

ELECTE UG 19 1991

AD

TECHNICAL REPORT 9111



A New Method for Determining Residues Generated During Munition Demilitarization Activities: Analysis of Environmental Soil and Water Samples

> M. A. Major R.T. Checkai C.T. Phillips R.O. Nwanguma R.S. Wentsel

> > April 1991

U S ARMY BIOMEDICAL RESEARCH & DEVELOPMENT LABORATORY

Fort Detrick

Frederick, MD 21701-5010

Approved for public release; distribution unlimited



U S ARMY MEDICAL RESEARCH & DEVELOPMENT COMMAND Fort Detrick Frederick, MD 21701-5012

91 8 19 011

NOTICE

<u>Disclaimer</u>

The findings in this report are not to be construed as an official Department of the Army position unless so designated by other authorized documents.

Disposition

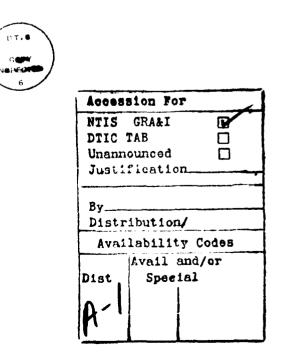
Destroy this report when it is no longer needed. Do not return it to the originator.

ia. NAME OF PERFORMING ORGANIZATION S. Army Biomedical Research (if applicable)		7a. NAME OF MONITORING ORGANIZATION						
		SGRD-UBG-E						
c. ADDRESS (City, State, and ZIP Code)			7b. ADDRESS (City	y, State, and ZIP C	ode)			
ort Detr		_						
rederick	, MD 2170)2-5010						
a. NAME OF FUNDING/SPONSORING ORGANIZATIONU.S. Army Corps (If applicable)			9. PROCUREMENT	INSTRUMENT ID	NTIFIC	ATION NU	MBER	
f Engine			CERD-M					
	City, State, and			10. SOURCE OF F		-		
	husetts Av			PROGRAM ELEMENT NO.	PROJECT	TASK NO.		WORK UNIT
ashingto	n, DC 201	514-1000		62720A	3E162720A83		00	026
New Met nalysis	of Environ	etermining Res	sidues Generated and Water Sample	l During Munit es	tion Demilit	ariz	ation A	ctivities:
2. PERSONAL	r, R.T. C	heckai, C.T. H	Phillips, R.O. M	Wanguma, R.S	. Wentsel			
3a. TYPE OF REPORT Cechnical 13b. TIME COVERED FROM 10/90 TO 12/91			14. DATE OF REPORT 1991 April	RT (Year, Month, 18	Day)	15. PAGE	COUNT	
6. SUPPLEM	ENTARY NOTA	TION						
7.	COSATI	CODES	18. SUBJECT TERMS	Continue on reverse	e if necessary and	identi:	fy by block	number)
FIELD	GROUP	SUB-GROUP	munitions		(00 (00)			
17	08		open burning/o	open detonati	on (OB/OD)	- \		
			polycyclic are		arbons (PAH	5)		
9. ABSTRACT	Continue on	reverse if necessary	and identify by block n	umber)			on of m	unitions
cocedures are presented for qualitative screening and subsequent quantitation of munitions								
ind related compounds present at sites contaminated by open burning/open detonation (OB/OD) lisposal of munitions. Methods were developed to recover munitions and polycyclic aromatic								
hydrocarbons (PAHs) from soil via sonic extraction into acetonitrile, and munitions from								
vater via trapping onto Cl8 disposable cartridges and eluting with methanol. Complementary								
IPLC procedures were used for identification and quantification of these compounds. A								
qualitative HPLC gradient method was developed, and used to screen samples for a wide range								
of munitions and PAHs. Quantitation of munitions and their environmental reduction products								
vas accomplished using an HPLC isocratic method. Recoveries of munitions and degradation								
products spiked into sandy loam soil were greater than 90% in all cases. Corresponding								
recoverie	recoveries from aqueous samples were comparable for the nitroaromatics, but were poorer for							
the nitra	the nitramines. Critieria of detection for individual muntions ranged from 0.067 mg L^{-1} to).37 mg L^{-1} .							

