

AD-A281 635



①

GRANT NO: DAMD17-90-Z-0020

TITLE: WORLD REFERENCE CENTER FOR ARBOVIRUSES

PRINCIPAL INVESTIGATOR: Robert E. Shope, M.D.

S DTIC
ELECTE
JUL 18 1994
F

CONTRACTING ORGANIZATION: Yale University School of Medicine
333 Cedar Street
P.O. Box 20846
New Haven, CT 06510-8047

REPORT DATE: June 7, 1994

TYPE OF REPORT: Final Report

PREPARED FOR: U.S. Army Medical Research, Development,
Acquisition and Logistics Command (Provisional),
Fort Detrick, Frederick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for public release;
distribution unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

94-21498



1380

94 7 12 1 58

REPORT DOCUMENTATION PAGE

Form Approved
OMB No. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503.

1. AGENCY USE ONLY (Leave blank)	2. REPORT DATE 7 June 1994	3. REPORT TYPE AND DATES COVERED Final Report (2/9/90 - 12/8/93)	
4. TITLE AND SUBTITLE World Reference Center for Arboviruses		5. FUNDING NUMBERS Grant No. DAMD17-90-Z-0020	
6. AUTHOR(S) Robert E. Shope		8. PERFORMING ORGANIZATION REPORT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Yale University School of Medicine 333 Cedar Street P.O. Box 20846 New Haven, CT 06510-8047			
9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research, Development, Acquisition and Logistics Command (Provisional), Fort Detrick Frederick, Maryland 21702-5012		10. SPONSORING/MONITORING AGENCY REPORT NUMBER	
11. SUPPLEMENTARY NOTES			
12a. DISTRIBUTION/AVAILABILITY STATEMENT Approved for public release; distribution unlimited		12b. DISTRIBUTION CODE	
13. ABSTRACT (Maximum 200 words) The World Reference Center for Arboviruses Characterized Sabia and Guanarito viruses, two new arenaviruses causing hemorrhagic fever in Brazil and Venezuela, respectively. A new genotype of Japanese encephalitis virus was discovered from Indonesia. Classifications of Venezuelan encephalitis and arenaviruses were revised. Outbreaks of arenavirus hemorrhagic fever in Venezuela and Brazil were investigated. Military watchdogs had antibody to Japanese encephalitis virus. Rapid PCR-based diagnostic techniques for encephalitides were developed. Neutralization tests of sera of army volunteers supported the licensing of the Japanese encephalitis inactivated vaccine. Recombinant vaccinia viruses expressing the prM/E genes were developed as candidate vaccines. Reagents were distributed to laboratories in 24 countries. <div style="text-align: right;">DTIC QUALITY INSPECTED 6</div>			
14. SUBJECT TERMS Arbovirus, Arenavirus, Japanese encephalitis, Dengue, Vaccinia Recombinant, Vaccines, Hemorrhagic fever, Rift Valley fever, Serosurvey, ELISA, Yellow fever, Toscana virus		15. NUMBER OF PAGES	
17. SECURITY CLASSIFICATION OF REPORT Unclassified		18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	16. PRICE CODE
19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT Unlimited		

FOREWORD

Opinions, interpretations, conclusions and recommendations are those of the author and are not necessarily endorsed by the U.S. Army.

Where copyrighted material is quoted, permission has been obtained to use such material.

Where material from documents designated for limited distribution is quoted, permission has been obtained to use the material.

Citations of commercial organizations and trade names in this report do not constitute an official Department of the Army endorsement or approval of the products or services of these organizations.

RES In conducting research using animals, the investigator(s) adhered to the "Guide for the Care and Use of Laboratory Animals," prepared by the Committee on Care and Use of Laboratory Animals of the Institute of Laboratory Animal Resources, National Research Council (NIH Publication No. 86-23, Revised 1985).

RES For the protection of human subjects, the investigator(s) have adhered to policies of applicable Federal Law 45CFR46.

RES In conducting research utilizing recombinant DNA technology, the investigator(s) adhered to current guidelines promulgated by the National Institutes of Health.

