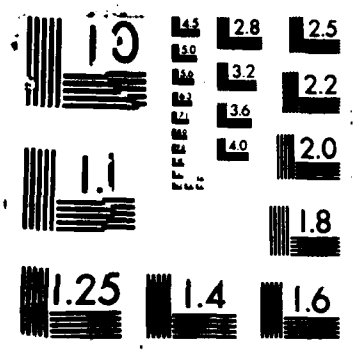
AD-A181 691 SYNTHESIS OF NUCLEOSIDE MONO- AND DIALDEHYDES AS ANTIVIRAL AGENTS(U) ROCHESTER INST OF TECH NY J P NEENAN 15 DEC 86 DAMD17-86-C-6002 1/1

UNCLASSIFIED F/G 6/15 NL


2.8
2.5
3.2
2.2
3.6
2.0
4.0
1.8
1.25
1.4
1.6

AD _____

SYNTHESIS OF NUCLEOSIDE MONO- AND DIALDEHYDES AS ANTIVIRAL AGENTS

Annual Report

John P. Neenan, Ph.D.

15 December 1986

Supported by

U.S. ARMY MEDICAL RESEARCH AND DEVELOPMENT COMMAND
Fort Detrick, Frederick, Maryland 21701-5012

Contract No. DAMD17-86-C-6002

Rochester Institute of Technology
Rochester, New York 14623

UNCLASSIFIED

SECURITY CLASSIFICATION OF THIS PAGE

REPORT DOCUMENTATION PAGE

1a. REPORT SECURITY CLASSIFICATION UNCLASSIFIED	1b. RESTRICTIVE MARKINGS
2a. SECURITY CLASSIFICATION AUTHORITY	3. DISTRIBUTION / AVAILABILITY OF REPORT Approved for public release; distribution unlimited
2b. DECLASSIFICATION / DOWNGRADING SCHEDULE	
4. PERFORMING ORGANIZATION REPORT NUMBER(S)	5. MONITORING ORGANIZATION REPORT NUMBER(S)
6a. NAME OF PERFORMING ORGANIZATION Rochester Institute of Technology 6b. OFFICE SYMBOL (If applicable)	7a. NAME OF MONITORING ORGANIZATION
6c. ADDRESS (City, State, and ZIP Code) Rochester, NY 14623	7b. ADDRESS (City, State, and ZIP Code)
8a. NAME OF FUNDING / SPONSORING ORGANIZATION U.S. Army Medical Research And Development Command 8b. OFFICE SYMBOL (If applicable)	9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER Contract No. DAMD17-86-C-6002
8c. ADDRESS (City, State, and ZIP Code) Fort Detrick Frederick, MD 21701-5012	10. SOURCE OF FUNDING NUMBERS PROGRAM ELEMENT NO. 62770A; PROJECT NO. 3M162. 770A871; TASK NO. AH; WORK UNIT ACCESSION NO. 374

11. TITLE (Include Security Classification)

Synthesis of Nucleoside Mono- and Dialdehydes as Antiviral Agents (U)

12. PERSONAL AUTHOR(S)
John P. Neenan, Ph.D.

13a. TYPE OF REPORT	13b. TIME COVERED	14. DATE OF REPORT (Year, Month, Day)	15. PAGE COUNT
Annual	FROM 11-15-85 TO 11-14-86	1986 December 15	

16. SUPPLEMENTARY NOTATION

17. COSATI CODES		
FIELD	GROUP	SUB-GROUP
06	01	
07	03	

18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number)
NUCLEOSIDE DIALDEHYDES
ANTIVIRAL AGENTS

19. ABSTRACT (Continue on reverse if necessary and identify by block number)

Seven nucleoside 2', 3'-dialdehydes were synthesized by periodic oxidation of inosine, 6-methylmercaptopurine riboside, thymine riboside, guanosine, 5'-fluoro-5'-deoxyadenosine, 8-bromoadenosine, and N^6,N^6-dimethylaminopurine riboside, respectively. The resultant nucleoside dialdehydes, as well as the intermediate 5'-fluoro-5'deoxyadenosine, were submitted to the USAMRIID antiviral screening program. Antiviral screening is still in progress as of this date. Preparation of the 4', 5'-unsaturated derivative of adenosine dialdehyde is in progress as of this date.

