AD\_\_\_\_\_

Award Number: DAMD17-98-1-8107

TITLE: Growth Inhibition of Breast Tumor Cells by Hypodense and Normodense Eosinophilic Cell Lines

PRINCIPAL INVESTIGATOR: Paulette Furbert-Harris, Ph.D.

CONTRACTING ORGANIZATION: Howard University Washington, DC 20059

REPORT DATE: July 2000

TYPE OF REPORT: Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release; Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

## 20010531 037

|  |  | GE  | Form Approved<br>OMB No. 074-0188   |   |
|--|--|---|---|---|
| REPORT DOCUMENTATION PAGE<br>Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing int<br>the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any of<br>the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any of<br>the data needed, and completing and reviewing this collection of information operations and Reports, 1215 Jefferson Davis |  |   | tructions, searching exi  | sting data sources, gathering and maintaining   |
| he data needed, and completing and reviewing this  | equices. Directorate for Information Operations an   | ding this burden estimate or any o<br>nd Reports, 1215 Jefferson Davis  | ther aspect of this collec<br>Highway, Suite 1204, A  | rlington, VA 22202-4302, and to the Office of   |
| Anagement and Budget, Paperwork Reduction Pro  | pject (0704-0188), Washington, DC 20503<br>2. REPORT DATE  | 3. REPORT TYPE AND  | DATES COVERE  | D   |
| iank)  | July 2000  | Annual (15 Jun  |   |   |
| . <b>TITLE AND SUBTITLE</b><br>Growth Inhibition of Br<br>Normodense Eosinophilic  | east Tumor Cells by Hy<br>Cell Lines   | podense and   | 5. FUNDING NU<br>DAMD17-98-   |   |
| A <b>UTHOR(S)</b><br>Paulette Furbert-Harris   | , Ph.D.  |   |   |   |
| 7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES)<br>Howard University  |  |   | 8. PERFORMING ORGANIZATION<br>REPORT NUMBER   |   |
| Washington, DC 20059   |  |   |   |   |
| E-MAIL:<br>pfurbert-harris@fac.howard.edu<br>9. SPONSORING / MONITORING AG   | ENCY NAME(S) AND ADDRESS(ES)   | 1   |   | NG / MONITORING<br>EPORT NUMBER   |
| U.S. Army Medical Research and<br>Fort Detrick, Maryland 21702-50  | Materiel Command   |   |   |   |
| 11. SUPPLEMENTARY NOTES  | This report contains colored p   | hotos   | <b>I</b>  |   |
| 12a. DISTRIBUTION / AVAILABILITY<br>Approved for public release; distr   | STATEMENT<br>ibution unlimited   |   |   | 12b. DISTRIBUTION CODE  |
| 13. ABSTRACT (Maximum 200 Word   | is)  |   |   | . <b>I</b>  |
| (mediators) that inhi<br>and 72hr) from eosin<br>231 breast tumor cell<br>and MDA-MB-231<br>their inhibitory capac<br>the presence of IL-4<br>exogenous cytokines<br>IL-4, IL-5 were mar<br>that eosinophils proc<br>growth. Moreover,<br>growth inhibition of   | were based on the hypoth<br>bit the growth of breast tume<br>ophil cell lines were examina-<br>l colony formation. Serial 2-<br>colony formation in vitro. The<br>city 72hr > 48hr > 24hr. Pri-<br>c, and current examination re-<br>s were used, TNF $\alpha$ potently<br>ginal in their activity (10-35<br>huce mediators (some of wh<br>the data suggest potential co-<br>tumor cells. The establishme<br>of eosinophil mediators and f | or cells <u>in vitro</u> . Cu<br>ed for growth inhib<br>fold dilutions (1:2, 1<br>The time of harvest<br>or testing of eosince<br>evealed the presen<br>inhibited colony for<br>%) inhibitory activition<br>ollaborative activition | iltured superr<br>ition of MCF<br>:4 and l:8) inh<br>ing of the sup<br>phil condition<br>ce of TNFα (<br>ormation of M<br>ity. These da<br>es), which can<br>y of eosinoph<br>ell lines offer | hatants (24hr, 48hr<br>-7 and MDA-MB-<br>hibited both MCF-7<br>bernatants affected<br>hed media revealed<br>>40pg/ml). When<br>1CF-7, while IL-3,<br>ta clearly illustrate<br>n inhibit tumor cell<br>hil mediators in the<br>a rich resource for |
| 14. SUBJECT TERMS  |  |   |   | 15. NUMBER OF PAGES   |
| Breast Cancer  |  |   |   | 47<br>16. PRICE CODE  |
| 17. SECURITY CLASSIFICATION<br>OF REPORT   | 18. SECURITY CLASSIFICATION<br>OF THIS PAGE  | 19. SECURITY CLASS  | IFICATION   | 20. LIMITATION OF ABSTRAC   |
| Unclassified<br>NSN 7540-01-280-5500   | Unclassified   | Unclassi  | -   | Unlimited<br>Indard Form 298 (Rev. 2-89)  |

| Standary L    | OUII | 720    | [ <b>TUY</b> . | 2-0 |
|---------------|------|--------|----------------|-----|
| Prescribed by | ANSI | Std. 2 | 239-18         |     |
| 298-102       |      |        |                |     |

#### FOREWORD

Opinions, interpretations, conclusions and recommendations are those of the author and are not necessarily endorsed by the U.S. Army.

