

Potent New Small-Molecule Inhibitor of Botulinum Neurotoxin Serotype A Endopeptidase Developed by Synthesis-Based Computer-Aided Molecular Design

Yuan-Ping Pang^{1*}, Anuradha Vummenthala¹, Rajesh K. Mishra¹, Jewn Giew Park¹, Shaohua Wang¹, Jon Davis^{2a}, Charles B. Millard^{2ab}, James J. Schmidt^{3*}

1 Computer-Aided Molecular Design Laboratory, Mayo Clinic, Rochester, Minnesota, United States of America, **2** Division of Biochemistry, Walter Reed Army Institute of Research, Silver Spring, Maryland, United States of America, **3** Integrated Toxicology Division, United States Army Medical Research Institute of Infectious Diseases, Frederick, Maryland, United States of America

Abstract

Botulinum neurotoxin serotype A (BoNTA) causes a life-threatening neuroparalytic disease known as botulism. Current treatment for post exposure of BoNTA uses antibodies that are effective in neutralizing the extracellular toxin to prevent further intoxication but generally cannot rescue already intoxicated neurons. Effective small-molecule inhibitors of BoNTA endopeptidase (BoNTAe) are desirable because such inhibitors potentially can neutralize the intracellular BoNTA and offer complementary treatment for botulism. Previously we reported a serotype-selective, small-molecule BoNTAe inhibitor with a K_i^{app} value of $3.8 \pm 0.8 \mu\text{M}$. This inhibitor was developed by lead identification using virtual screening followed by computer-aided optimization of a lead with an IC_{50} value of $100 \mu\text{M}$. However, it was difficult to further improve the lead from micromolar to even high nanomolar potency due to the unusually large enzyme-substrate interface of BoNTAe. The enzyme-substrate interface area of $4,840 \text{ \AA}^2$ for BoNTAe is about four times larger than the typical protein-protein interface area of $750\text{--}1,500 \text{ \AA}^2$. Inhibitors must carry several functional groups to block the unusually large interface of BoNTAe, and syntheses of such inhibitors are therefore time-consuming and expensive. Herein we report the development of a serotype-selective, small-molecule, and competitive inhibitor of BoNTAe with a K_i value of $760 \pm 170 \text{ nM}$ using synthesis-based computer-aided molecular design (SBCAMD). This new approach accounts the practicality and efficiency of inhibitor synthesis in addition to binding affinity and selectivity. We also report a three-dimensional model of BoNTAe in complex with the new inhibitor and the dynamics of the complex predicted by multiple molecular dynamics simulations, and discuss further structural optimization to achieve better *in vivo* efficacy in neutralizing BoNTA than those of our early micromolar leads. This work provides new insight into structural modification of known small-molecule BoNTAe inhibitors. It also demonstrates that SBCAMD is capable of improving potency of an inhibitor lead by nearly one order of magnitude, even for BoNTAe as one of the most challenging protein targets. The results are insightful for developing effective small-molecule inhibitors of protein targets with large active sites.

Citation: Pang Y-P, Vummenthala A, Mishra RK, Park JG, Wang S, et al. (2009) Potent New Small-Molecule Inhibitor of Botulinum Neurotoxin Serotype A Endopeptidase Developed by Synthesis-Based Computer-Aided Molecular Design. PLoS ONE 4(11): e7730. doi:10.1371/journal.pone.0007730

Editor: Narcis Fernandez-Fuentes, Leeds Institute of Molecular Medicine, United Kingdom

Received: September 2, 2009; **Accepted:** October 13, 2009; **Published:** November 10, 2009

This is an open-access article distributed under the terms of the Creative Commons Public Domain declaration which stipulates that, once placed in the public domain, this work may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose.

Funding: This work was supported by the United States Army Medical Research & Materiel Command (W81XWH-04-2-0001 and W81XWH-08-1-0154), the United States Army Research Office (W911NF-09-1-0095), the United States Defense Threat Reduction Agency (3.10023_07_RD_B and 3.10014_08_WR_B), and the University of Minnesota Supercomputing Institute. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. Opinions, interpretations, conclusions, and recommendations are those of the authors and are not necessarily endorsed by the United States Army.

Competing Interests: All authors are inventors of a filed provisional patent application titled "Small-Molecule Botulinum Toxin Inhibitors" that is related to the inhibitors described in this paper, but this does not alter the adherence to all the PLoS ONE policies on sharing data and materials.

* E-mail: pang@mayo.edu (YPP); james.schmidt@det.amedd.army.mil (JJS)

^a Current address: Integrated Toxicology Division, United States Army Medical Research Institute of Infectious Diseases, Frederick, Maryland, United States of America

^b Current address: United States Army Medical Research & Materiel Command, Fort Detrick, Maryland, United States of America

Introduction

Botulism is a life-threatening neuroparalytic disease that occurs in at least five forms—food-borne botulism, infant botulism, wound botulism, adult enteric infectious botulism, and inhalation botulism. The disease is caused by the specific action of botulinum neurotoxins. Seven antigenically distinct botulinum neurotoxins (serotypes A to G) are known, but primarily serotypes A, B, E, and F have been reported to cause botulism in humans.

Botulinum neurotoxin serotype A (BoNTA) is a protein produced by the spore-forming anaerobic bacterium, *Clostridium*

botulinum. The lethal doses of BoNTA for a 70-kg human are estimated as 0.9 μg by inhalation and 70 μg by ingestion [1]. BoNTA inhibits the release of acetylcholine from presynaptic nerve terminals at neuromuscular junctions, causing flaccid paralysis and frequently leading to prolonged mechanical ventilation with serious medical sequelae or death following respiratory arrest [2]. Despite its toxicity, the purified and diluted BoNTA has been used as medical treatment for cholinergic nerve and muscle dysfunctions [3,4]; BoNTA (marketed as Botox) also can temporarily reduce facial frown lines. However, overdose with BoNTA can develop systemic botulism [5].

Report Documentation Page

Form Approved
OMB No. 0704-0188

Public reporting burden for the collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to a penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number.

1. REPORT DATE 10 NOV 2009	2. REPORT TYPE N/A	3. DATES COVERED -	
4. TITLE AND SUBTITLE Potent new small-molecule inhibitor of botulinum neurotoxin serotype A endopeptidase developed by synthesis-based computer-aided molecular design. PLoS One (online) 4:e7730		5a. CONTRACT NUMBER	
		5b. GRANT NUMBER	
		5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Pang, Y-P Vummenthala, A Mishra, RK Park, JG Wang, S Davis, J Millard, CB Schmidt, JJ		5d. PROJECT NUMBER	
		5e. TASK NUMBER	
		5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) United States Army Medical Research Institute of Infectious Diseases, Fort Detrick, MD		8. PERFORMING ORGANIZATION REPORT NUMBER TR-09-116	
9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS(ES)		10. SPONSOR/MONITOR'S ACRONYM(S)	
		11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION/AVAILABILITY STATEMENT Approved for public release, distribution unlimited			
13. SUPPLEMENTARY NOTES The original document contains color images.			
14. ABSTRACT Botulinum neurotoxin serotype A (BoNTA) causes a life-threatening neuroparalytic disease known as botulism. Current treatment for postexposure of BoNTA uses antibodies that are effective in neutralizing the extracellular toxin to prevent further intoxication but generally cannot rescue already intoxicated neurons. Effective small-molecule inhibitors of BoNTA endopeptidase (BoNTAc) are desirable because such inhibitors potentially can neutralize the intracellular BoNTA and offer complementary treatment for botulism. Previously, we reported a serotype-selective, small-molecule BoNTAc inhibitor with a K1pp value of $3.8 \pm 0.8 \mu\text{M}$. This inhibitor was developed by lead identification using virtual screening followed by computer-aided optimization of a lead with an IC50 value of $100 \mu\text{m}$			
15. SUBJECT TERMS Clostridium botulinum, neurotoxin, serotype A, small-molecule inhibitors, computer-aided design			
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT
a. REPORT unclassified	b. ABSTRACT unclassified	c. THIS PAGE unclassified	SAR
			18. NUMBER OF PAGES 17
			19a. NAME OF RESPONSIBLE PERSON

Current post-exposure treatment of botulism relies on administration of antibodies against the toxins. The available antitoxin is an equine product that causes allergic reactions in ~10% of patients [6]. In late 2003, a solution of human antibodies was licensed as “BabyBIG” to treat infant botulism [7], but to date generic botulism immune globulin injections are still unavailable. Furthermore, antibodies are effective in neutralizing the extracellular toxin to prevent further intoxication but generally cannot rescue already intoxicated neurons. There is a need to develop complementary, post-exposure treatments such as small-molecule antidotes that potentially can neutralize the intracellular toxin.

Structurally, BoNTA consists of a light chain ($M_r \sim 50,000$) and a heavy chain ($M_r \sim 100,000$) that are linked by a disulfide bond. The light-chain is a zinc endopeptidase that specifically cleaves SNAP-25, a neuronal protein required for acetylcholine release [8]. The BoNTA endopeptidase (BoNTAe) has been a target for developing small-molecule inhibitors of the toxin [9–15]. However, developing potent inhibitors is as difficult as developing small-molecule inhibitors of protein-protein complexes. The latter has been a known challenge for decades [16]. The difficulty arises because, upon binding, the substrate wraps around the circumference of BoNTAe constituting an unusually large substrate enzyme interface area of $4,840 \text{ \AA}^2$ (Figure 1) [17]. This interface requires molecules possessing several functional groups to block it. Consequently, syntheses of such molecules are time-consuming and costly. The extent of the challenge can be appreciated by comparing the interface area ($4,840 \text{ \AA}^2$) of BoNTAe to the typical protein-protein interface area of $750\text{--}1500 \text{ \AA}^2$ [16]. Because of the large-interface challenge, small-molecule BoNTAe inhibitors reported to date typically have inhibitory potencies in the low micromolar range (see Section 3.1). To date, the most potent BoNTAe inhibitor has been an elegantly designed peptidic analog that mimics GlnArgAlaThrLysMetLeu, and this heptapeptidic mimetic (HPM) has a K_i value of 41 nM [18]. Nevertheless, some of the reported micromolar small-molecule inhibitors already demonstrate *ex vivo* efficacy in protecting cells against BoNTA [9,11]. Our micromolar small-molecule BoNTAe inhibitor leads also demonstrate *in vivo* efficacy at a concentration of 2 mg/kg in protecting mice against 5 LD_{50} challenge of BoNTA (manuscript in preparation). These results suggest that BoNTAe is a viable target for developing small-molecule antidotes to BoNTA.

Previously we reported a serotype-selective, small-molecule BoNTAe inhibitor with a K_i^{app} value of $3.8 \pm 0.8 \text{ \mu M}$ [10]. This inhibitor was developed by lead identification using virtual screening followed by optimization of a lead with an IC_{50} value of 100 \mu M [19]. However, we were unable to further improve the lead from micromolar to even high-nanomolar potency using conventional computer-aided molecular design (CAMD), despite the use of a high-resolution BoNTAe crystal structure (free and in complex with a peptidic substrate analog) [20] and advanced terascale computer simulation techniques [10].

In this article, we report the development of a serotype-selective, small-molecule, and competitive inhibitor of BoNTAe with a K_i value of $760 \pm 170 \text{ nM}$ by using a new approach, termed synthesis-based computer-aided molecular design (SBCAMD), to address the challenge in synthesis. We also report a three-dimensional (3D) model of the high-nanomolar inhibitor in complex with BoNTAe and the dynamics of the complex predicted by multiple molecular dynamics simulations (MMDSs). On the basis of the 3D model, we discuss insights into further structural optimization to achieve K_i values comparable to that of HPM and the prospect of small-molecule BoNTAe inhibitors as effective antidotes to neutralize the intracellular BoNTA. We also discuss the advantages of both SBCAMD and CAMD in drug discovery.

Results

Inhibitor Design

MMDSs on our early lead **1** (Figure 2) [10] suggested that structures like analog **2** (Figure 2) have higher affinities for BoNTAe than **1**. This is because (1) one diphenylmethanol group of **2** can adopt the Venus-flytrap-like conformation to grasp Arg363 of BoNTAe more tightly via cation- π interactions than the phenyl group of **1**; (2) the other diphenylmethanol group can fill the large void of the BoNTAe active site and interact with Arg231 via cation- π interactions, whereas **1** cannot reach Arg231; (3) two ammonium groups of **2** rather than only one ammonium group of **1** can interact with four carboxylates (Glu55, Glu64, Glu164, and Glu257) at the active site.

However, we could not make **2** using the scheme shown in Figure S1 in a short period, nor could we make analog **6** (Figures 2 and S2), a thiophene analog of **2**, given the same time constraint. We subsequently replaced the pyrrole ring of **2** with a benzene ring to avoid the problem of making **2**. This transformation resulted in analogs **3–5** (Figure 2) that can be made with facile syntheses described in the schemes shown in Figure S3, Figure S4, and Figure S5, although these molecules are more bulky than **2** and carry an aromatic hydroxamate that has a relatively low affinity for zinc due to π -electron delocalization. We initially hoped that the bulkiness problem could be remedied by the flexibility of the BoNTAe active site and therefore made **3–5** with ease. However, biological evaluation showed that none of these analogs was active in inhibiting BoNTAe at an inhibitor concentration of 10 \mu M (Table 1). Although change of an aromatic hydroxamate to an aliphatic hydroxamate does not require too much synthetic effort, we did not pursue this modification, as it would make these molecules even more bulky.

Alternatively, we made analog **7** (Figure 2) as a truncated analog of **6** according to the scheme shown in Figure S6. Unexpectedly, **7** precipitated in the inhibition assay buffer even though dimethyl sulfoxide was used to dissolve **7**. Gratifyingly, a different truncation of **6** led to the syntheses of analogs **8** and **9** (Figure 2) according to Schemes I–III of Figure 3. Encouragingly, both **8** and **9** showed 35% inhibition of BoNTAe at an inhibitor concentration of 10 \mu M (Table 1). These results inspired the design of analog **10a** (Figure 2). The replacement of the thiophene ring of **8** by a pyrrole ring in **10a** was due to our synthetic experience and available intermediates of the unsuccessful synthesis of **2**. In making **10a** according to Schemes I and IV of Figure 3, we had to protect the pyrrole nitrogen atom in order to improve the yield of the Sonagashira coupling [21]. However, we could not cleave the *p*-methoxybenzyl or benzyl group from the pyrrole nitrogen atom, nor could we protect the nitrogen atom using tosyl chloride. Nonetheless, according to the inhibitor-bound BoNTAe complexes generated by MMDSs, we conjectured that the benzyl group of **10b** could interact favorably with the serotype-specific Phe194 via the π - π interaction. We therefore completed the synthesis of **10a** without cleaving the benzyl group to give **10b**. More encouragingly, **10b** at 10 \mu M showed 64% inhibition of BoNTAe (Table 1). A structure-activity relationship study on a series of homologues of **10b** with two identical alkylamino chain lengths varying from 6 to 10 methylene groups identified *N,N*-bis(7-aminoheptyl)-1-benzyl-5-(3-(hydroxy-3,3-diphenylpropyl)-4-(3-hydroxyamino)-3-oxopropyl)-1*H*-pyrrole-2-carboxamide (AHP, see Figure 2) as the most potent of the series (see Section 2.4 for characterization of AHP). To confirm the conjecture about the interaction of the benzyl group with Phe194, we also made analog **11** with a methyl group attached to the pyrrole nitrogen atom of **11** (Figures 2 and 3). Indeed, **11** at 1 \mu M showed only 8% inhibition of BoNTAe,

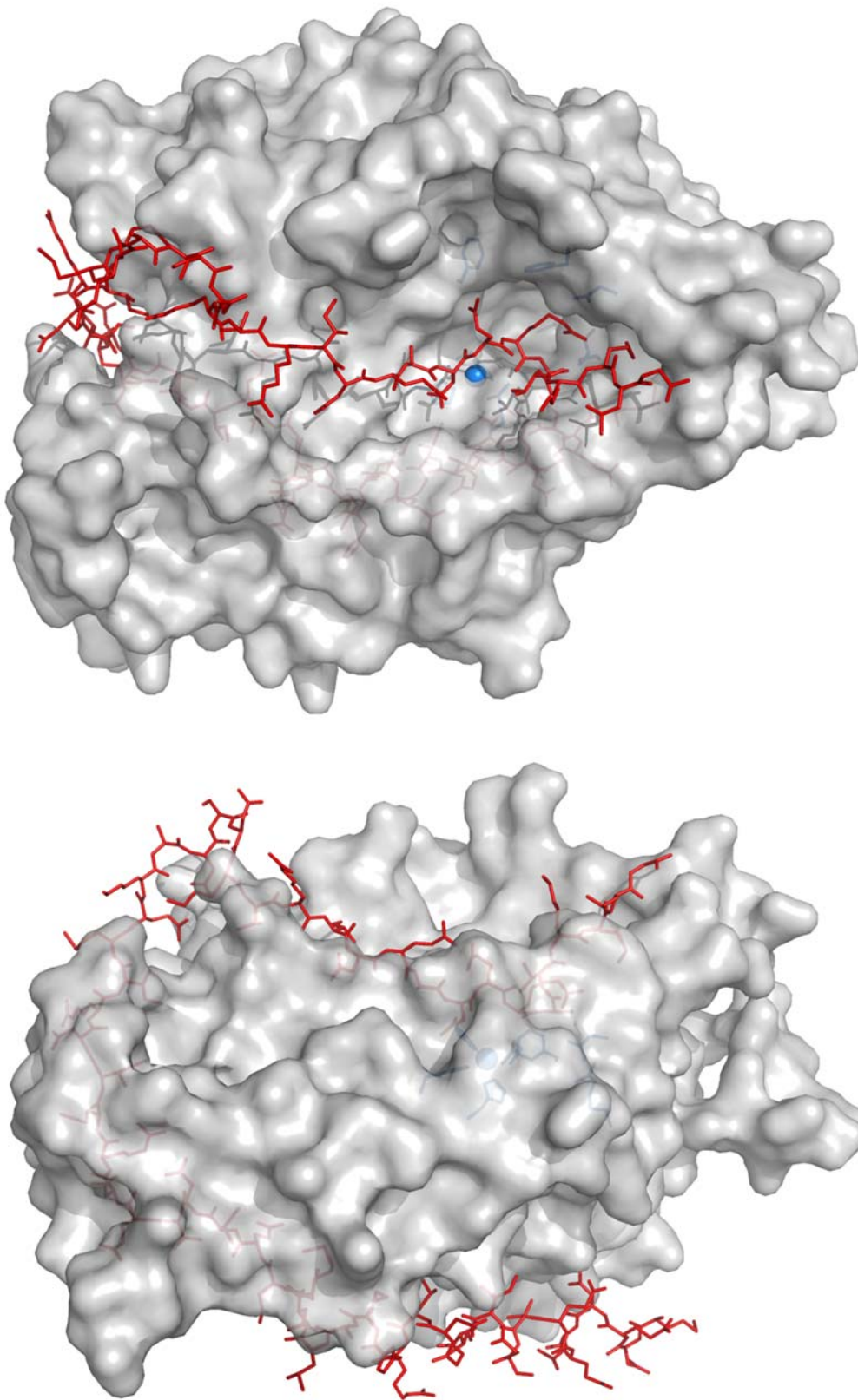


Figure 1. Large enzyme-substrate interface of BoNTAe. A: top view of the active site showing the substrate binding at the large pocket. B: side view of the active site showing the substrate wrapping around the circumference of BoNTAe. Active-site residues of BoNTAe (Zn^{+2} , H223, H227, E262, F163, F194, R363, and D370) are shown in light blue sphere or light blue stick model. The SNAP-25 substrate (146–204) is shown in red stick model. BoNTAe is shown in grey surface model with 15% transparency.
doi:10.1371/journal.pone.0007730.g001

