offline (Sante DICOM Editor, Athens, Greece).

Regional blood flow (ml/min) in the ICA and VA (ICA<sub>flow</sub> and VA<sub>flow</sub>) was determined using standard, validated ultrasound techniques (Hoskins, 2008), where:

 $X_{Flow} = \pi^*$ (diameter in cm/2)<sup>2</sup> \* time averaged peak velocity in cm/s \* 60 s. Average coefficients of variation determined from three repeated measurements of ICA and VA flow measurements in 7 subjects at SL were 4.0 ± 2.6% and 4.0 ± 2.1%, respectively. Global CBF (gCBF) was estimated assuming symmetrical bilateral flow in the major tributary arteries of the brain (Ogoh *et al.*, 2013; Willie *et al.*, 2013) as:

 $gCBF = (ICA_{flow} + VA_{flow})^*2.$ 

Regional and global measurements of CBF were also expressed relative to estimates of cardiac output (%Q) derived from simultaneous intra-arterial blood pressure tracings (Bogert *et al.*, 2010). Cerebral vascular resistance index (CVRi) was calculated as:

 $CVRi = mean ABP/X_{flow}$ 

### **Cerebral Oxygen Delivery**

Arterial blood was immediately analyzed for PaO<sub>2</sub>, PaCO<sub>2</sub> (Siemens RAPIDLab 248, Erlangen, Germany), [Hb], SaO<sub>2</sub> (Radiometer OSM3, Copenhagen, Denmark) and Hct (M24 Centrifuge, LW Scientific, Lawrenceville, GA, USA). Blood gases were Accepted Articl

temperature corrected (Kelman & Nunn, 1966; Severinghaus, 1966). CaO<sub>2</sub> (vol%) was calculated as:

 $CaO_2 = 1.39 \text{ x [Hb]} + PaO_2 * 0.003$ 

Regional and global cerebral  $DO_2$  were calculated as the products of  $CaO_2$  and  $ICA_{flow}$ ,  $VA_{flow}$ , and gCBF.

### **Data Analysis**

After verification of normality, mixed repeated measures ANOVA's were used to analyze the interaction of time by sex for each variable of interest ( $\alpha = 0.05$ ). Subsequent estimation-maximization and multiple-imputation (5 trials) analyses verified negligible effects of missing values (SPSS 20, IBM, Chicago, IL, USA). Paired t-tests (without imputation of missing values) were used for *post hoc* comparisons with the Holm procedure to control for Type I error. A priori power calculations ( $\alpha =$ 0.05,  $\beta = 0.20$ ) were integrated into the study design to limit Type II error. Pearson product moment correlations were used to describe shared variance between variables. Data are presented as mean ± SD.

Based on the hypothesis that increased CBF may play a role in the pathogenesis of acute mountain sickness [AMS (Jensen *et al.*, 1990; Baumgartner *et al.*, 1994; Baumgartner *et al.*, 1999)], a secondary analysis was performed to evaluate potential relationships (Spearman correlations) between changes in CBF and DO<sub>2</sub> with the severity of Lake Louise Questionnaire (LLQ) symptoms scores reported in these subjects on ALT1 (Subudhi et al. – in review). Paired t-tests were used to evaluate differences in CBF and  $DO_2$  between those with severe AMS (LLQ  $\ge 6$  including headache) and those remaining healthy.

# **Results**

### Subject Characteristics

Detailed baseline characteristics of the 21 (12 males and 9 females;  $21 \pm 1$  years old) subjects participating in AltitudeOmics are presented elsewhere (Subudhi *et al.*, In Review). Males exhibited greater [Hb], CaO<sub>2</sub> and DO<sub>2</sub> than females over the course of the study (all P < 0.05), but since no interactions in CBF or DO<sub>2</sub> were detected across acclimatization, combined data are presented below.

### **Cerebral Blood Flow and Oxygen Delivery**

Acute exposure to 5,260 m (Pbar = 408 ± 1 mmHg) decreased PaO<sub>2</sub>, SaO<sub>2</sub> and CaO<sub>2</sub> by 66.1 ± 5.4 mmHg, 22 ± 6%, and 4.1 ± 1.2 ml/dl, respectively (all P<0.001; Table 1). This severe degree of hypoxia increased heart rate 14 ± 11 bpm (P < 0.001) without affecting mean ABP (P=0.380). CBF increased 74 ± 81% in the ICA (P = 0.018), 59 ± 54% in the VA (P = 0.001), and 69 ± 57% globally (P = 0.003). Respective CVRi values fell (all P< 0.001; Table 2), allowing a larger percentage of cardiac output to perfuse the brain (P = 0.010). Increased ICA<sub>flow</sub> was characterized by increased ICA velocity (P = 0.004) without a change in diameter (P =0.068), while increased VA<sub>flow</sub> was explained by an increase in VA diameter (P = 0.005) without a change in velocity (P=0.120). MCA<sub>velocity</sub> was unchanged (P = 0.953). Increased gCBF offset the decrease in CaO<sub>2</sub> to maintain global cerebral DO<sub>2</sub> (Figure 1), although a small increase in VA DO<sub>2</sub> was observed (P=0.039, Figure 2). Observed changes in measures of regional and global CBF and DO<sub>2</sub> were not correlated with LLQ scores of AMS (r = -0.07 to -0.23, P = 0.38 to 0.78), nor were they different between those reporting severe AMS and those remaining healthy (P = 0.57 to 0.97).

Following acclimatization, a  $32 \pm 36\%$  rise in ventilation was accompanied by a 5.5  $\pm$  2.7 mmHg decrease in PaCO<sub>2</sub> and 9.2  $\pm$  4.1 mmHg increase in PaO<sub>2</sub> (ALT1 vs. ALT16; all P < 0.001). SaO<sub>2</sub> and [Hb] rose 6  $\pm$  5% and 1.8  $\pm$  0.9 g/dL, respectively, improving CaO<sub>2</sub> by 3.1  $\pm$  1.2 ml/dl (all P < 0.001; Table 1). ABP was unaffected by acclimatization (ALT1 vs. ALT16; P=0.211). ICA<sub>flow</sub>, VA<sub>flow</sub> and gCBF returned to SL values (SL vs. ALT16; P = 0.810, 0.977, 0.620, respectively; Table 2). Respective CVRi values increased as both ICA and VA diameters decreased from ALT1 to ALT16 (all P < 0.020) and restored the relative distribution of cardiac output back to SL values (SL vs. ALT16; P = 0.121). Cerebral DO<sub>2</sub> fell from ALT1 to ALT16 (ICA DO<sub>2</sub> P = 0.028, VA DO<sub>2</sub> P = 0.020, global DO<sub>2</sub> P = 0.011) as the reductions in CBF outweighed the increase in CaO<sub>2</sub> (Figure 1); however, neither global nor regional cerebral DO<sub>2</sub> values fell below that measured at SL (all P > 0.420; Figures 1 & 2).

# Discussion

This is the first study to assess regional cerebral oxygen delivery in the field over a period of acclimatization to high altitude. Our findings confirm that global cerebral DO<sub>2</sub> was preserved across acclimatization through a changing balance between CBF and CaO<sub>2</sub>, but there was slight increase in relative DO<sub>2</sub> to the posterior cerebral

circulation during acute exposure. Although changes in CBF and DO<sub>2</sub> were not associated with the incidence or severity of AMS, regional regulation of CBF may serve to support vital homeostatic cerebral functions in hypoxia.

### Preservation of Cerebral Oxygen Delivery

The increase in CBF upon arrival at high altitude and decrease back to sea level values with acclimatization was opposed by changes in  $CaO_2$  (Figure 2). These responses preserved cerebral  $DO_2$  close to sea level values and affirm that components of  $CaO_2$  (PaO<sub>2</sub>, SaO<sub>2</sub>, [Hb]) outweigh the influence PaCO<sub>2</sub> in regulating CBF in severe hypoxia. Increased CBF upon arrival at high altitude resulted from reduced cerebral vascular resistance rather than increased blood pressure (Tables 1&2). Although reduction in vascular resistance is commonly attributed to dilation of pial and parenchymal arterioles in the brain (Fog, 1938), we observed increased diameter of larger tributary arteries, supporting a global vascular response to this degree of hypoxia (Heistad *et al.*, 1978; Faraci & Heistad, 1990; Willie *et al.*, 2012). Mechanisms governing hypoxic vasodilation are complex, involving local (e.g. astrocyte regulation, nitric oxide) and diffuse (e.g. central chemoreception, autonomic nervous system) mechanisms, but all stem from a reduction in  $PaO_2$ (Severinghaus, 2001; Xu & Lamanna, 2006). When PaO<sub>2</sub> is above 60 mmHg, little vasodilation is evident (Mardimae et al., 2012; Willie et al., 2012). Below this threshold, the degree of vasodilation increases exponentially and outweighs the degree of hypocapnic vasoconstriction (Mardimae et al., 2012; Willie et al., 2012) presumably to provide greater blood flow in a time of need. While the correlation between changes in gCBF and  $CaO_2$  was not significant, the change in  $CaO_2$  from SL

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to ALT1 was similar among all subjects and may not have afforded an appropriate range of values to detect the relation that has previously been shown with progressive hemodilution (Korosue & Heros, 1992). Qualitatively, the ~70% increase in gCBF was within the expected range during acute hypocapnic hypoxia (Severinghaus, 1966; Jensen *et al.*, 1990; Severinghaus, 2001; Brugniaux *et al.*, 2007) and proportional to the ~60% reduction in PaO<sub>2</sub> that was responsible for the reduction in CaO<sub>2</sub>. This reciprocal relationship, whether evolved or serendipitous, is advantageous for survival in these extreme conditions as it mitigates negative consequences of cerebral hypoxemia.

Although increased CBF has been suggested to play a role in the pathogenesis of AMS (Baumgartner *et al.*, 1994), our results were more similar to those refuting the hypothesis (Jensen *et al.*, 1990; Baumgartner *et al.*, 1999). Regional and global CBF and DO<sub>2</sub> measurements were not correlated with AMS symptoms scores and did not differentiate between those with severe AMS and those who remained healthy after rapid ascent to high altitude. Nonetheless, our data should be interpreted with caution since it is possible that increased CBF contributes to the development of AMS when other, yet to be described, factors are present.

Increased  $PO_2$  and decreased  $PCO_2$  after 16 days at high altitude are hallmarks of ventilatory acclimatization that are addressed elsewhere (Fan et al. in review). As a result,  $PaO_2$ -mediated dilation was reduced and  $PaCO_2$ -mediated vasoconstriction was increased, thereby lowering CBF. Assuming a cerebral  $O_2$  reactivity of 3% CBF /

% SaO<sub>2</sub> and a CO<sub>2</sub> reactivity of 4% CBF / mmHg CO<sub>2</sub> from a previous duplex ultrasound study (Willie *et al.*, 2012), we could account for the entire decrease in gCBF across acclimatization. Specifically, the 5% increase in SaO<sub>2</sub> could be expected to reduce CBF by  $\sim 15\%$  and the 5.5 mmHg decrease in PaCO<sub>2</sub> could be expected to reduce CBF by ~22%, thus accounting for the 36% decrease in gCBF we observed from ALT1 to ALT16 (Table 2). We acknowledge that increased cerebrovascular CO<sub>2</sub> reactivity with acclimatization in our subjects (Fan et al. in review) may account for an even greater proportion of the net effect on CBF at ALT16. Also, the relative influence of other hematological factors, such as increased hematocrit and blood viscosity (Sorensen et al., 1974; Todd et al., 1994; Tomiyama et al., 1999) from erythropoiesis and plasma volume contraction, may have contributed to the reduction of CBF across acclimatization (data to be presented elsewhere). Yet our data suggest that the inherent vascular reactivities to O<sub>2</sub> and CO<sub>2</sub> are sufficient to maintain tight control over cerebral DO<sub>2</sub> in hypoxia. Consistent delivery of oxygen may help offset the decreased PO<sub>2</sub> gradient (plasma to mitochondria) and support the cerebral metabolic demand for oxygen at this altitude (Severinghaus *et al.*, 1966; Moller et al., 2002) to preserve cerebral function. Together, our data demonstrate that integrated mechanisms controlling cerebral blood flow are well suited to preserve global cerebral oxygen delivery at 5,260 m.

### **Regional Cerebral Oxygen Delivery**

We observed a small increase in DO<sub>2</sub> through the posterior cerebral circulation upon arrival at high altitude (Table 2) that dissipated with acclimatization. The acute increase in DO<sub>2</sub> was characterized by an increase in VA diameter and supports

recent findings of greater VA (vs. ICA) vasoreactivity during acute hypoxia (Willie et al., 2012; Ogoh et al., 2013). Of note, Ogoh et al. (Ogoh et al., 2013), showed that acute hypoxia (~15 min) increased VA, but not ICA, blood flow. Since the areas perfused by the VA include the brainstem, and posterior aspects of the thalamus and hypothalamus, increased blood flow and DO<sub>2</sub> to these regions during acute hypoxia (Buck et al., 1998; Binks et al., 2008) may be seen as necessary to maintain vital homeostatic functions (Sheldon et al., 1979; Bilger & Nehlig, 1993). Since increased cardiorespiratory drive with acclimatization was not associated with a continued elevation of VA DO<sub>2</sub>, we speculate that the increased VA DO<sub>2</sub> during acute hypoxia was protective, to defend against a potential threat in oxygen supply, rather than to merely support neuronal metabolic activity associated with heightened autonomic activity (i.e. neurovascular coupling). Although such hypothetical explanations for regional differences in the regulation of CBF and DO<sub>2</sub> are intriguing, our results must be interpreted with caution since measured differences were small and are not consistently reported in the literature (Huang et al., 1987; Willie et al., 2013). Future studies with more focal measurements of  $DO_2$  (e.g. PET and MRI) and neuronal activity in key regulatory regions of the brain, as well as measurements of neurovascular coupling (as an index of neuronal plasticity) during acute and prolonged hypoxia are needed to yield further insight into this question.

### **Brain Sparing**

Reduced cerebral vascular resistance associated with vasodilation upon arrival at altitude can explain the proportional increase in CBF and greater allocation of cardiac output. This effect could be magnified if there is net constriction in other

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vascular beds at rest. Previous studies have shown that superior mesenteric and renal artery blood flow decrease in acute hypoxia and could allow for greater perfusion of the brain (Greene & Roach, 2004). With acclimatization, cerebral vascular resistance and blood flow returned to sea-level values. These results are similar to fetal 'brain sparing' effects (Campbell et al., 1967; Peeters et al., 1979; Sheldon et al., 1979) that are presumed to preserve vital homeostasis during hypoxia in utero (Pearce, 2006; Salihagic-Kadic et al., 2006). Similar effects have also been shown in newborn dogs (Cavazzuti & Duffy, 1982), piglets (Goplerud *et al.*, 1989), and premature infants (Daven *et al.*, 1983). The largest response to hypoxia tends to occur in the brainstem during the early postnatal period and decreases with age (Bilger & Nehlig, 1993). We are the first to demonstrate that such a 'brain sparing' reaction exists in healthy human adults exposed to acute hypoxia and recedes with acclimatization. Preferential distribution of cardiac output to the brain upon acute altitude exposure may represent a conserved mechanism that protects against hypoxic brain damage in mammals, particularly in regions associated with basic cardiovascular and respiratory control during periods of acute hypoxia. Measurements of regional cerebral metabolism are needed to determine if 'brain sparing' effectively matches DO<sub>2</sub>, or if the increase in CBF represents a protective form of overcompensation.

### Limitations

Our rapid ascent profile in combination with supplemental oxygen during transport from low to high altitude was designed to induce an abrupt change in PaO2, similar to that which can be achieved in laboratory studies with hypoxic gas or hypobaric

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chambers. As such, our results must be interpreted in this context and thus may be expected to be different from other field studies that have followed more traditional progressive ascents (Huang *et al.*, 1987; Jensen *et al.*, 1990; Baumgartner *et al.*, 1994; Willie *et al.*, 2013).

