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Award Number: DAMD17-99-1-9428

TITLE: Apoptosis Based Gene Therapy of Breast Cancer

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REPORT DATE: August 2001

TYPE OF REPORT: Annual Summary

PREPARED FOR: U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release; Distribution Unlimited

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20020401 027

REPORT DOCUMENTATION PAGE			Form Approved	
Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing ins			tructions, searching existing data sources, gathering and maintaining	
the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Burden to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Burdent Resource Reports (2020) Washington Provide Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Burdent Resource Reports (2020) Washington Provide Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Burdent Resource Reports (2020) Washington Provide Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Burdent Resource R				
1. AGENCY USE ONLY (Leave blar	nk) 2. REPORT DATE	3. REPORT TYPE AND	DATES COVERED	
	August 2001	Annual Summary	(1 Aug 00 - 31 Jul 01)	
4. TITLE AND SUBTITLE	Therapy of Breast Cance	r	5. FUNDING NUMBERS	
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6 AUTHOB(S)				
Stephen E. Karp, M.D.				
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES)			8. PERFORMING ORGANIZATION	
Virginia Commonwealth University			REPORT NUMBER	
RICHMONG, VIIGINIA 23290-0500				
E-Mail: skarp@mail2.vcu.edu				
9. SPONSORING / MONITORING /	AGENCY NAME(S) AND ADDRESS(E	S)	10. SPONSORING / MONITORING	
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U.S. Army Medical Research and Materiel Command Fort Detrick Maryland 21702-5012				
11 SUPPLEMENTARY NOTES	· · · · · · · · · · · · · · · · · · ·			
TT. SOFFELMENTANT NOTES				
12a. DISTRIBUTION / AVAILABILITY STATEMENT			12b. DISTRIBUTION CODE	
Approved for Public Release; Distribution Unlimited				
13. ABSTRACT (Maximum 200 W	ords)			
This project is an attempt to develop a gene therapy model for the treatment of metastatic				
breast cancer. Our goal is to develop adenovital vectors that deriver a cycooldar gond coloctively to breast cancer cells. We have employed recombinant molecules of caspase-3, a				
distal effector protein in the apoptotic cascade. Recombinant caspase-3 molecule, unlike				
the parent molecule, is constitutively active; expression of it in breast cancer cells				
rapidly induces apoptosis. As expression of caspase-3 in non-malignant cerrs could be				
specific promoters. We have demonstrated that the promoter of the Hexokinase-II gene is				
tumor specific and the mammoglobin promoter is expressed only in breast cells. In order to				
tightly control expression to cancer cells we are attempting to employ the CRE / Lox				
system. One adenovirus would express the CRE gene under the control of the henominate in a promotor adenovirus would express an inactivated recombinant caspase-3 under				
control of the mammog	lobin promoter. Only the	he co-infection	of these two viruses in cells	
in which both of these promoters are active, i.e. breast cancer cells, would permit the				
expression of caspase-	3. These viruses are pr	esently being cor	istructea.	
			15 NUMBER OF PAGES	
Breast Cancer, gene th	nerapy, apoptosis		6	
			16. PRICE CODE	
17. SECURITY CLASSIFICATION	18. SECURITY CLASSIFICATION	19. SECURITY CLASSIFI	ICATION 20. LIMITATION OF ABSTRACT	
OF REPORT	OF THIS PAGE	OF ABSTRACT Unclassifi	ed Unlimited	
NSN 7540-01-280-5500	0110103511160	011014333111	Standard Form 298 (Rev. 2-89)	
· -			Prescribed by ANSI Std. Z39-18 298-102	

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Introduction

Conventional therapy for patients with metastatic breast cancer is unsuccessful as despite therapy with hormonal agents and chemotherapy virtually all patients die of their disease. This project is an attempt to develop a model for gene therapy of breast cancer that acts by selectively inducing apoptosis in breast cancer cells. Caspase-3, (c3), is a distal effector protein in the cascade that induces apoptosis. Expression of the c3 gene product is regulated by post translational cleavage and reassembly of the two fragments into the active molecule. Our proposal is to develop a recombinant c3 that is constitutively active (1). This recombinant c3 activates the apoptosis cascade and rapidly induces cell death. As recombinant c3 activates apoptosis far "downstream" it bypasses the mechanisms of resistance to chemotherapy and other cytotoxic agents that most cancer cells have developed by the time they have developed metastatic potential. Therefore this represents a powerful cytotoxic agent for gene therapy of cancer.

