

AD _____

Award Number:
W81XWH-07-1-0028

TITLE:
Inhibition of Prostate Cancer Skeletal Metastases by Targeting Cathepsin K

PRINCIPAL INVESTIGATOR:
Jian Zhang, MD., PhD

CONTRACTING ORGANIZATION:
University of Pittsburgh

Pittsburgh, PA 15260

REPORT DATE:
May 2009

TYPE OF REPORT:
Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT:

Approved for public release; distribution unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

REPORT DOCUMENTATION PAGE

Form Approved
OMB No. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. **PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.**

1. REPORT DATE (DD-MM-YYYY) 01-05-2009		2. REPORT TYPE Annual		3. DATES COVERED (From - To) 4/15/2008-4/14/2009	
4. TITLE AND SUBTITLE Inhibition of prostate cancer skeletal metastases by targeting Cathepsin K				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER W81XWH-07-1-0028	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Jian Zhang Email: zhangj2@upmc.edu				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) University of Pittsburgh Pittsburgh, PA 15260				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research And Material Command Fort Detrick, Maryland 21702-5012				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for public release; distribution unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT We have previously shown that CatK is expressed not only by osteoclasts but also by prostate cancer (PCa) cells and stromal cells. Zoledronic acid (ZA), a bisphosphonate which exerts beneficial effects in PCa patients with bone metastases, reduces both pain and skeletal related events. We hypothesized that combination of a bisphosphonate with the CatK inhibitor, through different inhibitory mechanisms, could diminish PCa progression in vivo. Accordingly, PCa C4-2B cells were inoculated into the tibiae of SCID mice. The mice were randomized into the following treatment groups (n=10/group): ZA, CatK inhibitor, the combination of ZA and CatK inhibitor, and saline vehicle alone. Test drugs were initiated at 4 weeks after the tumor cell inoculation and treatments were continued for 8 weeks. We found that CatK inhibitor significantly reduced skeletal tumor burden as determined by both tumor volume vs soft tissue volume ratio and serum PSA levels. In contrast, ZA less effectively diminished the skeletal tumor volume. ZA failed to decrease PSA levels. Importantly, combinations of the two agents were more effective in decreasing tumor volume than any single agent. Using one PCa cell line, we show for the first time that a CatK inhibitor diminishes PCa growth in bone and the inhibitory effects are enhanced in combination with ZA. More cell lines need to be tested.					
15. SUBJECT TERMS Cathepsin K inhibitor, Zoledronic Acid, prostate cancer, bone metastasis, animal model					
16. SECURITY CLASSIFICATION OF: U			17. LIMITATION OF ABSTRACT UU	18. NUMBER OF PAGES 47	19a. NAME OF RESPONSIBLE PERSON USAMRMC
a. REPORT U	b. ABSTRACT U	c. THIS PAGE U			19b. TELEPHONE NUMBER (include area code)

Table of Contents

	Page
Introduction.....	4
Body.....	4-7
Key Research Accomplishments.....	7
Reportable Outcomes.....	7
Conclusion.....	7
References.....	8
Appendices.....	9-47

directly injected C4-2B cells into the tibia of SCID mice. The experimental protocols are summarized in figure 1A. In protocol 1, the mice were administered orally either CatK inhibitor or vehicle either at the time of tumor cell injection. In protocol 2, four weeks after the tumor cells injection, the mice were randomized into groups: ZA, CatK inhibitor, the combination of ZA and CatK inhibitor, and saline vehicle alone. Test drugs were given for 8 weeks. We found that CatK inhibitor significantly inhibited the establishment and development of PCa tumor in bone (Figure 1B and C). C4-2B caused dominantly osteolytic areas with some areas of osteoblastic activity (Figure 1B). It significantly reduced tumor volume (Figure 1C), serum PSA levels (Figure 1D) and osteoclast formation at bone-tumor interface (Figure 4E) compared to the vehicle-treated animals. These results indicated tumor burden was reduced by administration of the CatK inhibitor. In contrast, ZA less effectively diminished the skeletal tumor volume compared to the administration of the CatK inhibitor (Figure 1C). Interestingly, ZA failed to decrease PSA levels in these animals (Figure 1D). The inhibitory effects of the CatK inhibitor were enhanced in combination with zoledronic acid treatment (Figure 1B to 1D).

Note: one single human PCa cell line C4-2B was initially tested.

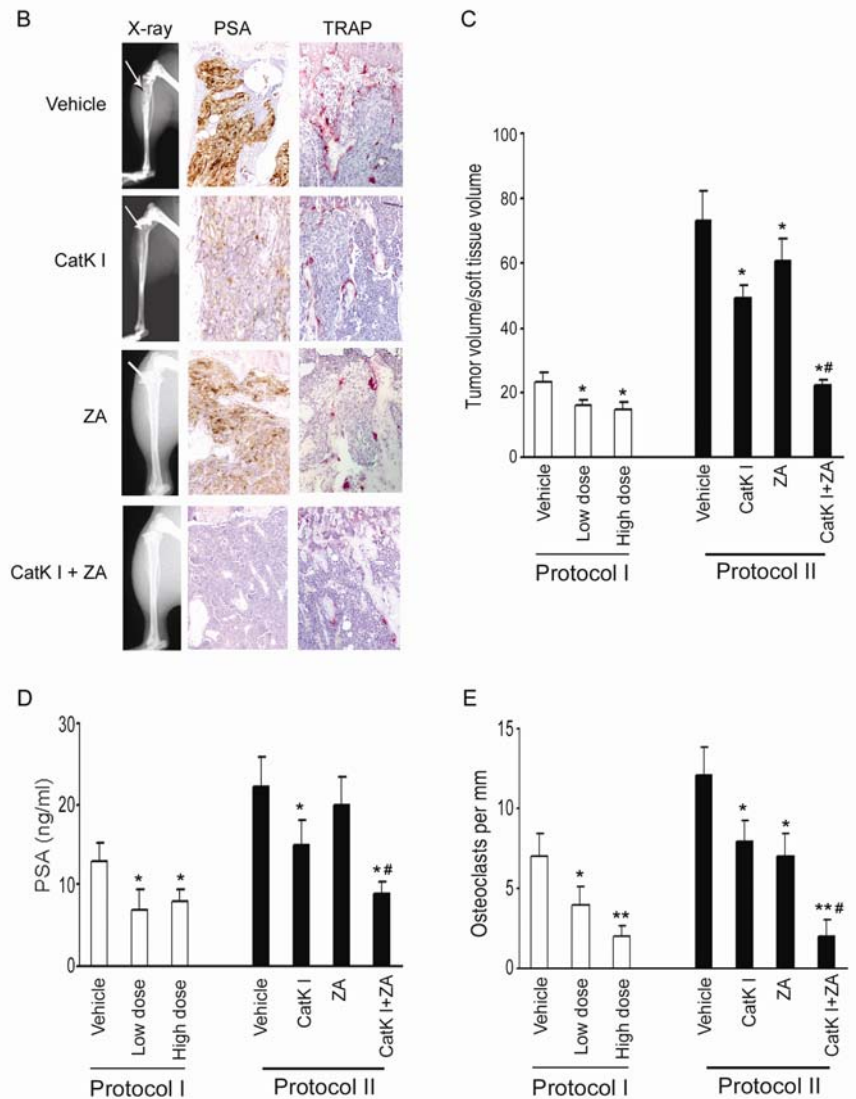
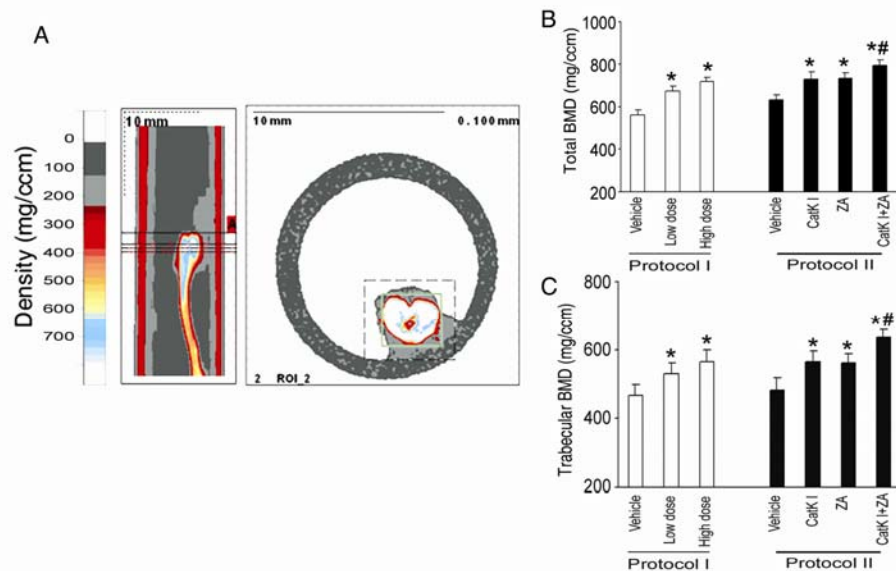


Figure 1B to 1E. CatK inhibitor prevented the PCa establishment and retarded progression of PCa tumor in mouse bone.

(B) In this representative figure, note the area of osteolysis and osteosclerosis of the vehicle-treated mouse compared to the radiograph of the CatK inhibitor-treated mouse. PSA is strong positive in all vehicle-treated mice compared to the CatK inhibitor-treated mice. (C) Tumor volume vs non-bone soft tissue volume was measured by bone histomorphometry. (D) Serum PSA levels in the mice model were measured by ELISA. (E) Osteoclast numbers per millimeter bone surface were quantified by bone histomorphometry. Results are reported as mean ± SD. *P<0.01 compared to vehicle group; #P<0.01 compare to basal group.

CatK inhibitor treatment diminished tumor-induced loss of bone mineral density (BMD) based on pQCT analysis

To further confirm the alteration of tumor-induced bone lesions by approaches other than bone histomorphometric analysis, total and trabecular BMD for the tibiae were also measured by pQCT (Figure 2A).



CatK inhibitor treatment, both at low and high doses, significantly diminished the tumor-induced loss of total and trabecular BMD (Figure 2B and C). As expected, the CatK inhibitor also increased total and trabecular BMD in both protocols. The combination of the CatK inhibitor with ZA significantly enhanced both total BMD and trabecular BMD compared to any of the single agent alone (Figure 2B and C).

Figure 2. CatK inhibitor protected bone mineral density in a mouse model. (A) Slices that

were scanned by pQCT. The small box in the right panel indicates the tibia. The reference slice and the slices examined were indicated on a representative tibia. (B) Total bone mineral density detected by pQCT of tibiae from each group. (C) Trabecular bone mineral density detected by pQCT of tibiae from each group. *P<0.01 compared to vehicle group; #P<0.01 compare to basal group.

CatK inhibitor had no effect on tumor growth at subcutaneous sites

As a parallel study for protocol 1, at the same time as the intratibial injection, C4-2B cells were injected into the subcutaneous sites. Subcutaneous tumor growth was measured for 8 weeks. Interestingly, we found CatK inhibitor had no direct effect on the tumor volume (Figure 3A) and the tumor cell proliferation *in vivo* (Figure 3B and C). These results suggest that the effect of CatK inhibitor maybe specific to the bone microenvironment.