We used duplex sonography primarily because it is a non-invasive technique that can be utilized in field settings. This technique yields volumetric measurements, in terms of ml/min, which, based on first principles, can be multiplied by  $CaO_2$  to yield DO<sub>2</sub>. Our low CVs were in line with a previous study showing similarity between duplex sonography and both PET and xenon inhalation methods of measuring gCBF (Schoning & Scheel, 1996). Nevertheless, we acknowledge that all these techniques are limited by the lack of an absolute standard for validating CBF. Our gCBF measurements were based on unilateral, left-sided measurements of the ICA and VA - the main arteries perfusing the brain. While left VA flow has been reported to be  $\sim$ 20% higher than the right (Schoning *et al.*, 1994), this was not expected to have an effect on global measurements since ICA flow represents the majority of gCBF (Schoning & Scheel, 1996). Yet, unilateral VA measurements may have influenced our finding of increased VA DO<sub>2</sub>. Future studies are needed to determine if 'brain sparing' effects are attenuated when independent measurements of left and right VA flow are summed.

Since the ICA feeds the MCA, we expected that changes in ICA flow would be reflected in MCA<sub>velocity</sub>. This was not the case: ICA flow increased  $\sim$ 70% while

MCA<sub>velocity</sub> was unchanged throughout the study. A similar discrepancy between ICA flow and MCA<sub>velocity</sub> has been previously described by Willie et al. (Willie *et al.*, 2012) and argued to support dilation of the MCA in hypoxia (Wilson *et al.*, 2011). We calculated that a 12% increase in MCA diameter could explain the measured discrepancy between ICA<sub>flow</sub> and MCA<sub>velocity</sub>. This exact degree of vasodilation has recently been demonstrated at high altitude with a color-coded ultrasound technique (Willie *et al.*, 2013), yet because additional studies are needed to clarify artery-specific responses to hypoxia and validate MCA-diameter measurement techniques, we chose to refrain from further interpretation of MCA<sub>velocity</sub>.

### **Summary & Implications**

Overall, our findings highlight the integrative nature of responses that preserve oxygen delivery to the brain at high altitude. Regional cerebral vasoreactivity to O<sub>2</sub> and CO<sub>2</sub> may favor oxygen delivery to posterior and inferior regions of the brain during acute hypoxia to sustain vital cerebral functions associated with homeostasis. Whether these mechanisms evolved to promote survival in conditions provoking cerebral hypoxia is not clear at present, but further research in this area may yield important insights into human tolerance and adaptation to chronic states of hypoxemia.

# **Competing Interests**

The authors have no conflicts or competing interests to disclose.

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# References

- Baumgartner RW, Bartsch P, Maggiorini M, Waber U & Oelz O (1994). Enhanced cerebral blood flow in acute mountain sickness. *Aviat Space Environ Med* **65**, 726-729.
- Baumgartner RW, Spyridopoulos I, Bartsch P, Maggiorini M & Oelz O (1999). Acute mountain sickness is not related to cerebral blood flow: a decompression chamber study. *J Appl Physiol* **86**, 1578-1582.
- Bert P (1943). Barometric Pressure. College Book Company, Columbus, Ohio.
- Bilger A & Nehlig A (1993). Regional cerebral blood flow response to acute hypoxia changes with postnatal age in the rat. *Brain Res Dev Brain Res* **76**, 197-205.
- Binks AP, Cunningham VJ, Adams L & Banzett RB (2008). Gray matter blood flow change is unevenly distributed during moderate isocapnic hypoxia in humans. *J Appl Physiol* **104**, 212-217.
- Bogert LW, Wesseling KH, Schraa O, Van Lieshout EJ, de Mol BA, van Goudoever J, Westerhof BE & van Lieshout JJ (2010). Pulse contour cardiac output derived from non-invasive arterial pressure in cardiovascular disease. *Anaesthesia* 65, 1119-1125.
- Brugniaux JV, Hodges AN, Hanly PJ & Poulin MJ (2007). Cerebrovascular responses to altitude. *Respir Physiol Neurobiol* **158**, 212-223.
- Buck A, Schirlo C, Jasinksy V, Weber B, Burger C, von Schulthess GK, Koller EA & Pavlicek V (1998). Changes of cerebral blood flow during short-term exposure to normobaric hypoxia. *J Cereb Blood Flow Metab* **18**, 906-910.
- Campbell AG, Dawes GS, Fishman AP & Hyman AI (1967). Regional redistribution of blood flow in the mature fetal lamb. *Circ Res* **21**, 229-235.
- Cavazzuti M & Duffy TE (1982). Regulation of local cerebral blood flow in normal and hypoxic newborn dogs. *Ann Neurol* **11**, 247-257.
- Daven JR, Milstein JM & Guthrie RD (1983). Cerebral vascular resistance in premature infants. *Am J Dis Child* **137**, 328-331.
- Faraci FM & Heistad DD (1990). Regulation of large cerebral arteries and cerebral microvascular pressure. *Circ Res* **66**, 8-17.

- Articl C Accepte
- Fog M (1938). The Relationship between the Blood Pressure and the Tonic Regulation of the Pial Arteries. *J Neurol Psychiatry* **1**, 187-197.
- Goplerud JM, Wagerle LC & Delivoria-Papadopoulos M (1989). Regional cerebral blood flow response during and after acute asphyxia in newborn piglets. *J Appl Physiol* **66**, 2827-2832.
- Greene ER & Roach RC (2004). Doppler ultrasound determination of the distribution of human cardiac output: effects of age and physical stresses. *Conf Proc IEEE Eng Med Biol Soc* **5**, 3704-3707.
- Heistad DD, Marcus ML & Abboud FM (1978). Role of large arteries in regulation of cerebral blood flow in dogs. *J Clin Invest* **62**, 761-768.
- Hoskins PR (2008). Simulation and validation of arterial ultrasound imaging and blood flow. *Ultrasound Med Biol* **34**, 693-717.
- Huang SY, Moore LG, McCullough RE, McCullough RG, Micco AJ, Fulco C, Cymerman A, Manco-Johnson M, Weil JV & Reeves JT (1987). Internal carotid and vertebral arterial flow velocity in men at high altitude. J Appl Physiol 63, 395-400.
- Jensen JB, Wright AD, Lassen NA, Harvey TC, Winterborn MH, Raichle ME & Bradwell AR (1990). Cerebral blood flow in acute mountain sickness. *J Appl Physiol* **69**, 430-433.
- Kelman GR & Nunn JF (1966). Nomograms for correction of blood Po2, Pco2, pH, and base excess for time and temperature. *J Appl Physiol* **21**, 1484-1490.
- Korosue K & Heros RC (1992). Mechanism of cerebral blood flow augmentation by hemodilution in rabbits. *Stroke* **23**, 1487-1492; discussion 1492-1483.
- Lucas SJ, Burgess KR, Thomas KN, Donnelly J, Peebles KC, Lucas RA, Fan JL, Cotter JD, Basnyat R & Ainslie PN (2011). Alterations in cerebral blood flow and cerebrovascular reactivity during 14 days at 5050 m. *J Physiol* **589**, 741-753.
- Luft UC, Clamann HG & Opitz E (1951). The latency of hypoxia on exposure to altitude above 50,000 feet. *J Aviat Med* **22**, 117-122; passim.
- Luft UC & Noell WK (1956). Manifestations of brief instantaneous anoxia in man. *J Appl Physiol* **8**, 444-454.
- Mardimae A, Balaban DY, Machina MA, Han JS, Katznelson R, Minkovich LL, Fedorko L, Murphy PM, Wasowicz M, Naughton F, Meineri M, Fisher JA & Duffin J (2012). The interaction of carbon dioxide and hypoxia in the control of cerebral blood flow. *Pflugers Arch* **464**, 345-351.

- Moller K, Paulson OB, Hornbein TF, Colier WN, Paulson AS, Roach RC, Holm S & Knudsen GM (2002). Unchanged cerebral blood flow and oxidative metabolism after acclimatization to high altitude. *J Cereb Blood Flow Metab* 22, 118-126.
  Ogoh S, Sato K, Nakahara H, Okazaki K, Subudhi AW & Miyamoto T (2013). Effect of acute hypoxia on blood flow in vertebral and internal carotid arteries. *Exp Physiol* 98, 692-698.
  - Pearce W (2006). Hypoxic regulation of the fetal cerebral circulation. *J Appl Physiol* **100**, 731-738.
  - Peeters LL, Sheldon RE, Jones MD, Jr., Makowski EL & Meschia G (1979). Blood flow to fetal organs as a function of arterial oxygen content. *Am J Obstet Gynecol* 135, 637-646.
  - Salihagic-Kadic A, Medic M, Jugovic D, Kos M, Latin V, Kusan Jukic M & Arbeille P (2006). Fetal cerebrovascular response to chronic hypoxia--implications for the prevention of brain damage. *J Matern Fetal Neonatal Med* **19**, 387-396.
  - Sato K, Sadamoto T, Hirasawa A, Oue A, Subudhi AW, Miyazawa T & Ogoh S (2012). Differential blood flow responses to CO(2) in human internal and external carotid and vertebral arteries. *J Physiol* **590**, 3277-3290.
  - Schoning M & Scheel P (1996). Color duplex measurement of cerebral blood flow volume: intra- and interobserver reproducibility and habituation to serial measurements in normal subjects. *J Cereb Blood Flow Metab* **16**, 523-531.
  - Schoning M, Walter J & Scheel P (1994). Estimation of cerebral blood flow through color duplex sonography of the carotid and vertebral arteries in healthy adults. *Stroke* **25**, 17-22.

Severinghaus JW (1966). Blood gas calculator. *J Appl Physiol* **21**, 1108-1116.

- Severinghaus JW (2001). Cerebral circulation at altitude. In *High altitude: an exploration of human adaptation*, vol. 161. ed. Hornbein TF & Schoene RB, pp. 343-375. Marcel Dekker, New York, New York.
- Severinghaus JW, Chiodi H, Eger EI, 2nd, Brandstater B & Hornbein TF (1966). Cerebral blood flow in man at high altitude. Role of cerebrospinal fluid pH in normalization of flow in chronic hypocapnia. *Circ Res* **19**, 274-282.
- Sheldon RE, Peeters LL, Jones MD, Jr., Makowski EL & Meschia G (1979). Redistribution of cardiac output and oxygen delivery in the hypoxemic fetal lamb. *Am J Obstet Gynecol* **135**, 1071-1078.

- Artic Accepte
- Smith ZM, Krizay E, Guo J, Shin DD, Scadeng M & Dubowitz DJ (2013). Sustained high-altitude hypoxia increases cerebral oxygen metabolism. *J Appl Physiol* **114**, 11-18.
  - Sorensen SC, Lassen NA, Severinghaus JW, Coudert J & Zamora MP (1974). Cerebral glucose metabolism and cerebral blood flow in high-altitude residents. *J Appl Physiol* **37**, 305-310.
  - Subudhi AW, Bucher J, Bourdillon N, Elliot JE, Evero O, Fan JL, Jameson-Van Houton S, Julian CG, Kark S, Kern J, Kim S, Laurie SS, Lovering AT, Kayser B & Roach RC (In Review). AltitudeOmics: The integrative physiology of the onset and retention of human acclimatization to hypoxia.
  - Todd MM, Wu B, Maktabi M, Hindman BJ & Warner DS (1994). Cerebral blood flow and oxygen delivery during hypoxemia and hemodilution: role of arterial oxygen content. *Am J Physiol* **267**, H2025-2031.
  - Tomiyama Y, Jansen K, Brian JE, Jr. & Todd MM (1999). Hemodilution, cerebral O2 delivery, and cerebral blood flow: a study using hyperbaric oxygenation. *Am J Physiol* **276,** H1190-1196.
  - Wade OL & Bishop JM (1962). *Cardiac output and regional blood flow*. Davis, Philadelphia.
  - Willie CK, Macleod DB, Shaw AD, Smith KJ, Tzeng YC, Eves ND, Ikeda K, Graham J, Lewis NC, Day TA & Ainslie PN (2012). Regional brain blood flow in man during acute changes in arterial blood gases. *J Physiol* **590**, 3261-3275.
  - Willie CK, Smith KJ, Day TA, Ray LA, Lewis NC, Bakker A, Macleod DB & Ainslie PN (2013). Regional cerebral blood flow in humans at high altitude: Gradual ascent and two weeks at 5050m. *J Appl Physiol*.
  - Wilson MH, Edsell ME, Davagnanam I, Hirani SP, Martin DS, Levett DZ, Thornton JS, Golay X, Strycharczuk L, Newman SP, Montgomery HE, Grocott MP & Imray CH (2011). Cerebral artery dilatation maintains cerebral oxygenation at extreme altitude and in acute hypoxia--an ultrasound and MRI study. *J Cereb Blood Flow Metab* **31**, 2019-2029.
  - Wolff CB, Barry P & Collier DJ (2002). Cardiovascular and respiratory adjustments at altitude sustain cerebral oxygen delivery -- Severinghaus revisited. *Comp Biochem Physiol A Mol Integr Physiol* **132**, 221-229.
  - Xu K & Lamanna JC (2006). Chronic hypoxia and the cerebral circulation. *J Appl Physiol* **100**, 725-730.

	Variable	
Articl	VE PaO2 PaCO2 SaO2 [Hb] CaO2 HR SV Mean ABP	l/min mmHg mmHg % g/dl ml/dl bpm ml mmHg
5	* Different f # Different f ALT1	rom SL rom
pte		
CO		
A		

Table 1. Cardiopulmonary and hematological values [mean ±SD (n)]

SL

12.05 ± 2.50 (21)

102.2 ± 5.5 (21)

38.1 ± 4.4 (21)

98 ± 1 (21)

13.9 ± 1.4 (21)

19.4 ± 1.9 (21)

76 ± 12(21)

91 ± 27 (21)

79 ± 8 (21)

ALT1

11.93 ± 2.92 (17)

36.1 ± 2.8 (18)\*

26.5 ± 3.1 (18)\*

76 ± 6 (18)\*

14.2 ± 1.5 (18)\*

15.2 ± 2.1 (18)\*

90 ± 16 (16)\*

85 ± 20 (16)

76 ± 13 (16)

ALT16

14.88 ± 2.65 (21)\*#

45.3 ± 3.2 (20)\*#

20.9 ± 2.5 (20)\*#

82 ± 3 (20)\*#

16.0 ± 2.0 (20)\*#

18.4 ± 2.4 (20)\*#

96 ± 13 (20)\*

83 ± 21 (20)

80 ± 10 (20)

Variable		SL	ALT1	ALT16
ICA Dia	cm	0.51 ± 0.08 (21)	0.54 ± 0.07 (16)	0.50 ± 0.07 (20)#
ICA Vel	cm/s	29.8 ± 8.2 (21)	38.9 ± 8.1 (16)*	32.1 ± 5.4 (20)#
ICA Flow	ml/min	384 ± 197 (21)	556 ± 203 (16)*	379 ± 97(20)#
ICA CVRi	mmHg/ml/min	0.25 ± 0.12 (21)	0.16 ± 0.09 (16)*	0.23 ± 0.07 (19)#
VA Dia	cm	0.36 ± 0.06 (20)	0.41 ± 0.06 (16)*	0.36 ± 0.06 (19)#
VA Vel	cm/s	21.4 ± 4.4 (20)	24.4 ± 6.4 (16)	19.3 ± 7.1 (19)#
VA Flow	ml/min	133 ± 47 (20)	206 ± 98 (16)*	122 ± 55 (19)#
VA CVRi	mmHg/ml/min	0.66 ± 0.24 (20)	0.46 ± 0.28 (16)*	0.84 ± 0.58 (19)#
aCDF	ml/min	1057 ± 412 (20)	1524 + 456 (16)*	001 ± 000 (10)#
	1111/11111 	$1057 \pm 415 (20)$	$1524 \pm 450 (10)^{\circ}$	961 ± 225 (19)#
gCBF CVRI	mmHg/mi/min	0.09 ± 0.03 (20)	0.05 ± 0.02 (16)*	0.08 ± 0.02 (19)#
DO2 ICA	ml/min	75 ± 37 (21)	84 ± 32 (16)	68 ± 19 (19)#
DO2 VA	ml/min	26 ± 10 (20)	31 ± 16 (16)*	22 ± 11 (19)#
DO2 gCBF	ml/min	206 ± 79 (20)	230 ± 74 (16)	181 ± 51 (19)#
_				
MCAv	cm/s	59.5 ± 10.3 (21)	61.1 ± 13.3 (17)	57.7 ± 7.1 (21)
MCA CVRi	mmHg/cm/s	1.36 ± 0.25 (21)	1.28 ± 0.32 (17)	1.41 ± 0.24 (20)
ICA %O	%	5.4 + 2.7 (21)	7.6 + 2.7 (15)*	4.8 + 1.4 (18)#
VA %O	%	19 + 08(20)	$2.6 \pm 1.1 (15)^*$	$15 \pm 0.7 (18)$ #
	%	15 0 + 5 8 (20)	$2.0 \pm 1.1 (13)$ 20 4 + 6 2 (15)*	12 6 + 3 <u>4</u> (18)#
BCDI /0Q	/0	13.0 - 3.0 (20)	20.4 ± 0.2 (13)	12.0 - 3.4 (10)#

Table 2. Cerebrovascular values [mean± SD (n)]

\* Different from SL

# Different from ALT1

Figure 1. Reciprocal changes in global cerebral blood flow (gCBF) and arterial oxygen content (CaO2) maintained global cerebral oxygen delivery (DO2) across the study. \* Different from sea level (SL). # Different from arrival at altitude (ALT1).



