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FINAL REPORT
OF
THE NATURE OF AIRBORNE PARTICULATES
AT TROPIC EXPOSURE SITES

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Airborne particulates were collected at five exposure sites in Panama using cascade impactor air samplers. Scanning electron microscopy, energy dispersive x-ray analysis, and culture identification techniques were used to analyze the particulates. Analysis revealed that the particulates consist of silicates, chlorides, and sulfur-rich and phosphorus-rich particles. Atmospheric particle levels were higher in the dry season than in the rainy season, and the predominant fungal species varied at each exposure site. The		

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open direct exposure of culture plates served as a simple, appropriate method for monitoring atmospheric fungal spores.

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THE NATURE OF AIRBORNE PARTICULATES AT TROPIC EXPOSURE SITES

1. BACKGROUND

a. Microbial deterioration of natural polymers is a major material problem in the humid tropics. However, the role of microorganisms in the deterioration of many synthetic polymers is unclear. Polymers which are not broken down directly by microorganisms may be affected adversely by the microbial products of surface contaminant metabolism. The presence of such surface contaminants may be a primary distinguishing factor between natural environment and chamber testing. Little information is available on the nature and source of surface contaminants and their contribution to tropic materiel degradation.

b. The US Army Tropic Test Center (USATTC) first studied surface deposits, their sources, and their roles in microbiological deterioration of materials in the 1960's (reference 1). This effort centered on volatile organic materials produced by vegetation and associated microflora as the source of the organic surface deposits. These volatile organic molecules, it was hypothesized, were present in the atmosphere under the canopy in concentrations high enough that they would either condense on exposed surfaces or be used directly from the air by fungi. Much of this work was theoretical and based largely on laboratory studies.

c. A subsequent USATTC methodology investigation, TECOM Project No. 9-CO-049-000-002 (reference 2), inventoried volatile and condensed organic materials at seven sites in the Panama Canal Area. USATTC found that the volatile organic components of the air were primarily fossil fuel combustion products in low concentrations (comparable to levels in unpolluted temperate areas). USATTC did not find any volatile effluents from vegetation. USATTC traced some components of condensed materials to local vegetation sources, but the mode of transfer was not determined because the components were not volatile. These condensed materials supported fungal growth.

d. Past studies have concentrated on volatile atmospheric organics as the source of surface contaminants. Results from USATTC's second study (reference 2) suggest that the source may be atmospheric particulates rather than volatiles. This investigation focused on the nature of atmospheric particulates at USATTC test sites.

2. OBJECTIVE

Determine the nature of airborne particulates at various USATTC exposure sites and determine whether these particulates differ between the rainy and dry seasons.

3. PROCEDURES

a. Particulate Sample Collection

(1) USATTC collected airborne particulates using Misco-Sierra High

Volume Air Samplers with constant air flow controller attachments. The air flow controllers were set at a constant flow of 40 cubic feet per minute (CFM) ($1.9 \times 10^{-2} \text{m}^3/\text{sec}$). A 5-stage collector (High Volume Cascade Impactor) separated the collected particles according to size. The manufacturer provided particle size cut-off values for each stage at 50-percent collection efficiency for spherical particles at 25° C and 1 atm. pressure:

<u>Stage No.</u>	<u>Mass Median Diameter (microns) at 40 CFM</u>
1	to 7.2
2	7.2 to 3.0
3	3.0 to 1.5
4	1.5 to 0.95
5	0.95 to 0.49
High Volume Standard Filters (6)	0.49 to 0.00 (remaining particles)

(2) USATTC used both cellulose and fiberglass filters during this study. The filters were weighed before and after sampling, and the weight gain noted when the two weights were compared is considered to be the weight of the particles collected. Before each weighing, the filter papers were conditioned to laboratory humidity for at least 24 hours.

(3) A detailed sampling schedule is presented in Appendix A, table 1. Sampling times at the exposure sites were normally 24 hours, but sampling periods of less or more than 24 hours were used occasionally. The variable sampling intervals were used to determine whether or not overloading occurred during the regular 24-hour sampling periods. USATTC checked for the possibility of daytime and nighttime sampling result differences by changing filters during a 24-hour period and comparing the results.

(4) The exposure sites used for testing were the Fort Sherman Coastal Exposure Site (FSCES), Fort Sherman Open Exposure Site, MacKenzie Forest Exposure Site (MFES), Fort Clayton General Purpose Test Area (FCGPTA), and the Rodman Munition Surveillance Site (RMSS). Because electrical power outlets for the air samplers were not available at RMSS, limited sampling was done there. Most sampling was done at the FCGPTA because it is near USATTC's main installations. To determine if there are seasonal differences in atmospheric particulates, air sampling was carried out during both the rainy and dry seasons.

b. Microscopic and Energy Dispersive X-ray (EDX) Analysis of Particulates.

(1) An International Scientific Instrument Super II Scanning Electron Microscope (SEM) was used to perform the microscopic and EDX analysis. The SEM operates by focusing a high voltage electron beam on the sample, generating a secondary electron (SE) emission which is picked-up by a SE detector. The SE detector gives details of the sample's surface morphology. Simultaneously, x-rays are emitted by the surface constituents of the sample which are detected by a Si-Li drifted detector. By means of an EDX analysis, the elemental composition of the sample surface is then obtained. A Si-Li detector can detect only the x-rays from elements with an atomic number higher than 10 (i.e., sodium and higher).

(2) Three sections of about 0.8 cm² were cut from each filter paper and mounted on carbon SEM stubs with an isopropyl alcohol-base graphite glue. The samples from the high volume standard filter were randomly selected. The samples to be analyzed and a blank sample from an unexposed filter were sputter-coated together with gold and graphite. The x-ray spectrum of the blank sample was used for background correction in the analysis of the air filters' EDX spectrum.

(3) The sample was examined with the SEM at a low magnification (usually 100x) and an area representative of the sample was selected. An EDX analysis of this area was performed. The surface coverage of the area was about 5x10⁵ μm². To determine whether or not the particulate composition of this area was homogeneous, a second EDX analysis was performed on a partial section of the analyzed area. The surface coverage of the smaller section was about 1x10⁵ μm². Normally, a third EDX analysis was performed on an equally small section located elsewhere in the analyzed area. Many more EDX analyses were required when the sample was heterogeneous or contained many large particles (with diameters greater than 5 μm). The SEM spot mode was useful for the EDX analysis of large particles. This mode allowed the collection of x-rays from a selected spot on the particle surface, thus avoiding the collection of extraneous x-rays from the surrounding particle. The acquisition time for an EDX analysis was generally set for 100 seconds.

(4) At the latter stages of the test, a Robinson Back-scattered Electron (BSE) detector was used instead of the SE detector. The BSE detector allowed samples to be used without a gold coating. In the absence of the gold peak, the new normalization peak was a section of the spectrum 50 eV wide and centered at 350 KeV. This is a region where no M, L, or K x-ray peaks are found.

c. Microbiological Assay

(1) Portions (0.5 cm²) of the filter papers containing particulate samples were cut and placed in carrot-agar culture plates that were left standing at room temperature (75° F/24° C) for several days. The fungi that grew in the culture media were either identified or described when identification was not possible. As a control for the microbiological assays, portions of unexposed filter papers were placed on culture plates and incubated with the exposed samples.

(2) A second procedure, a membrane test, was used to collect particulates for microbiological assay and its results were compared to those obtained using the filter papers. This procedure is described in detail in Test Operations Procedure (TOP) 8-2-514 (reference 3) and was used mainly at FCGPTA during the final test stages. During this procedure, air was pulled through a membrane filter of 0.47 μm pore size for five minutes. Air flow was set at 11.5 liters per minute ($1.9 \times 10^{-4} \text{m}^3/\text{sec}$). A sufficient number of organisms was collected, while avoiding overcrowding on the membrane filter, in 5 minutes of sampling time (reference 3). The entire membrane filter was placed on carrot-agar medium in a petri dish, and the cultures were kept at room temperature ($75^\circ \text{F}/24^\circ \text{C}$) for several days. The fungi and bacteria that grew on the culture plates were either identified or described when identification was not possible. Unexposed membrane filters, which were placed on culture plates and incubated at the same time as the exposed membrane filters, served as controls for the microbiological assay.