	21. ABSTRACT SECURITY CLASSIFICATION
UNCLASSIFIED/UNLIMITED SAME AS RPT.	0110011100
28. NAME OF RESPONSIBLE INDIVIDUAL	22b TELEPHONE (Include Area Code) 22c. OFFICE SYMBOL
Michael A. Major. Ph.D.	(301) 663–2014 SGRD–UBG–E

TABLE OF CONTENTS

Introduction1
Materials and Methods1
Discussion
Results
References10
Distribution List12



.

LIST OF TABLES

		Page
Table	I	3
Table	II	4
Table	III	9
	LIST OF FIGURES	

Page

Figure	I	1
Figure	11	3

INTRODUCTION

Out-of-date and out-of-specification munitions have commonly been disposed of by burning and detonation on unprotected ground (1). This practice, open burning/open detection (OB/OD), generates a mixture of contaminants into the immediate area at high concentration (2). The mixture consists primarily of unburned munitions but environmental weathering and microbial action are known to produce modifications (3). Brueggemann's analysis (4) of the ash from munitions burned in incinerators revealed substantial concentrations of polycyclicaromatic hydrocarbons (PAHs) but these compounds are not mentioned in Jenkin's report describing a method for quantitation of explosive residues in soil (5).

Nitroglycerin (NG) is a component of several military munitions (6) but is not generally found as a contaminant at OB/OD sites. Failure to detect NG may be attributed either to an actual absense of this compound or to an inability of current detection methods. Failure to detect NG could be due to its weak absorbance at UV wavelengths greater than 220 nm, where most munition analyses are performed.

Nitroguanidine (NQ) is used exclusively in M30 propellant, which is a mixture of NQ, NG and nitrocellulose. Since NQ is more polar than other munitions, it will leach rapidly and may therefore have a short residence time at the soil surface. It is also readily degraded by ultraviolet (UV) light (7). In addition, because of its high polarity, methods which utilize reversed phase high performance liquid chromatography (HPLC) and UV absorbance often have difficulty separating NQ from extractable natural soil components (8).

The nitramine munitions cyclotrimethylenetrinitramine (RDX) and cyclotetramethylenetetranitramine (HMX) are chemically similar and are widely used as explosives and as propellants. Industrial synthesis methods of these munitions do not produce pure compounds. All military grades of HMX contain RDX as an impurity and most RDX contains HMX (6). They are normally found together as environmental contaminants. Nitramines are easily extracted from soil samples and readily lend themselves to quantitation by reverse phase HPLC and UV absorbance.

Trinitrotoluene (TNT) and other nitroaromatics are the most common contaminants at many OB/OD sites. These compounds are less polar than the preceding and have excellent UV absorbance. They are readily quantitated by reverse phase HPLC and UV detection methods. Nitroaromatics undergo a variety of modifications in the environment but tend to remain identifiable as related forms because frequently their ring structure is not degraded (3).

MATERIALS AND METHODS

The method employed for sample preparation and extraction was adapted from the method developed and extensively tested by Jenkins (9, 10, 11). The modified procedure entailed grinding dried soil samples, and extracting into acetonitrile with 18 hours of sonication at 20 $^{\circ}$ C. Extracts were centrifuged at 3900 X G for 15 min, passed under piston pressure through a Gelman 0.45 Mm Acrodisc-CR disposable filter, and analyzed by HPLC. The latter portion of the sequence differs from Jenkins method in that a step requiring mixing the acetonitrile extract with an aqueous floculating solution was eliminated, and that an internal standard (1,3-dinitrobenzene, DNB) is incorporated.