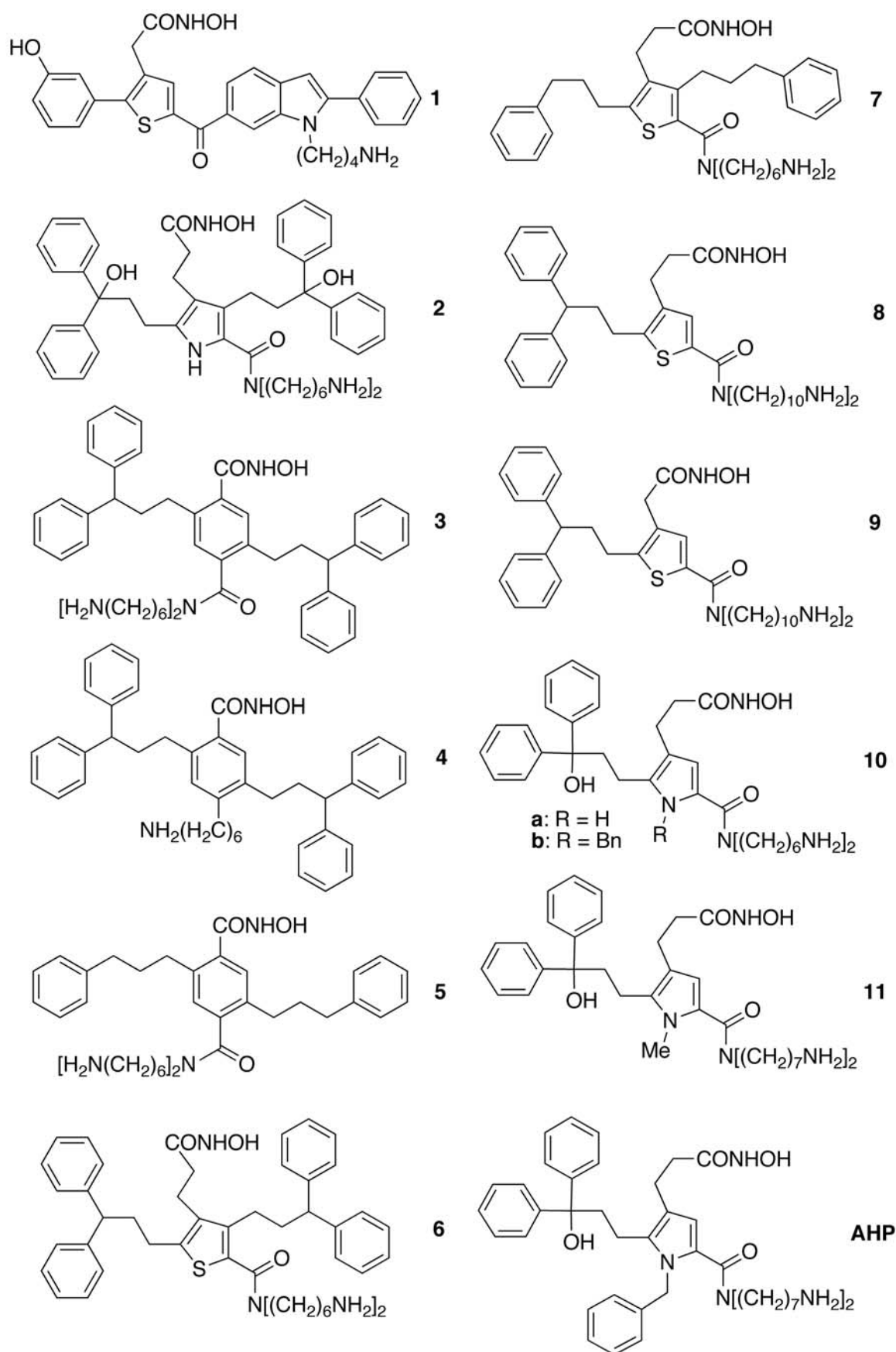


Figure 2. Chemical structures of BoNTAe inhibitors and their analogs resulted from SBCAMD.

doi:10.1371/journal.pone.0007730.g002

Table 1. Relative BoNTAe inhibition activities of AHP and its analogs.

Inhibitor	% Inhibition of BoNTAe
1	46% at 10 μ M
2	Not made
3	0% at 10 μ M
4	3% at 10 μ M
5	1% at 10 μ M
6	Not made
7	Precipitated in assay buffer
8	35% at 10 μ M
9	35% at 10 μ M
10a	Not made
10b	64% at 10 μ M
11	8% at 1 μ M
AHP	54% at 1 μ M

doi:10.1371/journal.pone.0007730.t001

whereas **AHP** exhibited 54% inhibition under the same conditions (Table 1).

Computer Simulation

Twenty different molecular dynamics simulations (each lasted 10 ns) were carried out for the **AHP**•BoNTAe complex to probe the intermolecular interactions of the design inhibitor using the PMEMD module of the AMBER 8 program [22] (see Section 4.4). The initial complex structure was generated by manually docking **AHP** into the BoNTAe active site with (1) the hydroxamate group near the active-site zinc ion, (2) the diphenylmethanol group near Arg363, (3) the benzyl group near Phe194, (4) one ammonium group near Glu55 and Glu164, and (5) the other ammonium group near Glu257. The BoNTAe structure used for the docking was taken from the crystal structure of an inhibitor-bound BoNTAe (Protein Data Bank Code: 3BOO [20]) in which conformations of missing residues 62–67 were taken from the crystal structure of a BoNTAe mutant in complex with SNAP-25 (Protein Data Bank Code: 1XTG [17]).

For each of the 20 simulations, 200 instantaneous conformations were saved at 5-ps intervals during the last 1-ns period. A total of 4,000 instantaneous conformations of **AHP**•BoNTAe from the 20 simulations were subjected to a cluster analysis using the averagelinkage algorithm (epsilon = 2.0 Å and RMS on alpha-carbon atoms) [23] implemented in the PTRAJ module of the AMBER 10 program [22]. Only one cluster of the BoNTAe conformations was identified.

All 4,000 instantaneous conformations of **AHP**•BoNTAe were subjected to a second-round cluster analysis using the averagelinkage algorithm (epsilon = 2.0 Å and RMS on all atoms of **AHP**). This analysis identified five clusters of the **AHP** conformations. The numbers of the **AHP** conformations in Clusters 1–5 are 2400, 600, 721, 200, and 79, respectively. Of the three most populated clusters (Figure 4), the **AHP** conformations in Clusters 1 and 3 are similar, and the one in Cluster 2 is unique. All inhibitor conformations in Clusters 1–3 have one ammonium group forming a salt bridge to Asp370, although this group was placed away from Asp370 but close to Glu257 in the initial conformation used in the MMDSs. For the inhibitor conformations in Cluster 2, the other ammonium group moved away from Glu56 and Glu164,

and the ammonium group of Lys66 moved close to the two carboxylates to form salt bridges, although the ammonium groups of **AHP** and Lys66 were placed close to and away from the carboxylates in the initial conformation, respectively. For the inhibitor conformations in Clusters 1 and 3, the Glu56-interacting ammonium group of **AHP** and the Lys66 ammonium group remained in the same locations as those in the initial conformation.

To investigate whether **AHP** adopts two different conformations found in Clusters 1 and 2 or one of the two, a second round of 20 10-ns-long simulations on **AHP**•BoNTAe were carried out using the representative conformation of **AHP**•BoNTAe in Cluster 2 as an initial conformation. The same cluster analyses of the second-round simulations identified again one cluster of the BoNTAe conformations and five clusters of the **AHP** conformations. The numbers of the inhibitor conformations in Clusters 1–5 of the second-round simulations are 459, 201, 2762, 178, and 400, respectively. Interestingly, the most populated inhibitor conformation (Cluster 3) of the second-round simulations is nearly identical to the third most popular inhibitor conformation (Cluster 2) of the first-round. However, the most popular inhibitor conformation (Cluster 1) of the first-round simulations disappeared in the second-round, suggesting that **AHP** adopts only one conformation in the BoNTAe active site.

Deviations for all alpha-carbon atoms between the representative conformation of Cluster 3 of the second-round simulations and the free BoNTAe crystal structure (Protein Data Bank Code: 3BON [20]) that was aligned over the representative conformation were computed and plotted against temperature factors of the corresponding alpha-carbon atoms (Figure 5). A similar plot was also obtained from the average conformation of Cluster 3 of the second-round simulations (Figure 5). It is worth noting that only alpha-carbon atoms were used for the alignment and that no energy minimization was performed on the representative or average conformation. As apparent from these correlation plots, the alpha-carbon deviations of the representative and average conformations are consistent with temperature factors for all residues except for terminal residues or residues that are involved in crystal contact such as residues 197–213, 244–257, 283–295, and 375–387. The representative conformation of **AHP**•BoNTAe in Cluster 3 of the second-round simulations has a free-and-cool root mean square deviation (C&F RMSD) of 1.06 Å for all alpha-carbon atoms of the protein relative to those of the crystal structure. The C&F RMSD [24] is the RMSD of residues that are not involved in crystal contact and have temperature factors lower than the average temperature factors, and it is used to minimize the structural difference caused by crystal packing [25–27] and high temperature factors. Consistently, the average conformation of **AHP**•BoNTAe in Cluster 3 of the second-round simulations has a C&F RMSD of 0.85 Å. These results suggest that the average or representative conformation in Cluster 3 of the second-round simulations best represents the **AHP**•BoNTAe complex structure in water. The coordinates of the average and representative conformations are available from Datasets S1 and S2 of Supporting Information.

In the representative conformation of **AHP**•BoNTAe (Figures 6A and 6B), the hydroxamate group coordinates the zinc ion and has (1) a hydrogen bond to Glu224, (2) a one-water-mediated hydrogen-bond network to Tyr366, and (3) a one-water-mediated hydrogen-bond network to Arg363; the two hydroxamate-linking methylene groups have van der Waals interactions with the methyl and methylene groups of Thr220 and Glu224, respectively; the diphenylmethanol group has (1) cation-pi interactions with Arg363, (2) pi-pi interactions with Tyr366, Phe194, and Phe196,

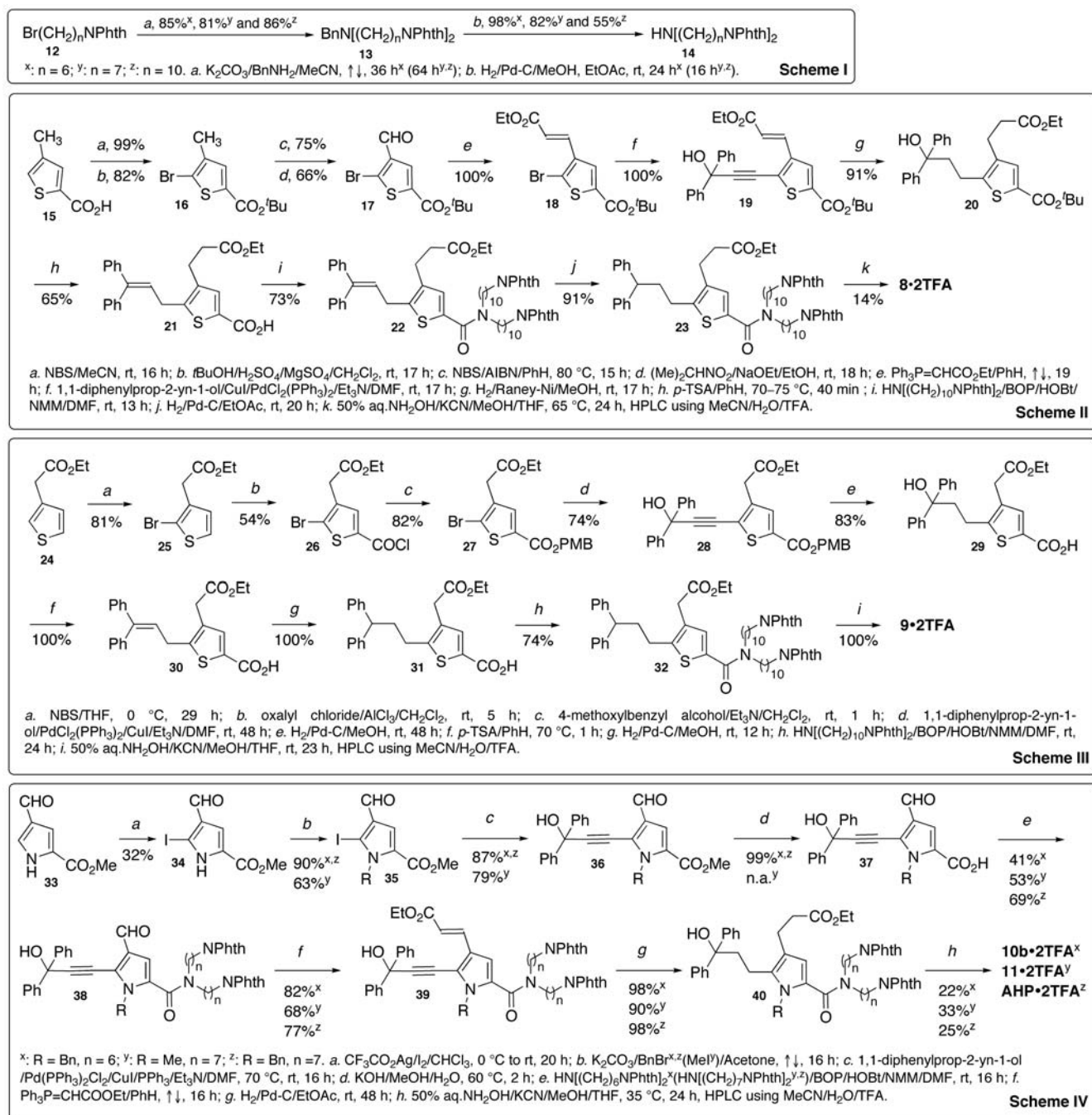


Figure 3. Syntheses of BoNTAe inhibitors 8, 9, 10b, 11, and AHP. Scheme I: synthesis of intermediate **14**; Scheme II: synthesis of **8**; Scheme III: synthesis of **9**; Scheme IV: syntheses of **10b**, **11**, and **AHP**. doi:10.1371/journal.pone.0007730.g003

(3) a hydrogen bond to Asp370, and (4) a three-water-mediated hydrogen-bond network to Asp203; the benzyl group has pi-pi interactions with Phe194 and Phe419; the pyrrole-substituted carbonyl oxygen atom has a one-water-mediated hydrogen-bond network to Gln162; one ammonium group has a salt bridge with Asp370 and an intramolecular cation-pi interaction with the diphenylmethanol group; the other ammonium group is placed at the mid point to the carboxylates of Glu55, Glu64, and Glu164 and has ionic interactions with these carboxylates; the alkyl chain of the Glu55-interacting ammonium group has van der Waals interactions with the methylene groups of Glu164, Lys66, and Gln162.

In the average conformation of **AHP**•BoNTAe (Figure 6C), there is significant contraction for the benzyl group and the alkyl ammonium group that interacts with Glu55, Glu64, and Glu164. The contraction of the benzyl group indicates that it spins in a hydrophobic pocket constituted by Val70, Ile161, Phe194, Val373, and Phe419. It also indicates that the alkyl ammonium group is highly mobile in interacting with Glu55, Glu64, and Glu164. The inhibitor dynamics revealed by the MMDSs suggests that the binding of **AHP** to BoNTAe is partly entropy-driven. It also suggests that the electron density for the benzyl group will be slightly disordered and that the electron

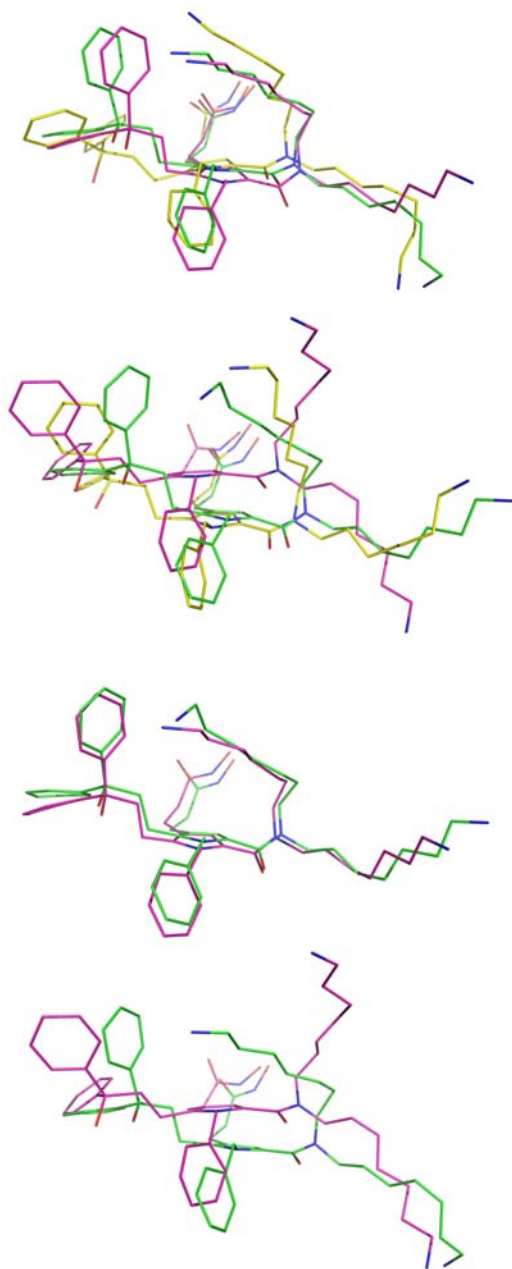


Figure 4. Overlays of AHP conformations of the three most populated clusters from the MMDSS. A (row 1): Clusters 1 (green), 2 (magenta), and 3 (yellow) from the first-round; B (row 2): Clusters 1 (yellow), 3 (green), and 5 (magenta) from the second-round; C (row 3): Cluster 2 (magenta) of the first-round and Cluster 3 (green) of the second-round; D (row 4): Cluster 1 (green) of the first-round and Cluster 5 (magenta) of the second-round.
doi:10.1371/journal.pone.0007730.g004

density for one alkyl ammonium group will be significantly disordered in the crystal structure of **AHP**•BoNTAe to be determined.