(3) Carrot-agar culture plates were opened and exposed at the test sites. Generally, 1 to 2 minutes of exposure was sufficient to collect a variety of spores without overcrowding the plate. This collection method measured the amount of microorganisms deposited on surfaces at a given site. As controls, a similar number of culture plates were opened in the laboratory, an air-conditioned environment with limited particulate fall-out. This direct exposure test was done to compare the microorganisms collected by suction methods (air samplers and membranes) with those from a non-suction method (free-fall deposition).

4. RESULTS

a. Particle Collection

(1) The results for particle collection are listed in Appendix A, table 1. They are listed according to the Julian date on which the sample was collected.

(2) The particle collection weight results indicate that most particles were collected by the final filter and the second stage of the cascade impactor. Exceptions were samples collected at the FSCES, where the particles were collected largely by the first two stages of the cascade impactor.

(3) The Appendix A, table 1 results show that higher collection rates were obtained when cellulose filters were used. However, collection rates were much more variable than those obtained with fiberglass filters. In the two instances when filter papers were changed after a 6- or 8-hour operation, it was found that more particles (by weight) were collected at night than during the day. Collection rates were higher in the dry season than in the rainy season.

(4) The color of the collected particulates ranged from light brown to black.

b. Energy Dispersive X-Ray Results

(1) The EDX analysis results are listed in Appendix A, table 2. The particles were classified into four groups based on their main component. These groups were silicates, chlorides, sulfur-rich, and phosphorus-rich particles. The silicates were particles without definite form and size; they were found on all stages of the cascade impactor. Chloride particles were cubic, large, and found mainly in the first two stages of the cascade impactor. Sulfur-rich particles were spherical, small, and found mostly in the last three stages of the cascade impactor. The phosphorus-rich particles were found largely in the first two stages of the cascade impactor. These phosphorus-rich particles were found in filters sampled at the FCGPTA and the MFES.

(2) No significant differences in EDX results were observed for particles sampled during daytime when compared with those sampled at night. However, the levels of phosphorus detected were much higher for particle samples obtained during the rainy season than during the dry season.

(3) The nature of the samples was problematic during SEM examination. This problem was caused by the non-conductive nature of the filter paper and the fact that the particles were not firmly attached to the paper. Even when gold and graphite coatings were used, charging was significant. The sample's thermal expansion while under the electron beam caused particles to move and occasionally to become dislodged from the filter paper.

c. Fungal Analysis Results

(1) Appendix A, table 3 lists the fungi found from each stage. Appendix A, table 4 lists the fungi found using the membrane air sampling procedures and Appendix A, table 5 lists the fungi found in the culture plates after direct exposure. The different sampling methods provided basically similar results.

(2) The fungi identified and listed in Appendix A, tables 3 thru 5, include 11 of the 15 species found by Hutton, et al. (reference 4) in their 1968 study. The isolation and identification of every fungus species observed were not within the scope of this test. Fungi that could not be readily identified were labeled as "unknowns." A description of these unknown fungi is presented in Appendix B. The main difference in the results of Appendix A, tables 3, 4, and 5 is the number of times Fusarium was observed. Fusarium was observed much more often in the cultures from the high volume sampler and from those that were exposed directly, than in those prepared from membranes filters.

5. DISCUSSION

a. The air sampler results must be interpreted with care, especially those results relating to the weight of the particulates collected, because of the tropical atmosphere's high humidity. Hutton, et al. (reference 5) reported that samples from the first three stages had the tendency to become wet when using a four-stage cascade collector. He found that only the smallest particles were dry enough for analysis. Therefore, conditioning the collecting filters to the laboratory humidity is critical before weighing the filters.

b. More consistent collection rates were obtained with fiberglass filters than with cellulose filters. For this reason, fiberglass filters are preferable to the cellulose filters for humid tropic use. Although high collection rates were obtained sometimes using cellulose filters, they may result from the interaction of air moisture with the filter material and may not accurately reflect atmospheric particulate concentration.

c. Gauger, et al. (reference 6) also reported higher rates of particulate collection during the dry season than the rainy season. A number of environmental factors contribute to the higher atmospheric particulate content during the dry season. These factors include the following:

- (1) Increase of trade winds, which can lift particulates into the air.
- (2) The absence of rainfall, capable of washing away litter and debris from the ground, and of precipitating atmospheric particulates which could serve as condensation nuclei for the rain droplets.
- (3) Extensive dry season forest and grassland fires which produce enough particulates to cause an atmospheric haze.

d. The exhaust from the air samplers was found to affect atmospheric particulates by disturbing and lifting up litter, dust and spores which could be picked up by the samplers. The air samplers collected copper metal chips at different exposure sites, including the laboratory. These copper chips appear to be originating from the air sampler itself, when copper is blown into the atmosphere through its exhaust. The sampler collected these chips which were suspended in the air by the force of the exhaust hitting the floor.

e. Many silicon-containing particulates of different sizes and without definite forms or shapes were collected at all exposure sites. Silicon, by weight, is the second most abundant element on the earth's crust. Combined with oxygen and other elements, it forms an enormous diversity of silicate minerals. Silicon is the main component of sands and clays.

f. Also, many chloride particulates were collected at all exposure sites, especially at the FSCES. In general, these were the largest particulates collected, had a cubic shape, and were water soluble. The marine environment surrounding the 56-mile-wide Isthmus of Panama is the source of the chloride salts.

g. The sulfur-rich particulates were found at all exposure sites. These were spherical and smaller than the chloride particulates and were found mainly in the last three stages of the cascade collector. Several analyses of the detected sulfur-rich particulates were done. Our EDX analysis results indicated only the presence of sulfur. Further EDX analyses were done with windowless Si-Li detectors during the advanced SEM course at Lehigh University, Allentown, Pennsylvania. These EDX analyses showed only the presence of carbon, oxygen, and sulfur. However, the cellulose filter could be the source of the carbon and oxygen signals. The absence of nitrogen in the windowless EDX results rules out the possibility of the particulates being ammonium sulfate. Ammonium sulfate has been reported by Junge (reference 7) as the form by which aerosol sulfur would travel. The particulates were insoluble in water and in carbon disulfide, and tests for sulfates using dilute barium chloride yielded negative results. These test results suggest that our sulfur-rich particulates are small particulates of silicon or carbon absorbed with sulfur, and not sulfuric acid droplets or pure elemental sulfur. This suggestion is further supported by the facts that silicon is found on all stages and that sulfur could use it as a transport media. Carbon-bound (organic) sulfur is produced by the decomposition of organic mercaptans. The binding that occurs between sulfur and carbon/silicon can be hypothesized as electrostatic. Its surface charge would make it insoluble in carbon disulfide.

h. The phosphorus-rich particulates were without any definite structural or geometrical form and were found mostly on samples from the FCGPTA and MFES sites. This is not surprising, since both sites are basically under canopy, and biological activity is higher in forested sites than in open sites (reference 8). The higher biological activity during the rainy season can also explain the higher levels of phosphorus detected at these sites during the rainy season. EDX analysis using the external window mapping mode showed the phosphorus to be associated with particles of biological origin. These particles include seeds, pollen, debris, leaves, litter, and spores.

i. The fungi identification results in Appendix A, tables 3 through 5, indicate that the sampling techniques used in this investigation provide basically similar results. More species of fungi were observed when culture plates underwent open exposure at the sites. Thus, the simple method of exposing culture plates is considered best for monitoring atmospheric fungal contaminants.

j. Test results indicate that atmospheric particulates contain a diversity of fungal species. Almost identical species of fungi were observed in culture plates prepared with particulates collected from the different cascade collection stages. This indicates that spores can travel and can be

found as individual spores, clusters, or attached to other particles. Individual spores may detach from the clusters, but are trapped by the subsequent filter stages.

6. CONCLUSION

a. Surface contaminants have been found to be both organic and inorganic in nature. The predominant organic particulates were fungal spores and phosphorus-rich particulates. Silicates, chlorides, and sulfur-rich particles were the main inorganic particulates. Silicates were found in all size ranges separated by the cascade impactor; chlorides generally had a diameter larger than 1.5 μm ; and sulfur-particles had diameters usually between 0.5 μm and 3.0 μm .

b. Higher collection rates were obtained for dry season sampling than for rainy season sampling, and when sampling was done at night rather than during the day.

c. Precautions are needed to prevent the air sampler exhaust from causing unnaturally high levels of phosphoric particulates.

d. Fiberglass filters were found to be best suited for humid tropic sampling.

e. The simple method of open, direct exposure of culture plates was found to be the most appropriate method for collecting atmospheric fungal spores.