20. DISTRIBUTION / AVAILABILITY OF ABSTRACT	21. ABSTRACT SECURITY CLASSIFICATION
☐ UNCLASSIFIED/UNLIMITED ☒ SAME AS RPT. ☐ DTIC USERS	UNCLASSIFIED
22a. NAME OF RESPONSIBLE INDIVIDUAL Virginia M. Miller	22b. TELEPHONE (Include Area Code) (301) 663-7325 22c. OFFICE SYMBOL SGRD-RMI-S

DD FORM 1473, 84 MAR — 83 APR edition may be used until exhausted. All other editions are obsolete.

SECURITY CLASSIFICATION OF THIS PAGE

UNCLASSIFIED

Summary

Prior to the start of this contract, a number of nucleoside 2',3'-dialdehydes were found to have in vitro activity against vaccinia virus, a DNA virus; as well as against RNA viruses of the arena-, bunya-, rhabdo-, and togaviridae families. Based on these findings, the first year of this project was devoted to the synthesis of nucleoside dialdehydes by periodate oxidation of the following starting compounds: inosine, 6-methylmercaptopurine riboside, thymine riboside, guanosine, 5'-fluoro-5'-deoxyadenosine, 8-bromoadenosine, and $\underline{N}^6$, $\underline{N}^6$-dimethylaminopurine riboside. The resultant nucleoside dialdehydes, as well as the intermediate 5'-fluoro-5'-deoxyadenosine, were characterized and submitted to the antiviral screening program at USAMRIID. Work is currently underway on the preparation of the 4',5'-unsaturated derivative of adenosine dialdehyde. This derivative is intended to serve as a precursor to nucleoside monoaldehydes. The limited antiviral screening results obtained thus far on the eight compounds submitted during the first year of this project will also be discussed.

Foreword

Citations of commercial organizations and trade names in this report do not constitute an official Department of the Army endorsement or approval of the products or services of these organizations.

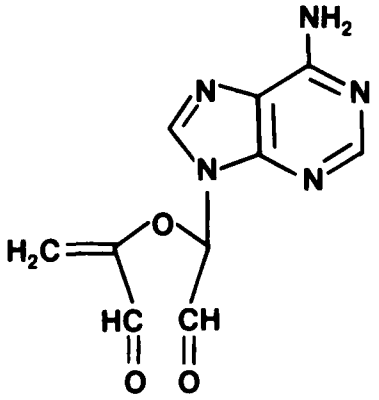
Table of Contents

Background

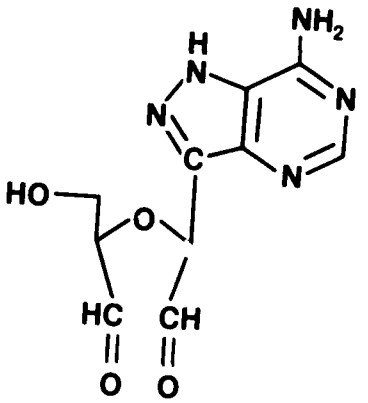
Prior to the start of this contract, a number of nucleoside dialdehydes had been found to have antiviral activity. Keller and Borchardt reported inhibition of vaccinia virus in vitro by adenosine dialdehyde.[1] The antiviral activity of adenosine dialdehyde has been attributed to its potent inhibition of the enzyme S-adenosyl-L-homocysteine (AdoHcy) hydrolase, with resultant blockage of methylation of the 5'-cap of viral mRNA[1-4]. Neenan et al. more recently reported in vitro inhibition of a number of RNA viruses in the USAMRIID screening system by adenosine dialdehyde and other nucleoside dialdehydes.[5] Based on these results the series of nucleoside dialdehydes reported herein was synthesized and submitted to the antiviral screening system during this past year (Table I).

Chemistry

Compounds 1-4, 7 and 9 were prepared by periodic acid oxidation[6] of: inosine, 6-methylmercaptopurine riboside, thymine riboside, guanosine, 8-bromoadenosine and N^6,N^6-dimethylaminopurine riboside, respectively. The requisite nucleoside starting materials were all obtained commercially. Compound 5 was obtained by periodic acid oxidation of 5'-fluoro- 5'deoxyadenosine (6), which had previously been synthesized as a candidate antiviral in a lengthly procedure described by Shen et al.[7] Instead, we prepared 6 in one step from commercially obtained 5'-O-tosyladenosine by method of Kowollik et al.[8] The synthesis of the 4'5'-unsaturated derivative (8) of adenosine dialdehyde, as previously described by Grant and Lehrner[9] is currently underway. An attempt to prepare the dialdehyde derivative of formycin A (10) was unsuccessful.