An estimation of the efficiency of extraction of each compound was obtained by doping subsamples of uncontaminated surface soil (A horizon, Wheeling sandy loam [fine-loamy, mixed, mesic Ultic Hapludalfs]) with acetonitrile containing a mixture of selected OB/OD compounds plus DNB. The soil was dried and extracted as above and the efficiency of extraction was calculated from the amount of each compound recovered. Because the efficiency of extraction of the OB/OD components at our test sites was similar to that of DNB, a simplified recovery correction system was possible. All soil samples were extracted with acetonitrile containing 5 mg L⁻¹ (ppm) of DNB as an internal standard. Concentrations of OB/OD components observed in the extraction mixture were corrected for any observed loss of internal standard during the extraction process. Corrections were also made for any increases in concentration due to evaporation of the extraction solvent, acetonitrile. Corrections for losses or gains were generally minor.

Aqueous leachates from soil columns were collected and subsamples were adjusted to pH 6.00 \pm 0.05, then made to contain 300 g L⁻¹ NaCl. Two hundred mL of the resultant solution was put through a J.T. Baker 40 μ m Sep-Pak Octadecyl (C18) disposable cartridge at a rate of 1.8 mL min⁻¹. Cartridges were prepared for use by wetting with 2 mL methanol, followed by 2 mL water. Munitions were eluted from the cartridges with 2 X 1-mL additions of methanol and eluates were analyzed by HPLC. Efficiencies of recovery were also determined for this procedure using aqueous standards of munitions.

HPLC analyses were performed with a Hewlett-Packard (HP) 1050 HPLC system that consisted of an autoinjector, pumping module, and UV detector. Signal integration was performed with an HP 3396A integrator. All analyses except screening tests for the presence of NG were done by UV absorbance at 244 nm. NG was determined at 220 nm. Extracts of uncontaminated (background) and highly contaminated surface soils (A horizons) were screened by a gradient (Table I) method developed for this investigation. This method was developed to identify a broad range of munitions, environmentally modified unitions, and polycyclicaromatic hydrocarbons (PAHs). A 15- μ L sample was injected onto a 4.6 X 250 mm Rainin Microsorb C18 column with a 5 μ m particle size, in series with a 4.6 X 250 mm Supelcosil LC-PAH column. Elution was accomplished with a methanol:water gradient (Table I). This method was used for preliminary identification of the components present in contaminated soils from OB/OD sites. A simpler isocratic method developed by Jenkins et al. (12) was used to substantiate the identification of contaminants, and for quantitation. A mobile phase of 70.7 percent water, 27.8 percent methano] and 1.5 percent tetrahydrofuran was utilized at a flow rate of 2 mL min⁻¹ with a 4.6 X 75 mm Supelco LC8 column of 3 μ m particle size. This mobile phase and column combination were also used to screen for the presence of NG.

Percent Methanol (% MeOH)
30
33.5
47.5
51.0
54.5
100.0
100.0

Table I. HPLC Time/Gradient (Methnol:Water Mixture) for Initial Screening of Samples for a Broad Range of Munition-Related Analytes and PAHs

	Percent Munition Recovered (%). +s			
	From soil with acet	extracted onitrile	From aqueous leachate concentrates in MeOH	
Compound	doped <u>uncontam</u> .	doped <u>contam.</u>		
HMX	99 <u>+</u> 6	112 ± 4	29 <u>+</u> 10	
TNB	102 <u>+</u> 2	114 ± 3	123 ± 3	
RDX	95 <u>+</u> 1	91 <u>+</u> 2	38 <u>+</u> 1	
TNT	107 <u>+</u> 1	94 <u>+</u> 9	90 <u>+</u> 4	
2,4-DNT	103 <u>+</u> 1	110 <u>+</u> 5	108 <u>+</u> 7	
2,6-DNT	103 <u>+</u> 1	103 ± 2	104 <u>+</u> 20	
2-Amino-DNT	100 <u>+</u> <1	103 <u>+</u> 1		
4-Amino-DNT	98 ± 3	102 ± 4	137 <u>+</u> 40	

Table II. Efficiencies of Recovery of Munitions from Soil and Water

DISCUSSION

Jenkins (5,9,12) has provided a workable method for the extraction and analysis of explosive residues in soils. It is quick and provides good resolution of contaminants common to OB/OD sites. However, several problems remain. His isocratic HPLC method was designed primarily for use with nitramine and nitroaromatic compounds. More polar munitions tend to coelute with a complex mixture of natural soil components. Acetonitrile elutes as a large negative peak immediately prior to or coeluting with HMX, which complicates the quantitation of this compound. Compounds which elute later than the dinitrotoluenes suffer from peak broadening, which impairs their quantitation.