APPENDIX A. TABULATED TEST RESULTS

TABLE 1. INDIVIDUAL STAGE PARTICLE RESULTS, BY WEIGHT AND PERCENT

OBS SITE	C. DATE	TIME (hr)	FILTER	CRATE (mg/hr)	WT1	WT2	WT3	WT4	WT5	WT6	WTOTAL	PC1	PC2	PC3	PC4	PC5	PC6
								mg									
1	FCG 2064	7	F	12.1	56.24	9.76	0.58	13.09	0.62	4.46	84.75	66.40	11.50	0.70	15.40	0.70	5.3
2	FCG 2069	23	F	4.3	9.34	13.51	4.31	1.81	0.62	22.49	52.08	17.90	25.90	8.30	3.50	1.20	43.2
3	FCG 2068	23	F	5.4	11.03	23.09	14.34	6.91	50.46	19.10	124.93	8.80	18.50	11.50	5.50	40.40	15.3
4	FCG 2070	24	C	3.2	9.49	15.37	8.28	5.54	3.19	34.39	68.26	12.40	20.10	10.90	7.30	4.20	45.1
5	ROD 2076	3	F	22.8	11.26	16.80	5.45	2.22	0.36	32.29	66.38	16.50	24.60	7.80	3.30	0.60	47.2
6	MCK 2081	24	F	2.5	15.67	22.03	4.57	2.19	2.34	13.85	60.65	25.80	36.30	7.50	3.60	3.90	22.8
7	MCK 2082	24	C	2.6	10.09	19.87	4.96	4.13	3.18	18.97	61.20	16.50	32.50	8.10	6.70	5.20	31.0
8	MCK 2123	23	C	2.7	2.88	15.38	1.55	1.92	1.72	39.07	62.52	4.60	24.60	2.50	3.10	2.80	62.4
9	FCG 2126	21	C	13.7	45.24	53.64	41.05	35.76	28.33	84.06	288.08	15.70	18.60	14.30	12.40	9.80	29.2
10	ROD 2140	25	C	8.3	30.69	42.47	27.18	24.08	21.65	60.71	206.78	14.80	20.50	13.10	11.70	10.50	29.4
11	MCK1 2160	6	C	59.6	51.48	50.99	47.78	46.09	47.06	114.28	357.68	14.40	14.30	13.40	12.90	13.10	31.9
12	MCK2 2160	9	C	88.4	98.02	126.72	92.66	104.29	137.82	236.25	795.76	12.30	15.90	11.60	13.10	17.30	29.8
13	MCK3 2161	7	C	23.9	22.87	26.35	21.93	22.16	18.62	55.44	167.37	13.70	15.70	13.10	10.90	11.10	33.1
14	FCG1 2203	6	C	22.1	18.31	22.47	16.98	14.43	15.09	45.30	132.58	13.80	17.00	12.80	10.30	11.40	34.1
15	FCG2 2203	6	C	30.5	20.92	25.52	23.26	24.44	25.04	63.66	182.84	11.40	14.00	12.70	13.40	13.70	34.8
16	FCG3 2204	6	C	38.3	32.07	38.27	24.76	27.61	28.32	78.82	229.85	13.90	16.70	10.80	12.00	12.30	34.3
17	FCG4 2204	6	C	15.2	13.26	17.62	15.00	12.70	2.60	30.26	91.44	14.50	19.30	16.40	13.90	2.80	33.1
18	F50 2207	96	C	2.1	32.00	55.17	17.24	16.71	9.71	68.86	199.69	16.00	27.60	8.60	8.40	4.90	34.5
19	F5C 2217	24	C	16.3	130.01	79.95	28.87	23.69	23.33	105.49	391.34	33.20	20.40	7.40	6.00	6.00	42.2
20	F5D 2217	23	C	13.5	45.45	48.70	30.11	26.53	28.15	130.49	309.43	14.70	15.70	9.70	8.60	9.10	42.2
21	FCG 2363	25	C	1.8	9.10	11.12	1.17	1.60	2.71	19.07	44.77	20.33	24.84	2.61	3.57	6.05	42.6
22	F5C 3018	25	F	2.1	16.97	21.11	7.08	0.89	1.79	4.70	52.54	32.29	40.18	13.48	1.69	3.41	8.9
23	F5D 3018	25	F	4.2	5.51	5.69	20.68	12.24	4.54	57.31	106.17	5.19	5.35	19.67	11.53	4.28	53.9
24	MCK 3025	22	F	5.1	9.82	45.57	28.16	14.62	4.81	10.23	113.21	8.67	40.25	24.87	12.92	4.25	9.0
25	F5C 3025	23	F	8.3	48.77	70.05	31.60	19.99	3.02	16.48	189.91	25.68	36.89	16.64	10.52	1.59	8.6
26	F5D 3038	24	F	7.5	69.43	54.56	14.32	6.92	2.05	33.70	180.98	38.37	30.15	7.91	3.82	1.13	18.6
27	MCK 3038	26	F	1.5	5.14	12.57	4.41	1.70	3.14	11.85	38.81	13.25	32.39	11.36	4.38	8.09	30.5
28	FCG 3041	25	F	1.8	7.55	11.71	3.67	1.24	1.75	19.95	45.87	16.46	25.53	8.00	2.70	3.82	43.4
29	FCG 3081	25	F	5.2	11.52	24.82	11.48	9.25	2.97	68.86	128.90	8.94	19.26	8.90	7.18	2.30	53.4
30	FCG 3087	24	F	22.0	9.76	13.92	5.10	3.27	3.41	18.06	53.52	18.24	26.01	9.53	6.11	6.37	33.7
31	FCG 3088	24	F	2.5	8.71	17.66	7.22	3.07	1.20	22.93	60.79	14.33	29.05	11.88	5.05	1.97	37.7
32	FCG 3095	24	F	2.8	7.34	16.27	7.09	4.01	3.74	27.85	66.30	11.07	24.54	10.70	6.05	5.64	42.0
33	FCG 3096	24	F	2.6	7.44	17.26	4.78	4.04	1.36	27.56	62.44	11.91	27.64	7.66	6.47	2.18	44.1
34	FCG 3097	24	F	3.7	10.21	16.93	7.25	5.88	5.64	42.38	88.29	11.56	19.18	8.21	6.66	6.39	48.0
35	FCG 3101	24	F	2.9	6.79	11.41	6.22	4.20	3.50	37.09	69.21	9.81	16.49	8.99	6.06	5.06	53.5
36	FCG 3102	23	F	2.5	5.62	10.54	3.51	2.53	1.74	33.22	57.16	9.83	18.44	6.14	4.43	3.04	58.1
37	FCG 3103	25	F	5.2	16.98	4.23	20.33	6.67	3.23	77.50	128.94	13.17	3.28	15.77	5.17	2.51	60.1
38	FCG 3104	24	F	3.4	9.43	15.29	5.45	5.83	4.17	42.29	82.46	11.44	18.54	6.61	7.07	5.05	51.2
39	FCG 3108	43	F	5.0	30.00	32.68	12.99	16.19	11.67	112.84	216.37	13.87	15.11	6.00	7.48	5.39	52.1
40	FCG 3110	24	F	4.7	14.74	18.69	7.43	5.47	6.24	59.70	112.27	13.13	16.64	6.62	4.87	5.56	53.1
41	FCG 3111	30	F	5.9	28.28	26.09	10.16	10.81	7.48	93.49	176.31	16.04	14.80	5.76	6.13	4.24	53.0
42	FCG 3115	26	F	1.4	1.70	12.18	3.96	1.72	1.23	15.90	36.69	4.63	33.20	10.79	4.69	3.35	43.3
43	FCG 3116	24	F	1.4	1.39	7.06	3.95	2.10	1.31	18.56	34.37	4.05	20.54	11.49	6.11	3.81	54.0
44	FCG 3117	25	F	1.5	1.33	11.11	2.76	1.01	1.08	20.86	38.15	3.49	29.12	7.23	2.65	2.83	54.6
45	FCG 3118	24	F	1.5	4.26	9.64	2.22	0.80	0.11	18.93	35.96	11.85	26.81	6.17	2.22	0.31	52.6
46	FCG 3124	24	F	1.5	3.48	10.90	4.73	2.72	0.13	12.89	34.85	9.99	31.28	13.57	7.80	0.37	36.9
47	FCG 3125	24	F	1.6	2.51	13.39	3.82	2.92	1.60	13.44	37.68	6.66	35.53	10.14	7.75	4.25	35.6

