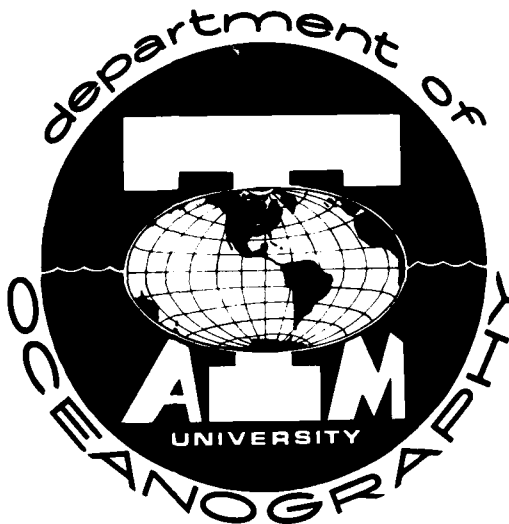




LEVEL

12

AD A103536



DTIC  
 S E L E C T E D  
 SEP 1 1981  
 H

**GEOCHEMISTRY OF DISSOLVED GASES IN THE  
 HYPERSALINE ORCA BASIN**

**TECHNICAL REPORT  
 by  
 Denis A. Wiesenburg**

**Reference 80-14-T  
 December 1980**

**Sponsored by  
 OFFICE OF NAVAL RESEARCH  
 Contracts N00014-75-C-0537 and N00014-80-C-00113**

**DISTRIBUTION STATEMENT A  
 Approved for public release;  
 Distribution Unlimited**

DTIC FILE COPY

81 9 01 023

MANDATORY DISTRIBUTION LIST

FOR UNCLASSIFIED TECHNICAL REPORTS, REPRINTS & FINAL REPORTS  
PUBLISHED BY OCEANOGRAPHIC CONTRACTORS  
OF THE OCEAN SCIENCE AND TECHNOLOGY DIVISIONS  
OF THE OFFICE OF NAVAL RESEARCH  
(REVISED NOV. 1978)

- 1 Deputy Under Secretary of Defense  
(Research and Advanced Technology)  
Military Assistant for Environmental Science  
Room 3D129  
Washington, DC 20301
- Office of Naval Research  
800 North Quincy Street  
Arlington, VA 22217
- 3 \* ATTN: Code 483\*  
1 ATTN: Code 460  
2 ATTN: 102B
- 1 ResRep (if any) - - - - - - - - - -Mr. Frank Lucas or Robert Prestridge  
ONR Resident Representative  
Room 582, Federal Building  
300 E. 8th Street  
Austin, TX 78701
- 6 ATTN: Library, Code 2627
- 12\*\* Defense Documentation Center  
Cameron Station  
Alexandria, VA 22314  
ATTN: DCA
- Commander  
Naval Oceanographic Office  
NSTL Station  
Bay St. Louis, MS 39522
- 1 Attn: Code 8100  
1 Attn: Code 6000  
1 Attn: Code 3300
- 1 NODC/NOAA  
Code D781  
Wisconsin Avenue, N.W.  
Washington, D.C. 20235
- \* Add one separate copy of  
Form DD-1473

\*\* Send with these 12 copies two completed forms DDC-50  
one self addressed back to the contractor, then the  
other addressed to ONR, Code 480.





20. ABSTRACT (cont.)

oxide. Physical stratification of the water column strongly influences Orca Basin gas distributions. The high salinity brine ( $\sim 250\%$ ) is internally well mixed due to convective overturning, but transfer across the brine-sea water interface is controlled by molecular diffusion. With a molecular diffusivity of  $10^{-5} \text{ cm}^2 \cdot \text{sec}^{-1}$ , it will take  $10^6$  years for all salts to diffuse from the basin. Heat diffuses faster than salt and is lost from the basin at a rate of  $0.5 \text{ } \mu\text{cal} \cdot \text{cm}^2 \cdot \text{sec}^{-1}$ . If geothermal heat input from the sediments is slightly higher, this input could account for the higher temperature in the brine ( $5.6^\circ\text{C}$ ) compared to the deep Gulf waters ( $4.2^\circ\text{C}$ ).

This study has shown the utility of dissolved gases in examining water chemistry of unusual areas. Since sources of dissolved gases are independent of the sources of major ions in solution, calculations of gas distributions on a salt-free basis are useful in examining production and consumption processes. The high stability of the Orca brine pool makes it an ideal natural laboratory for examining processes of anaerobic decomposition.

Texas A&M University  
Department of Oceanography  
College Station, Texas

Research conducted through the  
TEXAS A & M RESEARCH FOUNDATION

GEOCHEMISTRY OF DISSOLVED GASES IN THE  
HYPERHALINE ORCA BASIN

by

Denis Alan Wiesenburg

REFERENCE 80-14-T

December 1980

This research was sponsored by  
Office of Naval Research Contracts  
N00014-75-C-0537 & N00014-80-C-00113  
with the Texas A & M Research Foundation

Distribution of this report is unlimited

Preface

This report was written by Denis Alan Wiesenburg as partial fulfillment of the requirements for the degree of Doctor of Philosophy in Oceanography at Texas A&M University. Financial support was provided by the Office of Naval Research, under Contracts N00014-75-C-0537 and N00014-80-C-00113.

Accession For	
NTIS CRA&I	<input checked="" type="checkbox"/>
DTIC TAB	<input type="checkbox"/>
Unannounced	<input type="checkbox"/>
Justification	
By _____	
Distribution/ _____	
Availability Codes	
Dist	Avail and/or Special
A	

## ABSTRACT

Geochemistry of Dissolved Gases in the  
Hypersaline Orca Basin (December 1980)

Denis Alan Wiesenburg, A.B., Duke University  
M.S., Old Dominion University

Chairman of Advisory Committee: Dr. David R. Schink

Hypersaline, anoxic waters significantly affect the biogeochemistry of dissolved gases in the Orca Basin (northern Gulf of Mexico). The high stability of the Orca brine pool makes it an ideal laboratory for studying production and consumption of dissolved gases during anaerobic decomposition. Depth distributions were determined for nitrogen, oxygen, argon, methane, ethane, propane, ammonia, hydrogen sulfide, and nitrous oxide. Physical stratification of the water column strongly influences Orca Basin gas distributions. The high salinity brine ( $\sim 250$  ‰) is internally well mixed due to convective overturning, but transfer across the brine-sea water interface is controlled by molecular diffusion. With a molecular diffusivity of  $10^{-5} \text{ cm}^2 \cdot \text{sec}^{-1}$ , it will take  $10^6$  years for all salts to diffuse from the basin. Heat diffuses faster than salt and is lost from the basin at a rate of  $0.5 \mu\text{cal} \cdot \text{cm}^2 \cdot \text{sec}^{-1}$ . If geothermal heat input from the sediments is slightly higher, this input could account for the higher temperature in the brine ( $5.6^\circ\text{C}$ ) compared to the deep Gulf waters ( $4.2^\circ\text{C}$ ).

The high stability of the brine (due to increased density) prevents either reactants or products of anaerobic decomposition from escaping by other than molecular processes. Concentrations

of biogenic methane and ethane are higher there than in any other anoxic marine basin. Oxygen, nitrate, and nitrous oxide are absent from the brine, while phosphate and ammonia levels are 60 and 500  $\mu\text{mol}\cdot\text{liter}^{-1}$ , respectively. However, there is no hydrogen sulfide in the anoxic brine.

Rates of microbial activity are generally slower in the hypersaline Orca Basin. Organic matter is decomposed so slowly that fronds of Sargassum seaweed have been found buried at depths of 5 m in the sediment, the only sedimentary environment where this has been observed. The absence of free sulfide can be attributed to a slower rate of sulfate reduction. Not enough sulfide is being generated to complex the iron produced there. Iron has accumulated to levels of 30  $\mu\text{mol}\cdot\text{liter}^{-1}$  in the brine and only metastable iron sulfides, not pyrite, are found in the sediments.

Nitrogen and argon in the brine are supersaturated by 470 and 350 %, respectively, compared to atmospheric equilibrium solubilities. The  $\text{N}_2/\text{Ar}$  ratio, however, is 36.4, the estimated ratio from equilibrium of deep Gulf of Mexico waters with air. A comparison of brine and sea water nitrogen and argon concentrations on a salt-free basis showed that these brine gases are remnant gases from the original sea water that formed the Orca brine. This comparison also confirmed that the Orca brine formed by dissolution of a salt deposit at a temperature less than 5°C.

Production of nitrogen and light hydrocarbon gases (methane, ethane, and propane) was evident at the brine-sea water interface.

Methane maxima, similar to those observed in ocean surface waters, were found above the interface. Methane distribution at the interface was similar to the bacterial biomass profiles. Ethane at the interface exhibited a smaller maximum than methane, but the higher levels of ethane in the brine had the same distribution as methane, indicating bacterial production of both methane and ethane.

The carbonate system in the Orca brine closely resembles that of normal sea water. The lower Orca Basin pH (6.83) results mainly from the input of  $2.2 \text{ mmol} \cdot \text{liter}^{-1}$  of biogenic carbon dioxide. The carbonate system dissociation constants were determined at  $25^\circ\text{C}$  to be 5.5 for  $\text{pK}'_1$  and 8.3 for  $\text{pK}'_2$ . At the pH of the Orca Basin brine, the  $\text{CO}_3^{=}$  concentration of the brine is the same as normal sea water. Since the  $K'_{\text{sp}}$  of aragonite appears to be larger in the brine than in sea water, the increased calcium in the Orca brine must be responsible for the striking preservation of the aragonitic pteropod shells found in the basin sediments.

This study has shown the utility of dissolved gases in examining water chemistry of unusual areas. Since sources of dissolved gases are independent of the sources of major ions in solution, calculations of gas distributions on a salt-free basis are useful in examining production and consumption processes. The high stability of the Orca brine pool makes it an ideal natural laboratory for examining processes of anaerobic decomposition.

## ACKNOWLEDGEMENTS

Without the encouragement, assistance, and tutorage of Norman L. Guinasso, Jr., this research would not have been possible. I thank him and Dr. David R. Schink, chairman of my advisory committee, for their valuable help in shaping my view of marine chemistry. I also thank the other members of my committee, Dr. W.M. Sackett, Dr. L.M. Jeffrey, Dr. M.W. Rowe, Dr. C.S. Giam, and Dr. J.W. Foster (Graduate Council Representative) for their contributions. Dr. Sackett was especially helpful in educating me in the fundamentals of stable isotope geochemistry.

Special thanks are due to my colleagues and fellow students at Texas A&M for allowing me to participate in their research cruises and to use their data along with my own. These include: J.M. Brooks, B.B. Bernard, B.J. Presley, J.H. Trefry, R.F. Pflaum, C.S. Schwab, B.P. Boudreau, P.J. Setser, and J.L. Bullister. John W. Johnson and Robert M. Key are gratefully acknowledged for their advice and comradery in the laboratory. Tonalee Carlson provided the pH data and showed me the importance of alkalinity titrations. L.A. Barnard took the excellent SEM pictures of the pteropods.

Financial support was provided by the Office of Naval Research under Contracts N00014-75-C-0537 and N00014-80-C-00113 and National Science Foundation Grants OCE75-21275 and OCE-21009.

Special thanks go to my wife, Jean, for her understanding which remained intact long after her patience was exhausted.

## TABLE OF CONTENTS

CHAPTER	PAGE
I INTRODUCTION AND REVIEW . . . . .	1
Basic Concepts . . . . .	3
The Study Site . . . . .	7
Previous Investigations . . . . .	13
The Early Work . . . . .	13
Subsequent Investigations . . . . .	26
Anaerobic Decomposition Processes . . . . .	35
Research Objectives . . . . .	40
II DENSITY EFFECTS AND PHYSICAL MIXING PROCESSES . . . . .	43
Introduction . . . . .	43
Temperature Distribution . . . . .	47
Salinity Distribution . . . . .	49
Mixing Processes in the Orca Basin . . . . .	58
High Salinity Brine . . . . .	58
Oxygen Step Region and General Mixing . . . . .	64
III MAJOR ATMOSPHERIC GASES IN THE ORCA BASIN BRINE . . . . .	79
Methods . . . . .	83
Gas Solubilities . . . . .	83
Dissolved Oxygen . . . . .	86
Argon and Nitrogen Sampling . . . . .	87
Chromatographic Analysis of Argon and Nitrogen . . . . .	91
Results . . . . .	100
Solubility Comparisons . . . . .	100
Dissolved Oxygen . . . . .	104
Argon and Nitrogen . . . . .	107
Discussion . . . . .	110
Dissolved Oxygen . . . . .	110
Argon and Nitrogen . . . . .	114
Transition Zone . . . . .	114
High Salinity Brine . . . . .	121
IV DISSOLVED REDUCED GASES . . . . .	131
Methods . . . . .	134



## TABLE OF CONTENTS (continued)

CHAPTER	PAGE
Sampling . . . . .	134
Analytical Techniques . . . . .	136
Results . . . . .	142
Caseous Hydrocarbons . . . . .	142
Nitrous Oxide and Ammonia . . . . .	149
Hydrogen Sulfide . . . . .	149
Discussion . . . . .	152
Overall Effects of Decomposition . . . . .	156
Microbial Activity in the Orca Basin . . . . .	160
Methane and Hydrogen Sulfide in the Brine . . . . .	164
Mechanisms for H <sub>2</sub> S Removal . . . . .	178
Other Light Hydrocarbons in the Brine . . . . .	179
Methane at the Interface . . . . .	180
N <sub>2</sub> O Consumption During Denitrification . . . . .	182
 V CARBON DIOXIDE AND THE CARBONATE SYSTEM IN THE HYPERHALINE BRINE . . . . .	185
Alkalinity Considerations . . . . .	188
Carbonate System Interactions . . . . .	190
Methods . . . . .	196
Sampling . . . . .	196
Analytical Techniques . . . . .	197
pH . . . . .	197
Total Carbon Dioxide . . . . .	198
Dilution Techniques . . . . .	200
Titration with HCl . . . . .	201
NaHCO <sub>3</sub> Titration . . . . .	202
Model Calculations . . . . .	203
HCl Titration: General Equation . . . . .	203
NaHCO <sub>3</sub> Titration . . . . .	207
Buffer Intensity . . . . .	212
Results and Discussion . . . . .	213
pH and Total CO <sub>2</sub> Relationships . . . . .	213
Brine Dilution Experiments . . . . .	221
Alkalinity versus Total CO <sub>2</sub> . . . . .	226
Alkalinity in the High Salinity Brine . . . . .	230
Pteropod Preservation in the Sediments . . . . .	237
Excess Calcium and K' <sub>sp</sub> of Brine . . . . .	241

TABLE OF CONTENTS (continued)

CHAPTER	PAGE
VI SUMMARY AND CONCLUSIONS . . . . .	246
REFERENCES . . . . .	250
VITA . . . . .	265

## LIST OF TABLES

TABLE	PAGE
1. Abundance and properties of atmospheric gases . .	5
2. First report of hydrographic data from the Orca Basin (Tabulated from McKee and Sidner 1976)	15
3. Moles per kilogram of ions in sea water and various brines . . . . .	17
4. Concentration changes during evaporation of sea water and brine (after Collins 1969) . . . . .	25
5. Approximate stability across the sills of some basins and fjords (after Richards 1965) . . . . .	36
6. Temperatures and salinities taken from two STD casts on cruise 77-G-13. Note the large change in salinity over such a small depth . . . . .	52
7. Salinity zones in and above the Orca Basin . . .	54
8. Solubilities of N <sub>2</sub> , O <sub>2</sub> , Ar in sea water at 35‰ salinity based on Eq. 5 and coefficients from Weiss (1970) . . . . .	81
9. Measured gas solubilities of Orca Basin brine and sea water. All values are atmospheric equilibrium solubilities except those marked with an asterisk . . . . .	101
10. Calculated theoretical production of dissolved argon in the high salinity Orca Basin brine . . .	122
11. Dissolved gases in Red Sea water, Red Sea brines, Gulf of Mexico Deep Water and Orca Basin brine. .	127
12. Oxidation reactions of sedimentary organic matter	132
13. Analyses of Orca Basin water performed by various chemists on samples from different cruises aboard the R/V Gyre and aboard cruise EN32 on the R/V Endeavor. A dash indicates that measurements were not made on that cruise . . . . .	137

## LIST OF TABLES (continued)

TABLE	PAGE
14. Method of analysis used for Orca Basin samples .	138
15. Low-molecular-weight hydrocarbons in the Orca Basin during cruise 77-G-2. A dash indicates sample was lost . . . . .	144
16. Average concentrations of POC and PON in deep Gulf samples along with mole ratios (after Fredericks 1972) . . . . .	159
17. Molecular and isotopic composition of methane and other gaseous hydrocarbons in anoxic waters .	165
18. Ionic balance percentages and total ionic strength for standard sea water (SSW = 35 <sup>0</sup> /oo salinity), Dead Sea brine (DSB = 299 <sup>0</sup> /oo), Orca Basin brine (OBB = 258 <sup>0</sup> /oo), Red Sea Brine (RSB = 256 <sup>0</sup> /oo), and East Flower Garden brine (EFG = 218 <sup>0</sup> /oo) . . . . .	187
19. Notation for carbonate calculation . . . . .	192
20. Sass and Ben-Yaakov (1977) mathematical solution to the general titration formula. Primes on constants are omitted for simplicity . . . . .	208
21. Total carbon dioxide, $\delta^{13}\text{C}-\text{CO}_2$ and pH from several brine areas . . . . .	220
22. Ion enrichment factors relative to sea water for calcium and magnesium (referenced to chloride) .	227
23. Calculated alkalinity and carbonate dissociation constants for seawater and Orca Basin brine . . .	234
24. Calculations showing the possible changes in total calcium content resulting from the increases in alkalinity, total $\text{CO}_2$ , and ammonia in the Orca Basin . . . . .	244

## LIST OF FIGURES

FIGURE	PAGE
1. Index map of the Texas-Louisiana continental slope showing several small basins which have resulted from salt flow and dissolution. Location of the Orca Basin is indicated. Depth contours are in fathoms . . . . .	8
2. Bathymetric map of the Orca Basin based on the seismic data taken on cruise 76-G-10 of the R/V Gyre (after Shokes et al. 1977) . . . . .	9
3. Vertical temperature and salinity ( $g \cdot liter^{-1}$ ) profiles extending from the deep Gulf of Mexico water into the hypersaline Orca Basin brine . . . . .	11
4. One section of the minisparker record obtained by Shokes et al. (1977) during their mapping of the Orca Basin brine pool . . . . .	12
5. Composite vertical profiles of temperature, shipboard conductive salinity and dissolved oxygen measured in both lobes or ends of the Orca Basin on R/V Gyre cruise 76-G-10 (after Shokes et al. 1977) . . . . .	18
6. Composite vertical profiles of dissolved orthophosphate, silicate and nitrate across the sea water-brine interface for three stations in the Orca Basin (after Shokes et al. 1977) . . . . .	19
7. Plot of bromine versus chlorinity for pore waters with chlorinity anomalies . . . . .	27
8. Typical temperature and salinity depth profiles taken in the water column of the northwestern Gulf of Mexico above the Orca Basin . . . . .	44
9. Density profile for the Gulf of Mexico, extending down through the Orca Basin brine . . . . .	45

## LIST OF FIGURES (continued)

FIGURE	PAGE
10. Continuous temperature profile through the interface to the bottom of the Orca Basin made using a Plessey 9040 STD with the salinity probe disabled . . . . .	48
11. Detailed vertical profile of salinity through the interface region of the Orca Basin . . . . .	51
12. Vertical profiles of temperature and arbitrary salinity in the high salinity water of the Orca Basin. The salinity data are presented to show trends in the salinity distribution. Gulf salinity is 35 ‰ and brine is about 263 ‰ . . . . .	57
13. Salinity versus temperature profile for the Orca Basin . . . . .	66
14. Temperature and salinity profiles for the initial two-layer system in the Orca Basin if the brine entered the basin without mixing. . . . .	69
15. Modified temperature depth profile showing the linear thermal layers through the interface into the high salinity brine . . . . .	71
16. Plot of diffusion coefficients for temperature ( $D_T$ ) with increasing distance from the Orca Basin interface zone (layer 5) . . . . .	74
17. One section of the minisparker profile taken on Gyre cruise 76-G-10. The profile crosses the longest section of the southern lobe of the basin. The right side of the profile is 3% higher than the left hand side . . . . .	75
18. Oscillations of a seiche in steps of one-eighth of a cycle for one-half of a complete oscillation. The depth is exaggerated (after Neumann and Pierson 1966, with modifications to show the two-layered system) . . . . .	77

## LIST OF FIGURES (continued)

FIGURE	PAGE
19. Plot of temperature versus atmospheric solubility for argon and nitrogen, indicating that the relationship is not a linear one . . . . .	82
20. Schematic diagram of thermal jacketed purge cylinder used to equilibrate the brine with dissolved gases . . . . .	84
21. All-glass sample chambers constructed to take uncontaminated water samples from Nansen bottles for dissolved gas analysis . . . . .	92
22. Dissolved gas extraction and analysis system . .	93
23. Typical chromatograms showing the analysis of major dissolved gases in oxygenated sea water with (B) and without (A) the activated charcoal trap in the gas flow line . . . . .	97
24. Typical gas chromatograph for the anoxic water of the Orca Basin derived from the analytical techniques described in Fig. 15 . . . . .	99
25. Dissolved oxygen profiles measured by the micro-Winkler technique (Carpenter 1969) for four different Texas A&M cruises during 1977 and 1978	106
26. Vertical profiles of dissolved argon, nitrogen, and methane through the Orca Basin interface region into the high salinity brine. The $N_2/Ar$ ratio in the brine is 36.4 . . . . .	109
27. A schematic representation of the possible pathways through which organic nitrogen or ammonia may be converted to molecular nitrogen . . .	117
28. Nitrite profiles above the Orca Basin obtained during R/V Gyre cruise 77-G-3 and reported by Wiesenburg et al. (1977) . . . . .	120

## LIST OF FIGURES (continued)

FIGURE	PAGE
29. Argon versus nitrogen in the Orca Basin brine and solubility values for fresh water, 35‰ sea water and Orca Basin brine in equilibrium with the atmosphere . . . . .	125
30. Vertical distributions of temperature, salinity, dissolved oxygen, methane, ethane, propane, and $\delta^{13}\text{CH}_4$ in the Orca Basin . . . . .	143
31. Dissolved methane depth distributions in the Orca Basin taken from gas-tight piggyback samplers and standard Nansen bottles. Data are from cruises 77-G-13 and EN-32 . . . . .	146
32. Hydrocarbon, oxygen and $\sigma_t$ data from the basin pycnocline (cruise 77-G-2). Note the maxima in the methane and ethane profiles at the level where density begins to increase rapidly . . . .	147
33. Methane depth distributions in the pycnocline from different hydrocasts on two separate cruises . .	148
34. Vertical distribution of dissolved oxygen, nitrate, $\text{N}_2\text{O}$ , and ammonia in the Orca Basin. $\text{N}_2\text{O}$ is consumed in the anoxic brine along with nitrate while ammonia increases to about $500 \mu\text{mol}\cdot\text{liter}^{-1}$	150
35. Vertical profiles of ammonia measured by three separate methods (see text) on three different cruises. No obvious structures appear in the ammonia profile within the brine . . . . .	151
36. Depth distributions of sulfate, sulfide, and iron in the Orca Basin . . . . .	153
37. Sediment geochemistry data from a piston core taken in the Orca Basin ( $27^\circ 01.3' \text{N}$ , $91^\circ 16.5' \text{W}$ ) at a water depth of 2340 m . . . . .	169
38. pH versus log salinity dilution curves for the Dead Sea brine and two other waters (after Amit and Bentor 1971) . . . . .	194



## LIST OF FIGURES (continued)

FIGURE	PAGE
39. Schematic presentation of the gas chromatographic system used to measure total carbon dioxide in the Orca Basin brine . . . . .	199
40. Typical Deffeyes diagram for sea water (after Deffeyes 1965) . . . . .	209
41. Theoretical alkalinity (A) versus total CO <sub>2</sub> (C) plot showing lines of constant pH. Lines 1 and 2 represent the results of a NaHCO <sub>3</sub> titration with A > C (1) or A < C (2) . . . . .	211
42. pH and total carbon dioxide depth distribution in the northwestern Gulf of Mexico . . . . .	214
43. pH and total carbon dioxide profiles through the Orca Basin brine interface and into the high salinity brine . . . . .	215
44. Detailed plot of pH and dissolved oxygen through the interface region of the Orca Basin . . . . .	217
45. pH versus log salinity dilution curves for two samples of Orca Basin brine and for surface and deep water from the Gulf of Mexico . . . . .	222
46. Change in total carbon dioxide with dilution of Orca Basin brine. These CO <sub>2</sub> measurements were made on the same samples analyzed for pH and shown in Fig. 45. Total CO <sub>2</sub> values for each brine sample are shown in units of mmol·liter <sup>-1</sup> . . . . .	224
47. Change in pH of the Orca Basin brine and Dead Sea brines with addition of 0.5 N NaHCO <sub>3</sub> . . . . .	228
48. Representative HCl titration curves of sea water and Orca Basin brine. Volume of sample was 180 ml for each . . . . .	231
49. Buffer capacity (the incremental change in pH for a given addition of HCl) plotted as a function of pH . . . . .	233

## LIST OF FIGURES (continued)

FIGURE	PAGE
50. Carbonate system species distributions for sea water, the Orca Basin brine, and the Dead Sea brines. The vertical dashed lines indicate the pH ranges of the natural systems . . . .	236
51. Scanning electron photographs of three pteropod species preserved in the Orca Basin sediments. The number above the length scale on each micrograph gives the scale length in micrometers . . .	239
52. Three scanning electron photographs of a <u>Cavolinia longirostris</u> pteropod found in the Orca Basin sediments. The number above each length scale on each micrograph gives the scale length in micrometers. . . . .	240

CHAPTER I  
INTRODUCTION AND REVIEW

Every atmospheric gas is found dissolved in sea water. Oceanic gas concentrations are affected by the same equilibrium processes and transport mechanisms that control the overall geochemistry of the oceans. Unlike the major sea water components, however, the primary source for gases dissolved in sea water is not rivers, but the atmosphere. With a knowledge of the atmospheric input function and with an understanding of the rates and mechanisms of in situ production or consumption, dissolved gases provide a powerful tool for understanding physical and chemical processes in the ocean.

Much of the early work on dissolved gases in sea water has been reviewed by Richards (1957, 1965). Kester (1975) has summarized the more current studies. Historically, oxygen has been the most widely studied gas in the ocean. Oxygen is the only gas routinely measured by physical oceanographers (along with temperature and salinity) in evaluating water mass distributions. The value it has as a water mass tracer results from the relatively large variations in concentration which it experiences due to biological production or consumption. The fact that a precise, standardized technique (Winkler 1888) has existed for over 90 years has made oxygen measurements both routine and reliable, and thus widely accepted in the

---

The style and format of this dissertation follows that of the journal Limnology and Oceanography.

oceanographic community. The less variable, non-reactive gases have been intensely studied in sea water only in the last two decades. The application of gas chromatography and mass spectrometry has enabled oceanographers to study gases with small variations or in extremely low concentrations. As more information is obtained on dissolved oceanic gases other than oxygen, it has been realized that dissolved gases can be important geochemical tracers for examining both transport processes and reaction mechanisms.

While numerous studies of dissolved gas distributions have been made in fresh water and sea water of normal salinity (20-40 parts per thousand), little work has been done in hypersaline environments. Weiss (1969) measured argon, nitrogen and total carbon dioxide on two samples from the hot Red Sea brines. Using his data and calculated solubilities, he inferred that the source water for the Red Sea brines originated in the shallow coastal areas or near-surface waters of the southern portion of the Red Sea. Swinnerton and Linnenbom (1969) reported on the low-molecular-weight hydrocarbons in the same Red Sea samples and found the values to be much lower than in other anoxic environments. Brooks et al. (1979) described the hydrocarbon and major ion distributions in the East Flower Garden Brine in the Gulf of Mexico. Sackett et al. (1979) discussed the implications of methane and total carbon dioxide data from the Orca Basin waters and sediment. These few papers represent the total of previous work concerning dissolved gases in

hypersaline, oceanic waters. None of these studies have taken a systematic approach to determine the processes governing both the reactive and unreactive gases in hypersaline waters. It is my purpose in this dissertation to present a systematic study of the geochemistry of reactive and unreactive gases in the hypersaline Orca Basin and to show how these gas distributions can be used in interpreting other geochemical evidence of brine formation, reaction and interaction.

#### Basic Concepts

The Earth's atmosphere is a semi-homogenous mixture of water vapor, major gases ( $N_2$ ,  $O_2$ , Ar and  $CO_2$ ), unreactive minor gases (e.g., Ne, He, Kr, and Xe), and unstable minor gases (e.g.,  $CH_4$ , CO,  $H_2$ , and  $N_2O$ ). The concentrations of these gases in the standard atmosphere are presented in Table 1. The solubility of an atmospheric gas in sea water is described by Henry's Law which relates the concentration of an ideal gas in solution ( $C^*$ ) to a constant times the partial pressure of the gas above the solution,

$$C^* = \beta P_G \quad (1)$$

where  $\beta$  is the Bunsen solubility coefficient (a function of temperature and salinity) and  $P_G$  is the partial pressure of the specified gas in the atmosphere. The ideality assumption is reasonable for most gases since most non-ideal variations are less than 0.2%

(Table 1).  $P_G$  is related to the mole fraction of gas ( $f_G$ ) in dry air by the expression

$$P_G = \left[ P_t - \frac{h}{100} P_{vp} \right] f_G \quad (2)$$

where  $P_t$  is the total pressure (atm),  $h$  is the relative humidity (percent), and  $P_{vp}$  is vapor pressure of the solution (atm). Combining Eq. 1 and 2 gives an expression which can be used to calculate solubilities,

$$C^* = \beta \left[ P_t - \frac{h}{100} P_{vp} \right] f_G \quad (3)$$

From Eq. 3, it becomes obvious that gas solubilities are a function of the Bunsen solubility, atmospheric fraction, vapor pressure, relative humidity, and total atmospheric pressure at the time the water parcel in question equilibrated with the atmosphere. Bunsen solubility coefficients have been accurately determined for most major gases (Weiss 1970, 1974), noble gases (Weiss 1971), and trace reactive gases (Yamamoto et al. 1976; Crozier and Yamamoto 1974; Gordon et al. 1977; Wiesenburg and Guinasso 1979). Atmospheric gas concentrations are relatively constant. While water vapor pressure can be calculated as a function of temperature and salinity, the appropriate barometric pressure and relative humidity when air and water equilibrate are not easy to obtain. Once a water parcel is out of contact with the atmosphere, it is difficult to determine the conditions under which it was equilibrated. It is,

Table 1. Abundance and properties of atmospheric gases.

Gas	Mole fraction in dry air* ( $f_G$ )	Molar volume at STP+ (liter·mol <sup>-1</sup> )	Percent difference from 22.414
N <sub>2</sub>	0.78080 ± 0.00004	22.391	0.10
O <sub>2</sub>	0.20952 ± 0.00002	22.385	0.13
Ar	(9.34 ± 0.01) × 10 <sup>-3</sup>	22.386	0.12
CO <sub>2</sub>	(3.3 ± 0.1) × 10 <sup>-4</sup>	22.296	0.53
Ne	(1.818 ± 0.004) × 10 <sup>-5</sup>	22.421	0.03
He	(5.24 ± 0.004) × 10 <sup>-6</sup>	22.436	0.10
CH <sub>4</sub>	1.41 × 10 <sup>-6</sup>	22.356	0.26
Kr	(1.14 ± 0.01) × 10 <sup>-6</sup>	22.350	0.28
H <sub>2</sub>	0.58 × 10 <sup>-6</sup>	22.430	0.007
CO	0.11 × 10 <sup>-6</sup>	22.387	0.12
N <sub>2</sub> O	5 × 10 <sup>-7</sup>	22.228	0.56
Xe	(8.7 ± 0.1) × 10 <sup>-8</sup>	22.277	0.61

\* From Glueckauf (1951), Valley (1965), and Schmidt (1978).

+ Based on van der Waals equation for gases using constants from Weast (1969).

therefore, reasonable to assume a standard pressure of one atmosphere and a relative humidity of 100%. With these simplifying assumptions, Eq. 3 reduces to:

$$C^* = \beta(1 - P_{vp}) f_G \quad (4)$$

which is the equation most often used to express the atmospheric equilibrium solubility of a gas in solution.

Weiss (1970) has developed an equation to describe atmospheric equilibrium solubility from moist air as a function only of temperature and salinity. He expressed the temperature dependence of solubility at constant salinity by an integrated form of the van't Hoff equation and used the Setchénow relation to describe the salinity effect. Combining these two relationships and adding an additional term to account for the vapor pressure of water, he developed an atmospheric solubility equation of the form

$$\ln C^* = A_1 + A_2 (T/100) + A_3 \ln (T/100) + A_4 (T/100) + S\text{‰} [B_1 + B_2 (T/100) + B_3 (T/100)^2] \quad (5)$$

where  $C^*$  is the equilibrium solubility in  $\text{nmol}\cdot\text{liter}^{-1}$  (or other appropriate units),  $A_1$  and  $B_1$  are constants,  $T$  is the absolute temperature,  $S\text{‰}$  is the salinity in per mil. Weiss (1970, 1971, 1974) and Weiss and Kyser (1978) have fitted solubilities for oxygen, nitrogen, argon, neon, helium, carbon dioxide and krypton to Eq. 5



to describe the atmospheric solubility of those gases relative to water saturated air at one atmosphere total pressure. Recently, Wiesenburg and Guinasso (1979) have rewritten Eq. 5 expanding the  $A_1$  term to  $f_g + A_1$ , to include the atmospheric gas concentration as a variable. They have fitted solubility data for methane, hydrogen and carbon monoxide to the modified equation. All atmospheric solubility calculations in this work will be made using Eq. 5 or the modified version of Wiesenburg and Guinasso (1979).

#### The Study Site

The Orca Basin is located on the Louisiana continental slope in the Gulf of Mexico and is centered at 26°55'N, 91°20'W. In this region, salt tectonics have had an observable influence on the topography. Salt diapirs protrude almost to the surface from basement salt deposits in many areas near the Orca Basin. Salt flow and dissolution have created a relatively unstable sea-floor which has become pock - marked with intraslope basins (Fig. 1). The Orca Basin is but one of many of these types of basins which are situated within the growing margin of the Gulf Coast geosyncline.

The bathymetry of the Orca Basin is shown in Fig. 2. The basin itself encompasses an area of approximately 400 km<sup>2</sup> with an "L"-shaped trough along the eastern and southern portions. Enclosing slopes drop from an average depth of 1800 meters along the encircling continental slope to over 2400 meters within two separate

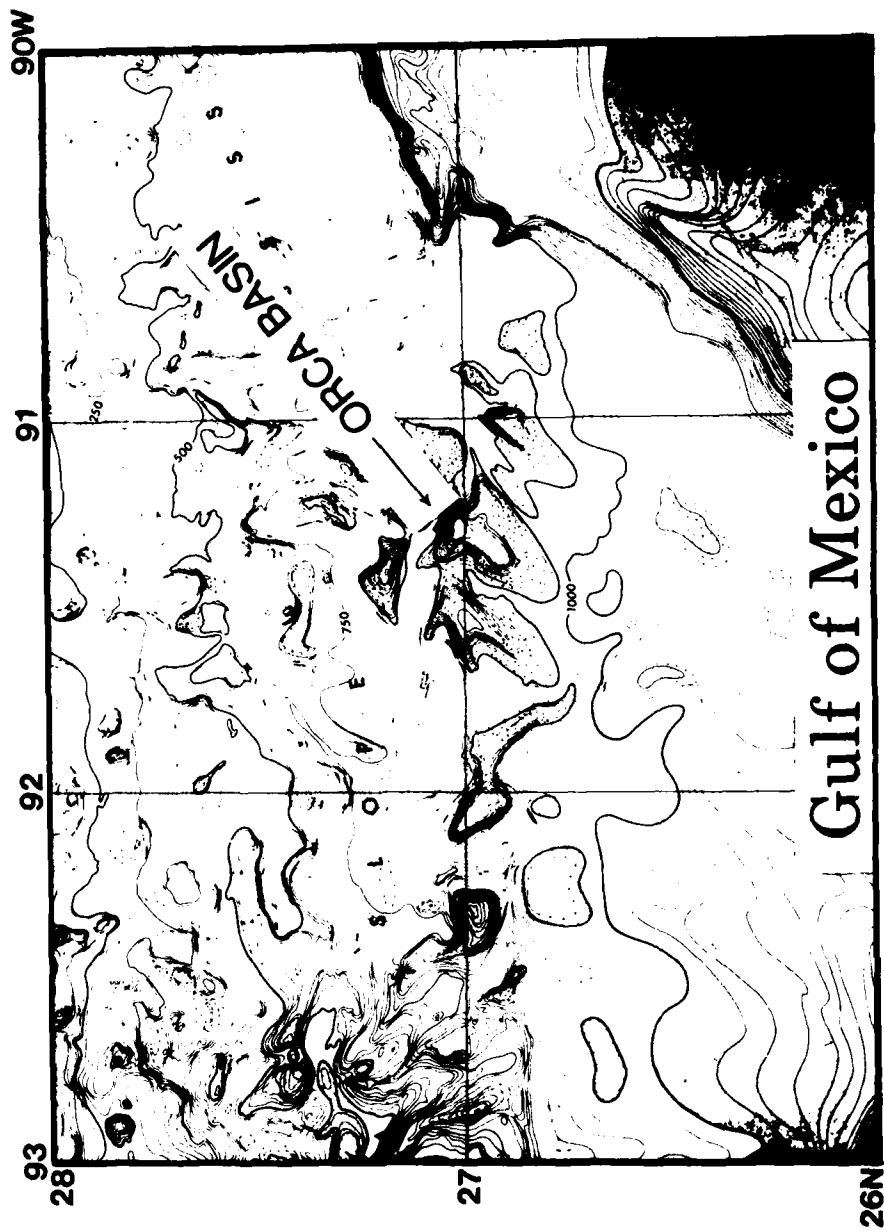


Fig. 1. Index map of the Texas-Louisiana continental slope showing several small basins which have resulted from salt flow and dissolution. Location of the Orca Basin is indicated. Depth contours are in fathoms.