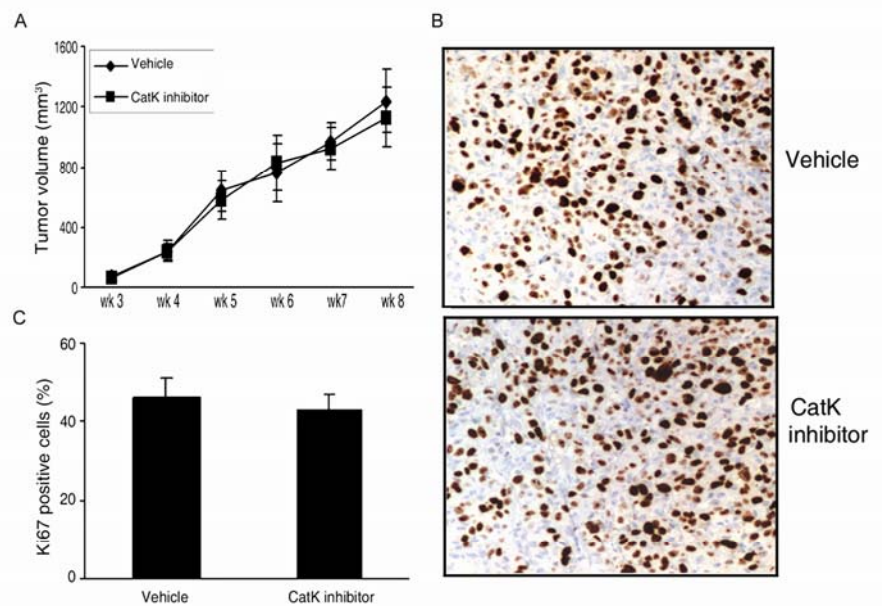


Figure 3. CatK inhibitor had no effect on subcutaneous tumor growth *in vivo*.

(A) Tumor volume were monitored twice weekly by caliper measurements. (B) Subcutaneous tumor sections were immunohistochemically stained with Ki67 antibody. (C) Quantified data were determined by the number

of Ki67 positive cells dividing the total numbers in five randomly selected fields under light microscopy ($\times 400$).

Therefore, as a preliminary in vivo study of using one single cell line, we initially showed that the CatK inhibitor significantly reduced skeletal tumor burden as determined by both tumor volume vs soft tissue volume ratio and serum PSA levels. In contrast, ZA less effectively diminished the skeletal tumor volume. ZA failed to decrease PSA levels. Importantly, combinations of the two agents were more effective in decreasing tumor volume than any single agent. Taken together, we show for the first time that a CatK inhibitor diminishes prostate cancer growth in bone and the inhibitory effects are enhanced by combination with ZA.

In our next step, we will extend this study using different tumor cells lines (LNCaP and PC3) and xenografts. The reason is that C4-2B cells caused a mixture of osteolytic and osteoblastic lesions. We have to test other cell lines that cause either osteolytic or osteoblastic lesions. As such, PC3 cells will cause osteolytic lesions; LuCaP 23.1, a PCa xenograft that, will induce highly osteoblastic lesion in bone.

In addition, the direct injection of tumor cells into the tibia has the advantage of nearly a 100% take rate, as opposed to a 20-30% take rate using the vascular approach (cardiac-injection). But we are aware that direct injection has the disadvantage of not modeling the entire metastatic cascade. Therefore, we will use cardiac-injection model that we have gained great improvement for the past 2 years.

Finally, we have collected serum and urine for measuring bone resorption makers as planned in our original application. These measurements are on-going: (a) Urinary deoxypyridinoline (Dpd), a breakdown product of collagen that indicates bone resorption. Urinary Dpd is measured using ELISA (Pyrilinks-D, Metra, Mountain View, CA). Urinary creatinine is measured to correct for urine concentration. (b) Bone-specific alkaline phosphatase (BAP), an osteoblast protein that indicates bone production, will be measured by EIA (Alkphase-B; Metra). (c) Intact osteocalcin (OCN), a non-collagenous matrix protein that indicates bone production, will be measured by competitive EIA (NovoCalcin, Metra). These will be the set of key data to successfully report the final results.

Key Research Accomplishments

1. Using one single cell line, we initially showed that the CatK inhibitor significantly reduced skeletal tumor burden as determined by both tumor volume vs soft tissue volume ratio and serum PSA levels.
2. In contrast, ZA less effectively diminished the skeletal tumor volume. ZA failed to decrease PSA levels.
3. Importantly, combinations of the two agents were more effective in decreasing tumor volume than any single agent, although these results should be determined in different cell lines and xenografts.
4. We have collected some serum and urine for measuring bone remodeling markers (on-going experiments)

Reportable Outcomes

An abstract, based on the up-to-date results, was selected as an oral presentation in Annual AACR meeting in Denver in April 2009 (Please see appendix 1) and the manuscript was submitted to the journal *Neoplasia* (Please see appendix 2).

Conclusion

From our preliminary studies, using one single cell line, we initially showed that the cathepsin K inhibitor diminishes prostate cancer growth in bone and the inhibitory effects are enhanced by combination with zoledronic acid.

References

Yi Lu, Qiuyan Chen, Eva Corey, Jinlu Dai, June Escara-Wilke, Johann Zimmermann, Evan T. Keller, Jian Zhang. A cathepsin K inhibitor diminishes prostate cancer growth in bone and the inhibitory effects are enhanced by combination with zoledronic acid. 2009 Denver AACR Proceedings abstract #2075

Targeting cathepsin K diminishes prostate cancer establishment and growth in murine bone. Yi Lu, Qiuyan Chen, Zhi Yao, Evan T. Keller, Jinlu Dai, June Escara-Wilke, Eva Corey, Johann Zimmermann, Jian Zhang *Neoplasia* (submitted)

Appendices

Please see the attached copy of AACR abstract and the manuscript.

Supporting data:

NA

2009 AACR Annual Meeting

April 18-22, 2009

Denver, CO



[Print this Page for Your Records](#)

[Close Window](#)

Abstract Number: 2075

Session Title: The Tumor Microenvironment: Opportunities for Therapeutic Intervention

Presentation Title: A cathesin K inhibitor diminishes prostate cancer growth in bone and the inhibitory effects are enhanced by combination with zoledronic acid

Presentation Start/End Time: Monday, Apr 20, 2009, 3:10 PM - 3:25 PM

Location: Room 405-407, Colorado Convention Center

Author Block: *Yi Lu, Qiuyan Chen, Eva Corey, Jinlu Dai, June Escara-Wilke, Johann Zimmermann, Evan T. Keller, Jian Zhang.* University of Pittsburgh, Pittsburgh, PA, University of Washington, Seattle, WA, University of Michigan, Ann Arbor, MI, Novartis Pharma Ltd., Basel, Switzerland

Prostate skeletal metastases are most often radiographically characterized as osteoblastic (increased bone mineral density) as opposed to osteolytic (decreased bone mineral density) lesions, it is clear that from histological evidence, however, prostate cancer skeletal metastases form a heterogeneous mixture of osteolytic and osteoblastic responses. Cathespin K (CatK) is a cysteine protease produced by osteoclasts. We and other researchers have shown that CatK is expressed not only by osteoclasts but also by prostate cancer cells and stromal cells. In addition, a highly selective CatK inhibitor has been shown to reduce the tumor-induced osteolysis and skeletal tumor burden in prostate and breast cancers in mouse models. Zoledronic acid (ZA), a bisphosphonate which exerts beneficial effects in prostate cancer patients with bone metastases, reduces both pain and skeletal related events. In this study, we hypothesized that combination of a bisphosphonate with the CatK inhibitor, through different inhibitory mechanisms, could diminish prostate cancer progression in vivo. Accordingly, human prostate cancer C4-2B cells were inoculated into the tibiae of SCID mice. The mice were randomized into the following treatment groups (n=10/group): ZA, CatK inhibitor, the combination of ZA and CatK inhibitor, and saline vehicle alone. ZA was injected s.c. twice weekly at 100ug/kg. CatK inhibitor was given at 50mg/kg by oral feeding 5 times per week. Test drugs were initiated at 4 weeks after the tumor cell inoculation and treatments were continued for 8 weeks. Histomorphometrical analysis, bone mineral density, and serum PSA measurement were used to determine the effects of treatment on the tumor growth in bone. We found that CatK inhibitor significantly reduced skeletal tumor burden as determined by both tumor volume vs soft tissue volume ratio and serum PSA levels. In contrast, ZA less effectively diminished the skeletal tumor volume. ZA failed to decrease PSA levels. Importantly, combinations of the two agents were more effective in decreasing tumor volume than any single agent. In a parallel study, C4-2B cells were injected subcutaneously into mouse flanks. The animals were given ZA, the CatK inhibitor, or vehicle (n=10). Subcutaneous tumor volume was monitored. Neither ZA nor CatK inhibitor inhibited the growth of sc tumors in animals suggesting the inhibitory effects of the two agents were bone-specific. Taken together, we show for the first time that a CatK inhibitor diminishes prostate cancer growth in bone and the inhibitory effects are enhanced by combination with ZA. This project is supported by DOD PC061231.

2009 AACR Annual Meeting

April 18-22, 2009

Denver, CO

Citation Format: {Authors}. {Abstract title} [abstract]. In: Proceedings of the 100th Annual Meeting of the American Association for Cancer Research; 2009 Apr 18-22; Denver, CO. Philadelphia (PA): AACR; 2009. Abstract nr {Abstract number}

[Disclosure Information for CME-Designated Sessions](#)

Targeting cathepsin K diminishes prostate cancer establishment and growth in murine bone¹

Yi Lu^{*, †, 2}, Qiuyan Chen^{*, ‡, 2}, Zhi Yao[†], Evan T. Keller[§], Jinlu Dai[§], June Escara-Wilke[§], Eva Corey[¶], Johann Zimmermann^{#, 3}, Jian Zhang^{*}

Abbreviations: cathepsin K, CatK; osteoprotegerin, OPG; prostate cancer, PCa; parathyroid hormone-related protein, PTHrP; zoledronic acid, ZA; Receptor activator of nuclear factor KappaB ligand, RANKL

Address all correspondence to: Jian Zhang, MD., PhD, Rm 2E110, Pittsburgh VA Healthcare Systems, R&D 151-U, University Dr., University of Pittsburgh, Pittsburgh, PA 15240. Phone: 412-527-8327; E-mail: zhangj2@upmc.edu

¹This work was supported, in part, by grants: DOD PC061231 (J Z); UPCI CCSG (J Z); SPORE for Prostate Cancer 1 P50 CA69568 (ET. K); NCI, P01 CA093900 (ET. K)

²These authors made equal contributions.

³Current address: Polyphor Ltd, Hegenheimermattweg 125, CH-4123 Allschwil, Switzerland

^{*}Department of Medicine, University of Pittsburgh, Pittsburgh, PA 15240; [†]Department of Immunology, Tianjin Medical University, Tianjin, China; [‡]Department of Nasopharyngeal Carcinoma, Cancer Center, Sun Yat-sen University, Guangzhou, China; [§]Department of Urology and Pathology, University of Michigan, Ann Arbor, MI 48109; [¶]Department of Urology, University of Washington, Seattle, WA 98195; [#]Novartis Pharma Ltd., Basel, Switzerland

Running Title: CatK inhibitor reduces prostate cancer growth in bone

Keywords: Cathepsin K, zoledronic acid, prostate cancer, skeletal metastasis, animal model

Abstract

The processes of prostate cancer (PCa) invasion and metastasis are facilitated by proteolytic cascade involving multiple proteases, such as matrix metalloproteinases, serine proteases and cysteine proteases including cathepsin K (CatK). CatK, degrades collagen I and expresses predominantly in osteoclasts, was recently demonstrated in breast cancer and PCa cells. Importantly, its expression level is greater in PCa bone metastatic sites compared to primary tumor and normal prostate tissues. The roles of CatK in PCa growth in bone, however, have not been studied. In this report, we first confirmed CatK expression in PCa LNCaP, C4-2B, and PC3 cells as well as in prostate cancer tissues. Second, we observed the inhibitory effects of a selective CatK inhibitor on PCa cell invasion. The CatK inhibitor dose-dependently inhibited PCa conditioned media-induced bone resorption. Third, we injected C4-2B cells into the tibiae of SCID mice. The animals received either vehicle or Cat K inhibitor for 8 weeks at the time of tumor cell injection (tumor establishment model; protocol 1) or 4 weeks after tumor cell injection (tumor progression model; protocol 2). Determined by radiograph, histology and bone histomorphometry, CatK inhibitor significantly prevented the tumor establishment in protocol 1, and reduced the tumor growth in bone in protocol 2. CatK inhibitor also decreased serum PSA levels in both animal models. Fourth, we demonstrated that the inhibitory effects of the CatK inhibitor were enhanced in combination with zoledronic acid (ZA). We conclude that the selective CatK inhibitor may be used as a novel therapeutic approach for advanced PCa.