Table 1 (cont)

OBS SITE	CJDATE	TIME (hr)	FILTER	GRATE (mg/hr)	WT1	WT2	WT3	WT4	WT5	WT6	WTOTAL	PC1	PC2	PC3	PC4	PC5	PC6
48	FCG	3130	F	1.8	6.77	12.73	4.66	1.85	2.06	13.45	41.52	16.31	30.66	11.22	4.46	4.96	32.3
49	FCG	3131	F	2.1	4.52	9.26	3.61	0.15	0.73	32.94	51.21	8.83	18.08	7.05	0.29	1.43	64.3
50	FCG	3132	F	2.1	6.24	13.45	7.43	3.58	1.53	19.72	51.95	12.01	25.89	14.30	6.89	2.95	37.9
51	FCG	3136	F	1.9	5.20	10.03	5.55	3.78	1.93	21.23	47.72	10.90	21.02	11.63	7.92	4.04	44.4
52	FCG	3137	F	1.0	1.89	6.66	1.61	1.59	1.38	11.76	24.89	7.59	26.77	6.46	6.39	5.54	47.2
53	FCG	3138	F	0.9	2.24	5.36	1.01	0.55	0.35	13.26	22.77	9.84	23.54	4.44	2.41	1.54	58.2
54	FCG	3139	F	1.4	2.33	10.71	0.64	1.36	1.45	16.70	33.19	7.02	32.27	1.93	4.10	4.37	50.3
55	FCG	3143	F	0.8	0.13	8.63	4.22	0.55	0.98	5.30	19.81	0.66	43.56	21.30	2.78	4.95	26.7
56	FCG	3144	F	1.2	7.36	22.67	9.56	4.33	0.69	13.68	58.29	12.63	38.89	16.40	7.43	1.18	23.4
57	FCG	3145	F	1.1	2.19	8.89	2.70	0.60	0.46	10.49	25.33	8.65	35.10	10.66	2.37	1.81	41.4
58	FCG	3151	F	1.1	0.37	8.20	0.52	1.32	0.94	15.39	26.74	1.38	30.67	1.94	4.94	3.52	57.5
59	FCG	3152	F	1.4	0.54	10.21	4.67	1.33	0.28	14.09	31.12	1.74	32.81	15.01	4.27	0.90	45.2
60	FCG	3153	F	1.2	1.28	8.77	1.36	0.42	0.64	14.51	26.98	4.74	32.51	5.04	1.56	2.37	53.7
61	FCG	3157	F	1.3	3.94	10.60	3.24	0.66	1.55	17.03	37.02	10.64	28.63	8.75	1.79	4.19	46.0
62	FCG	3158	F	1.1	2.82	6.30	1.84	0.73	0.02	14.60	26.31	10.72	23.95	6.99	2.77	0.08	55.4
63	FCG	3159	F	1.3	0.05	8.88	2.44	0.93	1.62	17.25	31.17	0.16	28.49	7.83	2.98	5.20	55.3
64	FCG	3160	F	1.4	2.73	8.76	3.02	1.06	0.03	14.44	30.04	9.09	29.16	10.05	3.53	0.10	48.0
65	FCG	3164	F	2.1	5.04	9.18	5.17	3.83	2.83	21.90	47.95	10.51	19.15	10.78	7.99	5.90	45.6
66	FCG	3165	F	0.9	-0.04	5.40	1.29	-0.15	-1.05	15.10	20.55	-0.20	26.28	6.28	-0.73	-5.11	73.4
67	FCG	3166	F	1.0	1.23	5.50	2.04	1.05	0.15	14.82	24.79	4.96	22.19	8.23	4.24	0.60	59.7
68	FCG	3171	F	1.1	3.39	6.89	2.87	0.38	-0.83	13.23	25.93	13.09	26.49	11.08	1.47	-3.21	51.0
69	FCG	3172	F	1.3	4.63	12.65	2.67	0.58	0.45	11.16	32.14	14.41	39.36	8.31	1.80	1.40	34.7
70	FCG	3172	F	1.2	3.21	9.25	2.12	1.48	1.18	12.07	29.31	10.95	31.56	7.23	5.05	4.03	41.1
71	F50	3192	F	1.0	1.10	4.21	0.57	1.76	2.05	14.93	24.62	4.47	17.10	2.31	7.15	8.33	60.6
72	F5C	3192	F	2.3	24.80	13.60	2.04	2.35	2.06	13.46	58.31	42.53	23.32	3.50	4.03	3.53	23.0
73	MCK	3193	F	2.2	5.41	17.70	9.02	1.80	1.47	18.31	53.71	10.07	32.96	16.79	3.35	2.74	34.0
74	FCG	2064	F	1.7	11.80	100.0
75	FCG	2069	F	2.2	50.10	100.0
76	FCG	2068	C	3.5	81.56	100.0
77	FCG	2070	C	4.0	96.41	100.0
78	F5C	2081	F	7.5	186.90	100.0
79	F5C	2082	C	9.6	220.88	100.0
80	F50	2125	C	3.9	85.85	100.0
81	FCG	2126	F	8.9	185.86	100.0
82	ROD	2140	C	5.4	133.27	100.0

Table 1 (concluded)

SYMBOLS:

- SITE: FCG = FORT CLAYTON GENERAL PURPOSE TEST AREA
ROD = RODMAN MUNITIONS SURVEILLANCE SITE
MCK = MCKENZIE FOREST EXPOSURE SITE
FSO = FORT SHERMAN OPEN EXPOSURE SITE
FSC = FORT SHERMAN COASTAL EXPOSURE SITE
- CDATE: THE FIRST DIGIT REFERS TO THE YEAR IN WHICH
THE SAMPLE WAS COLLECTED, EITHER 1982 OR 1983.
THE LAST 3 DIGITS REFER TO THE DAY OF THE YEAR
ACCORDING TO THE JULIAN DATE SYSTEM FOR A
REGULAR YEAR.
- TIME: COLLECTION TIME IN HOURS (hr).
- FILTER: FIBERGLASS (F) OR CELLULOSE (C).
- CRATE: COLLECTION RATE. WEIGHT TOTAL IN MILLIGRAMS
DIVIDED BY COLLECTION TIME IN HOURS, (mg/hr).
- WT: WEIGHT OF PARTICLES COLLECTED IN MILLIGRAMS, (mg).
- PC: PERCENT OF WEIGHT COLLECTED, %.
- 1-6: REFERS TO THE STAGE NUMBER. STAGE 6 IS THE HIGH
VOLUME FILTER THAT IS PLACED BENEATH THE SET
OF IMPACTOR STAGES.

Table 2 (cont)