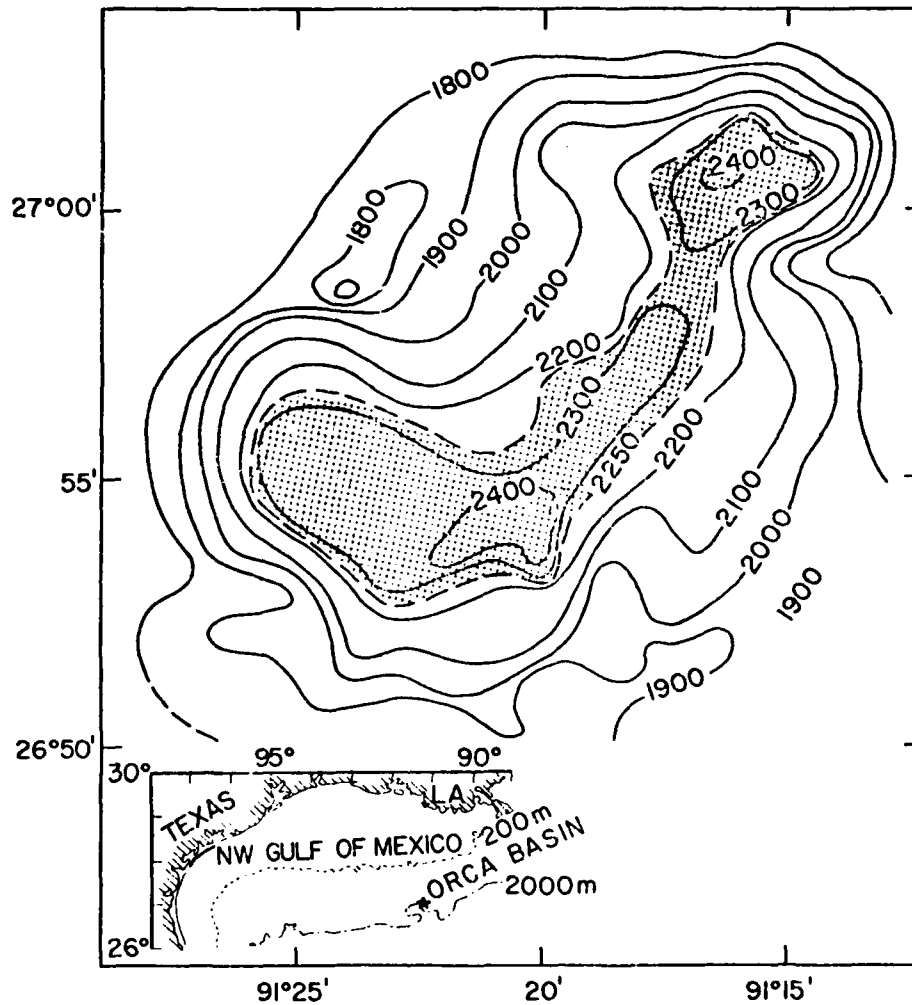


Fig. 2. Bathymetric map of the Orca Basin based on the seismic data taken on cruise 76-G-10 of the R/V Gyre (after Shokes et al. 1977). The stippled portion of the map represents the area covered by the high salinity brine.

areas. The bottom 200 meters of the Orca Basin is filled with water having an evaporative salinity of 250‰ and a temperature (5.6°C) more than 1.4°C higher than the overlying, freely circulating Gulf of Mexico water. The volume of brine in the Orca Basin is approximately 5 km<sup>3</sup>, a volume several times larger than all of the Red Sea brines combined.

The temperature and salinity structure in the Orca Basin is shown in Fig. 3. Bottom water at 2000 meters in this area of the Gulf of Mexico has a characteristic temperature of 4.2°C and an average salinity of 34.976‰. Both the temperature and salinity begin to increase rapidly in the Orca Basin at a depth of approximately 2240 meters, with the salinity gradient being more dramatic than the temperature gradient. The sea water to brine transition zone from normal Gulf of Mexico deep water to maximum salinity brine is only about 50 to 60 meters and provides a very sharp density contrast. The density difference of the brine (1.185 kg/l) as compared to the deep Gulf water (1.025 kg/l) provides such a sharp interface that it can be seen using low-frequency acoustical sounding devices. Shokes et al. (1977) used a 700-joule minisparker to generate low frequency sound pulses in order to map the extent of the brine. Part of the record that they obtained in that study is shown in Fig. 4. The brine appears as a distinct layer due to the sound reflection from the interface. On cruise EN-32 of the R/V Endeavor, the brine interface could also be seen by

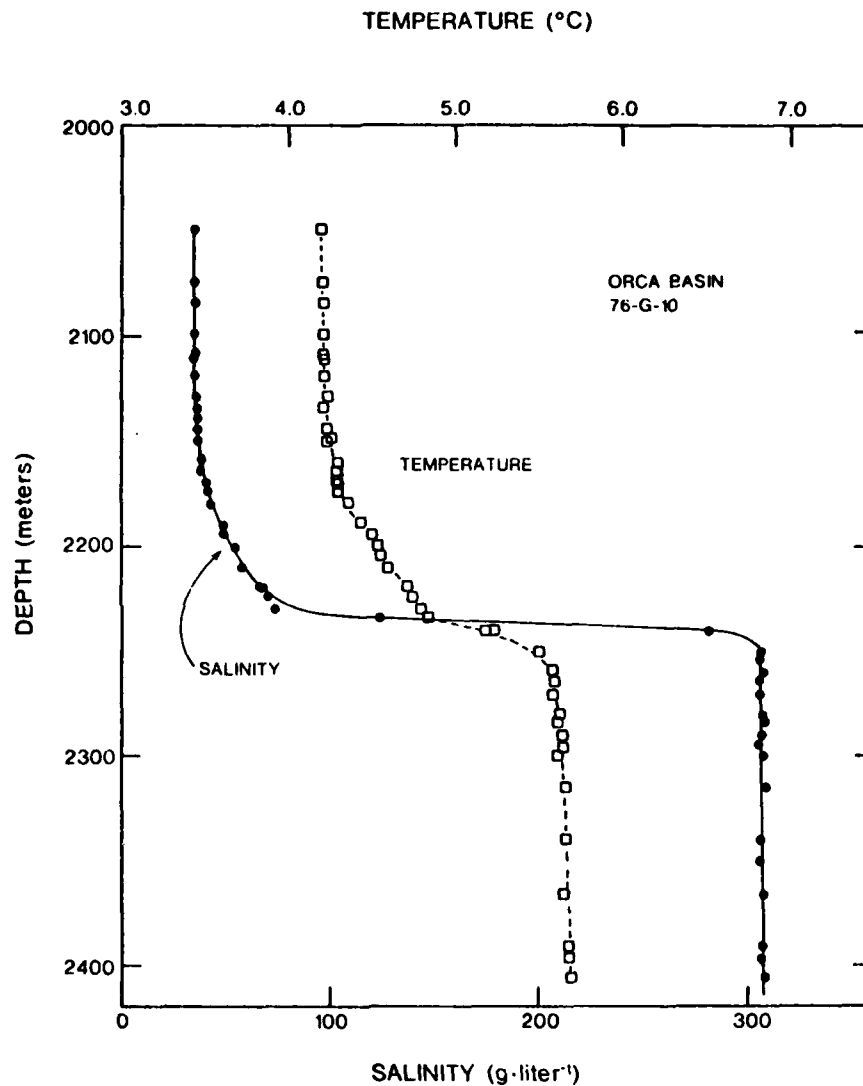


Fig. 3. Vertical temperature and salinity ( $\text{g}\cdot\text{liter}^{-1}$ ) profiles extending from the deep Gulf of Mexico water into the hypersaline Orca Basin brine. Note the sharp transition between the Gulf water ( $\rho = 1.025$ ) and the brine ( $\rho = 1.185 \text{ kg}\cdot\text{liter}^{-1}$ ).

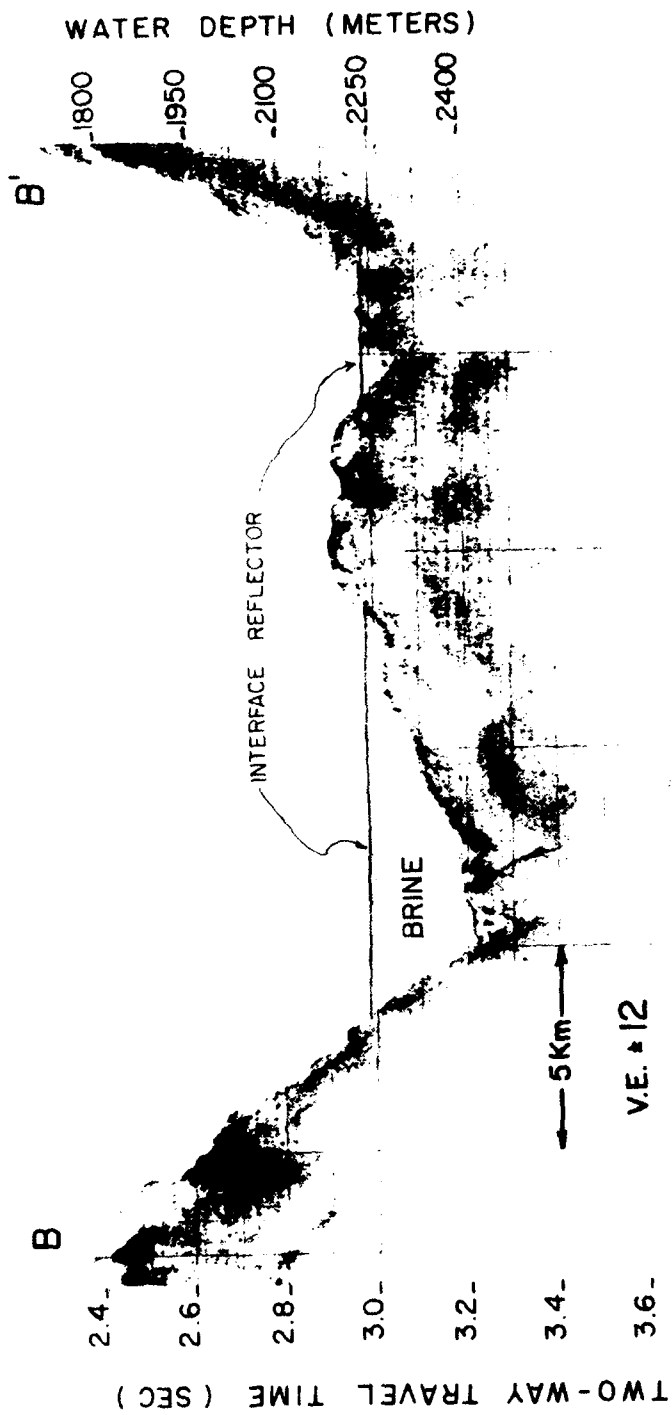


Fig. 4. One section of the minisparker record obtained by Shokes et al. (1977) during their mapping of the Orca Basin brine pool. The dark level line above the bottom is the result of sound reflections from the density interface.

using a 3.5 kHz bottom profiling system, which has a higher frequency than the minisparker of Shokes et al. (1977). This device was instrumental in positioning the ship above the brine for sampling. The brine-sea water interface could not be seen with the higher frequency 12 kHz bottom profiling system.

### Previous Investigations

#### The Early Work

The geochemical investigation of the Gyre Basin in the northwestern Gulf of Mexico (Bouma et al. 1975) prompted researchers at Texas A&M University to perform surveys of several other intraslope basins on the Texas and Louisiana continental slopes. Most of the other intraslope basins appear to be geochemically "normal" basins similar to the Gyre Basin (C. R. Schwab, personal communication). Only the Orca Basin was found to have water with above normal salinities.

The Orca Basin was first sampled as part of the intraslope basin study during cruise 75-G-16 of the R/V Gyre in November 1975. During this cruise, a 185-190 cm gravity core was taken from the shoaler northern end of the basin (Fig. 2). After the core was sliced and the interstitial water removed by squeezing, the water (brine) was placed in a freezer but it would not freeze completely! Examination of the interstitial fluid from this core revealed that the salinity was greater than 250‰ at a depth of 8 cm and

decreased slightly with depth in the core (McKee and Sidner 1976). Although similar hypersaline conditions had been found in deep borehole samples from the Gulf of Mexico (Manheim and Bischoff 1969), this was the first observation of hypersaline fluids in any open continental shelf or slope environment. With excitement running high the brave men of the Gyre returned to the Orca Basin on cruise 76-G-2 in early 1976. A second gravity core was taken in the deeper southern portion of the basin and water samples were taken to characterize the water in the basin. They noted that the basin had a local sill depth of 2000 m, with a 140 m thick bottom layer of anoxic bottom water with a salinity of 270‰. The observations from these two cruises have been briefly summarized by McKee and Sidner (1976). The first water column data (Table 2), a description of the sediment features which result from the unusual environment was described. Since their observations are relevant to all future studies, this author will briefly detail them here:

1. The sediments are high in organic matter.
2. The presence of fine laminations reflect the lack of burrowing organisms.
3. Sediments from the basin are black due to the presence of metastable metal sulfides which rapidly oxidize to a red brown color upon exposure to air.



Table 2. First report of hydrographic data from the Orca Basin  
(Tabulated from McKee and Sidner 1976).

<u>Depth</u>	<u>Temperature</u> ( °C )	<u>Salinity</u> ( ‰ )	<u>Oxygen</u> ( ml · liter <sup>-1</sup> )
S111	4.21	35.076	4.839
S111 + 100m	4.38	43	1.769
S111 + 200m (140m above bottom)	5.63	270	0.0

4. There was no noticeable hydrogen sulfide odor in the water or sediments.
5. The deeper water samples appeared gassy and the X-ray radiographs of the cores showed small gas bubbles which increased in concentration near the surface of the core.
6. The sediments show excellent organic preservation as evidenced by the presence of Sargassum throughout the entire length of the gravity cores.

These observations (McKee and Sidner 1976) were preliminary and there were no interpretations of results given.

The first study to make detailed observations of the water column chemistry was done by Shokes et al. (1977), using data obtained during R/V Gyre cruise 76-G-10 in October 1976. This work led to the landmark paper which first showed the extent of the Orca Basin and gave it its name. Shokes et al. (1977) did a complete bathymetric survey of the basin using a low frequency minisparker and estimated the brine volume to be  $40 \text{ km}^3$ . They made the first extensive geochemical analysis of the anoxic brine in the Orca Basin. In this analysis, detailed vertical measurements of temperature, salinity, oxygen, and nutrients were made in both ends of the basin (Fig. 5 and 6). The major ions in the brine were also measured and these values were compared to the Red Sea geothermal brines (Table 3). Shokes et al. (1977) concluded that the brine must originate by solution (presumably by

Table 3. Moles per kilogram of ions in sea water and various brines.

	<u>Na</u>	<u>K</u>	<u>Mg</u>	<u>Ca</u>	<u>Cl</u>	<u>SO<sub>4</sub></u>	<u>HCO<sub>3</sub></u>
Sea water *	0.478	0.0102	0.0531	0.0102	0.547	0.0283	0.0022
Orca Basin brine †	3.980	0.0161	0.0432	0.0272	4.217	0.0381	0.0053
Dead Sea brine ‡	1.596	0.163	1.559	0.493	5.684	0.0070	0.0034
Red Sea brine §	4.041	0.0552	0.0333	0.118	4.380	0.0078	0.0023
East Flower Garden	3.449	0.0093	0.0439	0.0403	3.695	0.0460	0.0024

\* Average sea water

† Shokes et al. (1977)

‡ Amit and Bentor (1971)

§ Brewer et al. (1969)

|| Brooks et al. (1979)

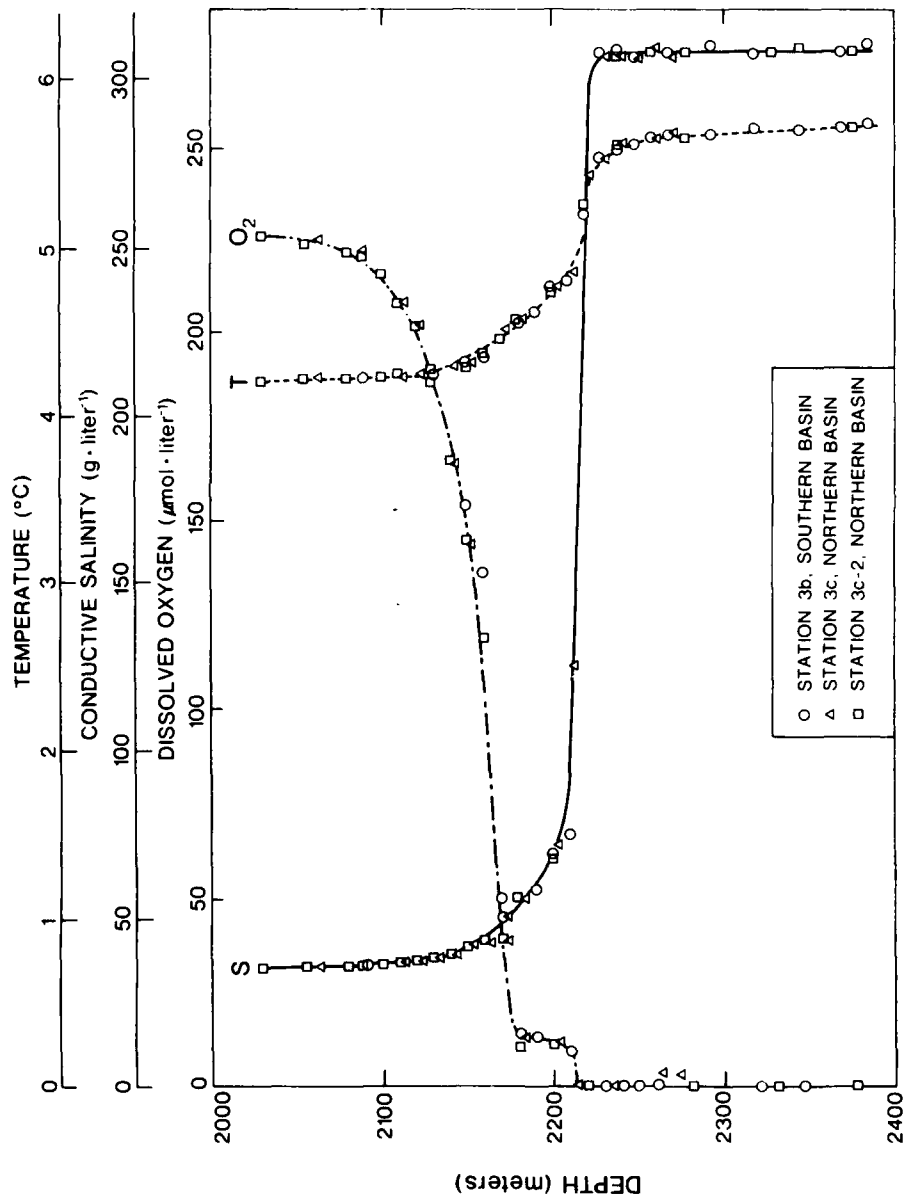


Fig. 5. Composite vertical profiles of temperature, shipboard conductive salinity and dissolved oxygen measured in both lobes or ends of the Orca Basin on R/V Gyre cruise 76-G-10 (after Shokes et al. 1977).

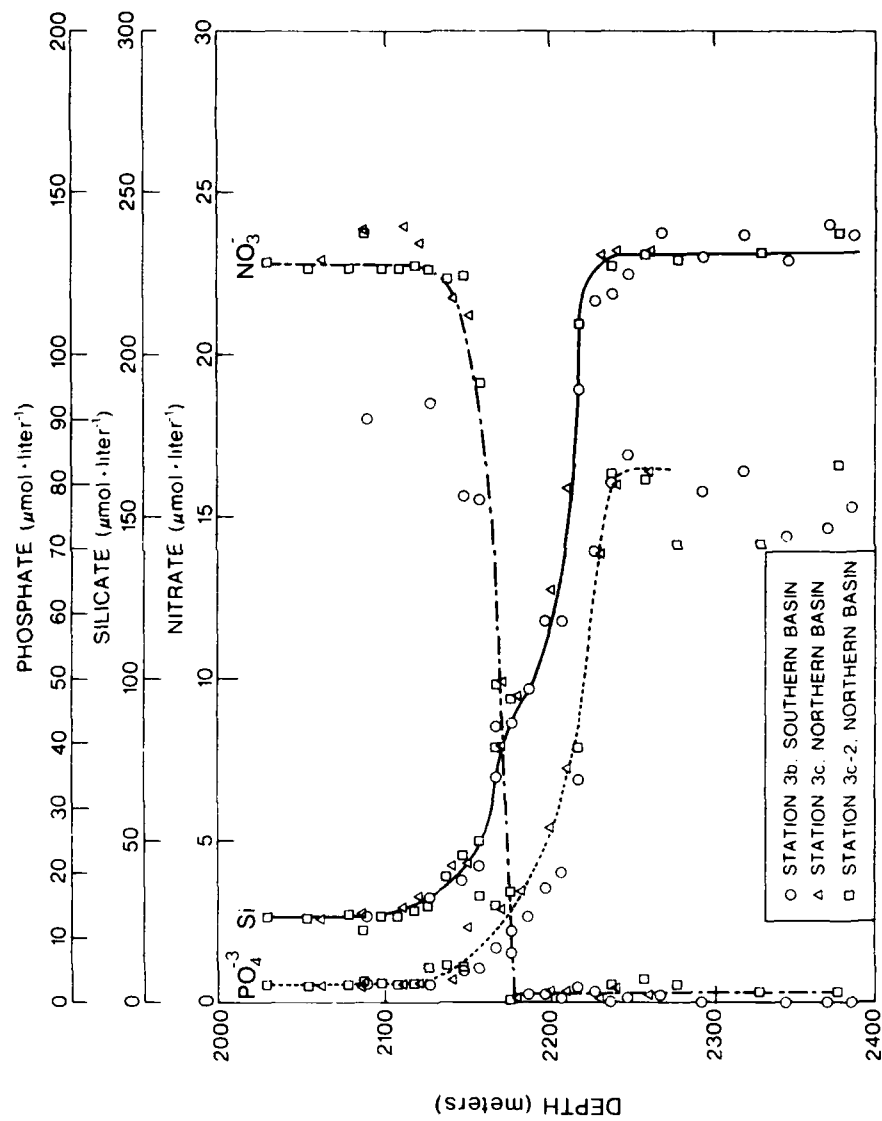


Fig. 6. Composite vertical profiles of dissolved orthophosphate, silicate and nitrate across the sea water-brine interface for three stations in the Orca Basin (after Shokes et al. 1977).

sea water) of a near-surface salt deposit (mainly NaCl). They reached this conclusion based on the absence of sulfate depletion in the brine and due to observations of minimal changes in temperature and ion compositions. Since there was a decreasing chloride gradient in the interstitial water of the basin sediment, it was suggested that the brine enters the basin laterally, rather than by diffusion upwards through the underlying sediments. Calculations of the heat input needed to raise the temperature of the  $40 \text{ km}^3$  brine from the overlying Gulf temperature of  $4.2^\circ\text{C}$  to the present  $5.6^\circ\text{C}$  could be added in about 325 years, assuming an average oceanic geothermal heat flow ( $1 \text{ } \mu\text{cal}\cdot\text{cm}^{-2}\cdot\text{sec}^{-1}$ ). The only dissolved gas measured was oxygen, which decreased to zero within the brine.

In a follow-up paper, Trabant and Presley (1978) presented a more detailed geological description of the Orca Basin. The brine volume estimate was lowered to  $5 \text{ km}^3$ . The geological setting of the Orca Basin was compared to that of several other intraslope basins which have been defined and discussed on the basis of seismic reflection profiles (Antoine and Bryant 1969; Garrison and Martin 1973). It was concluded that geologically the Orca Basin represents a typical intraslope basin. However, it has water with uncommon chemical composition and associated sediment geochemistry because of the presence of regional salt massifs which have led to salt flow into the brine. Trabant and Presley

(1978) examined a three meter gravity core taken in the deeper end of the basin and came to the same conclusions as McKee and Sidner (1976), viz. the absence of laminae and apparent textural homogeneity indicated pelagic deposition and no burrowing by organisms. The presence of Sargassum throughout the core was also noted. A complete description of the core was produced and the following sediment properties were discussed:

1. The sediment was a silty clay with a clay size fraction of 60-75%, with little or no vertical change of clay mineralogy.
2. The acid-leachable iron content is similar to other Gulf sediments, but with a relatively high sulfur content (1%) compared to other Gulf of Mexico sediments.
3. Water content of the sediment ranged from 140% to 280% (dry weight method).

In discussing the brine geochemistry in the Orca Basin, Traubant and Presley (1978) make one significant new note. They measured the bromide level in the brine and found a value of  $80 \text{ mg} \cdot \text{kg}^{-1}$ . Due to the well-known partitioning effects associated with bromine as a salt solution evaporates, bromine can be useful in determining the origin of the brine (see later discussion on bromine). LouAnn Salt (a logical source for salt in the Orca Basin) was observed to have 34 to 51 ppm bromide (Holser 1966).

Dissolving 308 g of this salt in a liter of sea water would give a bromide concentration equivalent to that found in the Orca Basin. Thus, Trabant and Presley (1978) offered strong evidence that the Orca Basin brine forms from the dissolution of previously deposited salt.

In the same volume, a paper on basin and slope sediments by McKee et al. (1978) further emphasized this point, while comparing the sediment geochemistry of intraslope basins with the geochemistry of shelf and slope sediments. In this same study, they noted that the more rapid sediment accumulation rate in intraslope basins caused basin sediments to have a more anoxic character than typical slope sediments. Of the basins studied, only the Orca had brine in the bottom. However, one other basin — the Gyre Basin — exhibited a higher concentration of salt in one of the cores that were studied (up to  $44.2 \text{ g}\cdot\text{kg}^{-1}$  of chloride compared to  $19.4 \text{ g}\cdot\text{kg}^{-1}$  for average sea water). The salinity in this basin core increased with depth in the sediment indicating an upward diffusion of salt from some deeper formation.

Another significant feature of the study by McKee et al. (1978) was a comparison of sulfate profiles of the normal slope sediments, intraslope basin sediments, and the Orca Basin sediments. Even though the sedimentation rate in the Orca Basin is three times as great as found on the slope, the sulfate gradient from the upper 120 cm of two Orca Basin cores was not significantly greater



than gradients found in most slope sediments. In one other Orca Basin core (9B), the slope of the sulfate profile indicated a  $dc/dx_0$  of  $0.27 \text{ mmol} \cdot \text{liter}^{-1} \cdot \text{cm}^{-1}$  which is higher than the value of  $0.02 \text{ mmol} \cdot \text{liter}^{-1} \cdot \text{cm}^{-1}$  found outside the basin (but still lower than some values found in the Gyre Basin which is not an anoxic basin). The variability of sulfate gradients in the cores from the Gyre and Orca Basins correlated with patterns of sediment redeposition within the basins. More evidence of slumping was found in Gyre Basin cores. However, those cores were more extensively studied than the Orca Basin cores.

These initial studies on the Orca Basin were primarily concerned with defining the origin of the brine. Two basic lines of evidence were pursued: the decreasing chloride gradient in the sediments and the bulk ion composition. The decreasing salt concentration with depth in the sediments is strong support for the salt influx coming from other than the basin floor. However, the ion composition is also an important piece of evidence in deciphering this puzzle. The ionic composition of the Orca Basin brine as reported by Shokes et al. (1977) is compared with standard sea water and several other oceanic brines in Table 3. Unlike evaporation brines such as the Dead Sea brine, the Orca Basin brine does not have tremendously increased concentrations of any ions other than sodium and chloride. This fact, and more recent evaluation of bromide data from the Orca Basin by McKee et al.

(1978), both support the contention that the Orca Basin formed as the result of the dissolution of a salt deposit. The bromide concept is important enough to describe in more detail.

Bromide does not form its own minerals when sea water evaporates. Some of it is lost from solution because it forms isomorphous admixture with chloride in the precipitates (Valyashko 1956; Braitsch and Herrman 1963). As sea water evaporates, the carbonates precipitate first, followed by the sulfates. During this stage of evaporation, little or no bromide precipitates, or if it does, it is occluded with the carbonates and sulfates (see Table 4).

Halite (NaCl) begins to precipitate when the chloride concentration reaches about  $275,000 \text{ mg}\cdot\text{liter}^{-1}$  (cf. normal sea water,  $19,000 \text{ mg}\cdot\text{liter}^{-1}$ ). Some small amount of bromide is entrained with chloride in the precipitate. However, as crystallization proceeds, more bromide is left in solution than is included in the precipitate. Sylvite (KCl) begins to precipitate when the chloride concentration has risen to about  $360,000 \text{ mg}\cdot\text{liter}^{-1}$ . Carnallite ( $\text{MgCl}_2\cdot\text{KCl}\cdot 6\text{H}_2\text{O}$ ) and bischoffite ( $\text{MgCl}_2\cdot 6\text{H}_2\text{O}$ ) precipitates follow. During evaporation the concentration of bromide in solution increases. Therefore, relative to chloride, the bromide concentration in a brine is a good indicator of the degree of sea water concentration. The above discussion assumes that appreciable quantities of biogenic bromide have not been introduced into the system. McKee et al. (1978) have discussed this idea and

Table 4. Concentration changes during evaporation of sea water and brine (after Collins 1969).

Element	Sea water	CaSO <sub>4</sub> ↓	NaCl↓	MgSO <sub>4</sub> ↓	KCl↓	MgCl <sub>2</sub> ↓
Lithium	0.2	2	11	12	27	34
Sodium	11,000	98,000	140,000	70,000	13,000	12,000
Potassium	350	3,600	23,000	37,000	26,000	1,200
Rubidium	0.1	1	6	8	14	10
Magnesium	1,300	13,000	74,000	80,000	130,000	153,000
Calcium	400	1,700	100	10	0	0
Strontium	7	60	10	1	0	0
Boron	5	40	300	310	750	850
Chloride	19,000	178,000	275,000	277,000	360,000	425,000
Bromide	65	600	4,000	4,300	8,600	10,000
Iodide	0.05	2	5	7	8	8
Total		295,000	517,000	469,000	538,000	602,000

illustrated it with a diagram (reproduced here as Fig. 7) that shows bromide versus chlorinity for various pore waters including the Orca Basin. As sea water evaporates, the chloride versus bromine distribution is represented by the solid line which separates the areas resulting from dissolving, evaporate-produced brine from residual source-solution brines. The Orca Basin is higher in chloride content without a significant increase in the bromide concentration indicating a solution source. This data seems to clearly establish the source of the Orca Basin as a dissolving evaporite deposit rather than a brine formed by direct evaporation.

#### Subsequent Investigations

These papers caused a flurry of activity in the oceanographic community. Besides cruises by Texas A&M University, expeditions to the Orca Basin have been staged by the University of Miami, the University of Texas and the University of Rhode Island. In the Fall 1977 American Geophysical Union meeting, seven papers were presented dealing with various geochemical aspects of the Orca Basin (see, for example, Guinasso 1977).

Of the papers now published from the initial cruises to the basin, I recognize two major categories: sediment geology and organic-microbial interactions in the brine. The sediment papers have been produced by geologists searching for the source of salt or are attempting to describe the unusual effects that the high

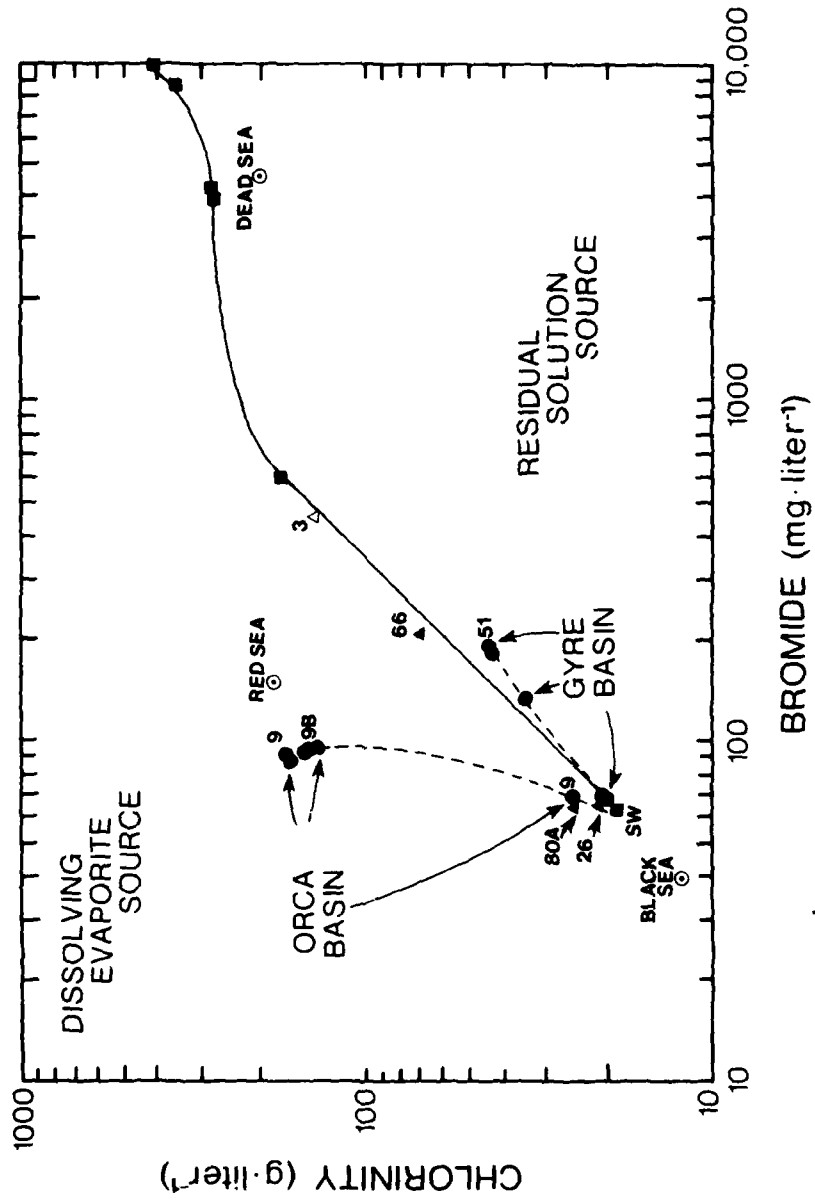


Fig. 7. Plot of bromine versus chlorinity for pore waters with chlorinity anomalies. The solid curve represents residual solution as sea water progressively evaporates, dividing two fields representing a dissolving evaporite source (above line) and a residual solution source (below line). Pore water data for the Orca Basin, Gyre Basin and Dead Sea are indicated.

salinity water has on various sediment properties (e.g., sedimentation rate, clay particle interaction, and preservation of carbonate and silicious materials). Kennett and Penrose (1978) used electron microscopy to examine the microfauna in the sediments. They noted that pteropod tests were extremely well preserved relative to other areas. Additionally, they noted the unusual occurrence of well preserved Sargassum particles deep in the core. While Sargassum has been found at the sediment surfaces as deep as 5300 m (Schoener and Row 1970), Kennett and Penrose (1978) noted that there are no previously documented cases of preserved seaweed in deep-sea sediments, especially with attached organisms as found in the Orca Basin.

Addy and Behrens (1980) also reported the presence of seaweed in their ten meter core from the knoll between two basins. They did a very complete study of the major ions in the interstitial water and for each 10 cm section of the core, determined grain size distributions. From radiocarbon dates, they determined that the bottom layer of the black mud in the sediments (which corresponds to the initial time of brine input to the basin) had an age of  $7910 \pm 170$  years. This age was reported at a depth of 4.85 meters which gave an implied sedimentation rate of  $61 \text{ cm} \cdot \text{ka}^{-1}$ . Measurements further down in the core in areas which were not deposited under anoxic (brine) conditions gave a sedimentation rate for that region of  $43 \text{ cm} \cdot \text{ka}^{-1}$ . This difference implies an

increased sediment accumulation rate due to the anoxic conditions existing in the brine. Carbonate percentages averaged four percent higher in the black mud laid down under anoxic conditions than in the gray mud that had an oxygenated origin. From this data, Addy and Behrens (1980) concluded that the enhanced sediment accumulation is due to increased preservation of carbonate test. The higher sand fractions in the basin core were almost entirely skeletal carbonate.

Besides the color of the mud in the sediments, Addy and Behrens (1980) used the presence of Sargassum fronds to ascertain the conditions (anoxic or aerobic) under which the sediments were deposited. Aerobic conditions allow more degradation of organic matter than in anoxic environments. The absence of recognizable Sargassum in Orca Basin gray muds indicates deposition in an oxygenated environment; while the well preserved Sargassum fronds in the black mud strongly suggest deposition in anoxic brine.

Addy and Behrens (1980) made several other measurements to determine the conditions of deposition, including water content (higher in the anoxic mud) and compaction (found to be less in the black mud sediments). Deposition in a brine basin would account for both of these observations.

The color of the black mud is due to very fine-grained, metastable iron sulfides such as greigite ( $\text{Fe}_3\text{S}_4$ ) and machinawite ( $\text{FeS}$ ), like those found in anoxic sediments of the Black Sea

(Berner 1974). Formation of metastable iron sulfides instead of pyrite ( $\text{FeS}_2$ ) will result in the absence of significant  $\text{H}_2\text{S}$  (from  $\text{SO}_4$  reduction) in the depositional environment. This observation has important implications for the work to be presented later.

Addy and Behrens (1980) approached the questions of brine origin by conducting seismic profiles of the slope area. If the Orca Basin brine is formed from dissolution of an exposed salt diapir, that feature should be found on the surrounding basin slope. In two multichannel subbottom profiles, Addy and Behrens (1980) noted several strong undulating reflectors with diapiric structures which they interpreted as salt surfaces. These salt surfaces become shallower to the north and possibly outcrop at the edge of the small basin east of the Orca Basin. A second salt structure similarly approaches the surface at the eastern margin north of the Orca Basin. A 3.5 kHz profile of the region showed hyperbolic reflections indicating a hummocky surface. Addy and Behrens noted that if salt were exposed, the dissolution would proceed along fractures and joints resulting in the uneven surfaces that were observed. Perhaps Addy and Behrens (1980) have located the source of the Orca Basin brine.

Another study dealing with sediment geology was that of Tompkins and Shephard (1979). They described possible depositional processes, geotechnical properties and clay mineralogy of two cores from the Orca Basin. One core was taken in the deepest



portion of the southern basin and gave sedimentation rates of over  $100 \text{ cm} \cdot \text{ka}^{-1}$ , based on microfaunal examinations. The conditions within the core suggested that the sediment was originally deposited under brine on the basin slope, and then slumped deeper into the basin. Addy and Behrens (1980) had avoided problems related to slumping by sampling only on the knoll between the two basins.

Tompkins and Shephard (1979) examined the clay mineralogy and clay mineral variations in the basin sediments. They noted an increase in smectite with increasing depth (and decreasing salt) in the sediments. They attributed the highly open sediment microstructure to the rapid sedimentation. Both the sediment microstructure and the levels of smectite seemed to control the diversity in clay mineral abundances in the brine. Sediments that were deposited outside the brine, then slumped into the basin, were easily discernable by their properties which were similar to normal continental slope sediments.

In a study which encompassed both water column and sediment data, Sackett et al. (1979) developed a carbon budget for the Orca Basin water column and sediments. Methane and carbon dioxide increased with depth in the sediments and both were assumed to result from the bacterial decomposition of organic matter. The  $\delta^{13}\text{C}$  values of carbon dioxide and methane in the sediments were very negative (isotopically lighter) relative to water column

values. The value of  $-105\text{‰}$  that Sackett et al. (1979) observed for the  $\delta^{13}\text{C}$  of methane is the lightest (natural) methane isotope ratio reported to date. The amounts of organic carbon found in the sediments were somewhat higher than for normal shelf and slope sediments. The  $\delta^{13}\text{C}$  values of the particulate organic carbon in the sediments ranged from  $-22\text{‰}$  (a normal value for open shelf sediments) down to a  $-26.7\text{‰}$ . They suggested that the lighter samples may have resulted from slumped material originally deposited in estuarine environments during the Pleistocene.

In the brine itself, Sackett et al. (1979) made concentration and isotopic ( $\delta^{13}\text{C}$ ) measurements of carbon dioxide, methane, and organic carbon. Sackett et al. (1979) used a mass balance equation of the following form:

$$\begin{aligned} &[(\text{mg C/L})(\delta^{13}\text{C})]_{\text{sea water}} + [(\text{mg C/L})(\delta^{13}\text{C})]_{\text{added carbon}} \\ &= [(\text{mg C/L})(\delta^{13}\text{C})]_{\text{brine}} \end{aligned} \quad (6)$$

to calculate the amount of isotopic composition of added carbon that would be required (starting from sea water) to produce the values for methane and carbon dioxide they observed in the brine and sediments. These calculations revealed that the carbon added to the Orca Basin had a concentration of  $27.4 \text{ mgC}\cdot\text{liter}^{-1}$  with a  $\delta^{13}\text{C}$  of  $-33\text{‰}$ . Sackett et al. (1979) were also able to partition the  $27.4 \text{ mgC}\cdot\text{liter}^{-1}$  between carbon dioxide added directly (via

fermentation) and carbon dioxide that had been cycled through an anaerobic methane consumption pathway ( $\text{CH}_4 \rightarrow \text{CO}_2$ ). Their estimates from these contributions were  $26.3 \text{ mgC}\cdot\text{liter}^{-1}$  from interstitial total carbon dioxide and  $26.3 \text{ mgC}\cdot\text{liter}^{-1}$  of carbon dioxide from methane oxidation. They suggested that the observed isotopic ratios of carbon can be explained by some complex mixture of three bacterially catalyzed processes: carbon dioxide fermentation, acetate fermentation, and sulfide reduction (encompassing methane consumption).

