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The marine adhesiv	e proteins of so	me six diffe	erent invert	tebrates hav	е	
having high isoelectric	points and high	lnese protei levels of ly	ins are char /sine and Do	racterized b opa. There	y are	
few unifying features in	the primary seq	uence. All	of the prot	ceins have r	epeat	
sequences with consensus peptides ranging from 4 to 10 amino acids. Most of						
The cross-linking of the	se proteins is m	ediated by c	atecholoxic	lase. This	enzyme	
catalyzes the conversion	of Dopa to o-qu	inone. The	latter taut	comerizes to	α,β-	
dehydro-functional group	s that are again	oxidized by	the enzyme	e, this time	to	
of nucleophilic addition	reactions.	expected to	undergo a c	liverse asso	rtment	
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ANNUAL AND FINAL REPORT ON CONTRACT: N00014-86-K0717 PRINCIPAL INVESTIGATOR: J. Herbert Waite CONTRACTOR: University of Delaware, College of Marine Studies CONTRACT TITLE: Polymerization of a Quinone-Crosslinked Marine Bioadhesive Protein

START DATE: 1 September 1986

RESEARCH OBJECTIVE: The main objectives of this research are A. to survey the diversity of DOPA-containing marine adhesive proteins or cements, and B. to examine the chemical mechanism of quinonetanning (i.e. cross-linking) of DOPA-containing proteins.

PROGRESS (Year 3): <u>SEQUENCE STUDIES</u> - The adhesive proteins of four organisms in addition to those from the original mussels, <u>Mytilus edulis and M. californianus</u>, have been characterized and partially sequenced. Consensus peptides from these proteins are shown in Table 1. Consensus peptides are those often repeated in the primary structure of the respective proteins but they are not necessarily the only sequences in the proteins. The three mussels <u>Mytilus edulis</u>, <u>M. californianus</u> and <u>G. demissa</u> all share the motif

Z-Dopa/Tyr-X-Y-Z-Dopa/Tyr-Lys

where Z is Ser at Thr in the two species of <u>Mytilus</u> and Gly in <u>Geukensia</u>. X and Y are usually 3- or 4-Hyp in <u>Mytilus</u> and variable in <u>Geukensia</u>, although Y is frequently pro/Hyp. The fourth mussel listed in Table 1, <u>Aulocomya ater</u>, from the southern coast of Chile, has a much shorter consensus sequence with only one DOPA per peptide seven amino acids long. The variable sixth position X is a hydrophobic amino acid. Lysine in position 7 is as often hydroxylated (δ -hydroxylysine) as not. The reef-building polychaete secretes a composite marine cement that has a glycine-rich consensus peptide with 2 DOPAs (per nine residues) separated from one another by a single glycine. In the trematode Fasciola, Dopas are commonly separated from one another by 2 amino acids (Glys, Asp, Ser) and from C-terminal lysine by a single Gly. The second Fasciola protein sequenced is distinct from all others in containing no Lys, in its low MW, and in having the lowest pI. A consensus peptide, GlyHisGlyDopa, repeated perhaps 6-8 times is inferred from products of digestion by pepsin.

The diversity of DOPA-bearing sequences encountered thus far in naturally occurring proteins that undergo quinone-tanning suggest that this process is not restricted to a single monolithic sequence and that further unique and as of yet unknown functions may be served by the sequences of these proteins.

CROSS-LINKING STUDIES

In an attempt to elucidate the nature of the chemical species, participating in curing of adhesive protein by covalent crosslinking, we have investigated the oxidation products of a synthetic peptide analogue, N-acetyl-DOPA ethyl ester (NAc-DEE), ssion For together with other related substances, under conditions felt to GRAAT approximate the salinity and pH found in the native environment of TAB ucumped rightion the mussel Geukensia demissa. We have observed that the quinones derived from DOPA-containing molecules such as NAc-DEE undergo a ribution/ novel oxidative rearrangement to form a catecholic enamic acid Lability Codes

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derivative. This derivative, N-acetyl- α , β -dehydroDOPA ethyl ester, was characterized by taking advantage of the unique difference spectra between borate and HCl solutions of catechols bearing, α , β -unsaturated carbons. Any loss of the unsaturation would result in the formation of a tetrahedral carbon adjacent to the ring, with the concomitant loss of the characteristic spectra. Provided no de-acetylation or de-esterification had occurred, α , β -dehydroDOPA ethyl ester then has the probable structure shown in Fig. 1C.

The rearrangement of the quinone to form an α,β -desaturated DOPA derivative was probably spontaneous, for the following reasons. Firstly, the rearrangement occurred following enzymic oxidation utilizating either mushroom tyrosinase or byssal catechol oxidase, which suggests that the precise nature of the active site is irrelevant for the rearrangement. Secondly, the reaction was inhibited by ascorbate, which suggests that the quinone must first dissociate from the enzyme, where an enzyme is used for oxidation, before the rearrangement occurs. Thirdly, the rearrangement was effected by non-enzymic oxidizing agents.