OBS	SITE	FILTER	CJDATE	STAGE	ANALYSIS	ELEMENTS																	
						Al	Ca	Cl	Cr	Cu	Fe	K	Mg	Mn	Na	P	Pb	S	Si	Sn	Ti	V	Zn
96	FCG	C	2126	2	2	8	4	.	.	7	6	2	9	.	.	1	.	3	5
97	FCG	C	2126	2	3	7	3	5	.	6	.	2	8	.	.	1	.	4
98	FCG	C	2126	3	1	2	.	4	5	3
99	FCG	C	2126	4	1	.	1	.	.	2	.	4	1	5
100	FCG	C	2126	4	2	.	3	.	.	4	.	2	1	5
101	FCG	C	2126	4	3	.	3	.	.	2	.	4	1	5
102	FCG	C	2126	6	1	4	1	3	.	8	7	6	.	.	9	10	.	5	2	.	6	.	.
103	FSO	C	2126	1	1	4	3	2	.	.	7	6	5	1
104	FSO	C	2126	1	2	3	7	5	1
105	FSO	C	2126	1	3	6	2	4	.	.	3	7	5	1
106	FSO	C	2126	2	1	5	3	2	.	7	8	9	6	.	10	.	4	1
107	FSO	C	2126	2	2	5	3	1	.	8	9	7	6	.	.	.	4	2
108	FSO	C	2126	2	3	2	5	3	.	6	7	4	1
109	FSO	C	2126	3	1	5	6	3	.	7	8	9	4	.	10	.	2	1
110	FSO	C	2126	3	2	9	3	1	.	5	6	8	7	.	10	.	2	1
111	FSO	C	2126	3	3	10	4	1	.	7	9	8	5	.	6	.	2	3
112	FSO	C	2126	4	1	4	3	.	.	5	6	7	1	2
113	FSO	C	2126	5	1	3	2	1	4
114	FSO	C	2126	5	2	.	4	.	.	6	5	3	1	2
115	FSO	C	2126	5	3	4	.	2	1	3
116	FSO	C	2126	5	4	1	3	1	.	.	4	5	.	1	.	2	.	.
117	FSO	C	2126	6	1	3	1	.	.	.	4	5	.	2	.	2	.	.
118	FSO	C	2126	6	2	3	1	.	.	.	4	5	.	2	.	2	.	.
119	FSO	C	2126	6	3	1	4	.	.	.	6	10	1	.	3	7	.	.
120	ROD	C	2140	1	1	8	5	4	.	6	.	7	3	7
121	ROD	C	2140	1	2	.	4	5	.	.	7	6	8	.	9	3	.	2	1	.	.	.	6
122	ROD	C	2140	1	3	.	4	5	.	.	7	6	8	.	9	3	.	2	1
123	ROD	C	2140	1	4	7	3	6	.	10	.	5	8	.	9	2	.	4	1
124	ROD	C	2140	2	1	7	5	4	.	11	10	2	8	.	9	1	.	3	6
125	ROD	C	2140	3	1	4	8	10	.	9	.	7	5	.	6	3	.	1	2
126	ROD	C	2140	3	2	3	7	.	.	9	8	6	4	.	5	.	1	2
127	ROD	C	2140	4	1	3	.	.	.	2	4	1	2
128	ROD	C	2140	5	1	3	5	2	1	3
129	ROD	C	2140	5	2	4
130	ROD	C	2140	6	1
131	MCK1	C	2160	2	1	4	.	3	1	.	2
132	MCK1	C	2160	2	2	7	5	4	.	.	3	8	9	1	.	2	6	.	.
133	MCK1	C	2160	2	3	.	8	4	.	7	.	3	5	.	6	1	.	2	1
134	MCK1	C	2160	4	1	6	4	5	.	2	.	3	1
135	MCK1	C	2160	4	2	.	1	.	.	.	6	3	.	.	5	3	.	4	2
136	MCK1	C	2160	4	3	2	5	2	.	.	4	2	.	3	1
137	MCK1	C	2160	4	4	1	5	.	.	.	4	.	3	1
138	MCK1	C	2160	6	1
139	MCK2	C	2160	1	1	.	4	.	.	.	3	5	1
140	MCK2	C	2160	2	1	7	5	.	.	6	1	.	2	3
141	MCK2	C	2160	2	2	7	5	.	.	6	1	.	2	3
142	MCK2	C	2160	2	3	.	6	4	.	.	5	1	.	3	2	.	.	.

Table 2 (concluded)

										ELEMENTS														
OBS	SITE	FILTER	CJDATE	STAGE	ANALYSIS	Al	Ca	Cl	Cr	Cu	Fe	K	Mg	Mn	Na	P	Pb	S	Si	Sn	Ti	V	Zn	
660	FS0	F	3192	3	3	.	.	1	.	2	3
661	FS0	F	3192	4	1	.	.	2	1
662	FS0	F	3192	4	2	.	.	2	1
663	FS0	F	3192	4	3	.	.	2	1
664	FS0	F	3192	5	1	4	3	2	.	1
665	FS0	F	3192	5	2	3	2	1
666	FS0	F	3192	6	1	.	.	2	1
667	FS0	F	3192	6	2	.	.	2	1
668	FS0	F	3192	6	3	.	.	2	1
669	FS0	F	3192	6	4	.	.	1	.	.	2	3

SYMBOLS:

- (.) : Element found absent.
- (1,2 ..) : Elements found present. Number one is assigned to the element exhibiting the strongest peak in spectrum.

Table 3 (concluded)

FUNGI SYMBOLS:

ABB - Aspergillus sp., BLACK-BROWN CONIDIA
 ABG - Aspergillus sp., BLACK-GREEN CONIDIA
 ADY - Aspergillus sp., YELLOW CONIDIA
 ALT - Alternaria sp.
 ASP - Aspergillus sp., BROWN CONIDIA
 ASS - Aspergillus sp., YELLOW-GREEN CONIDIA (SULPHUR)
 ASW - Aspergillus sp., WHITE CONIDIA
 CEP - Cephalosporium sp.
 CLA - Cladosporium sp.
 CUR - Curvularia sp.
 FUM - Fusarium sp.
 GLI - Gliocladium sp.
 GRA - Graphium sp.
 HYA - Hyalodendrum sp.
 MON - Monilia sp.
 NIG - Nigrospora sp.
 PAS - Paecilomyces sp.
 PEC - Penicillium sp.
 PES - Pestalotia sp.
 PHM - Phoma sp.
 RHN - Rhinocladella sp.
 RHZ - Rhizopus sp.
 SPI - Spicaria sp.
 STE - Stemphylium sp.
 STR - Streptomyces sp.
 TIL - Tiliachlidium sp.
 TRI - Trichoderma sp.
 CLD, FUL, UNA, UND, UNE, UNH, UNI, and UNK - (Unknowns, see Appendix B for their descriptions).

TABLE 5. LIST OF FUNGI OBSERVED, DIRECT EXPOSURE OF CULTURE PLATES

OBS SITE	SAMPLE TIME (min)	CJDATE	ABB	ABG	ADY	ASP	CEP	CLA	CUR	FUL	FUM	HYA	MON	NIG	PAS	PES	PHM	RHN	SPI	STE	STR	UNH	UNI	UNJ	UNK
1 FCG	1	11 3088	X	X	.	.	.	X	X	.	X	.	.	X
2 FCG	2	11 3088	X	X	.	.	.	X	X	.	X	.	.	X
3 FCG	3	11 3088	X	X	.	.	.	X	X	.	X	.	.	X
4 FCG	4	1440 3088	.	.	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X
5 FCG	5	1440 3088	.	.	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X
6 FCG	6	1440 3088	.	.	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X
7 FCG	1	2 3097	X	X	.	X	.	.	X
8 FCG	2	2 3097	X	X	.	X	.	.	X
9 FCG	3	4 3097	X	X	.	X	.	.	X
10 FCG	4	4 3097	X	X	.	X	.	.	X
11 FCG	5	8 3097	X	X	.	X	.	.	X
12 FCG	5	8 3097	X	X	.	X	.	.	X
13 FCG	1	1 3098	X	X	.	X	.	.	X
14 FCG	2	1 3098	X	X	.	X	.	.	X
15 FCG	3	2 3098	X	X	.	X	.	.	X
16 FCG	4	2 3098	X	X	.	X	.	.	X
17 FCG	5	5 3098	X	X	.	X	.	.	X
18 FCG	6	5 3098	X	X	.	X	.	.	X
19 FCG	1	1 3110	X	X	.	X	.	.	X
20 CTR	1	0 3110	X	X	.	X	.	.	X
21 FCG	1	1 3112	X	X	.	X	.	.	X
22 CTR	1	0 3112	X	X	.	X	.	.	X
23 CTR	1	1 3108	X	X	.	X	.	.	X
24 CTR	2	1 3108	X	X	.	X	.	.	X
25 CTR	3	1 3108	X	X	.	X	.	.	X
26 CTR	4	1 3108	X	X	.	X	.	.	X
27 CTR	5	1 3108	X	X	.	X	.	.	X
28 CTR	6	1 3108	X	X	.	X	.	.	X
29 CTR	7	1 3108	X	X	.	X	.	.	X
30 CTR	8	1 3108	X	X	.	X	.	.	X

FUNGI SYMBOLS:

- ABB - Aspergillus sp., BLACK-BROWN CONIDIA
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- ASP - Aspergillus sp., BROWN CONIDIA
- CEP - Cephalosporium sp.
- CLA - Cladosporium sp.
- CUR - Curvularia sp.
- FUM - Fusarium sp.
- HYA - Hyalodendrum sp.
- MON - Monilia sp.
- NIG - Nigrospora sp.
- PAS - Paecilomyces sp.
- PEC - Penicillium sp.
- PES - Pestalotia sp.
- PHM - Phoma sp.
- RHN - Rhinocladiella sp.
- SPI - Spicaria sp.
- STE - Stemphylium sp.
- STR - Streptomyces sp.
- FUL, UNH, UNI, UNJ and UNK - (Unknowns, see Appendix B for their descriptions).