The gradient chromatography method presented here can accurately detect munition compounds and PAHs with a very broad range of polarities if they are present at >1.0 mg L⁻¹ (ppm). This method was used in our Laboratory for the first analysis (screening) of the most highly contaminated surface samples from munition contaminated OB/OD sites. When very polar munitions and PAH compounds were not present, the isocratic method was used for substantiation of compound identification and for quantitation.

PAH compounds have occasionally been detected in the waste products from munition incinerators, but detectable quantities of these compounds were not found in the ash or soils from OB/OD sites. A possible explanation for this difference is that the high energy intermediates responsible for ring fusion may accumulate at higher concentrations in contained combustion, or may arise from residual petroleum products associated with shell casings and bursting devices.

TNT remaining on the surface at OB/OD sites is converted to TNB, with the latter's concentration often exceeding that of the parent compound (16). Quantitation of TNB in acetonitrile extracts of soil was somewhat limited by background absorption due to material coextracted from contaminated soils. But the quantitation of TNB in the methanol concentrates of the aqueous leachates contained compounds that often severely interfered with this determination. TNT is also microbially degraded, by reduction to aminodinitrotoluenes, and may also be transformed into phenolic compounds and diazo forms (3). Although the aromatic ring structure of these compounds is resistant to degradation, evidence exists of other environmental processes in which these compounds become strongly bound to soil (3). Unlike the nitroaromatics, nitramine munitions and NQ undergo reactions which may leave little trace of the original compound (16, 3). Therefore, it is not surprising that recovery of munitions from soil is generally poor, after aging under simulated field conditions. Green et al.(17) were able to recover only a small fraction of added TNT after soil columns were leached. Banwart and Hassett (18) found that extractable TNT declined from 2000 mg kg⁻¹ (ppm) to $<20 \text{ mg kg}^{-1}$ when plants were grown for 90 days in soil amended with straw. Care must be taken in the design of experiments to determine the fate of munition residues within soil columns, to ensure that concentrations in leachates and extracts remain within the detection limits of the analytical method. However, it is also important to note that soils differ in their binding capacities and that loading a soil in excess of its binding capacity for any given compound may yield remarkably high initial rates of movement.

RESULTS

The above procedures have proven effective in recovering and quantitating munitions at moderate to high concentrations in sandy loam soil (Table II); they have the additional advantage of being simple and reproducible. However, several shortcomings were encountered. Efforts to identify minor components of the OB/OD soil contaminant mixture were not successful due to interferences from natural soil components.

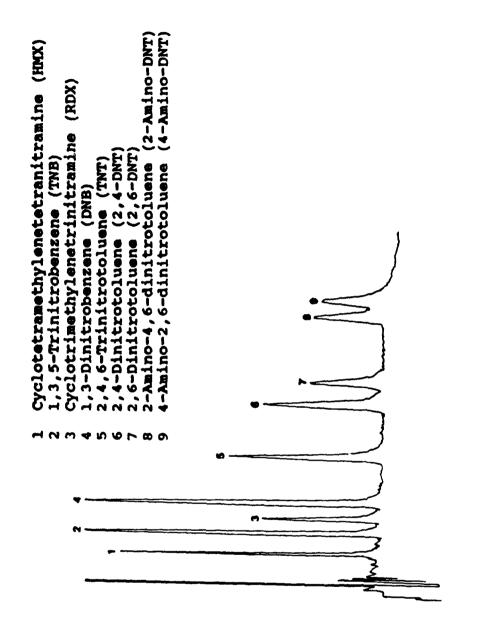
The isocratic HPLC method proved effective in quantitating intact munitions in water but performed less well with environmentally modified forms because they were later eluting and exhibited significant peak broadening (figure 1). We also observed that this solvent and column combination was unusually sensitive to temperature. At temperatures above 23 $^{\circ}$ C retention times were shortened and at 30 $^{\circ}$ C the system no longer resolved the two aminodinitrotoluenes.

