

SOUND Database of Marine Animal Vocalizations Structure and Operations

by

William A. Watkins, Kurt Fristrup, Mary Ann Daher, and Terrance Howald

August 1992

Technical Report

Funding was provided by the Office of Naval Research through the Ocean Acoustics Program (code 11250A) under Contract No. N00014-88-K-0273 and No. N00014-91-J-1445 with supplemental support by ORINCON/DARPA and NRL (code 211).

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> Woods Hole Oceanographic Institution Woods Hole, Massachusetts 02543

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ABSTRACT

The SOUND database system for marine animal vocalizations has been updated to include changes in the structure and operations that have evolved with use. These include more convenient operations, greater flexibility in analysis routines, and a revised database structure. The formats for data sorting and indexing, database structure, and analysis routines have developed into a convenient research tool. This report is a revision of the earlier operating manual for the SOUND databases (Watkins, Fristrup, and Daher 1991).

The interactive databases that comprise the SOUND system provide comprehensive means for quantitative analyses and statistical comparisons of marine animal vocalizations. These SOUND databases encompass (1) descriptive text databases cataloging the WHOI collection of underwater sound recordings of marine animals, (2) sets of files of digital sound sequences, (3) text databases organizing the digital sound cuts, and (4) software for analysis, display, playback, and export of selected sound files. The text databases index and sort the information about the sounds, and the digital sound cut files are accessed directly from the text record. From the text database, the sound cut data may be analyzed on screen, listened to, and compared or exported as desired.

The objective of this work has been the development of a basic set of tools for the study of marine animal sounds. The text databases for cataloging the recordings provide convenient sorting and selection of sounds of interest. Then, as specific sequences are digitized from these recordings, they become part of another database system that manages these acoustic data. Once a digital sound is part of the organized database, several tools are available for interactive spectrographic display, sound playback, statistical feature extraction, and export to other application programs.

KEY WORDS -- Sound database, Marine animals, Underwater sounds, Animal vocalizations.

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OBJECTIVES

This 1992 manual revises the description of the SOUND database system, and incorporates current formats for data sorting and indexing, for database structure, and for conducting data searches. Details of the operation of database associated programming are given for retrieval of selected digital sound files and for interactive display of text data along with waveform, FFT spectra, and RID spectral decomposition analyses. Other operations include concurrent playback of sound files and export of data for other uses.

The Marine Animal SOUND Database system (Watkins, Fristrup, and Daher 1991) provides convenient access and rapid analysis and display of acoustic data in the WHOI collection of recordings from marine species. The organization of the database structures follows that developed for convenient indexing and retrieval of data in our CETACEA database of literature references (Watkins, Daher, and Haley 1990). In addition, acoustic sequences from the various animal repertoires were digitized to provide a representative selection of sounds associated with the behaviors of the different animal species and sounds from a number of other non-biological sources. These digital sound files were organized and accessed directly from within the text databases.

The objective of this program has been the development of basic tools for the study of marine animal sounds, for accessing, analyzing, and comparing the acoustic patterns. This has included work on characterization of sound features for statistical comparison, and automatic recognition and diagnosis of marine animal vocalizations.

The SOUND databases have fulfilled these objectives of providing convenient means of relating the sounds to text data, sorting and retrieving them, allowing immediate analysis and playback of sequences of these sounds. This has opened the way for comprehensive, quantitative analyses and statistical comparisons of marine animal sounds.

ACKNOWLEDGEMENTS

The Marine Animal SOUND Database program is a joint effort by all who use these databases in their research, including William Schevill, Peter Tyack, Laela Sayigh, and This effort is shared also by previous Cheri Recchia. members of our bioacoustics program, including Nancy Haley, Karen Moore, and James Bird. The data collection at sea had expert contribution from all of those associated with the vessels and aircraft used for observations of marine animals over more than four decades. The organization and interpretation of the acoustic data encoded now by the SOUND databases are a result of the collective ideas and experiences of this entire group. The authors' contributions have emphasized basic structural organization (Watkins), innovative computer tools for augmenting the database functions (Fristrup), careful oversight of acoustic and annotation records (Daher), and analysis and software innovations (Howald). The central core of this program is the long standing collection of marine animal sound recordings -- tributes to the researchers that have made these important contributions for over 45 years.

Support for the Marine Animal SOUND Databases has been primarily from the Ocean Acoustics Program (code 11250A) of the Office of Naval Research with Contracts and Research Grants (most recent N00014-91-J-1445) and supplementary support through NRL (code 211, Stennis Center, MI) and DARPA/ORINCON (San Diego, CA). The program of bioacoustic studies that provided much of the previous work resulting in our acoustic recordings of marine life was supported for a considerable period by the Oceanic Biology Program of ONR. A wide range of other research programs and private support over this long period also have contributed to our work at sea and in the laboratory, providing understanding of the acoustic behavior of these marine species.

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INTRODUCTION

Database Organization --

This revision reflects changes to the SOUND database structure, organization, and associated analyses that have evolved since the original documentation (Watkins, Fristrup, and Daher 1991). These systems are in daily use, and the current arrangement is a result of its practical application for analysis and comparison of animal sounds.

The SOUND database system has a similar organization to that developed for the CETACEA databases described by Watkins, Bird, Moore, and Tyack (1988) with modified format revisions detailed by Watkins, Daher, and Haley (1990). Both SOUND and CETACEA databases use the same text systems for cataloging subjects, dates, geographic locations, and pertinent descriptive data. These notations are directly related to more than 150 species categories (Species List and List of Common Names for Marine Animals are appended). The acoustic data are referenced to text information about the species and the recording situation.

The data about the sound recordings cataloged in the SOUND databases are indexed and sorted by the database management program to provide convenient, rapid mechanisms for selecting sound recordings or digitized data of interest. The SOUND databases are presently installed on PC and AT compatible computers running Microsoft DOS, and the current organization for the system is based on INMAGIC 7.2 database software (INMAGIC INC., Cambridge, MA).

The text information for the SOUND databases are indexed in 31 fields (including date, location, recording data, sound class, species, number of animals, author, etc. -- see pages 19-22), and it allows independent sorting and retrieval of data in each field and unlimited subfields. The databases are searched by any combination of indexed or unindexed alphanumerical notations, by any combination of terms or partial phrases or stems of these notations. Searches may be complex and mixed with Boolean search statements. The data record for the desired sound file is selected by the text databases. The related digitized sound file may be immediately retrieved, analyzed, displayed, and played back, without leaving the text database. The databases continue to enlarge as the numbers of recordings increase, and as sound analyses have become more detailed. The databases (recordings, sound cuts, and literature) organize more than ten thousand records.

Marine Animal Recordings --

The library of sound recordings that provide the basis for the Marine Animal SOUND Databases were made during studies on acoustic behavior of marine species. This bioacoustic work was started in 1947 by William E. Schevill, and a continuous program of study of marine animal acoustic behaviors has been maintained at WHOI since that time. The initial set of tape recordings organized by the SOUND database included approximately 2000 magnetic tapes. These sounds on the tapes represented approximately 70 species of marine mammals, as well as fishes, crustaceans, and selected ambient noises.

The collection of marine animal sounds continues to enlarge as recordings are added from our own experiments at sea (two Arctic and four Caribbean cruises in 1989-1991), in addition to recordings by others. This library of recordings includes a variety of magnetic tapes, reel-to-reel with different widths and reel dimensions, VCR tapes, cassettes, phonograph records, computer digital media and optical disks. The combined SOUND databases now have more than 6000 entries.

The animal sounds in the recording collection and its database system incorporate a number of the historically important scientific contributions in bioacoustics, such as the first science recordings from marine mammals at sea by William Schevill (beluga, <u>Delphinapterus leucas</u>) and the early sound records of echolocating bats (<u>Eptesicus</u> and <u>Plecotus</u>) and oil birds (<u>Steatornis caripensis</u>) by Donald Griffin.

These original underwater bioacoustic studies were underway before tape recorders were readily available, so that recordings were made on a variety of different types of equipment. For example, the earliest records of the acoustic behavior of beluga were made on a "Gray Audograph" dictating machine, and signals were recorded on waxed paper disks. These sounds were then reproduced on transcription phonograph records. Copies of these recordings made on magnetic tape are retained in the collection.

The sound spectra recorded by the different recording media vary with each system that was used, from bandwidths of only a few kHz to 200 kHz or more. Recordings were made with single channel systems and with arrays of four or more hydrophones, using sensor depths varying from near surface to bottom mounted units. Recording equipment has varied considerably over the years, including standard audio systems, professional standards IRIG systems, and special purpose recorders. Most systems used "Direct" recording because of the wide bandwidths or high frequencies required to reproduce the natural animal sounds.

Marine mammals have been emphasized in the studies of animal acoustic behavior, therefore, much of the recorded data has focused on the sounds of cetaceans, pinnipeds, and sirenians. Recordings in the collection include underwater sounds from more than 70 species of marine mammals. Geographic locations for these recordings are scattered worldwide, with observations in many locations often taken over much of the year. Because of the long sequence of studies and the wide range of acoustic work represented by these continuing programs, the recording collection is representative for many areas of the world.

Prior to 1970, the attribution for sounds heard underwater was not always immediately certain, but with experience, more of the repertoire of the different species became recognized. As a result, there has been a change in the nature of the recordings over time. The use of multiple hydrophone arrays has increasingly allowed identification of vocalizations and sequences of sounds from individuals. Sources for the recorded sounds have been positively identified by these means. Many of the recordings were made especially to trace the usage or development of particular sequences of vocal activity.