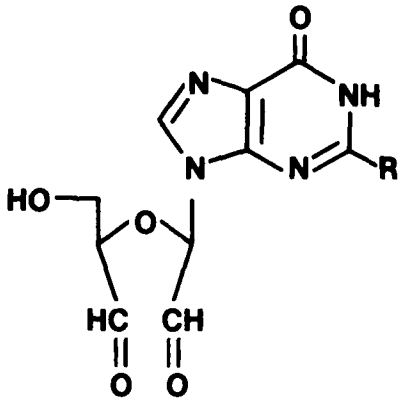


8

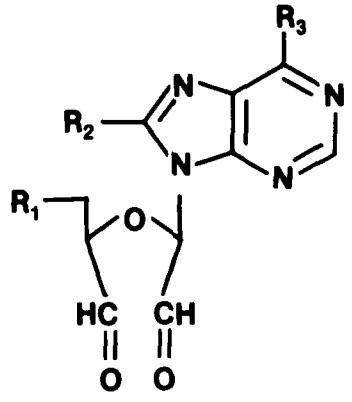


10

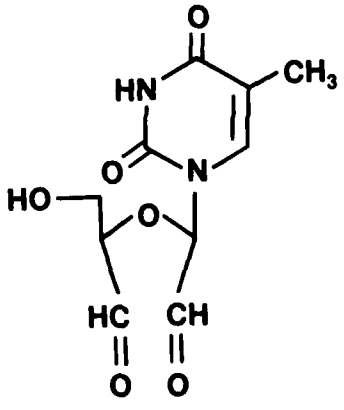
Table I. Structures of Compounds Submitted During Reporting Period



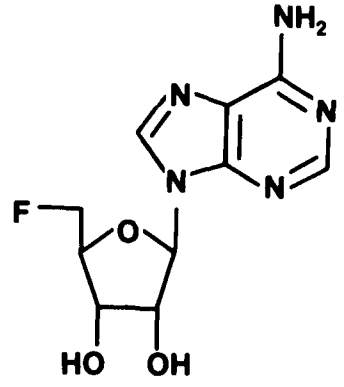
1, R = H
4, R = NH_2



2, R_1 = OH, R_2 = H, R_3 = SCH_3
5, R_1 = F, R_2 = H, R_3 = NH_2
7, R_1 = OH, R_2 = Br, R_3 = NH_2
9, R_1 = OH, R_2 = H, R_3 = $N(CH_3)_2$



3



6

Compound	Code No.	AVS No.	Name
1	OP-I-19	1915	Inosine-2',3'-dialdehyde
2	OP-I-9	1970	6-Methylmercaptopurine riboside-2',3'-dialdehyde
3	OP-I-25	1976	Thymine riboside-2',3'-dialdehyde
4	JDG-I-18	2151	Guanosine-2',3'-dialdehyde
5	OP-I-77	2275	5'-Fluoro-5'-deoxyadenosine-2',3'-dialdehyde
6	OP-I-63	2273	5'-Fluoro-5'-deoxyadenosine
7	OP-I-71	2274	8-Bromoadenosine-2',3'-dialdehyde
9	LME-I-39	2543	N^6,N^6-Dimethylaminopurine riboside-2',3'-dialdehyde

Biological Results.

As of this report date the only compound that the author has received any antiviral screening data on is compound 3 (AVS 1976). Compound 3 was essentially inactive against vesicular stomatitis virus in L929 cells.

Conclusions and Recommendations.

Once further antiviral data is obtained on the compounds already submitted, conclusions can perhaps be drawn with respect to future target compounds. Nevertheless, recommendations for next year include:

1. Completion of synthesis of compound 8. Once compound 8 is in hand, we will attempt to convert some of it reductively to the 2'- and/or 3'-monoaldehydes. Compound 8 and its monoaldehyde deriviatives might be good transition state analogs for the antiviral target enzyme AdoHcy hydrolase.

