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One mechanism by which is pathway. The yeast C-type cycl (Ume3p) is rapidly destroyed which conserved, the human C-type cycl tagged derivative of HcycC (Hcy Exponential cultures were subject HcycC levels were reduced comp yeast subjected to heat shock. The response to stress. Moreover, these	tin (HcycC) levels were monitor ycC-HA) was constructed and ted to either heat shock (42°C) ared to untreated controls. In a ese findings are consistent with	ress response genes. T I types of stress. To c red in cell lines exposed stability integrated into or exposed to tumor ne ddition, HcycC was rap a model that down regu	o relieve this letermine if thi to stress and a the human bi crosis factor al idly destroyed lation of Hcyco	repression, the <i>UME3</i> protein s novel regulatory strategy is poptotic inducers. An epitope reast cancer cell line MCF-7. pha (TNF α). In both studies, when ectopically expressed in C is part of the normal cellular	
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FOREWORD

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(5) Introduction.

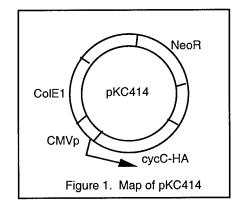
Disseminated malignancies are commonly treated with cytotoxic agents (e. g., chemotherapy, radiation) which target the unregulated growth associated with tumors. However, many of these procedures have proven unsuccessful due in part to the acquired resistance of cancer cells to these regimens. Mounting evidence suggests that one underlying mechanism by which malignancies are protected from cytotoxic agents is through aberrant activation of a pathway generally referred to as the "stress response". This system, which is found in all organisms from procaryotes to man, elicits the expression of several conserved gene families (heat shock proteins e. g., Hsp70, Hsp27) that protect the cell from cytotoxic agents. In human breast cancer, overexpression of Hsp's has been associated with tumors that are both more invasive and/or resistant to chemotheraputic drugs. We propose to expand studies initiated in the budding yeast *S. cerevisiae* to investigate the role of the human cyclin C in regulating stress response genes in breast cancer tissues.

(6) Body

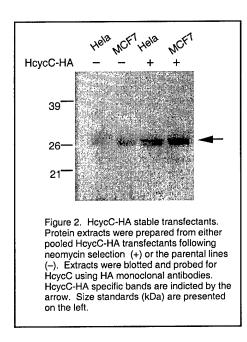
Object 1: Is HcycC down regulated as part of the normal cellular response to stress?

Task 1: Establish human cell lines stably expressing HcycC.

Stable HcycC transfectants were established in two separate cell lines, Hela and MCF7 breast cancer cells. The Hela cell line was chosen to distinguish between general effects on HcycC regulation from those specific for breast cancer cells. First, the HcycC cDNA was obtained from Dr. S. Reed, Scripts Institute (8). The cDNA was tagged with the HA epitope (15) using standard techniques. The tagged version of HcycC was inserted into the vector pCDNA3 to form pKC414 (Fig. 1) under the control of the CMV promoter which provides high, constitutive expression in many cell types.



Stable transfectants using this construct were made in both Hela and MCF7 breast cancer cell lines by lipofection. Transfectants were selected for neomycin resistance and subclones isolated. The clones (minimum of 10) were pooled and expanded for the experiments described below. To determine the expression levels of HcycC-HA, Western blots were performed on cell extracts prepared from log-phase cells. A band corresponding to 31 kDa HcycC-HA was observed in the HcycC-HA transfected cell lines but not in the mock transfected control (Fig. 2). These results indicate that HcycC-HA has been successfully integrated into these cell lines.

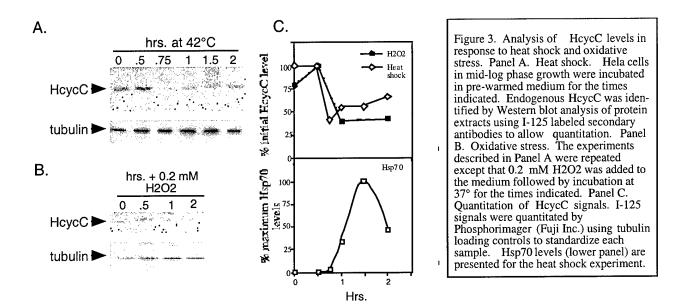


<u>Task 2: Examine HcycC regulation in response to</u> stress agents, heat shock, nutrient deprivation and hypoxia.