\_\_\_\_ Where copyrighted material is quoted, permission has been obtained to use such material.

\_\_\_\_ Where material from documents designated for limited distribution is quoted, permission has been obtained to use the material.

Citations of commercial organizations and trade names in this report do not constitute an official Department of Army endorsement or approval of the products or services of these organizations.

X In conducting research using animals, the investigator(s) adhered to the "Guide for the Care and Use of Laboratory Animals," prepared by the Committee on Care and use of Laboratory Animals of the Institute of Laboratory Resources, national Research Council (NIH Publication No. 86-23, Revised 1985).

X For the protection of human subjects, the investigator(s) adhered to policies of applicable Federal Law 45 CFR 46.

 $\underline{X}$  In conducting research utilizing recombinant DNA technology, the investigator(s) adhered to current guidelines promulgated by the National Institutes of Health.

 $\underline{X}$  In the conduct of research utilizing recombinant DNA, the investigator(s) adhered to the NIH Guidelines for Research Involving Recombinant DNA Molecules.

 $\underline{N/A}$  In the conduct of research involving hazardous organisms, the investigator(s) adhered to the CDC-NIH Guide for Biosafety in Microbiological and Biomedical Laboratories.

Date

#### **Table of Contents**

9 L

#### Page Number

----

| Front Cover                  | 1  |
|------------------------------|----|
| Standard Form 298            | 2  |
| Foreward                     | 3  |
| Table of Contents            | 4  |
| Introduction                 | 5  |
| Body                         | 5  |
| Key Research Accomplishments | 7  |
| Reportable Outcomes          | 7  |
| Conclusion                   | 8  |
| References                   | 9  |
| Appendices                   | 10 |

#### 5. Introduction

5

Eosinophils are nonimmune inflammatory cells principally involved in type I hypersensitivity and parasitic infections (1). In addition to their granular proteins which are toxic to surrounding tissues during inflammatory reactions, eosinophils also produce an array of cytokines which in addition to autologously regulating them, may also have anti-cancer activities (2). Eosinophils also produce perforin and granzymes and hence have the capacity to kill tumor targets similarly to cytotoxic T lymphocytes and natural killer cells (3). In this investigation we have hypothesized that eosinophils, like NK cells may be non specific anti-cancer effector cells and that their mechanism of killing of tumor cells is two-tiered; I) by binding to and releasing toxic granules like perforin and granzymes which cause the tumor cells to die by apoptosis and 2) by release of cytokines which inhibit tumor growth. We have immortalized subpopulations of eosinophils from individuals with mild to moderate eosinophilia. Hypo- and hyperdense eosinophils were collected from metrizamide density gradient fractions 22 and 24. These cells have been characterized by flow cytometry using a panel of fluorescein-labelled antibodies to surface markers found on eosinophils. Previous reports demonstrated that these cell lines like fresh peripheral blood eosinophils inhibit tumor cell growth. In this report we demonstrate the ability of conditioned media (cultured supernatants) and exogenous cytokines to inhibit tumor cell colony formation.

#### 6. Body

Collection of Cultured Supernatants from Eosinophil Cell Lines. Eosinophil cell lines (4) and sublines (3) were incubated at  $1 \times 10^6$  /ml for 24, 48 and 78hrs in RPMI medium supplemented with sodium pyruvate (1 mM), non-essential amino acids (1 mM), penicillin/streptomycin (50 units/ml, respectively), gentamycin (50 ug/ml) complete medium and 1% fetal bovine serum. The supernatants were collected after centrifugation at 2,000rpm for 30 mins, aliquoted and stored at  $-20^{\circ}$ C. Growth Inhibition of MCF-7 and MDA-MB-231 tumor cells by Eosinophil Cultured Supernatants.

a) Monolayer- MCF-7 and MDA-MB-231 tumor cells were seeded into the wells of a 6-well cluster plate at lx10<sup>5</sup> cells per well, and incubated overnight (16-24hr) at 37°C, 5% CO<sub>2</sub>. Supernatants from eosinophil cultures were added (l ml/well). Fresh complete RPMI medium containing 10% fetal bovine serum was added to triplicate wells per 6-well cluster. The plates were further incubated for an additional 48hrs or until the control wells were confluent. The eosinophils were removed from the wells which were washed 3x with phosphate buffered saline, then stained with hematoxylin and eosin. Photo microscopic documentation was carried out. Cell line GRC.014.24.S1 was unable to inhibit the growth of MRC-5 fibroblasts at 1:1, 2:1 and 5:1 E:T ratios (fig 2). On the otherhand, at 2:1 and 5:1, MCF-7 cells were inhibited. Cell line (fig 3) GRC.014.22S completely (fig 3) abrogated MCF-7 growth at 1:1 and 2:1 E:T ratios. Growth inhibition of MDA-MB-231 by these cell lines was more varied. The greatest inhibition with both cell lines was at the E:T ratio of 2:1.