Chemical Synthesis

AHP, an *achiral* molecule possessing five functional groups (hydroxamate, diphenylmethanol, benzyl, and two ammonium groups) designed to interact with the active-site residues of BoNTAe, was made readily with a 10-step synthesis outlined in

Schemes I and IV of Figure 3. This synthesis began with methyl 4-formyl-1*H*-pyrrole-2-carboxylate (**33**) that has a pyrrole ring as a frame to support the five functional groups in different orientations. This expensive commercially available starting material was, however, made in-house from 2-(trichloroacetyl)-pyrrole using a two-step synthesis according to published protocols [28,29]. To introduce the diphenylmethanol group to the pyrrole ring via the Sonagashira coupling [21], I_2 and CF_3CO_2Ag in $CHCl_3$ [29] were used to iodinate intermediate **33** to give the desired intermediate **34**, which was separable from methyl 4-formyl-3,5-diiodo-1*H*-pyrrole-2-carboxylate and methyl 4-formyl-3-iodo-1*H*-pyrrole-2-carboxylate through column chromatography. Another reported procedure [30] was also effective in converting **33** to **34** in a similar yield. *N*-Benzoylation of **34** followed by the Sonagashira coupling [21] with 1,1-diphenylprop-2-yn-1-ol gave intermediate **36x** with both diphenylmethanol and benzyl groups linked to the pyrrole ring, which was confirmed by the heteronuclear multiple bond coherence (HMBC) spectroscopy. Saponification of **36x** followed by amidation using intermediate **14y** gave intermediate **38z** with two protected alkylamino groups linked to the pyrrole ring, wherein **14y** was obtained from *N*-(7-bromoheptyl)-phthalimide in a high yield via a two-step synthesis outlined in Scheme I of Figure 3. Wittig olefination of **38z** gave intermediate **39z** with an ethyl ester as a precursor of the desired hydroxamate group. The two unsaturated bonds of **39z** were reduced by dry 10% Pd-C in ethyl acetate to give intermediate **40z**. The hydrogenation reaction was very sensitive to moisture; wet Pd-C or Raney nickel resulted in unidentified products. The final product (**AHP**) was obtained by treating **40z** with excess of 50% aqueous hydroxylamine for 24 hours [31]. This treatment converted the ethyl ester and *N*-phthalimides to hydroxamic acid and amines, respectively [32,33]. High performance liquid chromatography (HPLC) purification using TFA-containing solvent yielded pure **AHP** in its TFA salt form. Caution should be taken in removing TFA-containing eluent after HPLC, as the diphenylmethyl hydroxy group of **AHP** is prone to elimination under acidic conditions (see Section 4.2.7).

Biological Testing

According to HPLC-based kinetics assays described in Section 4.3 [34], **AHP** has a K_i value of 760 ± 170 nM, which was calculated from slopes of Dixon plots (Figure 7) with $K_i = K_M / [(slope)(V_{MAX})(S)]$, where (S) is the substrate concentration [35] and kinetic constants for the substrate were taken from reference [36]. Unlike inhibitor **1** as we reported previously [10], **AHP** has linear Dixon plots (Figure 7) indicating that **AHP** is a competitive inhibitor of BoNTAe and its K_i value can thus be obtained from the fractional inhibition method or the Cheng and Prusoff method [35,37]. Indeed, both methods yielded a K_i value of 760 ± 170 nM, consistent with the value from the Dixon plots. **AHP** did not inhibit botulinum neurotoxin serotype B endopeptidase (BoNTBe) at concentrations up to 5 μ M at which **AHP** showed >80% inhibition of BoNTAe. As described in Section 4.3, the BoNTAe inhibition assay buffer was supplemented by 25 μ M $ZnCl_2$, whereas the BoNTBe inhibition assay buffer contained no exogenous zinc ion. **AHP** at 5 μ M showed >80% inhibition of BoNTAe in the presence of 5-molar excess of exogenous zinc, but it did not show any BoNTBe inhibition in the absence of exogenous zinc. These results preclude the possibility that **AHP** nonspecifically inhibited BoNTAe through depleting the active-site zinc divalent cation via zinc chelation. Clearly, **AHP** is a serotype-selective, small-molecule, and competitive inhibitor of BoNTAe with a K_i value of 760 ± 170 nM.

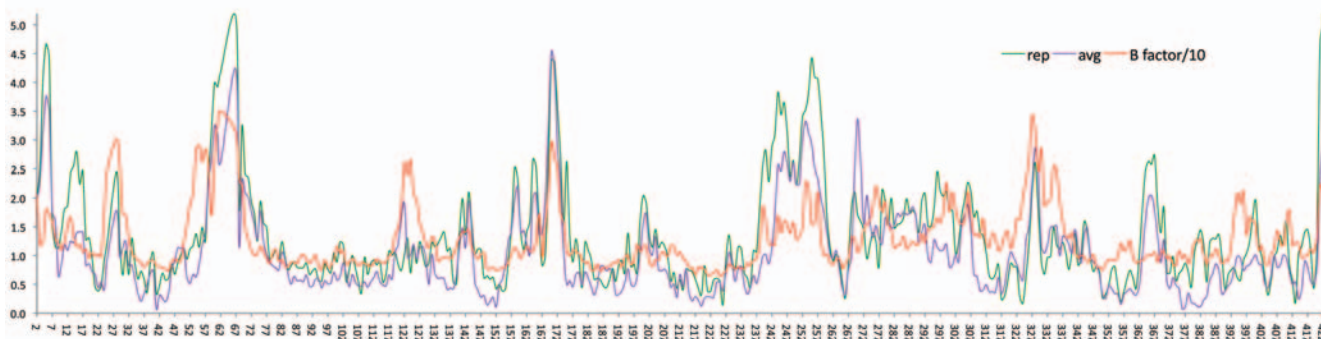


Figure 5. Deviations for alpha carbons between the BoNTAe crystal structure and the aligned MMDS-generated AHP-BoNTAe and B factors for the corresponding alpha carbons of the crystal structure. The crystal structure was taken from Protein Data Bank (Code: 3BON [20]). The representative (rep) and average (avg) conformations of AHP-BoNTAe were obtained from the second-round MMDSs. Horizontal axis: residue ID; vertical axis: alpha-carbon deviation in Å and B factors scaled by 10% in Å². doi:10.1371/journal.pone.0007730.g005

Discussion

Relative Potencies of BoNTAe Inhibitors

A mixture of two regioisomeric small molecules has been reported to have a K_i value of 600 ± 100 nM in inhibiting BoNTAe [38]; however, there has been no report for the separation of the mixture. To our knowledge, the most potent BoNTAe inhibitor has been **HPM**, a peptidic inhibitor with a K_i value of 41 nM [18]. A small-molecule BoNTAe inhibitor, 2,4-dichlorocinnamic hydroxamate (**DCCH**), was reported to have initially a K_i value of 300 ± 12 nM [39] and most recently an IC_{50} value of 59 or 81 μ M depending on assay conditions [40]. Using the assay conditions described in Section 4.3 that are different from those reported in [39,40], we found that the IC_{50} values of **AHP** and **DCCH** are <1 μ M and 34 μ M, respectively, and that **AHP** is >34 -fold more potent in inhibiting BoNTAe than **DCCH** [39] and one order of magnitude less potent than **HPM** [18]. Given this rank order of potency, below we discuss new insights into BoNTAe inhibitor design and new approaches that facilitate such design.

Insights into BoNTAe Inhibitor Design

While crystal structures of **HPM**-BoNTAe and **DCCH**-BoNTAe have been reported [18,41], the determination of the crystal structure of **AHP**-BoNTAe is currently underway. The results described in Section 2.2 suggest that the computer model of **AHP**-BoNTAe is suitable for qualitative comparison of the binding of **AHP** at the BoNTAe active site with those of **HPM** and **DCCH**.

As apparent from the cross-section view of the inhibitor-bound BoNTAe active site (Figure 8), **DCCH** occupies a small portion of the active-site region where the Arg198-containing hydrolysis product binds [17]; **AHP** occupies the bottom half of that region; **HPM** occupies nearly the entire region. The rank order of occupancy in the active site for the three inhibitors agrees well with the rank order of the inhibitory potency described in Section 3.1. From structural point of view, this agreement suggests that effective BoNTAe inhibitors have volumes that are comparable to or more than that of **AHP**. The “bulkiness” requirement observed is consistent with the reported nanomolar inhibitors of BoNTBe [42]. Therefore, we anticipate that optimization of known small-molecule BoNTAe inhibitors by small structural perturbations alone is unlikely to lead to effective BoNTAe inhibitors.

To obtain more potent nanomolar small-molecule BoNTAe inhibitors, we think that new functional groups have to be

introduced to **AHP** to occupy the upper portion of the active-site region. However, the more functional groups are introduced, the more challenging the inhibitor synthesis will become. In addition, adding more functional groups can hamper cell permeability. For these reasons, sophisticated computer simulations such as MMDSs employing the cationic dummy atom approach to simulating zinc proteins [19,43–45] are needed to identify new functional groups. These groups must have high affinities for the BoNTAe active site and low molecular weights to maintain cell permeability. In our view, for a realistic prospect of obtaining nanomolar BoNTAe inhibitors able to antagonize the intracellular BoNTA, the design and optimization of current BoNTAe inhibitor leads may need new approaches such as the nonbonded bivalence approach [46] that dimerizes composite inhibitors by intermolecular interactions (self-assembly) rather than covalent bonds in order to improve both affinity and permeability.

Synthesis-Based Computer-Aided Molecular Design

Despite the use of high-performance computing to predict inhibitor affinity and serotype selectivity in developing our small-molecule BoNTAe inhibitors, we recognized that the development process is still a *brute force* process for at least two reasons. First, exquisitely designed inhibitors such as inhibitor **2** may not be synthetically accessible within a reasonable amount of time. Second, like analog **7**, newly made inhibitors may not dissolve in an assay buffer under a high ionic strength condition. Identification of these problems *a priori* is beyond the scope of current computer simulations. The best way to circumvent these problems is to make and test more inhibitors with good affinities predicted by computer simulations. This requires facile synthesis of more of such inhibitors, which is, however, different from high-throughput screen of any available chemicals. Therefore, the ability to obtain nanomolar BoNTAe inhibitors relies more on expeditious syntheses of inhibitor analogs with good affinities than on exquisite design of high-affinity inhibitor analogs using free energy perturbation calculations that can compute the absolute binding free energy [10].

With these considerations, we pursued synthesis-based computer-aided molecular design (SBCAMD) that accounts the practicality and efficiency of inhibitor synthesis in addition to binding affinity and selectivity. The general form of our proposed approach comprises (1) identification of an inhibitor lead with the aid of computer simulations (*e.g.*, identifying **2** by MMDSs); (2) transformation of the lead to molecules that can be made in a

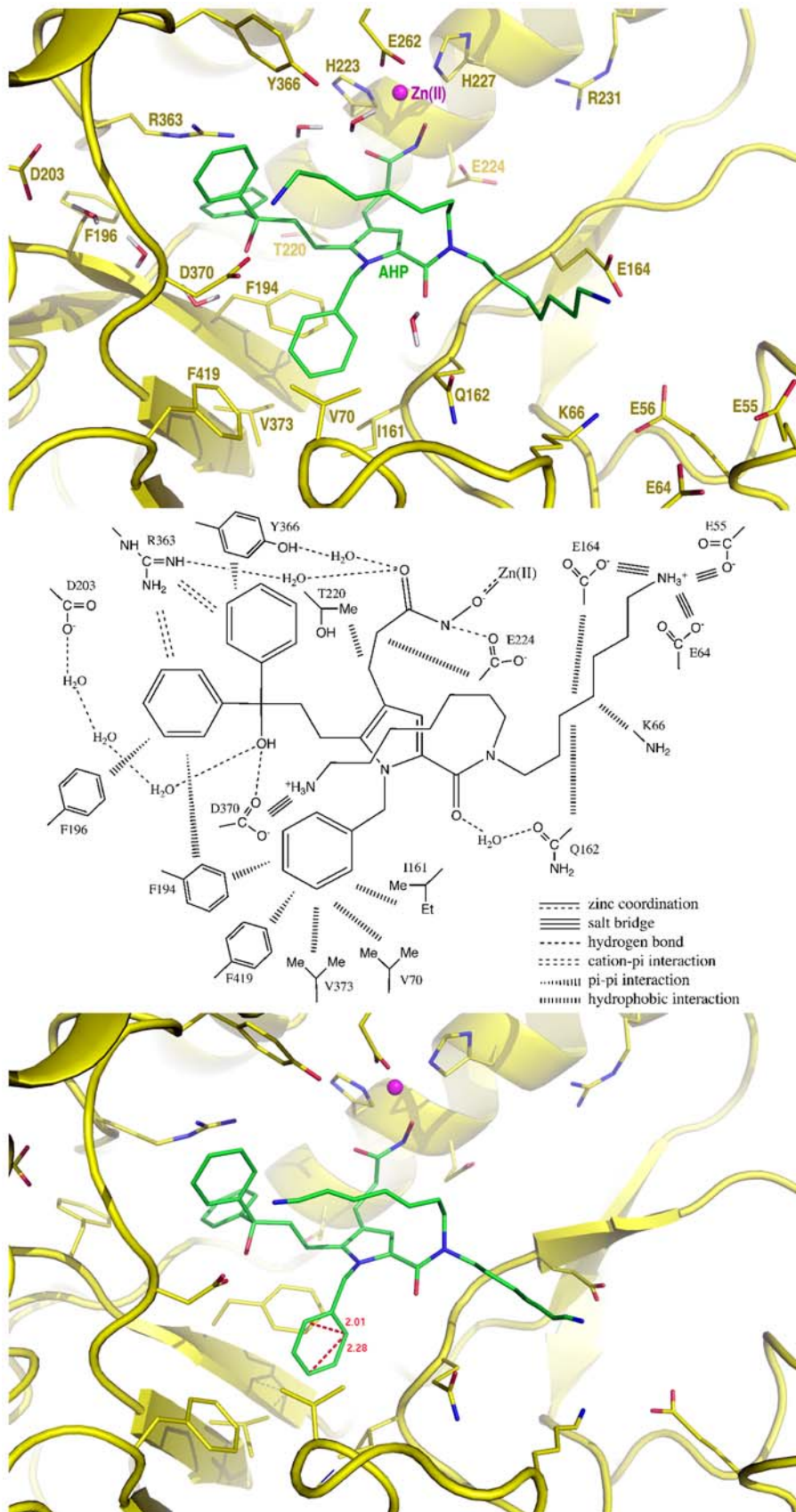


Figure 6. Close-up view of the intermolecular interactions of AHP with the active-site residues of BoNTAe. A (top): representative conformation generated by the second-round MMDSs; B (middle): diagram of the intermolecular interactions between **AHP** and BoNTAe; C (bottom): average conformation generated by the second-round MMDSs. doi:10.1371/journal.pone.0007730.g006

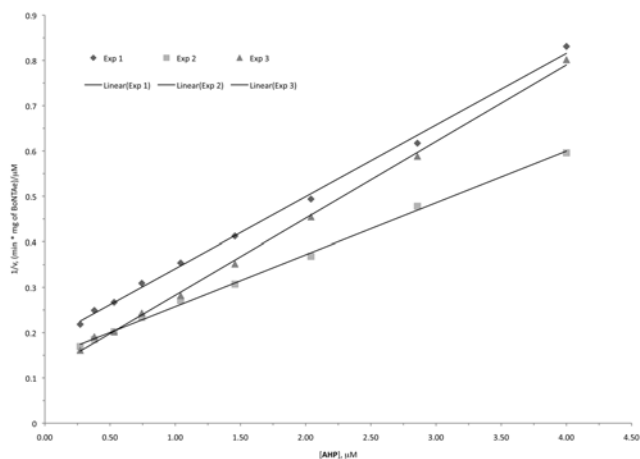


Figure 7. Dixon plots for competitive inhibition of BoNTAe by AHP.

doi:10.1371/journal.pone.0007730.g007

reasonable amount of time (*e.g.*, converting **2** to inhibitors **5–10**); (3) re-evaluation of the synthetically accessible molecules with computer simulations (*e.g.*, confirming the intermolecular interactions of **AHP** through MMDSs), this step may not be necessary if inhibitor synthesis takes less time than computer simulations; (4) synthesis and evaluation of the molecules from Step 2 or 3 (*e.g.*, synthesizing and testing **AHP**).

From synthetic chemistry point of view, SBCAMD uses a facile synthetic scheme to transform commercially available or known chemicals into a molecule that fits or likely fits the binding site of a drug target. In this case, the target molecule is determined with consideration of its synthetic effort. In contrast, CAMD uses retrosynthetic analysis [47] to repeatedly transform a target molecule as a potential inhibitor of a drug target into simpler

precursors until commercially available or known molecules are reached. In this case, like a natural product, the target molecule is pre-determined regardless of its synthetic effort.