The more recently recorded sounds deal less with species or population differences, and more with sounds that illustrate specific behaviors. Later recordings, therefore, are more focused on specific scientific objectives (vocal identity, mimicry, shared signalling, ontogeny of learned signals), and they relate to certain species (sperm whales, finbacks, bowheads, bottlenose dolphins, etc.). For example, our 1954-1968 sperm whale recordings led to specific studies of their activities, which in turn opened the way to work focused on coda signals produced by these whales, and then to analyses of distinctions between codas from individuals and those shared by the members a particular population.

INTERRELATED DATABASES

The textual data on marine animals and their sounds are organized by a series of interrelated computer databases currently using PC and AT workstations. These separate, related databases for organization of marine animal vocal (1) CETACEA, a comprehensive database of behavior include: references to the literature on marine animal sounds, (2) SOUND, a database for the descriptive information about each of the sound recordings, (3) SOUNDC, a database for the detailed information and acoustic descriptors for each of the separate digital sound cuts, (4) SOUND2 (SOUND2C, and other series), auxiliary databases for specific research emphases using these same data protocols, and (5) a set of matching digital sound cut files (filename matching the text database record). The digitized sound cuts are stored as separate files on optical disk, and they can be accessed, analyzed (time series and a variety of spectrographic portrayals), displayed, and played back from within the text (SOUNDC) databases.

Structures for all of the databases follow similar patterns, use the same systems of annotation, and relate to the same lists of species, geographic codes, etc. Separate, indexed fields within each of the database records describe the recording situation, the equipment used, the geographic locations and recording dates, species (and individuals) that were present, behavioral notes, sound types, etc. Records with any of these data or any alphanumeric combination in any field may be rapidly searched and selected. Then, the matching digital sound file can be retrieved for analysis and display or accessed for other purposes. Auxiliary databases provide separate organization for particular research projects on animal sounds (dolphin mother-calf signature whistles, beluga vocal behavior, sperm whale coda patterns, etc.).

A display-edit-export function for the sound files accesses the digital files from within the text databases. This is a pop-up, TSR utility for retrieving the stored digital sound file, and analyzing and displaying waveforms and color spectrograms (a variety of FFT and RID analyses) from within the text database. Cursors provide measurement of signal time and frequency, and select portions for expansion or export.

A header with identifying information is prefixed to each digital sound file. The header duplicates a portion of the ASCII text from the related (SOUNDC) text database record. This text information is also related to the original database record of the library collection of recordings, SOUND, thereby allowing the sorting and retrieval of tape information from subsidiary databases, SOUNDC, SOUND2C, etc. The identified digital sound files are accessed from within these text databases, retrieved directly from storage, such as from optical disks.

Equipment --

Sounds from the marine animal recording collection are reproduced on a variety of reel-to-reel, VCR, and cassette equipment. As possible, frequency responses and dynamic ranges at least as good as for the original recordings are maintained, so as to provide equivalent spectra to those of the original recording systems.

Sounds are analyzed with a variety of equipment, including Kay DSP Sonagraph 5500 (Kay Elemetrics, Pinebrook, NJ), WHOI VOICE analyzer (Martin, Catipovic, Fristrup, and Tyack 1990), RSIG and CSIG routines from within the SOUND databases (Fristrup, Daher, Howald, and Watkins 1992), and other special purpose analyses adapted to specific requirements. Digital conversion for sound cuts is performed at more than twice the highest frequency of the sequences of interest. PC and AT work-stations (optimized 286, 386, and 486 computers) are used for database operations.

The digital sound cut files are stored (archival) on optical disks (Maxtor OC-800), and large capacity hard disk drives are used for temporary deposit of sound files during database manipulations.

CETACEA --

The comprehensive, background database of references to the literature, CETACEA, emphasizes information on marine animal sounds. The data in about 4500 records (to date) are indexed and sorted by 300 subjects, 150 species categories, and a variety of other indexed notations

including dates, locations, sound spectral characteristics, environmental observations, etc. A unique feature of these database structures is the direct connection between species and all other indices, including subjects, locations, observation dates, notes, etc. In addition, codes have been adapted for ease in identifying and linking data in various fields. Searching the database is rapid, using a wide variety of simple and complex Boolean strategies. The CETACEA database and its operations have been described in two report? (Watkins, Bird, Moore, and Tyack 1988; and Watkins, Daher, and Haley 1990).

The CETACEA database and the related SOUND databases use an adaptation of INMAGIC software (INMAGIC Inc., Cambridge, MA) for the text data indexing, sorting, and searching routines. For each database, the programs operate with three interactive files: structure, index, and data records. They all share the direct linkage of species with other indices.

SOUND --

The databases for the marine animal collections of analog acoustic recordings organize the descriptive information about each of the sound records and sequences of sound. The SOUND databases (including auxiliary databases such as SOUND2, etc.) index and sort the data for more than 6000 recordings to date. These databases continue to grow as the collections increase in size and complexity. Separate databases are established for particular research needs (for example, one database is currently used for separate organization of the collection of VCR and cassette recordings of dolphins, and other related databases are for specific data on digitized cuts of signature sounds, etc.). Unique retrieval numbers for each record also provide the date and sequence of recording (for example, 83093 indicates the 93rd recording cataloged for the year 1983), and these retrieval codes are used in indexing and shelving of the library collections. Data about the recordings are sorted by 150 species, 30 indexed subjects that are also directly connected to species, in addition to a variety of other information, including recording locations, dates and times, identities of individuals, number of animals, sound levels, and sound types.

SOUNDC --

The digital cuts of marine animal sounds are organized by specific text databases, such as SOUNDC (sound cut). These text databases for the sound cuts have similar structure and design to the other SOUND databases, and they are used in the same ways to index and sort the data appropriate to the individual sound sequence on the digital file. The sound cut databases relate specifically to the sequences of sounds taken from the different recordings in the library collection. Unique retrieval numbers for each digital sound cut file identify the original recording as well as the sequence of digital cuts made from that original (for example, 83093034 identifies the 93rd recording catalogued for 1983 listed as tape number 83093, and this cut is sound cut number 34 from that recording). Added details indexed in the sound cut databases (such as SOUNDC) include particulars about that sound sequence, including reference to library tape cue, channel numbers analyzed, duration and sampling rate for the digital file, sound comparison characters, etc. These data are sorted by 150 species, 30 indexed subjects that are directly connected to species, such as recording location, dates, number of animals, sound types, etc.

A specific digital sound cut file selected by searching the SOUNDC databases is retrieved as desired. Then, using the pop-up display utility, RSIG, without leaving the database, the sound cut may be analyzed, displayed on screen, and played back by loudspeaker. At a keystroke, the text data for the cut is back on screen or another type of analysis is underway to provide further detail about the sound cut.

Digital Sound Files --

Digital sound cuts are stored as independent files, identified by filenames that match text database record numbers. The first 512 bytes of each file include the indexing information as a header for that sound cut file. The related text SOUNDC database provides the flexible search, selection, and retrieval capabilities for accessing these files from within the database. Sound sequences are selected and digitized as separate sound cut files for a variety of purposes, including that of illustration of a particular type of sound, detailing the repertoire of individual species, analyzing sounds related to certain

behaviors, comparing sequences of signals from different species, separating distinctive calls from different populations, or demonstrating variation with season and activity, etc.

ANALYSIS AND DISPLAY

The digital sound cut files identified by searching the SOUNDC database may be selected for later analysis, or as noted above, they may be immediately analyzed, displayed, and played back without leaving the SOUNDC text database. Immediate access to the digital acoustic data is provided by a pop-up utility, RSIG. RSIG retrieves the digital file identified by the current search results from SOUNDC, and the digital file is analyzed with both waveform and spectrogram displayed simultaneously on screen, with concurrent audio playback. The analyzed signals may be edited, re-analyzed, and expanded as desired. Or, the text search results may be re-examined and other signals rapidly retrieved for analysis and display. These digital sound files may also be reformatted and exported by RSIG as needed.

Alternatively, a stand alone analysis program, CSIG (with functions similar to RSIG), may be used directly, without interaction with the database, to analyze, display, and playback the digital sound files.

All of the SOUND databases, file handling routines, and signal analysis and display programs are related. They utilize common identifiers, data structures, field labels, search and sorting strategies, and display parameters. In addition to the primary databases and their analysis systems, other auxiliary databases are also maintained, such as SOUND2 and SOUND3C which organize data for different species and particular bioacoustic projects and analyses.

INMAGIC PROGRAM

The SOUND databases are currently used with an adaptation of INMAGIC database software (version 7.2, INMAGIC Inc., Cambridge, MA) for text databases. This is a

flexible text database system that has proven to be relatively simple and easily searched, while retaining the needed complexity of association within indexed data. A unique feature of the data association for these databases is the direct link between species and other indexed subjects. The combination of the coding system and the database software allows every component or alphanumeric entry in the record to be available for searching by a wide variety of simple and complex Boolean strategies.