One potential problem with the use of c3 as the "killing gene" is that expression in any normal cell would also be toxic. Therefore the expression of c3 must be very tightly regulated and exclusively limited to malignant cells. In order to accomplish this we intend to employ the following strategy. It has two essential parts. The first part is the use of two separate adenoviral vectors, (with different promoters); both adenoviral vectors must infect the target cell. The second regulatory step is the use of the CRE / Lox system(2). These two together will induce tumor targeted recombinant c3 expression and apoptosis.

One vector will induce the expression of the CRE gene under the control of the Hexokinase II promoter. We have previously shown that the Hexokinase II promoter is expressed in malignant breast and lung tumor cells but not in normal lung, breast and liver cells (3). Therefore only malignant cells will have functional CRE recombinase.

The gene expressed by the second adenoviral vector will be regulated by the promoter of the mammoglobin gene which is expressed solely in the breast (4). This vector codes for a construct that consists of 2 lox sites that flank "junk" DNA that includes a stop codon; downstream of this is the recombinant c3 that is constitutively active. This vector, by itself will have no functional effect on breast or any other cells, as the gene it delivers is inactive without the co-expression of CRE by the first vector. When a permissive cell is infected by both of these viruses the junk DNA is cut out, expression of constitutively active caspase-3 occurs, and apoptosis should be induced. This double system will tightly regulate recombinant c3 expression to only malignant cells.

Body

Progress to date:

Task 1: Expression of recombinant Caspase-3 induces apoptosis in Breast Cells

As stated in last year's report, we have shown that reverse caspase-3, a recombinant c3 molecule that has been engineered to have the p11 fragment 5' of the p20 fragment, no longer requires post translational cleavage and is constitutively active. As demonstrated last year recombinant c3 induces apoptosis in 5 different breast cancer cell lines. We are using this for our present viral constructs.

Task 2: Selectivity of our Site Specific Promoters

This task has as well been accomplished. As stated in last years report, we have shown that the Hexokinase II promoter can induce high level expression in malignant cells but is poorly expressed in normal cells. Similarly, the mammoglobin promoter is selectively expressed in breast cells, benign or malignant.

Task 3: Generation of Adenoviral vectors

This year's work has been particularly frustrating. We have worked hard to clone the required plasmids necessary for vector production. Several constructs were made that appeared to be the required plasmids, but then were later determined to be the wrong constructs. This has caused a great deal of lost time.

The problems we have had relate to technical problems with the cloning. We have tried a variety of cloning techniques to create the adenoviral transfer plasmids. These are multistep cloning exercises. One problem is that the fragments to be cloned are fairly large. For example the Hexokinase II promoter itself is almost 4 kB. All of them are less than the approximately 10 kB allowable size for inserts in the second generation adenovectors that we have been employing.

A second problem relates to the fact that shortly after this year began my technician left my lab and I have been unable to find a suitable replacement. Therefore we are still in the phase of transfer plasmid production, and are not even ready to create the first viral vectors. As such we have no significant new data to deliver in this report.

A third issue relates to my move from Virginia Commonwealth University to the Lahey Clinic in Burlington, Mass, this summer. This has profoundly changed the direction of my work. These gene therapy experiments have been terminated as of August 2001. A revised statement for the continuation of this research award has been submitted.

Key Research Accomplishments

• Demonstrated that recombinant revCaspase-3 is constitutively active, and induces apoptosis.

• Demonstrated that the mammoglobin and Hexokinase II promoters are regulatory elements that can control gene expression in breast cancer cells.

Reportable outcomes

None this year

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Conclusions

See above

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Appendices

none