**b)**Colony formation - Tumor cells were seeded into 6-well plates at 100 cells per well. The plates were incubated overnight at 37°C. Conditioned media supernatants from eosinophil cell line cultures (24, 48 and 72hrs) were added to the wells in total volume of 2mls. Serial 2-fold dilutions of the conditioned supernatants were made and each dilution was added in triplicate to the cells. Each plate contained triplicate control wells (10% RPMI medium). The plates were then incubated for 10 days at 37°C, 5% CO<sub>2</sub>. The wells were washed carefully with phosphate buffered saline, then stained with hematoxylin and eosin. Manual counts were taken and percent inhibition determined.

Concentrated undiluted supernatants completely inhibited colony formation. With the 1:2 dilution of cultured supernatant from the hypodense cell line GRC.014.22 (fig 5), there was 73% inhibition of MCF-7 colony formation and 93% with both the 48hr and 72hr supernatants. At the 1:4 dilution there was 69% inhibition with the 24hr supernatant, 81% with the 48hr supernatant and 88% with the 72hr preparation. At the 1:8 dilution 30% inhibition occurred with the 24hr supernatant, 52% with the 48hr supernatant and 71% with the 72hr preparation.

When MDA-MB-231 tumor cells were analyzed, both the concentrated and the 1:2 dilution inhibited colony formation by 100% (fig 6). However, with the 1:4 and 1:8 dilutions, 24hr, 48hr and 72hr were quite similar in pattern in that 30-45% inhibition was obtained. When the 1:2 diluted supernatants from the eosinophil cell line GRC.0l4.24 (fig7) were assayed, there was an increase in inhibition (81% to 93%) with the 24hr and 48hr supernatants. This decreased slightly to 87% with the 72hr sample. With the 1:4 dilution, there was 65% inhibition with the 24hr supernatant, 91% and 92% with the 48hr and 72hr preparations. With the 1:8 dilutions, MCF-7 colony growth was inhibited by 34% with the 24hr supernatant, 72% and 73% with the 48hr and 72hr preparations. MDA-MB-231 tumor cells (fig 8) were more resistant than MCF-7 cells to the 1:4 and 1:8 dilutions from all 3 culture times (24hr, 48hr and 72hr). The percent inhibition of colony formation for the 1:4 dilutions of the three culture preparations were 37, 50 and 60%, respectively and very similar for the 1:8 at 37, 40 and 60%, respectively. The 1:2 dilution by 100%.

A similar pattern of activity though lower was observed with cell line SD.031.22 (fig 9) in that when the 24hr supernatant was tested on MCF-7 cells, at 1:2 dilution there was 60% inhibition, 55% at 1:4 and 25% at 1:8. The pattern of activity was the same for both the 48hr and 72hr cultured supernatants however the percent inhibition was higher, 74%, 70%, 38% (48hr); and 97%, 74% an 38% (72hr). Activity against MDA-MB-231 was very similar (fig 10). Cultured Supernatants from eosinophil cell line SD.031.24 inhibited MCF-7 (fig 11) tumor cells similarly to the previous cell lines GRC.014.22 (fig 5) and GRC.014.24 (fig 7) and SD.031.22 (fig 9). Inhibition by the 24hr supernatants was 92%, 60% and 54% for the 1:2, 1:4 and 1:8 dilutions, respectively. Inhibition by the 48hr supernatant was 98%, 88% and 75% for the same dilutions, respectively. The 72hr cultured supernatant showed the same pattern of inhibition. The inhibitory activity of SD.031.24 supernatants against MDA-MB-231 (fig 12) was less than that against MCF-7, similarly to the GRC.014.24 supernatants. Sublines of the GRC.014 cell lines were developed by sterile sorting with a Fluorescent Activated Cell Sorter using CCR3 (Eotaxin Receptor) and CD49d as markers. Supernatants (24hr, 48hr and 72hr) from FACS sorted eotaxin receptor positive GRC 014.22 cells (referred to as GRC 014.22 S) (Fig 13) inhibited MCF-7 colony formation by 100%, 88% and 71% (1:2, 1:4 and 1:8, respectively), 100%, 91% and 64% (48hr); 100%, 84% and 77% (72hr). At 1:4 and 1:8 dilutions (fig 14) MDA-MB-231 cells were inhibited by  $\geq$  50%. Against MCF-7, the GRC.014.24S eotaxin receptor positive eosinophil

supernatants markedly inhibited colony formation 90%, 92% and 96% from 1:2 dilutions of 24hr, 48hr and 72hr cultured supernatants (fig 15). At 1:4 dilutions, inhibition was 50%, 75% and 85% for 24, 48 and 72hr cultures, respectively and at the 1:8 dilution percent inhibition was 26%, 54% and 63% for 24, 48 and 72hr cultured preparations, respectively. These supernatants also inhibited MDA-MB-231 cells (fig 16).

The last cell line tested was established from the GRC.014.24S subline, using the FACS Sorter and second eosinophil marker, CD49d. These cells are CCR3<sup>+</sup> Cultured supernatants from these cell line also inhibited MCF-7 (fig 17) and MDA-MB-231 (fig 18) in a dose-dependent manner.

#### Cytokine Presence in Eosinophil Cultured Supernatants.

24hr, 48hr and 72hr cultured supernatants from 7 eosinophil cell lines (parent and sublines) were evaluated by enzyme-linked immunoassay (ELISA) analysis for the presence of TNF $\alpha$ . TNF $\alpha$  was found in all samples at >40 pg/ml concentration.