2. Synthesis of the dialdehyde derivative of tubercidin.

3. Submission of coumound 3 to the AIDS screening system. Compound 3 is somewhat similar in structure to azidothymidine (AZT), which is currently being used for treatment of AIDS. Preliminary work in our laboratory (at Rochester) indicates that 3 can inhibit reverse transcriptase.

4. Selection of other target compounds based on antiviral screening results obtained with compounds 1-9 in this report.

Experimental Section

IR spectra were recorded on a Perkin Elmer 681 Infrared Spectrophotometer. ^1H NMR spectra were recorded on a Hitachi Perkin Elmer R-600 High Resolution NMR Spectrometer. Chemical shift values are reported in δ relative to Me_4Si. UV spectra were recorded on a Varian Cary 219 Spectrophotometer. Mass spectra were obtained in the electron impact mode using a direct insertion probe and recorded on a Hewlett Packard 5995 Gas Chromatograph/Mass Spectrometer. Elemental analyses were performed by Galbraith Laboratories, Knoxville, TN.

Inosine-2',3'-dialdehyde (1); WR 220 078; NSC 118994

Compound 1 was prepared by modification of the method of Dvonch *et al*[6]. To a stirred solution of 3.0 g (11.2 mmol) of inosine in 130 mL of water was added 2.81 g (12.3 mmol) of paraperiodic acid (H_5IO_6). The reaction mixture was kept in the dark for 50 min and then applied to a 1.4 x 18 cm column of AG 1-X8 anion exchange resin (acetate form). The column was washed with water. The self-eluate and washings (320 mL) were found to be free of iodate and periodate by starch iodide test paper and lyophilized to give 2.68 g (84%) of inosine dialdehyde as a flocculent white powder.

Physical and Analytical Data

Melting Point: Dec. > 205°, shrinking at 185°.

Analysis: For $C_{10}H_{10}N_4O_5 \cdot H_2O$ (284.24)

	Calcd	Found
C	42.26	42.34
H	4.26	4.60
N	19.71	19.12

Infrared Spectrum:

KBr. Compatible with structure. Broad band at 1100 cm^{-1} typical of nucleoside dialdehydes.[10]

NMR:

(D_2O) δ 8.70-8.23 (m, 2 H, H-2 and H-8), 6.20-5.07 (several overlapping m, 3 H, H-1', H-2', H-3'), 4.35-3.64 (several overlapping m, 3 H, H-4', 2 H-5').

UV Spectrum: λmax (H_2O) 248 nm (ε 12,880)

Thin-layer Chromatography: (Eastman 13254 Cellulose)

Eluent	R_f	Comment
H_2O	0.85	Homogeneous
EtOH-1M NH_4OAc (7:3,v/v)	0.73	Homogeneous

Code No.: OP-I-19

Prepared By: S. M. Opitz

Materials:

Inosine	Sigma lot 34F-0656
Paraperiodic acid	Sigma lot 34F-0351
Anion exchange resin	Bio-Rad AG 1-X8 (acetate) Control no. 28817
Water	double distilled

6-Methylmercaptopurine riboside-2',3'-dialdehyde (2).

Compound 2 was prepared by modification of the method of Dvonch, et al.[6] To a stirred suspension of 2.9 g (9.72 mmol) of 6-methylmercaptopurine riboside in 140 mL of water was added a solution of 2.44 g (10.69 mmol) of paraperiodic acid in a few mL of water. The reaction mixture was stirred in the dark for 45 min. and then applied to a 1.4 X 17.5 cm column of Bio-Rad AG 1-X8 anion exchange resin (acetate form). The column was washed with water. The self-elulate and washings (350 mL) were found to be free of iodate and periodate by starch iodide test paper and lyophilized twice to give 2.5 g (87%) of 6-methylmercaptopurine riboside-2',3'-dialdehyde.

Physical and Analytical Data

Melting Point: 175° (dec), starts shrinking at 120°.

