

DND 1111 0000

2

AD _____

ANTIBODY TO THE RNA-DEPENDENT DNA POLYMERASE OF HTLV-III:
CHARACTERIZATION AND CLINICAL ASSOCIATIONS

AD-A231 466

FINAL REPORT

JEFFREY LAURENCE

MARCH 1, 1990

Supported by

U.S. ARMY MEDICAL RESEARCH AND DEVELOPMENT COMMAND
Fort Detrick, Frederick, Maryland 21702-5012

Contract No. DAMD17-87-C-7020

Cornell University Medical College
1300 York Avenue
New York, New York 10021

DTIC
ELECTE
JAN 24 1991
S E D

Approved for public release; distribution unlimited

The findings in this report are not to be construed as an
official Department of the Army position unless so designated
by other authorized documents

REPORT DOCUMENTATION PAGE

Form Approved
OMB No. 0704-0188

1. REPORT SECURITY CLASSIFICATION Unclassified		1b. RESTRICTIVE MARKINGS	
2. SECURITY CLASSIFICATION AUTHORITY		3. DISTRIBUTION / AVAILABILITY OF REPORT Approved for public release; distribution unlimited	
3. DECLASSIFICATION / DOWNGRADING SCHEDULE			
PERFORMING ORGANIZATION REPORT NUMBER(S)		5. MONITORING ORGANIZATION REPORT NUMBER(S)	
4. NAME OF PERFORMING ORGANIZATION Cornell University Medical College	6b. OFFICE SYMBOL (if applicable)	7a. NAME OF MONITORING ORGANIZATION	
6. ADDRESS (City, State, and ZIP Code) 300 York Avenue New York, New York 10021		7b. ADDRESS (City, State, and ZIP Code)	
8. NAME OF FUNDING / SPONSORING ORGANIZATION U.S. Army Medical Research & Development Command	8b. OFFICE SYMBOL (if applicable)	9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER DAMD17-87-C-7020	
10. SOURCE OF FUNDING NUMBERS			
10. ADDRESS (City, State, and ZIP Code) Fort Detrick Frederick, Maryland 21702-5012		PROGRAM ELEMENT NO. 603105A	PROJECT NO. 3M2- 63105DH29
		TASK NO. AB	WORK UNIT ACCESSION NO. 043
11. TITLE (Include Security Classification) U) Antibody to the RNA-Dependent DNA Polymerase of HTLV-III: Characterization and Clinical Associations			
12. PERSONAL AUTHOR(S) Jeffrey Laurence			
13a. TYPE OF REPORT Final Report	13b. TIME COVERED FROM 11/15/86 TO 11/14/89	14. DATE OF REPORT (Year, Month, Day) 1990 March 1	15. PAGE COUNT 18
16. SUPPLEMENTARY NOTATION			
17. COSATI CODES		18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number)	
FIELD	GROUP	SUB-GROUP	RA 1, AIDS, Antibody, Clinical, HTLV-III, Infectious Diseases
06	03		
06	13		
19. ABSTRACT (Continue on reverse if necessary and identify by block number)			
<p>This project was designed to explore B cell epitopes of the HIV-1 polymerase which are important to the identification of in vitro immunologic responses correlating with progression of HIV disease, and may provide targets for immune intervention. We first reported a heterogeneous immune response for inhibition of HIV reverse transcriptase activity by IgGs from HIV seropositive individuals. Loss of this inhibitor appeared to be associated with development of clinical disease, a finding which has been confirmed by at least three groups world-wide. We subsequently attempted to identify a linear sequence of the polymerase which could be used in a more rapid serologic assay to define the epitope reactivity of these anti-T antibodies. A statistically significant correlation between anti-RT serologic reactivity and RT enzymatic inhibition was found utilizing two peptides from conserved, amino-terminus residues of the polymerase. Further epitope mapping is in progress, in concert with development of a in vivo mouse model to evaluate the efficacy of antibody reagents in concert with cell activation inhibitors to block progression of HIV infection.</p>			
20. DISTRIBUTION / AVAILABILITY OF ABSTRACT <input type="checkbox"/> UNCLASSIFIED / UNLIMITED <input checked="" type="checkbox"/> SAME AS RPT <input type="checkbox"/> DTIC USERS		21. ABSTRACT SECURITY CLASSIFICATION Unclassified	
22a. NAME OF RESPONSIBLE INDIVIDUAL Mary Frances Bostian		22b. TELEPHONE (Include Area Code) 301-663-7225	22c. OFFICE SYMBOL SGRD-RMI-S

Purpose: To explore epitopes related to the HIV-1 polymerase which are important both to the identification of in vitro cellular responses correlating with disease progression, and as targets for immune intervention in HIV infection.