Figure 2. Regional oxygen delivery  $(DO_2)$  increases in the vertebral artery (VA), but not internal carotid artery (ICA) at ALT1. Regional  $DO_2$  is reduced with acclimatization, but not below sea level (SL) values. \* Different from SL. # Different from arrival at altitude (ALT1).

ALT 1

ALT 1

#

ALT 16

#

ALT 16


1	AltitudeOmics: Enhanced	cerebrovascular	reactivity an	nd ventilatory	response to	$\rm CO_2$	with hi	gh
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2 altitude acclimatisation and re-exposure

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#### 25 Abstract

26 The present study is the first to examine the effect of high altitude acclimatisation and re-27 exposure on the responses of cerebral blood flow and ventilation to  $CO_2$ . We also compared the 28 steady-state estimates of these parameters during acclimatisation with the modified rebreathing 29 method. We assessed changes in steady state responses of middle cerebral artery velocity 30 (MCAv), cerebrovascular conductance index (CVCi) and ventilation (VE) to varied levels of  $CO_2$  in 31 21 lowlanders (9 females;  $21 \pm 1$  years), at sea-level (SL), during initial exposure to 5,260m (ALT1), 32 after 16 days of acclimatisation (ALT16) and upon re-exposure to altitude following either 7 33 (POST7) or 21 days (POST21) at low altitude (1,525m). In the non-acclimatised state (ALT1), MCAv 34 and VE responses to  $CO_2$  were elevated compared to SL (by 79±75% and 14.8±12.3 L/min, 35 respectively, P=0.004 & P=0.011). Acclimatisation at ALT16 further elevated both MCAv and VE 36 responses to  $CO_2$  compared to ALT1 (by 89±70% and 48.3±32.0 L/min, respectively, P<0.001). The 37 acclimatisation gained for VE responses to  $CO_2$  at ALT16 was retained by 38% upon re-exposure to 38 altitude at POST7 (P=0.004 vs. ALT1), while no retention was observed for the MCAv responses 39 (P>0.05). We found good agreement between steady-state and modified rebreathing estimates of 40 MCAv and VE responses to  $CO_2$  across all three time points (P<0.001, pooled data). Regardless of 41 the method of assessment, altitude acclimatisation elevates both the cerebrovascular and 42 ventilatory responsiveness to CO<sub>2</sub>. Our data further demonstrates that this enhanced ventilatory 43 CO<sub>2</sub> response is partly retained after 7 days at low altitude.

#### 45 Introduction

46 The ability to maintain adequate oxygen transport to the brain by cerebral blood flow 47 (CBF) in hypoxic environments is vital. The CBF responsiveness to CO<sub>2</sub>, termed cerebrovascular 48 CO<sub>2</sub> reactivity, provides a useful, non-invasive index of cerebrovascular function (3, 19). To date, 49 only a handful of studies have investigated the effect of acclimatisation to high altitude on 50 cerebrovascular CO<sub>2</sub> reactivity (1, 16, 17, 24, 30, 49). The interpretation of findings from these 51 studies is difficult due to the timing of measurements at high altitude (1, 16, 17, 24, 25), the 52 confounding effects of previous high-altitude exposure (1), artificial normobaric hypoxia (28, 46), 53 and the method used to assess reactivity (24, 30, 49). Data from Fan et al., (16, 17), obtained on 54 subjects at different stages of altitude acclimatisation, suggest that cerebrovascular CO<sub>2</sub> reactivity 55 is elevated with prolonged exposure to high altitude when using a modified rebreathing 56 technique. In contrast, Lucas et al., (30) reported a reduced cerebrovascular  $CO_2$  reactivity in the 57 same subjects that at the end of a 14 day stay at 5,050 m, when assessed with a steady-state 58 technique (poikilocapnic hypoxia). More recently, Rupp et al., (49) reported a reduced 59 cerebrovascular CO<sub>2</sub> reactivity during steady-state hypoxic hypercaphia following 5 days at 4,350 60 m. Thus, the effect of altitude acclimatisation on cerebrovascular CO<sub>2</sub> reactivity remains unclear.

In addition, it is unknown if and for how long changes in cerebrovascular CO<sub>2</sub> reactivity from acclimatisation persist after descent. Repetitive seven-month exposures to high altitude were reported to improve arterial O<sub>2</sub> saturation (SaO<sub>2</sub>), lower resting heart rate (HR) and decrease susceptibility to acute mountain sickness (AMS) upon subsequent re-exposures (59). Remarkably, these prior-exposure adaptations persisted despite a five-month deacclimatisation period. The specific effect of high altitude re-exposure on cerebrovascular and ventilatory responsiveness to CO<sub>2</sub> has yet to be examined.

68 Changes in cerebrovascular  $CO_2$  reactivity with high-altitude acclimatisation depend on the 69 method of assessment. At sea level, the steady-state method results in higher cerebrovascular  $CO_2$ 

70 reactivity (40-42) and lower ventilatory  $CO_2$  sensitivity (6, 18, 23, 55) compared to the modified 71 rebreathing test. These differences have been attributed to the presence of a  $PCO_2$  gradient 72 (between alveolar, arterial, and cerebrospinal fluid compartments) during the steady-state 73 method, which is supposedly abolished or minimised during rebreathing (6). Meanwhile, elevated 74 basal VE and subsequent underestimation of the ventilatory CO<sub>2</sub> sensitivity has been proposed as 75 one possible explanation for lower steady-state estimates (34). No studies have directly compared 76 the steady-state and modified rebreathing test estimates of cerebrovascular and ventilatory CO<sub>2</sub> 77 responsiveness following ascent or acclimatisation to high altitude.

The purpose of the present study was therefore two-fold: first, we wished to assess the effect of altitude exposure on cerebrovascular and ventilatory responsiveness to  $CO_2$  in acute conditions, after acclimatisation and upon re-exposure to high altitude after a period spent at low altitude; second, we wished to compare the steady-state and modified rebreathing methods for assessing the ventilatory and cerebrovascular responsiveness to  $CO_2$  at high altitude.

83

84

#### 85 Methods

86 Subject recruitment and screening

This study was conducted as part of the AltitudeOmics project. Following institutional ethics approval, young (19-23 years old), healthy, sea level residents were recruited from the greater Eugene, Oregon area (elevation 130 m). Potential subjects were screened to exclude anyone who was born or had lived at altitudes >1500 m for more than one year or had travelled to altitudes >1000 m in the past 3 months. A detailed description of subject recruitment procedures, including inclusion and exclusion criteria have been presented elsewhere (54).

93

94 Ethical approval

The study was performed according to the *Declaration of Helsinki* and was approved by the Institutional Review Boards of the Universities of Colorado and Oregon and by the Human Research Protection Office of the U.S. Department of Defense. All participants were informed regarding the procedures of this study, and written informed consent was given prior to participation.

100

101 Experimental Design

After familiarisation with the experimental procedures outlined below (visit one), the subjects underwent experimental trials near sea level (SL: 130 m, barometric pressure: 749 mmHg) and three times at high altitude (5,260 m, Mt Chacaltaya, Bolivia; barometric pressure 406 mmHg); on the 1<sup>st</sup> and 16<sup>th</sup> days at high altitude (ALT1 and ALT16) and again after either 7 (POST7; n=14) or 21 (POST21; n=7) days at low altitude (1,525 m, barometric pressure: 639 mmHg). An overview of the entire experimental design and protocol has been described in detail elsewhere (54).

109

110 Experimental protocol

For each subject, all ALT measurements were carried out around the same time of day to minimise any confounding effect of circadian rhythm. Measurements were taken upon arrival at ALT1 to minimise the influence of AMS. Likewise, no symptoms of AMS were observed at ALT16 or POST7.

For this study, following 10-15 min of quiet rest in a seated position, each experimental testing session comprised of: a) instrumentation; b) 10 min room air baseline; and c) cerebrovascular  $CO_2$  reactivity tests. The cerebrovascular  $CO_2$  reactivity tests consisted of: i) 10 min with end-tidal  $PCO_2$  (PETCO<sub>2</sub>) clamped at 40 mmHg; ii) 3 min voluntary hyperventilation to lower PETCO<sub>2</sub> to ~20 mmHg; iii) the modified rebreathing test (details below); and iv) 3 min with

120 PETCO<sub>2</sub> clamped at 50 mmHg. The entire cerebrovascular CO<sub>2</sub> reactivity protocol was carried out 121 in background of hyperoxia (end-tidal PO<sub>2</sub> [PETO<sub>2</sub>] > 250 mmHg).

122

#### 123 Experimental setup

124 Throughout the protocol, the subjects sat upright and breathed through a mouthpiece 125 attached to a two-way non-rebreathing valve (Hans-Rudolph 2700, Hans-Rudolph Inc., Shawnee, 126 KS, USA). The breathing circuit allowed switching from room air to either an end-tidal clamping 127 system or a rebreathing system. The end-tidal clamping setup used in the present study is a 128 modified version of the system previously described by Olin et al., (39). The setup allowed 129 stabilising PETCO<sub>2</sub> at 40 and 50 mmHg. Throughout the end-tidal PCO<sub>2</sub> clamping, we maintained 130 PETO<sub>2</sub> at >250 mmHg by titrating 50% or 100% O<sub>2</sub> into the inspiratory reservoir at SL and ALT, 131 respectively.

132

## 133 Modified rebreathing method

134 The modified rebreathing method is a well-established method for assessing both 135 ventilatory and cerebrovascular CO<sub>2</sub> reactivities (14, 16, 34, 41). By using hyperoxia (PETO<sub>2</sub> > 250 136 mmHg) the test minimises peripheral chemoreceptors' output (11, 21) and the ventilatory 137 response to the modified rebreathing method can thus be interpreted as the ventilatory  $CO_2$ 138 sensitivity primarily from the central chemoreflex. The details of the modified rebreathing method 139 have been previously described in Fan et al., (16, 17). The rebreathing bag was filled with gas to 140 achieve inspired  $PCO_2$  and  $PO_2$  of 0 mmHg and 300 mmHg, respectively, at each altitude. Subjects 141 were instructed to hyperventilate for 3 min (part ii) to lower and then maintain  $PETCO_2$  at 20 142 mmHg at both sea level and 5,260 m (in background  $PETO_2 > 250$  mmHg). Subjects were then 143 switched to the rebreathing bag, and following two initial deep breaths to mix the gas from the 144 bag with that in the respiratory system, they were instructed to breathe ad libitum (part iii). The

rebreathing tests were terminated when PETCO<sub>2</sub> reached 50 mmHg, PETO<sub>2</sub> dropped below 200
 mmHg or the subject reached the end of his/her hypercapnic tolerance.

147

148 *Measurements* 

149 Cerebrovascular variables: Middle cerebral artery velocity (MCAv, an index of cerebral 150 blood flow) was measured in the left middle cerebral artery using a 2-MHz pulsed Doppler 151 ultrasound system (ST3, Spencer technology, Seattle, WA, USA). The Doppler ultrasound probe 152 was positioned over the left temporal window and held in place with an adjustable plastic 153 headband (Marc 600 Headframe, Spencer technology, Seattle, WA, USA). The signal was acquired 154 at depths ranging from 43 to 54 mm. Signal quality was optimised and an M-mode screen shot 155 was recorded to facilitate subsequent probe placements. Peripheral saturation was measured on 156 the right side of the forehead by pulse oximetry (N-200, Nellcor Inc., Hayward, CA, USA).

157 *Cardiovascular variables:* Beat-to-beat mean arterial blood pressure (MAP) was measured 158 from an arterial catheter inserted in a radial artery, and connected to a calibrated, fluid-filled, 159 disposable pressure transducer positioned at the level of the heart (DELTRAN II, Utah Medical, Salt 160 Lake City, UT, USA). Heart rate (HR) was determined with a three-lead ECG (ADInstruments 161 BioAmp & Micromaxx, SonoSite Inc., Bothell, WA, USA). Cerebrovascular conductance index (CVCi) 162 was calculated using the equation CVCi = MCAv/MAP and normalised to values obtained at a 163 PETCO<sub>2</sub> of 20 mmHg and expressed as percentage change.

*Respiratory variables*: VE was measured using a pneumotachograph (Universal Ventilation
Meter, Vacu•Med, Ventura, CA, USA; Ultima<sup>™</sup> series, Medgraphics CPX, Minneapolis, MN, USA)
and expressed in units adjusted to BTPS. PETO<sub>2</sub> and PETCO<sub>2</sub> were measured using fast responding
gas analysers (O<sub>2</sub>Cap Oxygen analyser, Oxigraf, Mountain View, CA, USA). The pneumotachograph
was calibrated using a 3-L syringe (Han-Rudolph 5530, Kansas City, KS, USA) and the gas analysers

were calibrated using gas mixtures of known concentrations of O<sub>2</sub> and CO<sub>2</sub> prior to each testingsession.

171 Arterial blood gas variables: A 20-22 gauge arterial catheter was placed into a radial artery 172 and blood samples (2 mL) were taken over approximately 5 cardiac cycle periods. Core body 173 temperature was telemetrically recorded from an ingestion pill (CorTemp, HQInc, Palmetto, FL, 174 USA). All samples were analysed immediately for arterial pH, PO<sub>2</sub> (PaO<sub>2</sub>), PCO<sub>2</sub> (PaCO<sub>2</sub>) (Rapidlab<sup>™</sup> 175 248, Siemens Healthcare Diagnostics Inc., Henkestrasse, Germany), haemoglobin concentration 176 ([Hb]) and O<sub>2</sub> saturation (SaO<sub>2</sub>) (Radiometer OSM3, Radiometer Medical ApS, Copenhagen, 177 Denmark). The blood gas values were analysed in triplicate and temperature corrected (26, 53). 178 Arterial bicarbonate concentration ([HCO<sub>3</sub>]) was subsequently calculated using the Henderson-179 Hasselbalch equation.