#### Effect of Exogenous Cytokines on MCF-7 Colony Formation

IL-3 (fig 19) at 10ng/ml inhibited MCF-7 colony formation by 15%; at 50ng/ml by 30%, 0% at 100ng/ml and 26% at 200ng/ml. IL-4 inhibited colony formation by 10% at 10ng/ml, 25% at 50ng/ml and 14 at 100ng/ml. IL-5 at 10ng/ml failed to inhibit colony formation at 50 and 100ng/ml exerted 20% inhibition. At 10ng/ml IL-3 inhibited MDA-MB-231 (fig 20) colony formation by 10%, but had little to no effect any of the higher concentrations. At 10ng/ml, IL-4 inhibited colony formation by 17%, 28% at 50ng/ml, 20% at 100ng/ml and 16% at 200ng/ml. At 100ng/ml IL-5 inhibited tumor growth by 36%. TNF $\alpha$  on the other hand, dramatically inhibited colony inhibition of MCF-7 cells (fig 21)

#### 7. Key Research Accomplishments

- Promotion to Associate Professor
- Development of eosinophil sublines from 2 parental lines (3) based on FAC Sorting, using eosinophil markers
- Established inhibition of tumor growth by all 4 parent lines as well as 3 sublines

#### 8. Reportable Outcomes

,

1

- Activated eosinophil subpopulations kill breast tumor cells in vitro. Furbert-Harris PM, Howland C, Hunter KA, Laniyan I, Vaughn T, Dunston GM, Parish-Gause D, Harris D, Abdelnaby A, Anderson D, Brown R, Awich J, Sumner s and Oredipe O. Era of Hope Department of Defense Breast Cancer Research Program, Proceedings Vol. II p. 674.
- Stimulation by swainsonine of myocardial sulfhydryl levels in high-dose doxorubicin chemotherapy. OA Oredipe, I Laniyan, WM Griffin, D Parish-Gause, T Vaughn, WR Green and PM Furbert-Harris. FASEB 14(8) 1155.

#### 9. Discussion/Conclusion

We hypothesized that activated esoinophils and eosinophil cell lines inhibit breast cancer cell growth by releasing inflammatory substances that slow down the growth of the cells or are toxic to the cells, thereby causing their death. In this phase we have analyzed seven eosinophil cell lines, parent and sublines which we have developed by EBV-immortalization of metrizamide density gradient fractions of hypo-(M22) and hyperdense (M24) eosinophils. These cells were obtained from the peripheral blood of individuals (prior study) with mild to moderate eosinophilia. We have collected cultured supernatants from these cell lines, (which have been determined to inhibit the growth of breast tumor cells in vitro), and analyzed them for their inhibitory action on MCF-7 and MDA-MB-231 tumor cells. Cultured supernatants (24hr, 48hr and 72hr) from all 7 cell lines dramatically inhibited colony formation of both MCF-7 and MDA-MB-231 tumor cells. Undiluted, preparations of the cultured supernatants completely abrogated the colony growth. Serial two-fold dilutions, 1:2, 1:4 and 1:8 inhibited tumor colony formation in a dose-dependent manner. Supernatant preparations from 24hr, 48hr and 72hr cultures demonstrated inhibition dose-dependently. All supernatants were examined for TNFα and all contained >40 pg/ml. Exogenous cytokines which are known to regulate eosinophil differentiation and activation (I L-3, IL-5, GM-CSF) and IL-4 and TNFa, all of which can be produced by activated eosinophils, were examined for tumor cell growth inhibitory activity. There was marginal growth inhibition with all except TNFa. IL-3 at 50 ng/ml inhibited MCF-7 colony formation by 30% while IL-5 at 100 ng/ml inhibited MDA-MB-231 colony formation by 36%. The data were variable with regards to dose with IL-3, IL-4, IL-5 and GM-CSF. Overall, inhibition ranged from 0-35%. TNFa was markedly potent in its inhibitory activity.

These data confirm our hypothesis that mediators released by eosinophil cell lines inhibit tumor cell growth. Further analysis of eosinophil supernatants will be performed in order to better characterize the mediator participants active in eosinophil anti-cancer cytotoxic activity. These cell lines offer an excellent resource for studying the biology and functional activity of eosinophils.

#### 10. References

1

- 1. Gaga M, Frew AJ, Veronica AV and Kay AB. 1991.
- 2. Molet S, Gosset P, Vanhee D, Tillie-Leblond I, Wallaert B, Capron Ma and Tonnel AB. Modulation of cell adhesion molecules on human endothelial cells by eosinophil-derived mediators. Jl Leuk Biol 63:351-58, 1998.
- 3. Kuby J. Immunology (4<sup>th</sup> ed) RA Goldsby, Thomas J Kinat, Barbara A Osborne (eds). WH Freeman & CO., 2000.