Robert E. Slope June 7, 1994
PI Signature Date

TABLE OF CONTENTS

FRONT COVER.....1

SF 298 -- REPORT DOCUMENTATION PAGE.....2

FOREWORD.....3

TABLE OF CONTENTS.....4

INTRODUCTION.....5

BODY OF REPORT.....5

 1. Virus identification.....5

 a. TOGAVIRUSES.....5

 b. FLAVIVIRUSES.....5

 c. BUNYAVIRUSES.....5

 d. ARENAVIRUSES.....5

 2. Classification of arboviruses and arenaviruses.....5

 3. Diagnosis of disease.....6

 4. Serological surveys.....6

 5. Development of new techniques.....6

 6. Flavivirus vaccine development.....7

 a. EVALUATION OF JAPANESE ENCEPHALITIS PRE- AND POST-
VACCINATION PLAQUE REDUCTION NEUTRALIZATION IN
MILITARY SUBJECTS.....7

 b. DEVELOPMENT OF RECOMBINANT VACCINIA VIRUSES AS
FLAVIVIRUS CANDIDATE VACCINES.....7

 7. Low passage collection of arbovirus strains.....8

 8. Distribution of reagents.....8

CONCLUSIONS.....8

PERSONNEL SUPPORTED BY THE PROJECT.....8

PUBLICATIONS.....9

Accession For	
NTIS CRA&I	<input checked="" type="checkbox"/>
DTIC TAB	<input type="checkbox"/>
Unannounced	<input type="checkbox"/>
Justification	
By	
Distribution /	
Availability Codes	
Dist	Avail and/or Special
A-1	

INTRODUCTION

The World Reference Center for Arboviruses was established at the Yale Arbovirus Research Unit in 1965 as an outgrowth of The Rockefeller Foundation program on arboviruses which was moved in 1965 to Yale University from New York City. The U.S. Army has supported this program since 1972, initially through joint Navy-Army funding, then through separate contracts and grants, and during the past 3 1/2 years by this grant. The progress of this period is included in this report; it covers the work for the entire project which received support from the World Health Organization and NIH in addition to that of this grant.

BODY OF REPORT

1. Virus identification. Viruses were identified from Japan, Sudan, Venezuela, Brazil, United States, Indonesia, Thailand, Mexico, Egypt, Peru, Colombia, China, Malaysia, India, Italy, Senegal, Central African Republic, France, and Angola. Among these were:

a. TOGAVIRUSES. A variety 1E Venezuelan encephalitis virus was identified from a horse during an epizootic in Mexico. This was the first demonstration of this variety implicated in epizootic equine disease.

b. FLAVIVIRUSES. A new genotype of Japanese encephalitis virus was identified in Indonesia from among viruses isolated by the U.S. Naval Medical Research Unit Detachment in Jakarta. This genotype was recognized by sequencing 240 base pairs of the prM gene. The new genotype was represented in Bali, Flores, and Java.

c. BUNYAVIRUSES. Batai virus was identified for the first time from sera of human fever cases in Sudan. The virus was referred to the Center by the U.S. Naval Medical Research Unit #3 in Cairo.

d. ARENAVIRUSES. Sabia virus was a newly recognized arenavirus isolated from a fatal human case of hemorrhagic fever in Sao Paulo State, Brazil. The virus was identified in Brazil using reference reagents supplied from the Yale Center.

Guanarito virus was a newly recognized arenavirus isolated in Venezuela from a fatal human case and identified at Yale. This virus was shown to be the cause of Venezuelan hemorrhagic fever.

2. Classification of arboviruses and arenaviruses. The classification of the Venezuelan encephalitis virus complex was revised. During a study of the 1F variety from Sao Paulo Brazil, neutralization tests revealed that subtype 2 (Everglades) virus was more closely related to variety 1AB than previously described. It was proposed that subtype 2 be reclassified as a variety of subtype 1.

The classification of arenaviruses was revised to include the new members, Sabia and Guanarito from the New World. Complement fixation test results indicated that there are 2 sets of New World arenaviruses; one set contained Guanarito, Junin, Tacaribe, and Amapari; the other set contained Latino, Parana, Tamiami, Flexal, and Pichinde viruses.

3. **Diagnosis of disease.** An outbreak of hemorrhagic fever was investigated in Portuguesa State, Venezuela. Fifteen cases were confirmed by virus isolation and the clinical syndrome was described. Nine of the 15 cases were fatal. This was apparently a new disease for this region with a clinical picture very similar to that of Argentine and Bolivian hemorrhagic fevers.

A single fatal case of hemorrhagic fever in Sao Paulo State, Brazil was shown to be caused by a new arenavirus, Sabia virus. The exact site of exposure of the patient was not determined and is still not known.