Realizing the importance of bacteria in an anoxic environment such as the Orca Basin, LaRock et al. (1979) examined bacterial activity in the brine. Total biomass was measured in both the deep Gulf water and in the brine using direct counts of bacteria, ATP levels, and uridine uptake. ATP concentration and uridine uptake both decreased just below the interface and then increased with increasing depth. This pattern was similar to the ATP distribution in the Cariaco Trench (Karl et al. 1977). In the deepest sample from the brine an ATP value of  $15.4 \text{ ng}\cdot\text{liter}^{-1}$  was found. This high level indicates that an active microbial population has developed within the brine of the Orca Basin.

The most interesting pattern shown by the microbial data was at the brine/sea water interface. Both ATP levels and uridine uptake values displayed similar patterns in the interface region. They increased in magnitude (relative to deep Gulf levels) above

the interface but decreased below the interface in the anoxic zone. This pattern, showing the existence of an interface population (or ATP subsurface maximum) at the brine/sea water interface, is consistent with the concept of particles resting on the density gradient. Bacteria and particles are closely associated in the deep ocean and conditions at the interface are well suited to the development of an autotrophic bacterial population. The anoxic zone provides a source of both nutrients and other energy yielding substrates (by diffusion upward from the brine) and could provide a rich medium for bacterial reproduction.

Ignoring the decomposition processes that had intrigued most other workers, Millero, et al. (1979) concentrated on the physical-chemical properties of the brine. They measured conductivity, density, and the speed of sound in the brine and in various brine dilutions. The speed of sound in the brine is 16% greater in the brine than in sea water with a salinity of 35‰. This finding allows us to calculate the depth of the basin. The high sound velocity implies that the basin is deeper, and thus larger, than previously estimated by Shokes et al. (1977).

Millero et al. (1979) also found that salinity and density (under standard conditions) were consistent within the anoxic zone of the basin. These density and salinity measurements were done with exceptional care and corroborated the earlier data of Shokes et al. (1977). Millero et al. (1979) used the density measurements

to examine the physio-chemical nature of the brine. The ionic composition of the brine was used in additivity methods (of partial and molal volumes) to estimate the density of the brine. Measured values agreed with these estimates to within  $\pm 280 \times 10^{-6} \text{ g}\cdot\text{cm}^{-3}$ , from 15° to 35°C. Since this agreement was good, Millero et al. (1979) suggested that it seemed possible to make additional estimates of physio-chemical properties of brines using additivity methods.

#### Anaerobic Decomposition Processes

The salt that has flowed into the depths of the Orca Basin has created a large density gradient as was discussed earlier. Table 5 presents a list of various basins, both oxygen-bearing and anoxic, along with the stability of the water column in each basin. The hypersalinity in the Orca Basin has given the water in the basin water a stability that is nine orders of magnitude greater than the major anoxic basins in the ocean.

The stability of the Orca Basin has produced an isolated system removed from the general circulation of the oceans which provide renewal waters to other areas. With an absence of fresh, oxygenated water flowing into this geochemical system, oxygen becomes depleted and anaerobic decomposition processes begin. Little if any flushing can occur across its density gradient. The extreme stability of the Orca Basin thus makes it an ideal

Table 5. Approximate stability across the sills of some basins and fjords (after Richards 1965).

BASIN	$\Delta\sigma_T/\Delta Z \times 10^{-3} \text{ m}^{-1}$
<u>OXYGEN-BEARING BASINS</u>	
Gulf of Mexico	$2 \times 10^{-6}$
Catalina Basin	$3.1 \times 10^{-6}$
Santa Barbara Basin	$4.4 \times 10^{-6}$
Santa Monica Basin	$8.8 \times 10^{-6}$
<u>ANOXIC BASINS</u>	
Kaoe Bay	$2.4 \times 10^{-5}$
Cariaco Trench	$3.2 \times 10^{-5}$
Gulf of Cariaco	$3.5 \times 10^{-5}$
Saanich Inlet	$1.5 \times 10^{-5}$
Black Sea	$3.0 \times 10^{-4}$
Orca Basin*	$3.2 \times 10^{+3}$

\* Calculated from data of Shokes et al. (1977).

laboratory for examining the mechanisms of anaerobic decomposition.

An understanding of anaerobic decomposition processes is important to marine geochemists. The sequence of oxygen depletion, followed by nitrate, nitrite, and sulfate reduction is an oft repeated pattern in anoxic marine basins and fjords (Richards 1965), evaporite basins (Borchert and Muir 1964), and during the early diagenesis of marine sediments (see, for example, Martens et al. 1978). Many marine sediments are anoxic, especially along continental margins where the sedimentation rate is high. Such sediments receive a large flux of organic matter, its origin being the fixation of organic carbon in the overlying photosynthetic zone. Organic-rich sediments undergo early diagenesis in which microorganisms consume the labile organic matter using first oxygen, then nitrate, nitrite, and sulfate as oxidizers until each is depleted and methanogenesis takes place. The products of this anaerobic decomposition diffuse out of the sediments into the overlying sea water.

Early diagenesis in sediments overlain by oxygenated waters cannot be interpreted by examining reduced species concentration in the overlying waters. Diagenetic products diffusing out of the sediment into oxygenated sea water are diluted or are consumed by oxidizing bacteria. The oxidation products in each case are compounds (e.g.,  $\text{CO}_2$ ) that are in large abundances or are rapidly diluted in aerobic sea water. It thus becomes difficult to assess

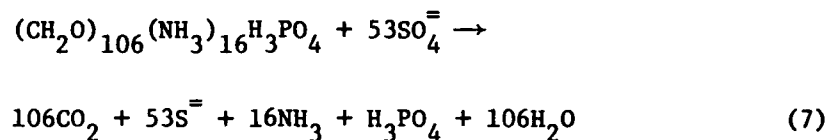
their input into the overlying water and consequently their geochemical impact on the oceans. For sediments at the bottom of anoxic basins or fjords, however, there is no free oxygen available to act as an oxidant to alter the decomposition products. Thus the terminal products of anaerobic decomposition accumulate in the overlying anoxic waters.

Such anoxic basins and fjords provide scientists with natural laboratories for examining the geochemistry and microbiology of organic matter decomposition. The Black Sea, being the largest anoxic basin in the world, has received the most attention. After the results of the 1967 U.S. expedition had been reported (Degens and Ross 1974), Laking (1974) compiled a bibliography with over 4,000 citations of research in the Black Sea. Since Richards (1955) reported the discovery of anoxic sea water in the Cariaco Trench, there have been over 120 papers published on this isolated Venezuelan basin (for a review, see Richards 1975). Such intense research activity is but another example of the scientific interest in anoxic environments. This interest is well justified. The chemical changes that led to the development of anoxic conditions are of interest beyond the immediate confines of the stagnating waters in basins and fjords where they are studied.

Examinations of regenerated nutrients and dissolved gases in anoxic environments has led to various models which explain the anaerobic decomposition of organic matter. During anaerobic



decomposition, organic material [represented here as  $(\text{CH}_2\text{O})_{106}$   $(\text{NH}_3)_{16}\text{H}_3\text{PO}_4$ ] is decomposed into the nutrient  $\text{H}_3\text{PO}_4$ , and the dissolved gases  $\text{CO}_2$ ,  $\text{CH}_4$ ,  $\text{NH}_3$ ,  $\text{N}_2$ , and  $\text{H}_2\text{S}$ . To describe this process, simple stoichiometric reactions have been used with some success (Richards et al. 1965; Gaines and Pilson 1972; Adams 1973). An example is the reaction of Richards (1965) for phosphorus and nitrogen liberation during bacterial sulfate reduction.



The C:N:P ratio of organic matter in this reaction is the average value of plankton of 106:16:1 given by Redfield (1934) and Fleming (1940).

Richards' model has been applied to the Cariaco Trench, Black Sea, Lake Nitinat, and Saanich Inlet. Most of these anoxic environments experience either periodic mass flushing or a slow continual input of oxygenated water into the anoxic waters. Unless the magnitude and time scale of the flushing can be accurately estimated for each area, it becomes difficult to apply the model. This is true since dissolved oxygen enters with renewal water. Input of oxygen can result in removal (oxidation) of reduced compounds and inhibition of anaerobic bacteria. With the discovery of the anoxic, hypersaline Orca Basin in the Gulf of Mexico (McKee

and Sidner 1976; Shokes et al. 1977) we have discovered a new natural laboratory to study anaerobic decomposition processes. As we shall see, this is a laboratory which does not experience the mass periodic or even slow gradual flushings of other anoxic basins.

#### Research Objectives

The unusual circumstances involved in the formation and chemistry of the Orca Basin make it a desirable area for oceanographic research. It is an unusual system when compared to the normal distributions of major ions and gases in the ocean. However, it probably is not unique when viewed in a geological perspective. Basins similar in nature to the hypersaline Orca Basin must have been common in the geological history of the Mediterranean Sea and other regions where salt tectonics has played an important role (N. L. Guinasso, personal communication). Remnants of basins similar to Orca Basin (as evidenced by rapid sedimentation rate) are found buried in the deeper sediments below the Mediterranean. Understanding such basins may be important on a geological time scale. Understanding decomposition processes is important on a daily basis. Dissolved gases can be useful geochemical tracers in such an environment. Inert gases can be used to elucidate formation processes, while biogenic gases (major products of anaerobic decomposition) yield information about current reactive processes.

Most of the published work on the Orca Basin has ignored the dissolved gases in the basin. Shokes et al. (1977) noted that gases effervesced from the water (brine) when it was brought to the surface, but they made no measurements of the effluent gases. Only dissolved oxygen, which decreases to zero in the brine, was measured. In their carbon budget study, Sackett et al. (1979) measured both methane and carbon dioxide of the basin water and sediments. Their approach was to concentrate on the carbon compounds in the basin. Sackett et al. (1979) did not consider all the dissolved gases resulting from anaerobic decomposition.

In examining the hot, salty Red Sea brines, Craig (1969) pointed out that dissolved gas distributions could be a powerful tool in evaluating the mechanisms of brine formation. This is true since, unlike the major ions, initial gas concentrations are independent of both the ion ratios of the brine and the bulk ion composition of the salt from which the brine formed. Weiss (1969) used nitrogen, argon, and carbon dioxide values from the Red Sea brine to gain valuable information concerning its source of origin. He showed the utility of dissolved gas measurements in such unusual environments. With these ideas in mind, this author has undertaken a systematic study of the dissolved gases in the Orca Basin. The objective of this study has been to use dissolved gases to address some of the questions relevant to both the Orca Basin and to anaerobic decompositions in general.

These questions include:

1. What is the source of the Orca Basin brine and can dissolved gas data be used to confirm the source?
2. How is anaerobic decomposition of organic material affected by high salinity water?
3. What are the origin and significance of the isotopically light methane in the brine and sediments of the Orca Basin?
4. Why is there no hydrogen sulfide in the Orca Basin in spite of evidence of sulfate reduction in the sediments?
5. What role does the input of biogenic carbon dioxide gas play in the carbonate chemistry of the brine?

CHAPTER II  
DENSITY EFFECTS AND PHYSICAL MIXING PROCESSES

Introduction

The dissolved gas distributions in the Orca Basin are directly affected by the density gradient and interface mixing processes which result from the presence of brine in the bottom of the basin. Thus, a consideration of the physical characteristics of the Orca Basin brine is important to this study. The Gulf of Mexico sea water above the Orca Basin exhibits temperature and salinity depth profiles (Fig. 8) which are typical of the expected distributions in this area of the western Gulf (cf. Nowlin 1972). Below about 2000 m, however, the salinity concentrations begin to increase slowly until an extremely rapid increase in temperature and salt content are observed at about 2250 m (see Fig. 3). Salinity increases from about 70‰ to over 250‰ over a depth interval of only 17 m. This rapidly changing salinity has produced the distinct pycnocline at this level.

A true appreciation of the significance of the pycnocline in the Orca Basin can be obtained by comparing the density gradient in the Orca Basin with the pycnocline near the surface of the Gulf of Mexico (Fig. 9). The surface pycnocline separates the homogeneous mixed layer from the bulk of the oceans and it is this pycnocline which is responsible for maintaining the nutrient and

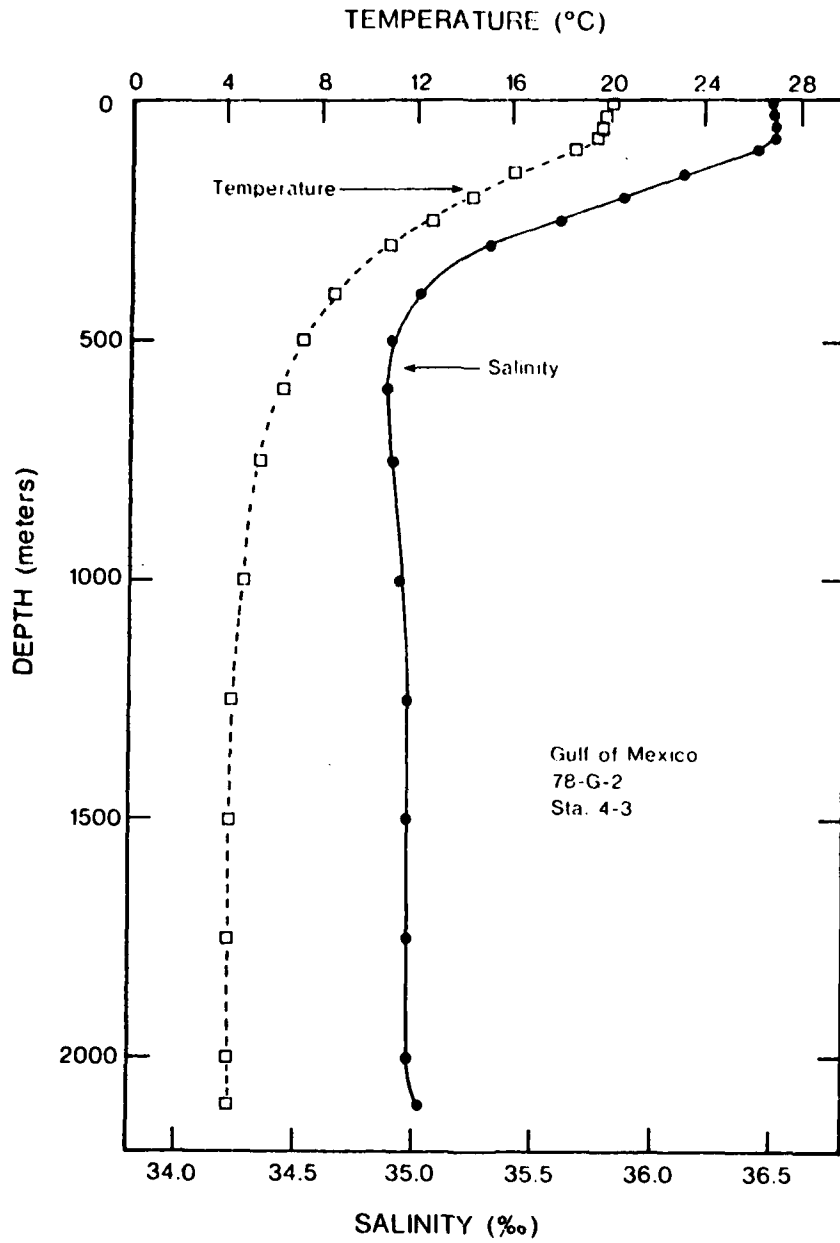


Fig. 8. Typical temperature and salinity depth profiles taken in the water column of the northwestern Gulf of Mexico above the Orca Basin.

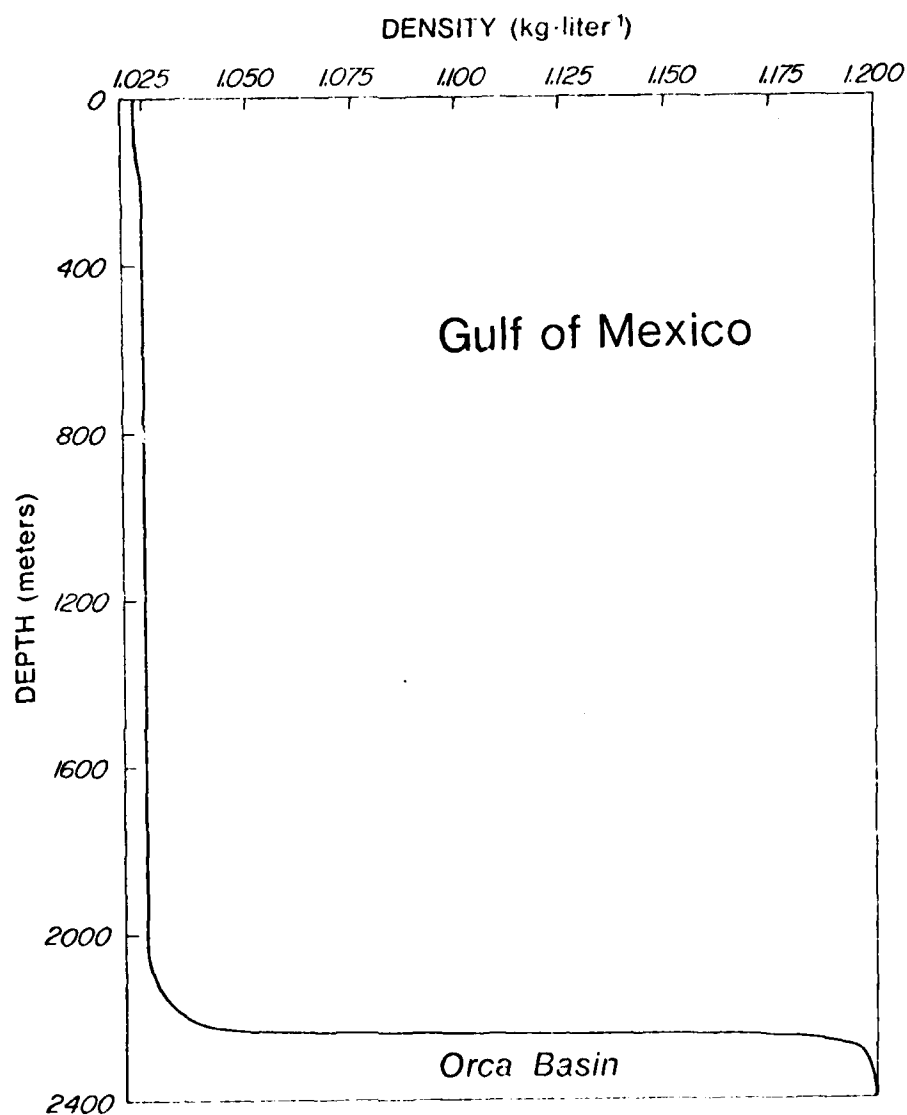


Fig. 9. Density profile for the Gulf of Mexico, extending down through the Orca Basin brine. Density at the brine interface increases from 1.025 to 1.185 kg·liter<sup>-1</sup> over a depth interval of less than 50 meters. This is one of the most striking density gradients of an oceanic area (see Table 5).

biological distributions in surface waters. That surface pycnocline is dwarfed by the density gradient at the surface of the Orca Basin brine layer. It is this gradient that reflects the low frequency sound waves that can be recorded even at a frequency as high as 3.5 kHz.

The Orca Basin pycnocline must act as a temporary trap for many of the particles which fall into the Basin. As particles settle through the water column, most have a density only slightly greater than the water through which they fall. Upon reaching the density gradient above the Orca Basin, their downward progress will be impeded. These particles probably stop at the density interface and begin to soak up salts (or dehydrate due to osmosis) until they reach isotonic balance with the brine. This may not be a continuous process. Particles may have to go through several layers or steps before their density will have increased sufficiently to allow them to proceed on their journey to the sediments. Even at the sediment interface, particles do not have a sufficient density, relative to the encompassing brine, to compact completely at first. As a result, the surface sediments are soupy, and they are difficult to sample since they tend to run out of the smallest hole in a box corer or other sampling device.

Support for the concept of an increased particle abundance at the brine-sea water interface comes from the bacterial biomass data of LaRock et al. (1979). They observed an increase in both



adenosine triphosphate levels and total bacterial cell counts in samples taken from the Orca Basin interface, as opposed to samples taken from the brine or from waters above the interface. Bacteria in the deep ocean are usually attached to particles rather than free-floating. Thus higher microbial biomass at the interface would be indicative of an increased particle density at that level. This increased microbial biomass was consistent with findings from another stratified basin, the Cariaco Trench, where Karl et al. (1977) observed a similar ATP distribution. The degree of stratification in the Orca Basin can be discerned by a detailed examination of the temperature and salinity distributions through the interface.

#### Temperature Distribution

Examination of a continuous temperature profile through the interface to the bottom of the Orca Basin (Fig. 10) with a Plessey Environmental Model 9040 STD shows that the Orca Basin brine is not thermally homogenous. Temperature begins to increase at a depth of around 2150 meters. The temperature depth profile goes through several steps between 2150 and 2250 meters before a more rapid increase begins. In the highest salinity brine (>250‰), there are at least two distinct layers having a temperature difference of about 0.05°C. This two-layer system within the brine was not observed by Shokes et al. (1977), who used reversing

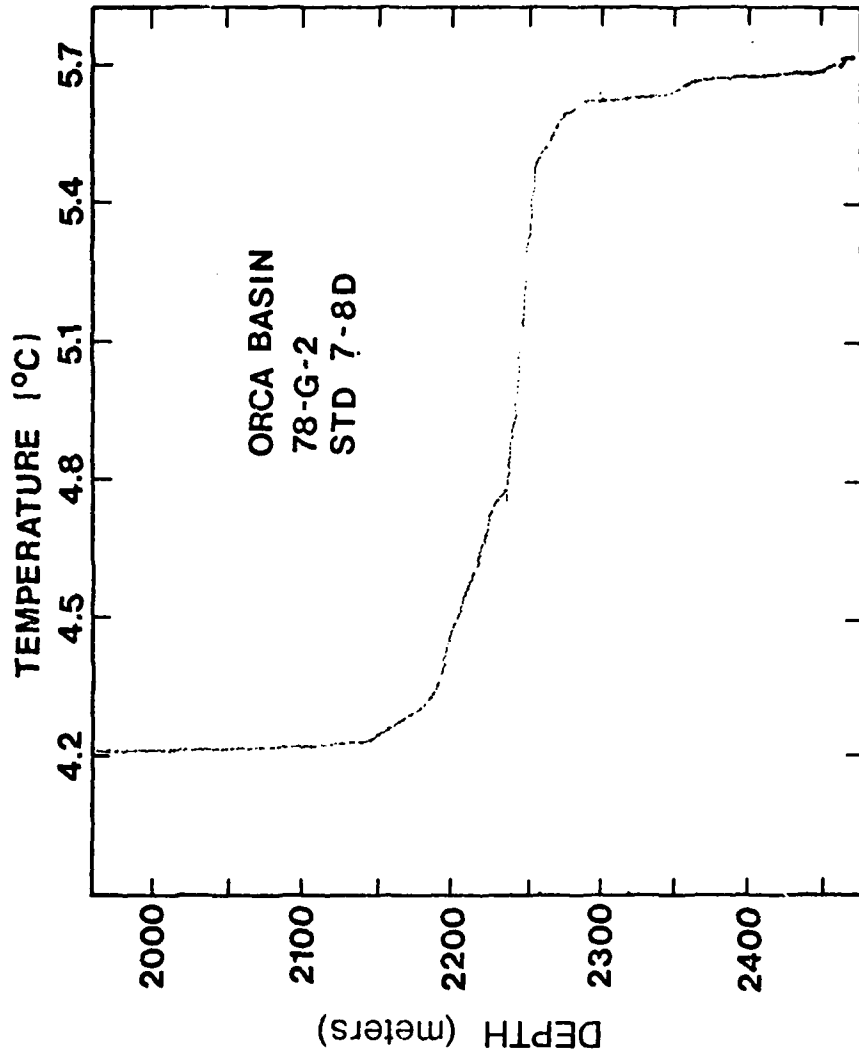


Fig. 10. Continuous temperature profile through the interface to the bottom of the Orca Basin made using a Plessey 9040 STD with the salinity probe disabled. Five thermal layers are observed above the high salinity brine, with at least two additional layers in the brine itself.

deep-sea thermometers to obtain their temperature profile.

Temperature of the Orca Basin sediment was measured (accidentally) on cruise 77-G-3 of the R/V Gyre when the same STD used to make the profile in Fig. 10 was lowered into the mud. Due to the small density difference between brine and sediment particles falling into the basin, the top layer of sediment in the Orca Basin is very soft and unconsolidated, with about the same consistency as clam chowder (New England style). The surface sediments have a temperature of 5.77°C, a value 0.03°C higher than the bottom water temperature. This temperature differential between the brine and the sediment indicates that the brine is heated from below and would account for the heat flow required to raise the initial brine temperature from the original 4.2°C (Deep Gulf water) to the 5.6°C found today. Heat flow calculations and comparison with the estimates of Shokes et al. (1977) are given in a later section.

#### Salinity Distribution

The importance of the density gradient which results from brine in the bottom of the basin was discussed in Chapter One. This density gradient (Fig. 9) is dependent not on temperature, but salinity. The temperature of the high salinity (>35‰) water increases with depth (Fig. 10). A temperature increase would result in a corresponding density decrease without the effect of

salt on density. Since salt is the major factor influencing density, salinity can be equated with density for the following discussion.

The salinity depth distribution in the Orca Basin falls logically into three zones. I have classified these zones (proceeding from deep to shallow) as the high salinity brine, the interface zone and the transition zone (Fig. 11). The high salinity brine is a semihomogenous mixture (Millero et al. 1979) with a salinity of 262‰, relative to standard sea water. The brine, which occupies the bottom 200 m of the Orca Basin, appears to have two thermal layers (Fig. 10) with a slightly higher temperature in the deeper region. This thermal gradient makes the lower layer slightly unstable with respect to the upper layer of the brine and thus could cause it to mix (see mixing discussion). The high salinity brine is very stable, however, relative to the water above it.

The interface above the high salinity brine is the most marked salinity gradient in the basin. Salinity increases from 70‰ to over 260‰ with a depth decrease of only 17 m. This change ( $dS/dz = 11.5\text{‰} \cdot \text{m}^{-1}$ ) is an order of magnitude greater than the zone either above or below it. It was very difficult to obtain samples in this region. The points plotted on Fig. 11 were obtained using the STD-rosette sampling system that was used to produce Fig. 10. The data points that were plotted (Table 6) reveal the narrow range of depths involved. Data were taken from

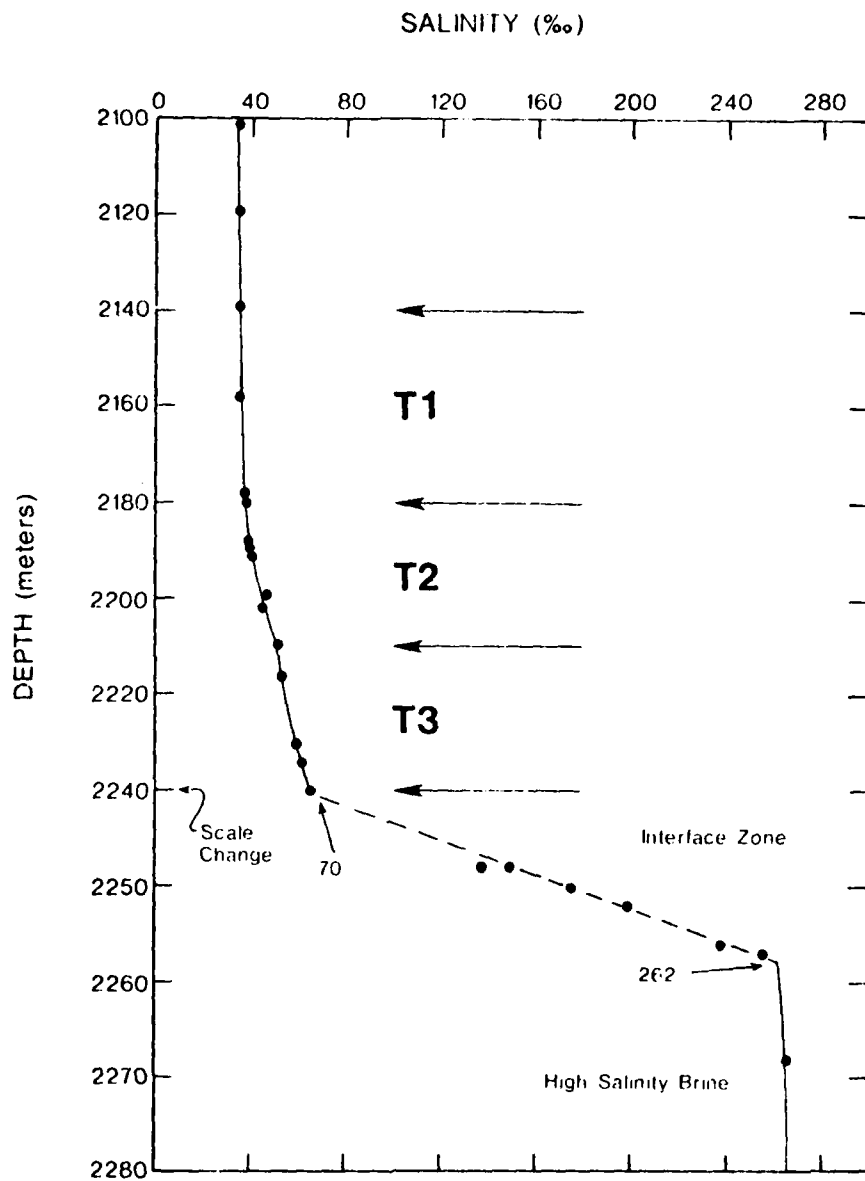


Fig. 11. Detailed vertical profile of salinity through the interface region of the Orca Basin. Three zones are identified: the transition zone from Gulf water, the true interface zone where salinities increase from 70 to 262‰, and the high salinity brine with a constant salinity of 262‰. The transition zone is subdivided into three regions depending on the change in slope of salinity and dissolved oxygen concentrations with depth (see Table 6).

Table 6. Temperatures and salinities taken from two STD casts on cruise 77-G-13. Note the large change in salinity over such a small depth.

Depth (meters)	Temp (°C)	Salinity* (‰)	Cast
2204	4.26	46	4
2230	4.54	60	4
2240	4.63	66	4
2248	4.77	138	4
2248	4.72	150	9
2250	4.81	176	9
2252	4.88	199	9
2256	4.96	238	4
2257	5.15	256	9
2268	5.38	266	9

\* Salinity samples were volume diluted to a salinity near  $35\text{g}\cdot\text{kg}^{-1}$  and analyzed using standard Copenhagen Water as a reference. Volumes were converted to weights using a brine density of  $1.185\text{g}\cdot\text{kg}^{-1}$ .

two separate casts using the same system, yet samples from identical depths of 2248 m from each cast show a salinity difference of 12 ‰ (138 cf. 150 ‰, Table 6).

The interface zone probably results from a mixing of the high salinity brine below with the transition zone water above. The lower layer of the transition zone (T3) has a salinity at the bottom of 70 ‰. A two point mixing curve for the interface zone would have 70 ‰ and 262 ‰ as the end points (Fig. 11). The two point mixing model seems to fit the salinity data in Fig. 11 reasonably well. Linear regression analysis of the data produces a straight line with a coefficient of variation of 0.991. Salinity changes by one part per thousand for each nine cm in this zone.

An STD system was the only reliable device capable of taking numerous samples in this narrow region on a single cast. The paucity of data from this region in other papers (e.g., LaRock et al. 1979) shows how difficult it is to obtain samples there. When closely spaced additional salinity samples are obtained, the two point mixing curve for this interface zone can be used to accurately calculate sample position relative to the high salinity brine.

The transition zone can be classified into three regions. The characteristics of each region are listed in Table 7. The upper region (T1) is the area where density first begins to increase from the constant values of the deep Gulf water. The

Table 7. Salinity zones in and above the Orca Basin.

Zone	Region	Salinity Range (‰)	Depth Range (m)	$\frac{\partial S}{\partial Z}$ (‰·m <sup>-1</sup> )	O <sub>2</sub> Range (μmol·L <sup>-1</sup> )	$\frac{\partial O_2}{\partial S}$ (μmol·L <sup>-1</sup> ‰)
Transition	T1	35.2	2130	0.031	225	30.5
		37.0	2178		170	
Transition	T2	37.0	2178	0.42	170	12.3
		50.0	2209		10	
Transition	T3	50.0	2209	0.62	10	0.5
		70.0	2241		0	
Interface	I	70.0	2241	11.53	0	-
		262.0	2258			
High Salinity Brine	B	262.0	2258	0	0	-
		+0.2	2430 (bottom)*			

\* Exact bottom of Orca Basin varies.



salinity change is small, but the effects are apparent. The depth of this region is about 50 meters. Transition region T2 spans a depth interval of 31 m with a  $dS/dz$  of 0.42, an order of magnitude greater than the region above. The salinity at the bottom of this region is 50‰. The deepest transition region has a salinity of between 50 and 70‰. Salinity in this region increases about 1.5 times as fast as in the region above. Region T3 abutts the interface zone below. It is in this region (T3) that Shokes et al. (1977) observed the step in the oxygen profile.

An examination of the temperature profile in the basin (Fig. 9) indicates that each salinity region is associated with a different slope in the temperature gradient. It is of interest to note here, that the slope of zone T3 for temperature is not much less than the thermal slope for the interface zone. Salinity changes, on the other hand, represent an order of magnitude increase in slope between region T3 and the interface zone.

The salinity distribution in the high salinity brine (Fig. 3) shows no obvious differences with depth. These data were obtained by volume dilutions of the brine before analysis by conductive salinometry (Shokes et al. 1977). Millero et al. (1979) performed precise weight dilutions on six samples from the brine and found an average salinity of  $262.84 \pm 0.14$ ‰. The bottom samples had an average of 263.02‰ ( $\sigma = 0.03$ ) and the four above then averaged 262.75‰ ( $\sigma = 0.02$ ). This slight difference gives an

indication of a two-layered salinity distribution. The difference in the means of these two levels is significant at the 95% confidence level, but only six samples were compared. More detailed salinity sampling is necessary to precisely define the salinity layering in the brine.

An attempt to obtain a detailed salinity profile was made on Gyre cruise 78-G-2. The Plessey 9040 STD was modified to simultaneously measure temperature and salinity in the high salinity brine. This STD has a normal salinity range of only 30-40 ‰. In more saline waters, the salinity output frequency overlaps into the temperature frequency range, obscuring both the temperature and salinity measurements. To measure salinity, a nylon plug was placed in the center of the salinity-sensing induction toroid. Insertion of this plug, with a 0.95 cm diameter hole drilled through its center, reduced the effective cross-sectional area of the toroid. The area was reduced sufficiently to make the measured inductance of the high salinity water the same as the inductance of 37 ‰ sea water in an unmodified STD, while not interfering with the temperature measurements.

The modified STD was lowered into the deepest portion of the basin on March 28, 1978. The rough untreated data from this attempt is shown in Fig. 12, along with an expansion of the temperature trace in the high salinity brine. The salinity profile shows a slight increase, with depth to the bottom, with two zones having separate slopes. The observed change in slope of the

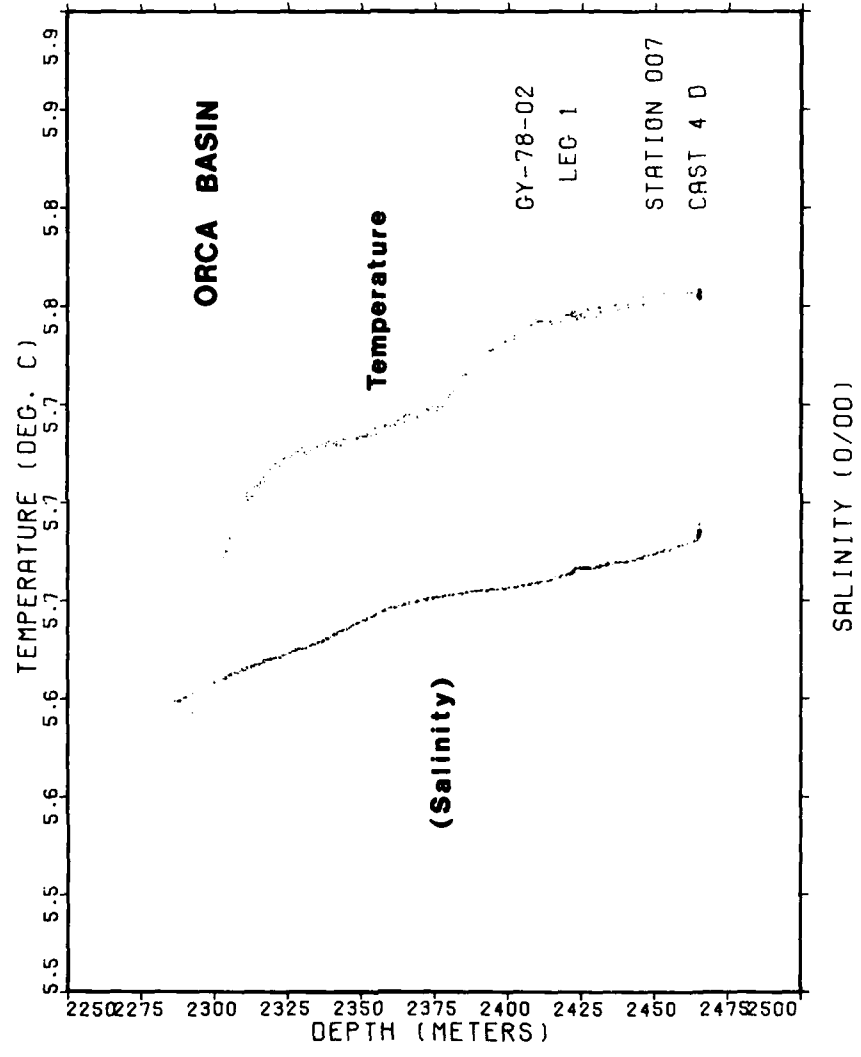


Fig. 12. Vertical profiles of temperature and arbitrary salinity in the high salinity water of the Orca Basin. The salinity data are presented to show trends in the salinity distribution. Gulf salinity is 35 ‰ and brine is about 263 ‰.

salinity profile, however, does not occur where the temperature profile shows an increase. Thus the continuous data do not indicate that there are two separate layers with homogeneous temperature and salinity. It must be remembered that the validity of this salinity trace has to be questioned. The technique that was used to produce the salinity profiles is rather unorthodox, and no calibrations were made to determine the true salinities. The flushing volumes of the smaller toroid also require investigation. The data does indicate, however, that a detailed examination of the salinity structure in the Orca Basin would be profitable in interpreting water structures in the basin.

#### Mixing Processes in the Orca Basin

##### High Salinity Brine

In spite of the above discussion, the salinity distribution in the high salinity brine is considered to be essentially uniform. This homogeneity is also true for several other parameters that have been reported for the high salinity brine (see, for example, Sackett et al. 1979). It follows that some mixing must be taking place to create this homogeneous mixture in the brine. There are several possible mechanisms for mixing. One mechanism would be mixing due to input of new brine. Other possibilities would be mixing resulting from thermal processes in the high salinity waters or from seiching the brine.