Introduction

It is estimated that over 350,000 patients a year in the United States die with bone metastasis [1]. Prostate, breast, lung, and nasopharyngeal carcinoma frequently metastasizes to skeletons, and among these patients, for example, skeletal metastases were identified in up to 90% of patients dying from prostate cancer (PCa) [2-5]. PCa skeletal metastases are most often radiographically characterized as osteoblastic (increased bone mineral density) as opposed to osteolytic (decreased bone mineral density) lesions, it is clear from histological evidence, however, PCa skeletal metastases form a heterogeneous mixture of osteolytic and osteoblastic responses [6-9]. Emerging evidence shows that osteoblastic metastases form on trabecular bone at sites of previous osteoclastic resorption, and that such resorption may be required for subsequent osteoblastic bone formation [10, 11]. The mechanisms through which PCa cells promote bone resorption and subsequent woven-bone formation remain poorly understood.

A number of reports have shown that osteoclast activities are important to the development of bone metastases in several cancer types including breast, lung, and PCa [12, 13]. As such, anti-resorptive approaches such as administration of bisphosphonates, i.e. zoledronic acid (ZA) or parathyroid hormone-related protein (PTHrP) neutralizing antibody have been reported in cancer animal models to block the tumor expansion in bone [14, 15]. In PCa skeletal metastasis animal models, we and other researchers have demonstrated antiresorptive agents such as soluble receptor activator of NF κ B (sRANK) [16], osteoprotegerin (OPG) [11], and overexpression of OPG diminish the tumor growth in bone [17]. In clinics, although ZA effectively reduces metastasis-related bone pain and skeletal complications in patients with metastatic PCa and breast cancer [18-20], within a two-year period more than 30% of patients will experience at least one skeletal complication under the therapy with ZA [19-21]. Therefore, new treatments are urgently needed for patients who have bone metastases and those who are at high risk for developing bone metastases.

Cathepsin K (CatK), identified as an osteolytic and protease enzyme [22-25], degrades bone matrix proteins including type I collagen, osteopontin, and osteonectin [26, 27]. It has been revealed that CatK is

highly expressed in osteoclasts, but that cathepsins B, L and S are expressed at negligible levels [22, 23]. Patients with pycnodysostosis, a disease characterized by abnormal bone remodeling [28-30], have mutations in the *CatK* gene [31] and mice with a null mutation in the *CatK* gene develop osteopetrosis of the long bones and vertebrae [32]. Since CatK has one of the highest matrix degradation activities with higher efficiency than any other cathepsins and metalloproteinases (MMPs) [27, 33] and its ability to destroy matrix components, CatK and some of its family members have been implicated in diseases involving bone and cartilage destruction, including tumor invasion [34-36] and rheumatoid arthritis [37, 38].

CatK was reported in breast cancer cells that are capable of causing bone resorption [39] and CatK mRNA was also detected in PCa cell lines and in primary PCa and metastases [40]. Importantly, CatK expression in bone metastases was significantly greater than primary PCa, while CatK in normal prostate tissues was negative [40] suggesting that CatK may play a role in PCa skeletal metastases . Recently, a selective human CatK inhibitor has been described to potently inhibit osteoclast resorption both in vitro and in vivo [41-44]. In this study, we report that CatK contributes to PCa-induced osteoclast activity at bone metastatic sites, and inhibition of CatK by a selective inhibitor may prevent the establishment and progression of PCa in bone. We finally demonstrated that the inhibitory effects of the CatK inhibitor were enhanced in combination with zoledronic acid (ZA).

Materials and Methods

Cell Lines and Cell Culture

PCa PC3 and LNCaP cells were purchased from the American Type and culture Collection (ATCC, Manassas, VA) and were cultured in RPMI 1640 medium. C4-2B cells (UroCor, Oklahoma City, OK), derived from its parental LNCaP but with characteristics of skeletal metastasis, were maintained in T medium [80% DMEM, 20% Ham's F12 medium (Invitrogen, Carlsbad, CA), 5 µg/mL insulin, 13.6 pg/mL triiodothyronine, 5 µg/mL transferrin, 0.25 µg/mL biotin, and 25 µg/mL adenine (Sigma, St. Louis, MO)]. Primary murine bone marrow cells (MBMC) were cultured in α MEM medium. All cell cultures were supplemented with 1% penicillin/streptomycin (Invitrogen, Carlsbad, CA) and 10% fetal bovine serum (FBS) (HyClone, Pittsburgh, PA). Prostate epithelial cells (PrEC) are primary human epithelial cells (Cambrex, Walkersville, MD) and were maintained in PrEGM BulletKit media (Cambrex). All cells were maintained in a 37 °C incubator equilibrated with 5% CO₂.

Animals

Male SCID mice (Charles River, Wilmington, MA) at 6 weeks of age were housed under pathogen-free conditions in accordance with the NIH guidelines. The animal protocol was approved by the Institutional Animal Care and Use Committee, both from the University of Pittsburgh and the University of Michigan.

CatK Inhibitor

Cathepsin K selective inhibitor and zoledronic acid used in these studies was provided by Novartis Pharma Ltd., (Basel, Switzerland). The inhibitor dosages in this *in vivo* study were chosen as 50mg/kg/day and 100mg/kg/day based on the renal toxicity of dose 500mg/kg/day in rat and preclinical efficacy and tolerability dose of 50mg daily used in human.

Immunohistochemical Staining for CatK

Tissue microarray of PCa specimens (83 Asian patients who underwent radical prostatectomy with Gleason score 5-10: n=83) were purchased from IU Abxis (Seoul, Korea) with corresponding non-neoplastic tissues (42 specimens). Slides were heated at 55°C for 30 minutes, deparaffinized and rehydrated, then antigen retrieval was performed with Target Retrieval kit from Dako (Carpinteria, CA). Slides were incubated for 24 h at 4°C with goat anti-human CatK polyclonal antibody (Santa Cruz Biotech, Santa Cruz, CA; 1:200 dilution), or isotype control goat IgG (Santa Cruz; 1:200 dilution). Biotinylated anti-goat (Dako) at a dilution of 1:500 was used as the secondary antibody. After incubation in avidin-biotin complex solution (Dako), the staining was developed by the diaminobenzidine method, followed by counterstaining with hematoxylin.

Obtaining Conditioned Media (CM)

CM were obtained from PCa cells as previously described [45]. Briefly, 2×10^6 cells were plated in 10-cm tissue culture dishes for 12 hours in RPMI 1640 with 10% FBS. The media were then changed to 10 ml of RPMI 1640 plus 1% FBS, and supernatants were collected 48 hours later. To normalize for differences in cell density because of proliferation during the culture period, cells from each plate were collected and total DNA content/plate was determined. CM were then normalized for DNA content between samples by adding RPMI.

CatK mRNA Expression and Quantification CatK mRNA

Total RNA was extracted from LNCaP, C4-2B, PC3 and PrEC cells using TRIzol reagent (Life Technologies, Gaithersburg, MD), then subjected to RT-PCR for detection Cat K mRNA. PCR primers used for detection of CatK consisted of sense 5'-CAG CAA AGG TGT GTA TTA TGA TGA AAG C-3', and antisense 5'-ATG GGT GGA GAG AAG CAA AGT AGG AAG G-3' resulting in a PCR product of 399 bp (Genebank accession no. X82153). Beta-actin cDNA was amplified as a control for RNA quality. The PCR products were subjected to eletrophoresis on a 1.5% agarose gel, stained with ethidium bromide. For quantification of CatK

mRNA expression, real-time RT-PCR was performed in an iCycler iQ multicolor real-time RT-PCR detection system (Bio-Rad, Hercules, CA) using the iScript one-step RT-PCR kit with SYBR Green (Bio-Rad). Melting curve analysis was performed to evaluate purity of the PCR products. Triplicate samples were run for each primer set. The relative expression of CatK to GAPDH (as housekeeping gene control) was calculated using the Δ CT method as previously described [46]. CatK primers were: sense 5'-CAG CAG AGG TGT GTA CTA TG-3' and antisense 5'-GCG TTG TTC TTA TTC CGA GC-3'. GAPDH primers were: sense 5'-CCA TGG AGA AGG CTG GGG-3' and antisense 5'-CAA AGT TGT CAT GGA TGA CC-3'.

Immunoblot Analysis

To evaluate the CatK protein expression in PCa cells, confluent LNCaP, C4-2B, PC3 and PrEC cells were washed with PBS and lysed. Protein concentration of the cell lysates was measured by BCA kit (Pierce, Rockford, IL) and proteins (40 μ g/lane) were applied to SDS-PAGE followed by Western blot with goat anti-human CatK polyclonal Antibody (diluted 1:100 in blocking solution) (Santa Cruz Biotech, Santa Cruz, CA), or mouse anti-human β -actin monoclonal antibody (Sigma; diluted 1:2000 in blocking solution)). The antibody binding was revealed using an HRP-conjugated anti-goat IgG, or HRP-conjugated anti-mouse IgG1 (Santa Cruz) at 1:5000 and enhanced chemiluminescence (ECL) blot detection system (Amersham Biosciences, Piscataway, New Jersey).

ELISA

To evaluate CatK protein production by PCa cells, CM collected from LNCaP, C4-2B, PC3 and PrEC cell cultures were measured by CatK ELISA kit (Alpco, Windham, NH) following the manufacturer's protocol. Means \pm standard errors were calculated from triplicates.

Knockdown Experiment by siRNA

The designed CatK siRNA, or scrambled siRNA was transfected into C4-2B cells using the transfection reagents from Santa Cruz. In brief, C4-2B cells were cultured in 6-well plates (3×10^5 /well) with antibiotic-free cell growth medium 1 day prior to transfection. For each transfection, 1 μ g of siRNA duplex was diluted in siRNA transfection medium to a final volume of 100 μ l. Six microliter siRNA transfection reagent was diluted into 100 μ l siRNA transfection medium and added directly into the siRNA duplex solution. After 30 min incubation at room temperature, 0.8 ml transfection medium was added to each tube containing the siRNA and transfection reagent mixture, then overlaid the mixture onto the cells which had been washed by siRNA transfection medium. Cells were incubated for 6 hours at 37°C and cell growth media containing 2x serum and antibiotics were then added. The cells were incubated for an additional 24 hours and the media was replaced with cell growth media for 48 hours. Cell lysates were collected for western blot to confirm CatK expression was knockdown.

Cell Proliferation

Cell proliferation was measured by a CellTiter 96 AQueous Non-Radioactive Cell Proliferation Assay (Promega, Madison, WI). Briefly, C4-2B cells were plated in 96-well plates at a density of 5000 cells/well in 200 μ L of T medium with 5% FBS. After 12 hours of culture, the media was changed to RPMI 1640 plus 0.5% FBS and a different concentration (0-100 μ M) of CatK inhibitor was added. The cells were incubated at 37 °C in a humidified 5% CO₂ atmosphere for 24 hours, then 20 μ L of MTS/PMS solution was added. After incubation for 2 hours at 37 °C, the absorbance of each well at 490 nm was recorded by using an ELISA plate reader. Data represent the average absorbance of six wells in one experiment.