Relevance to WRAIR Plan: 1. Definition of viral epitopes which should serve as targets for vaccine development, passive immunization, and cytotoxic T cell priming.

2. Continued exploration of the relevance of viral activation to immune intervention.

Collaboration with WRAIR Programs: 1. Use of the large bank of serial serum samples from a population in which clinical and laboratory data are available.

2. Possibility of using our newly developed transgenic mouse model to evaluate polymerase epitopes identified in this project as targets for immune intervention.

PUBLICATIONS, ALL ACKNOWLEDGING USAMRDC SUPPORT AND RELEVANT TO THIS PROPOSAL:

1. Laurence J, Freidman SM, Chartash EK, Crow MK, Posnett DN. Human immunodeficiency virus infection of helper T cell clones. Early proliferative defects despite intact antigen-specific recognition and interleukin 4 secretion. J Clin Invest 83:1843-48, 1989.

2. Laurence J, Sellers MB, Sikder SK. Effect of glucocorticoids on chronic human immunodeficiency virus (HIV) infection and HIV promoter-mediated transcription. Blood 74:291-297, 1989.

3. Laurence J, Cooke H, Sikder SK. Effect of tamoxifen on regulation of chronic HIV-1 infection and HIV LTR-directed transcription. Blood, 75:696-703, 1990.
4. Laurence J, Sikder SK, Jhaveri S, Salmon JE. Phorbol ester-mediated rescue of HIV from a chronically infected promonocyte clone blockade by protein kinase inhibitors and relationship to tat activation. Biochem. Biophys. Res. Commun. 166:349-357, 1990.
5. Laurence, J. Novel vaccination and anti-receptor strategies against HIV. AIDS Res. Human Retrov. 6:3-8, 1990.
6. Laurence J, Kulkosky J, Friedman SM, Posnett DN, Ts'o POP. PolyI.polyC12U-mediated inhibition of loss of alloantigen responsiveness and viral replicaton in human CD4+ T cell clones exposed to human immunodeficiency virus in vitro. J Clin Invest 80:1631-39, 1987.
7. Laurence J, Saunders A, Kulkosky J. Characterization and clinical association of antibody inhibitory to HIV reverse transcriptase activity. Science 235:1501-4, 1987.
8. Laurence J, Kulkosky J, Dong B, Early E, Mann W, Snyderman R, Cianciolo GJ. A soluble inhibitor of T lymphocyte function induced by HIV-1 infection of CD4+ T cells: Characterization of a cellular protein and its relationship to p15E. Cell Immunol, 128:337-352, 1990.
9. Laurence J. Molecular interactions among herpesviruses and human immunodeficiency viruses. J. Infect. Dis. 162:338-346, 1990.
10. Laurence J, Saunders A, Early E, Salmon JE. Human immunodeficiency virus infection of monocytes: relationship to Fc-gamma receptors and antibody dependent viral enhancement. Immunology 70:338-343, 1990.

Accession For	
NTIS GRA&I	<input checked="" type="checkbox"/>
DTIC TAB	<input type="checkbox"/>
Unannounced	<input type="checkbox"/>
Justification	
By _____	
Distribution/	
Availability Codes	
Dist	Avail and/or Special
A-i	

I. MAJOR PROJECT

Background: The serum of many animals naturally infected with retroviruses contain antibodies capable of blocking the enzymatic activity of particulate reverse transcriptases. Stimulated by the correlations of antibody to reverse transcriptase (RT) with clinical status first identified in cattle and cats, we reported a heterogeneous immune response for inhibition of HIV-associated RT function by IgGs from HIV-infected individuals (1). Loss of this inhibitor appeared to be associated with development of clinical disease (1).

