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Role of MEKK3 signaling pathway in the resistance of breast cancer cells to $TNF\alpha$ -mediated apoptosis

Introduction

Breast cancer is the most commonly diagnosed malignancy and one of leading causes of death in American women. So far the chemotherapy and radiotherapy are still common treatments for breast cancer. However, the efficiency of the treatment usually has been limited because breast cancer develops resistance to chemotherapeutic drugs and radiation treatment. TNF α is one of the most pleiotropic cytokine acting as a cytotoxic agent against a variety of tumor cell lines and also play a role in tumor regression mediated by cytotoxic T cells. TNF α is released by cytotoxic T cells and significantly contributes to the local immune response to the tumor. Tumor cells including breast cancer cells were naturally or acquire resistance to TNF α -mediated apoptosis yet the mechanism is still not fully understood. We recently created MEKK3 knockout mice to investigate its in vivo function and demonstrated that MEKK3 plays a crucial role in TNF α induced NF- κ B activation and apoptosis (Yang et al. 2001). Our study suggests that MEKK3 may be involved in breast cancer cells' resistance to TNF α -mediated apoptosis.

Body

To test our hypothesis, we have been working on the conditions to alter the MEKK3 activities in normal and in breast cancer cells and then determine how TNFresponses will be affected. The strategy being used is to block the MEKK3 activity using dominant interfering mutants, inhibitors and siRNA technique in breast cancer cells. During this funding period, we constructed the dominant MEKK3 mutants and tested their effects on NF- κ B reporter gene expression (figure 1). We also determined that MEKK3 is a specific activator of the NF- κ B, JNK and p38 MAPKs, but not the ERK1/2 MAPK (figure 2). In addition, we tested MEKK3 specific siRNA lentiviral vectors to infect cell lines (figure 3) (Qin et al. 2003). We are still in the process of defining optimal conditions for infection. Finally, we prepared a unique antibody that recognize the activated MEKK3 but not inactive MEKK3 mutants (figure 4). This reagent will be a great assert for our investigation of MEKK3 activity.

Key research accomplishment

The key accomplishments are listed above. In brief, we have generated most of the reagents that are crucial to test our working hypothesis and in the process of optimizing the experimental conditions for achieving our goals of our proposal work. **Reportable outcomes**

The major progress in the last funding period is that we generated a number of new reagents that will facilitate our investigation for the activity of MEKK3. We prepared an antibody that specifically detects active MEKK3 but not inactive mutant MEKK3 (fig 4). These studies will allow us to assess the endogenous MEKK3 activity in breast cancer cells and its modulation by inhibitors and dominant negative MEKK3 mutants. We are still working on the condition for gene delivery efficiency to tumor cells. We have successfully generated MEKK3 siRNA lentiviral vectors and obtained promising data showing that MEKK3 siRNA worked on endogenous MEKK3 protein albeit less effective than the co-transfected MEKK3.

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Conclusions

We have made good progress toward our planed research goal in understanding the molecular mechanism of NF- κ B and MAPK activation through the MEKK3 pathway. In this regard, Dr. Huang published one manuscript that is related to this research (Huang et al, NI 2004). During the course of this work, we have a unexpected interruption because Dr. Huang was unable to obtain her visa extension to stay in US to finishing her work. Dr. Huang thus left the country in Dec. 2003 and her work is continued now by another postdoctoral fellow, Dr. Ling Yu, who joined our Institution from Japan on April 1st, 2004. Dr. Yu have started testing the conditions set-up by Dr. Huang and she quickly measured the expression and activities MEKK3 mutants in various cells and in the process of making retrovirus. Dr. Yu is also working on MEKK3 siRNA to inhibit the endogenous MEKK3 expression in breast cancer cells for further testing their effects on cancer cell sensitivity to TNF α .

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Appendices fig 1-fig 4

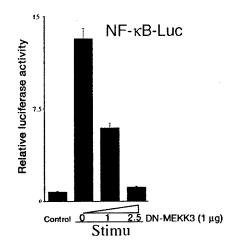


Figure 1 Dominant negative MEKK3 (DN-MEKK30 inhibits NF-κB reporter gene expression. One microgram of NF-κB-Luc reporter plasmid was transfected with either empty vector or with increasing amounts of DN-MEKK3. Transfected cells were either unstimulated (control) or stimulated (stimu) with IL-1 for 24 h before being assayed for the luciferase activity. DN-MEKK3 inhibited the reporter gene expression significantly.

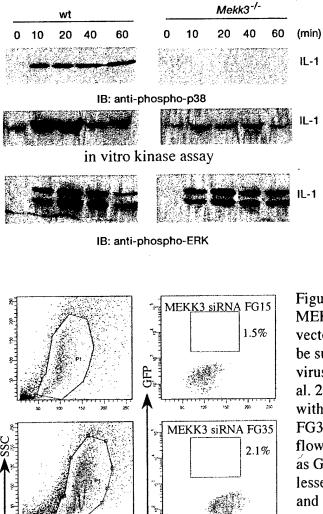


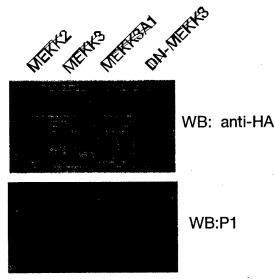
Figure 2. Activation of JNK, ERK, and p38 MAPK in wildtype and *Mekk3^{-/-}* MEFs. Wildtype and *Mekk3^{-/-}* MEFs either untreated or stimulated with IL-1 were harvested at the indicated time points. JNK activation was determined by an *in vitro* kinase assay, ERK and p38 MAPK activation was measured by immunoblotting with antiphospho-p38 and anti-phospho-ERK1/2 antibodies.

Figure 3 Infection of tumor cells with MEKK3 siRNA lentiviral expression vector. The MEKK3 siRNA cassette will be subcloned into the lentiviral vector the virus were prepared as described (Qin et al. 2003). Tumor cell line was infected with MEKK3 siRNA lentivirus FG15 and FG35, and analyzed 48 h later by FACS flow cytometry. Infected cells are shown as GFP positive. Both infections yield lesser than 3% positive cells (about 1.5 and 2.1% each).

Figure 4 Characterization of anti-phosphor-MEKK3 antibody P1. COS-1 cells were transfected with HA-tagged MEKK2, MEKK3, MEKK3 phosphorylation site mutant MEKK3A1, and DN-MEKK3 as indicated. Cells lysates were prepared 36 hr later and analyzed by immunoblotting with anti-HA antibody (top panel) and P1 antibody (bottom panel).

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