The INMAGIC text database system includes the following features: (1) permits records of any length, (2) relates 75 or more fields, (3) provides for unlimited numbers of defined subfields, (4) indexes and sorts fields and subfields, (5) indexes as each data record is entered, (6) allows independent sorting and retrieval of data in subfields as well as fields, (7) supports search strategies developed with Boolean operators (and, or, not) and nested arguments, (8) uses searches with qualifiers (greater than, less than, equal to, from/to), (9) provides for convenient right-hand truncation in search statements, (10) saves and combines search results, (11) allows use with user-defined formats for display or re-ordering of data, (12) prints any number of selected records in any of these formats, (13) lists any indexed terms or fields and subfields with their frequency of occurrence, (14) permits the use of extended characters in records, (15) provides for development of flexible on-line thesaurus of terms, search operators, and definitions for help in searching the records, (16) permits rapid copying of data records, and (17) allows importation of ASCII records created elsewhere. Records in the CETACEA database, for example, are indexed by more than 300 subjects, 150 species categories, and a variety of other notations including dates, locations, sound features, environmental observations, etc. With approximately 5000 records in the CETACEA database, searching and retrieval of data records or combinations of records are rapid, usually less than a second.

The databases may be searched by any combination of indexed or unindexed alphanumeric notations. Detailed searches may be made using specific indexed fields, such as genus/species (searchable by order/suborder and family as well). Searching may use any combination of terms and text words or even stems of words or partial phrases and parts of any alphanumeric entry. In addition, codes have been adapted for ease in identifying and searching species, subjects, geographic areas, etc. The alphanumeric coding of marine animal species allows indexing, sorting, and

retrieval of most subject fields, geographic locations, dates, and events in direct relationship to specific species. Detailed searches may be made using genus/species, record number, identification, age, gender, observation date, geographic locations (including area names and latitude and longitude), author of recording, sound type, etc.

Detailed descriptions of search operations and potential combinations of search statements appropriate to these databases are reviewed in more detail in the previous SOUND database report (Watkins, Fristrup, and Daher 1991) and in the CETACEA database reports (Watkins, Bird, Moore, and Tyack 1988, Watkins, Daher, and Haley 1990). See the INMAGIC Manual (INMAGIC INC., Cambridge, MA 02140-1338).

Search features include simple and complex BOOLEAN relations (such as, equals, less/greater than, less than or equals, starts with, contains stem, from...to, etc.). Codes assigned to species and subject categories provide direct associations of most indexed fields. This allows generalized searches or increasingly specific searches by truncation of the codes (refer to Organization to Species List, p. 41).

The design of the database takes advantage of INMAGIC's system for right-hand truncation, so that the placement of codes at the end of fields allows searches by codes or elements of codes, as well as by the other record data. Although a bit slower, searches are also possible using any alphanumeric combination contained in any record, whether in indexed or unindexed fields.

ORGANIZATION OF DATABASE RECORDS

Database records are organized for convenience in entering the data, for relative ease of utilization of the information in records, and for reorganization of the data for display.

The organizational detail is indicated by the list of field names in the database structure and descriptions of these fields (next sections). In brief, the records are organized as follows:

-- Each record is given a unique retrieval number (RECNO). This number gives the year of recording, the recording series, and the sequence of digital analysis. In addition, it serves as a pointer for identification of the recording in the library collection.

-- Separate fields give the recording cue for specific sounds, number and sequence of channels, equipment, author, sampling rate, duration of the sound sequence, etc.

-- Separate fields also are used for entering a variety of data about the animals and the particular vocal sequence, such as activity, animal identity, age, interaction, sound class and type.

-- Genus/species names and alphanumeric codes are entered for all species recorded -- order/suborder, family, genus, and species are indicated by each code.

-- Codes for genus/species are appended to data in many other fields for direct association with location, observation date and time, sound type, etc.

-- Location for the recording is given by name, by geographic code, as well as by latitude and longitude.

-- Notes, and annotations may be included, and are related to species.

-- Data on the recording situation are also entered, such as hydrophone depths, recording conditions, received signal levels, and sound type.

Most fields in the SOUND database records are indexed and may be searched separately or in combination to provide rapid selection of these records. Notes and similar fields are not indexed, but these too may be searched (more slowly) for any words, phrases, or alphanumeric notation.

The List of Species is provided at the end of this report to assist in identification of interrelated codes and to aid in database searches. In addition, a List of Common Names for Marine Animals provides reference to some of the more frequently used vernacular names for these species.

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STRUCTURE FOR SOUND DATABASES Revised July 1992

INMAGIC Program Format

(Index: T = term, Y = both term and key-word, N = no index) See INMAGIC Manuel for Sort and Emphasis codes.

LABEL	NAME	INDEX	SORT	EMPHASIS
RN	RECNO	т	3	1
CU	CUE	Т	3	1
NC	NOCHAN	т	3	1
SR	SAMRATE	т	1	1
CS	CUTSIZE	ТЗ	1	1
PL	PLAYBA	ХУ	7	1
SC	SIGCLAS	S Y	5	1
ID	IDENT	Y	7	1
AG	AGE	Y	7	1
IA	INTERAC		7	1
GS	GENSP	Y	5	1
GA	GEOA	Y	5	1
OD	OBSDATE		4	1
NT	NOTE	N		
DA	DATE	Т	4	1
IP	IDPRES	Y	7	1
AP	AGEPRES		7	1
BH	BEHAV	Y	5	1
OS	OTHERSE	-	5	1
NA	NOANIM	Т	3	1
GB	GEOB	Y	5	1
GC	GEOC	Y	7	1
OT	OBSTIME	E Y	7	1
SH	SHIP	Y	5	1
AU	AUTHOR	Y	5	1
LO	LOCATE	Y	5	1
HY	HYDEPTH	IТ	3	1
RC	RCOND	Y	5	1
RG	RGEAR	Y	5	1
SL	SIGLEVI		7	1
ST	SIGTYPE	E Y	5	1

Abbreviated List of Fields --

SOUND Databases (July 1992)

Retrieval number of record year/tape#/cut#) RN CU Cue or time on tape, buffer (B) size, sec in buffer NC Number channels recorded, digitized, chan/side ID SR Sample rate (convert Kay input freq. to sample rate) Cut size -- digital cut in sec (2 or 3 dec. places) CS PL Playback equip., filter setting low (L) high (H) kHz SC Signal class - <u>S</u>ignature, <u>M</u>imic..., qual 1-5, <u>O</u>verlap ID ID of individual vocalizing, species code AG Age, sex (M/F) prefix, birth year, ID, species code IA Interaction (MC = male-calf, etc.), ID Genus/Species animals producing sounds, species code GS GA Geographic location A = ASFIS code, species code OD Observation date of original recording, species code NT Note = Species code, observation or recording details DA Date of this record entry (latest modification) IP Identification conspecifics present, species code AP Age consp. present: sex-pref., birth yr., ID, sp. code BH Behavior of the animals, species code OS Other species present, species codes NA Number of animals vocalizing, species code GB Geographic location B = name of area, species code --/2 Location of birth/capture area, species code, (ID) Geographic location C = lat. & long., species code GC OT Observation time of original recording, species code SH Ship/cruise, aquarium, or recording platform AU Author, originator of the recording LO Location of original recording HY Hydrophone depth in m RC Recording conditions, weather, salinity, etc. RG Recording gear, equipment SL Signal level (dB received or source), species code ST Signal type codes, species code

--/2 Signal type names (coda, slow clicks), species code

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SOUND Database Field Description -- (July 1992)

- LABEL -- NAME -- Description with example.
- RN RECNO Retrieval number of record (Year/tape#/cut#) Year (2 digits), tape no. (3 digits), cut # (3 digits, alphanumeric base 36). Example: 850430B6
- CU CUE Cue or time (min:sec) on tape at signal end, (B) analyzer buffer size (min:sec), decimal time sec from start of Kay buffer to cursor at beginning of sound. Example: 542 B2:8 8.130 or 1:03:12 B2:8 8.130
- NC NOCHAN Number of channels recorded (1st digit), number of channels multiplexed (2nd digit), channel ID letter(s) -- side one A & B, side two C & D. Example: 42CD
- SR SAMRATE Sample rate (Kay input frequency converted).
- CS CUTSIZE Cut size -- sec (2 or 3 decimal places).
- PL PLAYBAK Playback recorder/filter type, settings in kHz (pass-band), L = low (HP) and H = high (for LP). Example: PEMTEK/KROHN-HITE L0.1 H12
- SC SIGCLAS Signal class, letter codes (Signature, Mimic, <u>Variant, Deletion, Uncharacteristic, Calf,</u> etc.), quality 1 to 5 (best), <u>0</u> = overlap in <u>Time</u> or <u>Frequency</u>. <u>N</u> = no overlap. Example: S 4 OTF
- ID IDENT Identification of vocal animal, species code.
- AG AGE Age with sex (M or F) prefix (decimal for part year) and ID, birth year of vocalizing animal with sex prefix and ID, species code. Example: M0.3FB19 M1985FB19 BD19D
- IA INTERAC Interaction, male-male (MM)/ female-calf (FC), etc., with ID's. Example: FCFB10 FCFB19

Fields in SOUND Structure (continued)