SBCAMD is proposed to work with large-binding-pocket targets, while CAMD works best with small-binding-pocket targets. There are knowingly a number of factors (such as solubility in an assay buffer) that are beyond the scope of current computer simulations, but govern the potency of an inhibitor. Therefore, the more inhibitors are tested, the more likely a better inhibitor can be identified. The CAMD approach works well with small-binding-pocket targets, even though it does not address practicality and efficiency of chemical synthesis for each “computer-identified” inhibitor to be tested. This is because inhibitors for this kind of targets are structurally simple and often commercially available or can be made without too much synthetic effort. When bulky inhibitors are needed to bind a large protein pocket, such inhibitors are structurally complex and generally commercially unavailable. Therefore, inhibitor syntheses inevitably become the rate determine step of the development process, and hence SBCAMD comes into play.

More case studies are certainly needed, however, in our view, SBCAMD holds promise in developing small-molecule BoNTAe inhibitors or other small-molecules inhibitors that bind at large interfaces, because it aims to reduce the number of chemicals to be tested relative to the number of chemicals used in the high throughput screen and the time and cost to make each chemical to be tested.

Conclusion

We developed a serotype-selective, small-molecule, and competitive inhibitor of BoNTAe with a K_i of 760 ± 170 nM. We also developed a 3D model of BoNTAe in complex with the inhibitor, which supports the concept underlying the design of the inhibitor. These developments were made possible by the use of a new approach (SBCAMD) that aims to reduce the number of

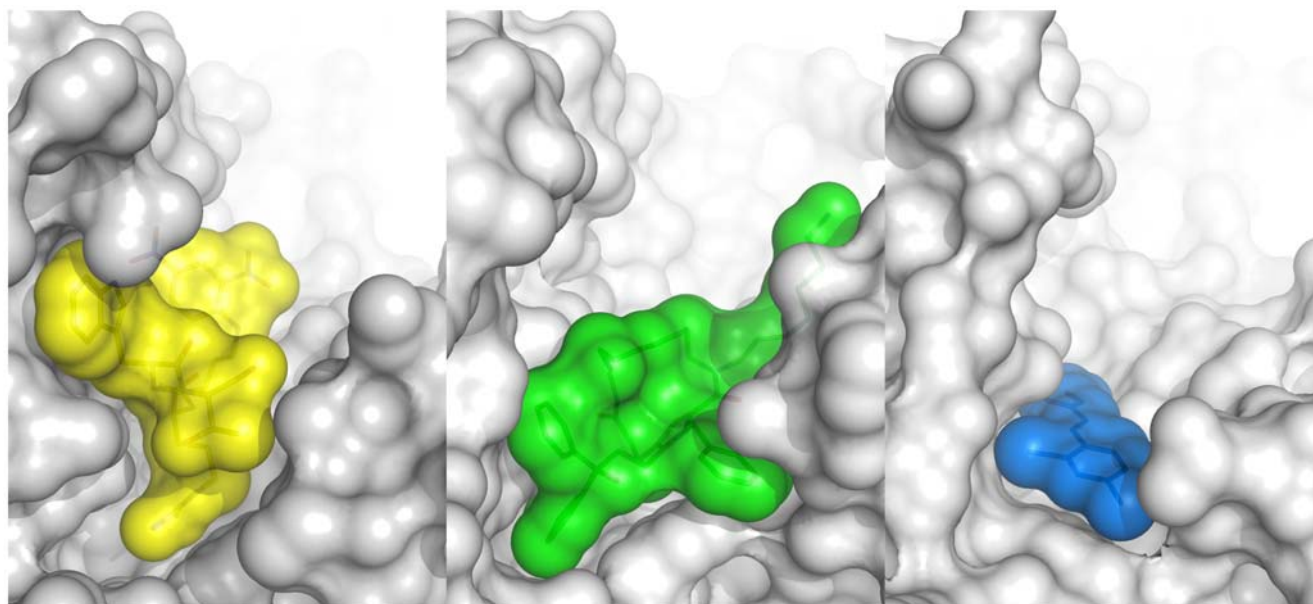


Figure 8. Cross-Section Views of HPM, AHP, and DCCH bound in the BoNTAe active site. Left: **HPM** with carbon in yellow obtained from the crystal structure of 3DS9 [18]; middle: **AHP** with carbon in green obtained from the MMDS-generated model; right: **DCCH** with carbon in blue obtained from the crystal structure of 2IMA [41]. Oxygen, nitrogen, and zinc are shown in red, blue, and light blue, respectively. BoNTAe is shown in white surface model with 15% transparency.

doi:10.1371/journal.pone.0007730.g008

chemicals to be tested as well as the time and cost to make each chemical to be tested. The results provide insights into developing effective small molecules that can neutralize the intracellular BoNTA to provide a therapy complimentary to current antibody therapy for treating botulinum. This work offers an example of the use of SBCAMD to improve expeditiously the potency of an inhibitor lead by nearly one order of magnitude for BoNTAe that is arguably one of the most challenging protein targets.

Materials and Methods

Reagents

Hexanes (Hex), ethyl acetate (EtOAc), and TFA were purchased from Fisher Scientific (Pittsburgh, PA) and used without purification. $\text{CF}_3\text{CO}_2\text{Ag}$ and 1,1-diphenylprop-2-yn-1-ol were purchased from Aldrich Chemical Co Inc (Milwaukee, WI). Dry Pd-C catalyst was purchased from Strem Chemicals Inc (Newburyport, MA). All other commercially available reagents were used as received. Recombinant BoNTAe was provided by Dr. Leonard Smith of the United States Army Medical Research Institute of Infectious Diseases, Fort Detrick, MD. BSA, HEPES buffer, and zinc chloride were purchased from Sigma-Aldrich (St. Louis, MO). Dithiothreitol was obtained from BioRad (Hercules, CA).

Chemical Synthesis

General description. The ^1H (400 MHz) and ^{13}C NMR (100 MHz) spectra were recorded on a Mercury 400 spectrometer from Varian (Palo Alto, CA). Chemical shifts are reported in ppm using either tetramethylsilane or the solvent peak as an internal standard. Data are reported as follows: chemical shift, multiplicity (s = singlet, brs = broad singlet, d = doublet, t = triplet, brt = broad triplet, q = quartet, m = multiplet), coupling constant, and integration. High-resolution mass spectra were obtained on a Bruker BioTOF II ESI. IR spectra were obtained on a Nicolet Continuum Infrared Microscope FT-IR using KBr pellet. Medium pressure liquid chromatography (MPLC) was performed with Biotage SP-1 (Charlottesville, VA) using silica gel (EM Science, 230–400 mesh). HPLC was carried out on a 5- μm C18 column (analytical: 4.60 \times 250 mm, HyperClone; semi-preparative: 21.2 \times 250 mm, Gemini) from Phenomenex (Torrance, CA) eluting with linear gradient of 80% of solution A (1000 mL of H_2O and 1 mL of TFA) to 100% of solution B (100 mL of H_2O , 900 mL of MeCN and 1 mL of TFA) over 20 minutes at a flow rate of 1.0 mL/min (analytical) or over a specified amount of time at a flow rate of 10 mL/min (semi-preparative) with UV detection at 254 nm on a Beckman Coulter System Gold HPLC system (166P detector and 125P solvent module) from Beckman Coulter (Brea, CA). Benzotriazol-1-yloxytris(dimethylamino)phosphonium hexafluorophosphate, 1-hydroxy benzotriazole, *para*-toluene sulfonic acid hydrate, and *N*-methyl morpholine are abbreviated as BOP, HOBT, *p*-TSA hydrate, and NMM, respectively. KCN is highly toxic and must be handled with extreme care by trained personnel.

Synthesis of intermediate 14. 2,2'-(6,6'-(Benzylazanediy)bis(hexane-6,1-diyl))diisoindoline-1,3-dione (**13x**). To a stirred solution of **12x** (10.73 g, 30.8 mmol) in MeCN (250 mL) was added K_2CO_3 (7.74 g, 56.0 mmol) and benzylamine (1.50 g, 14.0 mmol). The resulting solution was refluxed for 36 hours. The reaction mixture was cooled to room temperature, filtered and the filtrate was concentrated *in vacuo*. MPLC purification of the residue (Hex:EtOAc/70:30) gave **13x** as a yellow viscous oil (6.7 g, 85%). ^1H NMR (CDCl_3) δ 7.83–7.77 (m, 4H), 7.71–7.65 (m, 4H),

7.29–7.23 (m, 4H), 7.20–7.14 (m, 1H), 3.63 (t, J = 7.2 Hz, 4H), 3.48 (s, 2H), 2.34 (t, J = 7.2 Hz, 4H), 1.68–1.58 (m, 4H), 1.48–1.36 (m, 4H), and 1.33–1.24 (m, 8H); ^{13}C NMR (CDCl_3) δ 168.65, 140.32, 134.04, 132.37, 129.04, 128.27, 126.83, 123.35, 58.74, 53.77, 38.24, 28.86, 27.21, and 27.01.

2,2'-(6,6'-Azanediy)bis(hexane-6,1-diyl)diisoindoline-1,3-dione (**14x**). To a stirred solution of **13x** (1.00 g, 1.8 mmol) in MeOH:EtOAc (1:1, 20 mL each) was added 10% Pd-C (0.35 g). The resulting mixture was stirred at room temperature for 24 hours under a balloon of hydrogen. The reaction mixture was filtered through Celite and the filtrate was concentrated *in vacuo*. MPLC purification of the residue (Hex:EtOAc/70:30) gave **14x** as an amorphous white solid (0.82 g, 98%). ^1H NMR (CD_3OD) δ 7.84–7.78 (m, 8H), 3.67 (t, J = 7.2 Hz, 4H), 2.97 (t, J = 7.6 Hz, 4H), 1.74–1.62 (m, 8H), and 1.50–1.34 (m, 8H); ^{13}C NMR ($\text{DMSO-}d_6$) δ 168.65, 135.08, 132.25, 123.68, 47.27, 37.95, 28.47, 26.49, 26.28, and 26.20.

2,2'-(7,7'-(Benzylazanediy)bis(heptane-7,1-diyl))diisoindoline-1,3-dione (**13y**). Intermediate **13y** was obtained as a yellow oil (6.29 g, 81%) from **12y** (9.40 g, 29.0 mmol), K_2CO_3 (7.60 g, 55.0 mmol) and benzylamine (1.48 g, 13.8 mmol) following the procedure described for the synthesis of **13x**. ^1H NMR (CDCl_3) δ 7.86–7.80 (m, 4H), 7.72–7.66 (m, 4H), 7.30–7.24 (m, 4H), 7.24–7.16 (m, 1H), 3.65 (t, J = 7.2 Hz, 4H), 3.50 (s, 2H), 2.35 (t, J = 7.2 Hz, 4H), 1.70–1.62 (m, 4H), 1.48–1.38 (m, 4H), and 1.38–1.18 (m, 12H); ^{13}C NMR (CDCl_3) δ 168.61, 140.42, 134.04, 132.37, 129.03, 128.24, 126.79, 123.33, 58.80, 53.91, 38.24, 29.33, 28.80, 27.48, 27.13, and 27.07.

2,2'-(7,7'-Azanediy)bis(heptane-7,1-diyl)diisidoindoline-1,3-dione (**14y**). Intermediate **14y** was obtained as an amorphous white solid (1.05 g, 82%) from **13y** (1.50 g, 2.5 mmol) following the procedure described for the synthesis of **14x**. ^1H NMR (CD_3OD) δ 7.85–7.81 (m, 4H), 7.81–7.76 (m, 4H), 3.65 (t, J = 7.2 Hz, 4H), 2.56 (t, J = 7.6 Hz, 4H), 1.70–1.60 (m, 4H), 1.55–1.45 (m, 4H), and 1.45–1.30 (m, 12H); ^{13}C NMR ($\text{DMSO-}d_6$) δ 168.59, 135.03, 132.24, 123.65, 49.96, 38.03, 29.99, 29.21, 28.55, 27.37, and 26.93.

2,2'-(10,10'-(Benzylazanediy)bis(decane-10,1-diyl))diisoindoline-1,3-dione (**13z**). Intermediate **13z** was obtained as a yellow viscous oil (5.67 g, 86%) from **12z** (7.5 g, 20.5 mmol), K_2CO_3 (5.36 g, 38.8 mmol) and benzylamine (1.04 g, 9.7 mmol) following the procedure described for the synthesis of **13x**. ^1H NMR (CDCl_3) δ 7.86–7.80 (m, 4H), 7.72–7.69 (m, 4H), 7.32–7.19 (m, 5H), 3.67 (t, J = 7.2 Hz, 4H), 3.52 (s, 2H), 2.36 (t, J = 7.2 Hz, 4H), 1.70–1.60 (m, 4H), 1.47–1.38 (m, 4H), and 1.38–1.18 (m, 24H). ^{13}C NMR (CDCl_3) δ 168.62, 140.06, 134.03, 132.36, 129.10, 128.27, 126.86, 123.33, 58.69, 53.89, 38.25, 29.76, 29.72, 29.66, 29.55, 29.39, 28.81, 27.61, 27.07, and 27.02.

2,2'-(10,10'-Azanediy)bis(decane-10,1-diyl)diisoindoline-1,3-dione (**14z**). Intermediate **14z** was obtained as an amorphous white solid (602 mg, 55%) from **13z** (1.27 g, 1.87 mmol) following the procedure described for the synthesis of **14x**. ^1H NMR (CD_3OD) δ 7.86–7.81 (m, 4H), 7.81–7.76 (m, 4H), 3.65 (t, J = 7.2 Hz, 4H), 2.96 (t, J = 7.6 Hz, 4H), 1.72–1.60 (m, 8H), and 1.44–1.28 (m, 24H). ^{13}C NMR ($\text{DMSO-}d_6$) δ 168.61, 135.06, 132.23, 123.66, 47.28, 38.03, 29.44, 29.18, 28.54, 26.91, 26.66, and 26.04.

Synthesis of analog 8. *tert*-Butyl 5-bromo-4-methylthiophene-2-carboxylate (**16**). 4-Methylthiophene-2-carboxylic acid [48] (1.42 g, 10.00 mmol) was dissolved in MeCN (30 mL), followed by NBS (1.78 g, 10.00 mmol) at room temperature. After 16-hour stirring, solvent was removed *in vacuo*, the residue purified by MPLC (gradient Hex to EtOAc) to give 5-bromo-4-methylthiophene-2-

carboxylic acid (2.19 g, 99%). ^1H NMR (CDCl_3) δ 9.60 (brs, 1H), 7.56 (s, 1H), and 2.22 (s, 3H); ^{13}C NMR (CDCl_3) δ 166.92, 139.17, 136.66, 131.62, 119.99, and 15.49. The resulting acid was converted to **16** (82%) following the literature procedure [49]. ^1H NMR (CDCl_3) δ 7.38 (s, 1H), 2.18 (s, 3H), and 1.55 (s, 9H); ^{13}C NMR (CDCl_3) δ 160.90, 138.42, 134.71, 134.35, 116.95, 82.33, 28.41, and 15.46.

tert-Butyl 5-bromo-4-formylthiophene-2-carboxylate (**17**). Compound **16** (0.80 g, 2.89 mmol) and a catalytic amount of AIBN were dissolved in benzene (10 mL), followed by NBS (0.51 g, 2.89 mmol) at room temperature. The resulting mixture was heated to reflux for 15 hours. Solvent was removed *in vacuo*, the residue purified by MPLC (gradient Hex to EtOAc) to give an inseparable mixture of the starting material/desired product/dibromination by-product at a ratio of 2:7:1 in total 75% yield. ^1H NMR (CDCl_3) δ 7.58 (s, 1H), 4.40 (s, 2H), 1.55 (s, 9H). The mixture (1.68 g) was subjected to the Hass-Bender reaction [50]. MPLC purification (gradient Hex to EtOAc) afforded **17** as a pale yellow solid (0.91 g, 66%). ^1H NMR (CDCl_3) δ 9.90 (s, 1H), 7.87 (s, 1H), and 1.56 (s, 9H).

(*E*)-*tert*-Butyl 5-bromo-4-(3-ethoxy-3-oxoprop-1-enyl)thiophene-2-carboxylate (**18**). Aldehyde **17** (0.91 g, 3.13 mmol) was dissolved in benzene (12 mL), followed by (ethoxycarbonylmethylene)triphenylphosphorane (2.18 g, 6.26 mmol), and then heated at 80°C for 19 hours. Solvent was evaporated, and MPLC purification of the residue (gradient Hex to EtOAc) afforded **18** as a viscous oil (1.13 g, quantitative). ^1H NMR (CDCl_3) δ 7.71 (s, 1H), 7.60 (d, $J = 16.0$ Hz, 1H), 6.34 (d, $J = 16.0$ Hz, 1H), 4.27 (q, $J = 7.1$ Hz, 2H), 1.56 (s, 9H), and 1.34 (t, $J = 7.1$ Hz, 3H).