APPENDIX B. DESCRIPTION OF UNKNOWN FUNGI

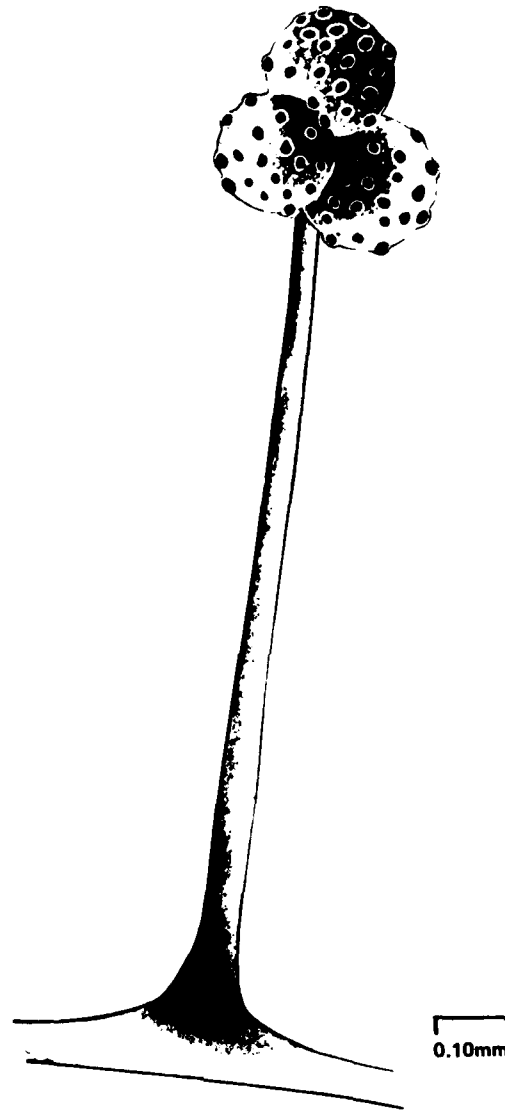


Figure B-1. UNA. White Colony. Conidiophore straight, hyaline, branched at the tip with white vesicles (usually 3) of different sizes. Oval Conidia, hyaline.

Appendix B (cont)

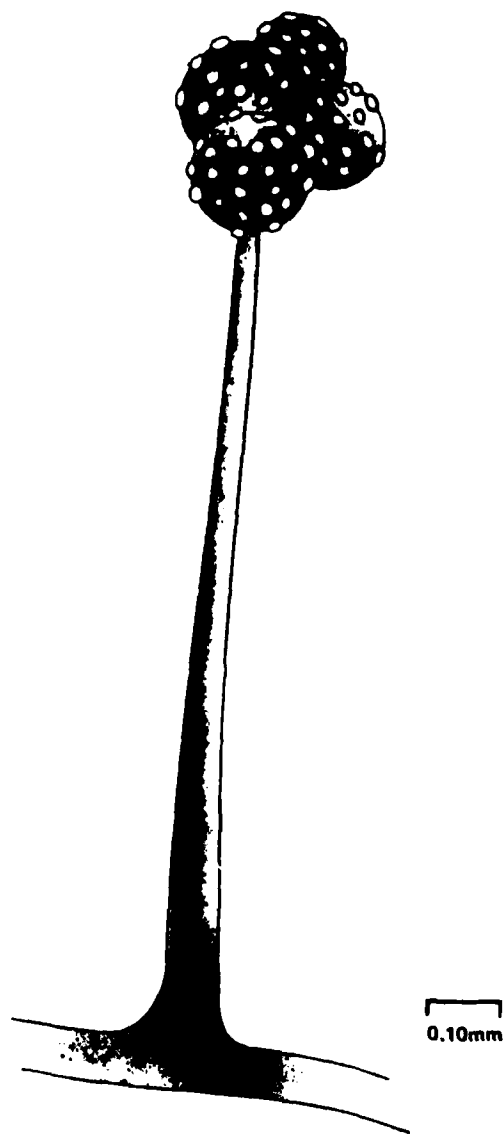


Figure B-2. UNB. Sporangiophores hyaline and erect, ending usually in 4 globose vesicles. Brown, Oval Conidia [similar to (UNA) but with pigmented spores].

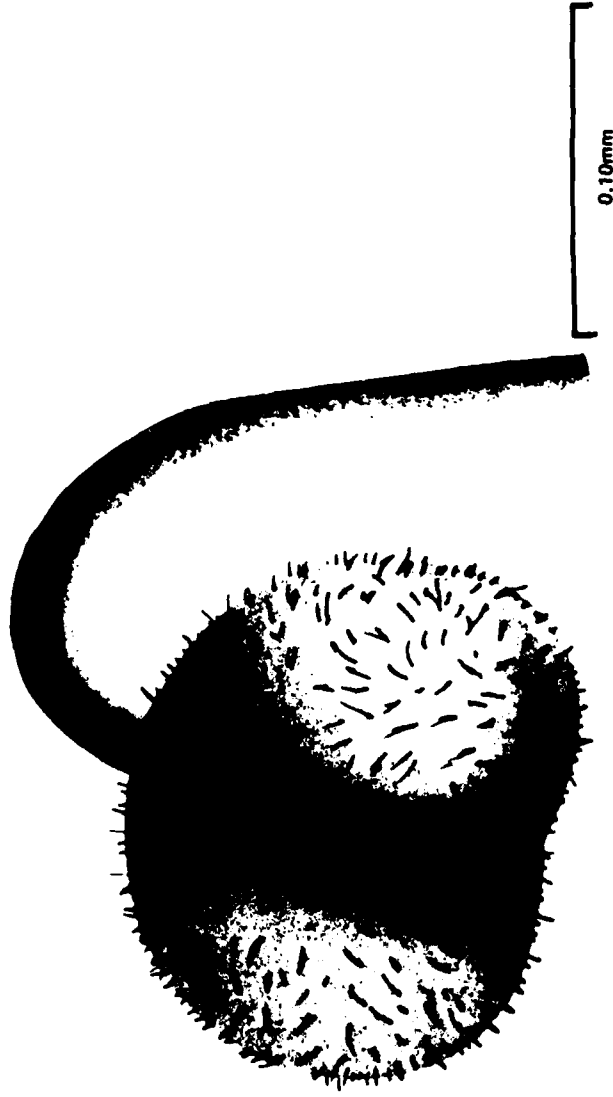


Figure B-3. UNC. Sporangioophore hyaline, single and erect from the mycelium but curved at the end. Dark globose vesicle at the tip. The wall of the vesicle is thick with a closed dehiscence in the center that opens to release the spores. Large oval spores with cuticularized and striated surface.

Appendix B (cont)

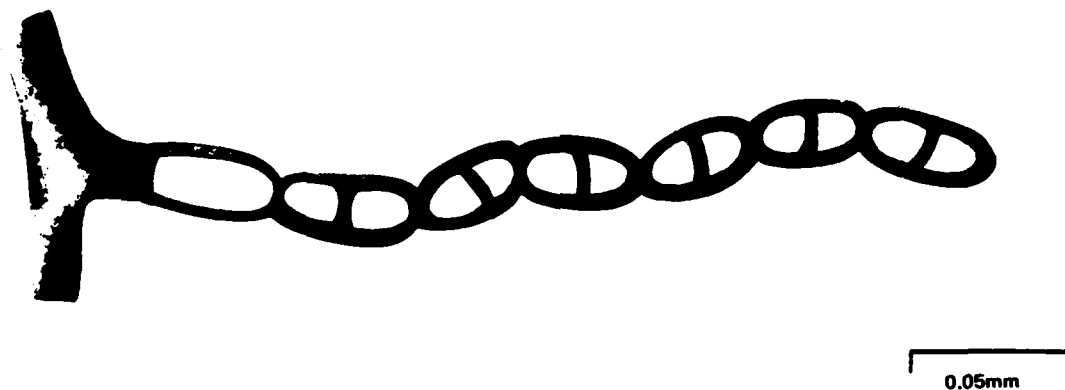


Figure B-4. UND. Conidiophore Inconspicuous. Conidia catenulate in acropetalous chains, cylindrical, hyalin, 2-celled.