In Vitro Cell Invasion Assay

The *in vitro* invasion assay was performed using C4-2B cells in the presence or absence of CatK inhibitor and C4-2B cells transfected with either CatK siRNA or control siRNA. The invasiveness of cells was

evaluated in 24-well matrigel invasion chamber (BD Biosciences, Bedford, MA), as directed by the manufacturer. Briefly, the upper and lower culture compartments of each well are separated by polycarbonate membranes (8- μ m pore size). The membranes in some wells were pre-coated with 100 μ g/cm² of collagen matrix (Matrigel) that was reconstituted by adding 0.5 ml of serum-free T medium to the well for 2 hours. To assess the ability of the cells to cross the polycarbonate membrane (i.e., baseline migration), 2.5×10^4 cells in 0.5 ml of T medium containing 5% FBS was placed into the upper compartment of wells that did not contain collagen matrix, and 0.75 ml of T medium containing 10% FBS was placed into the lower compartment. In parallel, we assessed the ability of the same cells to penetrate a collagen matrix by placing 2.5×10^4 cells in 0.5 ml of T medium containing 5% FBS in the upper compartment of wells that were coated with the reconstituted matrix and 0.75 ml of T medium containing 10% FBS. In upper compartments, different concentration of CatK inhibitor (0, 1, 10, 50, 100 μ M) was added. The transwell chambers were incubated for 24 hours at 37 °C in 95% air and 5% CO₂. Cell penetration through the membrane was detected by staining the cells on the porous membrane with a Diff-Quik stain kit (Dade Behring, Newark, DE) and quantified by counting the numbers of cells that penetrated the membrane in five microscopic fields (at x200 magnification) per filter. Invasive ability was defined as the proportion of cells that penetrated the matrix-coated membrane divided by the number of cells that migrated through the uncoated membrane (baseline migration). The results are reported as the mean of triplicate assays.

In Vitro Bone Resorption Assay

CM (25%, v/v) from C4-2B cells was added to non-adherent primary murine bone marrow cells (1×10^5 from C57BL/6 mouse femurs) that seeded into the 24-wells of BD Biocoat Osteologic Bone cell culture system (BD Bioscience) that consist of sub-micro synthetic calcium phosphate thin films coated on to the culture vessels and on dentin slices in 96-wells plate. Soluble RANKL (50ng/ml) and MCSF (10ng/ml) were used as positive controls or different concentration of the CatK inhibitor was added. The osteoclast culture was

maintained for 10 days and half of the media was changed every 3 days. Then the cells were fixed and stained for tartrate-resistant acid phosphatase (TRAP) (Kamiya Biomedical, Seattle, WA). Resorptive area on the digital images of osteologic or dentin slices were measured with a BIOQUANT system (R&M Biometrics, Inc., Nashville, TN). Samples were evaluated in triplicates.

Experimental Protocols for Intraosseous Tumorigenesis

The animal studies are summarized in figure 4A. In protocol 1, mice were randomized to receive either vehicle or CatK inhibitor (five times per week, 50mg/kg/day as low dose and 100mg/kg/day as high dose, orally, n=10) at the time of C4-2B cell injection (3×10^5 cells) and continued for 8 weeks. In protocol 2, four weeks after the tumor cells injection, the mice were randomized into the following treatment groups (n=10/group): ZA, CatK inhibitor, the combination of ZA and CatK inhibitor, and saline vehicle alone. ZA was injected s.c. once every 4 weeks at 100ug/kg. CatK inhibitor was given at 50mg/kg. Test drugs were given for 8 weeks. One group was sacrificed as basal control. Before sacrifice, the animals were anaesthetized, and magnified flat radiographs were taken with a Faxitron (Faxitron X-Ray Corp, Wheeling, Illinois). At sacrifice, the blood was collected and measured for PSA levels. All of the major organs and lumbar vertebrae were harvested for histological analysis.

Histopathology and Bone Histomorphometry

Histopathology was performed as described previously [16]. Briefly, bone specimens were fixed in 10% formalin and decalcified in 10% EDTA for 6 days. The specimens were then paraffin embedded, sectioned (5 μ M), and stained with hematoxylin and eosin (H&E). Nonstained sections were deparaffinized and rehydrated then stained for prostate-specific antigen (PSA) with anti-human PSA antibody using standard immunohistochemistry techniques. To perform TRAP staining, nonstained sections were stained for TRAP. For routine histopathology, soft tissues were preserved in 10% formalin, embedded in paraffin, sectioned (5 μ M),

and stained with H&E. To evaluate xenografts proliferation, sections were deparaffinized, rehydrated and stained with Ki67 monoclonal antibody following a modified protocol [47]. The Ki-67 labeling index (KI) was calculated as the percentage of positive tumor nuclei divided by the total number of tumor cells examined. At least 1,000 tumor cells per specimen were examined in five randomly selected fields by light microscopy (x400) by an investigator who was blinded to the animal groups as we have previously described [48].

Histomorphometric analysis was performed on a BIOQUANT system (BIOQUANT-R&M Biometrics Inc) as described previously [16]. Four discontinuous random regions of interest were examined within each tibia, without knowledge of treatment group, by examination at $\times 100$. Tumor volume was determined as the proportion of tumor area in the total nonmineralized portion of the bone.

Bone Mineral Density Analysis

Total bone mineral density and trabecular bone mineral density of tibiae were measured using peripheral quantitative computed tomography (pQCT) (Stratec, Ontario, CA). Scans from three slices of the proximal metaphases region were performed to obtain trabecular bone mineral densities.

Subcutaneous Tumors

As a parallel study for protocol 1, at the same time as the intratibial injection, C4-2B cells were resuspended in T media. Two million cells were mixed 1:1 with Matrigel (Collaborative Biomedical Products, Bedford, Massachusetts), and then injected into the right flank at 100 μl /site using a 23-g needle (n=10/group). Subcutaneous tumor growth was monitored by palpation, and two perpendicular axes were measured; the tumor volume was calculated using the formula as: $\text{volume} = \text{length} \times \text{width}^2/2$.

PSA Measurement

Total PSA levels in serum were determined using the Accucyte Human PSA assay (Cytimmune Sciences, College Park, MA). The sensitivity of this assay is 0.488 ng/ml.

Statistical Analysis

Statistical analysis was performed using Statview software (Abacus Concepts, Berkley, CA). For comparing two groups, Student's t test was used. ANOVA was used for initial analyses of comparing multiple groups, followed by Fisher's protected least significant difference for *post hoc* analyses. Differences with a $P < 0.05$ were determined as statistically significant.

Results

CatK is Highly Expressed in aggressive PCa Cells

Brubaker et al. reported that CatK protein were positively stained in up to 40% of primary PCa and nearly 100% of metastatic PCa cells at skeletal sites in human samples, whereas normal prostate glands were negative [40]. To confirm CatK expression in PCa, we performed immunohistochemistry on a tissue microarray that was consisted of 83 specimens from PCa patients with corresponding non-neoplastic prostate tissues. As a representative staining slide (figure 1A), CatK positive staining was at non- or minimal level (13%) in the non-neoplastic prostate glands, but at higher level (82%) in aggressive PCa. These results were consistence with those previously reported by Brubaker et al. [40]. To test whether CatK protein was expressed in PCa cells, total RNA and cell lysates were collected from PCa LNCaP, C4-2B, PC3 cells and normal prostate PrEC cells. All PCa LNCaP, C4-2B, and PC3 cells expressed higher levels of CatK mRNA than that of PrEC cells (Figure 2B-C). More aggressive C4-2B and PC3 cells expressed greater levels of both CatK mRNA and protein than the less aggressive LNCaP cells (Figure 1B-D). Cat K was present in the CM collected from all the PCa cells determined by ELISA (Figure 1E). C4-2B and PC3 cells produced greater levels of CatK protein than LNCaP cells (Figure 1E). In addition, CatK enzyme activity was also detected in LNCaP, C4-2B, and PC3 cells using an assay for degradation of collagen I as previously described (data not shown) [40].

CatK Inhibitor Diminishes PCa Tumor Invasion in vitro

To test the possible mechanism of tumor growth inhibition by CatK inhibitor in bone, the *in vitro* cell invasion assay was performed. We observed that CatK inhibitor diminished invasiveness of C4-2B cells *in vitro* in a dose-dependent fashion (Figure 2A). We examined the C4-2B cell viability using the cells treated with various doses of CatK inhibitor. We did not observe any significant differences among these cells (Figure 2B) suggesting that the CatK inhibitor had no toxicity for the tumor cells in our chosen doses. To further

confirm the role of CatK in the tumor cell invasion, we used C4-2B cells in which CatK expression was knocked down by siRNA and scrambled siRNA control cells to measure the ability of cell invasion and proliferation. CatK protein expression was significantly (95%) knocked down (Figure 2C) and the CatK siRNA knockdown, as expected, inhibited C4-2B cell invasion but did not affect the cell proliferation (Figure 2C-E). This result suggests that blocking the tumor cell invasiveness by CatK inhibitor could be a novel mechanism of CatK's function in PCa progression.

CatK Inhibitor Diminishes PCa-CM-Induced Bone Resorption in vitro

To evaluate the ability of the CatK inhibitor to diminish the PCa-CM-induced bone resorption, we first collected CM from C4-2B cell culture as we previously published [11]. We seeded non-adherent primary murine bone marrow cells into the 24-wells of BD osteologic bone cell culture system that consist of sub-micro synthetic calcium phosphate thin films coated onto the culture vessels. As a parallel study, a 96-well plate which contained dentin slice was also tested for bone resorption assay. Soluble RANKL (50ng/ml as a positive control) or the indicated concentration of CatK inhibitor was added. We observed that C4-2B CM-induced bone resorption in both systems (Figure 3A and B) and this induction was significantly diminished by the CatK inhibitor in a dose-dependent manner when we quantified bone resorption area on the dentin slides (Figure B).

CatK Inhibitor Prevents Establishment and Retards Progression of PCa Tumor in Murine Bone

To determine if inhibition of osteoclastogenic activity by the CatK inhibitor could prevent establishment or retard progression of PCa *in vivo*, we directly injected C4-2B cells into the tibia of SCID mice. The experimental protocols are summarized in figure 4A. Additionally, we injected C4-2B cells into subcutaneous sites of the same mice in protocol 1 to evaluate differences in response in the bone versus a non-osseous site. In protocol 1, the mice were administered orally either CatK inhibitor or vehicle either at the time of tumor cell injection. In protocol 2, four weeks after the tumor cells injection, the mice were randomized into groups: ZA,

CatK inhibitor, the combination of ZA and CatK inhibitor, and saline vehicle alone. Test drugs were given for 8 weeks. We found that CatK inhibitor significantly inhibited the establishment and development of PCa tumor in bone (Figure 4B and C). C4-2B caused dominantly osteolytic areas with some areas of osteoblastic activity (Figure 4B). It significantly reduced tumor volume (Figure 4C), serum PSA levels (Figure 4D) and osteoclast formation at bone-tumor interface (Figure 4E) compared to the vehicle-treated animals. These results indicated tumor burden was reduced by administration of the CatK inhibitor. In contrast, ZA less effectively diminished the skeletal tumor volume compared to the administration of the CatK inhibitor (Figure 4C). Interestingly, ZA failed to decrease PSA levels in these animals (Figure 4D). The inhibitory effects of the CatK inhibitor were enhanced in combination with zoledronic acid treatment (Figure 4B to 4D).