(*E*)-*tert*-Butyl 4-(3-ethoxy-3-oxoprop-1-enyl)-5-(3-hydroxy-3,3-diphenylprop-1-ynyl)thiophene-2-carboxylate (**19**). To a stirred mixture of compound **18** (0.34 g, 0.94 mmol), 1,1-diphenylprop-2-yn-1-ol (0.39 g, 1.87 mmol), CuI (25 mg, 0.13 mmol), and $\text{PdCl}_2(\text{PPh}_3)_2$ (46 mg, 0.07 mmol) in DMF (7 mL) was added Et_3N (1.25 mL) at room temperature. After 17-hour stirring, H_2O (5 mL) was added, extracted with EtOAc (3 \times 10 mL), the extracts were combined, washed with brine, dried over MgSO_4 , filtered, concentrated, and purified by MPLC (gradient Hex to EtOAc) to give **19** in a quantitative yield. ^1H NMR (CDCl_3) δ 7.73 (s and d, for d, $J = 15.9$ Hz, 1H each), 7.64 (d, $J = 7.6$ Hz, 4H), 7.38–7.34 (m, 4H), 7.29 (t, $J = 7.2$ Hz, 2H), 6.37 (d, $J = 15.9$ Hz), 4.21 (q, $J = 7.2$ Hz, 2H), 1.56 (s, 9H), and 1.28 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (CDCl_3) δ 166.96, 160.51, 144.42, 140.75, 136.45, 135.92, 130.05, 128.84, 128.70, 128.65, 128.57, 128.22, 126.27, 120.51, 83.16, 78.52, 75.33, 60.92, 28.36, 21.29, and 14.52.

tert-Butyl 4-(3-ethoxy-3-oxopropyl)-5-(3-hydroxy-3,3-diphenylpropyl)thiophene-2-carboxylate (**20**). To a stirred solution of **19** (0.46 g, 0.94 mmol) in MeOH (15 mL) Raney-Ni (1.0 mL as MeOH slurry) was added, and the stirring was continued under a balloon of hydrogen for 17 hours. The metal catalyst was removed with a magnet, the catalyst was washed with MeOH. The washings and the aliquot were combined, concentrated *in vacuo*, purified by MPLC (gradient Hex to EtOAc) to give **20** as an oil (0.42 g, 91%). ^1H NMR (CDCl_3) δ 7.45 (d, $J = 7.8$ Hz, 4H), 7.41 (s, 1H), 7.34–7.30 (m, 4H), 7.23 (t, $J = 7.2$ Hz, 2H), 4.09 (q, $J = 7.2$ Hz, 2H), 2.77–2.70 (m, 4H), 2.62–2.58 (m, 3H), 2.47 (t, $J = 7.5$ Hz, 2H), 1.54 (s, 9H), and 1.21 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (CDCl_3) δ 172.93, 161.79, 147.52, 146.65, 136.99, 134.24, 131.66, 128.56, 127.30, 126.15, 81.65, 78.03, 60.82, 44.10, 35.05, 28.47, 23.36, 23.08, and 14.41.

5-(3,3-Diphenylallyl)-4-(3-ethoxy-3-oxopropyl)thiophene-2-carboxylic acid (**21**). Intermediate **20** (0.38 g, 0.76 mmol) and *p*-TSA hydrate (0.15 g, 0.76 mmol) were dissolved in benzene (10 mL) at room temperature. The resulting mixture was heated at

70–75°C for 40 minutes. The reaction mixture was washed with water (10 mL), dried over MgSO_4 , concentrated, and the residue was purified by MPLC (gradient Hex to EtOAc) to give **21** as an oily residue (0.23 g, 65%). ^1H NMR (CDCl_3) δ 10.80 (brs, 1H), 7.65 (s, 1H), 7.42–7.35 (m, 4H), 7.29–7.24 (m, 6H), 6.21 (t, $J = 7.4$ Hz, 1H), 4.11 (q, $J = 7.2$ Hz, 2H), 3.61 (d, $J = 7.4$ Hz, 2H), 2.78 (t, $J = 7.6$ Hz, 2H), 2.51 (t, $J = 7.6$ Hz, 2H), and 1.22 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (CDCl_3) δ 172.72, 167.85, 149.21, 144.52, 141.95, 139.35, 137.59, 136.58, 129.94, 128.72, 128.46, 127.77, 127.68, 124.99, 60.88, 34.83, 29.21, 23.52, and 14.43.

Ethyl 3-(5-(bis(10-(1,3-dioxoisindolin-2-yl)decyl)carbamoyl)-2-(3,3-diphenylallyl)thiophen-3-yl)propanoate (**22**). To a stirred mixture of **21** (109 mg, 0.26 mmol), **14z** (152 mg, 0.26 mmol), BOP (115 mg, 0.26 mmol), and HOBT (35 mg, 0.26 mmol) in dry DMF (3 mL) was added NMM (34.2 μL , 0.31 mmol) at room temperature. After 13 hours stirring, H_2O (5 mL) was added, extracted with EtOAc (3 \times 5 mL). The extracts were combined, washed with brine, dried over MgSO_4 , filtered, concentrated, and then purified by MPLC (gradient Hex to EtOAc) to give **22** as an oil (187 mg, 73%). ^1H NMR (CDCl_3) δ 7.80–7.78 (m, 4H), 7.67–7.65 (m, 4H), 7.37–7.29 (m, 3H), 7.24–7.17 (m, 7H), 7.05 (s, 1H), 6.15 (t, $J = 7.4$ Hz, 1H), 4.06 (q, $J = 7.2$ Hz, 2H), 3.63 (t, $J = 7.1$ Hz, 4H), 3.52 (d, $J = 7.5$ Hz, 2H), 3.40–3.39 (m, 4H), 2.71 (t, $J = 7.5$ Hz, 2H), 2.44 (t, $J = 7.5$ Hz, 2H), 1.62–1.59 (m, 8H), and 1.23–1.51 (m, 24H); ^{13}C NMR (CDCl_3) δ 172.72, 168.62, 164.20, 143.60, 142.11, 142.05, 139.49, 135.75, 134.77, 134.86, 134.06, 132.36, 130.48, 130.00, 128.60, 128.36, 127.61, 127.55, 125.96, 123.33, 60.67, 38.22, 35.07, 29.69, 29.61, 29.57, 29.34, 28.80, 28.58, 27.03, 23.57, and 14.44.

Ethyl 3-(5-(bis(10-(1,3-dioxoisindolin-2-yl)decyl)carbamoyl)-2-(3,3-diphenylpropyl)thiophen-3-yl)propanoate (**23**). To a stirred solution of compound **15** (187 mg, 0.19 mmol) in EtOAc (5 mL) was added 10% Pd-C (40 mg). The resulting suspension was stirred under a balloon of hydrogen for 20 hours at room temperature. The catalyst was removed by filtration through Celite, and concentration of the filtrate gave **23** (171 mg, 91%). ^1H NMR (CDCl_3) δ 7.84–7.81 (m, 4H), 7.70–7.68 (m, 4H), 7.30–7.23 (m, 8H), 7.18–7.15 (m, 2H), 7.04 (s, 1H), 4.07 (q, $J = 7.2$ Hz, 2H), 3.98 (t, $J = 7.8$ Hz, 1H), 3.66 (t, $J = 7.2$ Hz, 4H), 3.44–3.40 (m, 4H), 2.71–2.63 (m, 4H), 2.44–2.35 (m, 4H), 1.65–1.61 (m, 8H), and 1.54–1.19 (m, 27H); ^{13}C NMR (CDCl_3) δ 172.80, 168.71, 164.26, 144.43, 142.95, 135.92, 134.33, 134.08, 132.38, 130.41, 128.81, 128.62, 128.04, 126.58, 123.38, 60.69, 50.68, 38.27, 37.49, 35.17, 29.73, 29.64, 29.60, 29.37, 28.83, 27.07, 26.33, 23.47, and 14.46.

N,N-Bis(10-aminodecyl)-5-(3,3-diphenylpropyl)-4-(3-(hydroxyamino)-3-oxopropyl)thiophene-2-carboxamide (**8**). A stirred solution of **23** (171 mg, 0.17 mmol) in MeOH:THF (2 mL each) containing 50% aqueous NH_2OH (1 mL) and a catalytic amount (two crystals) of KCN was heated at 65°C for 24 hours. The resulting solution was filtered through Celite, concentrated, redissolved in MeOH, and purified by HPLC (eluting time: 20 minutes) to give of **8-2TFA** (23 mg, 14%). Both semi-preparative and analytical HPLC retention times of **8-2TFA** are 14.83 minutes (see Figure S7 for chromatograms of **8-2TFA** before and after the HPLC purification). ^1H NMR (CD_3OD) δ 7.28–7.27 (m, 8H), 7.20–7.15 (m, 2H), 7.11 (s, 1H), 4.00 (t, $J = 7.8$ Hz, 1H), 3.50–3.40 (brs, 4H), 2.88 (t, $J = 7.2$ Hz, 4H), 2.74 (t, $J = 7.6$ Hz, 2H), 2.67 (t, $J = 7.2$ Hz, 2H), 2.39 (q, $J = 7.6$ Hz, 2H), 2.23 (t, $J = 7.2$ Hz, 2H), 1.70–1.52 (m, 8H), and 1.40–1.20 (m, 24H); ^{13}C NMR (CD_3OD) δ 170.24, 165.32, 160.34 (q), 144.59, 143.32, 136.41, 133.15, 130.77, 128.44, 128.24, 128.18, 127.73, 126.24, 125.90, 116.58 (q), 50.59, 39.54, 37.44, 33.48, 29.32, 29.20, 29.15, 28.98, 27.36, 26.58, 26.23, 25.82, and 23.61; IR cm^{-1} 3060, 3028, 2921, 2852,

1679, 1589, 1205, and 1136; HRMS-ESI calculated for $C_{43}H_{66}N_4O_3S [M+H]^+$ 719.4928, found 719.4956.

Synthesis of analog 9. Ethyl 2-(2-Bromothiophen-3-yl)acetate (**25**). To a solution of **24** (20.30 g, 119.25 mmol) in THF (150 mL) was added NBS (21.23 g, 119.25 mmol) over a period of 5 hours at 0°C, and then the mixture was warmed up to room temperature, stirring continued for 24 hours. The solvent was removed *in vacuo*, and the residue was dissolved in EtOAc (150 mL), washed with brine (4×30 mL). The organic layer was dried over $MgSO_4$, filtered, and concentrated *in vacuo*. MPLC purification (Hex:EtOAc/5:1) of the residue gave **25** as a colorless oil (24.00 g, 81%). 1H NMR ($CDCl_3$) δ 7.23 (d, $J=5.4$ Hz, 1H), 6.93 (d, $J=5.4$ Hz, 1H), 4.17 (q, $J=7.2$ Hz, 2H), 3.61 (s, 2H), and 1.27 (t, $J=7.2$ Hz, 3H); ^{13}C NMR ($CDCl_3$) δ 170.37, 133.84, 128.90, 125.93, 111.78, 61.32, 35.30, and 14.43.

Ethyl 2-(2-bromo-5-(chlorocarbonyl)thiophen-3-yl)acetate (**26**). A solution of oxalyl chloride (2.00 g, 16 mmol) in CH_2Cl_2 (3 mL) was added dropwise into a suspension of $AlCl_3$ (1.064 g, 8 mmol) in CH_2Cl_2 (6 mL) at 0°C. After the mixture was stirred for 1 hour, a solution of **25** (1.00 g, 4 mmol) in CH_2Cl_2 (2 mL) was added into the mixture during a period of 1 hour at 0°C. The mixture was allowed to warm up to room temperature and stirred for additional 3 hours. The reaction mixture was poured into ice cold H_2O (20 mL) and extracted with Et_2O (100 mL). The organic layer was washed with brine (3×15 mL), dried over $MgSO_4$, and concentrated *in vacuo*. MPLC purification (Hex:EtOAc/20:1) of the residue gave **26** as a colorless oil (682 mg, 54%). 1H NMR ($CDCl_3$) δ 7.82 (s, 1H), 4.19 (q, $J=7.2$ Hz, 2H), 3.64 (s, 2H), and 1.28 (t, $J=7.2$ Hz, 3H); ^{13}C NMR ($CDCl_3$) δ 169.30, 159.01, 139.01, 136.64, 136.46, 126.06, 61.79, 35.18, and 14.40.

4-Methoxybenzyl 5-bromo-4-(2-ethoxy-2-oxoethyl)thiophene-2-carboxylate (**27**). To a solution of 4-methoxybenzyl alcohol (220 μ L, 1.75 mmol) and Et_3N (244 μ L, 1.75 mmol) in CH_2Cl_2 (12 mL) was added dropwise a solution of **26** (367 mg, 1.17 mmol) in CH_2Cl_2 (3 mL) at room temperature. After the mixture was stirred for 1 hour at room temperature, it was poured into H_2O (20 mL) and extracted with Et_2O (50 mL). The organic layer was washed with brine (2×10 mL), dried over $MgSO_4$, and concentrated *in vacuo*. MPLC purification (Hex:EtOAc/8:1) of the residue gave **27** as a white solid (400 mg, 82%). 1H NMR ($CDCl_3$) δ 7.61 (s, 1H), 7.34 (d, $J=8.4$ Hz, 4H), 6.90 (d, $J=8.4$ Hz, 2H), 5.23 (s, 2H), 4.16 (q, $J=7.2$ Hz, 2H), 3.80 (s, 3H), 3.59 (s, 2H), and 1.26 (t, $J=7.2$ Hz, 3H); ^{13}C NMR ($CDCl_3$) δ 169.76, 161.30, 159.99, 135.14, 134.73, 133.55, 130.45, 127.80, 119.89, 114.22, 67.14, 61.56, 55.53, 35.28, and 14.40.

4-Methoxybenzyl 4-(2-ethoxy-2-oxoethyl)-5-(3-hydroxy-3,3-diphenylprop-1-ynyl)thiophene-2-carboxylate (**28**). CuI (7 mg, 0.037 mmol) was added into a mixture of **27** (214 mg, 0.51 mmol), $PdCl_2(PPh_3)_2$ (20 mg, 0.028 mmol), Et_3N (1.046 g, 10.36 mmol) and 1,1-diphenylprop-2-yn-1-ol (237 mg, 1.14 mmol) in anhydrous DMF (4 mL). The mixture was stirred at room temperature for 2 days. The mixture was poured into H_2O and extracted with Et_2O (100 mL). The organic layer was washed with brine (4×20 mL), dried over $MgSO_4$, and concentrated *in vacuo*. MPLC purification (Hex:EtOAc/6:1) of the residue gave **28** as a brown oil (206 mg, 74%). 1H NMR ($CDCl_3$) δ 7.67 (s, 1H), 7.62 (d, $J=7.6$ Hz, 4H), 7.38–7.25 (m, 8H), 6.90 (d, $J=8.8$ Hz, 2H), 5.24 (s, 2H), 4.10 (q, $J=7.0$ Hz, 2H), 3.81 (s, 3H), 3.67 (s, 2H), and 1.20 (t, $J=7.2$ Hz, 3H); ^{13}C NMR ($CDCl_3$) δ 170.18, 161.56, 159.98, 144.60, 139.49, 134.65, 133.54, 130.46, 128.64, 128.19, 127.83, 126.73, 126.24, 114.23, 101.51, 78.92, 75.28, 67.21, 61.54, 55.54, 35.33, and 14.37.

4-(2-Ethoxy-2-oxoethyl)-5-(3-hydroxy-3,3-diphenylpropyl)thiophene-2-carboxylic acid (**29**). To a solution of **28** (76 mg,

0.14 mmol) in dry MeOH (4 mL) was added 10% Pd-C (38 mg). The mixture was stirred under a balloon of hydrogen for 2 days at room temperature and filtered through Celite. Concentration of the filtrate *in vacuo* gave **29** as a colorless oil (50 mg, 83%). 1H NMR ($CDCl_3$) δ 7.66 (s, 1H), 7.44 (d, $J=7.6$ Hz, 4H), 7.33 (t, $J=7.6$ Hz, 4H), 7.28–7.22 (m, 2H), 4.11 (q, $J=7.2$ Hz, 2H), 3.44 (s, 2H), 2.82–2.75 (m, 2H), 2.67–2.60 (m, 2H), and 1.22 (t, $J=7.2$ Hz, 3H); ^{13}C NMR ($CDCl_3$) δ 170.97, 167.28, 151.87, 146.49, 137.33, 131.03, 128.88, 128.60, 128.01, 127.37, 126.71, 126.14, 78.05, 61.50, 43.73, 34.09, 29.95, 23.52, and 14.37.

5-(3,3-Diphenylallyl)-4-(2-ethoxy-2-oxoethyl)thiophene-2-carboxylic acid (**30**). To a solution of **29** (22 mg, 0.052 mmol) in benzene (6 mL) was added *p*-TSA hydrate (9 mg, 0.052 mmol). After the mixture was stirred at 70°C for 1 hour, it was poured into H_2O (20 mL) and extracted with Et_2O (50 mL). The organic layer was washed with brine (2×15 mL), dried over $MgSO_4$, and concentrated *in vacuo* to give **30** as a colorless oil (22 mg, quantitative yield). 1H NMR ($CDCl_3$) δ 7.71 (s, 1H), 7.44–7.20 (m, 10H), 6.20 (t, $J=7.4$ Hz, 1H), 4.09 (q, $J=7.2$ Hz, 2H), 3.60 (d, $J=7.2$ Hz, 2H), 3.46 (s, 2H), and 1.19 (t, $J=7.2$ Hz, 3H); ^{13}C NMR ($CDCl_3$) δ 170.63, 167.35, 150.56, 144.62, 141.88, 139.31, 137.52, 131.02, 129.93, 128.73, 128.44, 127.80, 127.68, 124.68, 104.60, 61.42, 34.13, 29.40, and 14.35.

5-(3,3-Diphenylpropyl)-4-(2-ethoxy-2-oxoethyl)thiophene-2-carboxylic acid (**31**). To a solution of **30** (22 mg, 0.054 mmol) in dry MeOH (5 mL) and EtOAc (1 mL) was added 10% Pd-C (4 mg). The mixture was stirred under a balloon of hydrogen for 12 hours at room temperature and filtered through Celite. Concentration of the filtrate *in vacuo* gave **31** as a colorless oil (22 mg, quantitative yield). 1H NMR ($CDCl_3$) δ 7.69 (s, 1H), 7.32–7.16 (m, 10H), 4.10 (q, $J=7.2$ Hz, 2H), 3.98 (t, $J=8.0$ Hz, 1H), 3.37 (s, 2H), 2.75 (t, $J=7.8$ Hz, 2H), 2.43 (q, $J=8.0$ Hz, 2H), and 1.21 (t, $J=7.2$ Hz, 3H); ^{13}C NMR ($CDCl_3$) δ 170.69, 167.27, 151.15, 144.20, 137.27, 131.21, 129.05, 128.88, 128.02, 126.72, 61.39, 50.75, 37.13, 34.05, 27.20, and 14.37.