#### 11. Appendices

1

Figure 1. Cytospin Preparation of Eosinophil Cell Line Stained with H&E

- Figure I. Legend
- Figure 2. Eosinophil Cytotoxic Activity on MRC-5 Fibroblast Cell Growth
- Figure 2. Legend
- Figure 3. Eosinophil Cytotoxic Activity on MCF-7 Cell Growth
- Figure 3. Legend
- Figure 4. Eosinophil Cytotoxic Activity on MDA-MB-231 Cell Growth
- Figure 4. Legend
- Figure 5. GRC.014.22 Eosinophil Supernatants Inhibit MCF-7 Colony Formation In Vitro
- Figure 5. Legend
- Figure 6. GRC.014.22 Eosinophil Supernatants Inhibit MDA-MB-231 Colony Formation In Vitro
- Figure 6. Legend
- Figure 7. The Effect of GRC.014.24 Eosinophil Cultured Supernatants on MCF-7 Breast Tumor Cells Colony Formation
- Figure 8. The Effect of GRC.014.24 Eosinophil Cultured Supernatants on MDA-MB-231 Colony Formation
- Figure 7, 8. Legend
- Figure 9. Effect of SD.031.22 Eosinophil Cultured Supernatants on MCF-7 Colony Formation
- Figure 10. Effect of SD.031.22 Eosinophil Cultured Supernatants on MDA-MB-231 Colony Formation

Figure 9, 10. Legend

| Figure 11.            | Effect of SD.031.24 Eosinophil Cultured Supernatants on MCF-7 Colony   |  |
|-----------------------|--|--|
| riguite II.           | Formation  |  |
| Figure 12.            | Effect of SD.031.24 Eosinophil Cultured Supernatants on MDA-MB-231 Colony Formation  |  |
| Figure 11, 12.        | Legend   |  |
| Figure 13.            | Inhibition of MCF-7 Colony Formation by GRC.014.22S Supernatants   |  |
| Figure 14.            | Inhibition of MDA-MB-231 Colony Formation by GRC.014.22S Supernatants  |  |
| Figure 13, 14.        | Legend   |  |
| Figure 15.            | Cultured Supernatants from an Eotaxin Positive (CCR3 <sup>+</sup> ) Eosinophil Cell Line (GRC.014.24S) Inhibit MCF-7 Colony Formation      |  |
| Figure 16.            | Cultured Supernatants from an Eotaxin Positive (CCR3 <sup>+</sup> ) Eosinophil Cell Line (GRC.014.24S) Inhibit MDA-MB-231 Colony Formation |  |
| Figure 15, 16. Legend |  |  |
| Figure 17.            | Cultured Supernatants from CCR3 <sup>+</sup> , CD49d <sup>+</sup> Eosinophil Cell Line Inhibit MCF-7 Colony Formation                      |  |
| Figure 18.            | Cultured Supernatants from CCR3 <sup>+</sup> , CD49d <sup>+</sup> Eosinophil Cell Line Inhibit MDA-MB-231 Colony Formation                 |  |
| Figure 17, 18.        | Legend   |  |
| Figure 19,            | The Effect of Cytokines on MCF-7 Colony Formation  |  |
| Figure 19.            | Legend   |  |
| Figure 20.            | The Effect of Cytokines on MDA-MB-231 Colony Formation   |  |
| Figure 20.            | Legend   |  |
| Figure 21.            | Effect of TNFa on MCF-7 Colony Formation   |  |
| Figure 21.            | Legend   |  |

, I

Fig.1 Cytospin preparation of Eosinophil Cell Line stained with H&E

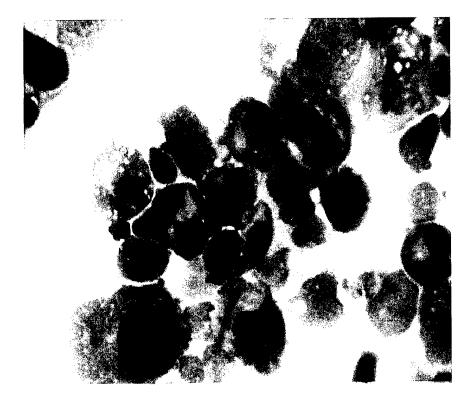
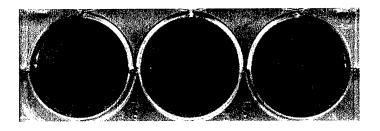


Fig 1. Cytospin preparation of eosinophil cell line stained with H & E

1 I

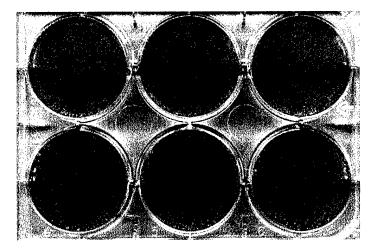
#### Fig.2 Eosinophil Cytotoxic Activity on MRC-5 Fibroblast Cell Growth





### Control - MRC-5

GRC.014.24.S1 (1:1)



GRC.014.24.S1 (2:1)

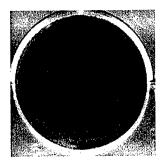
GRC.014.24.S1 (5:1)

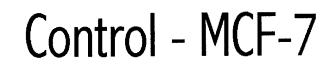
Fig. 2. MRC-5 human lung fibroblasts were seeded into the wells of a 6-well plate at  $1.5 \times 10^5$  cells/well and incubated overnight at 37°C. The eosinophil subline GRC.014.24 Sl was then added to the monolayers at effector to target (E:T) ratios of 1:1, 2:1, and 5:1. RPMI complete medium containing 10% fetal bovine serum was added to control wells. The experiments were performed in triplicate. The plates were further incubated for 48hrs or until the control wells were confluent.