180

181 Data acquisition

182 All analog data were sampled and recorded at 200Hz on a PC for off-line analysis 183 (ADInstruments Powerlab 16/30, Bella Vista, Australia).

184

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185 Data analysis
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186 Steady-state responses

Since the subjects could not tolerate PETCO<sub>2</sub> clamping at 50 mmHg at ALT16, the steadystate MCAv-CO<sub>2</sub>, MAP-CO<sub>2</sub> and CVCi-CO<sub>2</sub> slopes were estimated from the difference in mean MCAv, MAP and CVCi at the end of 20 and 40 mmHg PETCO<sub>2</sub> clamp (20 sec averages), plotted against the change in PaCO<sub>2</sub> between these two conditions across all time points (SL, ALT1, ALT16, POST7 and POST21). The absolute value of VE at clamp 40 mmHg was used as an estimate of steady-state VE responsiveness to CO<sub>2</sub>, since voluntary hyperventilation was necessary to reduce PETCO<sub>2</sub> to 20 mmHg.

## 195 Modified rebreathing

196 The rebreathing data were first reduced to one-second averages across the entire 197 rebreathing period. The  $VE-CO_2$  slopes were analysed using a specially-designed programme 198 (Analyse VE Rebreathing programme rev11, University of Toronto, Toronto, Canada), as previously 199 described (15, 16, 34). The MCAv-CO<sub>2</sub> slopes were analysed using a commercially available 200 graphing programme (Prism 5.0d, GraphPad Software Inc., San Diego, CA, USA), whereby 201 segmental linear regression (least squares fit) was used to estimate the MCAv-CO<sub>2</sub> slope during 202 the modified rebreathing. For comparison, we plotted the MCAv-CO<sub>2</sub> slopes using a sigmoid curve 203 as described by Battisti-Charbonney et al., (4), using the Prism programme. To minimise the sum 204 of squares for non-linear regression (Levenberg-Marquardt algorithm) we used the equation:

205 
$$MCAv = a + (b/(1 + exp(-(PETCO_2 - c)/d)))$$

Where MCAv is the dependent variable in cm/s,  $PETCO_2$  is the independent variable in mmHg, *a* is the minimum MCAv determined from the mean MCAv of the hypocapnic (hyperventilation) region, *b* is the maximum MCAv value, *c* is the mid-point value of MCAv, and *d* is the range of the linear portion of the sigmoid (inverse reflection of the slope of the linear portion).

We found good agreement in the MCAv-CO<sub>2</sub> slope obtained from these two models ( $R^2$ =0.71). However, due to the range of PETCO<sub>2</sub> used in this study, segmental linear regression generally provided better fit across all conditions, whereas the sigmoidal curve model was the preferred model for only 12 out of 58 trials. As such, only the MCAv-CO<sub>2</sub> slopes obtained using the segmental linear model are presented.

215

#### 216 Statistical analysis

217 Due to logistics impacts on planning and transportation, not all subjects were able to 218 participate in all high-altitude studies, please see the Figures and Table for complete sample size

219 reporting for each procedure. Most data are reported as the improvement over the time of 220 acclimatization (change from ALT1 to ALT16) and as the amount of that improvement that was 221 retained after time at low altitude, calculated as % retention = (POST7 or POST21 – ALT1)/(ALT16 222 – ALT1)\*100 (5). The effects of altitude acclimatisation and re-exposure (between SL, ALT1, ALT16, 223 POST7 and POST21) on the steady-state MCAv-CO<sub>2</sub> slope, CVCi-CO<sub>2</sub> slopes and VE at 40 mmHg, 224 were analysed using mixed model linear regression (IBM® SPSS® Statistics version 21, IBM® 225 Corporation, Armonk, NY, USA). To assess the effects of altitude acclimatisation (between SL, ALT1 226 and ALT16) on the rebreathing estimates of MCAv-CO<sub>2</sub> and  $\dot{V}$ E-CO<sub>2</sub> slopes, we used mixed model 227 linear regression analysis (Diagonal repeated covariance assumed). The interactions between 228 variables of interest were assessed using correlational (Pearson) analysis (IBM® SPSS®, Statistics 229 version 21). Data are shown as mean  $\pm$  SD. Results were considered significant at the alpha level 230 <0.05. Trends were consider at the alpha <0.10 level. A priori power calculations ( $\alpha$  = 0.05,  $\beta$  = 231 0.20) were used to determine sample size and limit Type II error.

232

233

#### 234 <u>Results</u>

Detailed baseline characteristics of the 21 (9 females; 21 ± 1 years old) subjects participating in AltitudeOmics are presented elsewhere (54). All 21 subjects completed the protocol at SL. Due to logistical issues, 4 of 21 subjects were unable to complete the entire experimental protocol at ALT1. Upon re-exposure to altitude, 14 of 14 subjects completed the protocol at POST7 and 5 of 7 at POST21. Due to low n, no comparison was carried out between ALT1 and POST21

241

242 Resting variables

The resting variables across acclimatisation and re-exposure have already been reported in
detail elsewhere (54) and will not be reproduced in this paper.

245

## 246 Steady-state method (Table 1)

Acclimatisation: Compared to SL, the steady-state MCAv-CO<sub>2</sub> slope was elevated at ALT1 (by 79  $\pm$  70%, P<0.001), and was further elevated at ALT16 (by 89  $\pm$  70% vs. ALT1, P=0.001). Similarly, the steady-state MAP-CO<sub>2</sub> slope was elevated at ALT1 (by 256  $\pm$  265%, P=0.013) and further elevated at ALT16 (by 164  $\pm$  1370% vs. ALT1, P<0.001). The steady-state CVCi-CO<sub>2</sub> slope was elevated at ALT1 (by 82  $\pm$  79%, P<0.001), and remained higher at ALT16 (by 93  $\pm$  81%, P<0.001 vs. SL, no difference with ALT1). VE at 40 mmHg was elevated at ALT1 compared to SL (by 14.8  $\pm$ 

253 12.3 L/min, P=0.011), and further elevated at ALT16 (by 48.3 ± 32.0 L/min vs. ALT1, P<0.001).

254 Re-exposure: Upon re-exposure to altitude, it appears that the acclimatisation gained in the 255 steady-state MCAv-CO<sub>2</sub> slope was not retained at POST7 (P=0.145 vs. ALT1). Compared to ALT16, 256 the steady-state MCAv-CO<sub>2</sub> slope was lowered at both POST7 and POST21 (P=0.029 & P=0.003, 257 respectively), but nevertheless remained higher compared to SL (P<0.001 & P=0.024, 258 respectively). Similarly, 49% of the acclimatisation gained in the MAP-CO<sub>2</sub> slope was retained at 259 POST7. Specifically, the MAP-CO<sub>2</sub> slope remained higher at POST7 compared to ALT1 (P=0.005). 260 When compared to ALT16, the MAP-CO<sub>2</sub> slope was lowered at both POST7 and POST21 (P<0.001 261 for both). Nevertheless, MAP-CO<sub>2</sub> slope were higher at POST7 and POST21 compared to SL 262 (P<0.001 & P=0.020, respectively). In contrast, no difference was observed in the CVCi-CO<sub>2</sub> slope 263 at POST7 when compared to ALT1 or ALT16 (P=0.980 & P=0.804, respectively), but it remained 264 higher compared to SL (P<0.001). Likewise, CVCi-CO<sub>2</sub> slope tended to remain higher at POST21 265 compared to SL (P=0.058), but was not different from ALT16 (P=0.715).

266 Upon re-exposure, the effect of acclimatisation on the VE at 40 mmHg was retained by 267 38% at POST7 (P=0.004 vs. ALT1). Compared to ALT16, VE at 40 mmHg was lower at POST7 and

POST21 (P=0.001 & P<0.001, respectively), but these values remained higher when compared to</li>
SL (P<0.001 & P=0.001, respectively).</li>

- 270
- 271 Modified rebreathing method (Table 1)

Similar to the steady-state method, the rebreathing MCAv-CO<sub>2</sub> slope was elevated at ALT1 (by 137 ± 117%, P<0.001), and further elevated at ALT16 (by 35 ± 33% vs. ALT1, P=0.040). The rebreathing  $\dot{V}E$ -CO<sub>2</sub> slope was elevated at ALT1 compared to SL (by 1.61 ± 1.14 L/min/mmHg, P=0.038), and further elevated at ALT16 (by 2.86 ± 2.61 L/min/mmHg vs. ALT1, P=0.004). The ventilatory recruitment threshold was lowered at ALT1 (by 4.4 ± 4.0 mmHg, P<0.001 vs. SL) and further lowered at ALT16 (by 4.4 ± 3.2 mmHg vs. ALT1, P<0.001).

278

279 Acid-base buffering capacity correlations (Figure 2)

Based on previous findings (16), we performed correlations between the pooled steadystate data with  $[HCO_3^-]$  and found resting  $[HCO_3^-]$  correlated with steady-state MCAv-CO<sub>2</sub> slope (R=-0.771) and  $\dot{V}E$  at 40 mmHg (R=-0.723, P<0.001 for both).

283

284 Steady-state vs. modified rebreathing (Figure 3)

We observed correlations between the steady-state and rebreathing MCAv-CO<sub>2</sub> slope at SL (R=0.609, P=0.003), ALT1 (R=0.817, P<0.001) and ALT16 (R=0.596, P=0.007), while the pooled MCAv-CO<sub>2</sub> slopes (combined SL, ALT1 and ALT16) between the two methods also correlated well (R=0.860, P<0.001). Likewise, there were significant correlations between  $\dot{V}E$  at 40 mmHg and the rebreathing  $\dot{V}E$ -CO<sub>2</sub> slope at SL (R=0.476, P=0.029), ALT1 (R=0.506, P=0.038) and ALT16 (R=0.927, P<0.001), while the pooled ventilatory data across all time points were also correlated (R=0.904, P<0.001).

## 294 Discussion

295 The present study is the first to assess the effect of altitude acclimatisation and re-296 exposure on cerebrovascular  $CO_2$  reactivity using both the steady-state and modified rebreathing 297 methods. We demonstrate that cerebrovascular  $CO_2$  reactivity was elevated immediately upon 298 arrival to 5,260m and is further elevated following 16 days acclimatisation, regardless of the 299 method of assessment. In addition, we found that cerebrovascular and ventilatory responsiveness 300 to  $CO_2$  remains elevated upon re-exposure to altitude, despite 7 or 21 days at low altitude. Since 301 these changes in cerebrovascular and ventilatory responsiveness to CO<sub>2</sub> correlated with the 302 changes in resting arterial [HCO<sub>3</sub>] across all time points, we speculate that these changes might be 303 partly due to an altered pH buffering capacity associated to exposure high altitude. Our data thus 304 demonstrate that the changes in cerebrovascular and ventilatory control gained due to altitude 305 acclimatisation over a period of 16 days are partially preserved upon subsequent exposure to 306 altitude, at least for up to a period of 3 weeks spent at low altitude.

307

#### 308 Effects of acclimatisation on cerebrovascular CO<sub>2</sub> reactivity

309 Our findings extend those from Fan et al., (16, 17) by demonstrating that the MCAv-CO<sub>2</sub> 310 slope is elevated upon arrival at 5,260 m and further elevated following 16 days of acclimatisation 311 (Fig. 1A). Importantly, previous studies by Fan et al., (16, 17) assessed MCAv-CO<sub>2</sub> slope in subjects 312 whom spent 8 days ascending to 5,050 m, while the subjects in the present study ascended rapidly 313 to altitude (•3 hours), thus making direct comparison difficult. Our findings contradict those of 314 Lucas et al., (30), who found that the MCAv-CO<sub>2</sub> slope was initially elevated at 5,050 m, but had 315 returned towards sea level values following two weeks at 5,050 m. However, because PETO<sub>2</sub> was 316 not controlled, the MCAv-CO<sub>2</sub> slopes reported by Lucas et al., (30) reflect MCAv changes from 317 hypoxic hypocapnia (room air breathing at 5,050 m; PETO<sub>2</sub> •48 mmHg & PETCO<sub>2</sub> 26-22 mmHg) to

318 hypercapnic hyperoxia (PETO<sub>2</sub> > 310 mmHg & PETCO<sub>2</sub>  $\bullet$  30 mmHg), and thus do not represent 319 isolated reactivity to  $CO_2$ . Rupp et al., (49) recently found the MCAv response to steady-state 320 hypoxic hypercapnia (PETO<sub>2</sub> = 55 mmHg) to be reduced following 5 days at 4,350 m. Therefore, 321 discrepancies between findings Rupp et al., (49) and those of the present study can be attributed 322 the differences in PETO<sub>2</sub> (55 mmHg vs. >200 mmHg), altitude (4,350 m vs. 5,260 m), and the 323 acclimatisation state of the subjects (5 days vs. 16 days). The results from the present study 324 demonstrate, for the first time, that cerebrovascular  $CO_2$  reactivity per se is enhanced with 325 acclimatisation to high altitude when studied using a background level of hyperoxia. Furthermore, 326 discrepancy between studies highlights how methodological differences can yield vastly different 327 results. Thus future studies are warranted to clarify the effect of hypoxic and hyperoxic 328 background on assessing cerebrovascular functions at both sea-level and following ascent to high 329 altitude.

330

## 331 Altered acid-base buffering capacity?

332 During altitude acclimatisation, there is a progressive and parallel reduction in arterial and 333 cerebrospinal fluid (CSF) bicarbonate concentration which serves to compensate for the changes 334 in pH associated with hyperventilation-induced hypocapnia (12, 13, 20). These changes in acid-335 base buffering capacity, in both the arterial and CSF compartments, would lead to a greater rise in 336 arterial and CSF  $[H^{\dagger}]$  for a given rise in PaCO<sub>2</sub>. In support of this notion, lowering CSF bicarbonate 337 concentration elevates the cerebrovascular  $CO_2$  reactivity in an anaesthetised dog model (27), 338 while bicarbonate infusion increases cerebral perfusion pressure in post-traumatic head injury 339 patients (9), elevates cerebral blood volume in preterm infants (57), and lowers ventilation in 340 healthy exercising humans at sea-level (44). As such, it has been suggested that the MCAv 341 responses to  $CO_2$  at high altitude are linked to changes in arterial acid-base balance (16, 25). In the 342 present study, we observed concomitant increases in cerebrovascular and ventilatory

343 responsiveness to  $CO_2$  with acclimatisation to high altitude and re-exposure (Fig. 1), which 344 occurred in parallel to the changes in  $[HCO_3^-]$  (Fig. 2). While it should be acknowledged that such 345 correlations do not imply causality, the possible role for acid-base status changes on 346 cerebrovascular and ventilatory responsiveness to  $CO_2$  at high altitude remains to be further 347 studied.

348

#### 349 Interaction between cerebrovascular and ventilatory responsiveness to CO<sub>2</sub>

350 Interaction between cerebrovascular CO<sub>2</sub> reactivity and the central chemoreceptor 351 activation was first alluded to by Heyman et al., (22) and has been subsequently expanded upon 352 by others (10, 16-18, 38, 43, 60-62). It was postulated that changes in cerebrovascular  $CO_2$ 353 reactivity affect the stability of ventilatory response to  $CO_2$  by modulating the degree of H<sup>+</sup> 354 washout at the level of the central chemoreceptor (38). Accordingly, a blunted cerebrovascular 355  $CO_2$  reactivity would lead to less central H<sup>+</sup> washout and subsequently greater central 356 chemoreceptor activation. Conversely, an enhanced cerebrovascular CO<sub>2</sub> reactivity would result in 357 lower central  $[H^+]$  and therefore lower ventilatory  $CO_2$  sensitivity. In agreement with previous 358 altitude studies (16, 17), we observed concomitant increases cerebrovascular and ventilatory 359 responsiveness to  $CO_2$  (Fig. 1). These findings seem to contradict the modulating role of 360 cerebrovascular CO<sub>2</sub> reactivity on central chemoreceptor activation, possibly due to other 361 overriding factors such as enhanced central chemosensitivity and changes in acid-base balance 362 associated with ascent to high altitude. Future work is necessary to further unravel the interaction 363 between the regulation of cerebral blood flow and ventilation.