CatK inhibitor treatment diminished tumor-induced loss of bone mineral density (BMD) based on pQCT analysis

To further confirm the alteration of tumor-induced bone lesions by approaches other than bone histomorphometric analysis, total and trabecular BMD for the tibiae were also measured by pQCT (Figure 5A). CatK inhibitor treatment, both at low and high doses, significantly diminished the tumor-induced loss of total and trabecular BMD (Figure 5B and C). As expected, the CatK inhibitor also increased total and trabecular BMD in both protocols. The combination of the CatK inhibitor with ZA significantly enhanced both total BMD and trabecular BMD compared to any of the single agent alone (Figure 5B and C).

CatK inhibitor had no effect on tumor growth at subcutaneous sites

As a parallel study for protocol 1, at the same time as the intratibial injection, C4-2B cells were injected into the subcutaneous sites. Subcutaneous tumor growth was measured for 8 weeks. Interestingly, we found CatK inhibitor had no direct effect on the tumor volume (Figure 6A) and the tumor cell proliferation *in vivo*

(Figure 6*B* and *C*). These results suggest that the effect of CatK inhibitor maybe specific to the bone microenvironment.

Discussion

Prostate cancer skeletal metastases cause significant complications including severe bone pain, impaired mobility, pathological fracture, spinal cord compression and hypercalcemia [49, 50]. In spite of the radiographic osteoblastic appearance it is clear from histological evidence that PCa metastases form a heterogeneous mixture of osteolytic and osteoblastic lesions although osteoblastic lesions are predominate [6-9]. Recent evidence shows that osteoblastic metastases form on trabecular bone at sites of previous osteoclastic resorption, and that such resorption may be required for subsequent osteoblastic bone formation [10, 11].

Cysteine proteases have been implicated in the cancer progression, including tumor cell aberrant proliferation and apoptosis, angiogenesis, invasion, and metastasis. This suggests that cysteine proteases are relevant drug targets for treating cancer [51]. In addition, cysteine cathepsins' upregulation has been reported for different types of human tumors, including breast, lung, brain, gastrointestinal, PCa, and melanoma [52]. CatK, a key component in bone remodeling, was recently identified with the highest matrix degradation activities. It also has higher efficiency than any other cathepsins and metalloproteinases (MMPs) [27, 33]. CatK knockout mice showed an osteopetrotic phenotype associated with a severe impairment of resorptive activity of osteoclasts [53]. Several lines of evidence have shown that osteoclastic lesions are important to the development of bone metastases [12]. Anti-resorptive approaches such as administration of bisphosphonates or anti-PTHrP neutralizing antibody have been reported in breast cancer models to be able to block the tumor expansion in bone [14, 15]. Bisphosphonates could prevent bone loss by inducing cell death of osteoclasts. However, bisphosphonates have several disadvantages, such as upper gastrointestinal irritation, fever, pain, and delayed fracture healing and have a very long half-life in human body (>10 years), which is a serious concern [54]. Furthermore, it has been recently reported that bisphosphonates are capable of causing osteonecrosis of the jaws [55-57] and bisphosphonates was shown to induce inflammation and rupture of atherosclerotic plaques in apolipoprotein-E null mice [58]. Therefore new treatments are needed. Our findings in the current study show that a selective CatK inhibitor could provide an alternative approach and support a recent report that

showed using CatK inhibitors would be more beneficial than using bisphosphonates [59] because they have a positive effect on bone formation in addition to the inhibition of bone resorption.

In our experiments, we have found prostate cancer cell lines, C4-2B, LNCaP, and PC3 expressed CatK mRNA and protein, and prostate epithelial cells, PrEC expressed very low CatK. CatK's expressions were higher in metastatic prostate cancer cells—C4-2B and PC3 cells than in nonmetastatic cells—LNCaP cells either in mRNA level or protein level. We also found that CatK were expressed at low level in the normal prostate, increased in primary prostate tumor, and at high level in a skeletal metastatic tumor by immunohistochemistry of tissue samples (tissue microarray). Therefore, there is a possibility that the selective CatK inhibitor may target not only the bone cells, but also the tumor cells.

CatK inhibitor using in this study is an enzyme inhibitor which should not adversely affect the tumor cell and should leave the interplay between bone resorbing and bone forming cells intact. Accordingly, we examined the C4-2B cells viability that treated with indicated doses of CatK inhibitor. We didn't observe any significant differences. Interestingly, CatK inhibitor diminished invasion of C4-2B cells *in vitro*. Using a CatK siRNA technique, we obtained similar results in regard to tumor cell invasion as using a CatK inhibitor. Furthermore, we found that the CatK inhibitor dose-dependently diminished PCa CM-induced bone resorption *in vitro*. These results indicate that CatK may play an important role in both PCa progression and bony metastasis.

To further determine the effects of CatK inhibitor *in vivo*, we observed CatK inhibitor could prevent establishment of PCa tumor in mouse bone by using tumor establishment model, and retard tumor progression in murine bone by using tumor progression model. As expected, the CatK inhibitor increased bone mineral density in both mouse models. Critically, we showed, for the first time, that the inhibitory effects of the CatK inhibitor were enhanced in combination with ZA. These results provide a rationale to use the CatK inhibitor in combination with ZA in order to achieve better clinical outcomes for the treatment.

Taken together, we show that CatK inhibitor reduces prostate cancer-induced bone lesions, diminishes tumor burden in bone, and increases bone formation. Our data suggest that CatK inhibitor is a potential agent for the treatment of bone metastases. This novel and unique targeting strategy has established the rationale to inhibit tumor-induced bone resorption at bone metastatic sites.

Acknowledgements

The authors thank Dr. G David Roodman for his helpful discussion, Dr. Zhong Cai for his technical help, and Ms. Adrienne Mullins for her editing.

References

- [1] Roodman GD (2004). Mechanisms of bone metastasis. *N Engl J Med.* **350**, 1655-1664.
- [2] Rana A, Chisholm GD, Khan M, Sekharjit SS, Merrick MV, and Elton RA (1993). Patterns of bone metastasis and their prognostic significance in patients with carcinoma of the prostate. *Br J Urol.* **72**, 933-936.
- [3] Abrams HL, Spiro R, and Goldstein N (1950). Metastases in carcinoma; analysis of 1000 autopsied cases. *Cancer.* **3**, 74-85.
- [4] Landis SH, Murray T, Bolden S, and Wingo PA (1999). Cancer statistics, 1999. *CA Cancer J Clin.* **49**, 8-31, 31.
- [5] Bubendorf L, Schopfer A, Wagner U, Sauter G, Moch H, Willi N, Gasser TC, and Mihatsch MJ (2000). Metastatic patterns of prostate cancer: an autopsy study of 1,589 patients. *Hum Pathol.* **31**, 578-583.
- [6] Berruti A, Piovesan A, Torta M, Raucci CA, Gorzegno G, Paccotti P, Dogliotti L, and Angeli A (1996). Biochemical evaluation of bone turnover in cancer patients with bone metastases: relationship with radiograph appearances and disease extension. *Br J Cancer.* **73**, 1581-1587.
- [7] Urwin GH, Percival RC, Harris S, Beneton MN, Williams JL, and Kanis JA (1985). Generalised increase in bone resorption in carcinoma of the prostate. *Br J Urol.* **57**, 721-723.
- [8] Vinholes J, Coleman R, and Eastell R (1996). Effects of bone metastases on bone metabolism: implications for diagnosis, imaging and assessment of response to cancer treatment. *Cancer Treat Rev.* **22**, 289-331.
- [9] Roudier MP, True LD, Higano CS, Vesselle H, Ellis W, Lange P, and Vessella RL (2003). Phenotypic heterogeneity of end-stage prostate carcinoma metastatic to bone. *Hum Pathol.* **34**, 646-653.
- [10] Carlin BI, and Andriole GL (2000). The natural history, skeletal complications, and management of bone metastases in patients with prostate carcinoma. *Cancer.* **88**, 2989-2994.

- [11] Zhang J, Dai J, Qi Y, Lin DL, Smith P, Strayhorn C, Mizokami A, Fu Z, Westman J, and Keller ET (2001). Osteoprotegerin inhibits prostate cancer-induced osteoclastogenesis and prevents prostate tumor growth in the bone. *J Clin Invest.* **107**, 1235-1244.
- [12] Fazzalari NL, Kuliwaba JS, Atkins GJ, Forwood MR, and Findlay DM (2001). The ratio of messenger RNA levels of receptor activator of nuclear factor kappaB ligand to osteoprotegerin correlates with bone remodeling indices in normal human cancellous bone but not in osteoarthritis. *J Bone Miner Res.* **16**, 1015-1027.
- [13] Loberg RD, Gayed BA, Olson KB, and Pienta KJ (2005). A paradigm for the treatment of prostate cancer bone metastases based on an understanding of tumor cell-microenvironment interactions. *J Cell Biochem.* **96**, 439-446.
- [14] Guise TA, Yin JJ, Taylor SD, Kumagai Y, Dallas M, Boyce BF, Yoneda T, and Mundy GR (1996). Evidence for a causal role of parathyroid hormone-related protein in the pathogenesis of human breast cancer-mediated osteolysis. *J Clin Invest.* **98**, 1544-1549.
- [15] Sasaki A, Boyce BF, Story B, Wright KR, Chapman M, Boyce R, Mundy GR, and Yoneda T (1995). Bisphosphonate risedronate reduces metastatic human breast cancer burden in bone in nude mice. *Cancer Res.* **55**, 3551-3557.
- [16] Zhang J, Dai J, Yao Z, Lu Y, Dougall W, and Keller ET (2003). Soluble receptor activator of nuclear factor kappaB Fc diminishes prostate cancer progression in bone. *Cancer Res.* **63**, 7883-7890.
- [17] Corey E, Brown LG, Kiefer JA, Quinn JE, Pitts TE, Blair JM, and Vessella RL (2005). Osteoprotegerin in prostate cancer bone metastasis. *Cancer Res.* **65**, 1710-1718.
- [18] Saad F, Gleason DM, Murray R, Tchekmedyian S, Venner P, Lacombe L, Chin JL, Vinholes JJ, Goas JA, and Zheng M (2004). Long-term efficacy of zoledronic acid for the prevention of skeletal complications in patients with metastatic hormone-refractory prostate cancer. *J Natl Cancer Inst.* **96**, 879-882.

- [19] Lipton A, Small E, Saad F, Gleason D, Gordon D, Smith M, Rosen L, Kowalski MO, Reitsma D, and Seaman J (2002). The new bisphosphonate, Zometa (zoledronic acid), decreases skeletal complications in both osteolytic and osteoblastic lesions: a comparison to pamidronate. *Cancer Invest.* **20 Suppl 2**, 45-54.
- [20] Saad F, Gleason DM, Murray R, Tchekmedyian S, Venner P, Lacombe L, Chin JL, Vinholes JJ, Goas JA, and Chen B (2002). A randomized, placebo-controlled trial of zoledronic acid in patients with hormone-refractory metastatic prostate carcinoma. *J Natl Cancer Inst.* **94**, 1458-1468.
- [21] Rosen LS, Gordon D, Tchekmedyian NS, Yanagihara R, Hirsh V, Krzakowski M, Pawlicki M, De Souza P, Zheng M, Urbanowitz G, et al. (2004). Long-term efficacy and safety of zoledronic acid in the treatment of skeletal metastases in patients with nonsmall cell lung carcinoma and other solid tumors: a randomized, Phase III, double-blind, placebo-controlled trial. *Cancer.* **100**, 2613-2621.
- [22] Drake FH, Dodds RA, James IE, Connor JR, Debouck C, Richardson S, Lee-Rykaczewski E, Coleman L, Rieman D, Barthlow R, et al. (1996). Cathepsin K, but not cathepsins B, L, or S, is abundantly expressed in human osteoclasts. *J Biol Chem.* **271**, 12511-12516.
- [23] Tezuka K, Tezuka Y, Maejima A, Sato T, Nemoto K, Kamioka H, Hakeda Y, and Kumegawa M (1994). Molecular cloning of a possible cysteine proteinase predominantly expressed in osteoclasts. *J Biol Chem.* **269**, 1106-1109.
- [24] Shi GP, Chapman HA, Bhairi SM, DeLeeuw C, Reddy VY, and Weiss SJ (1995). Molecular cloning of human cathepsin O, a novel endoproteinase and homologue of rabbit OC2. *FEBS Lett.* **357**, 129-134.
- [25] Inaoka T, Bilbe G, Ishibashi O, Tezuka K, Kumegawa M, and Kokubo T (1995). Molecular cloning of human cDNA for cathepsin K: novel cysteine proteinase predominantly expressed in bone. *Biochem Biophys Res Commun.* **206**, 89-96.