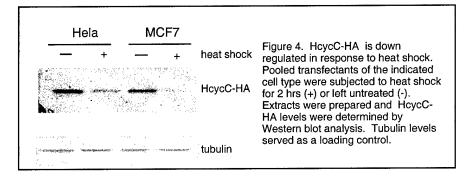
The human cyclin C is destroyed in response to stress.

The C-type cyclin subfamily shares several characteristics including association with the RNA polymerase holoenzyme and displaying a constant protein level throughout the cell cycle. To determine if the novel stress-induced destruction observed for Ume3p is also conserved, the levels of the human cyclin C (HcycC) were examined in mammalian cells exposed to stress. Antibodies raised against the endogenous HcycC (a gift from E. Lees, DNAX Inc.) were used to probe Hela cell extracts exposed to either heat shock (42°) or oxidative stress in the form of H2O2 (0.2 mM). To determine the decay kinetics of HcycC in cells exposed to stress, timecourse studies were performed. The Hela cells were grown to mid-log phase (approximately 70% confluent) prior to heat shock (42°). The cells were harvested at the indicated times and protein

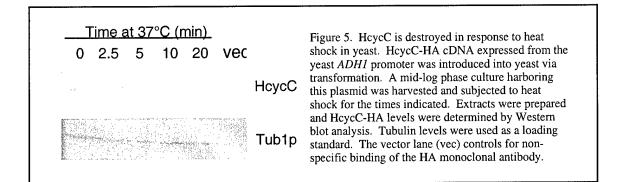
extracts prepared. Western blot analysis revealed that HcycC levels remained constant for 30 min. then fell to about 40% of their initial values for the two-hour duration of the experiment (Fig. 3A). The reduction in HcycC levels mirrored the increase in Hsp70 levels as indicated by Western blot analysis (Fig. 3C). The reduction of HcycC coincident with Hsp70 production mirrors our observations with Ume3p in yeast (2). Repeating these experiments with H2O2 treatment revealed decay kinetics for HcycC similar to heat shock (Fig. 3B and C). These results indicate that HcycC, similar to its yeast counterpart, is down regulated in response to stress suggesting that the C-type cyclin destruction pathway is conserved between yeast and man.



The next step to develop the HcycC-HA system is to determine whether the epitope tagged derivative is still destroyed in response to stress. The transfected Hela and MCF7 cell lines were grown to mid log phase, harvested, split and either left untreated or subjected to heat shock (42°C) for 2 hr. Western blots of protein extracts prepared from these samples revealed a significant reduction in HcycC-HA levels in both cell lines compared to the untreated controls (Fig. 4). These results indicate that HcycC-HA responds to stress in a manner similar to the endogenous protein. Moreover, the reduction in HcycC levels is promoter independent since both the endogenous and CMV driven HcycC proteins respond to stress in a similar manner. These results are consistent with the change in protein levels being the result of differences in stability. These studies are currently being repeated using pulse-chase experiments to confirm our hypothesis.

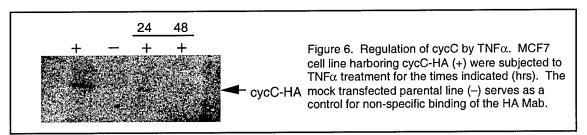


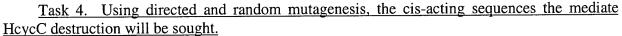
The final, and most stringent, test for the conservation of the stress-induced destruction pathway is the ability of the yeast system to recognize and destroy HcycC in response to stress. The HcycC-HA cDNA was placed under the control of the constitutive yeast *ADH1* promoter. Extracts prepared from yeast cultures harboring this construct contained an HA-dependent band the expected size for HcycC (33 kDa) but not in the vector control strain (data not shown). As an initial test, the HcycC-HA containing strain was subjected to heat shock (37°). Western blot analysis revealed that HcycC levels were reduced in response to heat shock (Fig. 5). As expected, no difference was observed in the HcycC mRNA accumulation from the *ADH1* promoter indicating that the reduction in HcycC-HA was most likely due to posttranscriptional mechanisms (data not shown). These findings indicate that not only is the pathway conserved, but to a remarkably high degree.