The gradient procedure presented here effectively separated components of a mixture that included most compounds likely to be encountered during analysis of soils from OB/OD contaminated sites (Figure 2). It was able to detect many compounds that would otherwise be missed by the isocratic method. However, this chromatographic procedure required 90 min to complete, and could not be run at high sensitivity because of problems with baseline drift.

Recovery of munitions doped into uncontaminated soil were nearly quantitative (Table II); whereas recoveries from the soil columns ranged from 20-25 percent for TNT, 2-5 percent for 2,4-DNT, and even less for 2,6-DNT. Due to the low recoveries from the soil columns, the munition concentrations in soil extracts and leachate water were often diminished to the extent that accurate quantitation was dificult at reported detection limits (Table III). These criteria of detection were calculated using the computerized Quality Assurance Program of the U.S. Army Toxic and Hazardous Materials Agency (USATHAMA), based on the methods of Hubaux and Vos (13, 14).

Concentration of the munitions in soil extracts was attempted by evaporating the acetonitrile into a stream of dry nitrogen at 60 $^{\circ}$ C. This procedure was not used because significant losses of TNT, the DNTs, and the early eluting PAH compounds occurred. Concentration of munitions by simple evaporation of the acetonitrile at ambient laboratory temperatures (20-25 $^{\circ}$ C) was also unsuccessful due to unacceptable losses of compound.

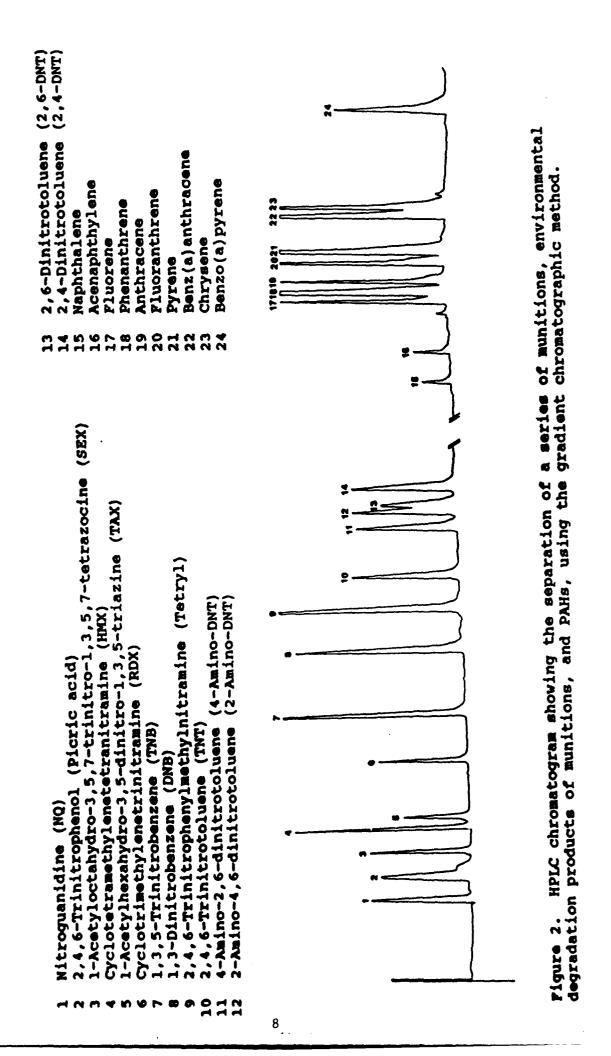
A procedure (15) developed by Brueggemann was used to concentrate munitions from our aqueous leachates. This method entails trapping munitions with disposable C18 columns, followed by elution with methanol. It was found to be useful for the nitroaromatics, however recoveries of the nitramine compounds were substantially lower (Table II). Passage of volumes of leachate greater than 30 mL had the effect of rinsing a portion of the nitramines from the column. In addition, this method concentrated naturally occurring soil components that interfered with determination of TNB and HMX.