- GS GENSP Genus and species of vocalizing animal. Scientific names and species code.
- GA GEOA Geographic location area code from ASFIS map, species code. Example: ANWABIA (ANW area, ABIA species)
- OD OBSDATE Observation date for original, species code. Example: 20-Dec-1960 AB1A
 - OBSDATE/2 Subfield for month and year, species code. Example: DecAB1A 1960AB1A
- NT NOTE Species code, comments, recording details. ID of animals in parentheses. Not indexed.
- DA DATE Date of this record entry, last modification. (Use F6 key) Example: 24-May-1991
- IP IDPRES ID of conspecifics present, species code. Separate subfields for each animal.
- AP AGEPRES Age of conspecifics present = sex prefix (M/F) with age (decimal for part year) and ID, birth year with sex prefix and ID, species code. Separate subfields each animal. Example: M0.3FB19 M1985FB19 BD19D
- BH BEHAV Behavior codes, species code.
- OS OTHERSP Other species present, species codes. Separate subfields for each species.
- NA NOANIM Number of animals vocalizing, species codes.
- GB GEOB Geographic location area name, species code.
 - GEOB/2 Location of birth/capture, species code, ID of animal (in parentheses).
- GC GEOC Geographic location latitude and longitude, "N" or "S" and two digits for lat., "E" or "W" and three digits for long. Each with species code. Example: N70AB1A E020AB1A

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Fields in SOUND Structure (continued)

- OT OBSTIME Observation time for original, species code. Example: 1430 AB1A
- SH SHIP Ship/cruise, aquarium, or other platform, (<u>SM</u>all boat, <u>AQ</u>uarium, <u>BE</u>ach, <u>IC</u>e, <u>UN</u>known)
- AU AUTHOR Author, originator of the recording.
- LO LOCATE Location of original recording.
- HY HYDEPTH Hydrophone depth in m.
- RC RECCOND Conditions, weather, salinity. Not indexed.
- RG RECGEAR Recording gear, equipment. Not indexed.
- SL SIGLEVL Signal level <u>R</u>eceived, <u>S</u>ource, (dB), species code. <u>E</u> = estimated. Example: S172AB1A E
- ST SIGTYPE Signal type, Long= > 0.1 sec; Short= < 0.1 sec BL Broadband long, noisy, many frequencies BS Broadband short, clicks, pulses Narrowband long, tonal, few harmonics Narrowband short, bell, short tones NL NS FM Frequency modulated, sweeps, "contours" CH Chirp, short FM sweep Pulsed, click bursts, sidebands PU SE Series, train, sequence of sounds Song, long, repetitive, patterned SO -- with species code. Example: NLABIA
 - SIGTYPE/2 Signal type, general: coda, slow clicks, etc. -- with species code.

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GEOGRAPHIC LOCATIONS

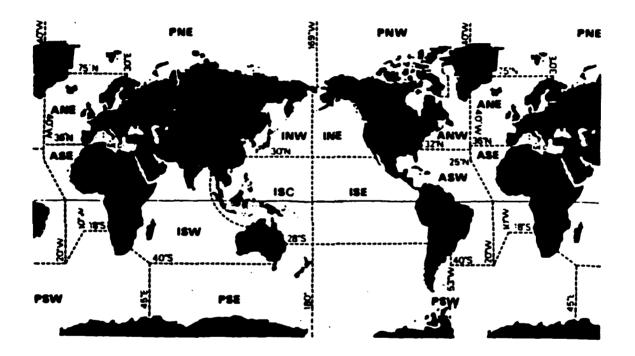
GEOA field -- The first of the geographic location fields in the structure listed above uses codes for the locations or places at which the original recordings were made. These location codes are adapted generally from the geographic codes used by the Aquatic Sciences and Fisheries Information System (Anon. 1980, ASFIS):

ANE	Northeast Atlantic
ANW	Northwest Atlantic
ASE	Southeast Atlantic
ASW	Southwest Atlantic
INE	Northeast Pacific
INW	Northwest Pacific
ISE	Southeast Pacific
ISC *	Southwest (central) Pacific
ISW	Indian Ocean
MED	Mediterranean
PNE	Eastern Arctic Ocean
PNW	Western Arctic Ocean
PSE	Eastern Antarctic Ocean
PSW	Western Antarctic Ocean
CSL *	Coastal Waters
FSR *	Freshwater
COS *	Cosmopolitan

* changes from the ASFIS codes

The species codes are combined with the ocean area codes. For example, a recording of <u>Megaptera novaeangliae</u> in the Indian Ocean is indexed with the codes ISWAC2A (ISW for the Indian Ocean, and AC2A for <u>Megaptera novaeangliae</u>).

ASFIS Map -- Aqua. Sci. Fish. Info. Sys. (Anon. 1980, ASFIS).



TEXT DATABASES

The use of text-based systems has allowed rapid association of our different existing data sets dealing with marine animals and their sounds. This includes the CETACEA database for literature references and the variety of observational information accompanying animal sightings, as well as the library of oceanic recordings of animal sounds and the SOUND cut databases. These text databases perform admirably for organizing, cataloging, and sorting such discrete pieces and sequences of information.

The text databases with INMAGIC software have provided descriptive detail for organization of the material, and we anticipate that numeric representations of acoustic features, for example, will be transitioned to other, more interactive, relational systems. For extensive comparisons of the sound data from large components of the acoustic files, work with other database and analysis systems is planned. Even so, the fundamental relationships between textual records about the sound recordings and the behavioral descriptors relative to the acoustic data will necessarily continue to be needed, regardless of the database systems that are employed. The text-based data will always be significant, and the means for intelligent organization of these data will continue to be important.

USING THE SOUND DATABASES

The following instructions for using the text program for the SOUND database provide a quick, step-by-step guide to its use, and they show the basic simplicity of database operations. The current database organization is adapted for use with INMAGIC 7.2 software (Anon. 1990).

1. Call up the directory containing the INMAGIC software and enter the database program by typing <u>INMAGIC</u>.

2. Choose <u>SELECT</u> from the Main Menu for searching the database, for printing or displaying entries. -- or choose <u>MAINTAIN</u> from the Main Menu for work within a database to edit old records or to add new ones.

3. Indicate the database that is desired, such as SOUND, SOUNDC, SOUND2, SOUND3C, etc.

4. Passwords may be indicated as required.

5. And then, searches and retrieval of records may be initiated. -- or records may be called up for modification and new records entered into the database.

Any number of indexed fields may be sorted and retrieved rapidly (much less than a second), and even unindexed fields, such descriptive notes, can be easily searched for any alphanumerical string. The time required for searching unindexed fields varies with the amount of stored data.

6. Records that are selected may be displayed, sorted, printed, or exported as desired (sequentially through the list of selections). Digital sound files for each text record may also be accessed from within the search display, then analyzed by waveform and spectrogram as well as played back (digital to analog conversion) on loudspeakers.

7. It is important to <u>EXIT</u> properly after maintenance work on the databases, because of potential disruption of indexing operations that could scramble the data.

These operations are similar to those described in somewhat more detail for the earlier arrangements of the SOUND databases (Watkins, Fristrup, and Daher 1991) and for

the literature reference CETACEA database (Watkins, Daher, and Haley 1990; Watkins, Bird, Moore, and Tyack 1988). See also the INMAGIC Manual (Anon. 1990) for detailed descriptions of the software functions and operations.

SEARCHING -- TEXT DATABASES

The SOUND database systems using INMAGIC software provide for organization and rapid handling of the textual data associated with the library of acoustic recordings of underwater sounds from animals. Indexed database fields are rapidly searched by specifying field labels or names by simple and by a variety of complex search strategies. Records (in any sequence) may be displayed, printed, or exported in a wide variety of formats defined by the user. Unlimited subfields may be indexed and searched in the same ways to provide extremely flexible and detailed sorting and retrieval of these text records.

Unindexed fields may be similarly searched for any word, phrase, or alphanumeric string, but at a slower rate.

Search commands may be combined with search relations to provide complex Boolean command sequences (including, excluding, broadening), word and word-stem searches, comparative searches (equals, starts with, greater/less than or equals, from...to), etc. Searchable, sorted data can be listed in a wide variety of ranges and formats.

The flexibility of the database software search system is combined with a system of coding assigned to the data. Codes are appended to each species, behaviors, sound types, etc. Therefore, the codes may also be used as a rapid search strategy. In addition, the species codes are used to make sure that different subjects, dates, places, etc. are always directly linked with species. Species codes have been assigned to each species by (1) order/suborder, (2) family, (3) genus, and (4) species. For example, a finback whale (<u>Balaenoptera physalus</u>) has the code "AC1F" -- indicating suborder Mysticeti, family Balaenopteridae, genus <u>Balaenoptera</u>, species <u>physalus</u>. See section on Organization of Species List (p. 41).

See the INMAGIC Manual (Anon. 1990) for additional search strategies.

DIGITAL SOUND FILES

Sequences of sound (cuts) from the repertoire of vocalizations of the different species of marine animals are selected from the original underwater recordings and digitized. These digital sound cuts (often 1 to 30 sec) are stored as separate files, and they are organized, indexed, and sorted by related text databases (SOUNDC, SOUND2C, etc.). The first 512 bytes of each digital sound cut file is a binary/ASCII header, including data from the SOUNDC database record for this sound sequence. This annotation includes such detail as the tape cue for the sound sample in the original recording, sampling rate, length of cut, observation date and place, species, associated behavior, etc. -- usually all of the database fields through "Note" (see sections on Data Structure on p. 18 and on Database Fields pp. 19-22). The digital sound cut files are labeled with an extension of the tape number from the original library recording to maintain continuity of the data (including year of recording, tape number, and number of this sound cut from this tape), as noted below for the sound cut numbering convention.

Example: <u>8821120F</u>

88211		complete library recording number.
88	The	year of the original recording.
211	The	recording number for that year.
20F	The	digital sound cut number.
8821120F	The	complete digital sound cut number.