Rift Valley fever returned to Egypt during 1993 after a 13-year absence. Sera of 3 cases of optic retinitis were submitted by the U.S. Naval Medical Research Unit #3. These were confirmed as Rift Valley fever by IgM capture ELISA.

Outbreaks of equine encephalitis in Colombia and Mexico were shown to be caused by Venezuelan encephalitis virus. These were the first epizootics of Venezuelan encephalitis recognized for two decades.

4. **Serological surveys.** A serosurvey was carried out of Egyptian human residents of a Nile Delta village that was gradually inundated over the past 10 years. High prevalence of West Nile and Sicilian sandfly fever virus antibodies is consistent with the hypothesis that the flooding conditions have created favorable breeding sites for arthropods that in turn increased the transmission of arboviruses.

Sera of 104 dogs, mostly patrol dogs, were referred by the U.S. Army Medical Research Unit, Korea. Sera of 60 dogs were positive in Korea by Japanese encephalitis neutralization test. Tests at Yale with other flaviviruses confirmed the probable specificity of the Japanese encephalitis tests and ruled out Zika, tick-borne flaviviruses, and West Nile (probably). There were some monotypic reactions to Tembusu and Pnom Penh bat viruses, indicating that some dogs were exposed also to other flaviviruses in addition to Japanese encephalitis.

Sera from Sao Paulo State, Brazil from patients suspected of leptospirosis were surveyed for antibody to Hantaan virus by immunofluorescence. One serum from Sao Paulo City was strongly positive.

5. **Development of new techniques.** A sensitive and rapid technique was developed for detection of St. Louis encephalitis and eastern encephalitis viruses using polymerase chain reaction and gel electrophoresis of amplified cDNA fragments.

The sensitivity and specificity of Vero cell lysate antigens of flaviviruses was examined and compared to those of antibody-captured mouse brain antigens for detection of IgG in yellow fever 17D vaccinees. While the classic mouse brain antigen was slightly more sensitive, the cell lysate method was cheaper, quicker, and of acceptable sensitivity.

Limited primer extension sequencing, approximately 240 base pairs, of dengue-1, dengue-2, and Japanese encephalitis viruses showed multiple genotypes for each virus and geographic clustering. This technique was applied to determine the molecular epidemiology (genotype distribution) of each of these flaviviruses.

6. Flavivirus vaccine development.

a. EVALUATION OF JAPANESE ENCEPHALITIS PRE- AND POST-VACCINATION PLAQUE REDUCTION NEUTRALIZATION IN MILITARY SUBJECTS. Serological studies supported the trial in U.S. Army personnel in Hawaii of the Biken inactivated mouse brain vaccine for Japanese encephalitis. Sera of 532 adult volunteers were tested by plaque reduction neutralization test. Two different dosing schedules produced substantial titers of neutralizing antibody at 60 and 180 days post-vaccination. Tests done at Yale correlated almost completely with tests done by different techniques at Biken and CDC, Fort Collins. The prevaccination sera were also tested at Yale by ELISA for yellow fever antibody (presumably secondary to vaccination). Those subjects with pre-existing yellow fever antibody had slightly better responses to Japanese encephalitis vaccine, but the difference was not statistically significant. The Japanese encephalitis test results were submitted to the Food and Drug Administration to support the request for licensure. The vaccine was subsequently approved and licensed for use in the U.S.

b. DEVELOPMENT OF RECOMBINANT VACCINIA VIRUSES AS FLAVIVIRUS CANDIDATE VACCINES. Japanese encephalitis, yellow fever, dengue-1, dengue-2, dengue-3, and dengue-4 recombinant vaccinia viruses were developed. Plasmids containing the prM/E genes were constructed and transfected into tissue culture cells infected with a rescuing vaccinia virus which then expressed the proteins. Candidate Japanese encephalitis vaccines were made in NYVAC attenuated vaccinia and ALVAC canary poxviruses. The NYVAC vaccinia was the basis of the candidate dengue vaccines.

Extensive preliminary studies showed that the prM gene was required for maturation of the Japanese encephalitis virus, and that recombinant vaccinia viruses with the prM/E genes secreted the proteins which formed 20 nm sub-viral particles that agglutinated goose cells and were immunogenic. No other flavivirus genes were needed to induce fully neutralizing antibodies.

The Japanese encephalitis and yellow fever recombinant vaccinia viruses immunized mice which resisted homologous virus challenge. The comparable NYVAC construct immunized pigs and markedly reduced viremia levels after challenge of the pigs.