Analysis: $C_{11}H_{12}N_4O_4S \cdot H_2O$ (314.33)

	Calcd	Found
C	42.04	42.30
H	4.49	4.83
N	17.83	17.69

IR Spectrum:

KBr. Compatible with structure. Broad band at 1100 cm^{-1} typical of nucleoside dialdehydes.[10]

NMR:

(D_2O) δ 8.94-8.30 (m, 2 H, H-2 and H-8), 6.37-5.10 (several overlapping m, 3 H, H-1', H-2', H-3'), 4.60-3.60 (several overlapping m, 3 H, H-4', 2 H-5'), 2.58 (s, 3 H, $-SCH_3$).

UV Spectrum: λmax (H_2O) 291 nm (ε 19,760)

Thin-Layer Chromatography: (Eastman 13254 Cellulose)

Eluent	R_f	Comment
H_2O	0.78	Homogeneous
EtOH-1M NH_4OAc (7:3)	0.94	Homogeneous

Code No.: OP-I-9

Prepared By: S. M. Opitz

Materials:

6-Methylmercaptopurine riboside	Sigma lot 44F-0496
Periodic acid	Sigma lot AG1-X8
AG 1-X8 resin (acetate form)	Bio-Rad control no. 28817
water	triple distilled

Thymine riboside-2',3'-dialdehyde (3)

Compound 3 was prepared by modification of the method of Dvonch et al.[6] To a stirred solution of 2.35 g (9.10 mmol) of thymine riboside in 100 mL of water was added a solution of 2.28 g (10.01 mmol) of paraperiodic acid in a few mL of water. The reaction mixture was stirred in the dark for 1.5 hr and then applied to a 1.4 X 18 cm column of Bio-Rad AG 1-X8 anion exchange resin (acetate form). The column was washed with water. The self-eluate and washings (310 mL) were found to be free of iodate and periodate by starch iodide test paper and lyphoilized twice to give 2.27 g (90%) of thymine riboside-2',3'-dialdehyde.

Physical and Analytical Data

Melting Point: 140° starts shrinking
158° opaque melt
180° transparent

Analysis: For $C_{10}H_{12}N_2O_6 \cdot 1\frac{1}{4}H_2O$ (278.75)

	Calcd	Found
C	43.08	43.22
H	5.25	5.42
N	10.05	10.03

IR Spectrum:

KBr. Compatible with structure. Broad band at 1100 cm^{-1} typical of nucleoside dialdehydes.[10]

NMR:

D_2O δ 7.86-7.46 (broad, 1 H, H-6), 6.06-4.99 (several overlapping m, 3 H, H-1', H-2', H-3'), 4.46-3.36 (several overlapping m, 3 H, H-4', 2 H-5'), 1.9 (s, 3 H, 5-CH_3).

UV Spectrum: λmax (H_2O) 264 nm (ε 9,760)

Thin-Layer Chromatography: (Eastman 13254 Cellulose)

Eluent	R_f	Comment
H_2O	0.91	Homogeneous
EtOH-1M NH_4OAc (7:3)	0.87	Homogeneous

Code No.: OP-I-25

Prepared by: S. M. Opitz

Materials:

Thymine riboside	Behring Diagnostics lots 91045 and 710162
Periodic acid	Sigma lot 34F-0351
AG 1-X8 resin (acetate form)	Bio-Rad control no. 28817
Water	triple distilled

Guanosine-2',3'-dialdehyde (4)

Compound 4 was prepared by modification of the method of Johnson et al.[11] To a stirred suspension of 5.7 g (20 mmol) of guanosine in 200 mL of water was added 5.02 g (22 mmol) of periodic acid. The reaction mixture was stirred in the dark at room temperature for 60 min. A 10 g portion of Bio-Rad AG 1-X8 anion exchange resin (acetate form) was then added to the reaction mixture. The resulting slurry was stirred briefly and then allowed to stand for one min. The resin was removed by vacuum filtration, and the procedure was repeated until a total of six 10 g portions of resin has been used and the filtrate was free of iodate and periodate by starch iodide test paper. A 20 g portion of Bio-Rad AG 11-A8 ion retardation resin was then added to the filtrate, slurried for one minute, and then removed by vacuum filtration. The filtrate was lyophilized to give 4.06 g of white powder, which was redissolved in 250 ml of water and treated with 30 g of ion retardation resin, followed by vacuum filtration. The filtrate was lyophilized to give 3.89 g (51%) of guanosine-2',3'-dialdehyde.