New brine entering into the depths of the basin would mix until there was a re-establishment of the density structure in the brine. This mixing process would be rapid however, and would probably result in mixing of the brine such that both temperature and salinity were well mixed. As can be seen in Figs. 10 and 12, temperature is not thermally homogeneous in the brine. Thus, a slower mixing process must be invoked to account for the thermal layers, as well as the homogeneous salinity and other parameters.

Such a process might be convective mixing due to heating from below. This mechanism could only account for mixing in the high salinity brine where the salinity content is approximately constant at 262‰. The temperature differences in this high salinity brine are about 0.05°C between the two layers and could not support density imbalances with a significant salinity gradient. Thus the following discussion of possible convective overturn processes in the Orca Basin is applicable only to the high salinity brine. This is the 262‰ brine in the bottom 140 m of the basin and not the lower salinity waters in the interface or transition zones that were described earlier.

It is quite possible that mixing in the basin deeps is driven by convective overturn resulting from the increasing temperature with depth in the high salinity brine. The bottom temperature measurement with the STD indicated that the Orca Basin is being heated from below. The bottom sediments are 0.03°C warmer than

the brine. The deep Gulf of Mexico water is  $1.4^{\circ}\text{C}$  cooler than the high salinity brine. The Orca Basin high salinity brine is similar to the classical thermal convection situation where there is heat-driven motion in a layer of fluid (the brine) contained between two horizontal planes, uniformly heated from below (the sediment) and cooled from above (Gulf water). In this type of system, deeper water becomes less dense than the water above it, due to increased heating from below, and can mix upward. Apparently, convective overturn is not uncommon in closed basins which are isolated from the more rapid advective mixing processes. For example, Newman (1976) examined the water structure in Lake Kivu and applied the same type of analysis to that system. Like the Orca Basin, he noted steps in the temperature profile in Lake Kivu and attributed the associated mixing in the lake to double-diffusive convective mixing. Turner (1968) also noted that there is increasing evidence that a step-like structure is characteristic of the ocean and some lakes. Instead of smooth gradients in temperature and salinity, well mixed layers separated by sharp interfaces are often observed. The layers would be the result of convective mixing of the waters due to heating from below. The same type of mixing may explain the homogeneity of salinity and other parameters in the high salinity brine of the Orca Basin.

The problem of convective overturn due to differential heating has been well studied in the realm of fluid mechanics. Benard

(1901) first showed that a thin layer of fluid becomes unstable when the temperature (i.e., density) differences exceeds a minimum value. Fluid motion takes place in a regular, steady pattern at first, but can become turbulent with large temperature differences. As described by Turner (1973), an unstable fluid layer of thickness  $\delta$  is driven upward (z direction) by buoyancy forces (density differences) but retarded by viscosity, and neglecting inertial effects there is a local imbalance. Unless the driving forces are much greater than the dissipative forces, the system will remain stable, in spite of the instability of a particular individual layer. The mathematics of the instability was first formulated and solved by Lord Rayleigh (Turner 1973). He showed that there is a critical value of Ra, the non-dimensional Rayleigh number, below which a fluid heated from below is stable to small disturbances. The Rayleigh number, Ra, is given as

$$Ra = (g\alpha/\kappa\gamma) \Delta T h^3 \quad (8)$$

where

g is acceleration due to gravity

$\alpha$  is the thermal coefficient of expansion

$\kappa$  is the thermal conductivity

$\gamma$  is the kinematic viscosity

$\Delta T$  is the temperature difference across a layer

h is the thickness of the layer

The Rayleigh number represents the balance between the driving buoyancy forces and the diffusive processes which retard the motion and tend to stabilize it. At a value of  $Ra_c$  of  $10^3$  to  $10^4$  cells which fill the gap between boundaries become unstable and convective overturn can occur (Turner 1968, 1965).

Since fluid mechanics usually deal with pure fluids (i.e., distilled water), the above description is normally applied to systems that only have a temperature gradient to cause the density imbalance. In the oceanic environment, however, there is usually a salinity gradient to consider also in interpreting buoyancy forces available to initiate convective overturn. The problem of convective overturn in linearly stratified salt solutions has been examined by Turner and Stommel (1964) and others. Turner and Stommel (1964) gave a qualitative description of the motions that result when a linearly stratified salt solution is heated uniformly from below. That description was clearly summarized by Turner (1968) and it could be used to describe the mixing regime in the high salinity brine of the Orca Basin. Rather than attempt to improve on his description, I simply quote:

Convective stirring first produces a layer at the bottom of the containing vessel which is well mixed in both heat and salt, and which grows by incorporating fluid from above. This layer does not continue to deepen indefinitely, however: at some point a second layer forms on top of it, and this behaves in the same way. In time, many such layers can form, with sharp interfaces separating the turbulent convecting regions, and these interfaces stay in nearly fixed positions until the bottom



one disappears as the lowest two layers merge. The system can be maintained in this state because of the very different molecular diffusivities of heat and salt: heat can more easily escape from the top of a layer to cause a convective overturning above, while the salt mostly stays behind and preserves a net stable density difference across the interface (Turner 1968).

The question to be considered here is whether heat input from the sediments into the Orca Basin brine is sufficient to create the density imbalance required to exceed the critical Rayleigh number,  $Ra_c$ . Unfortunately, various parameters such as the viscosity and thermal conductivity have not been determined for the Orca Basin brine. Thus in order to calculate  $Ra$  using Eq. 8, I have chosen the values for the thermal coefficient of expansion, thermal conductivity and kinematic viscosity that Newman (1976) used for Lake Kivu. The values for sea water would be equally applicable, since these terms are not the dominant terms in the equation. Increases in salinity tend to cause both the viscosity and the thermal coefficient of expansion to increase. The two combined increases tend to cancel since the ratio of the two is used in Eq. 8. Using Newman's (1976) data and the Orca Basin values of  $0.05^\circ\text{C}$  difference between the two layers with an interface distance of 25 m between the high salinity brine layers, a Rayleigh number on the order of  $10^{13}$  was calculated. This value is several orders of magnitude larger than the critical value of  $Ra = 10^4$ . Based on this calculation, it seems reasonable

to assume that the layering observed in the high salinity brine of the Orca Basin is due to double-diffusive convective mixing driven by geothermal heating from the basin sediments. Conclusive proof of this hypothesis requires a more sophisticated set of constants for the Rayleigh number calculation, as well as a better definition of the salinity and temperature microstructure in the high salinity brine.

#### Oxygen Step Region and General Mixing

One of the most striking features of the data presented by Shokes et al. (1977) was the anomaly (step) just above the high salinity brine in the oxygen depth profile. A comparison of the oxygen data (Fig. 5) and the temperature and salinity profiles (Figs. 10 and 11) indicates that the oxygen step is associated with one of the physical layers in the brine transition zone, region T3. There are many complex forces acting upon this interface region. Salinity increases from 50‰ to 70‰. Temperature increases from 4.25°C to 4.75°C. The brine interface may be oscillating with time. This region could be physically active. There are two plausible explanations for this step region. Either new, oxygenated brine is being put in at this level, or oxygenated water from above is mixing with water from the interface zone below to create this higher salinity, oxygenated water in region T3.

An examination of a temperature versus salinity plot for the Orca Basin is helpful in understanding the processes acting on the oxygen step region. Fig. 13 is a plot of the temperature and salinity data from Fig. 3. The salinity data are given in units of  $\text{g}\cdot\text{liter}^{-1}$ , since the samples were volume diluted. Correcting the data for density and replotting in salinity units of  $\text{g}\cdot\text{kg}^{-1}$  showed the same profiles, and would not alter the data interpretations. Fig. 13a shows a plot of all the temperature and salinity data taken from depths of 2050 m down to 2430 m, near the bottom of the basin. The solid line drawn through the data follows the data from a salinity of 35 ‰ to a salinity of 170 ‰, and then the second half of the curve (from 170 to 305 ‰) is simply a mirror image of the left hand portion of the solid curve. The curve was drawn in this manner since this data set (after Shokes et al. 1977) had only two data points in the high salinity region, other than in the high salinity brine itself. Fig. 13b is an expansion of the data between 35 and 70 ‰ from Fig. 13a, indicated by the area between arrows on Fig. 13a. The T-S profile in this region is almost a straight line, which is indicative of a two point mixing condition. The two points representing the extremes of the straight line drawn on Fig. 13b would be the temperatures and salinities of the two water masses that are mixing.

Fig. 13c is an expansion of the data between 45 and 75 ‰, and also exhibits the same straight line structure indicative of

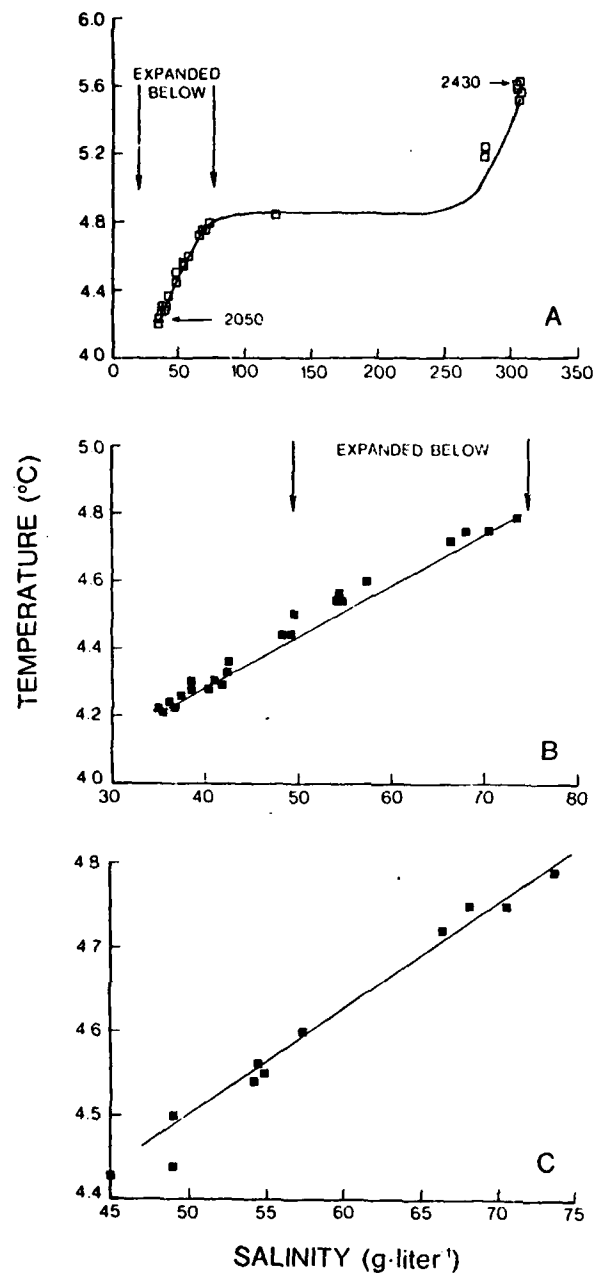


Fig. 13. Salinity versus temperature profile for the Orca Basin. Parts b and c are expanded plots of the data presented in part a. The data are taken from the same data shown in Fig. 3.

a two-point mixing model. Most of the points on Fig. 13c are from the oxygen step region shown on Fig. 5.

If the water in the oxygen step region is the result of a two point mixing process, this could explain the oxygen concentrations found in that zone. The oxygen would be oxygen mixed downward from above. This mixing, however, is probably not due to convective overturn as was discussed in the previous section for the high salinity brine. Salinity increases from 50 ‰ at the top of zone T3, to 70 ‰ at the bottom, 32 m below. For this zone to turn over due to heating from below, the 70 ‰ salinity water would have to reach a temperature of over 55°C. There is no evidence for such high temperatures at the Orca Basin interface. Yet, evidence for mixing of the oxygen step region(T3) is compelling.

The salinity-temperature plots presented in Fig. 13 might be explained by diffusive mixing processes alone, if one assumes that a double diffusive process is taking place. That is temperature and salinity are diffusing at different rates. For molecular diffusion, this is indeed the situation. Heat diffuses faster than salt (Munk 1966). Two questions to be considered in this discussion are what are the differences in the diffusion rates of these two parameters at the Orca Basin interface, and how does diffusivity change with distance from the interface.

If one assumes that the brine flowed without mixing into the depths of the Orca Basin at some initial time  $t = 0$ , then at that

time the brine was covered by Gulf of Mexico water with a sharp boundary between the two. In fluid dynamics, this initial case is described as one in which the distribution occupies a finite region with an initial state defined by:

$$C = C_0, \quad Z < 0, \quad C = 0, \quad Z > 0, \quad \text{at } t = 0 \quad (9)$$

where  $C$  = concentration,  $Z$  = distance,  $t$  = time and  $Z = 0$  at the interface. This distribution is analogous to one in which a long tube of clear water rests on a long column of solution, or when a long water column of Gulf of Mexico water rests on top of the Orca Basin brine. The case for the Orca Basin is shown diagrammatically in Fig. 14. At a time  $t_0$ , both the temperature and salinity profiles have only one value in the Gulf water ( $T_1, S_1$ ) and a single value in the brine ( $T_2, S_2$ ). The profile at time  $t_1$  is illustrated by the solid line in Fig. 14.

After initial diffusive mixing, the Orca Basin interface must reach steady-state to be maintained. This can be proved by performing a simple calculation of how long the temperature profile would be maintained at a constant thermal diffusivity of  $10^{-3} \text{ cm}^2 \cdot \text{sec}^{-1}$ .

Using the simple relationship,

$$L^2 = Dt \quad (10)$$

where  $L$  is length,  $D$  is the thermal diffusivity, and  $t$  is time, one can calculate the characteristic time for diffusion to spread an initial temperature step function. The interface thickness is 1700 cm. With  $D_T = 10^{-3}$ , the time required for diffusivity to spread an initial step function to the present thickness is

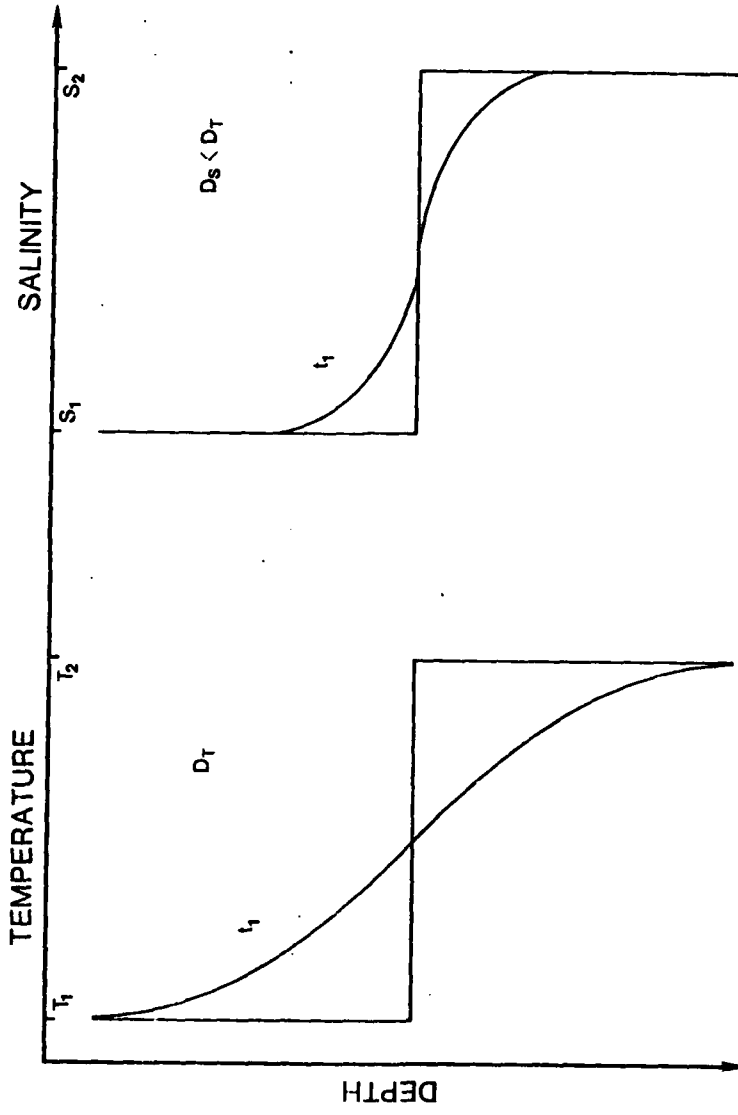


Fig. 14. Temperature and salinity profiles for the initial two-layer system in the Orca Basin if the brine entered the basin without mixing. The solid line indicates the degree of diffusion of each parameter at some characteristic time  $t_1$ .  $D_T > D_S$ .

$2.9 \times 10^9$  sec, or about 90 years. Since the time of initial input of brine into the Orca Basin is  $\sim 8000$  years, time dependent processes would have removed the interface long ago. Similarly, time-dependent diffusion of salt ( $D_s = 10^{-5}$ ) would spread the salinity interface in  $3 \times 10^{11}$  sec, or about 9000 years. The Orca Basin interface region would be unstable unless the system were steady-state.

A comparison of the STD temperature-depth profiles in the basin taken during three different years supports the steady-state concept. The first temperature-depth profile with an STD system was taken in 1977 by Guinasso (1977). The latest profile was taken in March, 1980, on Gyre cruise 80-G-4. The thermal layers throughout the basin (Fig. 15) are identical in both profiles, yet geothermal heat is entering the brine from below. This input would change the temperature interface without a balanced output. The Orca Basin diffusion must be in steady-state relative to heat.

Salt diffusion would not be steady-state without an input, but diffusion out would be very slow. Assuming a diffusion coefficient for salt of  $10^{-5} \text{ cm}^2 \cdot \text{sec}^{-1}$ , one can calculate the time for all salt in the basin to diffuse out. A one  $\text{cm}^2$  column in the deepest part of the basin would contain  $2 \times 10^5 \text{ cm}^3$  of brine, with each  $\text{cm}^3$  containing 0.25 g of salt. With  $D_s = 10^{-5}$ , a salt flux of  $1.25 \times 10^{-10} \text{ g} \cdot \text{cm}^{-2} \cdot \text{sec}^{-1}$  is calculated. Dividing the mass of salt in the water column ( $5 \times 10^4 \text{ g} \cdot \text{cm}^{-2}$ ) by the flux gives  $4 \times 10^{13}$  sec, or about 1,300,000 years, as the time for all the salt to diffuse out of the basin. In the 8000 years since the brine entered the



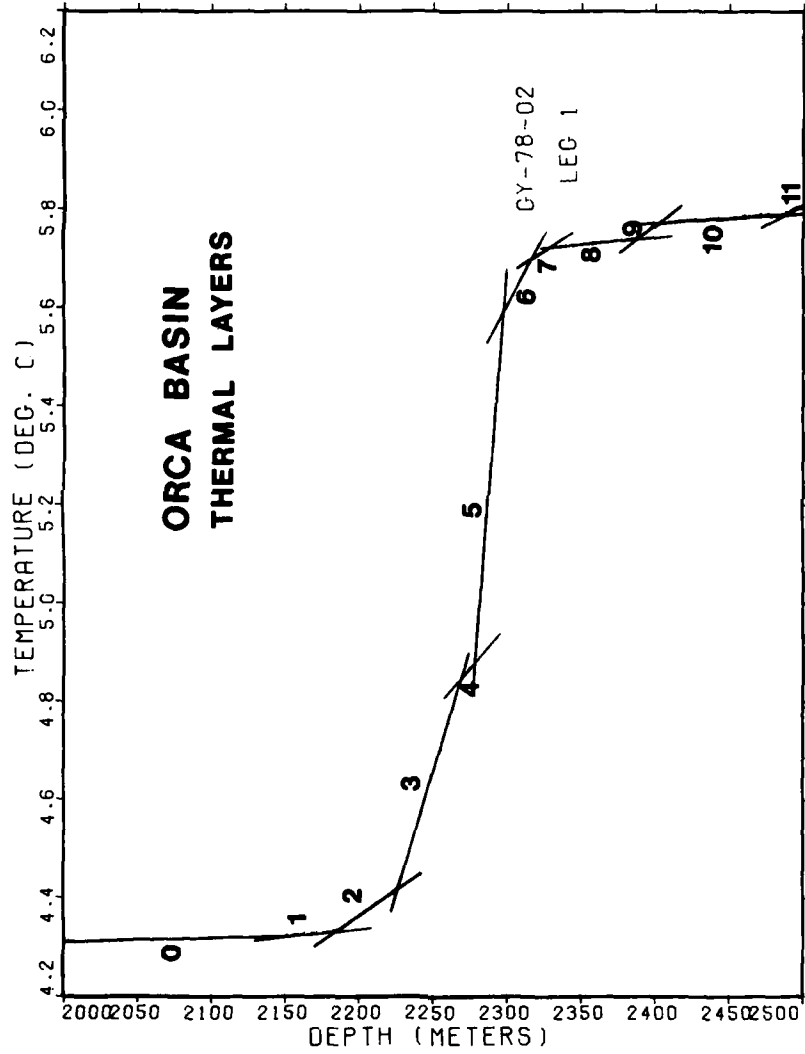


Fig. 15. Modified temperature depth profile showing the linear thermal layers through the interface into the high salinity brine. Each layer has a different thermal diffusivity. Depths have not been corrected.

basin, only 0.6% of the salt could have diffused out. The same would be true for other solutes produced there. Transport of solutes from the brine to the overlying Gulf waters would be very slow.

The slope of the twelve thermal layers in Fig. 15 can be used to calculate changes in thermal diffusivity. Through the interface (layer 5) diffusion must be entirely molecular. With such a large temperature and salinity increase over the short distance (17 m) of this zone, processes more rapid than molecular would rapidly distort the interface. With only molecular diffusion, the transport of heat through the interface is due only to the thermal conductivity of the brine. The average salinity in layer 5 is 167 ‰. A 17% NaCl solution would have a thermal conductivity of  $1.345 \times 10^{-3} \text{ cal} \cdot \text{cm}^{-1} \cdot \text{sec}^{-1} \cdot \text{C}^{-1}$  (Weast 1969). The temperature change across the 1700 cm interface is  $0.65^\circ\text{C}$ , giving a slope,  $dT/dz$ , of  $3.76 \times 10^{-4} \cdot \text{C} \cdot \text{cm}^{-1}$ . Heat flux through the interface (layer 5) can be calculated by the formula:

$$\text{Heat Flux} = \frac{dT}{dz} \cdot \kappa \quad (11)$$

where  $\kappa$  is the thermal conductivity. Heat flux through the interface was calculated as  $0.5 \mu\text{cal} \cdot \text{cm}^{-2} \cdot \text{sec}^{-1}$ . This flux is half of the world average geothermal heat flow used by Shokes et al. (1977) to calculate the time to heat the brine to  $5.6^\circ\text{C}$ . Since the Orca Basin brine is in approximate geothermal balance, their estimate is probably too high for this area.

If heat transport in the Orca Basin is steady-state, the heat flux through any layer in Fig. 15 will be the same. The heat flux through layer 5 has been calculated. Thus, using the slope of each

layer ( $dT/dz$ ) and Eq. 11, the thermal conductivity can be calculated for each layer. The thermal diffusivity ( $D_T$ ) is related to the thermal conductivity ( $\kappa$ ) by

$$D_T = \frac{\kappa}{C_p \rho} \quad (12)$$

where  $C_p$  is the heat capacity and  $\rho$  is density. Using an average density for each layer and assuming  $C_p$  was unity, thermal diffusivities were calculated for each of the twelve layers in Fig. 15. Thermal diffusion constants are plotted against depth in Fig. 16. The lowest diffusivity (molecular) is in layer 5, and the diffusivities increase in both directions away from the interface. The changes are regular and predictable above the interface (except layer 4), but very irregular in the high salinity brine. Where thermal diffusivity is greater than molecular, salt should diffuse at the same rate giving the linear temperature-salinity distributions that were observed.

The above discussion has shown that in addition to molecular processes, eddy diffusion is operative in the transition zone. This would require the input of energy into the basin to produce this type of mixing. One possibility for this input of energy into the basin would be a seiching of the brine in the basin. If the interface were oscillating, there would be sufficient energy input to cause turbulent mixing.

Evidence for seiching in the basin comes from the minisparker profiles obtained by Shokes et al. (1977). The dark line representing the brine interface appears to be elevated on the right side of the profile in Fig. 17. This profile could have been

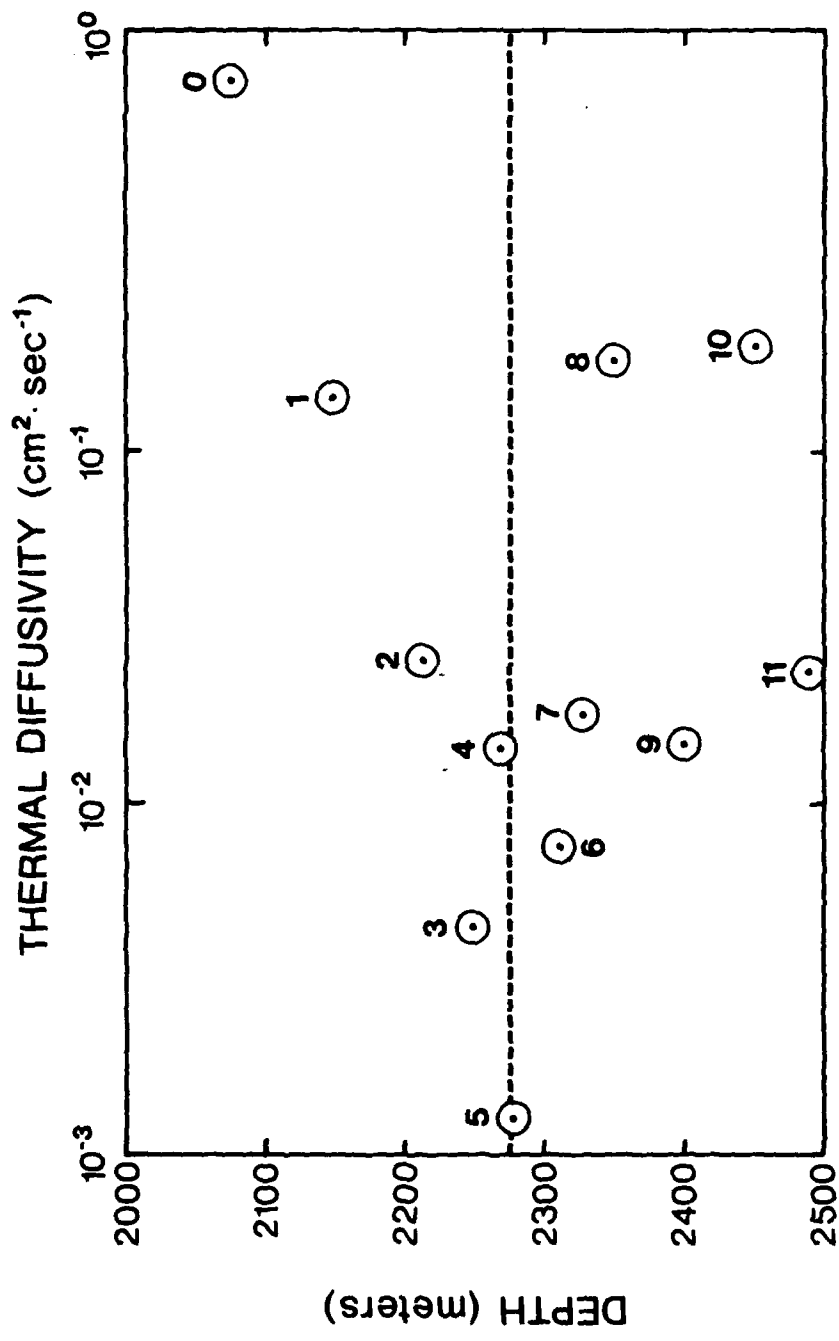


Fig. 16. Plot of diffusion coefficients for temperature ( $D_T$ ) with increasing distance from the Orca Basin interface zone (Layer 5). Layer numbers (given by each point) refer to thermal layers in Fig. 15.

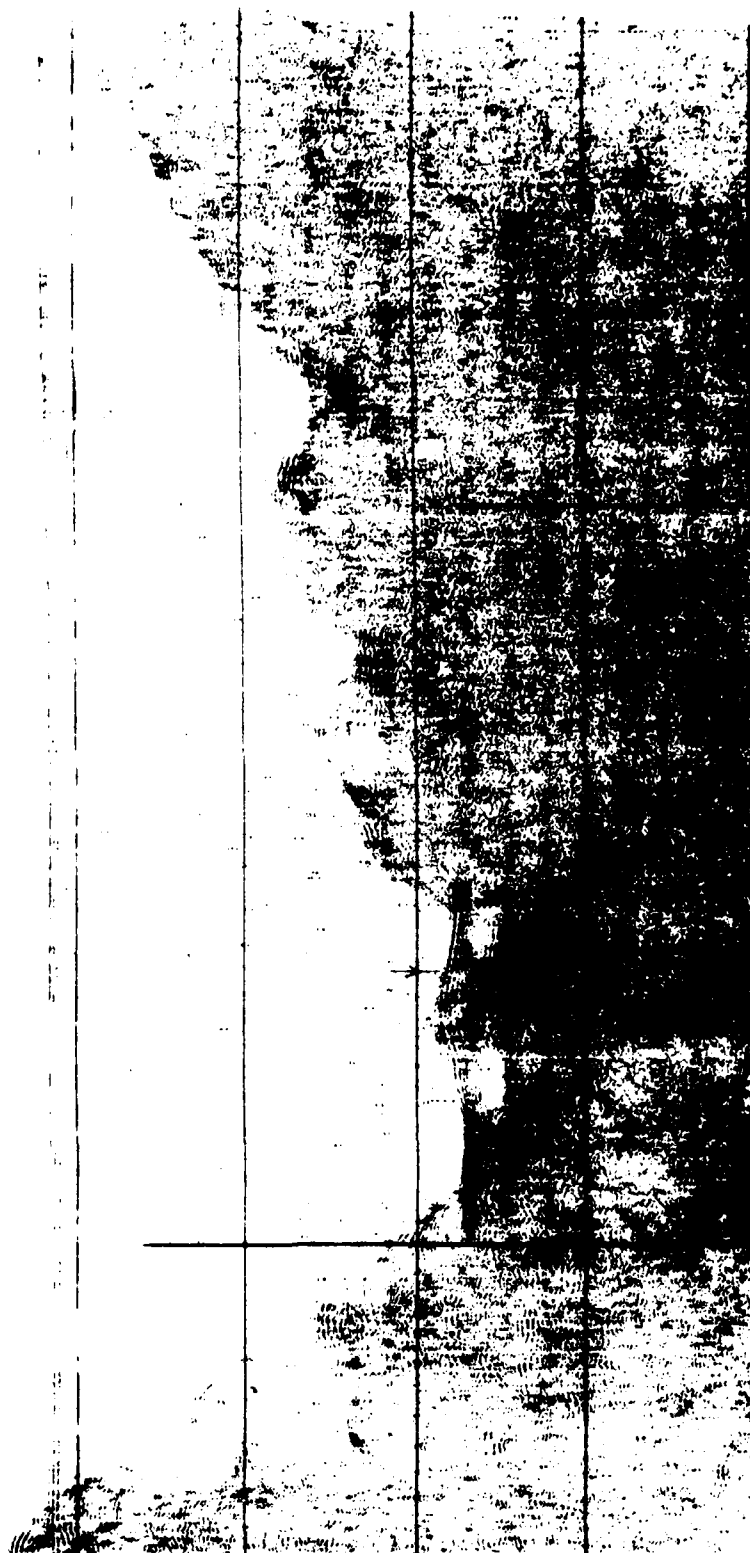


Fig. 17. One section of the minisparker profile taken on Gyre cruise 76-G-10. The profile crosses the longest section of the southern lobe of the basin. The right side of the profile is 3% higher than the left hand side.



taken when the interface was tilted due to seiching. Examination of several other of the minisparker records from cruise 76-G-10 showed that several transects exhibited a tilt in the interface. One of the profiles examined is shown in Fig. 17. The right hand side of this profile, which was taken along the longest section of the big basin is slightly higher than the left hand side of the profile. Energy to induce seiching could result from internal waves or even storms passing over with a reduction in barometric pressure. Merrell (1971) has calculated that energy from passing storms can influence bottom currents several thousand meters below.

When seiching occurs in the basin, turbulent mixing will occur on both sides of the brine sea water interface (layer 5). This is identical to the case in fluid mechanics of oscillations of the common boundary of two superimposed liquids. The motion will be damped more quickly than seiches in lakes since the low density contrast has the effect of diminishing the velocity of propagation of waves of any given length in the ratio  $[(1-s)/(1+s)]^{1/2}$ , where  $s$  is the ratio of the density of the Gulf water to that of the brine (0.88).

In oscillations of a water mass with two layers, there will be a discontinuity of motion at the interface. Fig. 18 shows mixing processes as a result of seiching in a two layer system with arrow vectors indicating the magnitude and direction of the water movements. The normal velocities are continuous, but the tangential

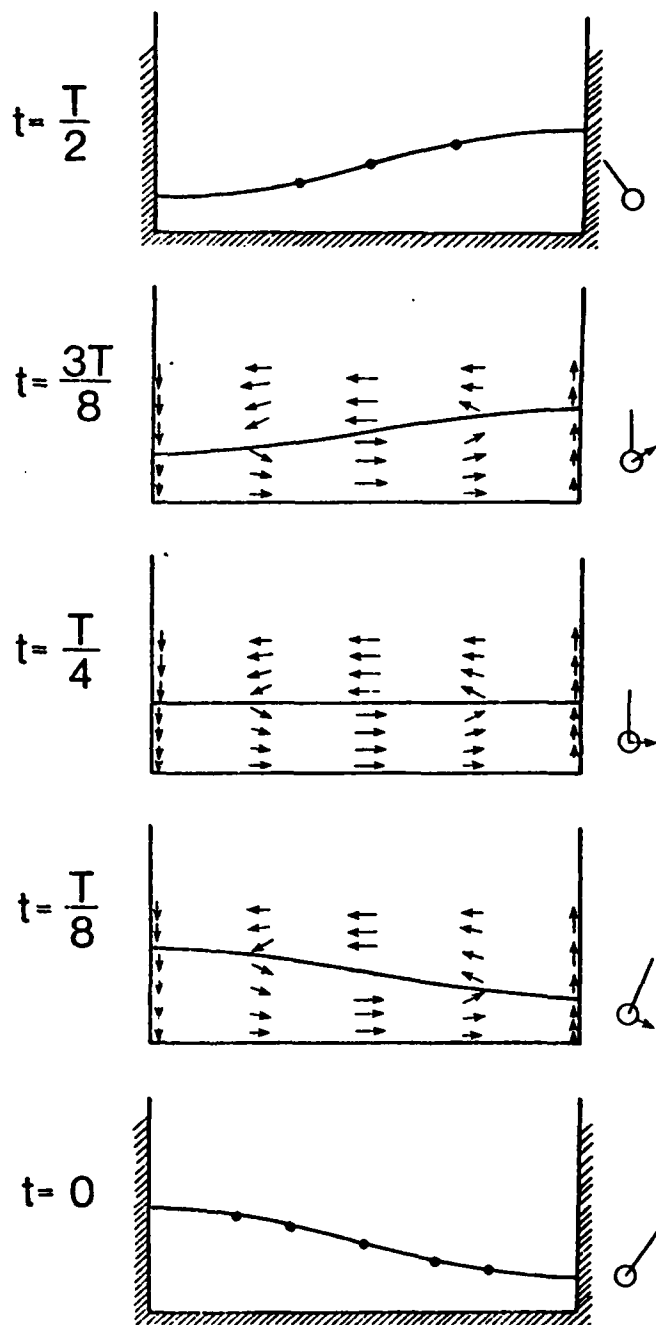


Fig. 18. Oscillations of a seiche in steps of one-eighth of a cycle for one-half of a complete oscillation. The depth is exaggerated (after Neumann and Pierson 1966, with modifications to show the two-layered system).



velocity changes direction as the interface is crossed (Lamb 1945). With the mixing at the interface, it is possible that the oxygenated water from above would mix with the brine from below and produce the oxygen step region that was observed. The diffusivities above the brine (Fig. 16) should be high enough to allow mixing of oxygen downward into this region.

CHAPTER III  
MAJOR ATMOSPHERIC GASES IN THE  
ORCA BASIN BRINE

Of the twelve atmospheric gases listed in Table 1, only nitrogen, oxygen and argon have concentrations of about one percent (Ar) or larger ( $N_2$ ,  $O_2$ ). These major, atmospheric gases have been measured in many oceanic environments. Their high atmospheric concentration results in dissolved levels in sea water which are great enough to be measured by simple gas chromatographic techniques. Oxygen is by far the most important gas in the ocean. It is required by all marine animals for survival and is produced during primary production by the phytoplankton in the surface oceans. In deeper waters, oxygen is consumed more rapidly than it is produced. The term apparent oxygen utilization (A.O.U.) has been defined (Redfield 1934) as the amount of oxygen removed due to respiration processes.

Argon and nitrogen are generally regarded as nonreactive in ocean waters. Some qualification of this assumption may be necessary for nitrogen, since it has been shown to be produced in the anoxic waters of the Cariaco Trench (Richards and Benson 1961; Linnenbom and Swinnerton 1969) and in some anoxic sediments (Wilson 1978). The oceanic distribution of argon and nitrogen is determined by physical mixing processes and by the effects of temperature and salinity on their solubility. In contrast, the

distribution of oxygen is influenced by biological as well as physical properties. Thus measurements of unreactive gases, along with oxygen, are useful in obtaining a quantitative separation of the effects of physical and biological processes on the distribution of oxygen in the ocean.

As was discussed in detail in Chapter I, the atmospheric equilibrium of dissolved gases in sea water is a function of both the temperature and salinity. The atmospheric equilibrium solubilities of oxygen, nitrogen and argon in sea water at 35‰ salinity are given in Table 8. These values were calculated from Eq. 5, using the constants determined by Weiss (1970). A plot of temperature versus solubility for argon and nitrogen (Fig. 19) shows that the relationship is not a linear one. This concept is important in interpreting the gas solubility data from a particular area, since the mixing of two water masses with the same salinity and different temperatures would produce an apparent supersaturation. For example, if two waters of 35‰ salinity, one having a temperature of 0°C and the other at 30°C, the apparent supersaturation would range from 7.8% for nitrogen to 8.6% for argon (Fig. 19).