The dengue NYVAC constructs induced antibody in mice, but at the end of the grant period had not yet been tested for ability to induce protection from challenge.

7. Low passage collection of arbovirus strains. A large collection of low passage arbovirus strains was developed and maintained lyophilized. Priority was given to yellow fever, dengue, chikungunya, California group encephalitis, Venezuelan encephalitis, St. Louis encephalitis, western encephalitis, eastern encephalitis, Japanese encephalitis, and other human and veterinary disease arboviruses. The original (or as close to original as was available) material was passaged once in C6/36 mosquito cells or in Vero cells. The resulting stock was lyophilized in aliquots. These were stored and distributed to any and all persons requesting material for study. The collection now contains in excess of 400 strains.

8. Distribution of reagents. Virus stocks, antigens, antibodies, cell lines, and live insects were distributed to laboratories in 24 countries. Data on reagents available for distribution were entered in a dBase-3+ data bank. More than 4,000 entries have been made.

CONCLUSIONS

The World Reference Center for Arboviruses received agents from the U.S. and foreign countries for characterization and identification. Sabia virus and Guanarito virus were two new arenaviruses that caused hemorrhagic fever in Brazil and Venezuela respectively. A new genotype of Japanese encephalitis virus was discovered from Indonesia. Batai virus was identified in sera of febrile humans for the first time from the Sudan. The classification of the Venezuelan encephalitis virus complex and the New World arenaviruses was revised. Outbreaks of arenavirus hemorrhagic fever in Venezuela and Brazil, of Rift Valley fever in Egypt, and of Venezuelan encephalitis in horses in Colombia and Mexico were investigated. Study of the epidemiology of Guanarito virus causing Venezuelan hemorrhagic fever implicated the cotton rat as a reservoir. Serosurveys found Japanese encephalitis and other flavivirus antibody in Korean watchdogs, hantavirus antibody in inhabitants of Sao Paulo, Brazil, and high prevalence of mosquito- and sand fly-borne viruses in Egyptians inhabiting a flooded village. Rapid PCR-based diagnostic techniques were developed for eastern encephalitis and St. Louis encephalitis. Neutralization tests of Army volunteers supported the licensing of the inactivated mouse brain vaccine for Japanese encephalitis. Recombinant vaccinia viruses expressing the prM/E genes were developed for Japanese encephalitis, yellow fever, and the four dengue serotypes. The Japanese encephalitis is a candidate vaccine and the dengue recombinants are potential candidate vaccines. The low passage collection of arboviruses was augmented, and reagents were distributed to laboratories in 24 countries.

PERSONNEL SUPPORTED BY THE PROJECT

Robert E. Shope, Principal Investigator

Shirley J. Tirrell, Associate in Research

Mark L. Wilson, Assistant Professor

PUBLICATIONS

Ansari, M.Z., Ajani, U.A., and Shope, R.E. Diagnosis of viruses by immunoassays. *Asian Pacific Journal of Allergy and Immunology* 11:167-175, 1993.

Ansari, M.Z., Shope, R.E., and Malik, S. Evaluation of Vero cell lysate antigen for the ELISA of flaviviruses. *J. Clin. Lab. Anal.* 7:230-237, 1993.

Arthur, R.R., El-Sharkawy, M.S., Cope, S.E., Botros, B.A., Oun, S., Morrill, J.C., Shope, R.E., Hibbs, R.G., Darwish, M.A., and Imam, I.Z.E. Recurrence of Rift Valley fever in Egypt. *Lancet* 342:1149-1150, 1993.

Barry, M., Patterson, J.E., Tirrell, S., Cullen, M.R. and Shope, R.E. The effect of chloroquine prophylaxis on yellow fever vaccine antibody response: Comparison of plaque reduction neutralization test and enzyme-linked immunosorbent assay. *Am. J. Trop. Med. Hyg.* 44:79-82, 1991.

Brown, S.E., Gorman, B.M., Tesh, R.B., and Knudson, D.L. Isolation of bluetongue and epizootic hemorrhagic disease viruses from mosquitoes collected in Indonesia. *Vet. Microbiol.* 32:241-251, 1992.

Brown, S.E., Gorman, B.M., Tesh, R.B., and Knudson, D.L. Coltiviruses isolated from mosquitoes collected in Indonesia. *Virology* 196:363-367, 1993.