Ethyl 2-(5-(bis(10-(1,3-dioxoisindolin-2-yl)decyl)carbamoyl)-2-(3,3-diphenylpropyl)thiophen-3-yl)acetate (**32**). A mixture of **31** (22 mg, 0.054 mmol), BOP (36 mg, 0.081 mmol), NMM (8 mg, 0.081 mmol), HOBT (11 mg, 0.081 mmol), and **14z** (47 mg, 0.081 mmol) in anhydrous DMF (4 mL) was stirred at room temperature under N_2 for ~24 hours. When the reaction was over by TLC monitoring, the mixture was poured into H_2O (10 mL) and then extracted with Et_2O (70 mL). The organic layer was washed with brine (3×10 mL), dried over $MgSO_4$, and concentrated *in vacuo*. MPLC purification (Hex/EtOAc 4:1) of the residue gave **32** as a yellow amorphous solid (32 mg, 74% based on recovery of starting material). 1H NMR ($CDCl_3$) δ 7.86–7.80 (m, 4H), 7.73–7.68 (m, 4H), 7.31–7.15 (m, 10H), 7.13 (s, 1H), 4.08 (q, $J=7.2$ Hz, 2H), 3.98 (t, $J=8.0$ Hz, 1H), 3.67 (t, $J=7.4$ Hz, 4H), 3.44 (brt, 4H), 3.33 (s, 2H), 2.70 (t, $J=7.6$ Hz, 2H), 2.40 (q, $J=7.6$ Hz, 2H), 1.70–1.57 (m, 8H), 1.36–1.23 (m, 24H), and 1.20 (t, $J=7.2$ Hz, 3H); ^{13}C NMR ($CDCl_3$) δ 170.91, 168.70, 164.02, 144.60, 144.40, 134.69, 134.08, 132.39, 130.87, 129.36, 128.80, 128.04, 126.59, 123.38, 61.15, 50.66, 38.27, 37.32, 34.18, 29.72, 29.63, 29.58, 29.37, 28.82, 27.06, 26.54, and 14.40.

N,N-Bis(10-aminodecyl)-5-(3,3-diphenylpropyl)-4-(2-(hydroxyamino)-2-oxoethyl)thiophene-2-carboxamide (**9**). To a stirred solution of **32** (18 mg, 0.018 mmol) in THF/MeOH (1.5 mL/1.5 mL), 1 mL of 50% aqueous NH_2OH was added, followed by catalytic amount (two crystals) of KCN. The resulting mixture was stirred for 23 hours at room temperature, and then filtered through a short Celite column. HPLC purification (eluting time: 30 minutes) of the filtrate gave **9-2TFA** as a white amorphous solid

(18 mg, quantitative yield). The semi-preparative and analytical HPLC retention times of **9-2TFA** are 17.70 and 15.07 minutes, respectively (see Figure S8 for chromatograms of **9-2TFA** before and after the HPLC purification). ^1H NMR (DMSO- d_6) δ 10.62 (s, 1H), 7.69 (brs, 7H), 7.35–7.10 (m, 11H), 3.98 (t, J = 7.8 Hz, 1H), 3.37 (brs, 4H), 3.10 (s, 2H), 2.72 (m, 4H), 2.64 (t, J = 7.6 Hz, 2H), 2.33 (q, J = 7.2 Hz, 2H), 1.60–1.40 (m, 8H), 1.30–1.15 (m, 24H); ^{13}C NMR (DMSO- d_6) δ 167.04, 163.29, 158.66 (q), 145.18, 143.86, 134.23, 132.01, 131.52, 129.21, 128.26, 126.91, 50.68, 39.50, 39.40, 37.26, 32.69, 29.57, 29.47, 29.44, 29.19, 27.68, 27.62, 26.89, and 26.44; IR cm^{-1} 2934, 2852, 1683, 1589, 1205, and 1136; HRMS-ESI calculated for $\text{C}_{42}\text{H}_{64}\text{N}_4\text{O}_3\text{S}$ [$\text{M}+\text{H}^+$] 705.4772, found 705.4761.

Synthesis of analog 10b. Methyl 4-formyl-5-iodo-1H-pyrrole-2-carboxylate (**33**). To a stirred solution of **33** (3.40 g, 22.2 mmol) in CHCl_3 (20 mL) at 0°C was added $\text{CF}_3\text{CO}_2\text{Ag}$ (5.40 g, 24.4 mmol) followed by adding I_2 in three portions each with 2.07 g (8.13 mmol) of the material. The resulting mixture was stirred for 16 hours in dark at room temperature. The reaction mixture was filtered through Celite and the filtrate was washed with saturated aqueous solution of $\text{Na}_2\text{S}_2\text{O}_3$. The organic layer was dried over anhydrous MgSO_4 , filtered and concentrated *in vacuo*. MPLC purification (CH_2Cl_2 : Et_2O /95:5) of the residue gave **34** as a white solid (1.90 g, 29%). Along with **34**, two minor compounds methyl 4-formyl-5-iodo-1H-pyrrole-2-carboxylate, methyl 4-formyl-3,5-diiodo-1H-pyrrole-2-carboxylate and the starting material **33** were also isolated. ^1H NMR (DMSO- d_6) δ 13.31 (brs, 1H), 9.52 (s, 1H), 7.04 (s, 1H), and 3.76 (s, 3H); ^{13}C NMR (DMSO- d_6) δ 187.13, 160.36, 128.78, 127.79, 115.58, 87.40, and 52.42.

Methyl 1-benzyl-4-formyl-5-iodo-1H-pyrrole-2-carboxylate (**35x**). To a stirred solution of **34** (0.72 g, 2.56 mmol) in acetone (20 mL) was added K_2CO_3 (0.71 g, 5.12 mmol). The resulting mixture was stirred for 30 minutes at room temperature. Benzyl bromide (0.46 mL, 3.84 mmol) was added dropwise to the reaction mixture and was refluxed for 16 hours. The reaction mixture was cooled to room temperature, concentrated and partitioned twice between ethyl acetate (10 mL) and water (10 mL). The aqueous layer was washed with EtOAc (10 mL). The combined organic layer was dried over anhydrous MgSO_4 , filtered and then concentrated *in vacuo*. MPLC purification (Hex:EtOAc/80:20) of the residue gave **35x** as a white solid (0.85 g, 90%). ^1H NMR (CDCl_3) δ 9.66 (s, 1H), 7.50 (s, 1H), 7.25–7.18 (m, 3H), 6.98 (d, J = 7.6 Hz, 2H), 5.75 (s, 2H), and 3.72 (s, 3H); ^{13}C NMR (CDCl_3) δ 186.98, 160.33, 136.43, 128.97, 128.22, 127.84, 127.10, 126.59, 119.32, 92.81, 52.81, and 52.11.

Methyl 1-benzyl-4-formyl-5-(3-hydroxy-3,3-diphenylprop-1-ynyl)-1H-pyrrole-2-carboxylate (**36x**). A solution of **35x** (0.47 g, 1.26 mmol), 1,1-diphenylprop-2-yn-1-ol (0.66 g, 3.15 mmol), CuI (0.048 g, 0.25 mmol), $\text{Pd}(\text{PPh}_3)_2\text{Cl}_2$ (0.18 g, 0.3 mmol), PPh_3 (0.066 g, 0.3 mmol), and Et_3N (4 mL) in anhydrous DMF (4 mL) was stirred for 16 hours at 70°C under N_2 atmosphere. The reaction mixture was partitioned between water and EtOAc. The combined organic layer was dried over MgSO_4 , filtered and concentrated *in vacuo*. MPLC purification (Hex:EtOAc/80:20) of the residue gave **36x** as a pale yellow solid (0.49 g, 87%). ^1H NMR (CDCl_3) δ 9.88 (s, 1H), 7.52–7.50 (m, 4H), 7.36 (s, 1H), 7.30–7.24 (m, 9H), 7.08–7.02 (m, 2H), 5.69 (s, 2H), 3.80 (s, 3H), and 3.65 (s, 1H); ^{13}C NMR (CDCl_3) δ 185.16, 160.72, 144.07, 136.84, 128.91, 128.72, 128.33, 127.91, 127.79, 127.05, 126.23, 124.96, 116.97, 103.35, 75.37, 75.27, 52.15, and 50.38. ^1H - ^{13}C HMBC (CDCl_3): there is a correlation between the pyrrole proton and the carbonyl carbon of the methyl ester, and there is no correlation between the pyrrole proton and the benzylic carbon.

1-Benzyl-4-formyl-5-(3-hydroxy-3,3-diphenylprop-1-ynyl)-1H-pyrrole-2-carboxylic acid (**37x**). To a solution of **36x** (0.49 g, 1.09 mmol)

in MeOH (20 mL) and water (6 mL) was added KOH (0.18 g, 3.27 mmol) and stirred at 60°C for 2 hours. The solvent was removed *in vacuo*. The residue obtained was acidified with 2 N HCl to pH 2–3 and extracted with EtOAc (3 \times 15 mL). The combined organic layer was dried over MgSO_4 , filtered and then concentrated *in vacuo* to give **37x** as a light brown solid. The crude product was subjected to the next reaction without further purification. ^1H NMR (CDCl_3) δ 9.82 (s, 1H), 7.52–7.50 (m, 4H), 7.45 (s, 1H), 7.30–7.18 (m, 9H), 7.06–6.99 (m, 2H), and 5.64 (s, 2H).

1-Benzyl-*N,N*-bis(6-(1,3-dioxoisindolin-2-yl)hexyl)-4-formyl-5-(3-hydroxy-3,3-diphenylprop-1-ynyl)-1H-pyrrole-2-carboxamide (**38x**). A solution of **37x** (0.35 g, 0.81 mmol), **14x** (0.46 g, 0.97 mmol), BOP (0.73 g, 1.61 mmol), HOBt (0.22 g, 1.61 mmol) in anhydrous DMF (3 mL) and NMM (2 mL) was stirred at room temperature for 16 hours. The reaction mixture was partitioned between water and CH_2Cl_2 (20 mL). The organic layer was dried over MgSO_4 , filtered and concentrated *in vacuo*. MPLC purification (Hex:EtOAc/40:60) of the residue gave **38x** as a pale yellow oil (0.30 g, 41%). ^1H NMR (CDCl_3) δ 9.86 (s, 1H), 7.82–7.75 (m, 4H), 7.70–7.62 (m, 4H), 7.56–7.54 (m, 4H), 7.27–7.21 (m, 6H), 7.15–7.14 (m, 3H), 7.02–6.98 (m, 2H), 6.58 (s, 1H), 5.34 (s, 2H), 3.68–3.52 (m, 4H), 3.28–3.18 (m, 2H), 3.04–2.94 (m, 1H), 1.68–1.59 (m, 2H), 1.58–1.46 (m, 2H), 1.42–1.27 (m, 6H), 1.18–1.04 (m, 4H), and 1.02–0.94 (m, 2H); ^{13}C NMR (CDCl_3) δ 185.34, 168.68, 162.62, 144.57, 136.84, 134.15, 132.26, 129.37, 128.87, 128.61, 128.11, 127.79, 127.29, 126.20, 125.18, 123.42, 109.14, 102.74, 75.31, 75.10, 49.57, 45.39, 37.96, 28.57, 27.13, 26.73, 26.48, and 26.28.

(*E*)-Ethyl 3-(1-benzyl-5-bis(6-(1,3-dioxoisindolin-2-yl)hexyl)carbamoyl)-2-(3-hydroxy-3,3-diphenylprop-1-ynyl)-1H-pyrrol-3-yl)acrylate (**39x**). To a stirred solution of **38x** (0.33 g, 0.37 mmol) in benzene (20 mL) was added (ethoxycarbonylmethylene)triphenylphosphorane (0.35 g, 0.92 mmol). The resulting mixture was stirred under reflux for 16 hours. The reaction mixture was cooled to room temperature and the solvent was removed *in vacuo*. MPLC purification (Hex:EtOAc/50:50) of the residue gave **39x** as a pale yellow oil (0.30 g, 85%). ^1H NMR (CDCl_3) δ 7.86–7.79 (m, 4H), 7.72–7.65 (m, 5H), 7.57–7.55 (m, 4H), 7.31–7.22 (m, 6H), 7.20–7.15 (m, 3H), 7.06–6.98 (m, 2H), 6.42 (s, 1H), 6.25 (d, J = 15.6 Hz, 1H), 5.33 (s, 2H), 4.18 (q, J = 7.2 Hz, 2H), 3.70–3.58 (m, 4H), 3.44 (s, 1H), 3.32–3.02 (m, 4H), 1.70–1.50 (m, 4H), and 1.49–1.12 (m, 15H); ^{13}C NMR (CDCl_3) δ 168.68, 167.65, 163.07, 144.63, 137.59, 136.76, 134.14, 132.32, 129.43, 128.98, 128.79, 128.66, 128.61, 128.06, 127.86, 127.77, 127.57, 126.24, 123.97, 123.42, 119.74, 115.04, 108.54, 102.26, 77.18, 75.34, 64.61, 60.36, 49.60, 38.00, 30.86, 28.64, 26.75, and 14.44.

Ethyl 3-(1-benzyl-5-(bis(6-(1,3-dioxoisindolin-2-yl)hexyl)carbamoyl)-2-(3-hydroxy-3,3-diphenylpropyl)-1H-pyrrol-3-yl)propanoate (**40x**). To a stirred solution of **39x** (0.15 g, 0.16 mmol) in EtOAc (20 mL) was added 10% dry Pd-C (0.015 g). The resulting mixture was stirred at room temperature for 48 hours under a balloon of hydrogen. The reaction mixture was filtered through Celite and the filtrate was concentrated to give **40x** as a colorless oil. The crude product was subjected to the next reaction without further purification. ^1H NMR of the crude product is as follows. ^1H NMR (CDCl_3) δ 7.77–7.75 (m, 4H), 7.64–7.62 (m, 4H), 7.27–7.06 (m, 13H), 6.71 (d, J = 7.2 Hz, 2H), 6.05 (s, 1H), 5.09 (s, 2H), 3.98 (q, J = 6.8 Hz, 2H), 3.57 (t, J = 7.2 Hz, 4H), 3.19 (t, J = 6.8 Hz, 4H), 2.56 (t, J = 7.6 Hz, 2H), 2.41–2.37 (m, 4H), 2.26–2.24 (m, 2H), 1.56–1.53 (m, 4H), and 1.30–1.11 (m, 15H).

N,N-Bis(6-aminohexyl)-1-benzyl-5-(3-(hydroxy-3,3-diphenylpropyl)-4-(3-hydroxyamino)-3-oxopropyl)-1H-pyrrole-2-carboxamide (**10b**). To a stirred solution of **40x** (0.15 g, 0.16 mmol) in MeOH:THF (3 mL each) was added 50% aqueous hydroxyl-

amine (4 mL) followed by KCN (0.04 g, 0.62 mmol). The resulting mixture was stirred at 35°C for 24 hours. After the solvent was removed *in vacuo*, the residue was subjected to HPLC purification (eluting time: 45 minutes). The eluent from HPLC was concentrated *in vacuo* at room temperature to remove the organic solvents. The resulting aqueous solution was lyophilized to give **10b-2TFA** as a white sticky solid (0.032 g, 22%). The semi-preparative and analytical HPLC retention times of **10b-2TFA** are 18.27 and 11.50 minutes, respectively (see Figure S9 for chromatograms of **10b-2TFA** before and after the HPLC purification). ¹H NMR (DMSO-*d*₆) δ 10.40 (brs, 1H), 7.70 (brs, 6H), 7.42–7.40 (m, 4H), 7.26–7.12 (m, 9H), 6.68 (d, *J* = 7.2 Hz, 2H), 6.10 (s, 1H), 5.15 (s, 2H), 3.26–3.18 (m, 4H), 3.14 (s, 1H), 2.74–2.68 (m, 4H), 2.45 (overlapped with DMSO-*d*₆, 2H), 2.34–2.28 (m, 4H), 2.17 (t, *J* = 7.2 Hz, 2H), 1.49–1.41 (m, 4H), 1.36–1.25 (m, 4H), 1.24–1.16 (m, 4H), and 1.14–1.02 (m, 4H); ¹³C NMR (DMSO-*d*₆) δ 169.44, 164.17, 158.86 (q), 148.59, 140.07, 133.78, 128.93, 128.54, 127.44, 126.86, 126.48, 126.28, 124.81, 118.30, 115.50 (q), 111.50, 76.72, 47.24, 42.30, 39.40, 34.50, 27.60, 26.41, 26.20, 21.75, and 19.05; IR cm⁻¹ 3407, 3089, 3060, 3032, 2946, 2860, 1683, 1597, 1495, 1209, and 1140; HRMS-ESI calculated for C₄₂H₅₇N₅O₄ [M+H⁺] 696.4483, found 696.4510.

Synthesis of analog 11. Methyl 4-formyl-5-iodo-1-methyl-1*H*-pyrrole-2-carboxylate (**35y**). To a stirred solution of **34** (0.30 g, 1.08 mmol) in acetone (10 mL) was added K₂CO₃ (0.30 g, 2.15 mmol). The resulting mixture was stirred for 30 minutes at room temperature. Methyl iodide (0.23 g, 1.62 mmol) was added drop wise to the reaction mixture and was refluxed for 16 hours. The reaction mixture was cooled to room temperature, concentrated and partitioned between ethyl acetate (10 mL) and water (10 mL). The aqueous layer was washed with EtOAc (2×10 mL). The combined organic layer was dried over anhydrous MgSO₄, filtered and concentrated *in vacuo*. MPLC purification (Hex:EtOAc/80:20) of the residue gave **35y** as a white solid (0.20 g, 63%). ¹H NMR (CDCl₃) δ 9.60 (s, 1H), 7.39 (s, 1H), 3.98 (s, 3H), and 3.80 (s, 3H); ¹³C NMR (CDCl₃) δ 186.84, 160.71, 128.32, 126.61, 118.76, 92.60, 52.05, and 38.00.