- [26] Bossard MJ, Tomaszek TA, Thompson SK, Amegadzie BY, Hanning CR, Jones C, Kurdyla JT, McNulty DE, Drake FH, Gowen M, et al. (1996). Proteolytic activity of human osteoclast cathepsin K. Expression, purification, activation, and substrate identification. *J Biol Chem.* **271**, 12517-12524.
- [27] Garnero P, Borel O, Byrjalsen I, Ferreras M, Drake FH, McQueney MS, Foged NT, Delmas PD, and Delaisse JM (1998). The collagenolytic activity of cathepsin K is unique among mammalian proteinases. *J Biol Chem.* **273**, 32347-32352.
- [28] Greenspan A (1991). Sclerosing bone dysplasias--a target-site approach. *Skeletal Radiol.* **20**, 561-583.
- [29] Cabrejas ML, Fromm GA, Roca JF, Mendez MA, Bur GE, Ferreyra ME, Demarchi C, and Schurman L (1976). Pycnodysostosis: some aspects concerning kinetics of calcium metabolism and bone pathology. *Am J Med Sci.* **271**, 215-220.
- [30] Meredith SC, Simon MA, Laros GS, and Jackson MA (1978). Pycnodysostosis. A clinical, pathological, and ultramicroscopic study of a case. *J Bone Joint Surg Am.* **60**, 1122-1127.
- [31] Gelb BD, Shi GP, Chapman HA, and Desnick RJ (1996). Pycnodysostosis, a lysosomal disease caused by cathepsin K deficiency. *Science.* **273**, 1236-1238.
- [32] Saftig P, Hunziker E, Wehmeyer O, Jones S, Boyde A, Rommerskirch W, Moritz JD, Schu P, and von Figura K (1998). Impaired osteoclastic bone resorption leads to osteopetrosis in cathepsin-K-deficient mice. *Proc Natl Acad Sci U S A.* **95**, 13453-13458.
- [33] Chapman HA, Riese RJ, and Shi GP (1997). Emerging roles for cysteine proteases in human biology. *Annu Rev Physiol.* **59**, 63-88.
- [34] Sinha AA, Gleason DF, Staley NA, Wilson MJ, Sameni M, and Sloane BF (1995). Cathepsin B in angiogenesis of human prostate: an immunohistochemical and immunoelectron microscopic analysis. *Anat Rec.* **241**, 353-362.
- [35] Szpadarska AM, and Frankfater A (2001). An intracellular form of cathepsin B contributes to invasiveness in cancer. *Cancer Res.* **61**, 3493-3500.

- [36] Yan S, Sameni M, and Sloane BF (1998). Cathepsin B and human tumor progression. *Biol Chem.* **379**, 113-123.
- [37] Dodds RA, Connor JR, Drake FH, and Gowen M (1999). Expression of cathepsin K messenger RNA in giant cells and their precursors in human osteoarthritic synovial tissues. *Arthritis Rheum.* **42**, 1588-1593.
- [38] Hummel KM, Petrow PK, Franz JK, Muller-Ladner U, Aicher WK, Gay RE, Bromme D, and Gay S (1998). Cysteine proteinase cathepsin K mRNA is expressed in synovium of patients with rheumatoid arthritis and is detected at sites of synovial bone destruction. *J Rheumatol.* **25**, 1887-1894.
- [39] Littlewood-Evans AJ, Bilbe G, Bowler WB, Farley D, Wlodarski B, Kokubo T, Inaoka T, Sloane J, Evans DB, and Gallagher JA (1997). The osteoclast-associated protease cathepsin K is expressed in human breast carcinoma. *Cancer Res.* **57**, 5386-5390.
- [40] Brubaker KD, Vessella RL, True LD, Thomas R, and Corey E (2003). Cathepsin K mRNA and protein expression in prostate cancer progression. *J Bone Miner Res.* **18**, 222-230.
- [41] Yamashita DS, and Dodds RA (2000). Cathepsin K and the design of inhibitors of cathepsin K. *Curr Pharm Des.* **6**, 1-24.
- [42] Marquis RW, Ru Y, LoCastro SM, Zeng J, Yamashita DS, Oh HJ, Erhard KF, Davis LD, Tomaszek TA, Tew D, et al. (2001). Azepanone-based inhibitors of human and rat cathepsin K. *J Med Chem.* **44**, 1380-1395.
- [43] Thompson SK, Halbert SM, Bossard MJ, Tomaszek TA, Levy MA, Zhao B, Smith WW, Abdel-Meguid SS, Janson CA, D'Alessio KJ, et al. (1997). Design of potent and selective human cathepsin K inhibitors that span the active site. *Proc Natl Acad Sci U S A.* **94**, 14249-14254.
- [44] Le Gall C, Bellahcene A, Bonnelye E, Gasser JA, Castronovo V, Green J, Zimmermann J, and Clezardin P (2007). A cathepsin K inhibitor reduces breast cancer induced osteolysis and skeletal tumor burden. *Cancer Res.* **67**, 9894-9902.

- [45] Lu Y, Cai Z, Xiao G, Keller ET, Mizokami A, Yao Z, Roodman GD, and Zhang J (2007). Monocyte chemotactic protein-1 mediates prostate cancer-induced bone resorption. *Cancer Res.* **67**, 3646-3653.
- [46] Lu Y, Cai Z, Xiao G, Liu Y, Keller ET, Yao Z, and Zhang J (2007). CCR2 expression correlates with prostate cancer progression. *J Cell Biochem.* **101**, 676-685.
- [47] Salas TR, Kim J, Vakar-Lopez F, Sabichi AL, Troncoso P, Jenster G, Kikuchi A, Chen SY, Shemshedini L, Suraokar M, et al. (2004). Glycogen synthase kinase-3 beta is involved in the phosphorylation and suppression of androgen receptor activity. *J Biol Chem.* **279**, 19191-19200.
- [48] Wallner L, Dai J, Escara-Wilke J, Zhang J, Yao Z, Lu Y, Trikha M, Nemeth JA, Zaki MH, and Keller ET (2006). Inhibition of interleukin-6 with CNTO328, an anti-interleukin-6 monoclonal antibody, inhibits conversion of androgen-dependent prostate cancer to an androgen-independent phenotype in orchietomized mice. *Cancer Res.* **66**, 3087-3095.
- [49] Moul JW, and Lipo DR (1999). Prostate cancer in the late 1990s: hormone refractory disease options. *Urol Nurs.* **19**, 125-131; quiz 132-123.
- [50] Coleman RE (1997). Skeletal complications of malignancy. *Cancer.* **80**, 1588-1594.
- [51] Turk V, Kos J, and Turk B (2004). Cysteine cathepsins (proteases)--on the main stage of cancer? *Cancer Cell.* **5**, 409-410.
- [52] Jedeszko C, and Sloane BF (2004). Cysteine cathepsins in human cancer. *Biol Chem.* **385**, 1017-1027.
- [53] Saftig P, Hunziker E, Everts V, Jones S, Boyde A, Wehmeyer O, Suter A, and von Figura K (2000). Functions of cathepsin K in bone resorption. Lessons from cathepsin K deficient mice. *Adv Exp Med Biol.* **477**, 293-303.
- [54] Grabowska U CT, Shiroo M. (2005). Recent developments in cathepsin K inhibitor design. *Curr Opin Drug Discov Devel* **8**, 619-630.
- [55] Melo MD, and Obeid G (2005). Osteonecrosis of the maxilla in a patient with a history of bisphosphonate therapy. *J Can Dent Assoc.* **71**, 111-113.

- [56] Marx RE (2003). Pamidronate (Aredia) and zoledronate (Zometa) induced avascular necrosis of the jaws: a growing epidemic. *J Oral Maxillofac Surg.* **61**, 1115-1117.
- [57] Ruggiero SL, Mehrotra B, Rosenberg TJ, and Engroff SL (2004). Osteonecrosis of the jaws associated with the use of bisphosphonates: a review of 63 cases. *J Oral Maxillofac Surg.* **62**, 527-534.
- [58] Shimshi M, Abe E, Fisher EA, Zaidi M, and Fallon JT (2005). Bisphosphonates induce inflammation and rupture of atherosclerotic plaques in apolipoprotein-E null mice. *Biochem Biophys Res Commun.* **328**, 790-793.
- [59] Vasiljeva O, Reinheckel T, Peters C, Turk D, Turk V, and Turk B (2007). Emerging roles of cysteine cathepsins in disease and their potential as drug targets. *Curr Pharm Des.* **13**, 387-403.

Figure legends

Figure 1. CatK mRNA and protein expression in PCa tissues and cell lines. (A) CatK expression in human PCa tissues. Immunohistochemical staining was performed for detection of CatK in human PCa and corresponding non-neoplastic tissues. (B) Total RNA was extracted from prostate epithelial cells (PrEC), LNCaP, C4-2B, and PC3 cells, and subjected to RT-PCR for detection CatK mRNA. PCR product of 399 bp was detected. (C) Quantification of CatK mRNA was determined by real-time PCR. Internal control was β -actin. * $P < 0.05$ compared to PrEC cells; # $P < 0.01$ compared to LNCaP cells. (D) Proteins were collected from PrEC, LNCaP, C4-2B, and PC3 cells and subjected to Western blot. (E) CM collected from PrEC, LNCaP, C4-2B, and PC3 cell cultures were measured by an ELISA kit. * $P < 0.01$ compared to PrEC cells; # $P < 0.01$ compared to LNCaP cells.

Figure 2. CatK inhibitor diminished the invasiveness of C4-2B cells. (A) The *in vitro* invasion assay was performed using C4-2B cells cultured in a transwell chamber. Cell penetration through the membrane was detected by staining the cells on the porous membrane with a Diff-Quik stain kit and quantified by counting the numbers of cells that penetrated the membrane in five microscopic fields (at $\times 200$ magnification) per filter. Invasive ability was defined as the proportion of cells that penetrated the matrix-coated membrane divided by the number of cells that migrated through the uncoated membrane (baseline migration). The results are reported as the mean of triplicate assays. * $P < 0.05$ compared to the vehicle; # $P < 0.001$ compared to the vehicle. (B) The C4-2B cells viability were examined by MTS assay. C4-2B cells were treated with various doses of CatK inhibitor (the doses that were used in the tumor invasion assay). (C) The designed CatK siRNA, or scrambled control siRNA were transfected into C4-2B cells. Cell lysates were collected for Western blot to confirm CatK expression was knocked down. (D) Cell viability was measured by MTS assay. (E) The cell invasion was measured for the CatK knockdown cells and compared to the control cells. The results are reported as the mean of triplicate assays. * $p < 0.01$ compared to the control siRNA-transfected cells.