All three places of the sound cut extension number use a 36-base character set -- employing numbers 0-9 and letters A-Z, in this order (20A follows 209, 5F0 follows 5EZ).

A different character is used for the first place of the sound cut extension (last three characters) to designate different series, channels, animals, sound types, etc.

The sound cut extension number <u>000</u> is not used (88211 and 88211000 are considered to be the same by the database program).

Digitizing the Sound Cuts --

A variety of systems may be used to provide the detailed scrutiny of the analog library recordings, to select sound sequences of interest, and to convert the sounds to digital format. Two signal analyzers used most often for creation of the files for the SOUND database include VOICE (Martin, Catipovic, Fristrup, and Tyack 1990) and a KAY DSP Sonagraph 5500 (Kay Elemetrics, Pine Brook, The analog library recordings are played back on NJ). appropriate equipment, and the sound is monitored aurally by good quality loudspeakers and visually by the real-time waveform and spectrographic displays of the signal To provide an indication of the methodology for analyzers. selection and digitizing of sound cuts, procedures and operations are listed using a PC computer, tape playback, and a KAY (DSP):

1. Computer -- call up the INMAGIC program and select the sound recording of interest. (See previous section on Searching the SOUND database.)

- a. Identify the recording number for digitizing.
 - $(Ex: \underline{G}et \underline{RN} = \underline{65005}).$
- b. Write (copy) this record into an ASCII file using an annotation format, such as ANN (Ex: write using ANN in 65005.asc).
- Computer -- call up the KAY (K) File Management Program.
 a. From the Main Menu select "System".
 - b. Choose "Data Path" and indicate Drive and Path for downloading of digitized signal files.
 - c. Choose "Interface" and select "SCSI".
 - d. Choose "I/O" and select "From 5500".
 - (To open "Data Transfer Menu"):
 - 1. "File Format" = 5500.
 - 2. "Number of Bits" = 8, 10, 12 or 16.
 - 3. "Channel" = 1 or 2.
 - 4. "Source" = Buffer, Between Cursors, Screen, or Highlighted List.
 - 5. "Name of File" = <u>Sound cut number</u>.KAY
 - Digital filename (Ex: 65005001.KAY).

Leave cursor blinking until download.

3. Playback -- Mount and play selected recording. Locate sound sequence for digitization.

- 4. KAY DSP-- set the analyzer for real time analysis.
 - a. Copy (digitize) sound onto KAY buffer and scroll through (waveform and spectrographic displays).
 - b. Locate signal within parameters to be downloaded (Buffer, Between Cursors, Screen).
- 5. Computer -- still in the <u>KAY</u> File Management Program: a. Press [Ret] to accept the KAY file selected
 - (named above with .KAY extension).
 - b. Press [F2] to transfer digital signal from the KAY to the designated Drive and Path.
 - c. Exit KAY program if no more is to be transferred <u>or</u> remain in KAY program for more signals.
- 6. Computer -- call up SideKick Plus (TSR editor utility sharing KAY program on screen).
 - a. Access the ASCII text file created in #1 above.
 - b. Rename this record, giving it the same number as the digital sound cut (in #2), TXT/ASC extension.
 - c. Update the text information appropriate to sound cut (digital file just downloaded #5).
 - d. Save the changes made to this file (press [F2]) and exit SideKick Plus program.
- 7. Computer -- call up the MASSEDIT program
 - (to attach header information to digital files). a. Indicate the ASCII file (made in #6) containing the
 - updated text information for the sound cut files. b. Designate the Drive/Directory for the digital files (Ex: MASSEDIT 65005.TXT D:).
- 8. Computer -- call up the INMAGIC database program (to index sound cut records in SOUNDC database).
 - a. Choose MAINTAIN from the Main Menu.
 - b. Select appropriate database (SOUNDC, SOUND2C, etc.)
 - c. <u>A</u>DD the ASCII text file modified (#6) in SideKick (Ex: <u>A</u>dd <u>65005.TXT</u>) to index the new edited data records into (sound cut) SOUNDC text database.
- 9. Computer -- from within the INMAGIC database program:
 - a. Search the SOUNDC (sound cut) text database for the sound sequence of interest (see Searching -- Text Databases, p. 29).
 - b. Display the text data for the sound sequence.
 - c. Using RSIG, analyze this sound on screen (a key stroke), waveform and spectrographic analyses.
 - d. Return to text data for this sound (a key stroke).
 - e. Listen to this sound, D-to-A conversion and playback of the sound (a key stroke).

PROGRAMS FOR THE SOUND DATABASES

A variety of programs have been designed or adapted for work with the SOUND databases and their associated digital sound cut files. The basic set of programs for these marine animal sound databases was described in detail by Fristrup, Daher, Howald, and Watkins 1992. Some of these are listed and discussed briefly below.

INMAGIC	 Text database index and search.
SIDEKICK PLUS	 Text ASCII data editor and transfer.
HEADEDIT	 Edit annotation of one sound cut file.
MASSEDIT	 Annotate one or more sound cut files.
KAYCHECK	 Check data in file headers.
CSIG	 Analyze and playback sound cut file without database involvement.
RSIG	 Analyze and playback sound cut file from within text database.

- INMAGIC is the current database software for organizing, indexing, sorting, and retrieving records containing information about the library collection of sound recordings, and the digital sound cuts. Program adapted from software by INMAGIC INC., Cambridge, MA (Anon. 1990).
- SIDEKICK PLUS is a pop-up utility (TSR) program used to access and modify ASCII text (database output), for example, from the SOUND database to provide appropriate text data for the sound cut files to be indexed in the SOUNDC databases. Software by Borland International, Scotts Valley, CA.

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- HEADEDIT program allows display and editing of ASCII text header on individual digital sound files (usage: HEADEDIT FILENAME.KAY). The binary data written by the download program for the KAY DSP 5500 Sonagraph (including number of samples, sampling rate, and number of bits per sample) are displayed also but are not available for editing. HEADEDIT may be used to read the header data, change existing header text, or enter new data. New text data for the header may be entered as blocks of new data by means of a utility program such as SKPLUS, or text may be typed in place directly with HEADEDIT. Program by Kurt Fristrup.
- MASSEDIT program is used to insert ASCII text into headers on one or a number of files at one operation (HEADEDIT works with one file at a time). The text data are inserted into the header space, the first 512 bytes of each digital sound file. This header space and some identifying binary code is provided by the KAY download program of the Kay Sonagraph (DSP 5500). The header data typically include the first fields of the sound cut text record (SOUNDC database) through at least the NOTE field, allowing direct connection of the information about the recording with the appropriate digital sound file. MASSEDIT matches the record retrieval numbers and ASCII data sets of a specified file (having .TXT or .ASC extension) to the appropriate files (with .KAY extension) in a specified directory (Usage: MASSEDIT FILENAME.TXT D:). See example in #7, p. 32. Program by Kurt Fristrup.
- KAYCHECK is a comparison program that searches for every .KAY file (format for KAY download program) in the current directory, and checks the header text data (first 512 bytes on the digital file) for accuracy of the text information on filename, sample rate, file size, etc. Files that are found to have discrepancies or are without a proper header are identified, with date, time, and size of file indicated. The output of KAYCHECK may be written to another file (usage: KAYCHECK>FILENAME) for printing or editing. Program by Kurt Fristrup.

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SOUND Database

- CSIG program provides access, analysis, and display of files of digitized sound (independent of databases). CSIG accesses the specified sound file, analyzes, and displays waveform (top of screen) and concurrent spectrographic analyses (bottom of screen). Analyses are FFT-based systems or point-by-point techniques using RID (W. Williams, Univ. Michigan). Frequency and time scales and variable cursors are provided. Portions of the displayed analysis may be selected and re-analyzed. All or any part of the data file may be exported. In addition, the digital file may be converted to an analog signal and played back repeatedly on a loudspeaker (a keystroke). See section below on Operating Commands, p. 36. Program by Kurt Fristrup and Terrance Howald.
- RSIG is a TSR (terminal stay resident) program used for access, analysis, display, and playback of the digital sound file from within the SOUNDC database. Just as for CSIG, above, spectrographic analyses are FFT-based systems or point-by-point techniques using RID (W. Williams, Univ. Michigan). Each sound cut may be immediately analyzed while the text data record for that cut is displayed on screen by the database. RSIG is loaded as a TSR utility before database selections are made, then file access and analyses are initiated (a key stroke) from within the database. At anytime, the screen display may be returned immediately to the text data record, re-analyzed, or any segment of the display expanded for further analysis. Any sequence or number of files selected by the database may be accessed, analyzed, and displayed in turn. All or any part of the data file may be exported. In addition, the digital file may be converted to an analog signal and played back repeatedly by loudspeaker. See section below on Operating Commands, p. 38. Program by Kurt Fristrup and Terrance Howald.

SOUND Database

Operating Commands for CSIG and RSIG --

CSIG operates independently from the SOUND database Usage: CSIG filename; [ESC] exits program.

RSIG is used during database operations and is first loaded as a TSR utility into the sound file directory.

From the text database display: [Alt] [Esc] Invokes RSIG which accesses the selected digital sound file, draws signal waveform and spectrogram, and sound playback. [Esc] Ends this analysis and re-displays text.