An example of the separation of a series of munitions and associated co-contaminants, by the isocratic HPLC method. Figure 1.

:

7



······	
Compound	mg_L ⁻¹
нмх	0.15
TNB	0.15
RDX	0.067
DNB	0.15
TNT	0.093
2,4-DNT	0.17
2,6-DNT	0.37
2-Amino-DNT	0.14
4-Amino-DNT	0.12
	•

Table III. Criteria of Detection^{*} for Quantitation of Munitions Determined by HPLC

* the x value on the standard regression line that has the same y value as y at x = 0 on the upper confidence limit (95% conf. level) curve (13).

REFERENCES

- Layton, D.W., T.E. McKone, C.H. Hall, M.A. Nelson, and Y.E. Ricker. 1986. Demilitarization of conventional ordnance: priorities for data-base assessments of environmental contaminants. Report UCRL-15902. Livermore, CA: Lawrence Livermore National Laboratory. AD-A182922.
- U.S. AEHA Hazardous Waste Study #37-26-0593-86. 1986. Summary of AMC open-burning open-detonation grounds evaluations. March 1981-1985. Aberdeen Proving Ground, MD: U.S. Army Environmental Hygiene Agency.
- Walsh, M.E. 1990. Environmental transformation products of nitroaromatics and nitramines. Special Report 90-2 CRREL. Hanover, NH: Cold Regions Research and Engineering Laboratory, U.S. Army Corps of Engineers. AD-A220610.
- 4. Brueggemann, E.E. 1986. Liquid chromatographic determination of explosives and polynuclear aromatic hydrocarbons (PAHs) in deactivation furnace ash. Technical Report 8604. AD-A171269.
- Jenkins, T.F., and D.C. Leggett. 1990. Comparison of extraction techniques and solvents for explosive residues in soil. Special Report 85-22. Hanover, NH: Cold Regions Research and Engineering Laboratory, U.S. Army Corps of Engineers. AD-A166474.
- Department of the Army Technical Manual. 1984. Military Explosives. TM 9-1300-214. Picatinny Arsenal, NJ: U.S. Army Armament, Munitions and Chemical Command.
- 7. Burrows, D.W., M.O. Schmidt, R.H. Chyrek, and C.I. Noss. 1988. Photochemistry of nitroguanidine. Technical Report #8808. AD-A203200.
- 8. Walsh, M.E. 1989. Analytical methods for determining nitroguanidine in soil and water. CRREL Report 89-35. Hanover, NH: Cold Regions Research and Engineering Laboratory, U.S. Army Corps of Engineers. AD-A216651.
- 9. Jenkins, T.F., and M.E. Walsh. 1987. Development of an analytical method for explosive residues in soil. CRREL Report 87-7. Hanover, NH: Cold Regions Research and Engineering Laboratory, U.S. Army Corps of Engineers. AD-A183738.
- 10. Jenkins, T.F., P.W. Schumacher, M.E. Walsh, and C.F. Bauer. 1988. Development of an analytical method for the determination of explosive residues in soil part II. CRREL Report 88-8. Hanover, NH: Cold Regions Research and Engineering Laboratory, U.S. Army Corps of Engineers. AD-A213045.
- 11. Bauer, C.F., T.F. Jenkins, S.M. Koza, P.W. Schumacher, P.H. Miyares, and M.E. Walsh. 1989. Development of and analytical method for the determination of explosive residues in soil part III. CRREL Report 89-9. Hanover, NH: Cold Regions Research and Engineering Laboratory, U.S. Army Corps of Engineers. AD-A213000.