Figure 3. CatK inhibitor diminishes PCa-CM-induced bone resorption *in vitro*. (A) Representative images of resorption pits on dentin slices or synthetic calcium phosphate thin films are shown. Left panels without a frame: BD Biocate osteologic bone cell culture system, right panels with a frame: pit assay. (B) Samples were evaluated in triplicates and the resorption area was quantified. Results are reported as mean \pm SD. *P<0.001 compared to Non-treatment (Non-TX) group; #P<0.01 compared to C4-2B CM-treated group; **P<0.001 compared to C4-2B CM-treated group.

Figure 4. CatK inhibitor prevented the PCa establishment and retarded progression of PCa tumor in mouse bone. (A) Schematic of experimental procedures to determine the effects of CatK inhibitor on the establishment and progression of prostate cancer. (B) In this representative figure, note the area of osteolysis and osteosclerosis of the vehicle-treated mouse compared to the radiograph of the CatK inhibitor-treated mouse. PSA is strong positive in all vehicle-treated mice compared to the CatK inhibitor-treated mice. (C) Tumor volume vs non-bone soft tissue volume was measured by bone histomorphometry. (D) Serum PSA levels in the mice model were measured by ELISA. (E) Osteoclast numbers per millimeter bone surface were quantified by bone histomorphometry. Results are reported as mean \pm SD. *P<0.01 compared to vehicle group; #P<0.01 compare to basal group.

Figure 5. CatK inhibitor protected bone mineral density in a mouse model. (A) Slices that were scanned by pQCT. The small box in the right panel indicates the tibia. The reference slice and the slices examined were indicated on a representative tibia. (B) Total bone mineral density detected by pQCT of tibiae from each group. (C) Trabecular bone mineral density detected by pQCT of tibiae from each group. *P<0.01 compared to vehicle group; #P<0.01 compare to basal group.

Figure 6. CatK inhibitor had no effect on subcutaneous tumor growth *in vivo*. (A) Tumor volume were monitored twice weekly by caliper measurements. (B) Subcutaneous tumor sections were immunohistochemically stained with Ki67 antibody. (C) Quantified data were determined by the number of Ki67 positive cells dividing the total numbers in five randomly selected fields under light microscopy ($\times 400$).

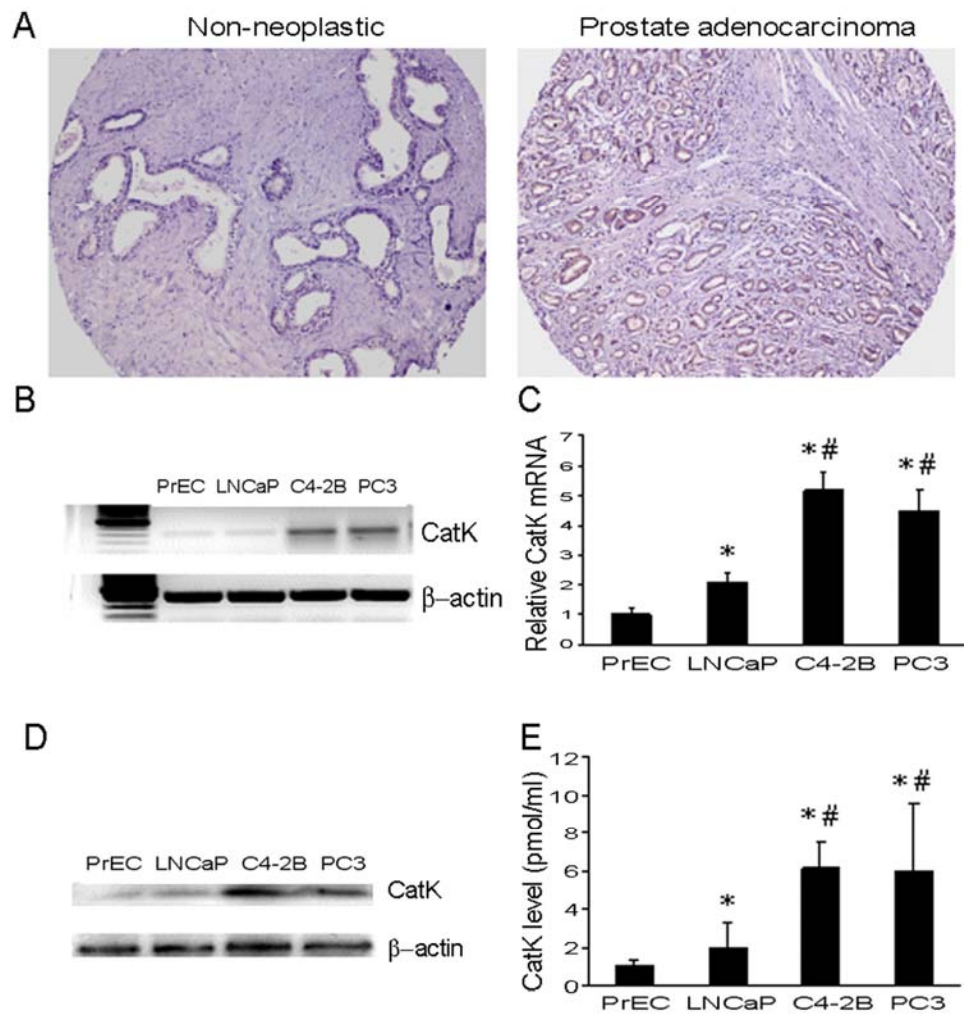


Figure 1

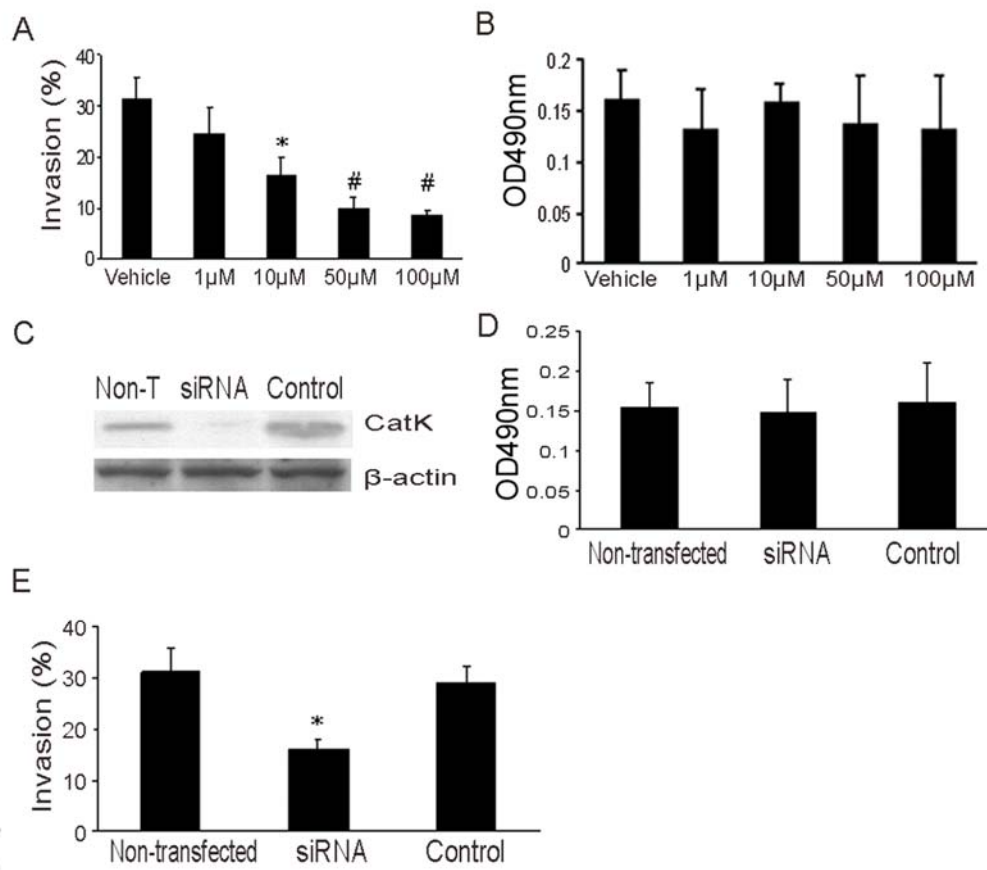


Figure 2

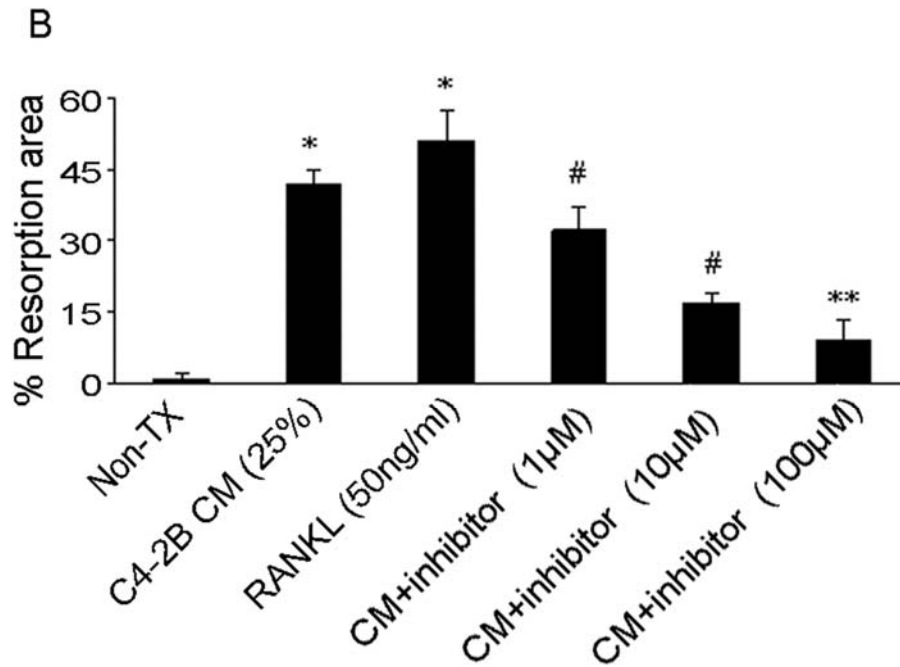
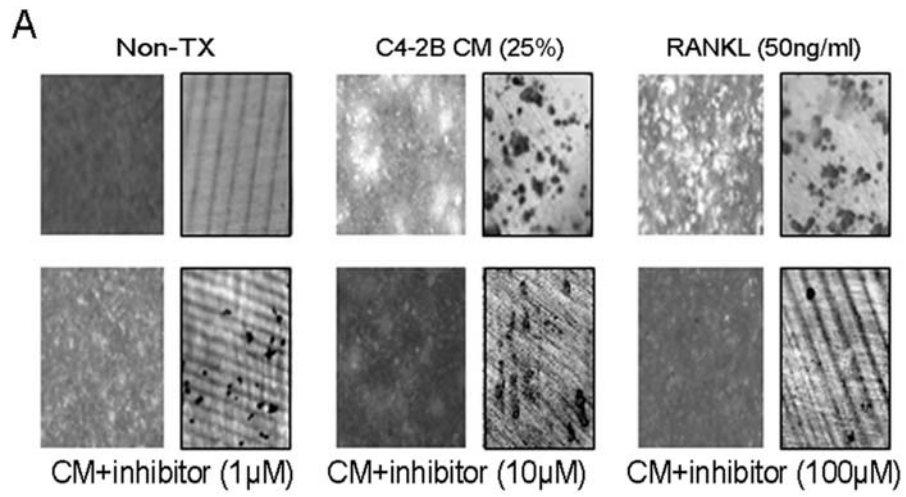