Brown, S.E., Morrison, H.G., Karabatsos, N. and Knudson, D.L. Genetic relatedness of two new Orbivirus serogroups: Orungo and Lebombo. *J. Gen. Virol.* 72:1065-1072, 1991.

Chapman, L.E., Wilson, M.L., Hall, D.B., LeGuanno, B., Dykstra, E.A., Ba, K., And Fisher-Hoch, S.P. Risk factors for Crimean-Congo hemorrhagic fever in rural northern Senegal. *J. Inf. Dis.* 164:686-692, 1991.

Chastel, C., Main, A.J., Bailly-Choumara, H., LeGoff, F., and LeLay, G. Essaouira and Kala Iris: Two new orbiviruses in the Kemerovo serogroup, Chenuda complex, isolated from Ornithodoros (Alectorobius) maritinus ticks in Morocco. *Acta Virol.* 37:484-492, 1993.

Chen, W.R., Rico-Hesse, R., and Tesh, R.B. a new genotype of Japanese encephalitis virus isolated in Indonesia. *Am. J. Trop. Med. Hyg.* 47:61-69, 1992.

Chen, W.R., Tesh, R.B. and Rico-Hesse, R. Genetic variation of Japanese encephalitis virus in nature. *J. Gen. Virol.* 71:2915-2922, 1990.

Chung, S.I., Livingston, Jr., C.W., Edwards, J.F., Crandell, R.W., Shope, R.E., Shelton, S.A. and Collisson, E.W. Evidence that Cache Valley virus induces congenital malformations in sheep. *Vet. Microbiol.* 21:297-307, 1990.

Comer, J.A., and Tesh, R.B. Phlebotomine sand flies as vectors of vesiculoviruses: A review. *Parasitologia* 33 (Suppl.):143-150, 1991.

Comer, J.A., Tesh, R.B., Modi, G.B., Corn, J.L. and Nettles, V.F. Vesicular stomatitis virus New Jersey serotype: Growth in and transmission by Lutzomyia shannoni (Diptera: Psychoidae). Am. J. Trop. Med. Hyg. 42:483-490, 1990.

Deblinger, R.D., Wilson, M.L., Rimmer, D.W., and Spielman, A. Reduced abundance of deer ticks, Ixodes dammini (Acari: Ixodidae) following incremental removal of deer density. J. Med. Entomol., 1992.

de Souza, M. and Freier, J.E. Vertical transmission of dengue-1 virus by Haemagogus equinus mosquitoes. J. Am. Mosq. Control Assoc. 7:118-120, 1991.

Durden, L.A., Logan, T.M., Wilson, M.L., and Linthicum, K.J. Experimental vector incompetence of a soft tick, Ornithodoros sonrai (Acari: Argasidae), for Crimean-Congo hemorrhagic fever virus. J. Med. Ent. 30:493-496, 1993.

Fan, W. and Mason, P.W. Membrane association and secretion of the Japanese encephalitis virus NS1 protein from cells expressing NS1 cDNA. Virology 177:470-476, 1990.

Fonseca, B.A.L., Khoshnood, K., Shope, R.E. and Mason, P.W. Flavivirus type-specific antigens produced from fusion of a portion of the E protein gene with the Escherichia coli TRP-E gene. Am. J. Trop. Med. Hyg. 44:500-508, 1991.

Gonzalez, J.P., Camicas, J.L., Cornet, J.P., Faye, O., and Wilson, M.L. Sexual and transovarial transmission of Crimean-Congo haemorrhagic fever in Hyalomma truncatum. Res. Virol. 143:23-28, 1992.

Gonzalez, J.P., Cornet, J.P., Wilson, M.L., and Camicas, J.L. Crimean-Congo hemorrhagic fever virus replication in adult Hyalomma truncatum and Amblyomma variegatum ticks. Res. Virol. 142:483-488, 1991.

Greiser-Wilke, I.M., Moennig, V., Kaaden, O.-R., and Shope, R.E. Detection of alphaviruses in a genus-specific antigen capture enzyme immunoassay using monoclonal antibodies. J. Clin. Microbiol. 29:131-137, 1991.

Howe, D.K., Vodkin, M.H., Novak, R.J., Shope, R.E., and McLaughlin, G.L. Use of the polymerase chain reaction for the sensitive detection of St. Louis encephalitis viral RNA. J. Virol. Methods 36:101-110, 1992.