Methyl 4-formyl-5-(3-hydroxy-3,3-diphenylprop-1-ynyl)-1-methyl-1*H*-pyrrole-2-carboxylate (**36y**). A solution of **35y** (0.20 g, 0.68 mmol), 1,1-diphenylprop-2-yn-1-ol (0.36 g, 1.7 mmol), CuI (0.025 g, 0.14 mmol), Pd(PPh₃)₂Cl₂ (0.097 g, 0.14 mmol), PPh₃ (0.036 g, 0.14 mmol), and Et₃N (2 mL) in anhydrous DMF (3 mL) was stirred for 16 hours at 70°C under N₂ atmosphere. The reaction mixture was partitioned between water (10 mL) and EtOAc (10 mL). The aqueous layer was washed with EtOAc (2×10 mL). The combined organic layer was dried over MgSO₄, filtered and concentrated *in vacuo*. MPLC purification (Hex:EtOAc/70:30) of the residue gave **36y** as a pale yellow solid (0.20 g, 79%). ¹H NMR (CDCl₃) δ 9.74 (s, 1H), 7.65–7.63 (m, 4H), 7.35–7.32 (m, 4H), 7.28–7.27 (m, 2H), 7.23 (s, 1H), 4.75 (brs, 1H), 3.87 (s, 3H), and 3.78 (s, 3H); ¹³C NMR (CDCl₃) δ 185.27, 160.96, 144.45, 128.71, 128.28, 127.25, 126.24, 125.29, 116.40, 103.82, 75.19, 74.83, 52.10, and 34.88.

N,N-Bis(7-(1,3-dioxoisindolin-2-yl)heptyl)-4-formyl-5-(3-droxy-3,3-diphenylprop-1-ynyl)-1-methyl-1*H*-pyrrole-2-carboxamide (**38y**). To a solution of **36y** (0.10 g, 0.3 mmol) in MeOH (10 mL) and water (4 mL) was added KOH (0.06 g, 1.1 mmol) and stirred at 60°C for 2 hours. The solvent was removed *in vacuo*. The residue obtained was acidified with 2 N HCl to pH 2–3 and extracted with EtOAc (10 mL). The aqueous layer was washed with EtOAc (2×10 mL). The combined organic layer was dried over MgSO₄, filtered and concentrated *in vacuo* to give **37y** as a light brown solid. The crude product was subjected to the next reaction without further purification. A solution of **37y** (0.20 g, 0.6 mmol), **14x** (0.34 g,

0.7 mmol), BOP (0.49 g, 1.1 mmol), HOBT (0.15 g, 1.1 mmol) in anhydrous DMF (3 mL) and NMM (2 mL) was stirred at room temperature for 16 hours. The reaction mixture was partitioned between water and dichloromethane (20 mL). The aqueous layer was washed with dichloromethane (2×20 mL). The combined organic layer was dried over MgSO₄, filtered and then concentrated *in vacuo*. MPLC purification (Hex:EtOAc/50:50) of the residue gave **38y** as a pale yellow oil. (0.30 g, 53%). ¹H NMR (CDCl₃) δ 9.82 (s, 1H), 7.78–7.66 (m, 4H), 7.62–7.58 (m, 8H), 7.27–7.24 (m, 4H), 7.20–7.15 (m, 2H), 6.55 (s, 1H), 5.36 (s, 1H), 3.62–3.50 (m, 7H), 3.34 (t, *J* = 6.8 Hz, 4H), 1.66–1.38 (m, 8H), and 1.36–1.04 (m, 12H); ¹³C NMR (CDCl₃) δ 185.15, 168.66, 162.58, 144.92, 134.12, 132.21, 129.97, 128.53, 127.98, 127.28, 126.19, 125.49, 123.35, 108.30, 102.95, 74.90, 74.83, 49.20, 45.20, 38.04, 33.85, 31.60, 28.64, and 26.88.

(*E*)-Ethyl 3-(5-(bis(7-(1,3-dioxoisindolin-2-yl)heptyl)carbamoyl)-2-(3-hydroxy-3,3-diphenylprop-1-ynyl)-1-methyl-1*H*-pyrrol-3-yl)acrylate (**39y**). To a stirred solution of **38y** (0.30 g, 0.4 mmol) in benzene (10 mL) was added (ethoxycarbonylmethylene)triphenylphosphorane (0.31 g, 0.9 mmol). The resulting mixture was stirred under reflux for 16 hours. The reaction mixture was cooled to room temperature and the solvent was removed *in vacuo*. MPLC purification (Hex:EtOAc/50:50) of the residue gave **39y** as a pale yellow oil (0.22 g, 68%). ¹H NMR (CDCl₃) δ 7.75–7.66 (m, 4H), 7.63–7.57 (m, 9H), 7.28–7.25 (m, 4H), 7.19–7.18 (m, 2H), 6.31 (s, 1H), 6.17 (d, *J* = 16.0 Hz, 1H), 4.82 (s, 1H), 4.03 (q, *J* = 6.8 Hz, 2H), 3.62–3.53 (m, 7H), 3.35 (t, *J* = 7.2 Hz, 4H), 1.64–1.48 (m, 8H), and 1.36–1.15 (m, 15H); ¹³C NMR (CDCl₃) δ 168.64, 167.64, 163.06, 145.15, 136.84, 134.08, 132.20, 129.58, 128.49, 127.85, 126.25, 123.60, 123.32, 119.97, 115.60, 108.00, 102.65, 76.56, 75.07, 60.26, 49.20, 45.20, 38.03, 33.72, 28.86, 28.63, 26.90, and 14.58.

Ethyl 3-(5-(bis(7-(1,3-dioxoisindolin-2-yl)heptyl)carbamoyl)-2-(3-hydroxy-3,3-diphenylpropyl)-1-methyl-1*H*-pyrrol-3-yl)propanoate (**40y**). To a stirred solution of **39y** (0.11 g, 0.12 mmol) in EtOAc (10 mL) was added dry powdered 10% Pd-C (0.01 g). The resulting mixture was stirred at room temperature for 48 hours under a balloon of hydrogen. The reaction mixture was filtered through Celite and the filtrate was concentrated to give **40y** as a colorless oil (0.10 g, 90%). The crude product was subjected to the next reaction without further purification. ¹H NMR (CDCl₃) δ 7.82–7.80 (m, 4H), 7.70–7.68 (m, 4H), 7.46–7.44 (m, 4H), 7.32–7.20 (m, 6H), 6.03 (s, 1H), 4.04 (q, *J* = 6.8 Hz, 2H), 3.63 (t, *J* = 7.2 Hz, 4H), 3.44–3.36 (m, 7H), 2.60–2.52 (m, 4H), 2.46–2.39 (m, 4H), 2.04 (s, 1H), 1.68–1.60 (m, 4H), 1.58–1.49 (m, 4H), and 1.30–1.17 (m, 15H).

N,N-Bis(7-aminohexyl)-5-(3-hydroxy-3,3-diphenylpropyl)-4-(3-(hydroxyamino)-3-oxopropyl)-1-methyl-1*H*-pyrrole-2-carboxamide (**11**). To a stirred solution of **40y** (0.10 g, 0.11 mmol) in MeOH:THF (4 mL each) was added 50% aqueous hydroxylamine (4 mL) followed by KCN (0.04 g, 0.55 mmol). The resulting mixture was stirred at 35°C for 24 hours. After the solvent was removed *in vacuo*, the residue was subjected to HPLC purification (eluting time: 40 minutes). The eluent from HPLC was concentrated *in vacuo* at room temperature to remove the organic solvents. The resulting aqueous solution was lyophilized to give **11-2TFA** as a white sticky solid (0.032 g, 33%). The semi-preparative and analytical HPLC retention times of **11-2TFA** are 16.52 and 11.35 minutes, respectively (see Figure S10 for chromatograms of **11-2TFA** before and after the HPLC purification). ¹H NMR (DMSO-*d*₆) δ 10.38 (brs, 1H), 7.71 (brs, 6H), 7.51–7.49 (m, 4H), 7.29–7.25 (m, 4H), 7.17–7.14 (m, 2H), 5.96 (s, 1H), 3.35–3.29 (m, 7H), 2.73–2.70 (m, 4H), 2.43 (m, 2H), 2.35 (m, 4H), 2.08 (t, *J* = 7.2 Hz, 2H), 1.48–1.42 (m, 8H), and 1.23–1.18 (m, 12H); ¹³C NMR (DMSO-*d*₆) δ 169.47, 164.32, 158.98 (q), 148.65, 133.40, 128.54, 126.88, 126.36, 125.04,

117.84, 116.64 (q), 110.35, 76.85, 41.75, 39.47, 34.54, 32.05, 28.86, 27.58, 26.68, 26.39, 21.72, and 19.04; IR cm^{-1} 3428, 3085, 3056, 3023, 2934, 2856, 1679, 1597, 1495, 1201, and 1136; HRMS-ESI calculated for $\text{C}_{38}\text{H}_{57}\text{N}_5\text{O}_4$ $[\text{M}+\text{H}^+]$ 648.4483, found 648.4511.

Synthesis of AHP. 1-Benzyl-*N,N*-bis(7-(1,3-dioxoisindolin-2-yl)heptyl)-4-formyl-5-(3-hydroxy-3,3-diphenylprop-1-ynyl)-1*H*-pyrrole-2-carboxamide (**38z**). A solution of **37x** (0.49 g, 1.13 mmol), **14y** (0.68 g, 1.35 mmol), BOP (0.99 g, 2.25 mmol), HOBt (0.30 g, 2.25 mmol) in anhydrous DMF (3 mL) and NMM (2 mL) was stirred at room temperature for 16 hours. The reaction mixture was partitioned between water and CH_2Cl_2 . The organic layer was dried over MgSO_4 , filtered, and concentrated *in vacuo*. MPLC purification (Hex:EtOAc/50:50) of the residue gave **38z** as a pale yellow oil (0.72 g, 69%). ^1H NMR (CDCl_3) δ 9.90 (s, 1H), 7.82–7.74 (m, 4H), 7.68–7.64 (m, 4H), 7.56–7.54 (m, 4H), 7.28–7.17 (m, 9H), 7.03–7.00 (m, 2H), 6.59 (s, 1H), 5.36 (s, 2H), 4.37 (brs, 1H), 3.68–3.54 (m, 4H), 3.28–3.18 (m, 2H), 3.06–2.94 (m, 2H), 1.70–1.23 (m, 12H), and 1.18–0.91 (m, 8H); ^{13}C NMR (CDCl_3) δ 185.26, 168.69, 162.59, 144.52, 136.86, 134.11, 132.30, 129.55, 128.88, 128.61, 128.13, 127.84, 127.40, 126.19, 125.04, 123.39, 109.02, 102.66, 75.40, 75.13, 49.62, 38.11, 29.13, 28.69, 27.28, 27.09, 26.93, and 26.54.

(*E*)-Ethyl 3-(1-benzyl-5-bis(7-(1,3-dioxoisindolin-2-yl)heptyl)-carbamoyl)-2-(3-hydroxy-3,3-diphenylprop-1-ynyl)-1*H*-pyrrol-3-yl)acrylate (**39z**). To a stirred solution of **38z** (0.65 g, 0.71 mmol) in benzene (20 mL) was added (ethoxycarbonylmethylene)triphenylphosphorane (0.62 g, 1.78 mmol). The resulting mixture was stirred under reflux for 16 hours. The reaction mixture was cooled to room temperature and the solvent was removed *in vacuo*. MPLC purification (Hex:EtOAc/50:50) of the residue gave **39z** as a pale yellow oil (0.54 g, 77%). ^1H NMR (CDCl_3) δ 7.82–7.78 (m, 4H), 7.71–7.67 (m, 5H), 7.57–7.55 (m, 4H), 7.30–7.18 (m, 9H), 7.03–7.02 (m, 2H), 6.39 (s, 1H), 6.24 (d, $J = 16.0$ Hz, 1H), 5.32 (s, 2H), 4.17 (q, $J = 7.2$ Hz, 2H), 3.68–3.61 (m, 5H), 3.32–3.22 (m, 2H), 3.12–3.01 (m, 2H), 1.70–1.54 (m, 4H), and 1.46–1.00 (m, 19H); ^{13}C NMR (CDCl_3) δ 168.64, 167.62, 163.06, 145.11, 137.56, 136.88, 134.09, 132.21, 129.06, 128.70, 128.46, 127.79, 127.74, 126.28, 123.66, 123.32, 119.95, 115.76, 108.67, 102.81, 76.66, 75.02, 60.31, 49.38, 45.40, 38.06, 29.11, 28.64, 26.82, and 14.60.

Ethyl 3-(1-benzyl-5-(bis(7-(1,3-dioxoisindolin-2-yl)heptyl)carbamoyl)-2-(3-hydroxy-3,3-diphenylpropyl)-1*H*-pyrrol-3-yl)propanoate (**40z**). To a stirred solution of **39z** (0.12 g, 0.12 mmol) in EtOAc (20 mL) was added 10% dry Pd-C (0.012 g). The resulting mixture was stirred at room temperature for 48 hours under a balloon of hydrogen. The reaction mixture was filtered through Celite and the filtrate was concentrated to give **40z** as a colorless oil. The crude product was subjected to the next reaction without further purification. ^1H NMR (CDCl_3) δ 7.85–7.79 (m, 4H), 7.72–7.67 (m, 4H), 7.36–7.31 (m, 4H), 7.28–7.22 (m, 4H), 7.20–7.11 (m, 5H), 6.78 (d, $J = 7.2$ Hz, 2H), 6.10 (s, 1H), 5.14 (s, 2H), 4.05 (q, $J = 6.8$ Hz, 2H), 3.64 (t, $J = 7.2$ Hz, 4H), 3.25 (t, $J = 7.2$ Hz, 4H), 2.63 (t, $J = 7.6$ Hz, 2H), 2.50–2.40 (m, 4H), 2.35–2.26 (m, 2H), 1.68–1.58 (m, 4H), and 1.42–1.08 (m, 19H).

N,N-Bis(7-aminoheptyl)-1-benzyl-5-(3-(hydroxy-3,3-diphenylpropyl)-4-(3-hydroxyamino)-3-oxopropyl)-1*H*-pyrrole-2-carboxamide (**AHP**). To a stirred solution of **40z** (0.14 g, 0.14 mmol) in MeOH:THF (2 mL each) was added 50% aqueous hydroxylamine (4 mL) followed by KCN (0.04 g, 0.56 mmol). The resulting mixture was stirred at 35°C for 24 hours. After the solvent was removed *in vacuo*, the residue was subjected to HPLC purification (eluting time: 45 minutes). The eluent from HPLC was concentrated *in vacuo* at room temperature to remove the organic solvents. The resulting aqueous solution was lyophilized to give **AHP•2TFA** as a

white sticky solid (0.034 g, 25%). The semi-preparative and analytical HPLC retention times of **AHP•2TFA** are 18.60 and 12.93 minutes, respectively (see Figure S11 for chromatograms of **AHP•2TFA** before and after the HPLC purification). ^1H NMR ($\text{DMSO}-d_6$) δ 10.41 (brs, 1H), 7.71 (brs, 6H), 7.42–7.38 (m, 4H), 7.26–7.10 (m, 9H), 6.67 (d, $J = 7.2$ Hz, 2H), 6.09 (s, 1H), 5.14 (s, 2H), 3.26–3.18 (m, 4H), 2.78–2.62 (m, 4H), 2.48–2.41 (overlapped with $\text{DMSO}-d_6$, 2H), 2.35–2.28 (m, 4H), 2.11 (t, $J = 7.2$ Hz, 2H), 1.50–1.42 (m, 4H), 1.36–1.28 (m, 4H), 1.24–1.12 (m, 8H), and 1.11–1.04 (m, 4H); ^{13}C NMR ($\text{DMSO}-d_6$) δ 169.46, 164.21, 158.68 (q), 148.59, 140.08, 133.69, 128.91, 128.54, 127.39, 126.86, 126.44, 126.28, 124.93, 118.32, 117.57 (q), 111.44, 76.72, 55.61, 47.22, 42.32, 39.40, 34.57, 28.92, 27.58, 26.67, 26.34, 21.78, and 19.05; IR cm^{-1} 3416, 3085, 3056, 3023, 2934, 2860, 1687, 1614, 1495, 1450, 1205, and 1136; HRMS-ESI calculated for $\text{C}_{44}\text{H}_{61}\text{N}_5\text{O}_4$ $[\text{M}+\text{H}^+]$ 724.4796, found 724.4791.

In Vitro Evaluation of AHP

Assays of endopeptidase activities of BoNTAe and BoNTBe were done at 37°C and contained 0.5 mM substrate, 0.5–1.5 $\mu\text{g}/\text{mL}$ recombinant BoNTAe or BoNTBe, 40 mM HEPES, and 0.05% tween at pH 7.3. BoNTAe inhibition assays also contained 1 mM dithiothreitol, 25 μM ZnCl_2 , and 0.5 mg/mL BSA, while BoNTBe inhibition assays were supplemented with 1 mM dithiothreitol only. Substrate for BoNTAe was an SNAP-25 fragment containing residues 187–203 with *N*- and *C*-termini acylated and amidated, respectively [51], while that for BoNTBe was residues 60–94 of VAMP [52]. Inhibitors were dissolved in dimethyl sulfoxide at 10 times the final assay concentration, then diluted into the assay mixture containing substrate, followed by addition of the endopeptidase (*i.e.*, inhibitor and endopeptidase were not preincubated). Assay times and endopeptidase concentrations were adjusted so that less than 10% of the substrate was hydrolyzed. Assays were stopped by acidification with TFA and analyzed by reverse-phase HPLC as described [34].

Inhibition of BoNTAe by **AHP** was determined in three independent experiments using nine concentrations of **AHP** in each. K_i was calculated from slopes of Dixon plots with the equation $K_i = K_m / [(\text{slope})/(\text{V}_{\text{max}}(\text{S}))]$, where (S) was the substrate concentration [35]. Kinetic constants for the substrate were taken from reference [36].