Key Functions for CSIG and RSIG [Alt] [D] Re-draws analysis screen with defaults. [Up/Down Move (horizontal) frequency cursor up/down. Arrows] [Left/Right] Move (vertical) time cursor left/right. Arrows] [Ctrl] [Left/ Right Arrows] 5 X speedup time cursor movement. [Home] Toggle left/right (vertical) time cursor. [PqDn] Zoom in to re-display data between cursors. [PqUp] Return to previous display bounds. [F7] Move backward partial (1/3) screen. [F8] Move forward partial (1/3) screen. Move backward full screen. [F9] Move forward full screen. [F10] [Ctrl] [PrtSc] Write data between cursors to a file. [Alt] [X] Save cursor information to SIGCURSE.DAT. [Alt] [P] Playback data on screen, between cursors. Increase playback rate by two. [ALT] [F1] Decrease playback rate by half. [Alt] [F2] [Alt] Toggle full-screen waveform display. [F] Toggle noise compensation. [Alt] [N] Increase dynamic range by 3 dB. [Ins] [**Del**] Decrease dynamic range by 3 dB. [Ctrl] [Home] Increase signal level (attenuator) 3 dB. [Ctrl] [End] Decrease signal level (attenuator) 3 dB. [Alt] [F5] Increase FFT size two times for spectrogram. Decrease FFT size by half for spectrogram. [Alt] [F6] [Alt] [F8] RID 1st order spectral decomposition, raise to 9th, 17th order (additional keystrokes). [Alt] [F9] RID lower order spectral decomposition.

SOUND Database

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Organization of the Species List --

The Species List for the SOUND Databases have undergone only minor modification since the previous description of the system (Watkins, Fristrup, and Daher 1991). This is the same species list as that developed for the CETACEA literature database (Watkins, Bird, Moore, and Tyack 1988; Watkins, Daher, and Haley 1990). Species are listed alphabetically within families. Alphanumeric codes have been assigned by (1) order/suborder, (2) family, (3) genus, and (4) species, in ascending alphanumeric order.

The first place of the species code is a letter representing the order or suborder. The letter "A" denotes the suborder Mysticeti and includes 11 species. The letter "B" denotes the suborder Odontoceti and includes 68 species. The letter "C" denotes the order Carnivora, including 37 species of pinnipeds, sea otters, and polar bear. The letter "D" denotes Sirenia and includes 5 species. Other orders and suborders are included in general categories and are coded by letter (E-Z).

The second place of the species code is a letter representing the family. For example, in the Species List the Balaenopteridae are indicated as code "AC" -order/suborder Mysticeti "A" and family Balaenopteridae "C". The code for the Ziphiidae is "bc" -- order/suborder Odontoceti "B", family Ziphiidae "C".

The third place of the species code is a number containing one or two digits representing the genus. For example, the code for the genus <u>Mesoplodon</u> is "BC5" -order/suborder Odontoceti "B", family Ziphiidae "C", genus <u>Mesoplodon</u> "5". The code for the genus <u>Kogia</u> is "BA1"-order/suborder Odontoceti "B", family Physeteridae "A", genus <u>Kogia</u> "1". The code for the genus <u>Stenella</u> is "BD15" -- order/suborder Odontoceti "B", family Delphinidae "D", and genus <u>Stenella</u> "15").

The last place of the species code is a letter representing species. For example, the code for <u>Kogia</u> <u>breviceps</u> is "BA1A" (suborder Odontoceti "B", family Physeteridae "A", genus <u>Kogia</u> "1", species <u>breviceps</u> "A"). The code for the related species, <u>Kogia simus</u> is "BA1B". The code for <u>Cephalorhynchus hectori</u> is "BD1D" (suborder Odontoceti "B", family Delphinidae "D", genus <u>Cephalorhynchus</u> "1", species <u>hectori</u> "D"). Both the scientific names and the genus/species codes are given in the GENSP field -- for example, a finback whale will have "<u>Balaenoptera physalus</u> AC1F" in the genus/species field.

Mysticeti A	
Balaenidae AA	
<u>Balaena mysticetus</u> Linnaeus 1758	AA1A
<u>Eubalaena</u> <u>glacialis</u> (Borowski) 1781	AA3A
<u>Eubalaena australis</u> (Desmoulins) 1822	AA3B
Eschrichtiidae AB	
Eschrichtius robustus (Lilljeborg) 1861	AB1A
Balaenopteridae AC	
	AC1A
<u>Balaenoptera</u> <u>bonaerensis</u> Burmeister 1867	
<u>Balaenoptera</u> <u>borealis</u> Lesson 1828	AC1B
Balaenoptera edeni Anderson 1878	AC1C
	AC1E
	AC1F AC2A
<u>Megaptera novaeangliae</u> (Borowski) 1781 Neobalaenidae AD	AC2A
<u>Caperea</u> <u>marginata</u> (Gray) 1846	AD1A
<u>Caperea</u> <u>marginata</u> (Gray) 1840	ADIA
Odontoceti B	
Physeteridae BA	
Kogia breviceps (Blainville) 1838	BA1A
<u>Kogia simus</u> (Owen) 1866	BA1B
Physeter catodon Linnaeus 1758	BA2A
Monodontidae BB	
<u>Delphinapterus leucas</u> (Pallas) 1776	BB1A
Monodon monoceros Linnaeus 1758	BB2A
Ziphiidae BC	
<u>Berardius</u> <u>arnuxii</u> Duvernoy 1851	BC1A
<u>Berardius</u> <u>bairdii</u> (Stejneger) 1883	BC1B
<u>Hyperoodon ampullatus</u> (Forster) 1770	BC2A
<u>Hyperoodon planifrons</u> Flower 1882	BC2B
Indopacetus pacificus (Longman) 1926	BC3D
Mesoplodon bidens (Sowerby) 1804	BC5A
Mesoplodon bowdoini Andrews 1908	BC5B BC5C
<u>Mesoplodon carlhubbsi</u> Moore 1963 Mesoplodon denginastria (Plainwille) 1817	BC5D
	BC5D BC5E
<u>Mesoplodon europaeus</u> Gervais 1855 <u>Mesoplodon ginkgodens</u> Nishiwaki and Kamiya 1958	
<u>Mesoplodon gravi</u> von Haast 1876	BC5J
<u>Mesoplodon hectori</u> (Gray) 1871	BC5K
<u>Mesoplodon layardii</u> (Gray) 1871 <u>Mesoplodon layardii</u> (Gray) 1865	BC5L
<u>Mesoplodon mirus</u> True 1913	BC5M
<u>Mesoplodon peruvianus</u> Reyes, Mead, Waerebeek 1991	
<u>Mesoplodon stejnegeri</u> True 1885	BC5S
<u>Tasmacetus shepherdi</u> Oliver 1937	BC7A
Ziphius cavirostris G. Cuvier 1823	BC9A
strang villouter 0. Surici 1925	

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SPECIES LIST SOUND Databases	
Delphinidae BD <u>Cephalorhynchus</u> <u>commersonii</u> Lacépède 1804 <u>Cephalorhynchus</u> <u>eutropia</u> (Gray) 1846(9?)	BD1A BD1B
<u>Cephalorhynchus heavisidii</u> (Gray) 1828	BD1C
<u>Cephalorhynchus hectori</u> van Beneden 1881	BD1D
<u>Delphinus bairdii</u> Dall 1873	BD3A
<u>Delphinus delphis</u> Linnaeus 1758	BD3B
<u>Delphinus tropicalis</u> van Bree 1971	BD3C
<u>Grampus griseus</u> (Cuvier) 1812	BD4A
<u>Lagenodelphis</u> <u>hosei</u> Fraser 1957	BD5A
<u>Lagenorhynchus acutus</u> (Gray) 1828	BD6A
Lagenorhynchus albirostris Gray 1846	BD6B
Lagenorhynchus australis (Peale) 1848	BD6C
Lagenorhynchus cruciger (Quoy and Gaimard) 1824	
Lagenorhynchus obliquidens Gill 1865	BD6G
Lagenorhynchus obscurus (Gray) 1828 Liggadalabhig baraalig (Daala) 1848	BD6H
<u>Lissodelphis borealis</u> (Peale) 1848 Lissodelphis porenii (Lesápède) 1804	BD8A
<u>Lissodelphis peronii</u> (Lacépède) 1804 <u>Peponocephala</u> <u>electra</u> (Gray) 1846	BD8B BD10A
<u>Sotalia borneensis</u> Lydekker 1901	BD10A BD12A
<u>Sotalia brasiliensis</u> Van Beneden 1875	BD12R BD12C
<u>Sotalia fluviatilis</u> (Gervais) 1855	BD12C BD12B
<u>Sotalia</u> <u>guianensis</u> Van Beneden 1864	BD12D BD12D
<u>Sousa chinensis</u> (Osbeck) 1765	BD12D
<u>Sousa plumbea</u> (Cuvier) 1829	BD13B
<u>Sousa tëuszii</u> (Kukënthal) 1892	BD13C
<u>Stenella</u> <u>attenuata</u> (Gray) 1846	BD15A
<u>Stenella</u> <u>clymene</u> Gray 1850	BD15B
Stenella coeruleoalba (Meyen) 1833	BD15C
Stenella frontalis (G. Cuvier) 1829	BD15F
<u>Stenella longirostris</u> (Gray) 1828	BD15L
<u>Steno bredanensis</u> (Cuvier) 1828	BD17A
Tursiops aduncus (Ehrenberg) 1832	BD19A
<u>Tursiops catalania</u> (Gray) 1868	BD19B
<u>Tursiops gillii</u> Dall 1873	BD19C
<u>Tursiops</u> <u>truncatus</u> (Montagu) 1821	BD19D
Globicephalidae - BE	
<u>Feresa</u> <u>attenuata</u> Gray 1874	BE1A
<u>Globicephala edwardii</u> Smith 1934	BE3A
<u>Globicephala macrorhynchus</u> (Gray) 1846	BE3B
<u>Globicephala melaena</u> (Traill) 1809	BE3C
<u>Globicephala scammoni</u> Cope 1869	BE3D
<u>Orcaella brevirostris</u> (Owen 1866)	BE5A
<u>Orcaella</u> <u>fluminalis</u> Anderson 1871	BE5B
<u>Orcinus orca</u> (Linnaeus) 1758	BE7A
<u>Pseudorca crassidens</u> (Owen) 1846 Phocoenidae BF	BE9A
	DF1 X
<u>Australophocaena dioptrica</u> Lahille 1912 <u>Phocoena</u> phocoena (Linnaeus) 1758	BF1A BF2A
<u>Phocoena spinipinnis</u> Burmeister 1865	BF2B
THACACHE STUTATINITS DATWEISCEL 1002	DI 2D