364

365 Going back up

366 Despite the large body of literature regarding high altitude acclimatisation over the past 367 century, the effect of prior exposure on physiological parameters during subsequent exposures is

368 not well documented. Most attention focused on the effect of a recent altitude exposure on the 369 risk for AMS (7, 31, 45, 51), or the rate of ascent (56). However, the dose of previous altitude 370 exposure and acclimatisation were generally not controlled in these studies. Wu et al., (59) found 371 a progressive reduction in the incidence of AMS, lower HR and higher SpO<sub>2</sub> in lowland railroad 372 workers over the course of several seven-month exposures to high altitude interspersed with 5 373 months spent at low altitude. Similarly, MacNutt et al., (32) found faster rate of ascent, lower AMS 374 and higher SpO<sub>2</sub> in trekkers with a recent altitude exposure compared to altitude naive trekkers, 375 despite a 7-30 day de-acclimatisation period. In the present study, we compared the 376 cerebrovascular and ventilatory responsiveness to CO<sub>2</sub> with acclimatisation and upon re-exposure 377 to 5,260 m following a period of either 7 or 21 days at low altitude. We found that 38% of the gain 378 in ventilatory response to  $CO_2$  over acclimatisation was retained at POST7 (Fig. 1C), while 379 essentially none of the gain in MCAv-CO<sub>2</sub> reactivity over acclimatisation was retained at POST7 380 (Fig. 1A). Regardless of the underpinning mechanism(s), our findings suggest that the effect of 381 previous altitude acclimatisation over 16 days on ventilatory response to  $CO_2$  is partially retained 382 after 7 days at low altitude, while it is reversed in the cerebrovascular response to CO<sub>2</sub>. Our data 383 extends those by Muza et al., (36) which showed that ventilatory acclimatisation gained at 4,300 384 m is retained following 8 days spent at low altitude. Since we found the CVCi-CO<sub>2</sub> slope to be 385 consistently elevated by 60-80% across all time points (Fig. 1D), while the changes MAP-CO<sub>2</sub> slope 386 closely follows the changes in MCAv-CO<sub>2</sub> slopes (Fig. 1B), we speculate that the changes in MCAv-387  $CO_2$  slopes at high altitude can be primarily accounted for by an enhanced sensitivity of the 388 cerebral vessels to  $CO_2$ , whereas the remainder can be attributed to an enhanced perfusion 389 pressure response.

390

391 Steady-state or modified rebreathing method?

392 There has been much debate over the use of the steady-state or the modified rebreathing 393 method for the assessment of cerebrovascular and ventilatory control, and attempts at consensus 394 have produced no uniform agreement [(18, 40), also see (2, 14) for reviews]. The steady-state 395 ventilatory responses to  $CO_2$  were found to be either similar (34, 37, 40-42, 47) or lower (6, 18, 23, 396 55) when compared to rebreathing estimates., while steady-state cerebrovascular  $CO_2$  reactivity 397 has been shown to be consistently higher than rebreathing values (18, 40-42). The present study 398 demonstrates that the changes in cerebrovascular and ventilatory CO2 responsiveness with 399 altitude acclimatisation were similar between the steady-state and the modified rebreathing 400 method (Table 1) - possibly due to tight control of arterial PCO<sub>2</sub> and PO<sub>2</sub> with our end-tidal 401 clamping setup. Moreover, we observed strong correlations in these parameters between the two 402 methods across all time points (Fig. 3). We therefore conclude that both methods can be used to 403 assess the changes in cerebrovascular and ventilatory responses to  $CO_2$  with high altitude 404 exposure and acclimatisation, provided that the level of CO2 is comparable across all the 405 conditions, under identical level of background O<sub>2</sub>.

406

#### 407 Limitations

408 Although the present study provided the opportunity to assess the effects of 409 acclimatisation and re-exposure to 5,260 m on the cerebrovascular  $CO_2$  reactivity, an important 410 methodological consideration should be acknowledged when interpreting our findings. In the 411 present study, transcranial Doppler ultrasound (TCD) was used to measure the MCAv, as an index 412 of global CBF changes during initial exposure, acclimatisation and subsequent re-exposure to 413 5,260 m. This is based on the assumption that: i) the MCA carries approximately upwards of 80% 414 of the overall blood flow to the respective hemisphere (29); ii) changes in MCAv reflect changes in 415 global CBF (8, 52); iii) the changes in MCAv in response to  $PaCO_2$  changes are comparable to the 416 changes of internal carotid blood flow (50); and iii) the diameter of the MCA does not change during the observed changes in arterial blood gases (52). In support, MCAv has been shown to
reflect changes in CBF assessed with the direct Fick method, at least during initial exposure to high
altitude (33, 35, 48).

420 Recent findings by Wilson et al., (58) indicate that the diameter of MCA, as measured using 421 TCD, vary, depending on the altitude (e.g., 5.30 mm at 75 m, 5.51 mm at 3,500 m, 5.23 mm at 422 5,300 m and 9.34 mm at 7,950 m). Importantly, the results from Wilson et al., (58) demonstrate 423 that the MCA diameter remains relatively unchanged up to 5,300 m. It should be noted that the 424 MCA diameters measured with TCD in that study were 80-90% greater than the values obtained 425 using magnetic resonance imaging in the same subjects. Since our measurements were carried out 426 in background hyperoxia (PETCO<sub>2</sub> > 300 mmHg), it seems unlikely that our cerebral blood velocity 427 values would be confounded by any effect of hypoxia-induced vasodilation of the MCA. Further 428 studies are needed to evaluate MCAv responses to  $CO_2$  while holding PETO<sub>2</sub> at consistent levels of 429 hypoxia.

430

431 Conclusion

Findings from the present study clearly show that both cerebrovascular and ventilatory responsiveness to  $CO_2$  is elevated upon arrival at high altitude and further elevated with acclimatisation. We demonstrate, for the first time, that this effect of high altitude acclimatisation on the ventilatory response to  $CO_2$  is partially retained after a period at low altitude, while prior acclimatisation has no effect of the cerebrovascular response to  $CO_2$ . Our data suggest that the increased cerebrovascular  $CO_2$  reactivity with acclimatisation may be accounted for by the changes in acid-base balance in the blood and possibly the cerebrospinal fluid compartment.

439

440

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448

#### 449 Author contributions

450 JF contributed to conception and design of the experiments, data collection, data analysis, 451 data interpretation, manuscript drafting and editorial process. OE contributed to the design of the 452 experiments, data collection, data analysis and manuscript revision. NB contributed to the data 453 collection and the manuscript revision. BK contributed to the interpretation of the data and the 454 revision of the manuscript. AL, AS and RR conceived, designed and executed the AltitudeOmics 455 study of which the present study was a part, and contributed to manuscript revision. AS also 456 contributed to data collection and data interpretation. All authors approved the final version of 457 the manuscript.

458

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#### 474 **Reference**

- Ainslie PN, and Burgess KR. Cardiorespiratory and cerebrovascular responses to hyperoxic
   and hypoxic rebreathing: Effects of acclimatization to high altitude. *Respir Physiol Neurobiol* 161: 201-209, 2008.
- Ainslie PN, and Duffin J. Integration of cerebrovascular CO<sub>2</sub> reactivity and chemoreflex
   control of breathing: mechanisms of regulation, measurement, and interpretation. *Am J Physiol Regul Integr Comp Physiol* 296: R1473-1495, 2009.
- 481 3. Ainslie PN, and Ogoh S. Regulation of cerebral blood flow during chronic hypoxia: a matter
  482 of balance. *Exp Physiol 95*: 251-262, 2009.
- 483 4. Battisti-Charbonney A, Fisher J, and Duffin J. The cerebrovascular response to carbon
  484 dioxide in humans. *J Physiol* 589: 3039-3048, 2011.
- 485 5. Beidleman BA, Muza SR, Rock PB, Fulco CS, Lyons TP, Hoyt RW, and Cymerman A.

486 Exercise responses after altitude acclimatization are retained during reintroduction to 487 altitude. *Med Sci Sports Exerc* 29: 1588-1595, 1997.

- Berkenbosch A, Bovill JG, Dahan A, DeGoede J, and Olievier IC. The ventilatory CO<sub>2</sub>
  sensitivities from Read's rebreathing method and the steady-state method are not equal in
  man. J Physiol 411: 367-377, 1989.
- 491 7. Bircher HP, Eichenberger U, Maggiorini M, Oelz O, and Bärtsch P. Relationship of
   492 mountain sickness to physical fitness and exercise intensity during ascent. J Wilderness
   493 Med 5: 302-311, 1994.
- Bishop CC, Powell S, Rutt D, and Browse NL. Transcranial Doppler measurement of middle
  cerebral artery blood flow velocity: a validation study. *Stroke* 17: 913-915, 1986.
- 496 9. Bourdeaux C, and Brown J. Sodium bicarbonate lowers intracranial pressure after
  497 traumatic brain injury. In: *Neurocrit Care* 13: 24-28, 2010.

- 10. Chapman RW, Santiago TV, and Edelman NH. Effects of graded reduction of brain blood
- flow on chemical control of breathing. *J Appl Physiol* 47: 1289-1294, 1979.
- 500 11. Cunningham DJC, Lloyd BB, and Patrick JM. The relationship between ventilation and end-
- 501 tidal PCO<sub>2</sub> in man during moderate exercise with and without CO<sub>2</sub> inhalation. *J Physiol* 169:
- 502 104-106, 1963.
- 503 12. Dempsey JA, Forster HV, and DoPico GA. Ventilatory Acclimatization to Moderate
   504 Hypoxemia in Man. *J Clin Invest* 53: 1091-1100, 1974.
- 505 13. Dempsey JA, Forster HV, Gledhill N, and DoPico GA. Effects of moderate hypoxemia and
   506 hypocapnia on CSF [H<sup>+</sup>] and ventilation in man. *J App Physiol* 38: 665-674, 1975.
- 507 14. Duffin J. Measuring the respiratory chemoreflexes in humans. *Respir Physiol Neurobiol*508 2011.
- 509 15. **Duffin J, Mohan RM, Vasiliou P, Stephenson R, and Mahamed S**. A model of the 510 chemoreflex control of breathing in humans: model parameters measurement. *Respir* 511 *Physiol* 120: 13-26, 2000.
- Fan JL, Burgess KR, Basnyat R, Thomas KN, Peebles KC, Lucas SJ, Lucas RA, Donnelly J,
   Cotter JD, and Ainslie PN. Influence of high altitude on cerebrovascular and ventilatory
   responsiveness to CO<sub>2</sub>. *J Physiol* 588: 539-549, 2010.
- 515 17. Fan JL, Burgess KR, Thomas KN, Lucas SJ, Cotter JD, Kayser B, Peebles KC, and Ainslie PN.
- 516 Effects of acetazolamide on cerebrovascular function and breathing stability at 5050 m. J
- 517 *Physiol* 590: 1213-1225, 2012.
- 518 18. Fan JL, Burgess KR, Thomas KN, Peebles KC, Lucas SJ, Lucas RA, Cotter JD, and Ainslie PN.
- 519 Influence of indomethacin on ventilatory and cerebrovascular responsiveness to CO<sub>2</sub> and
- 520 breathing stability: the influence of PCO<sub>2</sub> gradients. *Am J Physiol Regul Integr Comp Physiol*
- 521 298: R1648-1658, 2010.

- Faraci FM, and Heistad DD. Regulation of large cerebral arteries and cerebral
   microvascular pressure. *Circ Res* 66: 8-17, 1990.
- 524 20. Forster HV, Dempsey JA, and Chosy LW. Incomplete compensation of CSF [H<sup>+</sup>] in man
   525 during acclimatization to high altitude (48300 M). *J Appl Physiol* 38: 1067-1072, 1975.
- 526 21. Gardner WN. The pattern of breathing following step changes of alveolar partial pressures
  527 of carbon dioxide and oxygen in man. *J Physiol* 300: 55-73, 1980.
- 528 22. **Heyman A, Birchfield RI, and Sieker HO**. Effects of bilateral cerebral infarction on 529 respiratory center sensitivity. *Neurology* 8: 694-700, 1958.
- Jacobi MS, Patil CP, and Saunders KB. Transient, steady-state and rebreathing responses
  to carbon dioxide in man, at rest and during light exercise. *J Physiol* 411: 85-96, 1989.
- 532 24. Jansen GF, Krins A, and Basnyat B. Cerebral vasomotor reactivity at high altitude in 533 humans. *J Appl Physiol* 86: 681-686, 1999.
- 534 25. Jensen JB, Sperling B, Severinghaus JW, and Lassen NA. Augmented hypoxic cerebral 535 vasodilation in men during 5 days at 3,810 m altitude. *J Appl Physiol* 80: 1214-1218, 1996.
- 536 26. Kelman GR, and Nunn JF. Nomograms for correction of blood PO<sub>2</sub>, PCO<sub>2</sub>, pH, and base
  537 excess for time and temperature. *J Appl Physiol* 21: 1484-1490, 1966.
- 538 27. **Koehler RC, and Traystman RJ**. Bicarbonate ion modulation of cerebral blood flow during 539 hypoxia and hypercapnia. *Am J Physiol* 243: H33-40, 1982.
- 540 28. Kolb JC, Ainslie PN, Ide K, and Poulin MJ. Effects of five consecutive nocturnal hypoxic
- 541 exposures on the cerebrovascular responses to acute hypoxia and hypercapnia in humans.
- 542 *J Appl Physiol* 96: 1745-1754, 2004.
- Lindegaard KF, Lundar T, Wiberg J, Sjoberg D, Aaslid R, and Nornes H. Variations in middle
   cerebral artery blood flow investigated with noninvasive transcranial blood velocity
   measurements. *Stroke* 18: 1025-1030, 1987.

- 546 30. Lucas SJE, Burgess KR, Thomas KN, Donnelly J, Peebles KC, Lucas RAI, Fan JL, Cotter JD,
- 547 **Basnyat R, and Ainslie PN**. Alterations in cerebral blood flow and cerebrovascular 548 reactivity during 14 days at 5050 m. *J Physiol* 589: 741-753, 2011.
- 54931.Lyons TP, Muza SR, Rock PB, and Cymerman A. The effect of altitude pre-acclimatization550on acute mountain sickness during reexposure. Aviat Space Environ Med 66: 957-962,
- 551 1995.
- MacNutt MJ, Laursen PB, Kedia S, Neupane M, Parajuli P, Pokharel J, and Sheel AW.
  Acclimatisation in trekkers with and without recent exposure to high altitude. *Eur J Appl Physiol* 112: 3287-3294, 2012.
- 555 33. **Milledge JS, and Sorensen SC**. Cerebral arteriovenous oxygen difference in man native to 556 high altitude. *J Appl Physiol* 32: 687-689, 1972.
- Mohan RM, Amara CE, Cunningham DA, and Duffin J. Measuring central-chemoreflex
  sensitivity in man: rebreathing and steady-state methods compared. *Respir Physiol* 115:
  23-33, 1999.
- 560 35. **Møller K, Paulson OB, Hornbein TF, Colier WN, Paulson AS, Roach RC, Holm S, and** 561 **Knudsen GM**. Unchanged cerebral blood flow and oxidative metabolism after 562 acclimatization to high altitude. *J Cereb Blood Flow Metab* 22: 118-126, 2002.
- 36. Muza SR, Fulco CS, Lyons T, Rock PB, Beidleman BA, Kenney J, and Cymerman A.
  Augmented chemosensitivity at altitude and after return to sea level: impact on
  subsequent return to altitude. *Acta Andina* 4: 109-112, 1995.
- 37. Nickol AH, Dunroy H, Polkey MI, Simonds A, Cordingley J, Corfield DR, and Morrell MJ. A
  quick and easy method of measuring the hypercapnic ventilatory response in patients with
  COPD. *Respir Med* 103: 258-267, 2009.