Figure 3

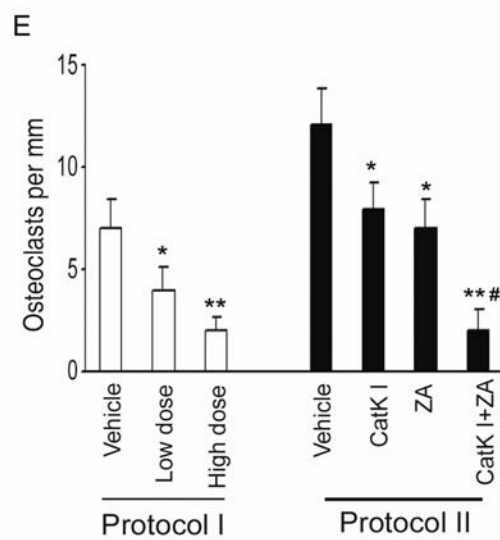
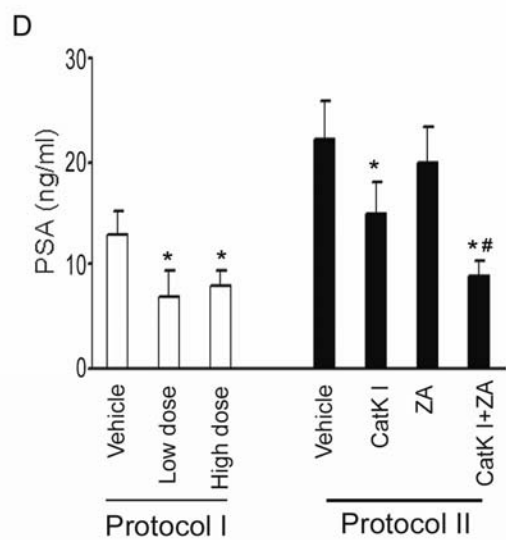
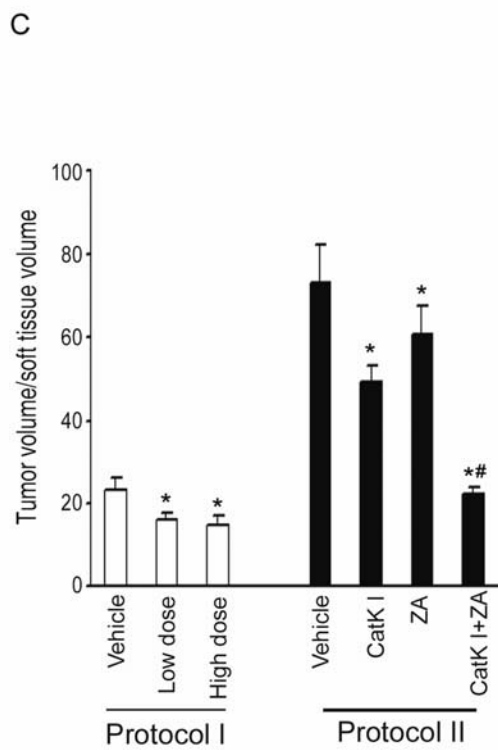
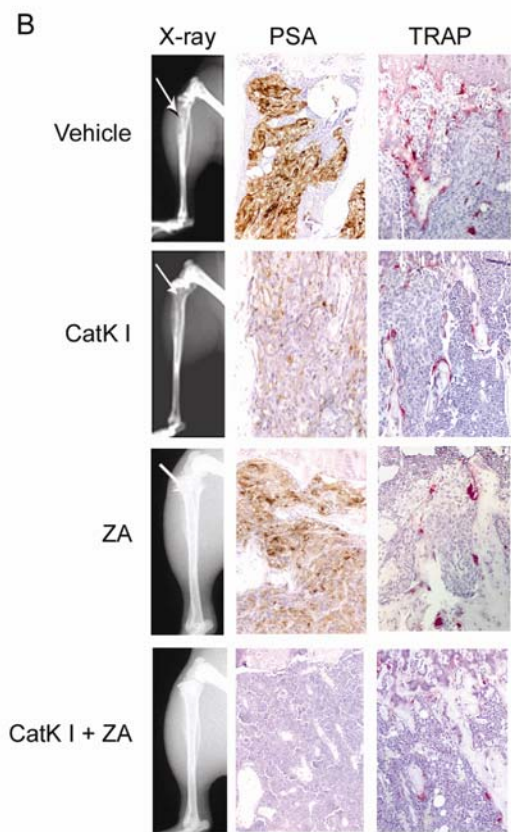


Figure 4B-E

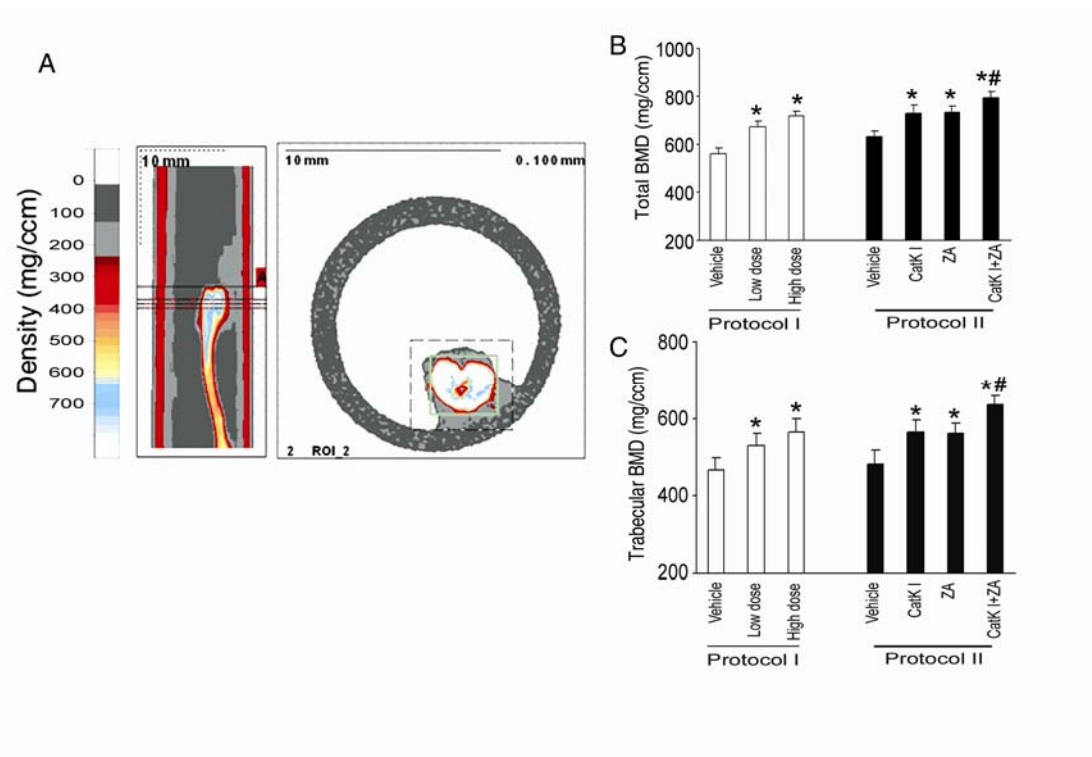


Figure 5

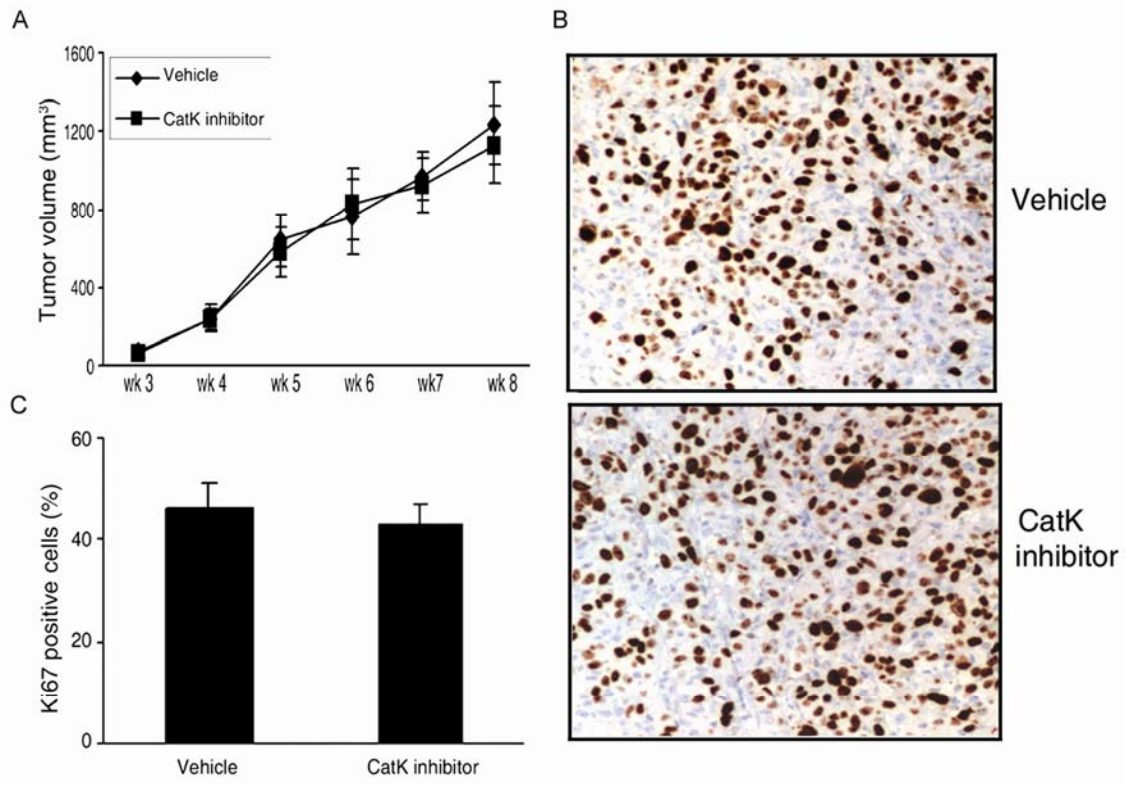


Figure 6

From: submissions@neoplasia.com [mailto:submissions@neoplasia.com]
Sent: Thu 2009-5-7 18:55
To: Zhang, Jian
Cc: edassist@neoplasia.com
Subject: Manuscript Received by Neoplasia 09-730 - Requires Action

Caution: This is a unique authorship confirmation link for Jian Zhang. If you are not Jian Zhang, please do not use this link to confirm authorship.

Your manuscript has been received by Neoplasia. To access your manuscript online, you must confirm authorship by

<html><a href=<http://www.neoplasia.com/jsoft2/authorship.php?uid=yel4a03632b3dd1a&msid=2623>>clicking here<P></html>

OR copying and pasting the following hyperlink to your web browser.

<http://www.neoplasia.com/jsoft2/authorship.php?uid=yel4a03632b3dd1a&msid=2623>

Notes:

1. When you click the above link, you will be prompted for a login name and password. If you are a registered user, just login. Then you will see a confirmation page indicating that you can access the manuscript online.

2. If you are a new user, please register first by clicking <http://www.neoplasia.com/jsoft2/register.php>. Then go to the first link.

MS #: 09-730

Title: Targeting cathepsin K diminishes prostate cancer establishment and growth in murine bone

First Author: Yi Lu

Corresponding Author: Jian Zhang

Email: zhangj2@upmc.edu

Files:

Cover Letter.Zhang.doc

Zhang.CatK inhibitor.text and figures.pdf