This finding, and the clinical associations, were subsequently confirmed by two other groups (2,3). The probability that this functional assay served to identify discrete subsets of anti-polymerase antibodies was strengthened by the fact the p66/p51 RT of HIV-1 (4,5) and HIV-2 (6) are highly immunogenic, detectable by immunoblotting in >90% of seropositive individuals, and do not correlate with clinical stages of HIV infection (4). Such serologic studies cannot, of course, detect changes in epitope specificity among antibody species as the disease progresses.

There is a need to explain why HIV RT, a core protein that is present at substantially lower concentrations in the virion than structural core proteins such as p24, is so uniquely immunogenic. Indeed, HTLV-I and HTLV-II are not commonly associated with anti-RT responses (4). Perhaps continuous cell

lysis in HIV infection constantly challenges the hosts immune system with RT. This could induce not only the strong humoral responses reported above, but also strong anti-pol cytotoxic T lymphocyte responses discovered by others (7). Longitudinal studies with this CTL anti-pol assay system are also ongoing, with preliminary reports demonstrating a correlation between loss of MHC class I restricted CD8+ T cell activity against RT targets and disease progression (7).

Present data: In an attempt to define a linear sequence of the HIV pol which could be used in a serologic assay to define these anti-RT inhibitory antibodies, synthetic peptides were prepared from nucleotide sequences of HIV pol regions homologous to the catalytic sites of other mammalian polymerases. The initial peptides, ranging in length from 8 to 15 amino acids, were synthesized at The New York Blood Center by the solid-phase method using an automated peptide synthesizer (Applied Biosystems, Foster City, CA). Peptide resins were cleaved by hydrogen fluoride, extracted, and analyzed for purity by HPLC.

The five sequences we initially employed were:

Code	Sequence	AA Position	Activity	Reference
I	LDVGDAYF	109-116	NTP binding site	8
II	VLPQGWKGSP	158-157	conserved region	8
III	IQKLVGKLNW	257-266	conserved region	8
IV	PENPYNTPVFAIKK	219-232	conserved region	9
V	DSRNPLWKGPALLW	496-509	COOH terminus	9

ELISA. Peptide solutions of 10mg/ml in PBS were air dried overnight at 37⁰C in polyvinyl, 96-well microtiter plates to yield 1-5 ug/well. Nonspecific binding sites were blocked with BSA. After a 6 h incubation at 25⁰C and a 16h incubation at 4⁰C, serum samples were added at 1:100, 1:200 and 1:400 dilutions. After 2 h at 25⁰C the wells were washed 10-20 times with 0.2% Tween 20 in PBS, after which affinity-purified, alkaline phosphatase conjugated goat antibody to human IgG was added. After 2 h the plates were washed, and substrate solution was added. After a 30 in. incubation the reaction was stopped by adding 1N H₂SO₄, and the optical densities were analyzed on an automated ELISA scanner. Seropositivity was defined as any value greater than three standard deviations obtained with negative controls.

SAMPLES. The samples analyzed included 100 sera obtained from our original longitudinal survey (1979-1985). An additional 150 samples have been obtained from Dr. Robert Redfield at WRAIR and are in the process of being analyzed.

A statistically significant but imperfect correlation was found between the presence of antibodies capable of blocking the catalytic activity of RT, as reported by our functional assay (1) and anti-peptide I, II or IV reactivity. The latter two residues correspond to the putative HIV catalytic site as determined by insertional mutagenesis (8, 9). Interestingly, they are outside the regions of the pol gene thought to be immunogenic by

hydrophobicity plots (10) or B cell eiptope computer analysis (11), yet are within the few discrete regions of pol predicted to be strong T cell epitopes (11).

The data, gathered on 100 HIV seropositive individuals at various clinical stages of infection and 15 seronegative controls were as follows:

0/15 HIV seropositive individuals were positive by ELISA with any of the 5 synthetic peptides.

38/100 HIV seropositive individuals were positive for one or more pol peptides. Of these 38, only 3 had WR stage 5-6 disease, with all others in the WR 1-2 category.

Of the 62 seropositive individuals who were non-reactive, all were more advanced than WR 2, and only 6 had anti-RT catalytic activity.