- 569 38. **Ogoh S, Hayashi N, Inagaki M, Ainslie PN, and Miyamoto T**. Interaction between the 570 ventilatory and cerebrovascular responses to hypo- and hypercapnia at rest and during 571 exercise. *J Physiol* 586: 4327-4338, 2008.
- 572 39. **Olin JT, Dimmen AC, Subudhi AW, and Roach RC**. A simple method to clamp end-tidal 573 carbon dioxide during rest and exercise. In: *Eur J Appl Physiol*2012, p. 3439-3444.
- 40. Pandit JJ, Mohan RM, Paterson ND, and Poulin MJ. Cerebral blood flow sensitivities to CO<sub>2</sub>
- 575 measured with steady-state and modified rebreathing methods. *Respir Physiol Neurobiol* 576 159: 34-44, 2007.
- 577 41. Pandit JJ, Mohan RM, Paterson ND, and Poulin MJ. Cerebral blood flow sensitivities to CO<sub>2</sub>
- 578 with the steady-state method and Read's rebreathing method. *Adv Exp Med Biol* 499: 279-
- 579284, 2001.
- Pandit JJ, Mohan RM, Paterson ND, and Poulin MJ. Cerebral blood flow sensitivity to CO<sub>2</sub>
  measured with steady-state and Read's rebreathing methods. *Respir Physiol Neurobiol* 137:
  1-10, 2003.
- 43. Peebles K, Celi L, McGrattan K, Murrell C, Thomas K, and Ainslie PN. Human
  cerebrovascular and ventilatory CO<sub>2</sub> reactivity to end-tidal, arterial and internal jugular
  vein PCO2. J Physiol 584: 347-357, 2007.
- 586 44. Péronnet F, and Aguilaniu B. Lactic acid buffering, nonmetabolic CO<sub>2</sub> and exercise
   587 hyperventilation: a critical reappraisal. *Respir Physiol Neurobiol* 150: 4-18, 2006.
- 45. Pesce C, Leal C, Pinto H, González G, Maggiorini M, Schneider M, and Bärtsch P.
  Determinants of acute mountain sickness and success on Mount Aconcagua (6962 m). *High*Alt Med Biol 6: 158-166, 2005.
- 46. Poulin MJ, Fatemian M, Tansley JG, O'Connor DF, and Robbins PA. Changes in cerebral
  blood flow during and after 48 h of both isocapnic and poikilocapnic hypoxia in humans. *Exp Physiol* 87: 633-642, 2002.

- 47. **Read DJ**. A clinical method for assessing the ventilatory response to carbon dioxide.
  595 *Australas Ann Med* 16: 20-32, 1967.
- 596 48. Roy SB, Guleria JS, Khanna PK, Talwar JR, Manchanda SC, Pande JN, Kaushik VS, Subba
- 597 **PS, and Wood JE**. Immediate circulatory response to high altitude hypoxia in man. *Nature*
- 598217: 1177-1178, 1968.
- 49. Rupp T, Esteve F, Bouzat P, Lundby C, Perrey S, Levy P, Robach P, and Verges S. Cerebral
  hemodynamic and ventilatory responses to hypoxia, hypercapnia, and hypocapnia during 5
  days at 4,350 m. *J Cereb Blood Flow Metab* 2013.
- 50. Sato K, Sadamoto T, Hirasawa A, Oue A, Subudhi AW, Miyazawa T, and Ogoh S.
   Differential blood flow responses to CO<sub>2</sub> in human internal and external carotid and
- 604 vertebral arteries. *J Physiol* 590: 3277-3290, 2012.
- 60551.Schneider M, Bernasch D, Weymann J, Holle R, and Bärtsch P. Acute mountain sickness:606influence of susceptibility, preexposure, and ascent rate. *Med Sci Sports Exerc* 34: 1886-6071001 2000
- 6071891, 2002.
- Serrador JM, Picot PA, Rutt BK, Shoemaker JK, and Bondar RL. MRI measures of middle
  cerebral artery diameter in conscious humans during simulated orthostasis. *Stroke* 31:
  1672-1678, 2000.
- 611 53. Severinghaus JW. Blood gas calculator. *J Appl Physiol* 21: 1108-1116, 1966.
- 612 54. Subudhi AW, Bourdillon N, Bucher J, Davis C, Eillot J, Eutermoster M, Evero O, Fan JL,
- 613 Jameson-Van Houton S, Julian CG, Kark J, Kark S, Kern J, Kayser B, Kim SE, Lathan C,
- 614 Laurie SS, Lovering AT, Paterson R, Polaner D, Ryan BJ, Spira J, Tsao J, Wachsmuth NB,
- and Roach RC. AltitudeOmics: The intergrative physiology of human acclimatization to
   hypobaric hypoxia and its memory on reascent. *Proc Natl Acad Sci* In review.
- 55. Tenney SM, Remmers JE, and Mithoefer JC. Interaction of CO<sub>2</sub> and hypoxic stimuli on
  ventilation at high altitude. *Q J Exp Physiol Cogn Med Sci* 48: 192-201, 1963.

- 619 56. Tsianos G, Woolrich-Burt L, Aitchison T, Peacock A, Watt M, Montgomery H, Watt I, and
- 620 Grant S. Factors affecting a climber's ability to ascend Mont Blanc. *Eur J Appl Physiol* 96:
  621 32-36, 2006.
- 622 57. van Alfen-van der Velden AAEM, Hopman JCW, Klaessens JHGM, Feuth T, Sengers RCA,
- and Liem KD. Effects of rapid versus slow infusion of sodium bicarbonate on cerebral
   hemodynamics and oxygenation in preterm infants. *Biol Neonate* 90:122-127, 2006.
- 625 58. Wilson MH, Edsell MEG, Davagnanam I, Hirani SP, Martin DS, Levett DZH, Thornton JS,
- 626 Golay X, Strycharczuk L, Newman SP, Montgomery HE, Grocott MPW, and Imray CHE.
- 627 Cerebral artery dilatation maintains cerebral oxygenation at extreme altitude and in acute
- hypoxia; an ultrasound and MRI study. *J Cereb Blood Flow Metab* 31: 2019-2029, 2011.
- 629 59. Wu TY, Ding SQ, Liu JL, Yu MT, Jia JH, Duan JQ, Chai ZC, Dai RC, Zhang SL, Liang BZ, Zhao
- 630 JZ, Qi DT, Sun YF, and Kayser B. Reduced Incidence and Severity of Acute Mountain
- 631 Sickness in Qinghai–Tibet Railroad Construction Workers after Repeated 7-Month
- 632 Exposures despite 5-Month Low Altitude Periods. *High Alt Med Biol* 10: 221-232, 2009.
- 633 60. Xie A, Skatrud JB, Barczi SR, Reichmuth K, Morgan BJ, Mont S, and Dempsey JA. Influence
  634 of cerebral blood flow on breathing stability. *J Appl Physiol* 106: 850-856, 2009.
- 635 61. Xie A, Skatrud JB, Khayat R, Dempsey JA, Morgan B, and Russell D. Cerebrovascular
  636 response to carbon dioxide in patients with congestive heart failure. *Am J Respir Crit Care*637 *Med* 172: 371-378, 2005.
- ,
- 638 62. Xie A, Skatrud JB, Morgan B, Chenuel B, Khayat R, Reichmuth K, Lin J, and Dempsey JA.
  639 Influence of cerebrovascular function on the hypercapnic ventilatory response in healthy
  640 humans. *J Physiol* 577: 319-329, 2006.
- 641

**Table 1.** Cerebrovascular and ventilatory reactivities parameters during the steady-state and modified rebreathing (mean ± SD).
 643

	SL	ALT1	ALT16	POST7	POST21	
	(n=21)	(n=17)	(n=20)	(n=14)	(n=5)	
Steady-state						
MCAv-PaCO₂ slope (cm/s/mmHg)	1.19 ± 0.42	2.16 ± 1.05*	3.39 ± 0.89*†	2.68 ± 0.88*§	2.06 ± 0.57*§	
CVCi-PaCO <sub>2</sub> slope (%/mmHg)	3.35 ± 1.21	5.87 ± 2.60*	5.75 ± 1.85*	5.89 ± 1.23*	5.41 ± 1.78*	
MAP-PaCO <sub>2</sub> slope (L/min)	0.03 ± 0.24	0.28 ± 0.19*	1.06 ± 0.45*†	0.56 ± 0.29*§	0.32 ± 0.18*§	
└E at 40 mmHg (L∕min)	19.15 ± 4.89	34.06 ± 12.23*	80.05 ± 32.32*†	49.03 ± 13.68*§†	43.25 ± 7.56*§	
Modified rebreathing						
MCAv-PETCO₂ slope (cm/s/mmHg)	1.34 ± 0.60	2.95 ± 1.11*	3.67 ± 0.87*†	-	-	
VE-CO₂ slope (L/min/mmHg)	1.90 ± 0.81	3.49 ± 1.51*	6.28 ± 3.56*†	-	-	
VE recruitment threshold (mmHg)	38.7 ± 3.4	33.7 ± 3.7*	29.2 ± 2.1*†	-	-	

644 \* different from SL (P<0.05); + different from ALT1 (P<0.05); § different from ALT16 (P<0.05).

## 647 Figure legend

- 648 **Figure 1** Changes in steady-state estimates of cerebrovascular, cardiovascular and ventilatory
- responsiveness to CO<sub>2</sub> with acclimatisation and re-exposure to 5,260 m. Values expressed as mean
- 450 ± SD. \* different from SL (P<0.05), † different from ALT1 (P<0.05), § different from ALT16 (P<0.05).

- 652 **Figure 2** Relationship between standard basic excess and steady-state cerebrovascular, ventilatory
- and cardiovascular responsiveness to CO<sub>2</sub> with acclimatisation to altitude. \* significant
- 654 correlations (P<0.05).
- 655
- 656 **Figure 3** Comparison of steady-state and rebreathing estimates of cerebrovascular and ventilatory
- responsiveness of CO<sub>2</sub> with acclimatisation to 5,260 m. \* significant correlations (P<0.05).





Figure 2







Figure 3



AltitudeOmics: Cerebral autoregulation during ascent, acclimatization, and re-exposure to high altitude and its relation with acute mountain sickness Andrew W. Subudhi<sup>1,2</sup>, Jui-Lin Fan<sup>3,4</sup>, Oghenero Evero<sup>1</sup>, Nicolas Bourdillon<sup>3</sup>, Bengt Kavser<sup>3</sup>, Colleen G. Iulian<sup>1</sup>, Andrew T. Loverina<sup>5</sup>, Ronnev B. Panerai<sup>6</sup>, Robert C. Roach<sup>1</sup> 1. University of Colorado Altitude Research Center, Department of Emergency Medicine, Anschutz Medical Campus, Aurora, Colorado, USA. 2. University of Colorado Colorado Springs, Department of Biology, Colorado Springs, Colorado, USA 3. University of Lausanne, Institute of Sports Sciences, Lausanne, Switzerland. 4. University of Geneva, Lemanic Doctoral School of Neuroscience, Geneva, Switzerland. 5. University of Oregon, Department of Human Physiology, Eugene, Oregon, USA. 6. University of Leicester, Leicester Royal Infirmary, Department of Cardiovascular Sciences, United Kingdom Running Head: Cerebral autoregulation at altitude Key Words: transcranial Doppler, cerebral blood flow, cerebral oxygenation, transfer function analysis, hypoxia Corresponding Address: Andrew W. Subudhi Department of Biology 1420 Austin Bluffs Parkway Colorado Springs, CO 80918 Phone: 719-255-3938 Email: asubudhi@uccs.edu 

## 46 Abstract

Cerebral autoregulation (CA) acts to maintain brain blood flow despite fluctuations 47 48 in perfusion pressure. Acute hypoxia is thought to impair CA, but it is unclear if CA is 49 affected by acclimatization or related to the development of acute mountain 50 sickness (AMS). We assessed changes in CA using transfer function analysis of 51 spontaneous fluctuations in radial artery blood pressure (indwelling catheter) and 52 resulting changes in middle cerebral artery blood flow velocity (transcranial 53 Doppler) in 21 active individuals at sea level (SL), upon arrival at 5,260 m (ALT1), 54 after 16 days of acclimatization (ALT16), and upon re-exposure to 5,260m after 7 55 days at 1,525 m (POST7). The Lake Louise Questionnaire (LLQ) was used to evaluate 56 AMS symptom severity. CA was impaired upon arrival at ALT1 (P<0.001) and did 57 not change with acclimatization at ALT16 or upon re-exposure at POST7. CA was not 58 associated with AMS symptoms (all R < 0.50, P > 0.05). These findings suggest that 59 alterations in CA are an intrinsic consequence of hypoxia and are not directly 60 related to the occurrence or severity of AMS.

# 62 Introduction

63	Cerebral autoregulation (CA) is a general term used to describe dynamic myogenic,
64	neurologic and metabolic responses that adjust cerebrovascular resistance to
65	maintain relatively constant cerebral blood flow across a wide range of perfusion
66	pressures (25). Dynamic CA is said to be impaired if fluctuations in mean arterial
67	blood pressure lead to concurrent fluctuations in mean cerebral blood flow.
68	Impairments in CA are associated with cerebrovascular disorders (3, 24, 31), yet the
69	relative importance of CA in the development and course of certain pathologies is
70	unclear.
71	
72	Our initial interest in CA stemmed from the hypothesis that impaired CA may be
73	involved in the development of acute mountain sickness (AMS), high-altitude
74	headache and cerebral edema (5, 7, 9, 16, 37). Conversely, we showed that
75	impairments in CA upon acute exposure to hypobaric hypoxia preceded, but were
76	not associated with, the development of AMS (2, 33, 35). Furthermore, since several
77	cross-sectional studies demonstrated that impairments in CA persist from 1 to 30
78	days of high-altitude exposure (1, 2, 11, 12, 17) - when AMS is not present – and are
79	evident in healthy, permanent high-altitude residents (12, 13), it seems reasonable
80	to suggest that a shift in CA may be an inherent and relatively benign consequence
81	of hypoxemia.
82	

83 To date, no longitudinal studies have characterized CA and tested its relation with
84 AMS during acute and chronic high-altitude exposures. Previous studies have either

85 omitted CA measurements upon arrival at high altitude (7, 11, 17), or followed slow 86 ascent profiles that allow for partial acclimatization prior to initial measurements 87 (1, 12, 39). In this study, we present novel data from sea-level residents who rapidly 88 ascended to high altitude (5,260 m), acclimatized for 16 days, and were 89 subsequently re-exposed to high altitude after spending 7 days at low altitude 90 (1,525 m). Specifically, we tested the hypotheses that CA would be: 1) impaired 91 upon rapid ascent to high altitude, 2) unaffected by 16 days of acclimatization, 3) 92 unaffected upon re-exposure to the same altitude, and 4) unrelated to the 93 occurrence or severity of AMS.
## 95 Methods

#### 96 Study overview

97 This study was conducted as part of the AltitudeOmics project. Briefly, institutional 98 ethics approval was obtained from the Universities of Colorado and Oregon and the 99 US Department of Defense Human Research Protection Office, Young, healthy sealevel residents were recruited from the greater Eugene, Oregon area (elevation 128 100 101 m) and screened to exclude anyone who was born or had lived at altitudes >1,500 m 102 for more than one year or had traveled to altitudes > 1,000 m in the past 3 months. 103 After obtaining written consent, physical exams and the Army Physical Fitness Test 104 (push ups, sit ups and 3.2 km run) were performed to verify health and fitness status. Approximately 4 weeks following sea-level (SL) measurements in Eugene. 105 106 Oregon, subjects were flown to La Paz, Bolivia. They spent two nights at low altitude 107 (Coroico, Bolivia, 1,525 m) before being driven to the Chacaltava research station at 108 5,260 m while breathing supplemental oxygen. Acute responses to high altitude 109 were assessed  $\sim 4$  hours after arrival and cessation of supplemental oxygen (ALT1). 110 Subjects acclimatized to altitudes ranging from 3,800 to 5,260 m over the next 15 111 days, with most of the time (75%) spent at 5,250 m. On the 16th day (ALT16), 112 measurements were repeated at 5,260 m before subjects were driven down to 113 Coroico for either 7 or 21 days. Subjects were driven back to the laboratory at 5,260 114 m for POST 7 or POST 21 re-exposure measurements. 115

This report focuses on novel data regarding resting CA evaluated immediately prior
to a series of cerebrovascular, respiratory and exercise interventions, as outlined
elsewhere (32). We have carefully avoided replication of data between reports,
except where common variables were necessary to describe subjects' basic
physiologic status at the time points of interest (e.g. heart rate, blood pressure,
arterial blood gases).