The significance of these observations is potentially farreaching. Our oxidation studies suggest that the yield of α , β dehydroDOPA moieties from DOPA analogues can be quite high. Side reactions, such as the inter- and intramolecular Michael-type addition reactions occurring when DOPA or dopamine are oxidized, were avoided by using only N-acetylated DOPA analogues, (or, where appropriate, substrates lacking a free amine group). These data and preliminary observation on synthetic DOPA-containing decapeptides



- A N-ACETYLDOPAETHYLESTER
- **B** N-ACETYLDOPAQUINONE ETHYLESTER
- C N-ACETYLTATSUIC ACID ETHYLESTER
- D N-ACETYLQUINONYLTATSUIC ACID ETHYLESTER
- I CATECHOLOXIDASE (CATECHOL:02 OXIDOREDUCTASE)
- II TAUTOMERIZATION REARRANGEMENT
- III CATECHOLOXIDASE

suggest that perhaps 50% of oxidized DOPA moieties (i.e. DOPA quinones) might undergo this rearrangement under appropriate conditions. Corrigently, since the rearrangement appears to be spontaneous, oxidized DOPA residues in adhesive proteins may yield significant amounts of α , β -dehydroDOPA, or derived substances. Furthermore, HPLC analyses of hydrolysates of mussel byssus, which contains much DOPA protein, gave results consistent with the presence of large amounts of byssal non-DOPA catecholic moieties similar in nature to α , β -dehydroDOPA.

Although the presence of these novel oxidation products of DOPA residues in natural adhesive peptides has not yet been unambiguously confirmed, nevertheless, current evidence strongly suggests that they exist. Since enamines, which are very reactive molecules, participate in reactions quite different from those of quinones, the nature of the hardening process in bioadhesives containing o-quinonyl enamine residues may be radically different from that hitherto anticipated, and may have important consequences for the structure and properties of cured adhesive.

INVENTIONS - one

PUBLICATIONS

A. This program:

Waite, J. H. (1986) Mussel glue from <u>Mytilus</u> <u>californianus</u>: a comparative study. J. Comp. Physiol. B. <u>156</u>, 491-496.

Mascolo, J. M. & Waite, J. H. (1986) Protein gradients in byssal threads of some marine bivalve molluscs. J. Exp. Zool. 240, 1-7.

Marumo, K. & Waite, J. H. (1987) Prolyl 4-hydroxylase in the foot of the marine mussel <u>Mytilus edulis</u> purification and characterization. J. Exp. Zool. <u>244</u>, 365-374.

Waite, J. H. (1987) Nature's underwater adhesive specialist. Int. J. Adhes. and Adhesives 7, 9-15.

Waite, J. H. & Rice-Ficht, A.C. (1987) Presclerotized eggshell protein from the liver fluke <u>Fasciola hepatica</u>. Biochem. <u>26</u>, 7819-7825.

Williams, T., Marumo, K., Waite, J. H. & Henken, R. (1988) Mussel glue has an open, extended conformation. ABB, in press.

Submitted Manuscripts

- Waite, J. H. & Rice-Ficht, A. C. (1988) A histidine-rich protein from the vitellarial cells of the liver fluke, <u>Fasciola hepatica</u>. To: J. Biol. Chem.
- Waite, J. H., Hansen, D. & Little, K. T. (1988) Adhesive protein of ribbed mussels: a natural glue with some features of collagen. To: J. Biol. Chem.
- Rzepecki, L., Nagafuchi, T. and Waite, J. H. (1988) Oxidative synthesis of α,β-dehydroDOPA derivatives and their potential involvement in auto-tanning of DOPAcontaining bioadhesive proteins. To: J. Biol. Chem.

Nagafuchi, T. and Waite, J. H. (1988) Adsorption of mussel adhesive protein to glass and polypropylene. To: Biomaterials.

TRAINING ACTIVITIES

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Bernardo Estupiñan and Karen Long received their M.S. degrees in the summer of 1988. Thesis titles were: "Study of polyphenolic protein as inducing agent in the settlement of <u>Mytilus edulis</u> larvae", and "Hemolytic toxins of two marine jellyfish", respectively.

Tatsuhiko Nagafuchi has completed two years of postdoctoral research and is now assistant professor of orthopaedics at Jikei University, Tokyo.

Five students are currently working on this research: Douglas Hansen, Stefan Samulewicz, Luke Huggins, and Kathy Little. Postdoctoral fellow: Leszek Rzepecki.

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Women/minorities - 1
noncitizen - 1 (Scotland)
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Table 1. Summary of consensus sequences from various Dopa-containing proteins in nature. Symbols used for Dopa and hydroxyproline are YOH and POH, respectively.

Species	Mr(kD)	pI	Sequences	#Repeats
PHYLUM MOLLUSCA				
Mytilus edulis byssal adhesive	130	>10	AKP(OH)SY(OH)POHPC AKPTY(OH)K	нттонк>60 13
M. californianus byssal adhesive	85	>10	ISYOHPOHPOHTYOHKPC	NHK 15
Geukensia demissa byssal adhesive	1 30	8.2	QTGYOH X1X2GY(OH)K A/TGYOHX1X2GY(OH)K X1 = hydrophobic a X2 = pro; hyp	50 >50 10 Mino acid
Aulocomya ater byssal adhesive	120	9.0	AGGYOHG X K(OH) X = hydrophobic, b	>20 pranched chair
PHYLUM ANNELIDA				
Phragmatopoma californica cement	35	8.4	X GGYOHGYOHGAK X = hydrophobic, branch chain	>20
PHYLUM PLATYHELMINTH	ES			
Fasciola hepatica eggshell precursor	31 10	7.2 6.6	GGGYOHXıX₂YOH G K GHGYOH	? 6

X1X2 = Gly, Ser, Asp

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