The raw data for the 38 reactive patient samples were:

Peptide No.	I	II	III	IV	V
No. sera +	31	26	3	8	3
% positive	81.6	68.4	7.9	21.1	7.9

Serologic Reactivity vs. Anti-RT inhibitory Activity

	Serologic Activity to peptides II and/or IV	No serologic activity for II and/or IV
No anti-RT activity	2	7
Anti-RT activity	20	9

Interestingly, Peptides III and V gave low frequencies of positive reactions similar to those for pol synthetic peptides selected on the basis of hydrophobicity plots (10).

II. Attempts to correlate anti-RT reactivity with standard neutralization assays or p24 antigenemia.

The possibility that these anti-RT IgGs correlated with existence of other antibodies with reactivity against neutralizing envelope epitopes was investigated. This latter area is controversial. Virus neutralizing factors have been found in >50% of serum samples obtained from AIDS and ARC patients (12), and in a higher number of HIV seropositive asymptomatic carriers (13). Clinical correlations in patients followed over time has been weak, however. The assays typically involve single HIV isolates as targets, albeit envelope variability is well documented among HIV strains, even from a single individual (14).

(a) We attempted to link HIV neutralization in vitro with anti-RT activity, using IgG from of the asymptomatic carriers. Using $\geq 75\%$ inhibition of enzyme activity as the criterion for neutralization, we found most samples effective at 10-100 ug IgG, without regard to the patient's clinical status or the anti-RT

capacity of the IgG.

(b) We have received 78 (38 HIV seropositive) serum samples from Dr. Zvi Bentwich, Kaplan Hospital, Rehovot, Isreal, together with clinical and immunologic data. We have correlated serum p24 antigen levels with anti-RT activity and related these to clinical course, as described below:

Group I: n = 24
HIV seropositive
Clinical data: CD4+ T cells < 400/mm³
lymphadenopathy or other clinical
manifestations of HIV infection

Group II: n = 14
HIV seropositive
Clinical data: CD4+ T cells ≥ 400/mm³
asymptomatic

Group III: n = 13
HIV seronegative
Clinical data: "non-HIV immune defects"

Group IV: n = 12
HIV seronegative
Clinical data: HIV at risk group, normal immune
function.

Group V: n = 15
HIV seronegative
Clinical data: non-HIV risk group, normal immune
function.

These sera were evaluated for antibody against HIV-1 RT catalytic activity, as well as for circulating p24 antigen by ELISA-based antigen capture (Abbott Labs, Chicago, IL). As shown in Table I, 6/24 (25.0%) of Group I samples were positive for p24 antigenemia, while 1/14 (7.1%) of Group II samples were as well. In contrast, anti-RT antibodies were noted in 2/24 (8.3%) of Group I, but 8/14 (57.1%) of Group II samples. These data

Table I. Correlation of p24 antigenemia and anti-RT antibodies with degree of clinical or immune deficit in HIV infection.

GROUP I: HIV seropositive,
symptomatic

GROUP II: HIV seropositive,
asymptomatic,
CD4+ T cells
>400/mm³

Sample code	p24 antigen (pg/ml)	anti-RT	Sample	p24 antigen (pg/ml)	anti-RT
778	1212	-	99	0	-
840	0	-	662	0	+
261	0	-	272	14	-
713	0	-	91	0	+
589	0	+	365	0	+
288	0	+	661	0	+
634	0	-	222	0	+
842	0	-	759	0	+
241	240	-	641	0	-
482	0	-	67	0	-
432	102	-	580	0	-
878	>8000	-	765	45	-
830	0	-	169	0	-
449	0	-	254	0	-
668	40	-			
320	0	-			
289	0	-			
815	0	-			
930	5152	-			
707	0	-			
966	0	-			
626	0	-			
469	0	-			
237	0	-			
% positive	25.0	8.3		7.1	57.1

support our original observation of association of such antibodies with improved clinical status. It also emphasizes the fact that while p24 antigenemia tends to be associated with advancing clinical stage of HIV infection, this correlation is far from perfect. Combination of anti-RT antibodies with other parameters such as absolute CD4+ T cell count, serum p24 levels, etc., may assist in forming an improved prognostic indication for state of HIV infection.