122

### 123 Physiology Protocol

124 All subjects were familiarized with study procedures during a practice session at 125 least 48 hours before experimental testing at SL. Subjects followed standardized 126 exercise and dietary regimens for 24 hours prior to each measurement period. At 127 each time point, a 22-gauge catheter was inserted into a radial artery at least 1 hour 128 prior to instrumentation. Subjects were seated in an upright position for 15 min 129 while sensors were placed to measure physiologic variables of interest. Limb lead 130 electrodes were used to measure ECG (BioAmp, ADInstruments, Colorado Springs, 131 CO, USA). Arterial blood pressure (ABP) was monitored via a fluid filled pressure 132 transducer (Deltran II, Utah Medical, Salt Lake City, UT, USA) attached to the radial 133 artery catheter. Core temperature was telemetrically recorded from an ingested pill 134 (CorTemp, HQInc, Palmetto, FL, USA). Cerebral blood flow velocity (CBFv) in the left 135 middle cerebral artery was measured by transcranial Doppler (2MHz Spencer 136 Technologies, Seattle USA) at depths ranging from 43 to 54 mm. Signal quality was 137 optimized and an M-mode screen shot was recorded to facilitate subsequent probe 138 placements and insonation angles.

139

140 After verification of signal quality, resting data were recorded for 6 min while 141 subjects breathed room air to assess CA at each altitude. Continuous analog data 142 (ABP, CBFv, ECG,  $O_2$  and  $CO_2$ ) were recorded at 200 Hz (ADInstruments Powerlab 143 16/30, Colorado Springs, CO, USA) for offline analysis. Core temperature and 144 arterial blood samples (2 ml) were taken during the last 30 s of measurement 145 periods. Blood samples were taken from the radial artery catheter and blood gases 146 were analyzed for PaCO<sub>2</sub> and PaO<sub>2</sub> in triplicate (RAPIDLab 248, Siemens, Erlangen, 147 Germany) and corrected for body temperature (15, 29).

#### 148 Acute Mountain Sickness

- 149 Self reported sections of the Lake Louise Questionnaire (LLQ) were used to assess
- 150 AMS on ALT1 and POST7 (~12 hours after arrival). Moderate and severe AMS were

151 defined as  $LLQ \ge 3$  and  $\ge 6$ , including headache, respectively (27).

#### 152 Data Analysis

- 153 Transfer function analyses were used to assess dynamic CA, based on spontaneous
- 154 fluctuations in the raw ABP and CBFv signals, as previously described (33, 34).
- 155 Briefly, 6-min recordings of instantaneous ABP and CBFv were reduced to beat-by-
- 156 beat averages, resampled at 5 Hz and transformed from the time to frequency
- domain using fast Fourier transformations (512 points per segment with 40%
- 158 overlap). The transfer function from mean ABP to CBFv was expressed in terms of
- 159 coherence, gain, and phase shift in the very low frequency range (0.02 0.07 Hz),
- 160 where dynamic CA is most active (21, 22), as well as in low (0.07 to 0.20 Hz) and

161 high (0.20 to 0.35 Hz) frequency ranges. All data were used in subsequent statistical 162 analyses. Reduction in phase shift was considered the primary criterion for 163 impaired CA because it signifies shorter delay in transmission of pressure (ABP) 164 into flow (CBFv), or a reduction in the ability of the cerebrovascular system to buffer 165 changes in ABP and maintain consistent blood flow. Yet, since increases in gain 166 (increase in CBFv relative to a change in ABP) and coherence (linear correlation 167 between ABP and CBFv) may also suggest CA impairment (8, 24, 41), all three 168 transfer function metrics are reported. To address difficulties in interpreting 169 possible permutations of these three variables, the inverse transfer function of the 170 resulting gain and phase shift was used to express results in the time domain as a 171 step function that could be fitted to one of 10 curves representing a single 172 autoregulation index (ARI) score (36). An ARI score of 0 indicates complete lack of 173 autoregulation and 9 indicates perfect autoregulation.

#### 174 Statistics

175 After calculating descriptive statistics (mean ± SD) and verifying normality

176 (D'Agostino and Pearson Test), variables were analyzed by repeated measures

177 ANOVAs to evaluate the effect of time on CA metrics with Fisher's LSD *post hoc* tests

and the Holm procedure to correct for multiple comparisons ( $\alpha = 0.05$ ).

179

180 Spearman rho correlations were run to evaluate relations between CA metrics and

181 the severity of LLQ symptom scores. Specifically, we tested the ability of CA

- assessments, measured at SL and upon arrival at ALT1, to predict ensuing
- 183 symptoms of AMS (7). Also, because AMS classification is dichotomous (i.e. positive

184	vs. negative), we used receiver operating characteristic (ROC) analyses (14, 18) to
185	evaluate the sensitivity (true positive rate) and specificity (true negative rate) of
186	ARI scores' ability to detect mild and severe AMS. The ROC area under the curve
187	(AUC) statistic was used as an indicator of test accuracy. An AUC of 1.0 signifies a
188	perfect test, with no chance of false positive or false negative results, while an AUC
189	of 0.5 signifies a meaningless test, where the probability of identifying a true
190	positive result is only 50%.

## 191 **Results**

### 192 Subjects

193

194 We studied 21 subjects at SL (12 males and 9 females; 21 ± 1 years old). Because of

195 logistical problems upon arrival in Bolivia, complete data sets were not obtained on

the first 7 subjects upon arrival at ALT1. Since the first 7 subjects comprised the

197 cohort studied at POST21, longitudinal assessments of CA were limited to the

remaining 14 subjects who completed the study at POST7.

199

201

### 200 Effect of Rapid Ascent to High Altitude

202 At SL, resting cardiovascular (HR, ABP, CBFv) and CA measurements (coherence,

203 gain, phase shift and ARI scores) were characteristic of young, healthy individuals

- with intact CA (Table 1, Figure 1). From SL to ALT1, PaO<sub>2</sub> and PaCO<sub>2</sub> decreased (65
- and 26%, respectively, P< 0.001, Table 1). This degree of hypoxia increased HR (P <
- 206 0.001), but did not affect mean ABP or CBFv. Very low frequency power spectral

207 density (PSD) of ABP and CBFv were unaltered, but increases in transfer function

208 coherence (P < 0.001) and decreases in phase shift (P < 0.050) and ARI score (P < 0.050) and (P < 0.050) an

209 0.001) were consistent (in 13 of 14 subjects) with the definition of impaired CA at

210 ALT1.

211

### 212 Effect of Acclimatization to High Altitude

- Acclimatization increased resting PaO<sub>2</sub> (27%) and decreased PaCO<sub>2</sub> (22%) from
- ALT1 to ALT16 (both P<0.001), without affecting HR, ABP or CBFv. Measures of CA
- at ALT16 were unchanged from ALT1 and remained impaired relative to SL in the
- 216 very low frequency range (all P < 0.010, Table 1, Figure 1).
- 217 Effect of Re-exposure to High Altitude
- 218 Resting PaO<sub>2</sub> and PaCO<sub>2</sub> at POST7 fell between ALT1 and ALT16 values (all P<0.050
- vs. ALT1 and vs. ALT16), indicating that the degree of acclimatization achieved at
- ALT16 was partially maintained at POST7. Assessments of CA at POST7 were similar
- to those at ALT1 and ALT16 and remained impaired relative to SL in the very low
- frequency range (P < 0.050, Table 1, Figure 1).
- 223

225

#### 224 Association between CA and AMS

- 226 Of the 21 subjects, 17 reported symptoms of at least moderate AMS at ALT1 (LLQ =
- $6.4 \pm 2.2$ ), 10 of who met the criteria for severe AMS (LLQ =  $7.8 \pm 1.7$ ). Correlations
- between CA metrics preceding the development of AMS symptom were weak (all
- r<0.50, P>0.050, Figure 2). The ROC analysis revealed that ARI scores measured at

230 SL were not sensitive or specific predictors of moderate (AUC=0.54, P=0.788) or

severe AMS (AUC=0.69, P=0.139). Additionally, the degree of impairment in CA

232 (measured as the change in ARI from SL to ALT1) was not a sensitive or specific

233 predictor of moderate (AUC=0.53, P=0.881) or severe AMS (AUC=0.72, P=0.124).

None of the 14 subjects studied at POST7 reported symptoms of AMS, thus

associations with CA could not be tested.

## 236 **Discussion**

237 The key findings of this study were that cerebral autoregulation, as assessed by

transfer function analysis, is 1) impaired upon rapid ascent to high altitude, 2)

unaffected by acclimatization, or 3) subsequent re-exposure to the same altitude,

and 4) not a sensitive or specific predictor of AMS. Based on our results we question

241 whether the so-called impairment in CA that persists at high altitude is

242 characteristic of pathological insufficiency in cerebrovascular regulation (16), or

243 alternatively reflects a relatively benign relaxation in autoregulation.

244

#### 245 Effect of high altitude on CA

246 This is the first longitudinal study of CA at high altitude, from rapid ascent through

247 acclimatization and upon re-exposure after a short period at low altitude. We show

that impairment of CA was a consistent characteristic across this high-altitude

249 exposure profile.

251 Increased transfer function coherence and gain along with reduced phase shift and 252 ARI score upon rapid ascent were all consistent with the classic definition of 253 impaired CA (Table 1) and outside the normal range of expected variability (6), 254 implying that changes in ABP were more readily transmitted into the cerebral 255 circulation as changes in CBFv at high altitude. Our finding of impaired CA after less 256 than one day of travel from low to high elevation is consistent with our previous 257 findings after 4 hours in a hypobaric chamber (35) and fills an important gap in the 258 literature between studies conducted in laboratories with hypoxic gas mixtures, 259 where normobaric hypoxia was achieved in a matter of minutes (5, 10, 26, 34), and 260 studies of trekkers, where several days of progressive ascent preceded initial high-261 altitude measurements (1, 2, 12, 37). Impaired CA at rest in acute hypoxia is a 262 consistent finding among all but one study (26), suggesting that neither the mode 263 nor rate of ascent appears to affect the general assessment. 264 265 By evaluating CA upon initial exposure and after 16 days at high altitude, we were 266 able to determine if changes in CA occur with acclimatization, as might be expected with increased  $PaO_2$  (2, 35), decreased  $PaCO_2$  (19, 23, 26), and further 267 268 sympathoexcitation (1). On the contrary, we found no change in CA over the course 269 of acclimatization (Table 1). Our longitudinal findings are consistent with other 270 cross-sectional studies demonstrating impaired CA at various time points after 271 arrival at high altitude (1, 2, 7, 11, 12, 37) and in permanent high-altitude residents 272 (12, 13). These results may indicate that assessments of CA are less sensitive to 273 changes in PaO<sub>2</sub> and PaCO<sub>2</sub> near their respective extremes. Alternatively, a slight

improvement in CA due to increased PaO<sub>2</sub> (2, 35) may have been masked if the
opposing effects of PaCO<sub>2</sub> (19, 23, 26) and/or sympathoexcitation (1) on CA were
heightened over time at altitude. Further testing with manipulation of arterial gases
and sympathetic activity is necessary to determine the relative influence of arterial
gases and neural stimulation on CA at high altitude, yet impaired CA remains a
consistent functional consequence across time at high altitude.

280

281 As an additional test of the hypothesis that impaired CA is a consistent response to 282 hypoxemia, we sent subjects down to low altitude for 7 days and re-evaluated their 283 CA response after a second rapid ascent back to high altitude. Upon re-exposure, the 284 measured impairment in CA was similar to that observed upon the first ascent 285 (ALT1) and after acclimatization (ALT16). Together, these results demonstrate that 286 impaired autoregulation was a consistent characteristic of hypoxemia across our 287 study and imply that slow fluctuations in arterial pressure were less effectively 288 dampened by the cerebral vasculature regardless of the state of acclimatization. 289 What remains to be determined is if such a tenuous pressure-flow relation may be 290 potentially harmful.

291

### 292 Relation of CA to AMS

Impairment of CA has been suggested to play a role in the development of AMS by
either permitting cerebral overperfusion and mechanical disruption of the blood
brain barrier (i.e. vasogenic cerebral edema) when mean ABP is elevated, or by
cerebral under-perfusion and exacerbation of cerebral hypoxia/ischemia when

297 mean ABP is lowered (9, 16). In the present study, we found no correlation between 298 measures of CA and subsequent AMS symptom scores (Figure 2), which opposes the 299 notion that lower CA predisposes people to AMS, or conversely, that higher CA 300 confers protection from AMS. Our additional ROC analyses of AMS status, confirmed 301 that ARI scores were neither sensitive nor specific indicators for the development of 302 moderate or severe AMS upon arrival at high altitude. These findings are congruent 303 with our previous report following the time course of changes in CA and AMS 304 symptoms over the first 10 hours of exposure to hypobaric hypoxia (35), where we 305 found similar levels of CA impairments in subjects who eventually developed AMS 306 or stayed healthy, but are at odds with other studies showing some association 307 between CA and AMS symptoms (5, 37). Our data also counter a recent finding that 308 sea-level assessments of CA predict ensuing severity of AMS (7).

309

310 Discrepancies between studies may be explained by the various methods used to 311 assess CA (transfer function vs. leg cuff – see Limitations), the questionnaires used 312 to assess AMS (LLQ vs. Environmental Symptoms Questionnaire), and the statistical 313 approach used to evaluate the relation between CA and AMS (correlation vs. ROC). 314 We acknowledge that caution should be exercised when interpreting correlations 315 with an ordinal level variable, such as the LLQ score, because by definition the scale 316 has limited mathematical meaning. For example, an LLQ score of 6 does not imply 317 symptom severity is exactly twice that of a score of 3. Due to the intrinsic level of 318 measurement, we believe that LLQ scores are best restricted to dichotomous 319 classification of positive or negative AMS status, and thus place more emphasis of

the negative results of our ROC analysis. We encourage others to consider thismethod of analysis for future AMS studies.

322

323 Overall, given the similarity in CA responses among individuals with a wide range of 324 AMS scores, we do not believe that changes in CA cause AMS. This assertion is 325 further supported by the complete lack of association between impaired CA at 326 POST7 when no symptoms of AMS were reported and previous reports 327 documenting impaired CA in healthy high-altitude natives (12, 13). Nonetheless, we 328 must acknowledge that the alteration in CA upon acute altitude exposure may set up 329 a tenuous pressure-flow relation which could permit AMS to develop, if other, yet 330 unidentified, factors are present at the same time. 331

332 Since impairment of CA appears to be a consistent physiological response in hypoxic 333 environments and unrelated to AMS status, it is tempting to speculate that the 334 underlying change in the cerebral pressure-flow relation may actually promote 335 successful acclimatization or adaptation to chronic states of hypoxemia (4). It is 336 possible that impairment of CA could promote cerebral oxygen delivery in a time of 337 need since it allows greater cerebral perfusion for a given increase in ABP. This 338 potentially beneficial consequence of impaired CA during hypoxemic stress might 339 outweigh the relative risk of reduced cerebral perfusion if ABP were to drop. We 340 therefore raise the possibility that the term impaired CA may be a misnomer 341 because it implies an association with pathology that has yet to be substantiated in 342 acute or chronic hypoxemia. We suggest that relaxation of CA might be a more

accurate term to describe changes in the cerebral pressure-flow relation fromnormoxia to hypoxia in the absence of pathology.