- 12. Miyares, P.H., and T.F. Jenkins. 1990. Salting-out solvent extraction method for determining low levels of nitromatics and nitramines in water. CRREL Special Report 90-30. Hanover, NH: Cold Regions Research and Engineering Laboratory. U.S. Army Corps of Engineers.
- USATHAMA Quality Assurance Program. 1990. USATHAMA PAM 11-41 (Rev. No. 0). Aberdeen Proving Ground, MD: U.S. Army Toxic and Hazardous Materials Agency.
- 14. Hubaux, A., and G. Vos. 1970. Decision and detection limits for linear calibration curves. <u>Anal. Chem</u>. 42(8):849-855.
- 15. Brueggemann, E.E. 1983. HPLC analysis of SEX, HMX, TAX, RDX, and TNT in wastewater. Technical Report 8206. AD-A127348.
- 16. Layton, D., B. Mallon, W. Mitchell, L. Hall, R. Fish, L. Perry, G. Snyder, K. Bogen, W. Malloch, C. Ham, and P. Dowd. 1987. Conventional weapons demilitarization: a health and environmental effects data base assessment; explosives and their co-contaminants. Final Report, Phase II, UCRL-21109. AD-A220588.
- 17. Greene, B., D.L. Kaplan, and A.M. Kaplan. 1985. Degradation of pink water compounds in soil - TNT, RDX, HMX. Natick, MA: U.S. Army Natick Research and Development Center. AD-A157954.
- 18. Banwart, W.L., and J.J. Hassett. 1990. Effect of soil amendments on plant tolerance and extractable TNT from TNT contaminated soils.<u>Agron.</u> <u>Abstr</u>, 82nd Annual Meeting, p 33.

DISTRIBUTION LIST

No. of Copies

4	Commander U.S. Army Medical Research and Development Command ATTN: SGRD-RMI-S Fort Detrick Frederick, MD 21702-5012
2	Commander U.S. Army Biomedical Research and Development Laboratory ATTN: SGRD-UBZ-IL Fort Detrick Frederick, MD 21702-5010
1	Commander U.S. Army Biomedical Research and Development Laboratory ATTN: SGRD-UBZ-P Fort Detrick Frederick, MD 21702-5010
2	Defense Technical Information Center ATTN: DTIC-FDAC Cameron Station Alexandria, VA 22304-6145
1	Commandant Academy of Health Sciences, U.S. Army ATTN: HSHA-IPM (Chief, Environmental Quality Branch) Fort Sam Houston, TX 78234-6100
1	Commandant Academy of Health Sciences, U.S. Army ATTN: HSHA-DCD Fort Sam Houston, TX 78234-6100
1	Chief Army Environmental Office ATTN: ENVR-EP The Pentagon Room 1E-669 Washington, DC 20310-2600
1	Commander U.S. Naval Facilities Engineering Command Environmental Quality Division 200 Stovall Street Alexandria, VA 22332-2300
1	Headquarters Air Force Engineering Service Center ATTN: RDV Tyndall Air Force Base, FL 32403-6001

•

1	Commander U.S. Army Materiel Command ATTN: AMSCG 5001 Eisenhower Avenue Alexandria, VA 22333-0001
1	Commander U.S. Army Environmental Hygiene Agency ATTN: HSDH-AD-L Aberdeen Proving Ground, MD 21010-5422
1	Dean School of Medicine Uniformed Services University of the Health Sciences 4301 Jones Bridge Road Bethesda, MD 20814-4799
1	HQDA (DASG-PSP-E) 5109 Leesburg Pike Falls Church, VA 22041-3258
1	Commander U.S. Army Toxic and Hazardous Materials Agency ATTN: CETH-TE-D Aberdeen Proving Ground, MD 21010-5401
1	Commander U.S. Army Health Services Command ATTN: HSPA-P Fort Sam Houston, TX 78234-6000