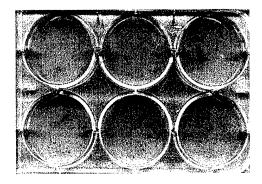
,

.

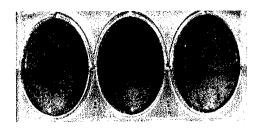
#### Fig.3 Eosinophil Cytotoxic Activity on MCF-7 Cell Growth







GRC.014.22.S (1:1) GRC.014.22.S (2:1)

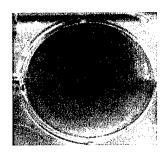


GRC.014.24.S1 (1:1)

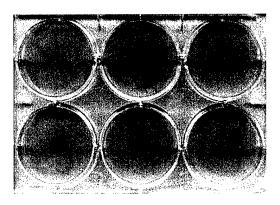
GRC.014.24.S1 (2:1) GRC.014.24.S1 (5:1) Fig. 3. MCF-7 tumor cells were seeded into the wells of 6-well cluster culture plates at  $1.5 \times 10^5$  cells/well and incubated overnight. Eosinophil cell lines GRC.014.22S and GRC.014.24SI at E:T ratios of 1:1, 2:1 and 5:1. The plates were incubated for an additional 48hr or until the control wells were confluent.

i 1

## Fig.4 Eosinophil Cytotoxic Activity on MDA MB 231 Cell Gowth

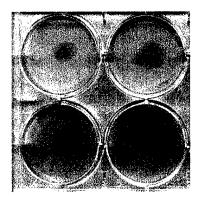


## Control - MDA MB 231



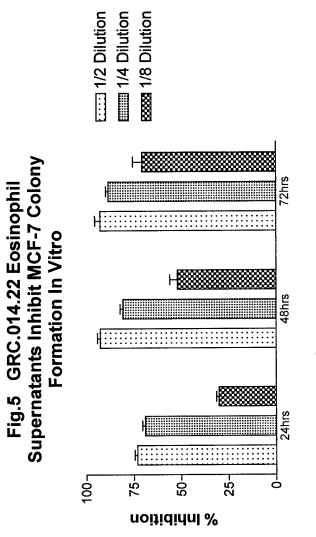
GRC.014.22.S (1:1) GRC.014.22.S (2:1)





GRC.014.24.S1 (1:1) GRC.014.24.S1 (2:1) GRC.014.24.S1 (5:1) Fig. 4. MDA tumor cells were seeded into the wells of 6-well cluster culture plates at  $1.5 \times 10^5$  cells/well and incubated overnight. Eosinophil cell lines GRC.014.22S and GRC.014.24Sl at E:T ratios of 1:1, 2:1 and 5:1. The plates were incubated for an additional 48hr or until the control wells were confluent.

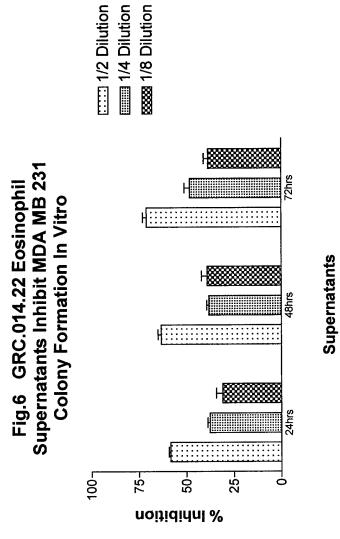
• •



,

Supernatants

Fig. 5. MCF-7 tumor cells were seeded into wells of a 6-well plate at 100 cells per well and incubated overnight. The cells were then incubated with cell free supernatants from eosinophil cell line GRC.014.22 cultures for 10 days at 37°C. Plates were washed 3x with phosphate-buffered saline; stained with H & E, then counted.



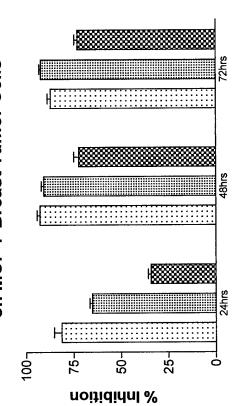
•

.

Fig. 6. MDA-MB-231 tumor cells were treated GRC.014.22 supernatants similarly to that described for MCF-7 in fig. 5. The colonies were stained and counted. Percent inhibition was determined.

,

L



Supernatants

## Fig.7 The Effect of GRC.014.24 Eosinophil Cultured Supernatants on MCF-7 Breast Tumor Cells

,

The contract of the contract o

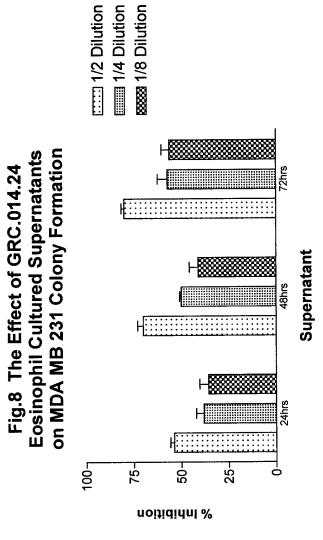
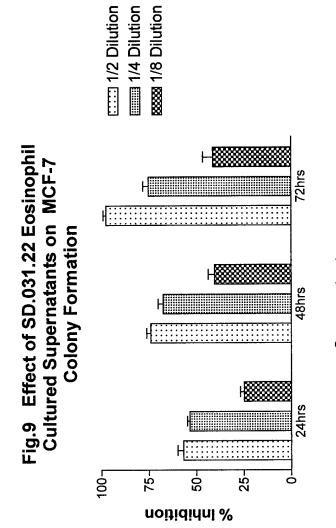


Fig. 7, 8. Cultured supernatants from GRC.014.24 eosinophils were examined for inhibitory activity against MCF-7 and MDA-MB-231 colony formation.