In studying the major atmospheric gases in the Orca Basin, I have examined both the concentrations and the solubilities of the gases in the brine. A major purpose of these measurements is to determine whether or not there has been dissolved nitrogen

Table 8. Solubilities of N<sub>2</sub>, O<sub>2</sub>, Ar in sea water at 35‰ salinity based on Eq. 5 and coefficients from Weiss (1970).

t°C	Solubilities ( $\mu\text{mol}\cdot\text{kg}^{-1}$ )		
	N <sub>2</sub>	O <sub>2</sub>	Ar
0	616.3	349.5	16.98
5	549.6	309.1	15.01
10	495.6	274.8	13.42
15	451.3	247.7	12.11
20	414.4	225.2	11.03
25	383.4	206.3	10.11
30	356.8	190.3	9.33

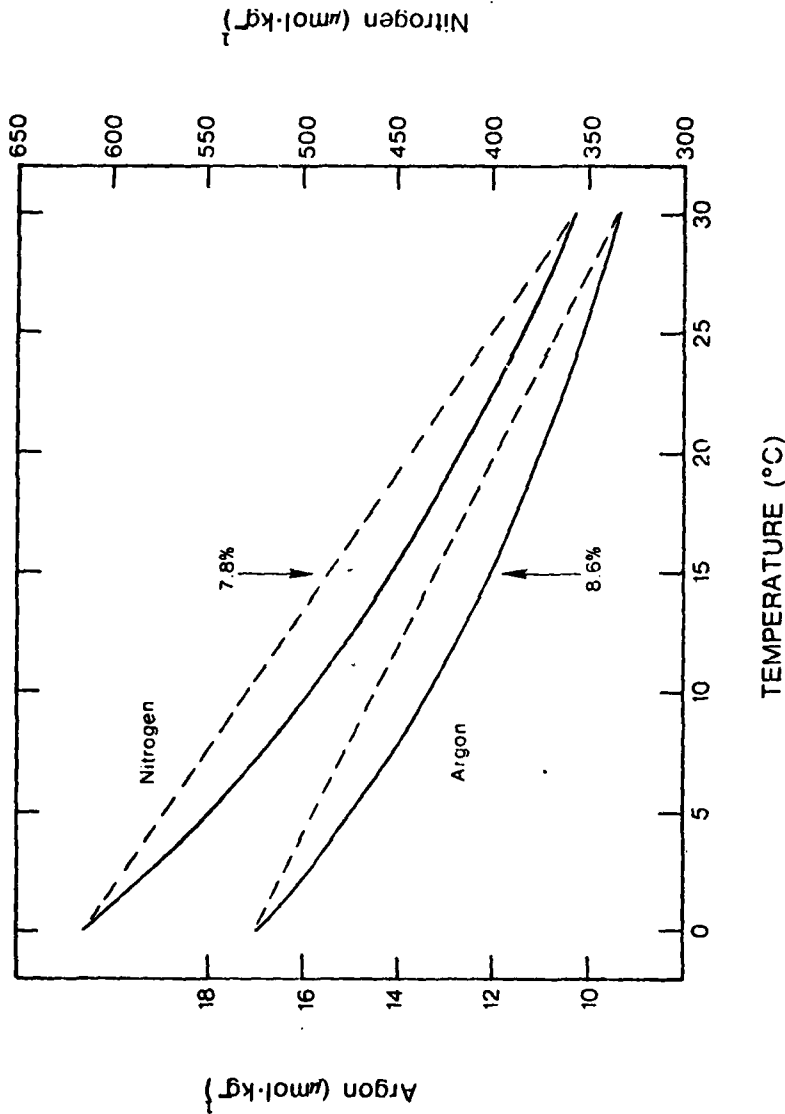


Fig. 19. Plot of temperature versus atmospheric solubility for argon and nitrogen, indicating that the relationship is not a linear one. Thus, if two water masses of equal salinity but different temperatures are mixed, an apparent dissolved gas supersaturation will be observed without the addition of any gas. Examples are shown for mixing of water at 0°C with water at 30°C (dashed line).

production in the brine. It will be observed that both nitrogen and argon are supersaturated in the brine relative to the brine temperature and salinity at one atmosphere total pressure. This chapter addresses the question of the origin of nitrogen and argon in the brine and what one can learn from these unreactive gases concerning the origin and geochemistry of the Orca Basin.

#### Methods

##### Gas Solubilities

Since gas solubilities had not been previously measured in hypersaline waters, there was a need to determine whether or not actual measured solubilities deviated significantly from the extrapolated data obtained using the equations of Weiss (1970). To equilibrate the brine with gases, a glass, thermally-regulated equilibration chamber was constructed using a design similar to one used by Schmidt (1979) for measuring atmospheric solubilities of hydrogen and carbon monoxide (Fig. 20). Into this chamber, approximately 400 ml of brine (or sea water for comparison) was introduced and gas was bubbled through at a rate of  $40 \text{ ml} \cdot \text{min}^{-1}$  for 30 min. The time required to saturate the brine with the equilibration gas should follow a simple first order exponential decay curve, assuming that equilibrium is reached between the bubbles in the equilibration gas and the bubbles in the stripper.

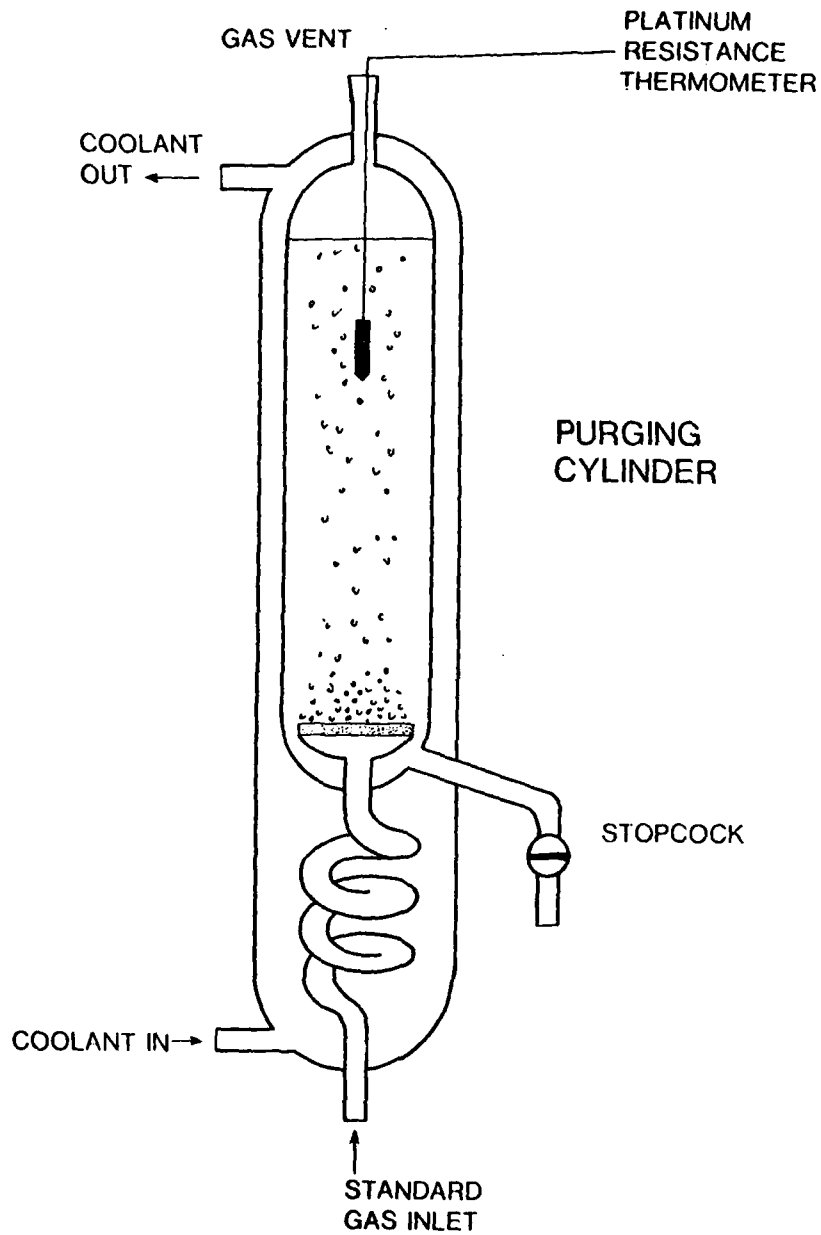


Fig. 20. Schematic diagram of thermal jacketed purge cylinder used to equilibrate the brine with dissolved gases.

The characteristic time required to increase the concentration of the gas  $i$  in solution by a factor of  $1/e$  is given by

$$t_i = \frac{B_i V_l P}{F_i^+} \quad (13)$$

where  $B_i$  is the Bunsen solubility coefficient of the gas  $i$  at the temperature of the liquid,  $V_l$  is the volume of the liquid,  $F_i^+$  is the volume rate of flow of  $i$  in the equilibration gas at STP, and  $P$  is the partial pressure of  $i$  inside the equilibration chamber, in atmospheres (1 atm = 101.325 kPa). To make this calculation, Bunsen coefficients were calculated for each gas, and other values (e.g. flow rate) were measured. At 0°C the characteristic time required to equilibrate sea water at 20°C was 15 sec for oxygen, 17 sec for argon and 8 sec for nitrogen. For each of the brine and sea water measurements, the equilibration gas was allowed to flow for a period of 30 min to assure that complete equilibration was reached. Air was used for the nitrogen source gas yielding atmospheric equilibrated values. Pure (>99.995%) argon and methane were used to measure Bunsen coefficients for those gases.

Since the bubbling of a gas through a liquid may tend to supersaturate the liquid with the gas used, one further precaution was taken during these measurements. After the 30 min equilibration period, the equilibration gas was shut off and the system sealed with one atmosphere of gas above the sample for two min before the brine was sampled. Analysis of the dissolved gas was



accomplished using the same gas chromatographic techniques as were used for Orca Basin samples (see below). As an additional check on possible supersaturation, sea water samples of a known salinity were analyzed using the same procedures as for the brine. Since the equilibrium solubilities of dissolved gases in sea water of normal salinity are known very precisely as a function of temperature and salinity, these measurements served as external standards for the equilibrium process.

The temperature of the brine in the equilibrator was monitored using a thin film platinum resistance electrode which was epoxy coated and immersed in the brine during the equilibration process. This probe was calibrated against the high precision platinum electrode (Leeds and Northrup, S/N 1762254) used to calibrate the Texas A&M University reversing deep-sea thermometers. The high precision probe is calibrated periodically against NBS temperature standards. The temperature measurements in this equilibration study are accurate to  $\pm 0.02^{\circ}\text{C}$ .

#### Dissolved Oxygen

Dissolved oxygen was measured on the sea water samples we collected from the Orca Basin by the microWinkler method of Carpenter (1965). No oxygen was found in the high salinity brine so no problems due to effervescing of the brine were observed here. Nor was the degassing effect noted in samples taken from the

interface where small amounts of oxygen had been previously measured (Shokes et al. 1977). As discussed previously, Shokes et al. (1977) had found an unusual step-like feature in the oxygen profile just above the brine interface, with values constant at about  $0.1 \text{ ml}\cdot\text{liter}^{-1}$ . After Wiesenburg et al. (1977) reported finding high nitrite concentrations in the interface area of the Basin, the standard Winkler technique was modified by adding one gram of sodium azide to the NaOH-NaI reagent. Any nitrite in a Winkler oxygen sample will reduce iodide to iodine just as oxygen does, thus erroneously causing a larger value to be reported. Adding sodium azide to the alkaline iodide reagent will prevent this nitrite interference (Broenkow and Cline 1969).

#### Argon and Nitrogen Sampling

Samples obtained from the Orca Basin brine for argon and nitrogen are supersaturated with these gases when sampled at one atmosphere total pressure. Since these gases will bubble out of the water during sampling, special precautions had to be taken to avoid both atmospheric contamination and loss of gas due to effervescence of the samples. A system was devised to capture the water sample before degassing took place.

A major problem with brine degassing was related to the type of sample bottles used. All plastic Niskin sampling bottles are the preferred water sampling device for marine geochemists. They

provide large volumes of water, are relatively inert and are trace metal free. These bottles, however, have a closing device comprised of a rubber band between two end caps. The pressure on the seal is minimal at best, and could not keep the Orca Basin gases from effervescing in the sample bottle, before they could be transferred to some suitable sample container. Sampling with teflon-coated, brass, 1.2 liter Nansen bottles proved more fruitful. These bottles have a metal, ball-valve type sealing system which closes off both ends of the bottle with a metal-to-metal seal when the bottle is tripped. The seal produced by these bottles was tight enough to prevent gas effervescence before transfer from the Nansen bottle. However, using Nansen bottles, there was more variability between samples in the brine than can be accounted for by analytical error (assuming the brine is a homogenous mixture) so some of the Nansen bottles probably worked more effectively than others.

To test the ability of the Nansen bottles to capture reproducible samples, samples were taken with gas tight "piggyback" bottles designed from the information given by Weiss (1968). These samplers consisted of a 0.95 cm O.D. (3/8 in) copper refrigeration tube placed between two high pressure ball valves (Hoke No.1874392). The ball valves of the piggyback sampler are attached to the ball valves on the Nansen bottles. This is accomplished by attaching a flange to the Nansen bottle to replace the nut which normally holds the Nansen ball valve together. The piggyback sampler is

attached to this flange. When the Nansen bottle is tripped, the ball valves on the piggyback sampler are closed simultaneously with the valves of the Nansen bottle. When the high pressure ball valves are closed, they trap a water sample at in situ pressure in the copper tube. With a tubing length between the valves of 56 cm and an 0.64 cm I.D., a sample of approximately 23 ml is collected. The sample is returned to the surface in the tubing at the in situ pressure. At the surface, a segment of the sample is sealed off in the copper tubing by placing refrigeration pinch off clamps (Imperial Eastman Corp., 105-FF pinch off tools) at either end of the tubing between the ball valves. The tubing and clamps can then be removed from the ball valves with a sealed off sample available for analysis in the laboratory.

Since the high pressure ball valves and the copper tubing have only a 0.63 cm (1/4 in) internal diameter, flow through the tubing can be impeded. Restriction of flow would require a larger flushing distance (volume) to assure a representative water sample. To insure proper flushing when lowering, a funnel is attached to each of the ball valves. These funnels have a maximum diameter of 5 cm, a size selected to give the greatest flushing with the minimum drag through the water. Weiss (1968) used smaller size funnels and tested the flushing efficiency of the samplers using fluorescein dye. The dye was placed in a 1.8 m curved copper tubing attached to the ball valves and lowered to various depths. After

tripping the sampler and retrieving the water, the remaining fluorescein dye was measured with a fluorometer. Casts to various depths were carried out. Extrapolations from the larger volume, curved tube to the normal 56 cm tube used in this study indicated that a normal piggyback sampler should flush 50% in approximately 2 m of lowering. To assure complete flushing in the samples from the Orca Basin brine, all bottles were placed at least 10 m apart, and they remained opened at depth for at least 15 min before they were closed (while the thermometers equilibrated). Comparison of samples in the narrow interface region indicated that sample flushing was essentially complete when these funnels and this sampling procedure were used.

The amount of water sampled by the piggyback sampler can be measured in either of two ways. The volume of water can be collected after analysis and measured volumetrically. This is the preferred method if the sample is to be analyzed aboard ship. If the sample is measured in the laboratory, the weight of the tubing, clamps and sample can be measured before analysis. After the water sample is removed from the tubing, the tubing and clamps can then be reweighed to determine the weight of the sample. Since the temperature and salinity of the water are usually available from the water sample taken in the companion Nansen bottle, a density can be calculated for the sample. With the mass and density, the volume of water that was analyzed can be determined. This second

procedure described here was used in obtaining sample volume during this work, since all piggyback samples were analyzed in the shore-based laboratory at Texas A&M University.

To obtain companion water samples for dissolved gas analysis from the Nansen bottles (both with and without piggybacks), several all-glass sampling chambers were constructed (Fig. 21). These devices were calibrated gravimetrically to a volume of 0.01 ml. Samples were taken from Nansen bottles by attaching a short piece of Tygon tubing between the spigot of the bottle and the lower port of the sample holder. Each end of the sample holder was equipped with a three port stopcock which was used both to seal the samples and to transfer them into a stripping chamber for gas analysis. The water from the Nansen bottle was allowed to flow into the glass water sampler through the lower port and to overflow a volume equal to two or three volumes of the sampler through the upper port. When the sampler had been flushed, the bottom and then the top stopcocks were closed, sealing the water sample in the container. This sample could then be attached to the gas chromatographic gas stream and analyzed as described in the next section.

#### Chromatographic Analysis of Argon and Nitrogen

The gas stripping and chromatographic system for the analysis of argon and nitrogen is shown in Fig. 22. The gas system is

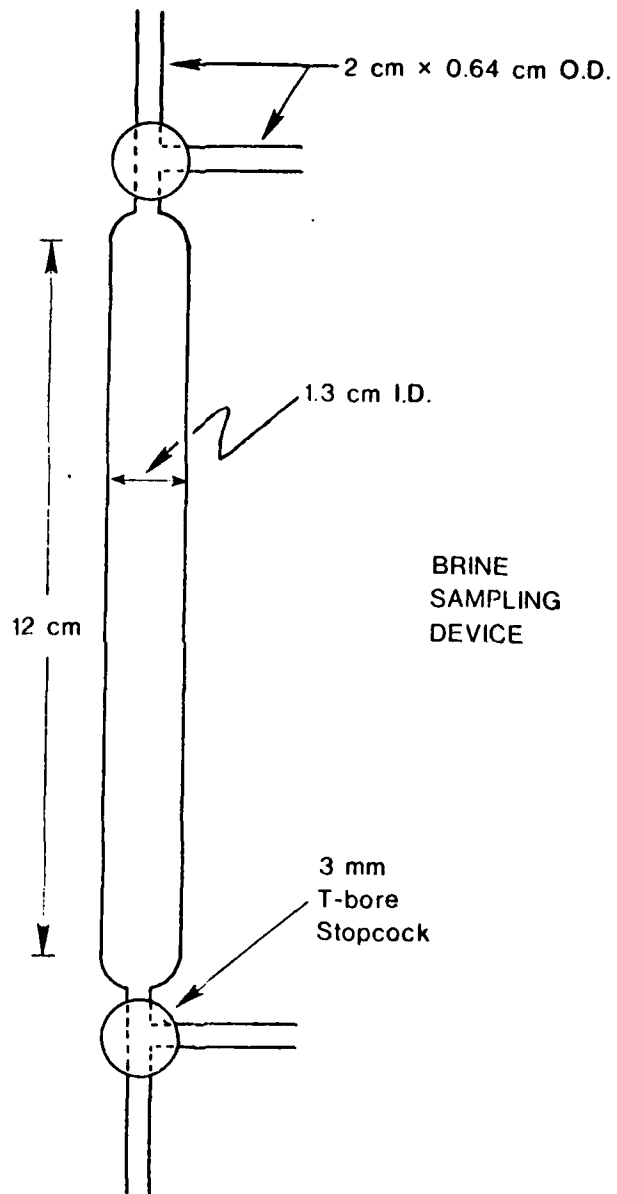


Fig. 21. All-glass sample chambers constructed to take uncontaminated water samples from Nansen bottles for dissolved gas analysis.

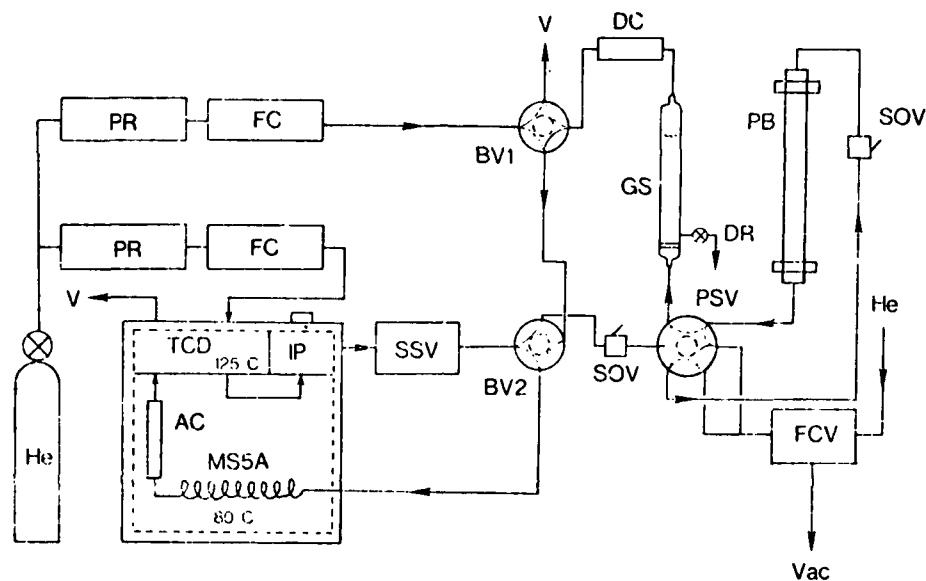


Fig. 22. Dissolved gas extraction and analysis system. The piggyback water sample (PB) is attached to the system using valve PSV. Any trapped air is removed from the gas lines by alternately evacuating the system and purging it with helium using valve FCV. With valve FCV and shut-off valve SOV closed, the piggyback clamps are removed and the water sample is transferred to the gas stripper (GS) using valve PSB (after reopening valve SOV). Gas dissolved in the water is extracted by the helium flow, dried on a drying column (DC), separated on a 5A molecular sieve column (MS5A), and analyzed using a thermal conductivity detector (TCD). An activated charcoal column (AC) is used before the detector to remove oxygen from the sample. Standard gas samples are introduced through a standard sample valve (SSV) with a 0.5 ml sample loop. Standards can be bypassed directly to the gas chromatograph using bypass valve two (BV2) or it can be passed through the stripper and drying columns before analysis. Bypass valve one (BV1) is used to provide helium to the thermal conductivity detector when the gas stripper is being drained and purged after sample analysis.



designed so that the piggyback sampler can be attached to the gas lines and all contaminant air can be removed before the pressurized brine sample is released to the gas stream. For this purpose, the piggyback sampler (PB) is attached to the gas lines using the same 3/8 in (0.95 cm) Swagelok fittings with which it was attached to the high pressure ball valves. When the sampler is connected at both ends, it is isolated from the stripping gas stream by a 6-port delrin plastic sampling valve (PSV). With a 3-port brass flow control valve (FCV), the gas stream connected to the isolated PB sampler can be alternately purged with helium and evacuated. The backpressure valve (SOV) is open during this air removal process. The gas lines were evacuated and purged with helium ten times to insure complete removal of atmospheric gases. Analysis of gas remaining in the lines after ten purgings and evacuations indicated no detectable argon or nitrogen. After the tenth purging, helium was left in the gas line and the system was isolated by moving valve FCV to the neutral position. Valve SOV was then closed and a positive helium pressure of 240 kPa was maintained in the system.

With valves FCV and SOV closed, the clamps are removed from the PB sampler and used to re-round the 3/8 in (0.95 cm) O.D. tubing, thus releasing the sample to the gas line. Immediately after the tube is opened, valve SOV is opened and the 6-port rotary valve (PSV) is turned clockwise transferring the brine into a 35 cm by 1.5 cm O.D. glass stripper chamber (GS) which has a

10 mm coarse glass frit at the bottom. Helium from the gas chromatographic stream pushes the brine downward from the PB sampler through the 6-port valve (PSV) and into the stripper. When all the brine has been pushed through the frit, the carrier gas begins to bubble through the sample thereby stripping the dissolved gases out of solution. Weiss and Craig (1973) have given a complete description of stripping characteristics and efficiencies of this type of gas extractor.

The effluent carrier gas and extracted gases exit at the top port of the stripper and pass through a 20 mesh indicating drierite ( $\text{CaSO}_4$ ) column to remove water vapor. After the drying process, the extracted gases pass through two bypass valves (BV1 and BV2) and are subsequently separated and detected using gas chromatographic methods. A Hewlett-Packard 5830A gas chromatograph with a thermal conductivity detector was used for separation and analysis. The sample gas is separated on a 30/60 mesh activated 5A molecular sieve column (2.0 m in length and 3.6 mm internal diameter) which retards  $\text{N}_2$  relative to the combined  $\text{O}_2 + \text{Ar}$  peak. At the  $80^\circ\text{C}$  operating temperature, this column also absorbs any evolved  $\text{CO}_2$ .

Argon and oxygen cannot be resolved with the column and temperature settings used here. In past studies, various techniques have been used to separate the argon and oxygen components. Without reacting the gases, they can be separated using either columns which are extremely long or which must be operated at very low

temperatures. Weiss and Craig (1973) used a hydrogen carrier gas and a palladium catalyst to convert oxygen to water, enabling the analysis of oxygen in the form of a water peak. Since there is no oxygen in the Orca brine and Winkler oxygen measurements of  $O_2$  were available in the overlying water, this author has chosen to use a simpler analytical technique. Oxygen is simply removed from the gas chromatographic stream, thus enabling the argon concentration to be measured without interference.

The removal of oxygen from the gas chromatographic stream was accomplished by placing a stainless steel column (0.5 m in length by 3.6 mm internal diameter) containing "superactivated charcoal" (Cooke 1973) between the MS5A column exit and the thermal conductivity detector. When activated charcoal is heated to temperatures greater than  $800^\circ\text{C}$  and then cooled to  $80^\circ\text{C}$ , it will quantitatively absorb oxygen passed over it. This method was found to be superior to removing  $O_2$  by reaction with a solution of chromous sulfate (Lingane and Pecsok 1948) that has been used by other investigators (Reeburgh 1969, 1976). Typical chromatograms for room air analyzed with (B) and without (A) the oxygen removal column are shown in Fig. 23.

Gas analysis after separation is performed using a thermal conductivity detector operating at  $125^\circ\text{C}$ . Flow rate through the detector was  $40 \text{ ml}\cdot\text{min}^{-1}$ . Detector response is monitored using the thermal printer-plotter associated with the chromatograph.

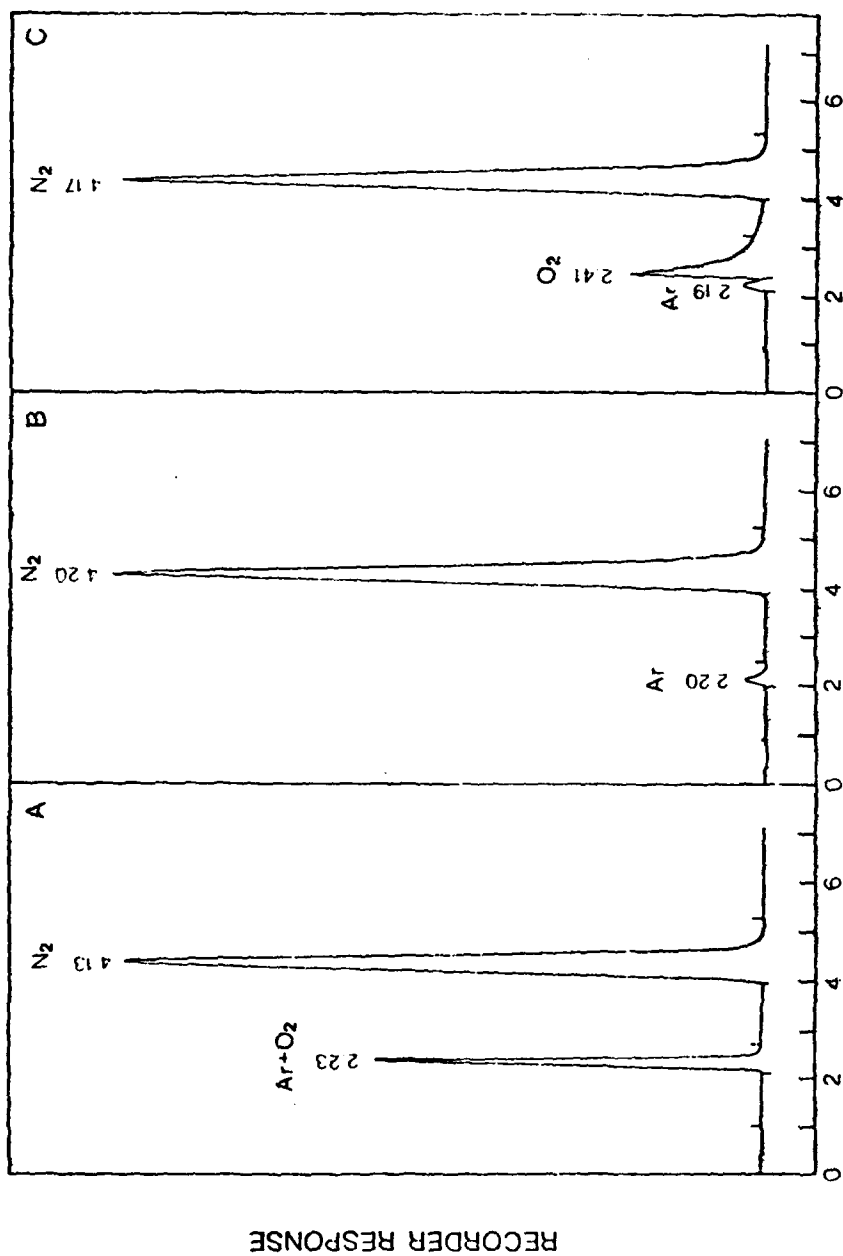


Fig. 23. Typical chromatograms showing the analysis of major dissolved gases in oxygenated sea water with (B) and without (A) the activated charcoal trap in the gas flow line. Deactivation of the trapping column can be easily recognized (C) when encroaching oxygen peaks begin to appear as tailing peaks after the argon peak.

Peak areas are automatically integrated and retention times recorded by the microprocessor system of the 5830A gas chromatograph and are automatically reported along with the peak area percentages. A typical chromatogram for Orca Basin brine is shown in Fig. 24.

The chromatographic system was standardized for argon and nitrogen analysis by injecting standard volumes of dry air into the column using a calibrated sampling loop (0.493 ml) and an automated sampling valve (SSV). The gas concentration in the sample in units of ml (STP)/ml is given by

$$C_x = \frac{A_x}{A_s} \cdot \frac{V_g}{V_l} \cdot \frac{T_o}{T_s} \cdot \frac{P_s}{P_o} \cdot C_s \quad (14)$$

where  $A_x$  and  $A_s$  are the peak area for the particular gas in the unknown sample and the calibration mixture, respectively;  $C_s$  is the standard gas concentration (v/v);  $V_g$  is the sample loop volume and  $V_l$  is the liquid volume.  $P_s$  and  $T_s$  are the pressure and absolute temperature in the sample loop; and  $P_o$  and  $T_o$  are the standard conditions of temperature (273.15°k) and pressure (1 atm) for gases. Corrections for nonideality are not necessary for argon, nitrogen or methane under these conditions (see Table 1).

The glass container water samples taken from the Nansen bottles were analyzed using the same chromatographic system used from the piggyback (PB) samples. The gas stripping system described above was also used, but it was modified slightly to account for the varying requirements of the glass samplers. The

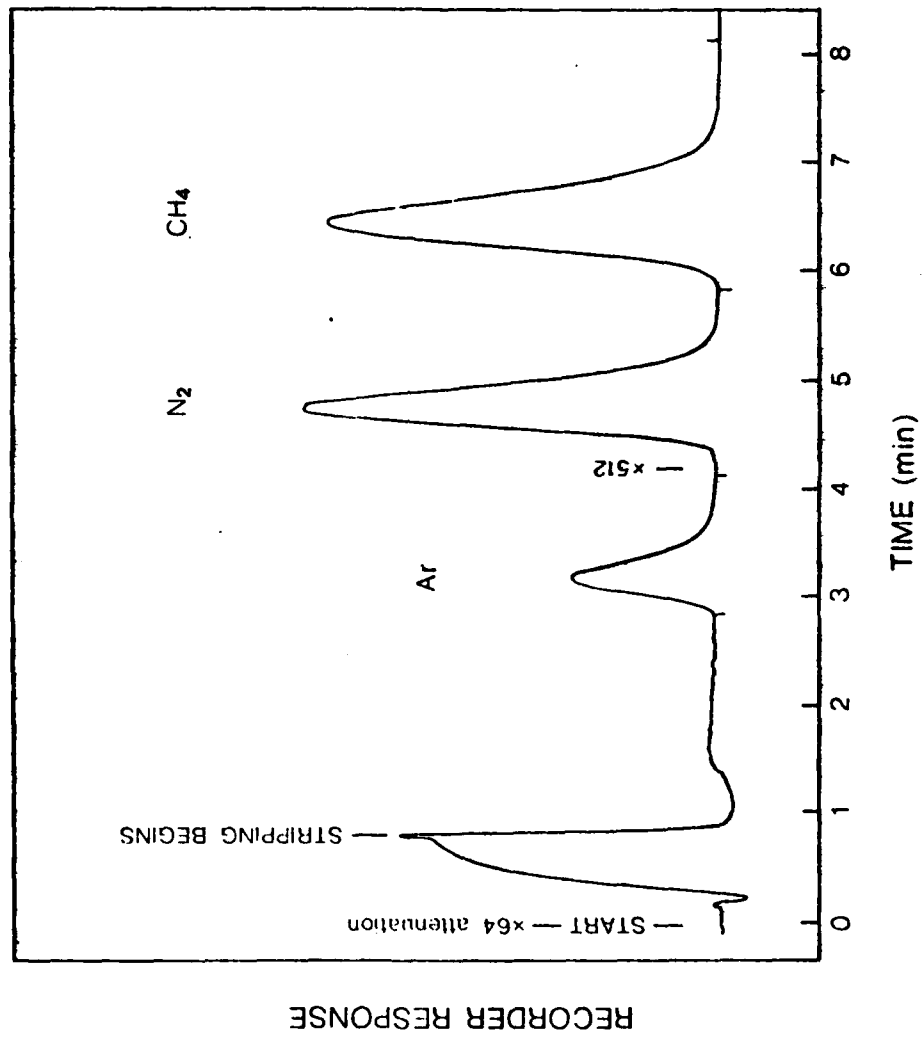


Fig. 24. Typical gas chromatograph for the anoxic water of the Orca Basin derived from the analytical techniques described in Fig. 15.

samplers (shown in Fig. 21) were attached to the gas stream between the PB sample valve (PSV) and the gas stripper (GS). The gas bypass valve (BV1) was turned clockwise passing clean helium to the TCD while the sampler gas lines were being purged. In purging the lines, the water sampler stopcocks are turned so the carrier gas passes around the sample, through the stripper and drying column and exits the vent port of bypass valve BV1 (see Fig. 22). After five minutes of purging, the lines are free of any contaminating atmospheric gases and valve BV1 is returned to its normal position (counter-clockwise). The carrier gas now flows again to the GC column and detector. After the detector baseline was restabilized from flow switching, the two three-way stopcocks of the glass samplers were turned to allow the helium carrier to push the brine into the stripper for analysis as described above.

## Results

### Solubility Comparisons

The solubility measurements made on the brine and corresponding sea water samples are given in Table 9. Only a few measurements were made, since the purpose here was not to establish a complete data set of brine solubility data. A high precision solubility data set would require microgasometric equipment (Douglas 1964) which was not available. The object of these measurements was simply to determine whether or not the equations defined

Table 9. Measured gas solubilities of Orca Basin brine and sea water. All values are atmospheric equilibrium solubilities except those marked with an asterisk.

Solution	Gas	Temp. (°C)	Sal. (‰)	Predicted Solubility ( $\mu\text{mol}\cdot\text{liter}^{-1}$ )	Measured Solubility <sub>-1</sub> ( $\mu\text{mol}\cdot\text{liter}^{-1}$ )	% Difference
Sea Water	N <sub>2</sub>	4.26	34.635	575.2	579.4	+0.7
	N <sub>2</sub>	11.17	34.635	498.3	515.5	+3.5
	N <sub>2</sub>	20.49	33.970	423.7	431.2	+1.8
	Ar	4.26	34.635	15.70	16.28	+3.7
	Ar	11.17	34.635	13.47	14.14	+5.0
	Brine	N <sub>2</sub>	4.21	250.0	121.9	100.7
N <sub>2</sub>		10.89	250.0	114.7	99.8	-13.0
N <sub>2</sub>		10.94	250.0	114.7	101.1	-11.8
N <sub>2</sub>		21.01	250.0	106.5	92.4	-13.3
N <sub>2</sub>		21.01	250.0	106.5	90.9	-14.7
Ar		4.21	250.0	3.75	3.48	-7.1
CH <sub>4</sub> *		25.06	250.0	306.6	269.9	-12%
CH <sub>4</sub> *		22.86	250.0	313.2	288.1	-8%
Ar*		4.21	250.0	402.0	314.1	-21%

\* Pure gas.



by Weiss (1970) could be extrapolated from sea water salinities to the higher salinity of the brine. This extrapolation was believed to be feasible since the major factor governing gas solubility in saline solution is the temperature. Salinity has a smaller effect. Yet, since the constants for salinity in Eq. 5 are small relative to the temperature coefficients, they can cause a large error to be propagated in any extrapolation process. The measured solubility data from this work is compared with the predicted solubility values in Table 9. Percentage differences between measured and calculated values are indicated.

Due to low solubility of argon in brine, it was difficult to measure precisely the argon concentration which resulted from equilibrating the brine with air in the equilibration chamber. One successful argon measurement of the atmospheric equilibrium solubility of argon was made at 4.21°C. At higher temperatures, however, the digital integrator would not sense the small argon peaks with enough regularity to integrate them accurately. Thus, pure argon was used to measure the Bunsen coefficient of argon. Also, the concentration of methane in the standard atmosphere (1.41 ppm; Prabhakara et al. 1974) does introduce enough methane into the equilibrated brine to measure with a thermal conductivity detector. Pure methane was used to determine the Bunsen coefficient of that gas as well.

The solubility data for the brine for all three gases

indicated that the equation of Weiss (1970) predicts a gas concentration in the brine that is over ten percent higher than the measured values. This agreement is not bad, considering the difficulties in extrapolating the equation to salinities an order of magnitude higher than was intended. Also, the Orca brine is not just a higher concentration of sea water. It is a sodium chloride saturated solution, with a mixture of other components. For example, total carbon dioxide of the brine is twice normal sea water values (Sackett et al. 1979). These other compounds also contribute to the fact that lower values were observed than were predicted.

The equilibrated normal sea water samples analyzed by this procedure show a slight supersaturation, in the range of one to five percent. This supersaturation was expected as has been discussed previously. These results indicate that it was not a complication of the analytical technique which caused the lower levels of argon, nitrogen, and methane solubility, relative to the calculated values. The true values are probably lower than predicted ones, due to the differences in salt content between sea water and the brine, and due to difficulties in extrapolating empirical equations beyond their working range. This information implies that either the equation does not follow the chemistry at the higher salt concentrations or that the nature of the interactions change under hypersaline conditions. From data on the

changes in ionic activity as a function of increased ionic strength (Harned and Owen 1950), there is some indication that the latter is the case. However, that problem is not germane to this discussion.

The solubilities of argon, nitrogen, and methane are low in the brine, yet the measured values are relatively close to the predicted values. These low solubilities and the gas levels in the brine explain the effervescence which occurs when the brine is brought to the surface. Effervescing gas bubbles were captured on four samples taken directly from Nansen bottles on cruise EN-32 of the R/V Endeavor. One ml of this gas was analyzed by direct injection into the gas chromatograph. The gas collected was found to contain 1.1% argon, 63.7% nitrogen, 34.6% methane and 0.6% carbon dioxide (average of four samples). Although the methane levels are higher in the brine than nitrogen (see next Chapter), there is more nitrogen in the effervescing gas, since nitrogen is less soluble in the brine than is methane (Table 9).

#### Dissolved Oxygen

Dissolved oxygen has been measured on every Texas A&M cruise to the Orca Basin. The original data of McKee and Sidner (1976) and the plot of Shokes et al. (1977) were presented in Chapter I. I shall not give all of the newer data. The data from four cruises (77-G-2, 77-G-13-I, 77-G-13-III, and 78-G-2) are presented

in Fig. 25. These four oxygen and sigma-t profiles were taken over the span of one year. The oxygen profiles from each cruise show oxygen values at the top of the basin (2000 m) which are fairly typical of deep Gulf oxygen concentrations in this area, approximately  $225 \mu\text{mol}\cdot\text{liter}^{-1}$ . As density begins to increase (along with salinity) from the deep Gulf levels, oxygen concentrations begin to decrease gradually from about  $200 \mu\text{mol}\cdot\text{liter}^{-1}$  at a depth of 2175 m, to near zero but not exactly zero at about 2215 m. This same pattern can be seen in all the profiles. Below this gradual oxygen gradient there is a region of approximately 20 to 30 m where the oxygen concentration is very low, but measurable. The oxygen levels are between 2 and  $10 \mu\text{mol}\cdot\text{liter}^{-1}$  in this narrow zone, and absent or less than the Winkler detection limit below.