Recent work: To strengthen our work in refining these epitopes, we sought to use recombinant peptides which extended the sequences derived from our preliminary data. We established a collaboration with Dr. Sean Nowlan, who had described three highly immunogenic regions of pol (15):

Epitope designation	Amino acid residues	HIV-1 nucleotides	Provisional epitope map
rI	9-214	2101-2774	144-191
rII	191-335	2700-3134	214-335
rIII	335-567	3134-3832	511-536

These pol sequences, and 5 additional non-reactive recombinant peptides from pol (15) which we have also received, will be valuable in refining the exact specificity of pol targets for both antibody and CTL.

III. RELATED PROJECT: Regulation of chronic HIV infection and its relationship to immune activation and immune modulators.

As is apparent from our list of WRAIR-assisted publications, while this work was in progress we came to realize

the importance of several biologic properties of HIV also relevant to immune intervention. HIV infection of CD4+ T lymphocytes and monocytes may lead to a low level chronic or latent phase during which there is little or no viral replication. Transition to a productive state can be triggered, at least in vitro, by immunologic activation of infected cells (16). These signals induce cellular trans-acting factors, which in turn stimulate transcription from the HIV long terminal repeat (LTR). The virus is then able to synthesize its own regulatory factors which further stimulate transcription from the LTR, leading to viral replication and cell death (17).

We explored phorbol ester-mediated rescue of virus from a clone of promonocyte cells, U1.1A, chronically infected with HIV-1. Cellular responses to phorbol esters mimic responses to growth factors, hormones, and immunologic activation. In terms of HIV induction via PMA or immunologic stimuli (antigen, alloantigen, lymphokine), PKC activation presumably initiates an intracellular phosphorylation cascade, the end result of which is to stimulate pre-existing nuclear trans-acting factor (18). NF- κ B is the enhancer protein first described as capable of upregulating HIV replication by interacting with the 5' LTR at a site different from that of the tat-responsive TAR region (19). However, other PMA responsive sequences have been identified in viral sequences shared with HIV that affect other transcription factors (20). We reported (21) that PMA and PDB (4 β -phorbol 12,13-dibutyrate) could lead to a productive infection not

elicited by congeners that bind to phorbol ester receptors but do not transduce an intracellular signal (Table II). Induction appeared specific to activation of PKC, and was blocked by three PKC inhibitors, H7, 2-aminopurine (Table III) and sangivamycin (data not shown). Its association with regulatory pathways involving the tat transcription unit was also explored. PMA-mediated enhancement of an HIV-LTR driven reporter gene was not blocked by H7 at concentrations capable of inhibiting HIV replication while the PMA effect was synergistic with tat-linked trans-activation.

There is the need to identify similar agents, with potential clinical utility, capable of blocking upregulation of virus in chronic HIV infection. Indeed, classic anti-virals such as 3'azido-3'deoxythymidine (AZT) have no effect in preventing induction of HIV expression in macrophage (U1.1) and other cell lines, while alpha-interferon may have a salutary effect (22).

As gonadal steroids and glucocorticoids can alter specific gene expression in at least one immunosuppressive murine retrovirus, mammary tumor virus (MMTV), we first explored the ability of dexamethasone (DXM) to upregulate latent HIV replication or to alter transcription at the HIV-1 long terminal repeat (LTR). Chronically infected lymphocytes as well as lymphocyte and monocyte (U1.1) cell lines were unperturbed by DXM used over broad concentrations (10^{-4} to 10^{-9} M) and time intervals (24-96 hrs.) (23). This unresponsiveness corresponded to the lack of a positive effect of DXM on HIV associated trans-

Effect of phorbol esters on induction of HIV-1 from chronically infected U1.1A promonocytic cells

<u>Phorbol Ester</u>	<u>Concentration (ng/ml)</u>	<u>HIV-1 p24 core antigen (pg/10⁴ cells)</u>
None	-	2920
PMA	5	13,040 ± 1240
PMA	50	32,440 ± 4920
PMA	500	11,200 ± 600
PDB	5	31,880 ± 4920
PDB	50	59,400 ± 6320
PDB	500	46,840 ± 2800
phorbol 13-acetate	5	2480
phorbol 13-acetate	50	3600
phorbol 13-acetate	500	1040
4B-phorbol	5	4640
4B-phorbol	50	5040
4B-phorbol	500	3800