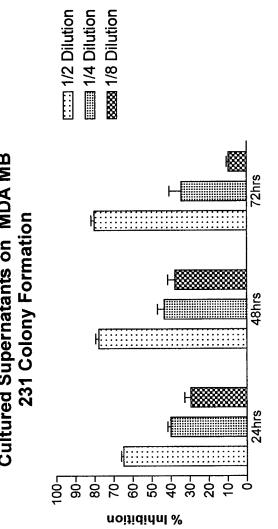
. .



,

t

Supernatant



Supernatant

# Fig.10 Effect of SD.031.22 Eosinophil Cultured Supernatants on MDA MB 231 Colony Formation

.

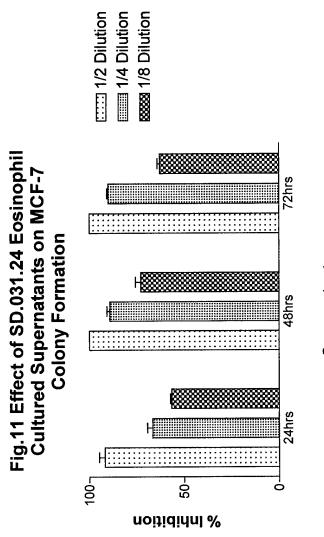
•

ı

Fig. 9, 10. Cultured supernatants from eosinophil cell line SD.031.22 were incubated with MCF-7 and MDA-MB-231 tumor cells. The cells were incubated for 10 days as described previously. After harvesting, and staining, the colonies were counted and the % inhibition determined.

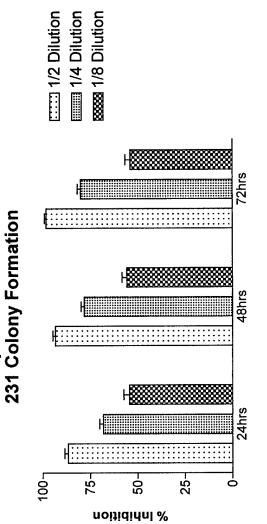
,

.



,

Supernatant

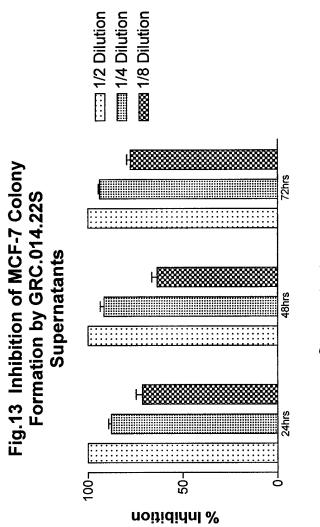


Supernatant

# Fig.12 Effect of SD.031.24 Eosinophil Cultured Supernatants on MDA MB 231 Colony Formation

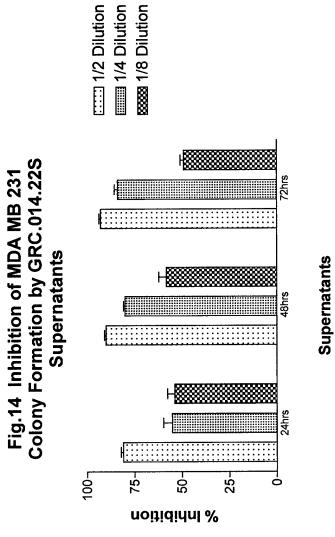
,

Fig.11, 12. MCF-7 (Fig 11) and MDA-MB-231 tumor cells were seeded into 6-well plates (100 cells/well) and incubated overnight. Control wells contained RPMI complete medium with 10% FBS. Serial toe-fold dilutions of cultured supernatants from the eosinophil cell line SD.031.24 were added and the plates were incubated for an additional 10 days. Colonies were stained with H & E and counted. Percent inhibition was determined.



,

Supernatants

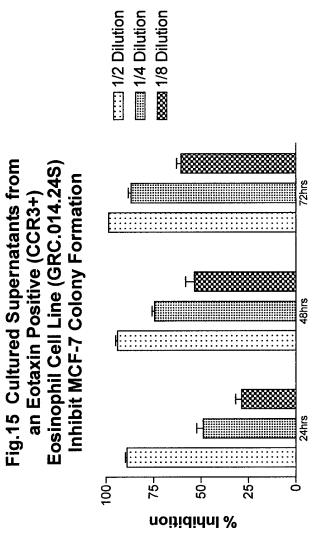


•

Fig. 13, 14. Supernatants from the eotaxin receptor positive subline GRC.014.22S were incubated with tumor cells MCF-7 and MDA-MB-231 (l00 cell/well) in 6-well plates as described previously. Percent inhibition of colony formation was determined.