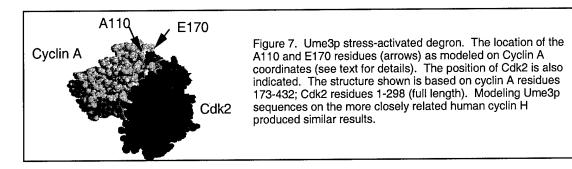
<u>Task 3: Examine HcycC regulation in response to cytotoxic agents doxorubicin, taxol, and</u> <u>5-fluoro-uracil.</u>

Due to the wide range of response kinetics observed for different stressors, an alternative approach was utilized to confirm our results from the heat shock experiments. Rather than subject the cells to stress, they were instead treated with the cytokine tumor necrosis factor alpha or TNFa. TNFa is produced by several cell types (e. g., macrophages, lymphoid cells) in response to inflammation, infection and other extracellular insults including ROS exposure (13). TNFα elicits a spectrum of cellular responses most notably fever, tumor necrosis and apoptosis (13, 14). One mechanism by which TNFa functions is through activation of the Jun N-terminal kinase/Stress Activated MAP kinase (JNK/SAPK) signal transduction cascade which represents a major stress-sensing pathway (7). In addition, $TNF\alpha$ appears to stimulate ROS production itself as part of its cell destruction function (10). To examine whether TNFa exposure affected HcycC levels, MCF7 cells transfected with HcycC-HA were treated with TNF α (2 µg/ml) for 24 and 48 hrs . A control flask was maintained for the same period but $TNF\alpha$ was omitted. Compared to the control, HcycC levels were significantly reduced in the MCF7 cell line exposed to TNF α at both the 24 and 48 hr timepoint (Fig. 6). Given the pleiotropic nature of TNF α response pathways, the precise nature of the signal leading to HcycC down regulation is not known. Although currently being confirmed by pulse chase experiments, these results are consistent with a model that HcycC is down regulated in response to stress. We are currently testing the impact of H2O2 on HcycC in both yeast and mammalian cells. Given that both cyclins are destroyed in yeast, it is possible that they share a common destruction element.





The results from these experiments indicate that the regulatory systems controlling the yeast and human C-type cyclin are well conserved. To identify cis-acting destruction elements required to mediate stress-induced degradation, we took advantage of a genetic approach in yeast that allows the selection of heat resistant derivatives of the yeast cyclin (3). Using a combination of directed and random mutagenesis, several cis-acting elements were identified (2). The two mutations identified in the genetic assay (A110V and E170K) are separated by 60 amino acids (Fig. 7). This finding is in sharp contrast to the relatively small, continuous regions (destruction box, PEST domains) required for destruction of cyclins that control cell cycle progression (9, 4, 12) or the deg1 destruction domain or degron required for Mat α 2 destruction (6). However, when the A110 and E170 residues are modeled onto either the human cyclin A (5) or cyclin H (1) structures, these two residues are brought together on the side of Ume3p away from Cdk binding (Fig. 7). These findings suggest the possibility that this region defines a new interactive face for cyclins.



Objective 2: Does HcycC activity effect drug sensitivity in transformed or nontransformed breast cell lines?

Task 5: Examine effect of overexpression of HcycC on drug resistance.

Two vectors were constructed that overexpressed HcycC in cos cells. These plasmids contained an SV40 origin for plasmid replication and expressed HcycC from the CMV promoter described above. The transient transfectants (and vector alone controls) were treated with three drugs used in breast cancer regimens namely doxorubicin, taxol, and 5-fluoro-uracil. No difference in viability was observed in pooled transfected exposed to either of these drugs compared to the mock transfected control (data not shown). These results indicate that overexpression of HcycC alone is insufficient to cause a significant change in drug sensitivity. However, since HcycC activates the cyclin dependent kinase Cdk8 (11), overexpressing the cyclin alone may not be functionally relevant. Therefore, experiments are ongoing in which the HcycC and Cdk8 will be simultaneously overexpressed and the drug sensitivities re-examined.

Task 6. Examine the role of HcycC in HSP gene expression.

No experiments have been completed toward this task due to grant time constraints.

Task 7: Examine drug resistance of tumor cells deleted for 6q21 with or without HcycC expression.

No experiments have been completed toward this task due to grant time contraints.

(7) KEY RESEARCH ACCOMPLISHMENTS:

- HcycC levels are reduced in response to heat shock and $TNF\alpha$ treatment
- A new destruction element (Stress-induced degron) has been identified that is required for the destruction of the yeast C-type cyclin in response to stress and differentiation cues.

(8) REPORTABLE OUTCOMES:

Cyclin C is destroyed in Response to stress in Breast Cancer cells. Daniel E. Egeland, Michael J. Mallory and Randy Strich. Manuscript in preparation.