Table III. Effect of protein kinase C (PKC) inhibitors on induction of HIV-1 from chronically infected U1.1A promonocytic cells*

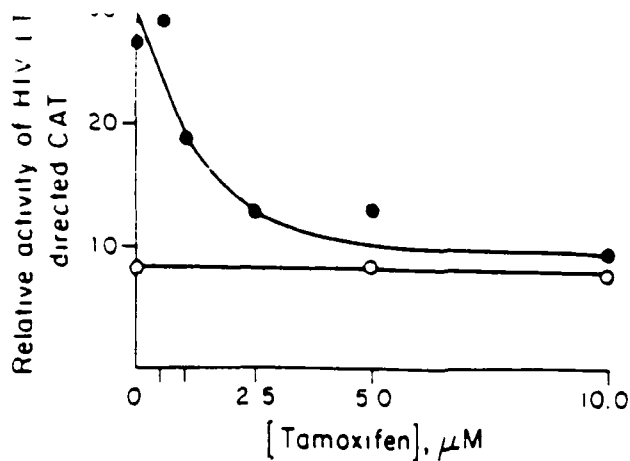
Expt.	PMA (5ng/ml)	<u>PKC Inhibitor</u>		<u>HIV-1 activity</u>	
		Agent	Concentration (μM)	[p24] core Ag (pg/10 ⁴ cells)	Inhibition (%)
A	-	-	-	1,880	-
	+	-	-	14,660 ± 1080	-
	+	H7	10	13,540 ± 650	7.6
	+	H7	50	5,840 ± 2590	60.2
	+	H7	75	2,150 ± 730	85.3
	+	H7	100	1,070	92.7
B	-	-	-	1,080	-
	+	-	-	13,770	-
	+	2-AP	10	17,930	0
	+	2-AP	50	2,890	79.0
	+	2-AP	75	710	94.8
	+	2-AP	100	640	95.4

*Cells were plated at 1 x 10⁴/microwell in 0.2ml of culture medium together with buffer or PKC inhibitor (either the isoquinoline H7 or 2-aminopurine) for 1h at 37°C, followed by addition of the phorbol ester PMA. Culture supernatants were harvested 48h later and HIV p24 core antigen concentrations assessed by ELISA.

activation. These cells possessed the appropriate steroid receptors, as DXM downregulated Fc gamma type I receptors in both normal and HIV-infected monocytic cells. In addition, DXM could block the transcriptional enhancement of an HIV-LTR linked reporter gene by phorbol ester, while leaving basal levels of HIV-LTR directed transcription unperturbed (23).

We subsequently showed that rescue of virus could be blocked by the triphenylethylene anti-estrogen tamoxifen (TMX), at concentrations which did not affect cellular DNA synthetic responses. This effect correlated with tamoxifen's ability to completely inhibit PMA enhancement of HIV-promoter mediated trans-activation in both monocytic and CD4+ T cells (Fig. 1). No interference with a primary infection was noted. Tamoxifen's mechanism of action may relate both to its capacity to inhibit PKC, as well as to consensus sequences for gonadal steroid responsive elements in the HIV LTR. Thus, TMX inhibited HIV-LTR-directed tat activity with an IC₅₀ of 2uM, and its IC₅₀ for inhibition of PKC is 6.1 uM (24); typical serum levels of this drug in man are on the order of 2uM (25).

Manipulation of gonadal HREs in relationship both to viral regulation and immune modulation may be especially pertinent in HIV infection for several reasons. First, TMX has a variety of immunologic effects. In vitro modulation of human lymphocyte reactivity includes increases in pokeweed mitogen-driven immunoglobulin production, inhibition of CD8+ T suppressor cell but not CTL phenomena, and augmentation of natural killer cell



B



	<u>1</u>	<u>2</u>	<u>3</u>	<u>4</u>	<u>5</u>	<u>6</u>	<u>7</u>
Conversion (%)	16.9	57.8	62.0	39.0	26.0	28.1	19.7
PMA (50ng/ml)	-	+	+	+	+	+	+
TMX (μM)	0	0	0.5	1.0	2.5	5.0	10

Fig.1 . Effect of tamoxifen on PMA-mediated enhancement of HIV-LTR-CAT. The CAT plasmid ($1\mu\text{g}$ DNA) was co-transfected into U1.1A cells (2×10^6) with a plasmid containing *tat* ($1\mu\text{g}$ DNA). PMA (50 ng/ml) and/or TMX (1-10 μM) was present throughout the 48h culture period. The percent conversion of [^{14}C]chloramphenicol (Cm) to its acetylated forms (ACm) was determined.