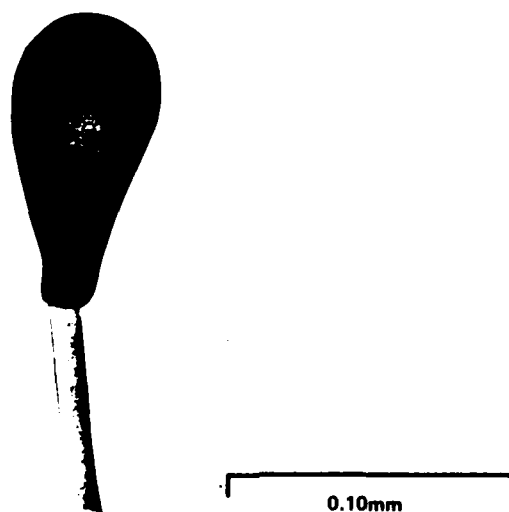


Figure B-5. UNE. Conidiophore short, simple, erect. Conidia dark brown, septate, pyriform with a funnel-shaped base.

Appendix B (cont)

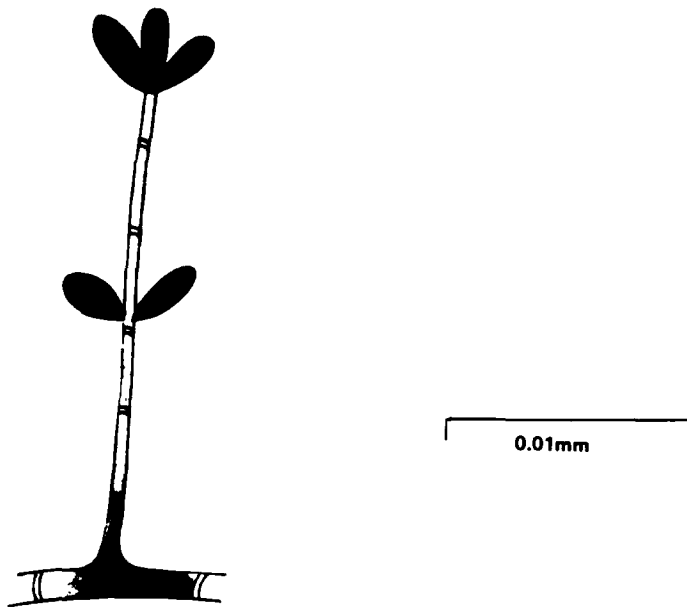


Figure B-6. UNF. Conidiophore erect and unbranched, bearing clusters of conidia on several nodes (verticillated).

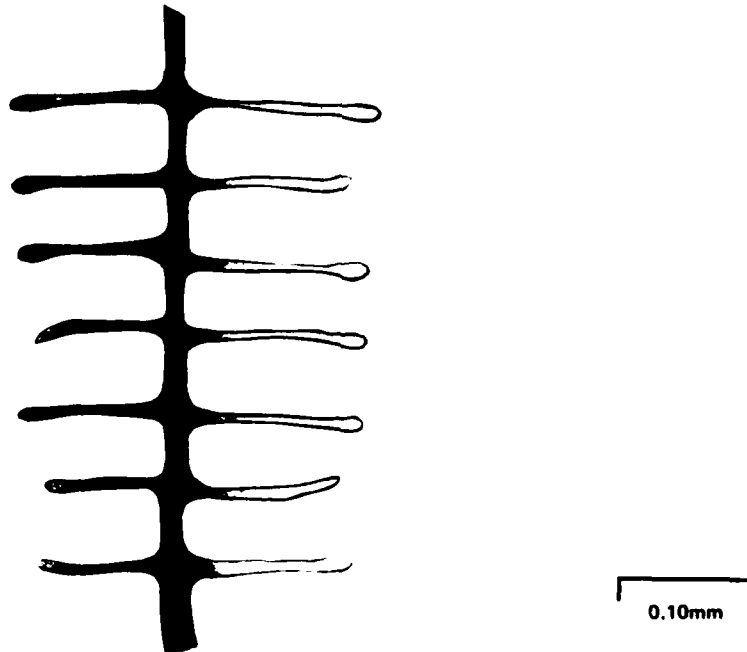


Figure B-7. UNG. Principal Hyphae with erect Conidiophores forming branches on both sides. A single oval conidia on tip of conidiophore. Hyaline colony.

Appendix B (cont)

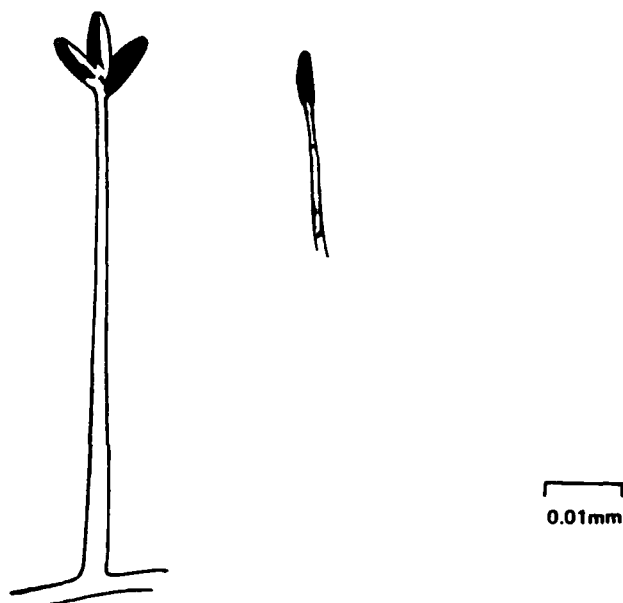


Figure B-8. UNH. Single, long and erect. Conidiophore arising from mycelia, brown colored. Terminal conidia, oblong and borne in heads at the apex of the conidiophore.

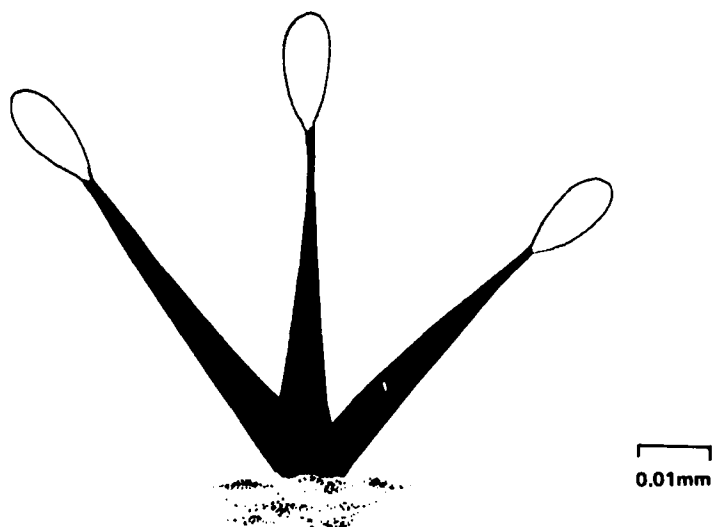


Figure B-9. UNI. Conidiophore arise single or in groups of 3 or more, erect. Conidia ovate, at the apex. All structures hyaline.

Appendix B (cont)

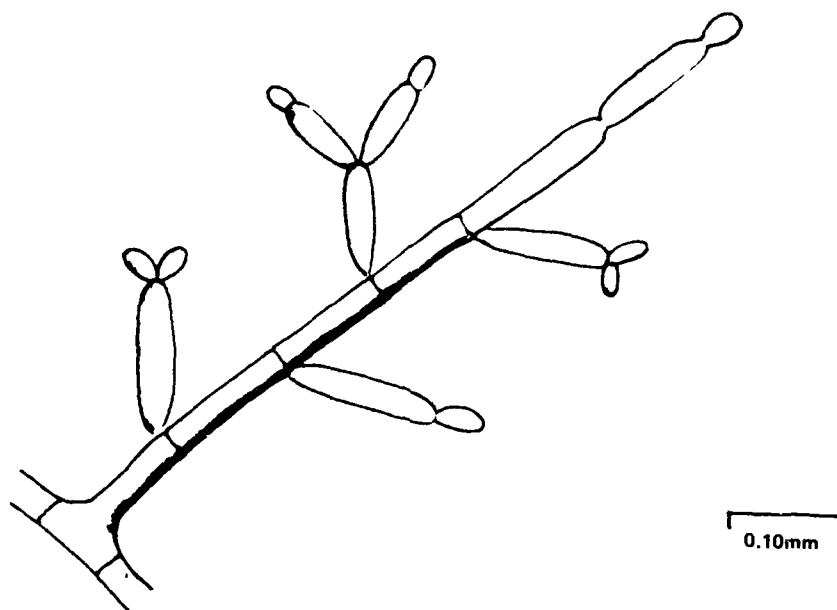


Figure B-10. UNJ. Conidiophore erect, with simple or forked branches occurring on both sides. Carrying 1 or 2 conidia at the tip. Hyaline structures.