The anomalously low dissolved oxygen values are paralleled by a change in the slope of the temperature profile (see Fig. 10). The change in temperature is not as obvious as the oxygen change (step) when viewed from temperature data taken only from deep-sea reversing thermometers. However, examination of the trace taken from the STD (Fig. 10) clearly shows the discontinuity in temperature that is associated with this step in the oxygen profile. The temperature gradient in this region is different from the gradient either above or below. This oxygen step-like feature is in the region where the salinity is increasing from 50 to 70‰. (Fig. 11). The feature is a persistent one and is not an artifact

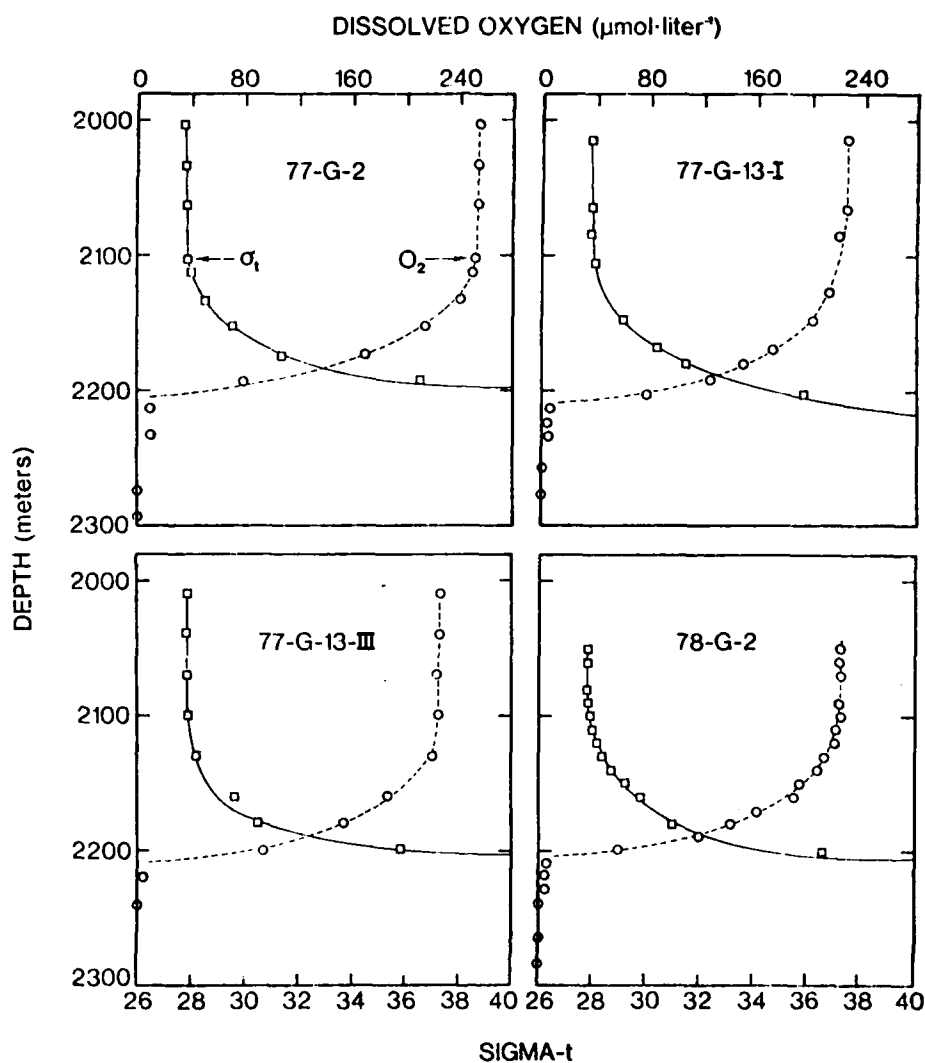


Fig. 25. Dissolved oxygen profiles measured by the micro-Winkler technique (Carpenter 1965) for four different Texas A&M cruises during 1977 and 1978. Note the persistent step in the oxygen profiles. There appears to be a systematic error in the 77-G-2 oxygen data. However, the shape of this profile is consistent with the other cruises.

of the Winkler dissolved oxygen technique.

#### Argon and Nitrogen

Attempts to sample and analyze the Orca Basin brine for argon and nitrogen were made on four cruises during 1977 and 1978. The early efforts yielded only limited success. Plastic syringes used on the first cruise allowed gas to diffuse out of the sample before analysis. The sampling systems described in the previous section were used on the last three cruises (77-G-13, 78-G-2 and EN-32). Even using these more sophisticated techniques, some samples were contaminated with atmospheric gases or depleted due to degassing of the brine. Criteria for determining the reliability of the data were established. One evaluative method was examining the methane concentration of each sample. Since methane is relatively constant in the brine (Sackett et al. 1979), extremely low methane concentrations in the brine samples would indicate gas loss before sample analysis. Nitrogen/argon ratios were also used in sample evaluation. Samples which had both an unusually high nitrogen/argon ratio high argon concentrations and very low methane levels were obviously contaminated with air ( $N_2/A$  in air = 83.6) and these data were discarded.

In the highest salinity brine ( $S = 262\text{‰}$ ), 25 measurements of nitrogen were reliable and 9 argon values were believed correct. The number of argon values is lower since this gas is found in very low concentrations in the brine and the gas chromatographic

peaks for this gas were not always integrated properly. Also, the standard for this gas was air and could only be determined when the activated charcoal column was in place in the analysis system (AC, Fig. 22). Since the column was not always used in the analysis system, fewer total determinations of the argon were made in the brine.

Nitrogen values from the brine averaged  $568 \mu\text{mol}\cdot\text{liter}^{-1}$  ( $\sigma = 70$ ) while the argon measurements averaged  $15.6 \mu\text{mol}\cdot\text{liter}^{-1}$  ( $\sigma = 1.2$ ). Measured atmospheric solubilities of these gases in the brine at  $4.21^\circ\text{C}$  (Table 9) were  $100.7 \mu\text{mol}\cdot\text{liter}^{-1}$  for nitrogen and  $3.5 \mu\text{mol}\cdot\text{liter}^{-1}$  for argon. These values represent a supersaturation in the brine of 560% and 450% for nitrogen and argon, respectively. The reliable measurements of argon and nitrogen are plotted in Fig. 26. Data in this figure are a composite of gas data taken with the pressurized piggyback samplers (open symbols) and Nansen samples collected in glass containers (closed symbols). Methane data analyzed at the same time (discussed in the next chapter) are presented in Fig. 26 as a reliability comparison and to indicate the demarkation line between Gulf water (no  $\text{CH}_4$ ) and the brine (average  $\text{CH}_4 = 714 \mu\text{mol}\cdot\text{liter}^{-1}$ ).

One nitrogen point is not plotted, although it could be significant to the following discussion. The nitrogen gas concentration at 2234 meters on cruise 77-G-13 was measured at  $1061 \mu\text{mol}\cdot\text{liter}^{-1}$  using a piggyback sampler. The argon concentration was  $9.86 \mu\text{mol}\cdot$

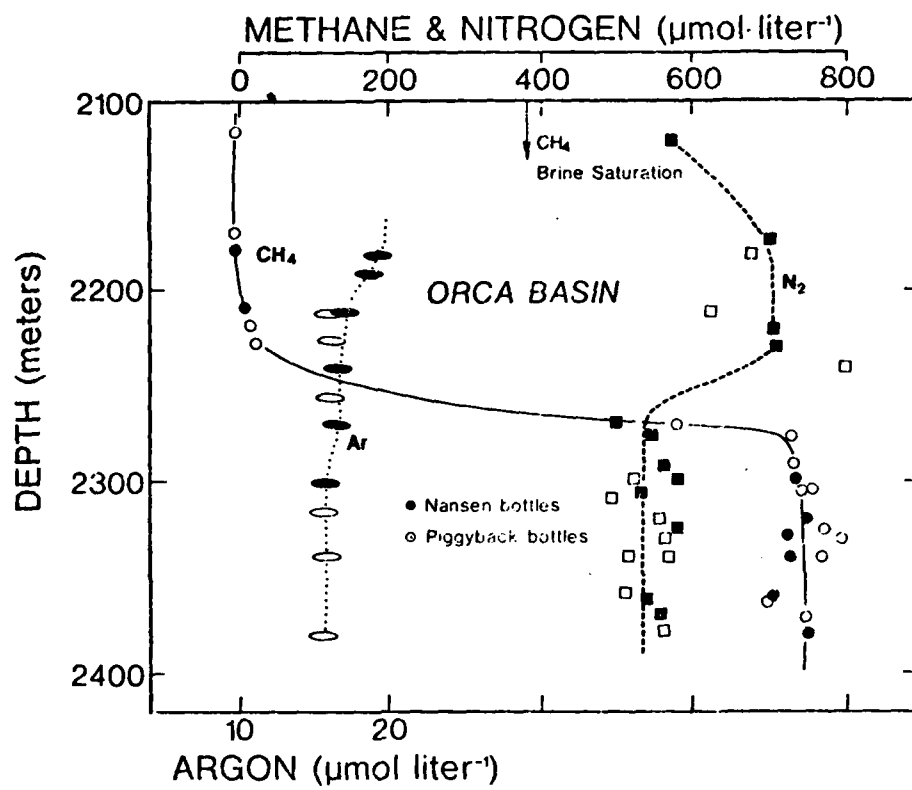


Fig. 26. Vertical profiles of dissolved argon, nitrogen, and methane through the Orca Basin interface region into the high salinity brine. The  $\text{N}_2/\text{Ar}$  ratio in the brine is 36.4.

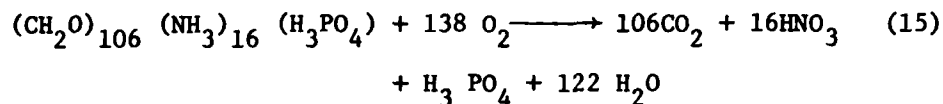


liter<sup>-1</sup> and the salinity of the water at this depth was 64.65‰. This sample is found in the oxygen step region where O<sub>2</sub> levels ranged from 4-20 μmol·liter<sup>-1</sup> (Fig. 18). Several other nitrogen measurements just above the high salinity brine also reflect levels higher than would be predicted from atmospheric equilibrium solubility. The bulge in the N<sub>2</sub> distribution between 2185 m and 2250 m (Fig. 26) also shows supersaturation, while the value at 2169 m (N<sub>2</sub> = 567 μmol·liter<sup>-1</sup>) is within 1.5% of the predicted sea water solubility value. The one very high nitrogen value in the oxygen step region may be questionable, since the associated Ar measurement is so low. The N<sub>2</sub>/Ar ratio (108) cannot, however, be accounted for by atmospheric contamination. The other N<sub>2</sub> and Ar data in this region are considered reliable for the following discussion.

### Discussion

#### Dissolved Oxygen

The dissolved oxygen profiles in the Orca Basin (Fig. 25) show a general decrease through salinity transition zones T1 and T2. A constant but low oxygen level was observed in transition zone T3. Oxygen was absent in the interface region and high salinity brine. Oxygen is the first oxidizing agent consumed in the organic matter decomposition process (Richards 1965). The equation used to describe oxygen consumption is



In this decomposition reaction, 138  $\mu\text{mol}$  of oxygen are consumed for every 106  $\mu\text{mol}$  of carbon dioxide that are produced. If the brine was originally Gulf sea water ( $T = 4.2$ ,  $S = 35\text{‰}$ ), it would have had an original oxygen concentration of 322  $\mu\text{mol}\cdot\text{liter}^{-1}$  (Weiss 1970). Removal of this amount of oxygen via Eq. 14 would produce 247  $\mu\text{mol}$  of carbon dioxide. The amount of carbon dioxide that has been added to the Orca brine due to decomposition has been calculated as 2290  $\mu\text{mol}\cdot\text{liter}^{-1}$  (Sackett et al. 1979). Thus, the  $\text{CO}_2$  input from oxygen consumption would account for only 11% of the total  $\text{CO}_2$  input. The major input of carbon dioxide must result from other, anaerobic decomposition processes which will be discussed in the next chapter.

The decrease in oxygen in the transition zone must be due to oxygen consumption as described in Eq. 14. The convex downward slope of the profiles in regions T1 and T2 are indicative of consumption in that region. The increase in excess total carbon dioxide through this region is about 470  $\mu\text{mol}\cdot\text{liter}^{-1}$  (Sackett et al. 1977), while oxygen decreases from 225 to 10  $\mu\text{mol}\cdot\text{liter}^{-1}$ . This oxygen decrease via Eq. 14 would result in the production of 165  $\mu\text{mol}\cdot\text{liter}^{-1}$  of carbon dioxide. That value represents 35% of the amount supposedly produced in this region. If oxygen is

consumed in producing this carbon dioxide, one can assume that 65% of the excess  $\text{CO}_2$  in this region comes from anaerobic decomposition or from diffusion of  $\text{CO}_2$  from the zone below.

The oxygen step zone has a low but measurable amount of oxygen. Water in this zone has a characteristic salinity between 50 and 70‰. All samples in this salinity range have measurable oxygen less than  $15 \mu\text{mol}\cdot\text{liter}^{-1}$ . No exceptions have been found. The microWinkler oxygen technique (Carpenter 1965) is not very precise in this area due to interferences at low concentrations. However, even with care to eliminate the possible effect of nitrite on the samples (Broenkow and Cline 1969), oxygen showed some variability in the step region, i.e., transition region T3. Although it is impossible to be certain, the oxygen levels in this region seem to be relatively constant. They decrease to zero within a very small depth region where salinity increases above 70‰, i.e., in the interface region.

This step-like feature in the oxygen profile is an unusual occurrence. Like other variables in the ocean, this feature must represent a balance between transport processes and removal mechanisms. Oxygen can be transported into this region by diffusive mixing from above, and by possible advection of new water/brine from the side. The change of oxygen with salinity in this zone (Table 7) is much lower than in either of the regions above. This may result from the low oxygen concentration, if the rate of

consumption is first order with respect to concentration. However, this is improbable. In other environments with low oxygen values, oxygen profiles decrease smoothly to zero without going through a step (see, for example, Richards 1975).

Nor does this step-like feature appear to be the result of input of new brine into the basin. The density of the high salinity brine is greater than the density of the water in the step region and would not remain on top of the interface as this feature does. A biological mechanism may explain the oxygen levels found there. In this region, the salinity increases from 50 to 70%. Just above this region (based on oxygen data), LaRock et al. (1979) observed a large increase in ATP and uridine uptake, with lower values below. It is possible that the microbial population which mediates consumption of oxygen in the ocean is inhibited by salinities greater than 50‰. If the microbes cannot metabolize oxygen effectively, the oxygen input due to diffusive mixing from above (see Chapter II) would be very slowly consumed and might remain in balance with oxygen consumption causing the observed feature.

Inhibition of bacterial activity in hypersaline waters exists in the Red Sea brines. However, most of this work has been done on the anaerobic bacteria. Trüper (1969) found that bacterial sulfate reduction is inhibited at high salinities, with significant decreases in microbial activity at 60‰ and greater. The

Orca Basin brine is not sterile (LaRock et al. 1979) like the hot metalliferous Red Sea brine (Watson and Waterbury 1969). A slowing of bacterial activity, however, combined with slow or periodic oxygen mixed into this region could produce the feature that was observed.

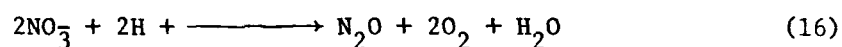
#### Argon and Nitrogen

The argon and nitrogen distributions in the Orca Basin can be divided into two zones of interest: the transition zone and the high salinity brine. In both of these zones, nitrogen gas concentrations are higher than would be predicted on the basis of solubilities. The nitrogen data in the high salinity brine show some random variability, with a relative standard deviation (RSD) of 12%. However, within the precision of the sampling and analytical techniques both nitrogen and argon levels (RSD = 8%) are considered constant in the high salinity brine (Fig. 26). On the other hand, there is an obvious trend in the nitrogen distribution in the transition zone.

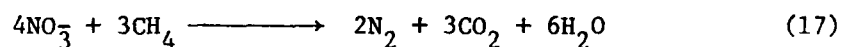
Transition Zone. Nitrogen values were higher in the transition zone than in either the brine, or the freely circulating Gulf water above. This increase in the nitrogen levels was found in the area where nitrate is being reduced to essentially zero. While the nitrate may not be the main source of excess nitrogen in the transition zone, denitrification processes must be the cause of

the excess nitrogen. The added nitrogen is calculated to be 140  $\mu\text{mol}\cdot\text{liter}^{-1}$ , which represents a supersaturation of 25%.

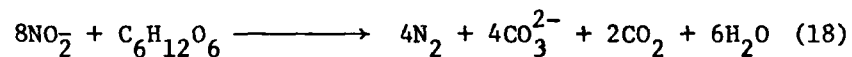
Denitrification is the biological reduction of nitrate or nitrite to nitrous oxide or free nitrogen. Denitrification can be carried on by a wide variety of facultative anaerobic bacteria (Brisou and Vargues 1961). The source for hydrogen in the reaction may come from either organic or inorganic substrates (e.g.,  $\text{H}_2$ ,  $\text{H}_2\text{S}$ ). Several feasible reactions at 25°C are given by Vaccaro (1965).



$$\Delta G^\circ = + 21 \text{ kcal.}$$



$$\Delta G^\circ = -475 \text{ kcal.}$$



$$\Delta G^\circ = -728 \text{ kcal.}$$

When an organic substrate supplies the hydrogen, the reactions are energetically feasible in aerobic waters. If the hydrogen comes from an inorganic source (Eq. 16), reaction is feasible only in the absence of oxygen.

The nitrate levels in the deep Gulf average 23  $\mu\text{mol}\cdot\text{liter}^{-1}$ . Reduction of 23  $\mu\text{mol}$  of nitrate would produce only 11.5  $\mu\text{mol}$  of

molecular nitrogen. Thus nitrate reduction alone cannot account for the nitrogen increase of  $140 \mu\text{mol}\cdot\text{liter}^{-1}$  that was observed in the transition zone. Another source of nitrogen substrate is needed to provide the starting material for nitrogen gas production. Two possible sources are ammonia produced from organic material decomposition at the brine interface or ammonia diffusing up from the brine. Ammonia levels in the brine (see Chapter IV) are about  $500 \mu\text{mol}\cdot\text{liter}^{-1}$ , but diffusion from the brine would be very small. Possible mechanisms of nitrogen gas formation are shown in Fig. 27.

Ammonia diffusing out of the brine or produced in the oxygen step region could be oxidized to nitrate and/or nitrite. These nutrients could be subsequently converted to nitrogen in this low oxygen environment. The biological oxidation of ammonia to nitrite then nitrate in sea water has been described in the classic laboratory experiments of von Brand and his co-workers (cited in Vaccaro 1965). The oxidation process is mediated by various nitrifying bacteria including Nitrosomonas and Nitrobacter (Zobell 1946) and Nitrosocystis (Murray and Watson 1963). These processes are well documented in the literature.

There is no evidence, however, for the direct conversion of ammonia to molecular nitrogen. While ammonia oxidation during denitrification is thermodynamically possible (Cline and Richards 1972), there is little, if any, experimental evidence for the direct biological oxidation of ammonia to free nitrogen. In the low oxygen waters of the eastern tropical Pacific, Goering (1968) detected no oxidation of ammonia during studies of natural

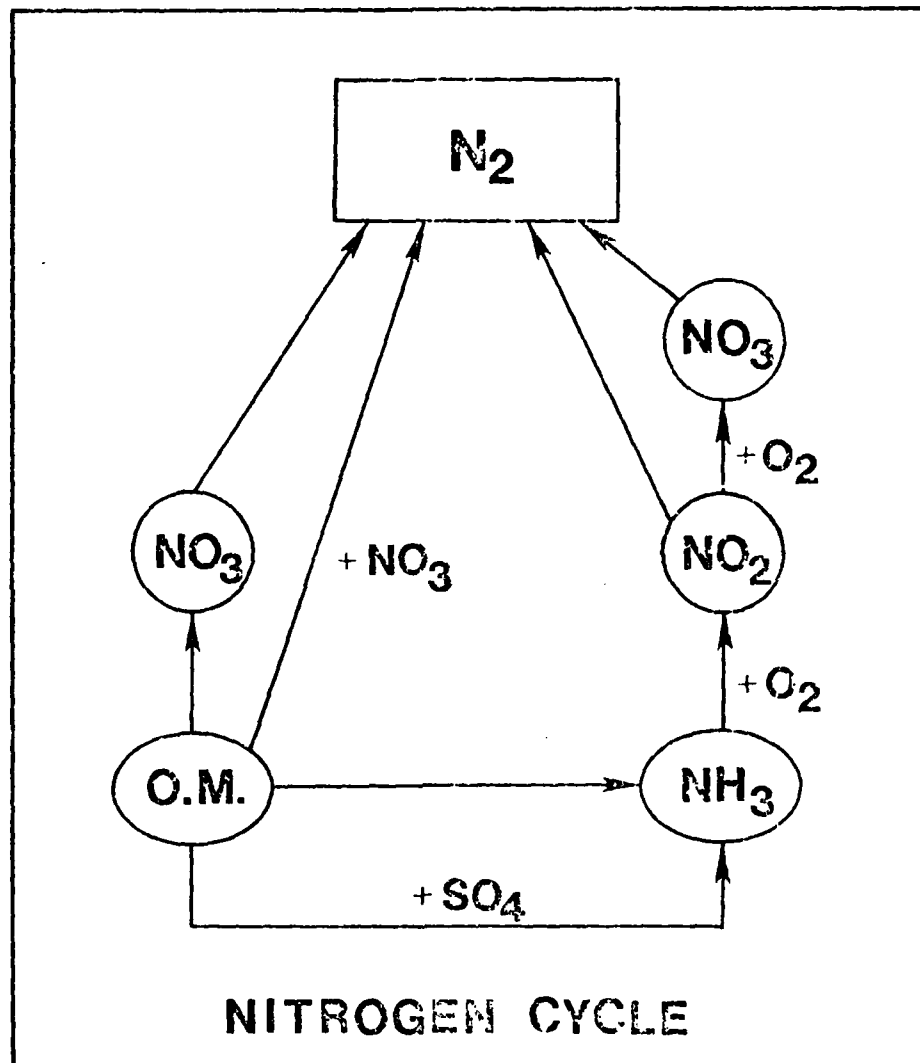
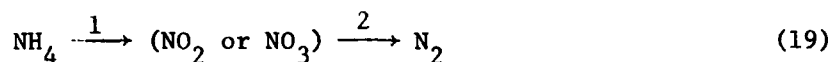


Fig. 27. A schematic representation of the possible pathways through which organic nitrogen or ammonia may be converted to molecular nitrogen. All of these pathways are thermodynamically possible, but not all under the same conditions.



microbial populations. Cline and Richards (1972) also reported no apparent oxidation of ammonia in their study of nitrate reduction in the same region. The low ammonia concentrations observed in the Pacific were attributed to removal of ammonia during bacterial assimilation, not nitrification to molecular  $N_2$ .

For production of molecular nitrogen from ammonia, a two-step process must exist:

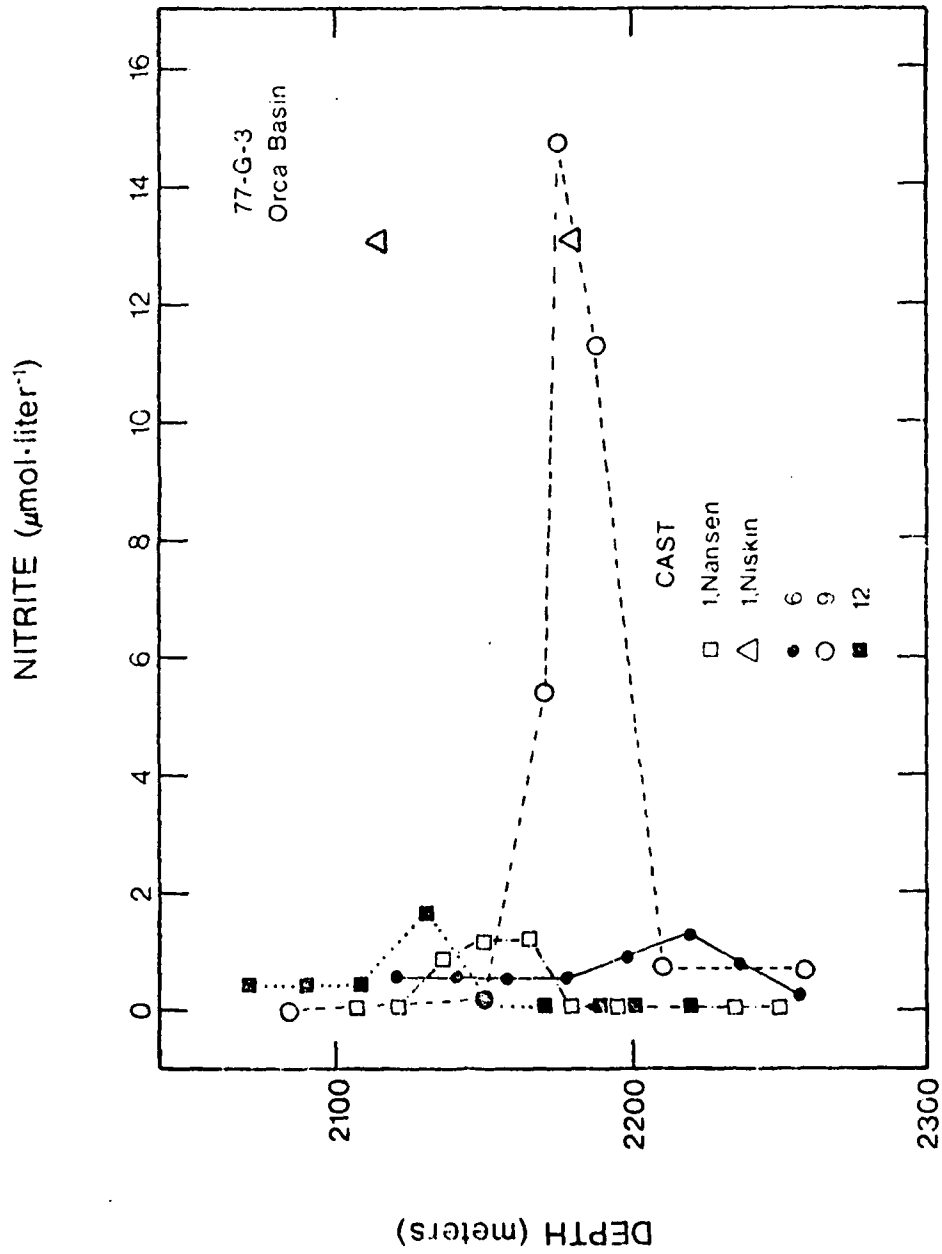


Oxygen concentration in the oxygen-step region of the Orca Basin (zone T3) averages  $10 \mu\text{mol}\cdot\text{liter}^{-1}$ . Carlucci and McNally (1969) have demonstrated that oxidation of ammonia to nitrite can occur under conditions of low oxygen tension, down to  $2 \mu\text{mol}\cdot\text{liter}^{-1}$ . Such conditions prevail in the Orca Basin interface region. In the eastern tropical Pacific, Cline and Richards (1972) observed the simultaneous occurrence of nitrite, along with the apparent loss of nitrate and suggested that nitrite is an intermediate in the reduction of nitrate to free nitrogen. Since nitrite does not accumulate in the Orca Basin from ammonia oxidation, it appears that the reduction of nitrate to nitrite and of nitrite to free nitrogen occur simultaneously (i.e., as coupled reactions (Eq. 19) which may be carried on by the same or different organisms).

Nitrate or nitrite may be the major intermediate in the production of nitrogen. Wiesenburg et al. (1977) noted the presence of high nitrite concentrations (up to  $15 \mu\text{mol}\cdot\text{liter}^{-1}$ ) in the

interface region (transition zone) of the Orca Basin. Their data (Fig. 28) indicated that the nitrite maximum was found well above the region of lowest oxygen values. The nitrite maximum may have resulted from nitrate reduction. A nitrite maximum above the brine was not observed on subsequent cruises, however. This maximum may have been a transient feature, with the nitrite being subsequently converted to molecular nitrogen as in Eq. 18.

A continuing supply of dissolved oxygen could be provided by mixing from above. This oxygen could be used in the oxidation of ammonia. The nitrate or nitrite so produced would in turn be reduced to  $N_2$  as in Eqs. 16-18. These conclusions are valid for the Orca Basin brine-sea water interface. They are also the same conclusion Barnes et al. (1975) suggested for the Santa Barbara Basin sediments. Barnes et al. (1975) examined nitrification and denitrification in suboxic sediments of the Santa Barbara Basin. Increases in both dissolved molecular nitrogen and the  $N_2/Ar$  ratio were observed in the sediment pore waters. Nitrogen increases (up to 17% above bottom water values) could not be accounted for by nitrite (or nitrate) reduction alone. It was concluded that ammonia, derived from organic matter containing nitrogen, was converted to molecular  $N_2$  through a nitrite intermediate under conditions of low oxygen tension. Since methane is also present in the sediments of this basin (Emery and Hogan 1958), the sediment-sea water interface geochemistry in the Santa Barbara



Basin may be similar to the brine-sea water interface geochemistry in the Orca Basin. In both areas, ammonia is being converted to molecular nitrogen via a nitrate or nitrite intermediate.

High Salinity Brine. Both argon and nitrogen are supersaturated in the high salinity Orca brine. The excess gas, relative to atmospheric solubilities (Table 9), must arrive with the entry brine into the basin, or be produced in the brine after deposition. Argon has no known biological source. Production of argon by radioactive decay of  $^{40}\text{K}$ , however, does occur. Craig (1969) attributed the 10% excess argon in the Discovery Brine of the Red Sea (relative to the Atlantis II brine) to excess radiogenic  $^{40}\text{Ar}$ . This concept is consistent with a model of the Discovery Brine as a stagnant overflow from the Atlantis Deep, as discussed by Weiss (1969). In the Orca Basin brine, however, radiogenic  $^{40}\text{Ar}$  is probably not an important constituent of the total argon observed. It can be calculated (Table 10) that radiogenic argon production would only be  $9 \times 10^{-7} \mu\text{mol}\cdot\text{kg}^{-1}$  during the 7900 year age of the brine in the Orca Basin. Even if the excess argon came from the Jurassic LouAnn salt (an unlikely case) an increase of only  $2 \mu\text{mol}\cdot\text{kg}^{-1}$  would be predicted. The observed excess argon is over  $10 \mu\text{mol}\cdot\text{kg}^{-1}$  (Fig. 26). Unlike the Red Sea brines, argon production is not important in the Orca Basin.

Molecular nitrogen in the Orca Basin remains relatively constant, below the zone where oxygen is present. There is abundant

Table 10. Calculated theoretical production of dissolved argon in the high salinity Orca Basin brine.

---

Assume the brine potassium content has been constant over time at the present concentration ( $0.63 \text{ g} \cdot \text{kg}^{-1}$ ) and that A is produced by radioactive decay of  $^{40}\text{K}$ .

K content of brine:

$$= 0.63 \text{ gK} = \text{kg}^{-1} = 16.1 \text{ mmol} \cdot \text{K} = \text{kg}^{-1} \text{ brine}$$

$$= 9.7 \times 10^{21} \text{ atoms K} \cdot \text{kg}^{-1} \text{ brine}$$

Atoms  $^{40}\text{K}$ : (Atomic abundance = 0.0118%)

$$= 1.1 \times 10^{18} \text{ atoms K} \cdot \text{kg}^{-1} \text{ brine}$$

Atoms  $^{40}\text{K}$  decaying to Ar: (Branching ratio = 0.11)

$$= 1.3 \times 10^{14} \text{ atoms} \cdot \text{kg}^{-1} \text{ brine}$$

Growth of Ar: ( $t = 7900\text{y}$ , decay const =  $\lambda = 5.4 \times 10^{-10} \text{ y}^{-1}$ )

$$\text{Ar} = ^{40}\text{K} (1 - e^{-\lambda t}) = 5.4 \times 10^{11} \text{ atoms Ar} \cdot \text{kg}^{-1} \text{ brine}$$

$$= 9.0 \times 10^{-7} \text{ } \mu\text{mol Ar} \cdot \text{kg}^{-1} \text{ brine}$$


---

ammonia in the high salinity brine (see next Chapter), but there is no apparent nitrogen gas production in this zone. Even though  $N_2$  is the thermodynamically stable species in sulfate reducing environments, a biochemical pathway for the direct conversion of ammonia to dissolved molecular nitrogen must not exist. Barnes et al. (1975) also came to this conclusion for the anoxic zone of the Santa Barbara Basin sediments. They observed near-uniform  $N_2$  distribution in the anoxic zone and noted that  $N_2$  production would increase the  $N_2/Ar$  ratio. In the Orca Basin, the  $N_2/Ar$  ratio in the brine was 36.4 (average of 25 points) while the predicted solubility values for  $N_2$  and Ar at 250‰ salinity and 5.65°C gave a  $N_2/Ar$  ratio of 36.5. There was no  $N_2/Ar$  increase.

Neither nitrogen or argon is being produced in the high salinity bottom water of the Orca Basin, yet elevated concentrations (relative to solubility) of both gases are observed. One can show that the observed nitrogen was present when the brine formed. The sea water which dissolved the salt exposure to form the brine must be deep Gulf water like that at the basin sill ( $T = 4.2^\circ C$ ,  $S = 35\text{‰}$ ). This deep Gulf water probably originates from the Upper North Atlantic Deep Water (UNADW), with a very small component of Mediterranean Sea water (J. Cochrane, personal communication). Assuming (the extreme) that the UNADW ( $2^\circ C$ ) is the initial component in the mixture which produces the Orca Basin sill water, it would have to be mixed equally with water at  $6.4^\circ C$

(and 35‰ salinity) to reach the values observed. Since mixing two waters of different temperatures can cause an apparent supersaturation (Fig. 19), this mixing effect was investigated as a possible cause of the supersaturation of  $N_2$  and Ar. Such a mixture, however, would only cause a supersaturation anomaly of 0.1% for Ar and 0.2% for  $N_2$ , far below the observed excesses. Having discounted all other possibilities, we are able to conclude that the Orca Basin brine is simply salt-saturated, deep-Gulf sea water with the  $N_2$  and Ar gases intact. This conclusion is substantiated by the following discussion.

Argon concentration is plotted against nitrogen concentration in Fig. 29. All concentrations in the plot are expressed as  $\mu\text{mol}$  per 1000 g  $H_2O$ , and are thus not affected by the dissolving or precipitation of salt. The observed concentrations for the Orca Basin were converted to these units by converting from  $\mu\text{mol}\cdot\text{liter}^{-1}$  to  $\mu\text{mol}\cdot\text{kg}^{-1}$  using the density equations of Millero et al. (1979), and to  $\mu\text{mol}$  per 1000g water using the value of 741.9 grams of  $H_2O$  per kg of brine. The Red Sea brine data, given for comparison, are from Weiss (1969).

Solubility values for fresh water and 35‰ sea water (Orca Basin sill water, OBSW), in equilibrium with air at one atmosphere total pressure (including vapor pressure) are also plotted in Fig. 29. These curves (in units of  $\mu\text{mol}\cdot\text{kg}^{-1}$ ) were calculated using Eq. 5 and the coefficients given by Weiss (1970), and were

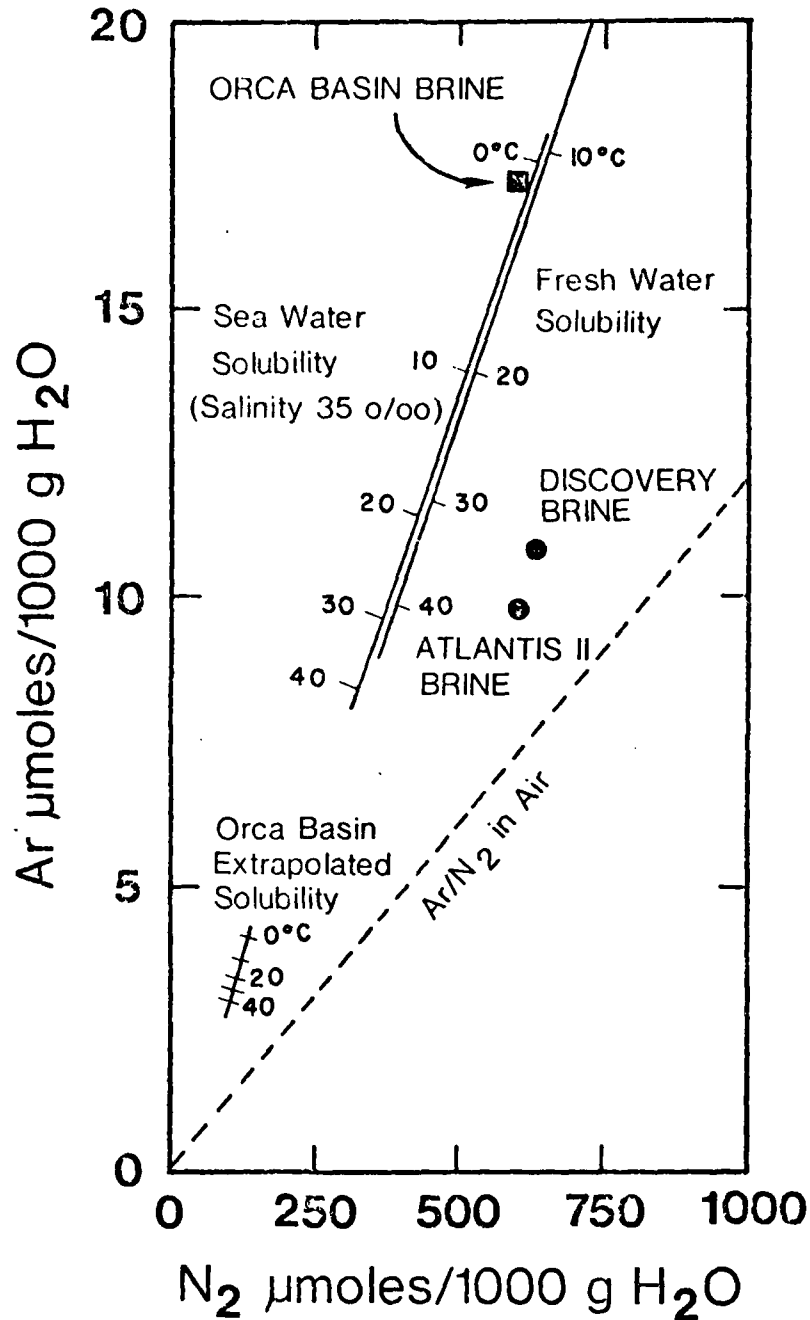


Fig. 29. Argon versus nitrogen in the Orca Basin brine and solubility values for fresh water, 35‰ sea water and Orca Basin brine in equilibrium with the atmosphere. Data for argon and nitrogen (Weiss 1969) from the Red Sea brines are given for comparison.



corrected using a value of 965.0 g water per 1000 grams of sea water. Using Eq. 5,  $N_2$  and Ar solubilities were extrapolated to the salinity of the Orca Basin brine. As was previously discussed, these extrapolated values are a few percent higher than the observed atmospheric solubilities in the brine (Table 7), but are close enough for the comparisons presented here. The Ar/ $N_2$  ratio in air is also plotted in Fig. 29.

Fig. 29 indicates that the observed  $N_2$  and Ar concentrations in the Orca Basin are supersaturated approximately 250% with respect to brine in equilibrium with air at 5.6°C. Although this difference could be produced by addition of radiogenic argon and large amounts of air, the existence of such a natural process is extremely improbable. Contamination can also be excluded. Bulk sample loss would move the brine  $N_2$ /Ar value in Fig. 29 along a straight line toward the origin, while air addition would move the point away from the origin along a line with a slope equal to the Ar/ $N_2$  ratio in air. Neither case is observed here.