SPECIES LIST SOUND Databases	
Phocoenidae (continued)	
<u>Phocoena</u> <u>sinus</u> Norris and McFarland 1958	BF2C
<u>Phocoenoides dalli</u> (True) 1885	BF4A
<u>Neophocaena phocaenoides</u> (G.Cuvier) 1829	BF6A
Susuidae BG	5643
Susu gangetica Lebeck 1801	BG1A
<u>Susu indii</u> Blyth 1859 Inio mooffmangia Blainwille 1017	BG1B
<u>Inia geoffrensis</u> Blainville 1817 Linetas usuillifan Millan 1919	BG2A
<u>Lipotes vexillifer</u> Miller 1918 Deptembrie blainwillei (Commune) 1044	BG3A
<u>Pontoporia blainvillei</u> (Gervais) 1844	BG4A
Carnivora C	
Otariidae CA	
<u>Arctocephalus</u> <u>australis</u> (Zimmerman) 1783	CA1A
<u>Arctocephalus</u> <u>forsteri</u> Lesson 1828	CAIF
<u>Arctocephalus</u> <u>galapagoensis</u> Heller 1904	CAIG
<u>Arctocephalus</u> <u>gazella</u> Peters 1875	CAIH
<u>Arctocephalus philippii</u> Peters 1866	CAIP
<u>Arctocephalus</u> <u>pusillus</u> (Schreber) 1776	CAIR
<u>Arctocephalus</u> townsendi Merriam 1897	CAIT
Arctocephalus tropicalis (Gray) 1872	CAIW
<u>Callorhinus</u> <u>ursinus</u> (Linnaeus) 1758	CA1W CA2A
<u>Eumetopias jubatus</u> (Schreber) 1776	CA2A CA3B
<u>Neophoca</u> <u>cinerea</u> (Peron) 1816	CA3B CA4A
<u>Otaria</u> <u>flavescens</u> (Shaw) 1800	CA4A CA6A
<u>Phocarctos hookeri</u> (Gray) 1844	CA8A
Zalophus californianus (Lesson) 1828	CA8A CA9A
Odobenidae CB	CAJA
<u>Odobenus</u> rosmarus (Linnaeus) 1758	CB1A
Phocidae CC	CDIA
<u>Cystophora</u> <u>cristata</u> (Erxleben) 1777	CC1A
<u>Erignathus</u> <u>barbatus</u> (Erxleben) 1777	CC2A
Halichoerus grypus (Fabricius) 1791	CC3A
<u>Hydrurga leptonyx</u> (Blainville) 1820	CC4A
	CC5A
<u>Lobodon</u> <u>carcinophagus</u> (Hombron & Jacquinot) 1842	
<u>Monachus</u> <u>monachus</u> (Hermann) 1779	CC8A
<u>Monachus</u> <u>schauinslandi</u> Matschie 1905	CC8B
Monachus tropicalis Gray 1850	CC8C
<u>Mirounga</u> <u>angustirostris</u> Gill 1866	CC10A
<u>Mirounga leonina</u> (Linnaeus) 1758	CC10A CC10B
<u>Phoca caspica</u> Gmelin 1788	CC10B CC12C
<u>Phoca</u> fasciata Zimmermann 1783	CC12C CC12F
<u>Phoca</u> <u>groenlandica</u> Erxleben 1777	CC12F CC12G
<u>Phoca hispida</u> Schreber 1775	CC12G CC12H
<u>Phoca largha</u> Pallas 1811	CC12H CC12L
<u>Phoca sibirica</u> Gmelin 1788	CC12L CC12S
<u>Phoca vitulina</u> Linnaeus 1758	CC12S CC12V
<u>Ommatophoca</u> <u>rossi</u> Gray 1844	CC12V CC14A
Mustelidae CD	CCT4N
Enhydra lutris (Linnaeus) 1758	CD1A
<u>Lutra felina</u> Molina 1782	CD1R CD2B
<u>BRUEN</u> <u>EVELIN</u> INTING 1/VE	

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Ursidae CE	-
<u>Ursus</u> <u>maritimus</u> Phipps 1774 CE1	A
Sirenia D	
Dugongidae DA	
Dugong dugon Muller 1776 DA1	Α
<u>Hydrodamalis</u> gigas Zimmermann 1780 DA2	В
Trichechidae DB	
<u>Trichechus</u> <u>inunquis</u> (Natterer) 1883 DB1	А
<u>Trichechus</u> manatus Linnaeus 1758 DB1	
Trichechus senegalensis Link 1795 DB1	
OTHER MAMMALS	
Primates E ChiropteraF	
Ungulates, <u>sensu lato</u> G Elephantidae G	
<u>Elephas maximus</u> Linnaeus 1758 GAL	А
Loxodonta africana (Blumenbach) 1797 GA2	
Other mammals H	
VERTEBRATES	
Aves I	
Reptilia J	
Reptilia J Amphibia K	
Fish, <u>sensu lato</u> L	
Other vertebrates S	
INVERTEBRATES Molluscs	
Cephalopoda M	
Other molluscs N	
Arthropods	
Crustacea 0	
Insecta P	
Other arthropods Q	
Other invertebrates R	
CENEDAL	
GENERAL Fossils T	
Uncertain (sea serpents and	
other indeterminate animals) U	
General pinniped V	
General cetacean W	
Ambient noise (ship, geologic, ice) X	

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COMMON NAMES FOR MARINE ANIMALS

List of Common Names for Marine Animals --

The "common" names given to different species of marine animals vary considerably, with only a few of these names consistently used by English speakers. Other language areas, of course, have their own (variable) set of common For accuracy, therefore, the databases always use names. the scientific nomenclature. However, for cross-reference, we have included the following list of some of the most commonly used names for these animals. The common names are listed in the same order, directly with the scientific names and the species codes used by the databases. Many species are not well known by any but their scientific names -- it is always acceptable, and often much easier, to refer to these marine animals by their scientific names. The scientific nomenclature is recognized worldwide, regardless of language.

COMMON NAMES FOR MARINE ANIMALS

Baleen whales (Mysticeti) A	
Family Balaenidae AA	
Bowhead whale <u>Balaena</u> <u>mysticetus</u>	AA1A
Northern right whale <u>Eubalaena</u> <u>glacialis</u>	AA3A
Southern right whale <u>Eubalaena</u> <u>australis</u>	AA3B
Family Eschrichtiidae AB	
Gray whale <u>Eschrichtius</u> robustus	AB1A
Family Balaenopteridae AC	
Minke whale <u>Balaenoptera</u> <u>acutorostrata</u>	AC1A
Southern minke whale <u>Balaenoptera</u> bonaerensis	AC1D
Sei whale <u>Balaenoptera</u> <u>borealis</u>	AC1B
Bryde's whale <u>Balaenoptera</u> <u>edeni</u>	AC1C
Blue whale <u>Balaenoptera musculus</u>	AC1E
Fin, Finback whale <u>Balaenoptera</u> physalus	AC1F
Humpback whale <u>Megaptera</u> <u>novaeangliae</u>	AC2A
Family Neobalaenidae AD	
Pygmy right whale <u>Caperea marginata</u>	AD1A
Toothed whales (Odontoceti) B	
Family Physeteridae BA	
Pygmy sperm whale <u>Kogia breviceps</u>	BA1A
Dwarf sperm whale <u>Kogia simus</u>	BA1B
Sperm whale <u>Physeter</u> <u>catodon</u>	BA2A
Family Monodontidae BB	
Beluga, white whale <u>Delphinapterus</u> <u>leucas</u>	BB1A
Narwhal <u>Monodon monoceros</u>	BB2A
Family Ziphiidae BC	
Arnoux bottlenose whale <u>Berardius</u> <u>arnuxii</u>	BC1A
Baird's bottlenose <u>Berardius</u> <u>bairdii</u>	BC1B
Northern bottlenose <u>Hyperoodon</u> <u>ampullatus</u>	BC2A
Southern bottlenose <u>Hyperoodon</u> <u>planifrons</u>	BC2B
<u>Indopacetus</u> <u>pacificus</u>	BC3D
Sowerby's beaked whale Mesoplodon bidens	BC5A
<u>Mesoplodon</u> bowdoini	BC5B
Hubb's beaked whale <u>Mesoplodon</u> <u>carlhubbsi</u>	BC5C
Blainville's beaked <u>Mesoplodon</u> <u>densirostris</u>	BC5D
<u>Mesoplodon</u> <u>europaeus</u>	BC5E
<u>Mesoplodon</u> <u>ginkgodens</u>	BC5H
<u>Mesoplodon gravi</u>	BC5J
Hector's beaked whale <u>Mesoplodon</u> <u>hectori</u>	BC5K
<u>Mesoplodon layardii</u>	BC5L
True's beaked whale <u>Mesoplodon mirus</u>	BC5M
<u>Mesoplodon</u> peruvianus	BC5P
<u>Mesoplodon</u> <u>stejnegeri</u>	BC5S
Tasman beaked whale <u>Tasmacetus</u> <u>shepherdi</u>	BC7A
Cuvier's, goose-beaked <u>Ziphius</u> <u>cavirostris</u>	BC9A