Physical and Analytical Data

Melting Point: 150° (dec)

Analysis: $C_{10}H_{11}N_5O_5 \cdot 2^3/_4H_2O \cdot 1^1/_2CH_3CO_2H$ (360.80)

	Calcd	Found
C	36.62	36.42
H	5.17	4.67
N	19.41	19.55

IR Spectrum:

KBr. Compatible with structure. Broad band at 1100 cm^{-1} typical of nucleoside dialdehydes.[10]

NMR:

Could not obtain. Insoluble in deuterated DMSO; forms gel in D_2O. Only acetate protons were detected at δ 1.9. See ref. 10.

UV Spectrum: λmax (H_2O) 252.5 nm (ε 13,375)

Thin-Layer Chromatography:

Eluent	R_f	Comment
H_2O	0.79	Homogeneous
EtOH-1M NH_4OAc (7:3)	0.80	Homogeneous

Code No.: JDG-I-18

Prepared by: J. D. Grinnell

Materials:

Guanosine	Sigma lot 34F-0198
Periodic Acid	Sigma lot 34F-03511
AG 1-X8 resin (acetate form)	Bio-Rad Control No. 28817
AG 11-A8 resin	Bio-Rad Control No. 29210
Water	Triple distilled

5'-Fluoro-5'-deoxyadenosine-2',3'-dialdehyde (5)

To a stirred suspension of 2.5g (8.96 mmol) of compound 6 in 150 mL of water was added 2.3g (9.86 mmol) of periodic acid. The reaction mixture was stirred in the dark for 1.5 hr and then applied to a 1.4 X 18 cm column of Bio-Rad AG 1-X8 anion exchange resin (acetate form). The column was washed with water. The self eluate and washings (450 mL) were found to be free of iodate and periodate by starch iodide test paper and lyophilized twice to give 2.45g (90%) of compound 5.

Physical and Analytical Data

Melting Point: > 187° dec

Analysis: For $C_{10}H_{10}FN_5O_3 \cdot 2H_2O$ (303.25)

	Calcd	Found
C	39.61	39.42
H	4.65	5.05
N	23.09	22.86

IR Spectrum:

KBr. Compatible with structure. Broad band at 1100 cm^{-1} typical of nucleoside dialdehydes.[10]

NMR:

(Me_2SO-d_6) δ 8.70-7.90 (2 br s, 2 H, H-2 and H-8), 7.70-6.80 (m, 2 H, 2 H-6'), 6.10-3.90 (several overlapping m, 6 H, H-1', H-2', H-3', H-4', 2 H-5').

UV Spectrum: λmax (H_2O 258 nm (ε 15,900).

Thin-Layer Chromatography: (Eastman 13254 cellulose)

Eluent	R_f	Comment
H_2O	0.62	Homogeneous
EtOH-1M NH_4OAc (7:3)	0.75	Homogeneous

Code No.: OP-I-77

Prepared by: S. M. Opitz

Materials:

Periodic acid	Sigma lot 34F-0351
AG 1-X8 resin (acetate form)	Bio-Rad Control No. 28817
Water	triple distilled

5'-Fluoro-5'-deoxyadenosine (6)

Prepared by modification of the general method of Kowollik et al.[8] A stirred solution of 5'-tosyladenosine (5.4 g, 12.8 mmol) in a 1.0 M solution of tetrabutylammonium fluoride (80 mL, 80 mmol) in THF was heated in an oil bath at 48-51° for 25 hr. The solvent was removed on a rotary evaporator. The residue was dissolved in water (500 mL) and applied to a 2.5 X 1.9 cm column of Bio-Rad AG 50W-X8 resin (hydrogen form). The column was washed with water until the eluate was neutral (pH paper). The resin and its contents were transferred to a beaker, stirred with 2M ammonumium hydroxide in a warm bath (40-45°) for a few min. The resin was removed by vacuum filtration, and repeatedly washed with 2 M ammonium hydroxide until the eluate showed no UV absorption. The combined eluate was lyophilized and to give a yellow powder which was dissolved in warm H_2O and kept at 4° overnight. The resultant precipitate was collected by filtration, washed with water and dried under suction. The crude product (2.45 g) was dissolved in 75 mL acetonitrile-water (11:89) and purified on a Waters Prep LC 500A system (column: 57 mm x 30 cm C_{18} cartridge; mobile phase: 11 vol% acetonitrile in water; flow rate: 100 mL/min; detector: refractive index). Solvents were removed by rotary evaporation under high vacuum to a small volume, followed by lyophilization to give 1.6 g (46%) of compound 6.