The large saturation anomalies in Ar and  $N_2$  can be best explained, not by gas production, but rather by the simple addition of salt to deep Gulf water - the mechanisms proposed by Shokes et al. (1972) for Orca brine formation. Comparison data for the Orca Basin - Gulf water and Red Sea brine - Red Sea water systems are given in Table 11. Weiss (1969) and Craig (1969) explained the  $N_2$  and Ar saturation anomalies in the Red Sea brine on  $N_2$

Table 11. Dissolved gases in Red Sea water, Red Sea brines, Gulf of Mexico Deep Water and Orca Basin brine. Gas concentrations are expressed as  $\mu\text{mol (STP)}/\text{kg}$  of 38.2‰ salinity (Red Sea) or 35.0‰ (Gulf of Mexico) calculated from data of Weiss (1969).  $\Delta$  values are excess concentrations relative to sea water,  $\delta$  values are relative to the PDB standard.  $\delta^{13}\text{C}$  ( $\Delta\text{CO}_2$ ) is the calculated  $\delta^{13}\text{C}$  of carbon dioxide that would have to be added to sea water to yield the observed  $\delta^{13}\text{C}$  of  $\Sigma\text{CO}_2$  in the brines.

	Red Sea		Gulf of Mexico		Orca Basin Brine
	Water S = 38.2 ‰ T = 28°C	Atlantis II Brine	Discovery Brine	Sea Water S = 35.0 ‰ T = 4.2°C	
$\text{N}_2$ $\mu\text{mol Kg}$	359	584	602	558	632
$\Delta\text{N}_2$	-	225	243	-	74
Ar	9.41	9.41	10.35	15.3	17.7
$\Delta\text{Ar}$	-	0.000	0.936	-	2.4
$\Sigma\text{CO}_2$ $\text{mmol/L}^{-1}$	(2.84)	4.20	0.90	2.28	4.57
$\Delta\text{CO}_2$	-	1.36	-1.94	-	2.28
$\delta^{13}\text{C}$ (‰)	(-1)	-5.6	-16.8	0.3	-16.4
$\delta^{13}\text{C}$ ( $\Delta\text{CO}_2$ ) (‰)	-	-15.2	+6.3	-	-32.2

production from organic matter in both brines and radiogenic Ar production for the Discovery Brine only. Considering the above discussion, these mechanisms are not operative in the Orca Basin brine. Both average  $N_2$  and Ar values are slightly higher than the predicted values (Table 10), but the uniform differences (13%) in each indicate a physical rather than a biochemical or radiogenic effect. This difference may be due to some of the assumptions (e.g. initial equilibrium) made in calculating the theoretical values. If error limits for  $N_2$  and Ar are taken as twice the standard deviation of the determinations, then the uncertainty envelope for the Orca Basin, Ar/ $N_2$  point on Fig. 29 encompasses 4.2°C and 35‰ on the sea water solubility curve. Thus, all the Ar and  $N_2$  in the Orca Basin high salinity brine can be accounted for by the addition of salt to deep Gulf water.

Shokes et al. (1977) surmised that the brine in the bottom 140 meters of the Orca Basin resulted from the underwater dissolution of a surface or near-surface salt deposit on the basin slope. This exposed diapir must be dissolving due to contact with deep Gulf water and the resultant brine flows down the slope into the bottom of the basin. The previous evidence for this mechanism of formation of the Orca Basin brine has been:

1. The decreasing salinity gradient in the sediments.
2. The relative ion abundances of potassium and calcium

relative to sodium, which are unlike the Red Sea brines and indicate a low temperature halite dissolution.

3. The bromine content of the Orca Basin water, which indicates that the brine is formed by the dissolution of an evaporite deposit rather than by an evaporative process.
4. The seismic data of Addy and Behrens (1980) which indicate that exposed salt diapirs do exist in the vicinity of the Orca Basin and support the earlier contentions of Shokes et al. (1977) concerning the location of the source salt.

The other piece of information which now emphasizes the origin of the brine is the nitrogen-argon relationships determined as part of this study. Dissolved argon and nitrogen data provide independent evidence that the Orca brine did form due to low temperature dissolution of a salt deposit by sea water. Since the gas data is not based on the major ions, which have been used in the other evaluation techniques, the conclusions based on these data are independent of the ionic balance percentages in the salt that was dissolved. The only assumption that was made, in using the gas data to make this evaluation, was that the sea water that participated in the dissolution of the salt diapir had an original salinity of 35‰. The temperature at which the salt water was

saturated with brine is estimated from Fig. 29 to be between 0 and 5°C and further supports the low temperature dissolution assumption of Shokes et al. (1977).

CHAPTER IV  
DISSOLVED REDUCED GASES

Organic matter is thermodynamically unstable in sea water and eventually decomposes into various small molecules in the C-N-S-H<sub>2</sub>O system. Decomposition of organic material in anoxic basins and sediments has been extensively studied (see for example, Richards 1965). These studies are important to an understanding of marine geochemistry since such environments enable us to determine the mechanisms by which organic material is recycled from the sediments back into the water column.

The most widely accepted model of organic matter decomposition is related to the free energy of the oxidant involved in the diagenesis process. Organic matter [assumed to have an idealized composition of  $(\text{CH}_2\text{O})_{106}(\text{NH}_3)_{16}(\text{H}_3\text{PO}_4)_1$  (Fleming 1940; Redfield 1958)] is oxidized by the oxidant having the greatest free energy yield per mole of organic carbon oxidized (McKinney and Conway 1957; Richards 1965; Claypool and Kaplan 1974). When an oxidant is depleted, the oxidant with the next highest energy yield is consumed until every oxidant is removed, or until all organic carbon has been depleted. Table 12 (Froelich et al. 1979) summarizes the predicted sequences of reactions. In anoxic marine environments, the decomposition reactions are mediated by denitrifying, iron oxide reducing, sulfate reducing and methanogenic bacteria

Table 12. Oxidation reactions of sedimentary organic matter.\*

- 
1.  $(\text{CH}_2\text{O})_{106}(\text{NH}_3)_{16}(\text{H}_3\text{PO}_4) + 138 \text{ O}_2 \longrightarrow 106 \text{ CO}_2 + 16 \text{ HNO}_3 + \text{H}_3\text{PO}_4 + 122 \text{ H}_2\text{O}$   
 $\Delta G^{\circ'} = -3190 \text{ kJ/mole of Glucose}$
  2.  $(\text{CH}_2\text{O})_{106}(\text{NH}_3)_{16}(\text{H}_3\text{PO}_4) + 236 \text{ MnO}_2 + 472 \text{ H}^+ \longrightarrow 236 \text{ Mn}^{2+} + 106 \text{ CO}_2$   
 $+ 8 \text{ N}_2 + \text{H}_3\text{PO}_4 + 366 \text{ H}_2\text{O}$   
 $\Delta G^{\circ'} = -3090 \text{ kJ/mole (BIRNESSITE)}$   
 $-3050 \text{ kJ/mole (NSUTITE)}$   
 $-2920 \text{ kJ/mole (PYROLUSITE)}$
  - 3a.  $(\text{CH}_2\text{O})_{106}(\text{NH}_3)_{16}(\text{H}_3\text{PO}_4) + 94.4 \text{ HNO}_3 \longrightarrow 106 \text{ CO}_2 + 55.2 \text{ N}_2 + \text{H}_3\text{PO}_4$   
 $+ 177.2 \text{ H}_2\text{O}$   
 $\Delta G^{\circ'} = -3030 \text{ kJ/mole}$
  - 3b.  $(\text{CH}_2\text{O})_{106}(\text{NH}_3)_{16}(\text{H}_3\text{PO}_4) + 84.8 \text{ HNO}_3 \longrightarrow 106 \text{ CO}_2 + 42.4 \text{ N}_2 + 16 \text{ NH}_3$   
 $+ \text{H}_3\text{PO}_4 + 148.4 \text{ H}_2\text{O}$   
 $\Delta G^{\circ'} = -2740 \text{ kJ/mole}$
  4.  $(\text{CH}_2\text{O})_{106}(\text{NH}_3)_{16}(\text{H}_3\text{PO}_4) + 212 \text{ Fe}_2\text{O}_3 \text{ (or } 424 \text{ FeOOH)} + 848 \text{ H}^+ \longrightarrow$   
 $424 \text{ Fe}^{2+} + 106 \text{ CO}_2 + 16 \text{ NH}_3 + \text{H}_3\text{PO}_4 + 530 \text{ H}_2\text{O (or } 742 \text{ H}_2)$   
 $\Delta G^{\circ'} = -1410 \text{ kJ/mole (HEMATITE, Fe}_2\text{O}_3)$   
 $- 1330 \text{ kJ/mole (LIMONITIC GOETHITE, FeOOH)}$
  5.  $(\text{CH}_2\text{O})_{106}(\text{NH}_3)_{16}(\text{H}_3\text{PO}_4) + 53 \text{ SO}_4^{2-} \longrightarrow 106 \text{ CO}_2 + 16 \text{ NH}_3 + 53 \text{ S}^{2-} + \text{H}_3\text{PO}_4$   
 $+ 106 \text{ H}_2\text{O}$   
 $\Delta G^{\circ'} = -380 \text{ kJ/mole}$
  6.  $(\text{CH}_2\text{O})_{106}(\text{NH}_3)_{16}(\text{H}_3\text{PO}_4) \longrightarrow 53 \text{ CO}_2 + 53 \text{ CH}_4 + 16 \text{ NH}_3 + \text{H}_3\text{PO}_4$   
 $\Delta G^{\circ'} = -350 \text{ kJ/mole}$
- 

\* See Froelich et al. (1979) for an explanation of how free energies were calculated.

(Reactions 3-6), resulting in the production of  $N_2$ ,  $NH_3$ ,  $HS^-$ ,  $CO_2$ ,  $HCO_3^-$ ,  $CH_4$ , and other related compounds. Interstitial waters from the upper meter of anoxic marine sediments often represent the early stages of the diagenetic process (Thorstenson and MacKenzie 1974; Froelich et al. 1979), while deeper sediments usually exhibit complete sulfate reduction and large increases in methane and carbon dioxide (Hammond 1974).

Anoxic waters overlying reduced sediments have chemical distributions which reflect the oxic and suboxic processes (Reactions 1-4, Table 12) taking place at their oxic-anoxic sea water boundary, as well as the sulfate-reduction-catalyzed reactions taking place in the water and sediments. The combined influence of these decomposition processes in anoxic marine waters has been described for the various nutrients (Richards 1965; Richards et al. 1965; Fanning and Pilson 1972). Only a few studies, however, (Atkinson and Richards 1967; Linnenbom and Swinnerton 1969; Reeburgh 1976; Cohen 1978) have considered the role of various reduced gases in anoxic waters.

In many anoxic areas (e.g., Cariaco Trench) the stability of the water column is relatively low (see Table 5), and periodic inputs of oxygenated water replace some of the anoxic waters. In these other basins, the oxygenated, renewal water both dilutes and oxidizes the reduced products (nutrients and gases) of anaerobic decomposition (see Table 12). In the Orca Basin,



however, the great stability of the hypersaline water (~250 ‰) precludes input of any significant amount of renewal water. With no input of renewal water, and with little (other than diffusive) loss of decomposition products from the brine, the Orca Basin is an ideal location for studying the processes on anaerobic decomposition.

Shokes et al. (1977) have described nutrient distributions resulting from the anoxic conditions in the basin. Phosphate levels increased from  $2.5 \mu\text{mol}\cdot\text{liter}^{-1}$  above the brine to between 60 and  $80 \mu\text{mol}\cdot\text{liter}^{-1}$  near the bottom, with no obvious trends in the depth profile within the brine. Silicate likewise increased from  $25 \mu\text{mol}\cdot\text{liter}^{-1}$  to over  $235 \mu\text{mol}\cdot\text{liter}^{-1}$ , while nitrate decreased from  $23 \mu\text{mol}\cdot\text{liter}^{-1}$  above the brine to zero level throughout the brine. This chapter presents a detailed description of the geochemistry of reduced gases ( $\text{CH}_4$ ,  $\text{C}_2\text{H}_6$ ,  $\text{C}_3\text{H}_8$ ,  $\text{N}_2\text{O}$ ,  $\text{NH}_3$  and  $\text{H}_2\text{S}$ ) in the waters of the Orca Basin, and describes the distributions which have resulted from the anoxic, hypersaline conditions.

## Methods

### Sampling

Observations of reduced gases were made on five cruises aboard the R/V Gyre during October 1976 (76-G-10, April 1977 (77-G-2 and 77-G-3), November 1977 (77-G-13), and one aboard the R/V

Endeavor in February 1979. Stations were taken in both the northern and southern lobes of the Orca Basin and samples were collected both above and in the brine between 2000 m and 2500 m. Most water samples for reduced gas analysis were taken with standard 1.2-liter teflon-coated Nansen bottles. As was previously discussed, sampling with 5-liter Niskin bottles proved ineffective since gases in the Orca Basin brine are supersaturated at atmospheric pressure and degas as hydrostatic pressure is reduced. Since Nansen bottles have a ball valve-type closing apparatus these bottles seemed to inhibit degassing and gave representative results for most gas measurements in the brine.

Sampling from the Nansen bottles was performed initially by transferring the brine to glass bottles (cruise 76-G-10) for later analysis of the reduced gases. This technique gave low values, due to the effervescence which occurred during gravity transfer of brine from the Nansen bottles. During subsequent cruises, brine samples were drawn directly from the Nansen bottles into helium purged glass syringes which were used either to inject samples directly into an all-glass stripping chamber (Swinnerton et al. 1962) or to equilibrate the water with helium (McAuliffe 1971). Samples for methane analysis were collected in piggyback samplers on cruises 77-G-13 and 78-G-2, to compare the Nansen samples with the pressurized samples. On cruises EN-32, samples for methane analysis were also collected in the all glass sampling containers shown in Fig. 21.

### Analytical Techniques

The data for this chapter on reduced gases in the Orca Basin brine include measurements of oxygen, nitrous oxide, hydrogen sulfide, ammonia, methane, ethane, and propane as well as determinations of temperature, salinity, iron, sulfate, nitrate, and phosphate. A complete set of these values was needed to make a comprehensive evaluation of anaerobic decomposition processes outlined in Table 12, but it was impossible for one researcher to make all of these measurements simultaneously on every cruise to the Orca Basin. Table 13 gives a description of the various parameters measured on each of the six cruises as well as the analyst who was responsible for providing the data for this comprehensive interpretation. A list of analytical techniques that were used is given in Table 14.

Briefly, temperature was measured using standard reversing thermometers attached to Nansen bottles. As previously mentioned, salinity was obtained by performing either weight or volume dilutions (density corrected) to near 35‰, followed by analysis using a Plessey Environmental Systems 6230N inductive salinometer which was standardized against standard sea water. Salinities measured by this method are only relative since the Orca Basin brine is not just a concentrated sea water with ion ratios equivalent to those of normal sea water.

Dissolved oxygen was measured by a modification (Broenkow and

Table 13. Analyses of Orca Basin water performed by various chemists on samples from different cruises aboard the R/V Gyre and aboard cruise EN32 of the R/V Endeavor. A dash indicates that measurements were not made on that cruise.

Cruise	Temp.	Salinity	O <sub>2</sub>	PO <sub>4</sub>	NO <sub>3</sub>	NH <sub>3</sub>	H <sub>2</sub> S	Fe	SO <sub>4</sub>	N <sub>2</sub> O	CH <sub>4</sub>	C <sub>2</sub> H <sub>6</sub>	C <sub>3</sub> H <sub>8</sub>
76-G-10	T*	T	T	T	T	-	-	-	-	-	B	B	B
77-G-2	T	T	T	T	T	B	-	P	P	-	B	B	B
77-G-3	T	T	T	T	T	W	W	-	-	-	W	-	-
77-G-13	T	T	T	T	T	B	W	-	-	B	W	-	-
78-G-2	T	T	T	T	T	-	-	-	-	-	W	-	-
EN-32	W	W	-	-	-	-	W	-	-	-	W	-	-

\* Key for analysts is as follows: Texas A&M Technical Operations (T), James M. Brooks and Bernie B. Bernard (B), Ronald Pflaum (P), Denis A. Wiesenburg (W) all of Texas A&M University.

Table 14. Method of analysis used for Orca Basin samples.

Component	Method Used	Reference
Salinity	Conductive salinometer	
Sulfate	Gravimetric as BaSO <sub>4</sub>	Vogel (1968)
Dissolved Oxygen	MicroWinkler titration	Carpenter (1965)
Hydrogen Sulfide	Mixed diamine method	Cline (1969)
Iron	Ferrozine colorometric complexation	Stookey (1970)
Phosphate	Photometric as molybdenum blue	Murphy & Riley (1962)
Nitrate	Reduction to nitrite, photometric	Amstrong et al. (1967)
Nitrite	Diazo complexation, photometric	
Ammonia -1	Phenol hypochlorite method	Solorzano (1969)
-2	Phenate colorometric	Weatherburn (1967)
-3	Electrode referenced to NH <sub>4</sub> Cl	
Nitrous Oxide	Trapping and electron capture GC	Cohen (1978)
Methane	Gas stripping and thermal conductivity GC	Swinerton et al. (1962)
Hydrocarbons	Trapping and flame ionization gas chromatography	Brooks et al. (1980)

Cline 1969) of the microWinkler method of Carpenter (1965) as was described in the previous chapter. Nutrients were measured colorometrically by AutoAnalyzer using dilutions to bring the concentration of salts down to a level where they did not inhibit the relative responses to the nutrients alone. Dilution was also necessary due to the high nutrient concentrations observed. Inorganic phosphate was determined by the ascorbic acid-potassium antimonyl tartrate method of Murphy and Riley (1962) as modified for use on the AutoAnalyzer by Atlas et al. (1971). Nitrate ions were determined by the cadmium reduction method of Armstrong et al. (1967).

Ammonia was measured by three different techniques. On cruise 77-G-2, ammonia was measured using an Orion Research Model 95-10 specific ion electrode standardized against  $\text{NH}_4\text{Cl}$ -ammonia solutions. This probe had been used previously for studies of interstitial waters and had proven to be very effective for samples of high concentration. There seemed to be very little, if any, salt effect associated with this probe at the high  $\text{NH}_3$  levels found in the Orca brine. On cruise 77-G-3, ammonia was determined manually using the phenol hypochlorite method of Solorzano (1969). Samples for ammonia were taken in syringes or pipettes filled directly from the Nansen bottles and introduced into 50 ml serum vials below the surface of the complexing reagents. Ammonia samples collected in this way seem to be little affected by the

degassing which caused inaccuracies with other reduced gas measurements. On cruise 77-G-13, ammonia was determined by AutoAnalyzer using the phenate method of Weatherburn (1967) modified for use on the AutoAnalyzer II. Samples for AutoAnalyzer analysis were transferred into sample cups before analysis. There was a possibility of degassing in these samples since they remain open to the air while they await analysis. However, no apparent loss of ammonia was observed after sample collection, as evidenced by the consistency of the three methods.

Hydrogen sulfide was determined using the mixed diamine reagent method of Cline (1969). Samples were drawn from Nansen bottles directly into pipettes and the brine sample was introduced into the acidified reagent under the liquid surface, thus avoiding any contact with air which would rapidly oxidize free sulfide. Sulfate was measured gravimetrically using a standard barium precipitate method described by Vogel (1968). Dissolved iron in the water column was also measured using the ferrozine method of Stookey (1970).

Nitrous oxide was determined on cruise 77-G-13 by a technique similar to the one described by Cohen (1978) with modifications: an Ascarite column was placed after the magnesium perchlorate column to remove  $\text{CO}_2$ . An electron capture ( $^{63}\text{Ni}$ ) detector operated at  $300^\circ\text{C}$ , was used in conjunction with a Hewlett-Packard 5830A gas chromatograph for sample analysis, after separation on

a Porapak Q column. Calibration was accomplished by comparing the measured values against a standard calibration mixture.

Methane in the brine was determined by gas chromatography using both thermal conductivity and flame ionization detectors. During cruise 76-G-10, samples were collected in 300 ml sample bottles and returned to the lab for analysis by the method of Brooks et al. (1980). Some gas was lost from these samples due to degassing, bubble formation and escape during transfer to the sample bottle. To prevent degassing on samples taken on cruise 77-G-2, the brine was drawn directly into 50 ml equilibration syringes as previously described. Gaseous hydrocarbons were determined using the techniques of McAuliffe (1971). Methane on samples taken on cruises 77-G-3 and 77-G-13 was analyzed in water transferred from the Nansen bottle directly via nylon tubing into 12 ml syringes. The water in these syringes was injected through a septum into a 20 ml stripping chamber. Methane was stripped by the carrier-gas helium flow, and swept directly onto a 5A molecular sieve column, for separation from dissolved atmospheric gases. Methane was measured with a Hewlett-Packard 5710A gas chromatograph using a thermal conductivity detector. Sample transfer and chromatographic analysis techniques for methane from the piggyback and glass-container samplers used on cruises 78-G-2 and EN-32 were the same as were used for  $N_2$  and Ar on those cruises (see Chapter III). Gaseous hydrocarbons, ethane, and propane were determined on



samples from cruises 76-G-10 and 77-G-2 using the technique described for methane (Brooks et al. (1980)). The  $^{13}\text{C}/^{12}\text{C}$  isotopic ratio of the methane was determined by B. B. Bernard by degassing the brine, removing  $\text{CO}_2$  and combusting methane to carbon dioxide at  $800^\circ\text{C}$ .  $\delta^{13}\text{C}$  was determined using a Nuclide mass spectrometer and referenced to the PDB standard (Sackett et al. 1970).

$$\delta^{13}\text{C} = \left[ \frac{(^{13}\text{C}/^{12}\text{C}) \text{ sample}}{(^{13}\text{C}/^{12}\text{C}) \text{ standard}} - 1 \right] \times 1000 \quad (20)$$

## Results

### Gaseous Hydrocarbons

Fig. 30 shows salinity, temperature, dissolved oxygen, methane, ethane, propane, and  $\delta^{13}\text{CH}_4$  profiles from three stations in the Orca Basin for cruise 76-G-10. Results are plotted as distance above the bottom. The bottom depth for the stations plotted here was 2436 m. These data were taken from the same hydrocasts that Shokes et al. (1977) used to characterize the Orca Basin brine. Methane in the brine from cruise 77-G-3 is also plotted (closed circles). Samples which were collected in the brine exhibited effervescence during gravity transferal from the Nansen bottles. Thus, these values of approximately  $580 \mu\text{mol}\cdot\text{liter}^{-1}$  of methane in the brine are lower than the  $750 \mu\text{mol}\cdot\text{liter}^{-1}$  levels measured on cruise 77-G-2 (Table 15), when samples were drawn directly from

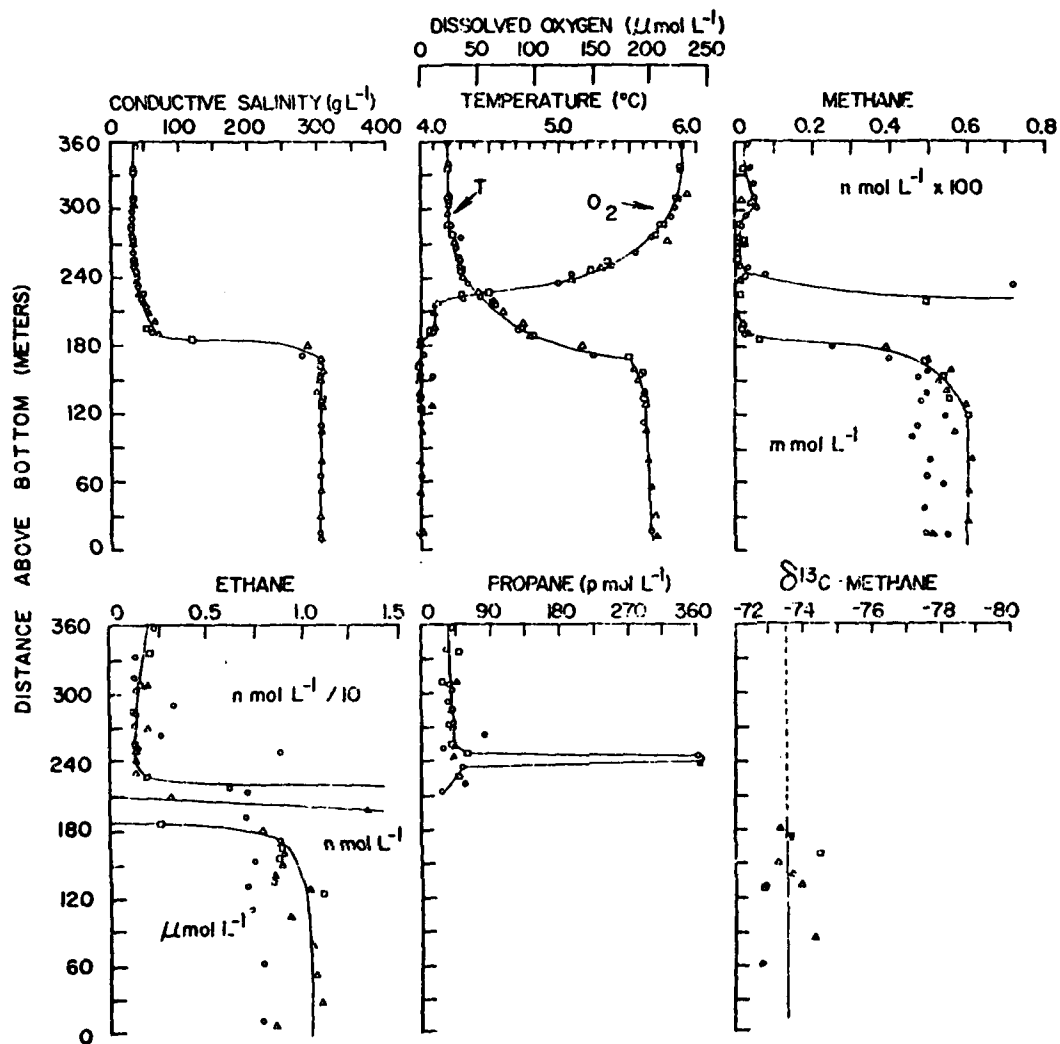


Fig. 30. Vertical distributions of temperature, salinity, dissolved oxygen, methane, ethane, propane and  $\delta^{13}\text{C}\text{H}_4$  in the Orca Basin. Data are plotted as distance above the bottom. Bottom depth = 2436 m. Gas measurements were made on stored samples from cruise 76-G-10, thus methane levels are slightly lower than true value, due to degassing of the supersaturated brine. The closed circles for methane are from cruise 77-G-3.

Table 15. Low-molecular-weight hydrocarbons in the Orca Basin during cruise 77-G-2. A dash indicates sample was lost.

Depth (m)	Salinity (‰)	Temp. (°C)	Oxygen ( $\mu\text{mol}\cdot\text{liter}^{-1}$ )	Methane ( $\mu\text{mol}\cdot\text{liter}^{-1}$ )	Ethane ( $\text{nmol}\cdot\text{liter}^{-1}$ )	Propane ( $\text{nmol}\cdot\text{liter}^{-1}$ )	$\frac{C_1}{C_2 + C_3}$
2023	34.98	4.20	255	0.0040	0.020	0.022	97
2033	34.99	4.21	254	0.0024	0.017	0.038	42
2063	35.02	4.20	254	0.0040	0.017	0.030	84
2103	35.10	4.20	252	0.0047	0.041	0.030	66
2113	35.24	4.21	259	0.0054	0.019	0.029	112
2133	35.92	4.20	240	0.0007	0.010	0.016	27
2153	37.16	4.22	214	0.0023	0.010	0.015	95
2173	39.48	4.27	169	-	0.007	0.007	-
2193	45.98	4.38	77.6	0.0011	0.013	0.019	35
2213	55.98	4.56	8.9	-	1.49	0.113	-
2233	68.88	4.75	9.8	-	0.97	0.430	-
2273	259.7 *	5.57	0	784	1320	<0.9	648
2293	258.5 *	5.62	0	711	1130	<0.9	684
2313	258.3 *	5.62	0	899	1320	<0.9	742
2363	258.4 *	5.65	0	674	1130	<0.9	649
2393	257.9 *	5.66	0	715	1249	<0.9	624
2403	258.2 *	5.65	0	645	1254	<0.9	562
2408	257.8 *	5.66	0	776	1204	<0.9	703
2413	257.8 *	5.66	0	830	1053	<0.9	859

\* These salinity data are from volume dilutions corrected for the density of the brine,  $\rho = 1.185$  at 22.5°C.

the Nansen bottles into equilibration syringes. Some small loss of methane may have occurred when the hydrostatic pressure was reduced on the bottles during their ascent from depth. However, the 750  $\mu\text{mol}\cdot\text{liter}^{-1}$  average of methane (Table 15) is believed to be actual methane concentration in the brine. The methane data from the pressurized samples taken on cruise 78-G-2 and EN-32 substantiate this conclusion. The 78-G-2 and EN-32 methane data are given in Fig. 31. The problems with sampling in the brine preclude observations of any trends in the data in the high salinity brine.

Some structure in the light hydrocarbon distributions was observed in the pycnocline above the brine. In the profiles from cruise 77-G-2 (shown in Fig. 32), a methane maximum was observed above the interface, just above the oxygen depletion zone, at a depth of 2120 m. Methane values from several cruises are given in Fig. 33. Methane concentrations begin to increase above deep ocean concentrations about 50 m above the brine interface. Less distinct ethane and propane maxima also appear above the brine in this region of oxygen depletion (Fig. 32).

The carbon isotopic ratios of methane are shown in Fig. 30. They range from -73 to -74‰ (relative to PDB) and relatively high  $C_1/(C_2 + C_3)$  ratios (average = 684) indicate that the methane is principally of biogenic origin.

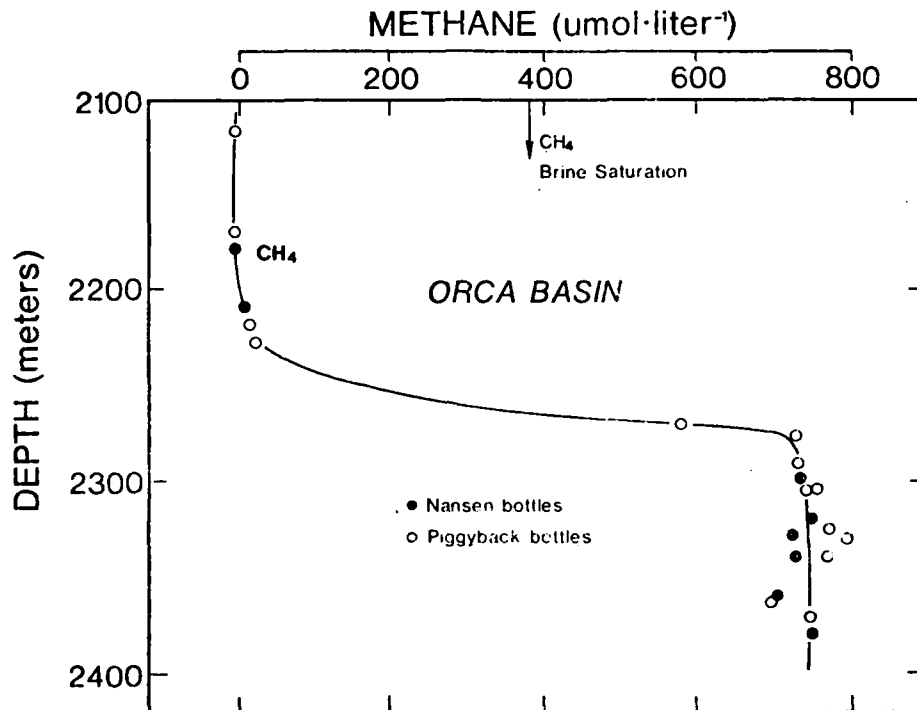


Fig. 31. Dissolved methane depth distributions in the Orca Basin taken from gas-tight piggyback samplers and standard Nansen bottles. Data are from cruises 77-G-13 and EN-32.

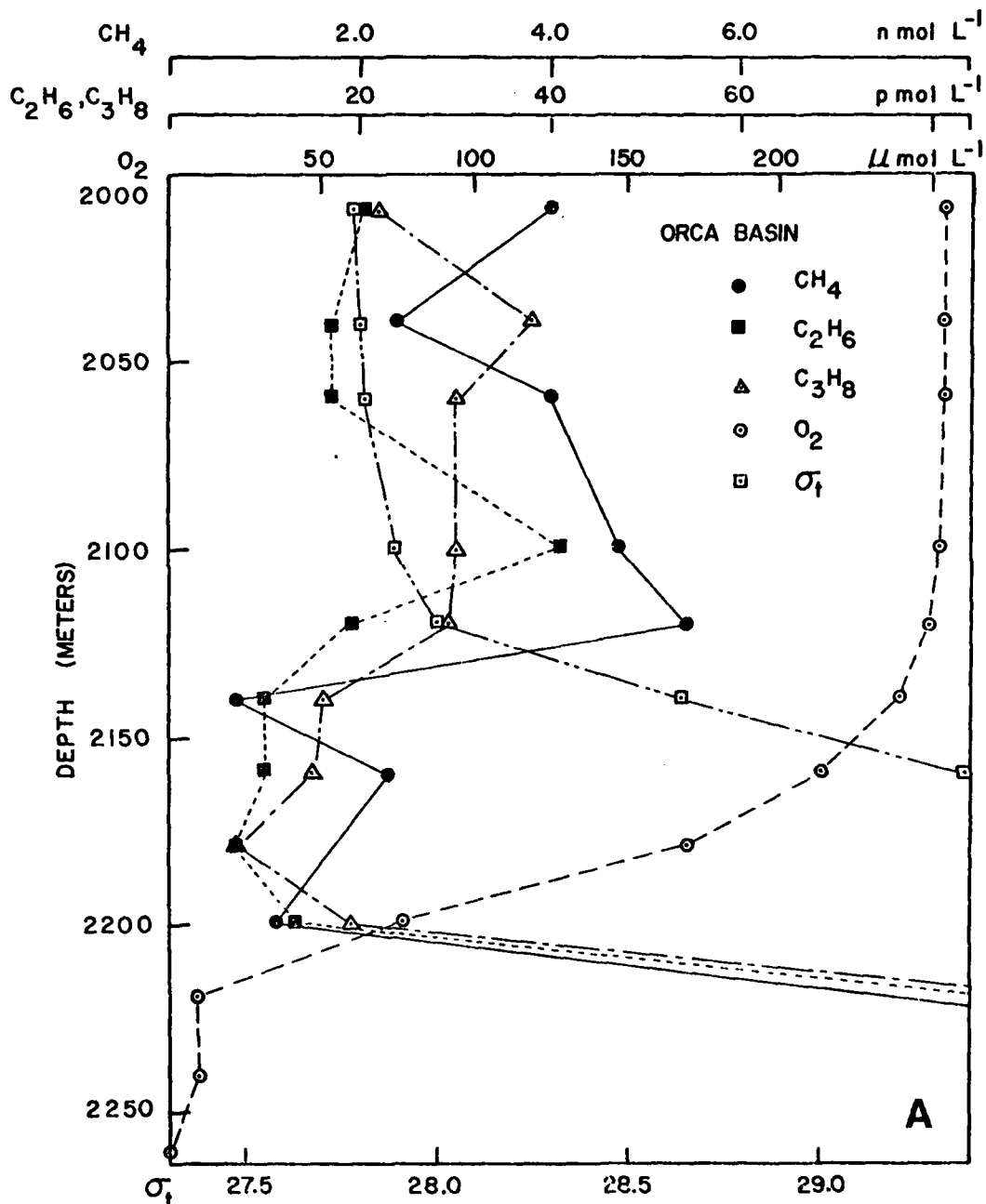


Fig. 32. Hydrocarbon, oxygen and  $\sigma_t$  data from the basin pycnocline (cruise 77-G-2 data). Note the maxima in the methane and ethane profiles at the level where density begins to increase rapidly.

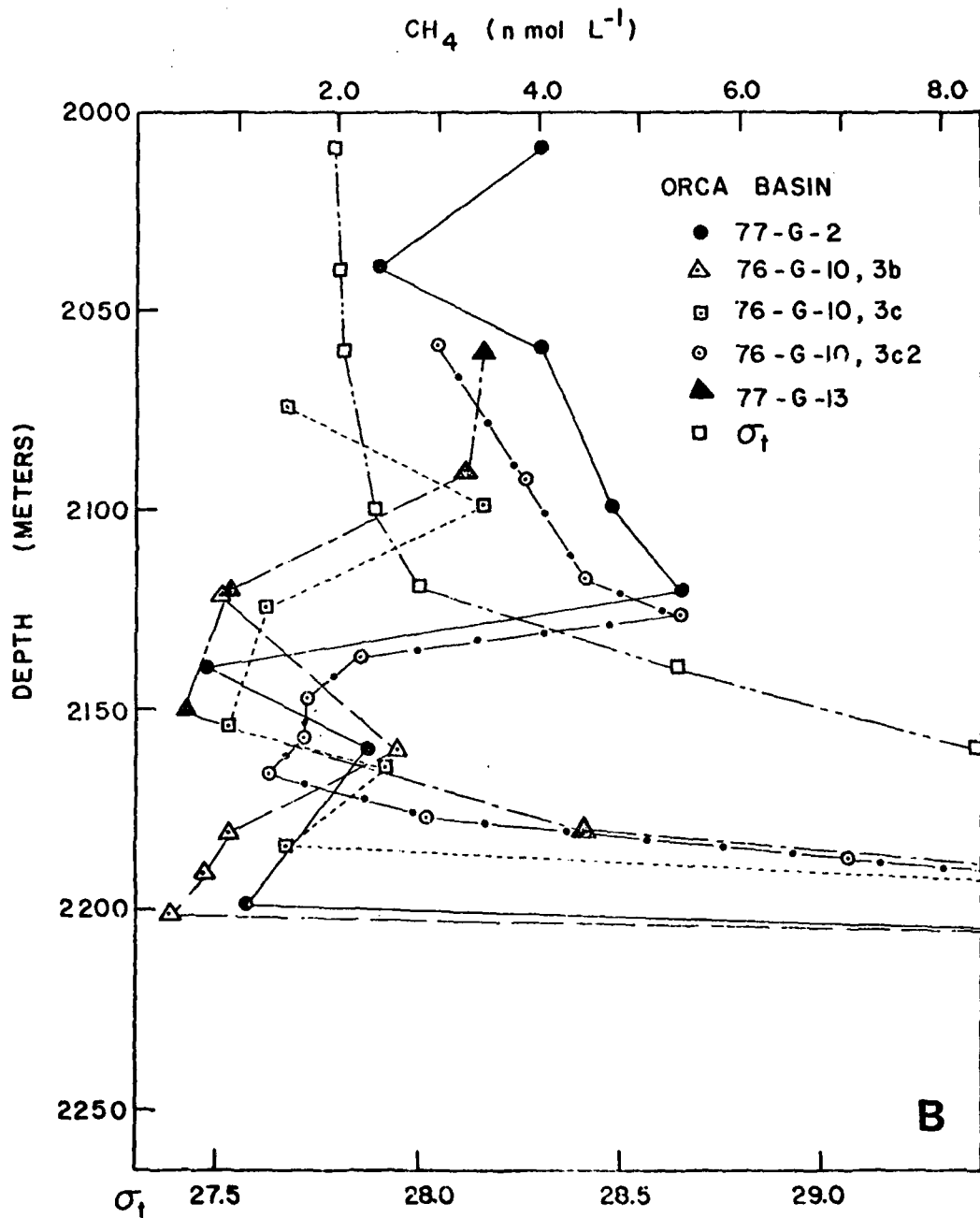


Fig. 33. Methane depth distributions in the pycnocline from different hydrocasts on two separate cruises.