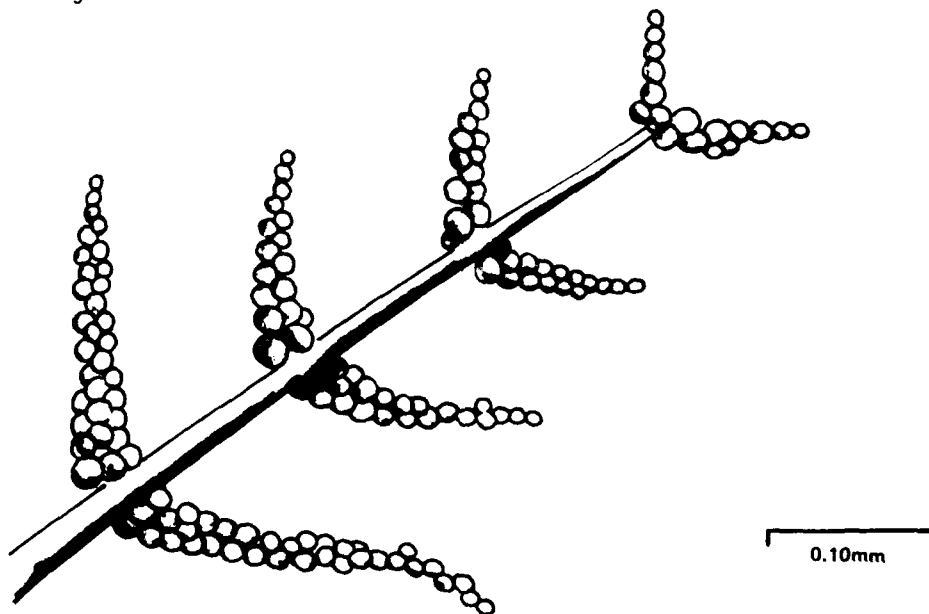


Figure B-11. UNK. Conidiophores occurring as side branches. Conidia borne laterally on the branches, very numerous, small, sessile, globose, hyaline, 1-celled.

Appendix B (cont)

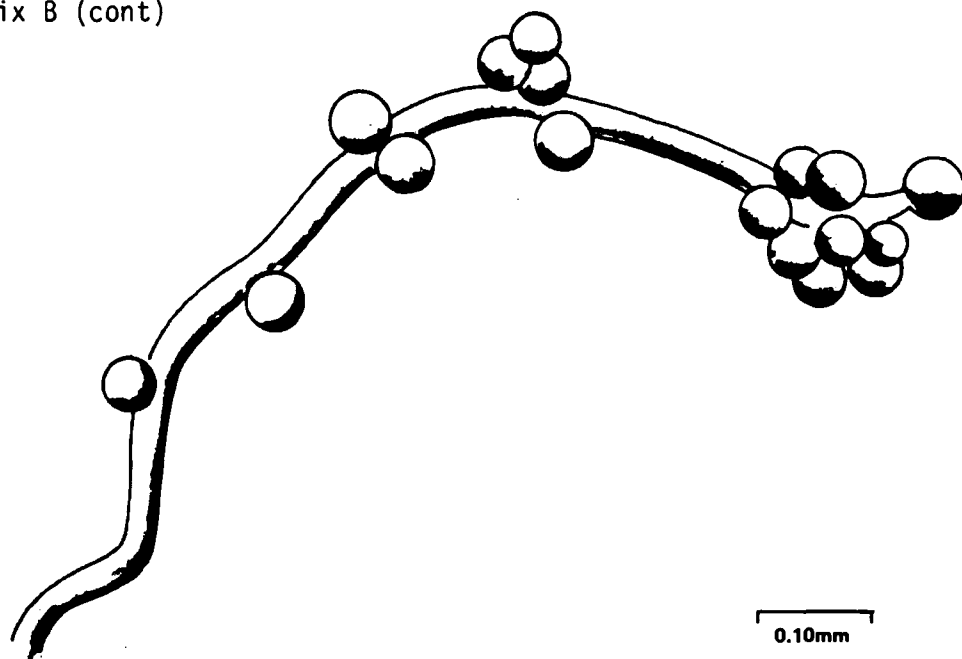


Figure B-12. UNL. Conidiophore long, hyaline, slender and proliferating. Bearing terminally or laterally conidia. Conidia hyaline, globose and 1-celled.

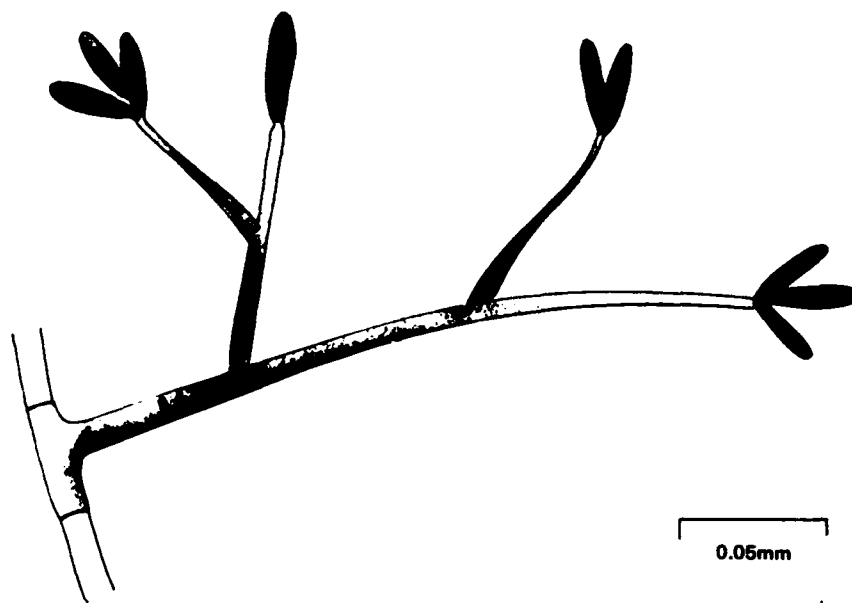


Figure B-13. FUL Fusarium-like Spores. Straight conidia, large, cylindrical, spores has no foot cell.

Appendix B (concluded)

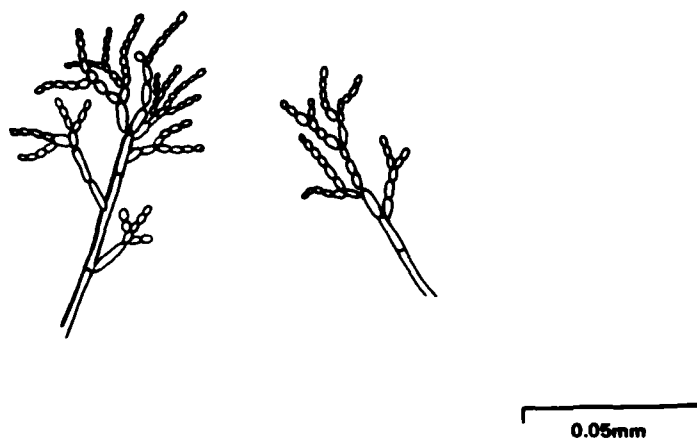


Figure B-14. CLD Cladosporium-like spores. Conidiophores erect, forming a dense turf, olive-green or brown. Conidia ovate in very long chains, fusoid, 1-celled, ten times smaller than common Cladosporium.

APPENDIX C. REFERENCES

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APPENDIX D. DISTRIBUTION LIST

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