Physical and Analytical Data

Melting Point: 204-205° dec.

Analysis: For $C_{10}H_{12}FN_5O_3 \cdot \frac{1}{2}H_2O$ (273.74).

	Calcd	Found
C	43.87	43.77
H	4.60	4.67
N	25.58	25.96

IR Spectrum: KBr. Compatible with structure.

NMR:

(Me_2SO-d_6) δ 8.27 (s, 1 H, H-8), 8.15 (s, 1 H, H-2), 5.95-5.92 (d, 1 H, H-1'), 5.63-5.40 (2 d, 2 H, OH-3' and OH-2'), 4.77-4.51 (m, 3 H, H-2' and 2 H-5', $J_{H-5'}$, F = 50 Hz), 4.29-4.04 (m, 2 H, H-3' and H-4', $J_{H-4'}$, F = 15 Hz).

UV Spectrum: λmax (H_2O) 258 nm (ε 14,670)

Thin-Layer Chromatography: (Eastman 13254 cellulose)

Eluent	R_f	Comment
H_2O	0.48	Homogeneous
EtOH-1M NH_4OA_c (7:3)	0.65	Homogeneous

Code No.: OP-I-63

Prepared By: S. M. Opitz

Materials:

5'-Tosyladenosine	Sigma lot 120F-4032
Tetrabutylammonium fluoride (1.0 M in THF)	Aldrich lot 00820BP
AG 50W-X8 resin, 50-100 mesh (hydrogen form)	Bio-Rad control. no. 29996

8-Bromoadenosine-2'3'-dialdehyde (7)

To a stirred suspension of 2.90 g (8.40 mmol of 8-bromoadenosine in 100 mL of water was added 2.11 g (9.24 mmol) of periodic acid. The reaction mixture was stirred in the dark for 1.5 hr and then applied to a 1.4 X 18 cm column of Bio-Rad AG 1-X8 resin (acetate form). The column was washed with water. The self-eluate and washings (400 mL were found to be free of iodate and periodate by starch iodide test paper, and lyophilized three times to give 2.77g (87%) of compound 7.

Physical and Analytical Data

Melting Point: >177° dec

Analysis: For $C_{10}H_{10}BrN_5O_4 \cdot 2H_2O$

	Calcd	Found
C	31.60	31.92
H	3.71	3.82
N	18.42	18.35

IR Spectrum:

KBr. Compatible with structure. Broad band at 1100 cm^{-1} typical of nucleoside dialdehydes.[10]

NMR:

(Me_2SO-d_6/D_2O) δ 8.17 (s, 1 H, H-2), 6.40-4.60 (several overlapping m, 4 H, H-1', H-2', H-3', H-4'), 3.6 (broad, 2 H, 2 H-5').

UV Spectrum: λmax (H_2O) 265 nm (ε 17,320)

Thin-Layer Chromatography: (Eastman 13254 cellulose)

Eluent	R_f	Comment
H_2O	0.61	Homogeneous
EtOH-1M NH_4OAc (7:3)	0.79	Homogeneous

Code No.: OP-I-71

Prepared By: S. M. Opitz

Materials:

8-Bromoadenosine	Sigma lot 84F-0602
Periodic acid	Sigma lot 34F-0351
AG 1-X8 resin (acetate form)	Bio-Rad control no. 28817

N^6,N^6-Dimethylaminopurine riboside-2',3'-dialdehyde (9).

To a stirred suspension of 2.4 g (8.13 mmoles) of N^6,N^6-dimethylaminopurine riboside in 100 mL of water was added 2.04 g (8.95 mmoles) of paraperiodic acid. The reaction mixture was stirred in the dark for 2-1/4 hours and then applied to a 1.4 cm x 16.24 cm column of Bio-Rad AG 1-X8 anion exchange resin (acetate form). The column was washed with water. The self-elute and washings (300 mL) were found to be free of iodate and periodate by starch iodide test paper, and were then lyophilized twice to give 2.19 grams (92%) of N^6,N^6dimethylaminopurine riboside-2',3'-dialdehyde.