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COMMON NAMES FOR MARINE ANIMALS Family Delphinidae (dolphins) ----- BD BD1A Commerson's dolphin <u>Cephalorhynchus</u> <u>commersonii</u> BD1B -- Cephalorhynchus eutropia -- Cephalorhynchus heavisidii BD1C Hector's dolphin -- Cephalorhynchus hectori BD1D Pacific common dolphin -- Delphinus bairdii BD3A Saddleback, common dolphin -- Delphinus delphis BD3B -- <u>Delphinus</u> tropicalis BD3C Grampus, Risseau's dolphin -- Grampus griseus BD4A Fraser's dolphin -- Lagenodelphis hosei BD5A White-sided dolphin -- Lagenorhynchus acutus BD6A White-beaked dolphin Lagenorhynchus albirostris BD6B Peale's dolphin -- Lagenorhynchus australis BD6C -- Lagenorhynchus cruciger BD6E BD6G Pacific white-sided Lagenorhynchus obliguidens Dusky dolphin -- Lagenorhynchus obscurus BD6H North. rt. whale dolphin - Lissodelphis borealis BD8A South. rt. whale dolphin -- Lissodelphis peronii BD8B -- Peponocephala electra BD10A Borneo dolphin -- Sotalia borneensis BD12A Amazon dolphin -- Sotalia brasiliensis BD12C -- <u>Sotalia</u> <u>fluviatilis</u> BD12B Guiana dolphin -- Sotalia guianensis BD12D Chinese river dolphin -- Sousa chinensis BD13A -- <u>Sousa plumbea</u> BD13B -- <u>Sousa tëuszii</u> BD13C Spotted dolphin -- Stenella attenuata BD15A -- <u>Stenella</u> <u>clymene</u> BD15B BD15C Striped dolphin -- Stenella coeruleoalba -- Stenella frontalis BD15F Spinner dolphin -- Stenella longirostris BD15L Rough-toothed dolphin -- Steno bredanensis BD17A Southern bottlenose dolphin -- Tursiops aduncus BD19A -- Tursiops catalania **BD19B** Pacific bottlenose dolphin -- Tursiops gillii BD19C Bottlenose dolphin, Atlantic -- Tursiops truncatus BD19D Family Globicephalidae (small toothed whales) -- BE BE1A -- Feresa attenuata Southern pilot whale -- Globicephala edwardii **BE3A** Short-finned pilot -- Globicephala macrorhynchus BE3B Long-finned pilot -- Globicephala melaena BE3C Pacific pilot whale -- Globicephala scammoni BE3D Irrawaddy dolphin -- Orcaella brevirostris BE5A -- Orcaella fluminalis BE5B Killer whale -- Orcinus orca BE7A False killer -- Pseudorca crassidens BE9A Family Phocoenidae (porpoises) -- BF -- Australophocaena dioptrica BF1A Harbor porpoise -- Phocoena phocoena BF2A Burmeister's porpoise -- Phocoena spinipinnis BF2B -- Phocoena sinus BF2C

COMMON NAMES FOR MARINE ANIMALS	
Family Phocoenidae (porpoises continued)	
Dall's porpoise <u>Phocoenoides</u> <u>dalli</u>	BF4A
Finless porpoise <u>Neophocaena</u> <u>phocaenoides</u>	BF6A
Family Susuidae (dolphins River & South) BG	DI UN
Ganges river dolphin <u>Susu gangetica</u>	BG1A
Indus river dolphin <u>Susu indii</u>	BG1B
Boutu (Amazon river) <u>Inia</u> <u>geoffrensis</u>	BG2A
Yangtze river dolphin Lipotes vexillifer	BG3A
Franciscana <u>Pontoporia blainvillei</u>	BG4A
Seals, Sea otters, etc. (Carnivora) C	
Family Otariidae (fur seals, sea lions) CA	
Southern fur seal <u>Arctocephalus</u> <u>australis</u>	CA1A
N. Z. fur seal <u>Arctocephalus</u> <u>forsteri</u>	CA1F
Galapagos fur <u>Arctocephalus</u> <u>galapagoensis</u>	CA1G
Kerguelen fur <u>Arctocephalus</u> <u>gazella</u>	CA1H
Juan Fernandez fur <u>Arctocephalus philippii</u>	CA1P
South African fur <u>Arctocephalus</u> <u>pusillus</u>	CA1R
<u>Arctocephalus</u> <u>townsendi</u>	CA1T
<u>Arctocephalus</u> <u>tropicalis</u>	CA1W
Northern fur seal <u>Callorhinus</u> <u>ursinus</u>	CA2A
Steller sea lion <u>Eumetopias jubatus</u>	CA3B
Australian sea lion <u>Neophoca</u> <u>cinerea</u>	CA4A
Southern sea lion <u>Otaria flavescens</u>	CA6A
N. Z. sea lion <u>Phocarctos</u> <u>hookeri</u>	CA8A
California sea lion <u>Zalophus</u> <u>californianus</u>	CA9A
Family Odobenidae (walrus) CB	
Walrus <u>Odobenus</u> Family Phocidae (seals) CC	CB1A
Hood seal <u>Cystophora</u> <u>cristata</u>	CC1A
Bearded seal <u>Erignathus barbatus</u>	CC2A
Gray seal <u>Halichoerus</u> grypus	CC3A
Leopard seal <u>Hydrurga leptonyx</u>	CC4A
Weddell seal <u>Leptonychotes</u> weddellii	CC5A
Crabeater seal <u>Lobodon</u> <u>carcinophagus</u>	CC6A CC8A
Med. monk seal <u>Monachus</u> Manachus	
Hawaiian monk seal <u>Monachus schauinslandi</u>	CC8B
Caribbean monk seal <u>Monachus tropicalis</u>	CC8C
North elephant seal <u>Mirounga</u> <u>angustirostris</u>	CC10A
South elephant seal <u>Mirounga</u> <u>leonina</u>	CC10B
Caspian seal <u>Phoca</u> <u>caspica</u>	CC12C
Ribbon seal <u>Phoca</u> <u>fasciata</u>	CC12F CC12G
Harp seal <u>Phoca</u> <u>groenlandica</u>	CC12G CC12H
Ringed seal <u>Phoca hispida</u>	CC12H CC12L
Spotted seal <u>Phoca largha</u> Baikal seal Phoca sibirica	CC12L CC12S
Harbor seal <u>Phoca</u> <u>Sibirica</u> Harbor seal <u>Phoca</u> <u>vitulina</u>	CC12S CC12V
Ross seal <u>Ommatophoca</u> <u>rossi</u>	CC12V CC14A
Family Mustelidae (sea otter) CD	CCT44
Sea otter Enhydra lutris	CD1A
	CD1R CD2B
<u>Lutra felina</u>	CD2D

COMMON NAMES FOR MARINE ANIMALS

Family Ursidae (polar bear) CE Polar bear <u>Ursus maritimus</u> Manatees (Sirenia) D	CE1A
Family Dugongidae (dugong) DA Dugong <u>Dugong dugon</u> <u>Hydrodamalis</u> gigas	DA1A DA2B
Family Trichechidae (manatee) DB Amazonian manatee <u>Trichechus</u> <u>inunguis</u> West Indies manatee <u>Trichechus</u> <u>manatus</u> West African manatee <u>Trichechus</u> <u>senegalensis</u>	DB1A DB1B DB1C
OTHER MAMMALS Primates E Chiroptera F Ungulates, <u>sensu lato</u> G Family Elephantidae (Elephants) GA Indian elephant <u>Elephas maximus</u> African elephant <u>Loxodonta africana</u> Other mammals H	GA1A GA2A
VERTEBRATES Aves I Reptilia J Amphibia K Fish, <u>sensu lato</u> L Other vertebrates S	
INVERTEBRATES Molluscs Cephalopoda M Other molluscs N Arthropods Crustacea O Insecta P Other arthropods Q Other invertebrates R	
GENERAL Fossils T Uncertain (sea serpents and other indeterminate animals) U General pinniped V General cetacean W Ambient noise (ship, geologic, ice) X General mammal Y Animals in general Z	

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The text databases index and sort the information about the sounds, and the digital sound cut files are accessed directly from the text record. From the text database, the sound cut data may be analyzed on screen, listened to, and compared or exported as desired. The objective of this work has been the development of a basic set of tools for the study of marine animal sound. The text databases for cataloging the recordings provide convenient sorting and selection of sounds of interest. Then, as specific sequences are digitized from these recordings, they become part of another database system that manages these acoustic data. Once a digital sound is part of the organized database, several tools are available for interactive specrographic display, sound playback, statistical

feature extraction, and export to other application programs.

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underwater sounds		
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