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Abstract: Stress-Induced Destruction of C-Type Cyclins in Yeast and Man (D-14). 1996. Yeast Genetics & Human Disease. Baltimore, MD

Abstract: Cyclin C Is Destroyed In Response To Stress In Breast Cancer Cells. 2000. Era of Hope Meeting. Atlanta, GA

Cell Lines. Two cell lines, Hela and MCF7, have been established that express a stably integrated HcycC-HA tagged construct.

(9) CONCLUSIONS:

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Similar to the findings in yeast, the human HcycC levels are influenced by stress. This conclusion has been shown in two ways. First, HcycC levels are reduced in cells exposed to elevated temperatures or reactive oxygen. Second, treating cells to TNF α , a known activator of the stress response pathway, also reduces HcycC concentrations. The next step will require the actual half life of HcycC be determined using pulse chase experiments to ascertain whether the loss in cyclin levels is due to changes in stability or in translation efficiency.

(10) REFERENCES:

1. Andersen, G., D. Busso, A. Poterszman, J. R. Hwang, J. M. Wurtz, R. Ripp, J. C. Thierry, J. M. Egly, and D. Moras. 1997. The structure of cyclin H: common mode of kinase activation and specific features. EMBO J 16:958-67.

2. Cooper, K. F., M. J. Mallory, J. S. Smith, and R. Strich. 1997. Stress and developmental regulation of the yeast C-type cyclin *UME3* (*SRB11/SSN8*). EMBO J. 16:4665-4675.

3. **Cooper, K. F., and R. Strich.** 1999. Functional analysis of the yeast C-type cyclin Ume3p/Srb11p- RNA polymerase II holoenzyme interaction. Gene Exp **8:**43-57.

4. Glotzer, M., A. W. Murray, and M. W. Kirschner. 1991. Cyclin is degraded by the ubiquitin pathway. Nature **349:**132-138.

5. Jeffrey, P. D., A. A. Russo, K. Polyak, E. Gibbs, J. Hurwitz, J. Messague, and N. P. Pavletich. 1995. Mechanism of CDK activation revealed by the structure of cyclin A-CDK2 complex. Nature **376**:3113-320.

6. Johnson, P. R., R. Swanson, L. Rakhilina, and M. Hochstrasser. 1998. Degradation signal masking by heterodimerization of MATα2 and MATa1 blocks their mutual destruction by the ubiquitin-proteasome pathway. Cell 94:217-227.

7. Leppa, S., and D. Bohmann. 1999. Diverse functions of JNK signaling and c-Jun in stress response and apoptosis. Oncogene 18:6158-62.

8. Lew, D. J., V. Dulic, and S. I. Reed. 1991. Isolation of three novel human cyclins by rescue of G1 cyclin (Cln) function in yeast. Cell 66:1197-1206.

9. Murray, A. W., and M. W. Kirschner. 1989. Cyclin synthesis drives the early embrionic cell cycle. Nature **339:**275-286.

10. **Obrador, E., J. Navarro, J. Mompo, M. Asensi, J. A. Pellicer, and J. M. Estrela.** 1998. Regulation of tumour cell sensitivity to TNF-induced oxidative stress and cytotoxicity: role of glutathione. Biofactors 8:23-6. 11. **Rickert, P., W. Seghezzi, F. Shanahan, H. Cho, and E. Lees.** 1996. Cyclin C/CDK8 is a novel CTD kinase associated with RNA polymerase II. Oncogene **12:**2631-2640.

12. Salama, S. R., K. B. Hendricks, and J. Thorner. 1994. G1 cyclin degradation: the PEST motif of yeast Cln2 is necessary, but not sufficient, for rapid protein turnover. Mol. Cell. Biol. 14:7953-7966.

13. **Tracey, K. J., and A. Cerami.** 1993. Tumor necrosis factor, other cytokines and disease. Annu Rev Cell Biol **9:**317-43.

14. Vandenabeele, P., W. Declercq, B. Vanhaesebroeck, J. Grooten, and W. Fiers. 1995. Both TNF receptors are required for TNF-mediated induction of apoptosis in PC60 cells. J Immunol 154:2904-13.

15. Wilson, I. A., H. L. Niman, R. A. Houghten, A. R. Charenson, M. L. Connolly, and R. A. Lerner. 1984. The structure of an antigenic determinant in a protein. Cell **37:**767-778.

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