Physical and Analytical Data

Melting Point: 170° (dec), starts shrinking at 120°.

Analysis: For $C_{12}H_{15}N_5O_4 \cdot 2H_2O$ (329.31)

	Calcd	Found
C	43.77	44.10
H	5.82	5.42
N	21.2	20.70

IR Spectrum:

KBr. Broad band at 1100 cm^{-1} typical of nucleoside dialdehydes[10].

NMR (D_2O):

δ8.53-8.00 (m, 2 H, H-2, H-8), 6.12-3.94 (several overlapping m, 3 H, H-1', H-2', H-3'), 4.74-3.53 (several overlapping m, 3 H, H-4', 2 H-5'), 3.48-3.24 (t, 6 H, CH_3 dimethylamino group).

Melting point (°C):

127° melts, opaque
170° decomposes into yellow liquid
>170° brown, charred solid

UV Spectrum: λmax (H_2O) 274 nm (ϵ 20,895)

Mass Spectrum (EI, methanol)

m/e 293 (M^+), 275 ($M^+ - H_2O$), 264 ($M^+ - CHO$)
246 [M - (H_2O + CHO)], 164 (N^6,N^6-dimethyladenine H^+)

Thin-layer Chromatography: (Eastman 13254 cellulose)

Eluent	RF
H_2O	0.77
EtOH-1 M NH_4OAc (7:3)	1.0

Code No.: LME-I-39

Prepared by: L.M. Eckel

Materials:

N^6,N^6-dimethylaminopurine riboside	Sigma lot 88C-0410
paraperiodic acid	Sigma lot 34F-0351
anion exchange resin (acetate form)	Bio-Rad AG 1-X8 control #28817

Literature Cited

1. B. T. Keller and R. T. Borchardt, Fed. Proc., Fed. Am. Soc. Exp. Biol., 42, Abstr. 2230 (1983).

2. J. L. Hoffman, "Transmethylation"; E. Usdin, R. T. Borchardt, and C. R. Creveling, Eds.; Elsevier/North Holland: New York, 1979, pp. 181-186.

3. J. L. Hoffman, Arch. Biochem. Biophys., 205, 132 (1980).

4. D. M. Houston, E. K. Dolence, B. T. Keller, U. Patel-Thombre, and R. T. Borchardt, J. Med. Chem., 28, 471 (1985).

5. J. P. Neenan, S. M. Opitz, R. M. Borges, J. D. Grinnell, P. G. Canonico, and M. A. Ussery, "Abstracts of Papers", 192nd National Meeting of the American Chemical Society, Anaheim, CA, September, 1986; American Chemical Society: Washington, D.C., 1986; MEDI 87.

6. W. Dvonch, H. Fletcher, III, F. J. Gregory, E-M. H. Healy, G. H. Warren, H. E. Alburn, Cancer Res., 26, 2386 (1966).

7. T-Y. Shen, W. V. Ruyle, T. Neilson, U.S. Patent 3,575,959 (to Merck & Co.), April 20, 1971.

8. G. Kowollik, K. Gaertner, G. Etzold, and P. Langen, Carbohyr. Res., 12, 301 (1970).

9. A. J. Grant and L. M. Lerner, J. Med. Chem., 23, 795 (1980).

10. F. Hansske and F. Cramer, Carbohydr. Res., 54, 75 (1977).

11. L. A. Johnson, R. B. Gordon, and B. T. Emmerson, Biochem. Med., 22, 33 (1979).

DISTRIBUTION LIST

5 copies	Commander US Army Medical Research Institute of Infectious Diseases ATTN: SGRD-UIZ-M Fort Detrick, Frederick, MD 21701-5011
1 copy	Commander US Army Medical Research and Development Command ATTN: SGRD-RMI-S Fort Detrick, Frederick, Maryland 21701-5012
2 copies	Defense Technical Information Center (DTIC) ATTN: DTIC-DDAC Cameron Station Alexandria, VA 22304-6145
1 copy	Dean School of Medicine Uniformed Services University of the Health Services 4301 Jones Bridge Road Bethesda, MD 20814-4799
1 copy	Commandant Academy of Health Sciences, US Army ATTN: AHS-CDM Fort Sam Houston, TX 78234-6100

END

7-87

DTIC