A. Dose-response curve for inhibition of PMA-driven upregulation of HIV-LTR-CAT activity. This activity was measured in the presence (closed circles) and absence (open circles) of PMA.

B. Chromatographic data for conversion of Cm to ACm.

activity (26). Second, a prominent manifestation of HIV infection, Kaposi's sarcoma, is most prevalent among men. A model for this disease, produced when tat under the control of the HIV LTR was introduced into the germline of mice, was elicited only in male transgenic animals, despite equivalent levels of tat mRNA expression their female counterparts (27). The possibility that this sexual difference in phenotype is hormonally based is under investigation (27). A further indication that hormonal therapy might have a therapeutic role in HIV infection was the preliminary observation that the synthetic progesterone megestrol acetate (Megace) led to significant improvements in appetite, lethargy, and body weight in some HIV infected patients (28), and is under clinical evaluation at Walter Reed.

References

1. Laurence J, Saunders A, Kulkosky J. Science 235:1501, 1987
2. Sano K, Lee MH, Morales F, Nishanian P, Fahey J, Detels R, Imagawa DT. J Clin Micro 25:2415, 1987.
3. Chatterjee R, Rinaldo CR Jr, Gupta P. J Clin Immunol 7:218, 1987.
4. DeVico AL, Veronese FDV, Lee SL, Gallo RC, Sarngadharan MG. AIDS Res Human Retrvir 4:17, 1986.
5. Veronese FDM, Copeland TD, DeVico AL, Rahman RR, Oroszlan S. Gallo RC, Sarngadharan MG. Science 231:1289, 1986.
6. Allan JS, Coligan JE, Lee T-H, Barin F, Kanki PJ, M'Boup S, McLane MF, Groopman JE, Essex M. Blood 69:331, 1987.
7. Walker BD, Flexner C, Paradis TJ, Fuller TC, Hirsch MS, Schooley RT, Moss B. Science 240:64, 1988.
8. Hizi A, Barber A, Hughes SH. Virology 170:326, 1989.
9. Prasad VR, Goff SP. Proc Natl Acad Sci 86:3104, 1989.
10. Gnann JW, Schwimmbeck PL, Nelson JA, Truax AB, Oldstone MBA. J Infect Dis 156:261, 1987.
11. Zvelebil MJJM, Sternberg MJE, Cookson J, coates ARM. FEBS Lett 242:9, 1988.
12. Rasheed, S, et al. Virology 150:1, 1986.
13. Robert-Guroft M, Brown M, Gallo RC. Nature 316:72, 1985.
14. Hahn BH, et al. Science 232:1548, 1986.
15. Padberg C, Nowlan S, Mermer B. AIDS Res Human Retrov 5:61, 1989.
16. Zagury, D, et al. Science 231:850, 1986.
17. Tong-Starksen SE, Luciw PA, Peterline BM. PHAS 84:6845, 1987.
18. Nelson B, Hellman L, Seu R. Mol Cellular Biol 8:3526, 1988.
19. Nabel GJ, Rice SA, Knipe AM, Baltimore D. Science 239:1299, 1988.
20. Chiu R, Imagawa M, Imbra RJ, Brockoven JR, Karin M. Nature

329:648, 1987.

21. Laurence J, Sikder SF, Jhivaris, Salmon JE. Clin Res 37:414A, 1989.

22. Poli G, et al. Science 244:575, 1989.

23. Laurence J, Sellers MS, Sikder SK. Blood 74:291, 1989.

24. Hogan F, Cooke E, Hallet MB, Manuel MR. Biochem Pharmacol 35:4463, 1986.

25. Daniel P, et al. Eur J Cancer Clin Oncol 17:1183, 1981.

26. Nagy E, Berczi I. Immunopharmacol 12:145, 1986.

27. Vogel J, et al. Nature 335:606, 1988.

28. vonRoenn JH, et al. Ann Intern Med 109:840, 1988.