345

#### 346 Limitations

347 One major limitation affecting the field is the lack of a gold standard method to 348 assess CA. We have chosen to evaluate rhythmical fluctuations in CA via transfer 349 function analysis, primarily because we believe it captures the natural cerebral 350 pressure-flow relation over time and thus has greater practical relevance over 351 methods which induce larger, more abrupt changes in ABP, as with leg cuff 352 inflation/deflation, rapid tilting, or more sustained changes in ABP, as with 353 pharmaceutical interventions. Still, we acknowledge that transfer function analysis 354 of resting data monitors relatively subtle fluctuations in ABP and CBFv, which, if 355 amplified, may not show impairment in CA (39). These factors may limit the 356 generalizability of resting CA assessments and lead to overstatement of the clinical 357 relevance of the findings. Additionally, there are no universal standards for the 358 parameter settings used in transfer function analysis or interpretation of 359 subsequent results, which makes comparisons between studies problematic. Future 360 work is needed to clarify differences in methods used to assess CA in hypoxemic 361 states and evaluate if these changes are generalizable to clinical settings. 362 363 Most CA studies rely on transcranial Doppler measurements of flow velocity and 364 assume that vessel diameter is unchanged, yet there is evidence to suggest that this

assumption may be invalid at extreme altitudes (39, 40). Dilation of the MCA at

366 ALT1 may explain why MCAv did not follow the expected increase in CBF upon 367 acute exposure to high altitude (30). We do not believe potential MCA dilation 368 affected our interpretation because the phase shift - our primary criterion for 369 assessing changes in CA – measures the relative timing of oscillations in ABP and 370 CBFv and thus is largely independent of absolute flow. However, since small changes 371 in diameter can have profound affects on flow (flow  $\sim$  radius<sup>4</sup>), future studies must 372 consider the use of continuous flow measurements, instead of velocity 373 measurements, to accurately assess CA in hypoxia. 374

375 Finally, our measurements of CA were limited to the MCA and relied on pressure

376 measurements taken in the radial artery. Since regional differences in

377 cerebrovascular regulation have recently been reported (20, 28, 38), more specific

378 measurements of regional pressure and flow are needed to fully characterize CA.

#### 379 **Conclusions**

380 Our data demonstrate that the initial impairment of CA upon acute exposure to high

altitude is invariant with acclimatization and re-exposure, suggesting that relaxation

- in the regulation of the cerebral pressure-flow relation is a characteristic response
- to hypoxia that is unaffected by the degree of acclimatization. Since changes in CA do
- 384 not follow the progression and resolution of AMS, we question the clinical relevance
- 385 of impaired CA at high altitude.

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# 408 **References**

409 (39)1. Ainslie PN, Lucas SJ, Fan JL, Thomas KN, Cotter JD, Tzeng YC, and 410 Burgess KR. Influence of sympathoexcitation at high altitude on cerebrovascular 411 function and ventilatory control in humans. J Appl Physiol 113: 1058-1067, 2012. 412 Ainslie PN, Ogoh S, Burgess K, Celi L, McGrattan K, Peebles K, Murrell C, 2. 413 Subedi P, and Burgess KR. Differential effects of acute hypoxia and high altitude on 414 cerebral blood flow velocity and dynamic cerebral autoregulation: alterations with 415 hyperoxia. J Appl Physiol 104: 490-498, 2008. 416 Aries MJ, Elting JW, De Keyser J, Kremer BP, and Vroomen PC. Cerebral 3. 417 autoregulation in stroke: a review of transcranial Doppler studies. Stroke 41: 2697-418 2704, 2010. 419 4. **Bailey DM**. Impaired cerebral autoregulation in acute mountain sickness: 420 incidental yet adaptive? *Stroke* 41: e571; author reply e572, 2010. 421 Bailey DM, Evans KA, James PE, McEneny J, Young IS, Fall L, Gutowski M, 5. 422 Kewley E, McCord JM, Moller K, and Ainslie PN. Altered free radical metabolism 423 in acute mountain sickness: implications for dynamic cerebral autoregulation and 424 blood-brain barrier function. J Physiol 587: 73-85, 2009. 425 Brodie FG, Atkins ER, Robinson TG, and Panerai RB. Reliability of 6. 426 dynamic cerebral autoregulation measurement using spontaneous fluctuations in 427 blood pressure. Clin Sci (Lond) 116: 513-520, 2009. Cochand NJ, Wild M, Brugniaux JV, Davies PJ, Evans KA, Wise RG, and 428 7. 429 Bailey DM. Sea-level assessment of dynamic cerebral autoregulation predicts 430 susceptibility to acute mountain sickness at high altitude. Stroke 42: 3628-3630, 2011. 431 432 Giller CA. The frequency-dependent behavior of cerebral autoregulation. 8. Neurosurgery 27: 362-368, 1990. 433 434 Hackett PH, and Roach RC. High-altitude illness. N Engl | Med 345: 107-114, 9. 435 2001. 436 10. Iwasaki K, Ogawa Y, Shibata S, and Aoki K. Acute exposure to normobaric 437 mild hypoxia alters dynamic relationships between blood pressure and cerebral 438 blood flow at very low frequency. J Cereb Blood Flow Metab 27: 776-784, 2007. 439 11. Iwasaki K, Zhang R, Zuckerman JH, Ogawa Y, Hansen LH, and Levine BD. 440 Impaired dynamic cerebral autoregulation at extreme high altitude even after 441 acclimatization. J Cereb Blood Flow Metab 31: 283-292, 2011. 442 12. Jansen GF, Krins A, Basnyat B, Bosch A, and Odoom JA. Cerebral 443 autoregulation in subjects adapted and not adapted to high altitude. *Stroke* 31: 444 2314-2318, 2000. 445 Jansen GF, Krins A, Basnyat B, Odoom JA, and Ince C. Role of the altitude 13. 446 level on cerebral autoregulation in residents at high altitude. J Appl Physiol 103: 518-447 523, 2007. 448 Katsogridakis E. Bush G. Fan L. Birch AA. Simpson DM. Allen R. Potter IF. 14. 449 and Panerai RB. Detection of impaired cerebral autoregulation improves by 450 increasing arterial blood pressure variability. J Cereb Blood Flow Metab 33: 519-523, 451 2013.

452 15. Kelman GR, and Nunn JF. Nomograms for correction of blood Po2, Pco2, pH, 453 and base excess for time and temperature. J Appl Physiol 21: 1484-1490, 1966. 454 Lassen NA, and Harper AM. Letter: High-altitude cerebral oedema. Lancet 2: 16. 455 1154, 1975. 456 Levine BD, Zhang R, and Roach RC. Dynamic cerebral autoregulation at 17. 457 high altitude. Adv Exp Med Biol 474: 319-322, 1999. 458 Metz CE. Basic principles of ROC analysis. Semin Nucl Med 8: 283-298, 1978. 18. 459 19. Ogoh S, Nakahara H, Ainslie PN, and Miyamoto T. The effect of oxygen on 460 dynamic cerebral autoregulation: critical role of hypocapnia. J Appl Physiol 108: 538-461 543, 2010. 462 Ogoh S, Sato K, Nakahara H, Okazaki K, Subudhi AW, and Miyamoto T. 20. 463 Effect of acute hypoxia on blood flow in vertebral and internal carotid arteries. *Exp* 464 Physiol 98: 692-698, 2013. 21. 465 **Panerai RB**. Cerebral autoregulation: from models to clinical applications. 466 Cardiovasc Eng 8: 42-59, 2008. 467 22. **Panerai RB**. Transcranial Doppler for evaluation of cerebral autoregulation. 468 Clin Auton Res 19: 197-211, 2009. 469 23. Panerai RB, Deverson ST, Mahony P, Hayes P, and Evans DH. Effects of 470 CO2 on dynamic cerebral autoregulation measurement. *Physiol Meas* 20: 265-275, 471 1999. 472 24. Panerai RB, White RP, Markus HS, and Evans DH. Grading of cerebral 473 dynamic autoregulation from spontaneous fluctuations in arterial blood pressure. 474 Stroke 29: 2341-2346, 1998. 475 Paulson OB, Strandgaard S, and Edvinsson L. Cerebral autoregulation. 25. 476 Cerebrovasc Brain Metab Rev 2: 161-192, 1990. 477 Querido JS, Ainslie PN, Foster GE, Henderson WR, Halliwill JR, Ayas NT, 26. 478 and Sheel AW. Dynamic cerebral autoregulation during and following acute 479 hypoxia: role of carbon dioxide. *J Appl Physiol* 2013. 480 27. Roach RC, Bartsch P, Hackett PH, and Oelz O. The Lake Louise acute 481 mountain sickness scoring system. In: *Hypoxia and Molecular Medicine*, edited by 482 Sutton JR, Coates J, and Houston CS. Burlinton, VT: Queen City Printers, 1993, p. 483 272-274. 484 28. Sato K, Sadamoto T, Hirasawa A, Oue A, Subudhi AW, Miyazawa T, and 485 **Ogoh S**. Differential blood flow responses to CO(2) in human internal and external 486 carotid and vertebral arteries. J Physiol 590: 3277-3290, 2012. 487 29. Severinghaus JW. Blood gas calculator. J Appl Physiol 21: 1108-1116, 1966. 30. 488 Severinghaus JW, Chiodi H, Eger EI, 2nd, Brandstater B, and Hornbein 489 **TF**. Cerebral blood flow in man at high altitude. Role of cerebrospinal fluid pH in 490 normalization of flow in chronic hypocapnia. *Circ Res* 19: 274-282, 1966. 491 Steinmeier R, Bauhuf C, Hubner U, Bauer RD, Fahlbusch R, Laumer R, 31. 492 and Bondar I. Slow rhythmic oscillations of blood pressure, intracranial pressure, 493 microcirculation, and cerebral oxygenation. Dynamic interrelation and time course 494 in humans. Stroke 27: 2236-2243, 1996. 495 Subudhi AW, Bourdillon N, Bucher J, Davis C, Elliot JE, Eutermoster M, 32. 496 Evero O, Fan JL, Jameson-Van Houten S, Julian CG, Kark J, Kark S, Kayser B,

497 Kern J, Kim SE, Lathan C, Laurie SS, Lovering AT, Paterson R, Polaner D, Ryan

498 BJ, Spira J, Wachsmuth NB, and Roach RC. AltitudeOmics: The integrative 499 physiology of human acclimatization to hypobaric hypoxia and its memory on 500 reascent. In Review. 501 Subudhi AW, Dimmen AC, Julian CG, Wilson MJ, Panerai RB, and Roach 33. 502 **RC**. Effects of acetazolamide and dexamethasone on cerebral hemodynamics in 503 hypoxia. I Appl Physiol 110: 1219-1225, 2011. 504 Subudhi AW, Panerai RB, and Roach RC. Acute hypoxia impairs dynamic 34. 505 cerebral autoregulation: results from two independent techniques. J Appl Physiol 506 107: 1165-1171, 2009. 507 35. Subudhi AW, Panerai RB, and Roach RC. Effects of hypobaric hypoxia on 508 cerebral autoregulation. Stroke 41: 641-646, 2010. 509 Tiecks FP, Lam AM, Aaslid R, and Newell DW. Comparison of static and 36. 510 dynamic cerebral autoregulation measurements. *Stroke* 26: 1014-1019, 1995. 511 37. Van Osta A, Moraine JJ, Melot C, Mairbaurl H, Maggiorini M, and Naeije R. 512 Effects of high altitude exposure on cerebral hemodynamics in normal subjects. 513 Stroke 36: 557-560, 2005. 514 38. Willie CK, Macleod DB, Shaw AD, Smith KJ, Tzeng YC, Eves ND, Ikeda K, 515 Graham J, Lewis NC, Day TA, and Ainslie PN. Regional brain blood flow in man 516 during acute changes in arterial blood gases. J Physiol 590: 3261-3275, 2012. 517 Willie CK, Smith KJ, Day TA, Ray LA, Lewis NC, Bakker A, Macleod DB, 39. 518 and Ainslie PN. Regional cerebral blood flow in humans at high altitude: Gradual 519 ascent and two weeks at 5050m. [Appl Physiol (1985) 2013. 520 40. Wilson MH, Edsell ME, Davagnanam I, Hirani SP, Martin DS, Levett DZ, 521 Thornton JS, Golav X, Strycharczuk L, Newman SP, Montgomery HE, Grocott 522 MP, and Imray CH. Cerebral artery dilatation maintains cerebral oxygenation at 523 extreme altitude and in acute hypoxia--an ultrasound and MRI study. *J Cereb Blood* 524 Flow Metab 31: 2019-2029, 2011. 525 Zhang R, Zuckerman JH, Giller CA, and Levine BD. Transfer function 41. 526 analysis of dynamic cerebral autoregulation in humans. Am J Physiol 274: H233-241, 527 1998. 528

## 530 Figure Captions

531

- 532 Figure 1. Arterial blood pressure to cerebral blood flow velocity transfer function
- 533 metrics (mean ± SD from 0 to 0.5 Hz) at sea level (SL), upon arrival at 5,260 m
- 534 (ALT1), after 16 days of acclimatization (ALT16), and upon re-exposure to 5,260 m
- after 7 days at low altitude (POST7). Similar impairments in cerebral
- autoregulation (increased coherence and gain and decreased phase shift) from SL
- 537 were seen in the very low frequency (0.02 to 0.07 Hz shaded area) at ALT1, ALT16,
- and POST7 (P < 0.05). \* Different from SL. & Different from SL.

- 540 Figure 2. Scatter plots showing no relation (P > 0.05) between autoregulation
- 541 indices (ARI), measured at sea level (SL, top) and as the change from SL to arrival at
- 542 high altitude (ALT1, bottom), and AMS symptoms scores from the Lake Louise
- 543 Questionnaire at ALT1.

Variable		SL	ALT1	ALT16	POST7
PaO <sub>2</sub>	mmHg	103 ± 5	36 ± 3*	45 ± 4*#	42 ± 4*#&
PaCO <sub>2</sub>	mmHg	37 ± 4	28 ± 2*	21 ± 3*#	24 ± 3*#&
HR	bpm	73 ± 9	90 ± 18*	95 ± 12*	85 ± 15*&
ABP	mmHg	77 ± 6	$76 \pm 14$	$81 \pm 10$	76 ± 8
CBFv	cm/s	62 ± 9	63 ± 14	59 ± 7	57 ± 9
PSD ABP	mmHg²/Hz	11 ± 13	9 ± 4	9 ± 5	6 ± 4
PSD CBFv	$(cm/s)^2/Hz$	13 ± 19	$14 \pm 16$	10 ± 6	11 ± 8
Coherence		$0.42 \pm 0.12$	0.64 ± 0.15*	$0.70 \pm 0.16^*$	0.55 ± 0.12*&
Gain	%/%	$0.64 \pm 0.24$	0.88 ± 0.35*	0.85 ± 0.25*	0.97 ± 0.33*
Phase Shift	radians	$0.48 \pm 0.28$	0.17 ± 0.21*	$0.27 \pm 0.09^*$	0.25 ± 0.19*
ARI		$4.4 \pm 1.0$	2.8 ± 0.9*	$2.8 \pm 1.0^{*}$	3.3 ± 1.6*

Table 1. Resting Data (n=14, mean ±SD)

\* Different from SL

# Different from ALT1

& Different from ALT16