٠

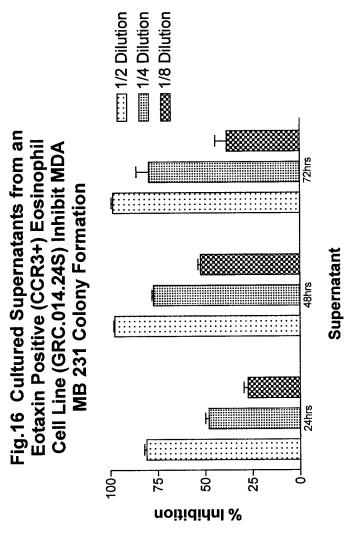
•



•

.

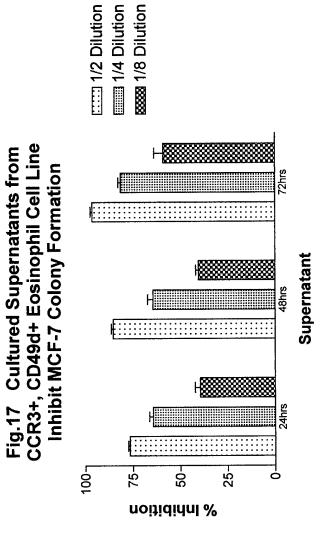




,

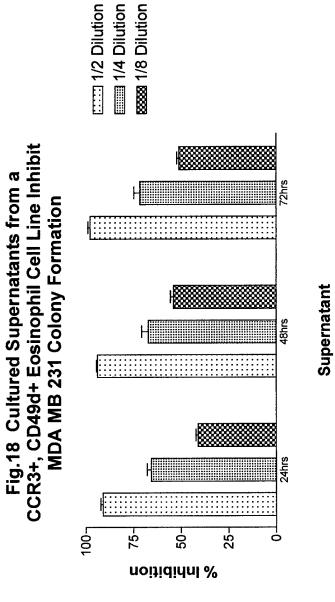
Fig. 15, 16. Cultured supernatants from the CCR3<sup>+</sup> eosinophil line GRC.014.24S similarly to that described in figures 13 and 14.

. .



•

,



a

,

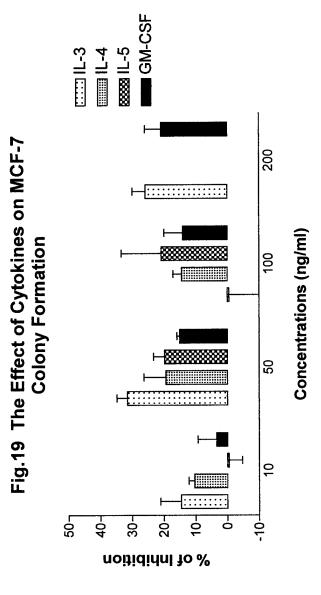
40

Fig. 17, 18. Cultured supernatants from CCR3<sup>+</sup> cells that were sorted using the CD49d marker, were incubated with MCF-7 and MDA-MB-231 tumor cells similarly to that described in figure 13 and 14.

r

· •

.



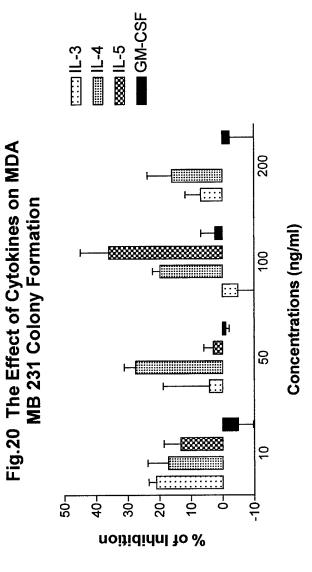
.

· · · •

.

Fig. 19. MCF-7 cells were seeded into 6-well plates (100 cell/well), then treated with cytokines IL-3, IL-4, IL-5, GM-CSF at 10, 50, 100 and 200ng/ml. The plates were incubated for 10 days; harvested; colonies stained then counted.

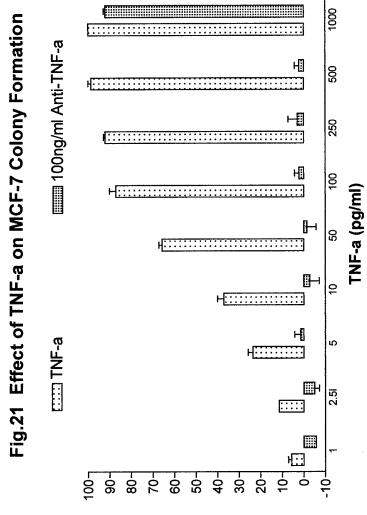
\* <sup>2</sup> •



J y

Fig. 20. MDA-MB-231 tumor cells were seeded; incubated with cytokines and examined for colony formation as described in fig. 18.

**ب ک** م



noitididal to %

46

Fig. 21. MCF-7 tumor cells were seeded into wells of 6-well plate at 100 cells/well and treated with different concentrations of TNF $\alpha$ , (200ul/well. Each concentration and media control was set up in triplicate. Anti-TNF $\alpha$  (100ng/ml) was also added to certain wells. The plates were further incubated for 10 days.

ور ۲۰ د