Konishi, E., and Mason, P.W. Proper maturation of the Japanese encephalitis virus envelope glycoprotein requires co-synthesis with the premembrane protein. J. Virol. 67:1672-1675, 1993.

Konishi, E., Pincus, S., Fonseca, B.A.L., Shope, R.E., Paoletti, E., and Mason, P.W. Comparison of protective immunity elicited by recombinant vaccinia viruses that synthesize E. and NS1 of Japanese encephalitis virus. Virology 185:401-410, 1992.

Konishi, E., Pincus, S., Paoletti, E., Laegreid, W.W., Shope, R.E., and Mason, P.W. A highly attenuated host-range restricted vaccinia virus strain, NYVAC, encoding the prM, E, and NS1 genes of Japanese encephalitis virus prevents JEV viremia in swine. Virology 190:454-458, 1992.

Konishi, E., Pincus, S., Paoletti, E., Shope, R.E., Burrage, T., and Mason, P.W. Mice immunized with a sub-viral particle containing the JEV M and E proteins are protected from lethal JEV infection. *Virology* 188:714-720, 1992.

Lederberg, J., Shope, R.E., and Oaks, Jr., S.C. *Emerging Infections. Microbial Threats to Human Health in the United States*, National Academy Press, Washington, DC, 294 pp, 1992.

Mason, P.W., Pincus, S., Fournier, M.J., Mason, T.L., Shope, R.E., and Paoletti, E. Japanese encephalitis virus-vaccinia recombinants produce particulate forms of the structural membrane proteins and induce high levels of protection against lethal JEV infection, *Virology* 180:294-305, 1991.

Mason, P.W., Zugel, M.U., Semprone, A., Fournier, M.J. and Mason, T.L. The antigenic structure of dengue type 1 envelope and NS1 proteins expressed in Escherichia coli. *J. Gen. Virol.* 71:2099-2105, 1990.

Pelz, E.G. and Freier, J.W. Vertical transmission of St. Louis encephalitis virus to autogenously developed; eggs of Aedes atropalpus mosquitoes. *J. Am. Mosq. Control Assoc.* 6:658-661, 1990.

Pincus, S., Mason, P.W., Konishi, E., Fonseca, B.A.L., Shope, R.E., Rice, C.M., and Paoletti, E. Recombinant vaccinia virus producing the prM and E proteins of yellow fever virus protects mice from lethal yellow fever encephalitis. *Virology* 187:290-297, 1992.

Rico-Hesse, R. Molecular evolution and epidemiology of dengue viruses of serotypes 1 and 2. *Virology*, 174:479-493, 1990.

Salas, R., Manzione, N., Tesh, R.B., Rico-Hesse, R., Shope, R.E. et al. Venezuelan haemorrhagic fever, *Lancet* 338:1033-1036, 1991.

Shope, R.E. Antigen and antibody detection and update on the diagnosis of dengue. *Southeast Asian J. Trop. Med. Public Health* 21:642-645, 1990.

Shope, R.E. Infectious diseases and atmospheric change. In: *Global Atmospheric Change & Public Health*, White, J.D. (ed.), pp. 47-52, Elsevier Science Publishing Co., 1990.

Shope, R.E. Research and surveillance on yellow fever and dengue: Antivirals. In: *Simposio Internacional Sobre Febre Amarela e Dengue*, Homma, A. (ed.), pp. 405-406, Fiocruz Press, Rio de Janeiro, 1991.

Shope, R.E. Global climate change and infectious diseases. *Environ. Hlth. Persp.* 96:171-174, 1991.

Shope, R.E. Impact of global climate change on human health: Spread of infectious disease. In: *Global Climate Change: Implications, Challenges and Mitigation Measures*. S.K. Majumdar et al., eds. The Pennsylvania Academy of Science, pp. 363-370, 1992.