MMDs of the AHP•BoNTAe Complex

The atomic charges of **AHP** were obtained according to the RESP procedure [53] with *ab initio* calculations at the HF/6-31G**/HF/6-31G* level using the Gaussian 98 program [54]. The starting structure of **AHP•BoNTAe** was generated by (1) manually docking **AHP** into the BoNTAe active site as described in Section 2.2 and (2) replacing the zinc ion with the tetrahedral zinc ion of the cationic dummy atom approach [19,43–45]. For BoNTAe, His223 and His227 were treated as HIN (histidinate) [44,55,56]; His39, His230, and His269 were treated as HID; all other His residues were treated as HIP; Glu261 and Glu351 were treated as GLH [44,55,56]. A total of 39 crystallographically determined water molecules (named HOH) located inside the enzyme were included for simulations. The topology and coordinate files of the water-containing **AHP•BoNTAe** complex were generated by the PREP, LINK, EDIT, and PARM modules of the AMBER 5.0 program [57]. The complex was refined by energy minimization using a dielectric constant of 1.0 and 100 cycles of steepest-descent minimization followed by 100 cycles of conjugate-gradient minimization. The refined complex was solvated with 12,098 TIP3P water molecules (named WAT) [58], leading to a system of 43,279 atoms. The WAT molecules were obtained from solvating the complex using a pre-equilibrated

box of 216,000 TIP3P molecules, whose hydrogen atom charge was set to 0.4170, where any water molecule was removed if it had an oxygen atom closer than 2.2 Å to any solute atom or a hydrogen atom closer than 2.0 Å to any solute atom, or if it was located further than 9.0 Å along the x-, y-, or z-axis from any solute atom. The solvated complex system were energy-minimized for 100 cycles of steepest-descent minimization followed by 100 cycles of conjugate-gradient minimization to remove close van der Waals contacts in the system, then heated from 0 to 300 K at a rate of 10 K/ps under constant temperature and volume, and finally simulated independently with a unique seed number for initial velocities at 300 K under constant temperature and pressure using the PMEMD module of the AMBER 8.0 program [22] with the AMBER force field (ff99SB) [59,60].

All simulations used (1) a dielectric constant of 1.0, (2) the Berendsen coupling algorithm [61], (3) a periodic boundary condition at a constant temperature of 300 K and a constant pressure of 1 atm with isotropic molecule-based scaling, (4) the Particle Mesh Ewald method to calculate long-range electrostatic interactions [62], (5) a time step of 1.0 fs, (6) the SHAKE-bond-length constraints applied to all the bonds involving the H atom, (7) saving the image closest to the middle of the “primary box” to the restart and trajectory files, (8) formatted restart file, and (9) default values of all other inputs of the PMEMD module. These simulations were carried out on a cluster of 300 Apple Xserves each equipped with two G5 processors at a clock rate of 2.0/2.3 GHz.

Average structures were obtained by using the CARNAL module of the AMBER 5.0 program. Cluster analyses were performed by using the PTRAJ module of the AMBER 10 program [22]. RMSDs were calculated by using the McLachlan algorithm [63] as implemented in the ProFit program (V2.6, <http://www.bioinf.org.uk/software/profit/>). The C&F RMSDs were obtained by (1) generating symmetry mates within 12 Å using PyMol 0.99rc6 [64], (2) identifying not-free residues that have a distance of <8.0 Å between any non-hydrogen atom of the structure in the primary cell and any non-hydrogen atom of the symmetry mates, (3) identifying hot residues that have alpha-carbon atoms with B factors greater than the average B factor for alpha carbons, (4) identifying short peptides with up to four residues that are between two hot residues, between two not-free residues, or between a hot and a not-free residue, (5) deleting the hot and not-free residues and the short peptides from the crystal structure and from the structure to be compared, and (6) computing the alpha-carbon RMSD of the truncated proteins using the ProFit program.

Supporting Information

Figure S1 Synthesis toward Analog 2.

Found at: doi:10.1371/journal.pone.0007730.s001 (0.21 MB PDF)

Figure S2 Synthesis toward Analog 6.

Found at: doi:10.1371/journal.pone.0007730.s002 (0.07 MB PDF)

Figure S3 Synthesis of Analog 3.

Found at: doi:10.1371/journal.pone.0007730.s003 (0.10 MB PDF)

References

- Arnon SS, Schechter R, Inglesby TV, Henderson DA, Bartlett JG, et al. (2001) Botulinum toxin as a biological weapon: medical and public health management. *JAMA* 285: 1059–1070.
- Shapiro RL, Hatheway C, Swerdlow DL (1998) Botulism in the United States - a clinical and epidemiologic review. *Ann Intern Med* 129: 221–228.
- Kessler KR, Benecke R (1997) Botulinum toxin—from poison to remedy. *Neurotoxicology* 18: 761–770.

Figure S4 Synthesis of Analog 4.

Found at: doi:10.1371/journal.pone.0007730.s004 (0.08 MB PDF)

Figure S5 Synthesis of Analog 5.

Found at: doi:10.1371/journal.pone.0007730.s005 (0.09 MB PDF)

Figure S6 Synthesis of Analog 7.

Found at: doi:10.1371/journal.pone.0007730.s006 (0.07 MB PDF)

Figure S7 Chromatograms of 8•2TFA before and after the HPLC purification.

Found at: doi:10.1371/journal.pone.0007730.s007 (6.12 MB PDF)

Figure S8 Chromatograms of 9•2TFA before and after the HPLC purification.

Found at: doi:10.1371/journal.pone.0007730.s008 (6.17 MB PDF)

Figure S9 Chromatograms of 10b•2TFA before and after the HPLC purification.

Found at: doi:10.1371/journal.pone.0007730.s009 (6.11 MB PDF)

Figure S10 Chromatograms of 11•2TFA before and after the HPLC purification.

Found at: doi:10.1371/journal.pone.0007730.s010 (6.45 MB PDF)

Figure S11 Chromatograms of AHP•2TFA before and after the HPLC purification.

Found at: doi:10.1371/journal.pone.0007730.s011 (6.03 MB PDF)

Dataset S1 The average conformation of AHP-bound BoNTAe in Cluster 3 of the second-round MMDSs.

Found at: doi:10.1371/journal.pone.0007730.s012 (0.38 MB TXT)

Dataset S2 The representative conformation of AHP-bound BoNTAe in Cluster 3 of the second-round MMDSs.

Found at: doi:10.1371/journal.pone.0007730.s013 (0.60 MB TXT)

Acknowledgments

We thank Christine M. McGuire for her invaluable assistance in preparing all chemicals outlined in Scheme I of Figure 3 and two anonymous reviewers for their comments and suggestions.

Author Contributions

Conceived and designed the experiments: YPP CBM JJS JD. Devised the syntheses of 4, 10b, 11, and AHP: AV. Devised the syntheses of 2, 5, 10b, 11, 14, and AHP: RKM. Devised the syntheses of 3, 6, 7, and 8: JGP. Devised the synthesis of 9: SW. Performed the experiments RKM AV JGP SW YPP JJS JD. Analyzed the data: all authors. Wrote the paper: YPP. Contributed with revisions: all authors.

7. FDA (2003) Botulism Immune Globulin Intravenous (Human): BabyBIG.
8. Simpson LL (1981) The origin, structure, and pharmacological activity of botulinum toxin. *Pharmacol Rev* 33: 155–188.
9. Boldt GE, Eubanks LM, Janda KD (2006) Identification of a botulinum neurotoxin A protease inhibitor displaying efficacy in a cellular model. *Chem Commun (Camb)*. pp 3063–3065.
10. Tang J, Park JG, Millard CB, Schmidt JJ, Pang Y-P (2007) Computer-aided lead optimization: improved small-molecule inhibitor of the zinc endopeptidase of botulinum neurotoxin serotype a. *PLoS ONE* 2: e761.
11. Burnett JC, Ruthel G, Stegmann CM, Panchal RG, Nguyen TL, et al. (2007) Inhibition of Metalloprotease Botulinum Serotype A from a Pseudo-peptide Binding Mode to a Small Molecule That Is Active in Primary Neurons. *J Biol Chem* 282: 5004–5014.
12. Johnson SL, Chen LH, Harbach R, Sabet M, Savinov A, et al. (2008) Rhodanine derivatives as selective protease inhibitors against bacterial toxins. *Chem Biol Drug Des* 71: 131–139.
13. Moe ST, Thompson AB, Smith GM, Fredenburg RA, Stein RL, et al. (2009) Botulinum neurotoxin serotype A inhibitors: Small-molecule mercaptoacetamide analogs. *Bioorg Med Chem* 17: 3072–3079.
14. Roxas-Duncan V, Enyedy I, Montgomery VA, Eccard VS, Carrington MA, et al. (2009) Identification and biochemical characterization of small-molecule inhibitors of Clostridium botulinum neurotoxin serotype A. *Antimicrob Agents Ch* 53: 3478–3486.
15. Larsen JC (2009) U.S. Army Botulinum Neurotoxin (BoNT) medical therapeutics research program: Past accomplishments and future directions. *Drug Develop Res* 70: 266–278.
16. Arkin MR, Wells JA (2004) Small-molecule inhibitors of protein-protein interactions: Progressing towards the dream. *Nat Rev Drug Dis* 3: 301–317.
17. Breidenbach MA, Brunger AT (2004) Substrate recognition strategy for botulinum neurotoxin serotype A. *Nature* 432: 925–929.
18. Zuniga JE, Schmidt JJ, Fenn T, Burnett JC, Arac D, et al. (2008) A potent peptidomimetic inhibitor of botulinum neurotoxin serotype A has a very different conformation than SNAP-25 substrate. *Structure* 16: 1588–1597.
19. Park JG, Sill PC, Makiyi EF, Garcia-Sosa AT, Millard CB, et al. (2006) Serotype-selective, small-molecule inhibitors of the zinc endopeptidase of botulinum neurotoxin serotype A. *Bioorg Med Chem* 14: 395–408.
20. Silvaggi NR, Wilson D, Tzipori S, Allen KN (2008) Catalytic features of the botulinum neurotoxin A light chain revealed by high resolution structure of an inhibitory peptide complex. *Biochemistry* 47: 5736–5745.
21. Vasilevskii SF, Sundukova TA, Shvartsberg MS, Kotlyarevskii IL (1980) Synthesis of acetylenyl-N-methylpyrroles. *Bulletin of the Academy of Sciences, USSR Division of Chemical Science (English)* 29: 1346–1350.
22. Case DA, Cheatham TE, 3rd, Darden T, Gohlke H, Luo R, et al. (2005) The Amber biomolecular simulation programs. *J Comput Chem* 26: 1668–1688.
23. Shao J, Tanner SW, Thompson N, Cheatham TE III (2007) Clustering molecular dynamics trajectories: 1. Characterizing the performance of different clustering algorithms. *J Chem Theory Comput* 3: 2312–2334.
24. Ekstrom F, Hornberg A, Artursson E, Hammarstrom LG, Schneider G, et al. (2009) Structure of HI-6-sarin-acetylcholinesterase determined by X-ray crystallography and molecular dynamics simulations: reactivator mechanism and design. *PLoS ONE* 4: e5957.
25. Eyal E, Gerzon S, Potapov V, Edelman M, Sobolev V (2005) The limit of accuracy of protein modeling: influence of crystal packing on protein structure. *J Mol Biol* 351: 431–442.
26. Wang Q, Pang Y-P (2007) Accurate reproduction of 161 small-molecule complex crystal structures using the EUDOC program: expanding the use of EUDOC to supramolecular chemistry. *PLoS ONE* 2: e531.
27. Wang Q, Pang Y-P (2007) Normal-mode-analysis-monitored energy minimization procedure for generating small-molecule bound conformations. *PLoS ONE* 2: e1025.
28. Garrido DA, Buldain G, Ojea MI, Frydman B (1988) Synthesis of 2-alkylputrescines from 3-alkylpyrroles. *J Org Chem* 53: 403–407.
29. Smith JA, Ng S, White J (2006) The regioselective synthesis of arylpyrroles. *Org Biomol Chem* 4: 2477–2482.
30. Farnier M, Fournari P (1973) Synthese d'iodoaldehydes pyrroliques. *Bulletin de la Societe Chimique de France*. pp 351–356.
31. Ho CY, Strobel E, Ralbovsky J, Galembo RA, Jr. (2005) Improved solution- and solid-phase preparation of hydroxamic acids from esters. *J Org Chem* 70: 4873–4875.
32. Mootoo DR, Fraiser-Reid B (1989) n-Pentenyl 2-amino-2-deoxy glycosides undergo stereoselective coupling under mild, chemospecific conditions. *Tetrahedron Lett* 30: 2363.
33. Ariffin A, Khan MN, Lan LC, May FY, Yun CS (2004) Suggested improved method for the Ing-Manske and related reactions for the second step of Gabriel synthesis of primary amines. *Synth Commun* 34: 4439–4445.
34. Schmidt JJ, Bostian KA (1997) Endoproteinase activity of type A botulinum neurotoxin: substrate requirements and activation by serum albumin. *J Protein Chem* 16: 19–26.
35. Segel IH (1975) Enzyme kinetics: behavior and analysis of rapid equilibrium and steady state enzyme systems. New York: Wiley xxii, 957 pp 109–111.
36. Schmidt JJ, Stafford RG (2003) Fluorogenic substrates for the protease activities of botulinum neurotoxins, serotypes A, B, and F. *Appl Environ Microbiol* 69: 297–303.
37. Cheng Y, Prusoff WH (1973) Relationship between the inhibition constant (K_i) and the concentration of inhibitor which causes 50 per cent inhibition (I₅₀) of an enzymatic reaction. *Biochem Pharmacol* 22: 3099–3108.
38. Burnett JC, Wang C, Nuss JE, Nguyen TL, Hermone AR, et al. (2009) Pharmacophore-guided lead optimization: the rational design of a non-zinc coordinating, sub-micromolar inhibitor of the botulinum neurotoxin serotype A metalloprotease. *Bioorg Med Chem Lett* 19: 5811–5813.
39. Boldt GE, Kennedy JP, Janda KD (2006) Identification of a potent botulinum neurotoxin a protease inhibitor using in situ lead identification chemistry. *Org Lett* 8: 1729–1732.
40. Pires-Alves M, Ho M, Aberle KK, Janda KD, Wilson BA (2009) Tandem fluorescent proteins as enhanced FRET-based substrates for botulinum neurotoxin activity. *Toxicol* 53: 392–399.
41. Silvaggi NR, Boldt GE, Hixon MS, Kennedy JP, Tzipori S, et al. (2007) Structures of Clostridium botulinum Neurotoxin Serotype A Light Chain complexed with small-molecule inhibitors highlight active-site flexibility. *Chem Biol* 14: 533–542.
42. Blommaert A, Turcaud S, Anne C, Roques BP (2004) Small tripeptide surrogates with low nanomolar affinity as potent inhibitors of the botulinum neurotoxin B metallo-proteolytic activity. *Bioorg Med Chem* 12: 3055–3062.
43. Pang Y-P (1999) Novel zinc protein molecular dynamics simulations: Steps toward antiangiogenesis for cancer treatment. *J Mol Model* 5: 196–202.
44. Pang Y-P, Xu K, El Yazal J, Prendergast FG (2000) Successful molecular dynamics simulation of the zinc-bound farnesyltransferase using the cationic dummy atom approach. *Protein Sci* 9: 1857–1865.
45. Pang Y-P (2001) Successful molecular dynamics simulation of two zinc complexes bridged by a hydroxide in phosphotriesterase using the cationic dummy atom method. *Proteins* 45: 183–189.
46. Pang Y-P (2004) Nonbonded bivalence approach to cell-permeable molecules that target DNA sequences. *Bioorg Med Chem* 12: 3063–3068.
47. Corey EJ (1988) Retrosynthetic thinking - essentials and examples. *Chem Soc Rev* 17: 111–133.
48. Smith K, Barratt ML (2007) Highly selective 5-substitution of 3-methylthiophene via directed lithiation. *J Org Chem* 72: 1031–1034.
49. Zhang Q, Shi C, Zhang HR, Wang KK (2000) Synthesis of 6H-indolo [2,3-b][1,6]naphthyridines and related compounds as the 5-Aza analogues of ellipticine alkaloids. *J Org Chem* 65: 7977–7983.
50. Hass HB, Bender ML (1949) The reaction of benzyl halides with the sodium salt of 2-nitropropane. A general synthesis of substituted benzaldehydes. *J Am Chem Soc* 71: 1767–1768.
51. Schmidt JJ, Stafford RG, Bostian KA (1998) Type A botulinum neurotoxin proteolytic activity - development of competitive inhibitors and implications for substrate specificity at the s-1' binding subsite. *FEBS Lett* 435: 61–64.
52. Shone CC, Roberts AK (1994) Peptide substrate specificity and properties of the zinc-endopeptidase activity of botulinum type B neurotoxin. *Eur J Biochem* 225: 263–270.
53. Cieplak P, Cornell WD, Bayly C, Kollman PA (1995) Application of the multimolecule and multiconformational resp methodology to biopolymers: charge derivation for DNA, RNA, and proteins. *J Comput Chem* 16: 1357–1377.
54. Frisch MJ, Trucks GW, Schlegel HB, Gill PMW, Johnson BG, et al. (1999) GAUSSIAN 98, Revision A.7. Gaussian, Inc Pittsburgh, PA.
55. El Yazal J, Pang Y-P (1999) *Ab initio* calculations of proton dissociation energies of zinc ligands: hypothesis of imidazole as zinc ligand in proteins. *J Phys Chem B* 103: 8773–8779.
56. El Yazal J, Roe RR, Pang Y-P (2000) Zinc's affect on proton transfer between imidazole and acetate predicted by *ab initio* calculations. *J Phys Chem B* 104: 6662–6667.
57. Pearlman DA, Case DA, Caldwell JW, Ross WS, Cheatham TE III, et al. (1995) AMBER, a package of computer programs for applying molecular mechanics, normal mode analysis, molecular dynamics and free energy calculations to simulate the structural and energetic properties of molecules. *Comput Phys Commun* 91: 1–41.
58. Jorgensen WL, Chandreskar J, Madura JD, Impey RW, Klein ML (1982) Comparison of simple potential functions for simulating liquid water. *J Chem Phys* 79: 926–935.
59. Hornak V, Abel R, Okur A, Strockbine B, Roitberg A, et al. (2006) Comparison of multiple Amber force fields and development of improved protein backbone parameters. *Proteins* 65: 712–725.
60. Wickstrom L, Okur A, Simmerling C (2009) Evaluating the performance of the ff99SB force field based on NMR scalar coupling data. *Biophys J* 97: 853–856.
61. Berendsen HJC, Postma JPM, van Gunsteren WF, Di Nola A, Haak JR (1984) Molecular dynamics with coupling to an external bath. *J Chem Phys* 81: 3684–3690.
62. Darden TA, York DM, Pedersen LG (1993) Particle Mesh Ewald: An N log(N) method for Ewald sums in large systems. *J Chem Phys* 98: 10089–10092.
63. McLachlan AD (1982) Rapid comparison of protein structures. *Acta Cryst A* 38: 871–873.
64. DeLano WL (2002) The PyMOL Molecular Graphics System. Palo Alto, CA, USA.