- Shope, R.E. and Evans, A.S. Assessing geographic and transport factors, and recognition of new viruses. In: *Emerging Viruses*, S.S. Morse (ed.), Chap. 11, pp. 109-119, Oxford, New York, 1993.
- Tesh, R.B. Undifferentiated arboviral fevers. In: *Tropical and Geographical Medicine*, second edition, K.S. Warren and A.A.F. Mahmoud (eds.), Chap. 73, pp. 685-691, McGraw-Hill, New York, 1990.
- Tesh, R.B. Arboviruses of Central Asia and the former Soviet Union. In: *Textbook of Pediatric Infectious Diseases*, Third Edition, R.D. Feignan and J.D. Cherry, eds. W.B. Saunders Co., Philadelphia, pp. 1462-1468, 1992.
- Tesh, R.B., and Andreadis, T.G. Infectivity and pathogenesis of iridescent virus type 22 in various insect hosts. *Arch. Virol.* 126:57-65, 1992.
- Tesh, R.B. and Guzman, H. Mortality and infertility in adult mosquitoes after the ingestion of blood containing ivermectin. *Am. J. Trop. Med. Hyg.* 43:229-233, 1990.
- Tesh, R.B., Lubroth, J., and Guzman, H. Simulation of arbovirus overwintering: Survival of Toscana virus (Bunyaviridae: Phlebovirus) in its natural sand fly vector, Phlebotomus perniciosus, *Am. J. Trop. Med. Hyg.* 47:574-581, 1992.
- Tesh, R.B., Wilson, M.L., Salas, R., de Manzione, N.M.C., Tovar, D., Ksiazek, T.G., and Peters, C.J. Field studies on the epidemiology of Venezuelan hemorrhagic fever. 1. Implication of the cotton rat Sigmodon alstoni as the probable rodent reservoir. *Am. J. Trop. Med. Hyg.* 49:227-235, 1993.
- Tignor, G.H., Casals, J. and Shope, R.E. The yellow fever epidemic in Ethiopia, 1961-1962: Retrospective serological evidence for concomitant Ebola or Ebola-like virus infection. *Trans. Roy. Soc. Trop. Med. Hyg.* 87:162, 1993.
- Vasconcelos, P.F.C., Travassos da Rosa, A.P.A., Rodriguez, S.G., Tesh, R., Travassos da Rosa, J.F.S., and Travassos da Rosa, E.S. Infecção humana adquirida in laboratorio causada pelo virus SPH 114202 (Arenavirus: familia Arenaviridae): Aspectos clinicos e laboratoriais. *Rev. Inst. Med. Trop. Sao Paulo* 35:521-525, 1993.
- Vodkin, M.H., McLaughlin, G.L., Day, J.F., Shope, R.E., and Novak, R.J. A rapid diagnostic assay for eastern equine encephalomyelitis viral RNA. *Am. J. Trop. Med. Hyg.* 49:772-776, 1993.
- Warrell, D.A. and Shope, R.E. Rabies. In: *Tropical and Geographical Medicine*, second edition, K.S. Warren and A.A.F. Mahmoud (eds.), Chap. 69, pp. 635-644, McGraw-Hill, New York, 1990.
- Weaver, S.C., Rico-Hesse, R., and Scott, T.W. Genetic diversity and slow rates of evolution in New World alphaviruses. *Curr. Topics Microbiol. Immunol.* 176:99-117, 1992.

Weaver, S.C., Scott, T.W., and Rico-Hesse, R. Molecular evolution of eastern equine encephalomyelitis virus in North America. *Virology* 182:774-784, 1991.

Weaver, S.C., Tesh, R.B., and Guzman, H. Ultrastructural aspects of replication of the New Jersey serotype of vesicular stomatitis virus in a suspected sand fly vector, Lutzomyia shannoni (Diptera: Psychodidae). *Am. J. Trop. Med. Hyg.* 46:201-210, 1992.

Wilson, M.L. and Deblinger, R.D. Vector management to reduce the risk of Lyme Disease, pp. 126-156. In: *Ecology and Environmental Management of Lyme Disease*. Ginsberg, H.S. (ed.) New Brunswick NJ: Rutgers University Press, 1993.

Wilson, M.L., Dykstra, E.A., and Schmidt, B.A. Temperature- and humidity-dependent longevity of unfed adult Hyalomma truncatum (Acari: Ixodidae). *J. Med. Ent.* 30:467-471, 1993.

Wilson, M.L., Gonzalez, J.P., Cornet, J.P., and Camicas, J.L. Transmission of Crimean-Congo haemorrhagic fever virus from experimentally infected sheep to Hyalomma truncatum ticks. *Res. Virol.* 142:395-404, 1991.

Zeller, H.G., Karabatsos, N., Calisher, C.H., Digoutte, J.P., Cropp, C.B., Murphy, F.A., and Shope, R.E. Electron microscopic and antigenic studies of uncharacterized viruses. II. Evidence suggesting the placement of viruses in the family Bunyaviridae. *Arch. Virol.* 108:211-217, 1989.