### Nitrous Oxide and Ammonia

The nitrous oxide data collected during cruise 77-G-13 are plotted in Fig. 34, along with dissolved oxygen, nitrate and ammonia measurements from the same cast. Nitrous oxide concentrations above the brine average  $15.3 \text{ nmol}\cdot\text{liter}^{-1}$ . This value is near the atmospheric solubility level that would be predicted ( $12 \text{ nmol}\cdot\text{liter}^{-1}$ ) from the constants of Weiss and Price (1980), Eq. 5 and a value of 290 ppbv as the atmospheric concentration of  $\text{N}_2\text{O}$ .  $\text{N}_2\text{O}$  decreases rapidly near the interface and reaches a near constant level of  $0.28 \text{ nmol}\cdot\text{liter}^{-1}$  within the brine. The  $\text{N}_2\text{O}$  level in the brine may be real, although possible contamination during sample transfer cannot be totally eliminated. Ammonia levels (Fig. 34) increase from essentially zero above the brine to over  $500 \text{ }\mu\text{mol}\cdot\text{liter}^{-1}$  within the brine itself. Ammonia values measured by the three different methods on different cruises (Fig. 35) are consistent, and indicate little structure in the brine itself. While ammonia increases in the brine interface region, nitrate decreases from a mean value in deep Gulf water of  $22.6 \text{ }\mu\text{mol}\cdot\text{liter}^{-1}$  to zero within the brine (Fig. 34). This is direct evidence of the denitrification occurring at the brine interface.

### Hydrogen Sulfide

Hydrogen sulfide, which was also measured on three separate cruises, was not present in the Orca Basin as free  $\text{H}_2\text{S}$ . Measure-



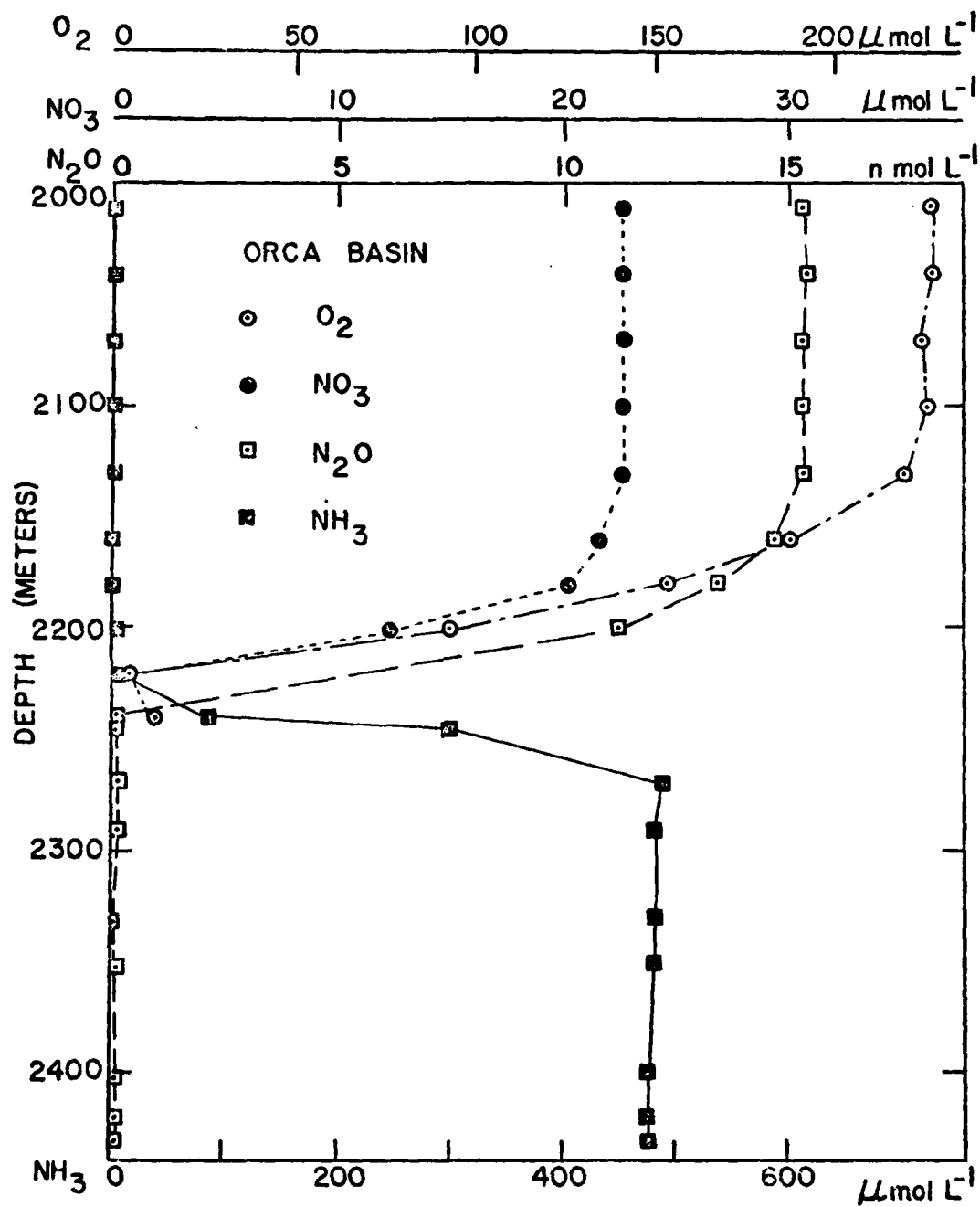


Fig. 34. Vertical distribution of dissolved oxygen, nitrate, N<sub>2</sub>O and ammonia in the Orca Basin. N<sub>2</sub>O is consumed in the anoxic brine<sub>1</sub> along with nitrate while ammonia increases to about 500 μmol·liter<sup>-1</sup>.

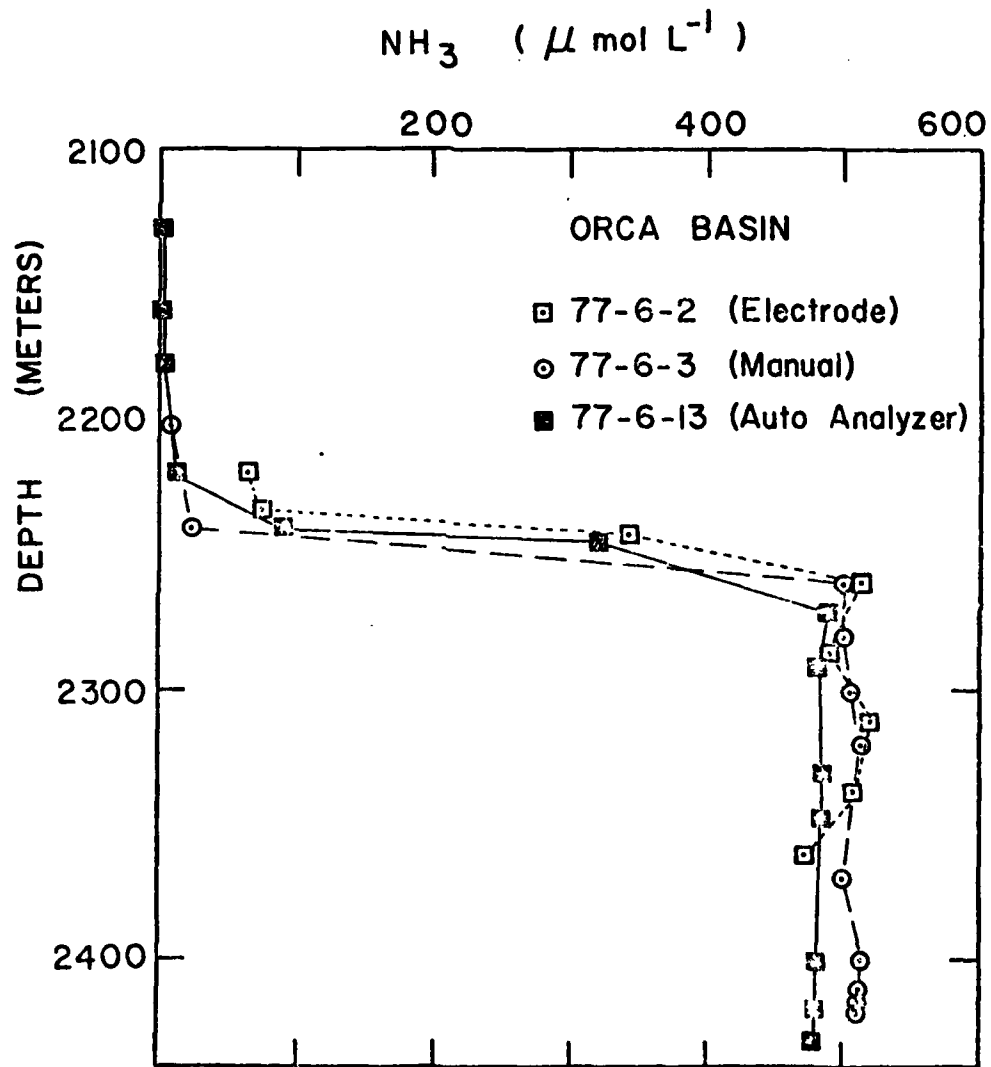


Fig. 35. Vertical profiles of ammonia measured by three separate methods (see text) on three different cruises. No obvious structures appear in the ammonia profile within the brine.

ments in the 3-1000  $\mu\text{mol}\cdot\text{liter}^{-1}$  range indicated that there was no sulfide in the Orca Basin, although some sulfate reduction must be taking place to produce the biogenic iron sulfide found in the Orca Basin sediments (Addy and Behrens 1980). Attempts to measure  $\text{H}_2\text{S}$  in the 1-3  $\mu\text{mol}\cdot\text{liter}^{-1}$  range showed no sulfide, except just below the interface where the highest concentration measured was 2.8  $\mu\text{mol}\cdot\text{liter}^{-1}$  (Fig. 36). Water that was filtered from the interface region contained black particles of iron sulfide. Thus it is plausible that the  $\text{H}_2\text{S}$  values that were measured at the interface were not free  $\text{H}_2\text{S}$ . The Cline (1969) method of  $\text{H}_2\text{S}$  analysis requires the analysis to be performed in an acid solution. Any particulate iron sulfides present in the brine would be converted to free sulfide and appear as  $\text{H}_2\text{S}$  in the measurement. Hydrogen sulfide, if present in Orca brine, may be lower than the sensitivity of the methodology.

Fig. 36 shows the distribution of  $\text{H}_2\text{S}$  sulfate and iron in the Orca Basin. The sulfate levels (33.5  $\mu\text{mol}\cdot\text{liter}^{-1}$  average) are about 18% higher than the sulfate concentrations in deep Gulf of Mexico water above the brine. Iron concentrations increase from trace amounts above the brine to over 30  $\mu\text{mol}\cdot\text{liter}^{-1}$  in the anoxic brine.

#### Discussion

The Orca Basin is a giant, flow-through chemical reactor which

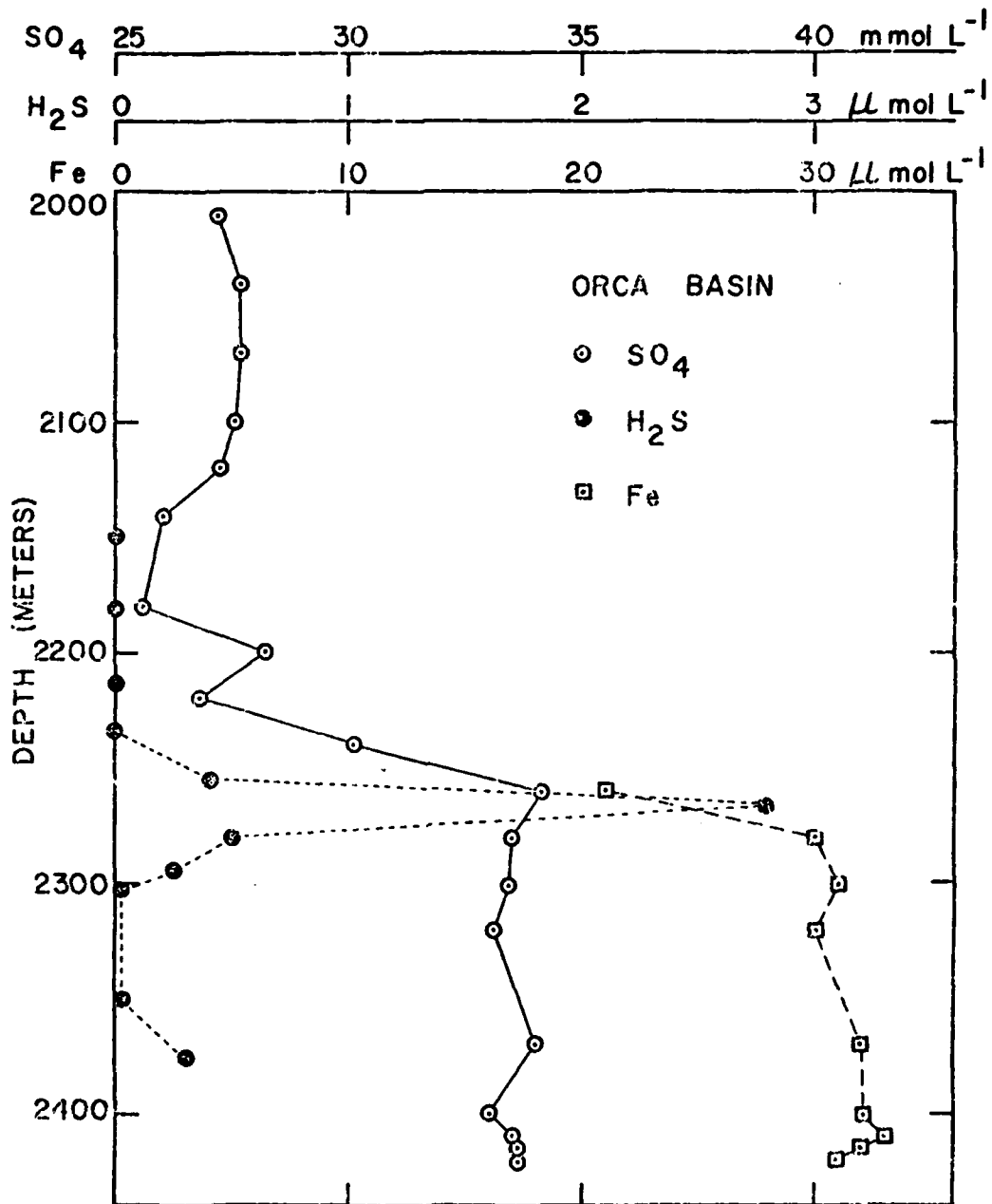


Fig. 36. Depth distributions of sulfate, sulfide and iron in the Orca Basin. Free H<sub>2</sub>S is probably absent from the brine with iron sulfides causing the apparent H<sub>2</sub>S distribution observed. Iron has accumulated in the Orca Basin, unlike other anoxic marine waters, due to the lack of available sulfide to precipitate the iron.

is only unusual in that a very small percentage of the reaction products escape. In any chemical reactor, processes that modify the concentration of a dissolved component can be classified as: external processes, which are active only at the boundaries and internal processes, which are active everywhere in the system. The external processes are of importance in determining the concentration at the boundaries, while the internal processes, together with the boundary concentrations, determine the distribution throughout.

The general equation for the rate of change of the concentration of a dissolved component at any point in a water column may be written as:

$$\frac{\partial c}{\partial t} = \frac{\partial}{\partial x} \left( A_x \frac{\partial c}{\partial x} \right) + \frac{\partial}{\partial y} \left( A_y \frac{\partial c}{\partial y} \right) + \frac{\partial}{\partial z} \left( A_z \frac{\partial c}{\partial z} \right) \quad (21)$$

$$- u \frac{\partial c}{\partial x} - v \frac{\partial c}{\partial y} - w \frac{\partial c}{\partial z} + R$$

where

$A_x$ ,  $A_y$  are horizontal eddy diffusion coefficients,

$A_z$  is the vertical eddy diffusion coefficient,

$u$ ,  $v$  are horizontal advection velocities,

$w$  is the vertical advection velocity with  $z$  positive downward, and

$R$  is the net rate of production or consumption of a component which is not conservative.

This equation describes the effects of diffusion and advection on

the local time rate of change,  $\frac{\partial}{\partial t}$ , of a nonconservative solute whose concentration,  $c$ , is changed chemically or biologically at rate  $R$ .

In the chemical reactor that is the Orca Basin, organic material raining down from the surface provides the fuel for the diagenetic reactions given in Table 12. The processes of decomposition in the Orca Basin are no different than in other anoxic basins, but the high salt content of the water can have a great influence on reaction rates and mechanisms. Eq. 22 also applies to the Orca Basin, but rates of advection, diffusion and reaction may be slower in this dense, hypersaline environment than in other anoxic environments.

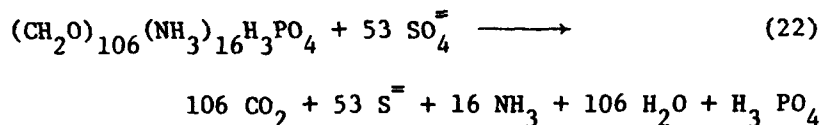
It will be argued in this discussion that the processes of decomposition in the Orca Basin are no different than in other anoxic environments, only slower. Briefly summarized the evidence for reduced microbial activity in the Orca Basin is

- (1) The presence of fronds of Sargassum seaweed in deep cores. Kennett and Penrose (1978) have noted that this type of preservation has never been observed before in the deep ocean.
- (2) A comparison of the initial sulfate gradients between the Orca Basin and the basin next to it indicates that sulfate is not as rapidly reduced in the Orca Basin sediments.

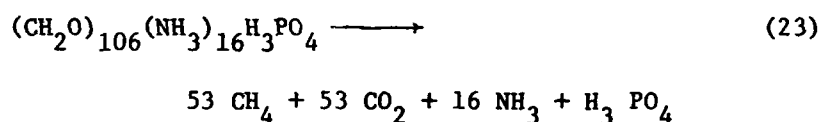
- (3) The isotopic composition of carbon dioxide and methane is unusually light (relative to PDB standard) in the Orca Basin. Sackett et al. (1979) suggested that the isotopically light methane was the result of an unknown lighter source material. However, slower rates of methanogenesis would produce greater fractionation and could account for the observed  $\delta^{13}\text{C}$  values starting from the organic material found in the brine.
- (4) No pyrite is found in the Orca Basin sediments. If there is only a slow production of sulfide in the Orca Basin, the high iron levels would rapidly remove the sulfide from solution and inhibit the continued reaction of metastable iron sulfides to pyrite. Metastable iron sulfides are found in the Orca Basin sediments, pyrite has not been found.

#### Overall Effects of Decomposition

Organic matter in the marine environment has an ideal C:N:P ratio of 106:16:1 by atom which is close to the statistical average of planktonic material, the primary source of organic carbon to the marine ecosphere. Anaerobic decomposition of this organic matter can be expressed by the reactions in Table 12. The most important reaction is sulfate reduction, given by the equation:



This reaction yields mole ratios for  $\text{CO}_2:\text{S}^{2-}:\text{NH}_3:\text{H}_3\text{PO}_4$  of 106:53:16:1. Methane production during anaerobic decomposition will alter the ratios predicted by this model by either liberating or consuming carbon dioxide. Richards et al. (1965) proposed that one representation of methanogenesis might be



yielding mole ratios for  $\text{CO}_2:\text{CH}_4:\text{NH}_3:\text{H}_3\text{PO}_4$  of 53:53:16:1 for the products. This reaction would alter the ratios predicted by Eq. 7 by an amount dependent upon the quantity of methane produced. In the anoxic waters of the Orca Basin, the average concentrations observed for excess decomposition products are  $2.28 \text{ mmol}\cdot\text{liter}^{-1}$  for  $\text{CO}_2$ ,  $0.7 \text{ mmol}\cdot\text{liter}^{-1}$  for  $\text{CH}_4$  (Sackett et al. 1979), less than  $1 \text{ }\mu\text{mol}\cdot\text{liter}^{-1}$  for  $\text{S}^{2-}$ ,  $500 \text{ }\mu\text{mol}\cdot\text{liter}^{-1}$  for  $\text{NH}_3$  and  $60 \text{ }\mu\text{mol}\cdot\text{liter}^{-1}$  for  $\text{PO}_4^{3-}$  (this study). These values result in  $\Delta\text{CO}_2:\text{CH}_4:\text{NH}_3:\text{H}_3\text{PO}_4$  mole ratios of 38:12:8.3:1, far different than predicted by Richard's (1965) model.

How can these differences be explained? It is doubtful that these differences can be accounted for by compositional differences between Gulf plankton, and the world average planktonic ratios.



Plankton falling through the water column decomposes into particulate organic matter (POM). The POM in the deep water should have the same characteristics as the plankton debris which reach the bottom. Particulate organic carbon (POC) and particulate organic nitrogen (PON) have been reported by Fredericks (1972), in both surface waters of the Gulf and at depth. The average values he measured for POC and PON at the surface, and at 200 to 2200 m, are shown in Table 16. When converted to a mole carbon and mole nitrogen ratio, the C/N ratio in surface waters was 3.62 and increased to a deep water C/N ratio of 7.78. The average of C/N ratio for the model of Richards (1965) is 6.6, while the ratio of excess carbon ( $\text{CO}_2 + \text{CH}_4$ ) to nitrogen ( $\text{NH}_3$ ) in the Orca Basin is 6.0. Considering variations in Fredericks' (1970) data with depth all of these ratios are approximately the same.

Since the excess carbon to nitrogen ratio in the Orca Basin approximates the theoretical ratio, there must be excess phosphate in the brine (brine  $\text{PO}_4 = 60 \mu\text{mol}\cdot\text{liter}^{-1}$ ). Phosphate could be added to the brine from organic matter decomposition or by input of phosphate associated with the brine coming from the LouAnn salt, from which the Orca brine originated. Unfortunately, a detailed study of phosphorus content in natural solution brines has not been made. Of the few brines that have been analyzed, most have contained less than  $1 \text{ mg}\cdot\text{liter}^{-1}$  (Collins 1975). For phosphorus,  $1 \text{ mg}\cdot\text{liter}^{-1}$  is equal to  $32 \text{ mmol}\cdot\text{liter}^{-1}$ .

Table 16. Average concentrations of POC and PON in deep Gulf samples along with mole ratios (after Fredericks 1972).

	Gulf Surface $\mu\text{g}\cdot\text{liter}^{-1}$	$\mu\text{g}\cdot\text{liter}^{-1}$	Deep Gulf $\mu\text{mol}\cdot\text{liter}^{-1}$	Orca Basin
PON	6.5	0.46	1.5	0.11
POC	20	1.66	10	0.83
C/N		3.62	7.78	6.03

The excess C/N ratio in the brine can be used to calculate the predicted phosphate from anaerobic decomposition alone. Using the brine C/N value of 6.0, the decomposition-produced phosphate would be  $27.8 \mu\text{mol}\cdot\text{liter}^{-1}$ . This value would give a C:N:P ratio of 106:18:1, which is very near the predicted values from Richard's model. This calculation suggests that there is an excess  $32 \mu\text{mol}\cdot\text{liter}^{-1}$  of phosphate in Orca Basin. Considering the high phosphorus content of natural brine, the excess could be from the original brine that entered the basin before any decomposition of organic matter had occurred.

#### Microbial Activity in the Orca Basin

Distributions of the reduced species in the brine are controlled either by in situ processes within the brine and sediment and/or by the evaporite dissolution and migration of the brine into the basin. Since the brine originates from solution by sea water of a near-surface salt deposit within the basin, the reduced gases should be controlled primarily by in situ processes within the brine, after migration into the basin. The inflowing brine should have initial reduced gas concentrations similar to overlying sea water, as was shown for argon and nitrogen. Thus biological action by anaerobic bacteria in the Orca Basin brine must produce the concentrations of reduced species that were observed.

LaRock et al. (1979) have reported that ATP concentrations and uridine uptake show an increase in magnitude above the brine interface, followed by a decrease below the interface into the anoxic zone. Substantial ATP increases were found further down in the anoxic zone. As in other anoxic environments, the bacterial populations required to mediate the decomposition of organic matter are present in the hypersaline waters of the Orca Basin. One might expect the high salt content of the Orca Basin to inhibit microbial activity. However, the observed ATP levels ( $3-15 \text{ ng ATP}\cdot\text{liter}^{-1}$ ) indicate a larger bacterial population than in the overlying deep Gulf waters, where values from  $1-3 \text{ ng ATP}\cdot\text{liter}^{-1}$  were observed (LaRock et al. 1979). This large bacterial population is probably associated with the high concentration of dissolved organic carbon ( $3.5 \text{ mg C}\cdot\text{liter}^{-1}$ ) and particulate organic carbon ( $70 \text{ }\mu\text{g C}\cdot\text{liter}^{-1}$ ) observed in the brine (Sackett et al. 1979). Although the total bacterial biomass is larger in the brine than in the deep Gulf, one must consider the inhibitory effects of salt on the reaction rates during bacterial decomposition.

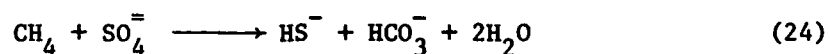
Unfortunately, bacteria activity at high sodium chloride concentrations has received limited attention in the literature. Larsen (1967) and Kushner (1968) have reviewed the literature on halophilic bacteria and noted that they do not exclude salts, but have an internal environment as high in ionic strength as their external environment. Many bacteria are facultative halophiles,

and the sulfate reducers, methanogens, and other anaerobes have probably accommodated themselves to the Orca Basin brine well enough to produce or consume the reduced gases observed. Like increasing pressure or decreasing temperature, the increasing salt content of a solution will reduce the rate at which microbes perform their metabolic activities (Kushner 1978). Most bacteria in hypersaline waters grow slowly, with mean generation times of 7-15 hours at 37°C (Kushner 1968), compared with generation times of less than an hour for bacteria at sea water salt concentrations and 37°C. The large concentrations of biogenic methane in the Orca Basin ( $750 \mu\text{mol}\cdot\text{liter}^{-1}$ ,  $\delta^{13}\text{C}_{\text{CH}_4} = -74\text{‰}$ ), relative to other anoxic basins, and the evidence of sulfate reduction in the sediments suggests that anaerobic bacteria are able to tolerate the hypersaline conditions and actively reduce organic material even at 5.6°C, a temperature much lower than the 30-40°C preferred by most halophilic bacteria. Their slower rate of metabolism in the Orca Basin, however, has a profound influence on the compositions of the decomposition products found there.

Another bacterially-mediated decomposition reaction must be considered in addition to those presented in Table 12. That is the process of anaerobic methane consumption during sulfate reduction. Although the process of methane oxidation under aerobic conditions is well documented (Vary and Johnson 1967; Wertlieb and Vishniac 1967; Quale 1972; Rudd, Hamilton and Campbell 1974)

and bacteria are known to anaerobically utilize hydrocarbons higher than methane (Novelli and Zobell 1947; Rosenfeld 1947; McKenna and Kalio 1965), the actual consumption of methane by bacteria under anoxic conditions is not well understood. Davis and Yarbrough (1966) have suggested that methane could be an additional carbon source for sulfate-reducing bacteria, and Fuhs (1961) has shown several reactions for anaerobic methane consumption that are energy-yielding. Generally, the longer the carbon chain, the more energy the bacteria can obtain.

Although bacteria prefer to use the energy source with the highest potential, it appears that methane consumption can occur during sulfate reduction. In studies of the anaerobic oxidation of  $^{14}\text{C}$ -labeled methane by the sulfate-reducing bacteria, Desulfovibrio desulfuricans, Davis and Yarbrough (1966) noted that these bacteria can consume methane, although at a slow rate. The following reaction was proposed by Fuhs (1961)



with the reaction having a free energy of  $-4.5 \text{ kcal} \cdot \text{mole}^{-1}$ . The consequences of anaerobic methane consumption would be decreased methane levels in the immediate vicinity of consumption, and an increase in the  $\delta^{13}\text{C}$  of the methane. Isotopically lighter methane would be preferentially consumed.

### Methane and Hydrogen Sulfide in the Brine

The concentration of methane in the Orca Basin is extremely high when compared to other anoxic areas (Table 17). The methane levels are over 100 times higher than the concentrations found in any other anoxic sea water, except for the oil-related East Flower Garden brine in the Gulf of Mexico (Brooks et al. 1979). The only natural water with higher methane content is Lake Kivu, an east African rift lake where Deuser et al. (1973) reported finding a methane concentration of  $22 \text{ mmol} \cdot \text{liter}^{-1}$ . The high methane levels in the Orca Basin probably result from the inability of methane produced there to easily escape from the brine.

The methane distribution across the brine-sea water interface is extremely sharp, with almost a six order of magnitude increase in methane concentration as the depth increases only 50 m. Such a sharp gradient is indicative of very slow mixing processes. Thus methane can probably be removed from the basin only by molecular diffusion, a very slow process. The gas that bubbles out of the brine at the surface has a large component of methane. Methane is supersaturated by several hundred percent (at one atm). This supersaturation is the result of methane production with little removal. The methane is produced in the sediments and perhaps at the brine-sea water interface.

It is the density structure of the Orca Basin (Fig. 9) which effectively inhibits removal of methane from the basin. Due to

Table 17. Molecular and isotopic composition of methane and other gaseous hydrocarbons in anoxic waters.

Location	Depth (meters)	Methane ( $\mu\text{mol}\cdot\text{liter}^{-1}$ )	Ethane ( $\text{nmol}\cdot\text{liter}^{-1}$ )	Propane ( $\text{nmol}\cdot\text{liter}^{-1}$ )	$\text{C}_1/\text{C}_2+\text{C}_3$ (by mole)	$\delta^{13}\text{C}$ (‰)	Reference
Lake Kivu	425	22000	16000		1000 <sup>†</sup>	-44	1
Black Sea	1900	5.5	13.7	0.15	400		2
Black Sea	2000	7.6	17.4	0.11	430		3
Cariaco Trench	1250	6.7	1.8	0.24	3200		4
Cariaco Trench	1200	10					5
Cariaco Trench	1400	7.1					6
Lake Nitinat	200	71	1.1	2.0	23000		5
Red Sea- Atlantis *	2050	2.6	23	1.5	104		5
East Flower Garden *	72	122	18500	0.65	6.6	-40	7
Orca Basin *	2300	750	1300	<1	730	-73	8

\* Brine areas.

<sup>†</sup> Estimated from other data in reference 1.

1 Deuser, Degens and Harvey (1973)

2 Hunt (1969)

3 Swinnerton and Linnenbom (1969)

4 Linnenbom and Swinnerton (1969)

5 Atkinson and Richards (1967)

6 Wiesenburg (1975)

7 Brooks, Bright, Bernard and Schwab (1979)

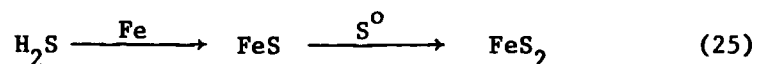
8 This study.



its stability (Table 5), the residence time of brine in the Orca Basin must be longer than in other anoxic basins. Sackett et al. (1979) found a  $\delta^{14}\text{C}$  value for the total  $\text{CO}_2$  in the brine of  $-501 \text{ ‰}$ . This figure would imply an age of one half-life of  $^{14}\text{C}$ , or about 5700 years. This residence time is much longer than for other anoxic basins. There is much uncertainty in the  $^{14}\text{C}$  age, however, due to a lack of knowledge of the  $^{14}\text{C}$  content of carbon compounds that entered the basin with the brine. Addy and Behrens (1980) used sedimentological data and  $^{14}\text{C}$  measurements of the carbonate in the sediments to calculate an age for the initial input of brine into the Orca Basin. They determined that the brine began flowing in to the basin 7900 years ago, a date even older than the age from the  $^{14}\text{C}$  measurements of Sackett et al. (1979). The age of Orca Basin water is greater than 800-year deep water residence time for the Cariaco Trench (Fanning and Pilson 1972) or the 2000-year residence time for bottom water in the Black Sea (Ostland 1974).

Unlike methane, the concentration of hydrogen sulfide in the Orca Basin brine (Fig. 36) is much lower than in other anoxic basins such as the Cariaco Trench or the Black Sea (Fanning and Pilson 1973; Brewer and Spencer 1974). Except at the brine sea water interface, sulfide is below the detection limits of the molybdenum blue technique used in this study (Cline 1969). In the

Black Sea at 2000 m, both methane and hydrogen sulfide are observed in high concentrations (Degens and Ross 1974). In the hot Red Sea brine at 2050 m, methane is present (Burke et al. 1980) but hydrogen sulfide is absent. There is little evidence that significant sulfate reduction is taking place in the Red Sea brine (Craig 1969). It can be argued that the rate of sulfide production in the Orca Basin is not sufficient to maintain a high dissolved sulfide level, due to the presence of iron with which it precipitates as metastable iron sulfides. Alternately, it has been suggested (Trabant and Presley 1978) that sulfate reduction proceeds normally in the Orca brine, and there is simply a large influx of iron from the sediments which is controlling the sulfide levels in the brine. This discussion will show that the rate of sulfate reduction is slow in the Orca Basin, and that reduced sulfur species are completely removed from solution by reactive iron. As such, the available sulfur is rapidly depleted from solution before any conversion of FeS to FeS<sub>2</sub> can occur, by the reaction



Addy and Behrens (1980) noted that there is no pyrite in the Orca Basin sediments.

Sulfate reduction is occurring in the Orca Basin. The sulfate in the pore waters decreases to zero at a depth of five to

seven meters in the sediment (Sackett et al. 1979). Fig. 37 shows a plot of the sediment data presented tabularly by Sackett et al. (1979). The open circles represent the actual sulfate profile that they observed. Also shown are the chloride gradient and a predicted sulfate profile ( $\text{SO}_4$ , closed circles) that would be found if sulfate were diffusing down through the sediment at the same rate as chloride, with no sulfate reduction. From the differences in the actual and predicted sulfate profiles, it is obvious that sulfate reduction is occurring in the Orca Basin sediments. The presence of hydrogen sulfide at the interface with the oxygenated Gulf of Mexico (Fig. 36) water also shows that sulfide is being produced there. However, the absence of  $\text{H}_2\text{S}$  in the brine itself may indicate that little sulfide is being produced in the anoxic waters or if so, it is rapidly removed. The only appreciable sulfate reduction is probably occurring at the interfaces, the sites of highest reactivity.

Although it is difficult to obtain rate data from concentration profiles alone, enough information is available about sulfate profiles worldwide to gain some insight into rates in the Orca Basin. The basis for the following discussion follows the review of Goldhaber and Kaplan (1974) on worldwide sulfate profiles and the more recent paper of Berner (1978). Both of these papers discussed a correlation between the sedimentation rate and the sulfate reduction rate, or the initial sulfate gradient in the near surface sediments.

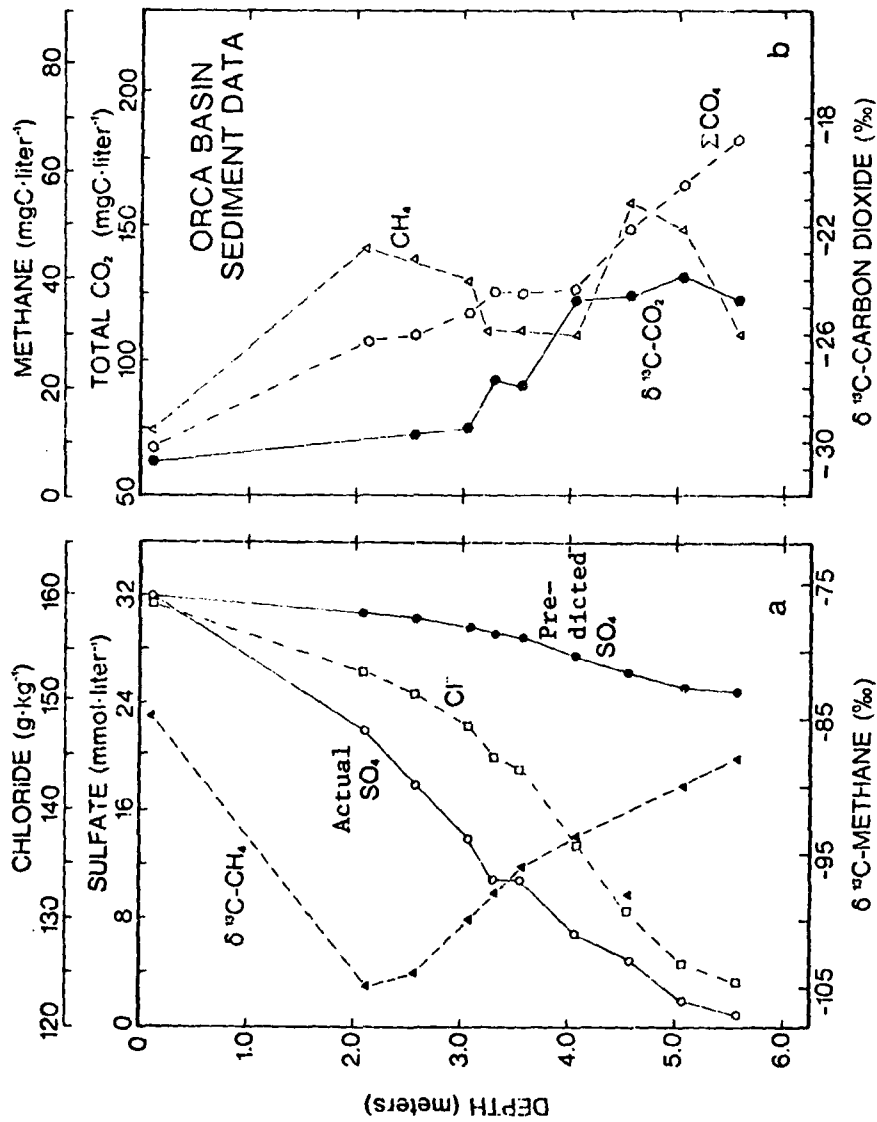


Fig. 37. Sediment geochemistry data from a piston core taken in the Orca Basin (27°01.3'N, 91°16.5'W) at a water depth of 2340 m. Data are plotted from tabulated values given in Sackett et al. (1979). Predicted SO<sub>4</sub> profile is calculated as described in the text.

Berner (1978) has shown that the log of the slope of the sulfate profile in marine sediments is directly correlated with the log of the sedimentation rate. Also the rate constant for sulfate reduction is directly correlated with the rate of sediment deposition. This trend has been shown to apply to diverse environments from nearshore sediments to abyssal Pacific sediments sampled during the Deep-Sea Drilling Project. The actual sulfate sediment profile in Fig. 37 (data from Sackett et al. 1979) has a slope of  $0.053 \text{ mM SO}_4 \cdot \text{cm}^{-1}$ . The sedimentation rate in the Orca Basin has been determined by Addy and Behrens (1980) to be  $61 \text{ cm} \cdot \text{ka}^{-1}$ . Approximately  $21 \text{ cm} \cdot \text{ka}^{-1}$  of this sedimentation is attributable to the preservation of excess carbonate material, as a result of the anoxic hypersaline conditions which exist in the Orca brine. Tompkins and Shephard (1979) estimated a sedimentation rate of  $100 \text{ cm} \cdot \text{ka}^{-1}$ . However, the core that they used for making this estimate was taken in the smaller lobe of the Orca Basin and had evidence of slumping, which would cause their rate to be high.

Taking the rate versus rate hypothesis one step further, Berner (1978) showed that there is a positive correlation, and a very predictable one, between the sedimentation rate and the rate of sulfate reduction. This point was also well illustrated by Goldhaber and Kaplan (1974). Goldhaber and Kaplan (1974) concluded that, besides temperature and pressure (which effect all reactions), the rate of sulfate reduction is dependent upon:

- (1) total organic carbon preserved in the sediment
- (2) the state of complexing of the organic matter
- (3) availability of organic matter for biogenic degradation
- (4) the environment of deposition
- (5) the sedimentation rate.

They noted that factors 1, 2 and 3 are directly influenced by factors 4 and 5, and that there was a direct relationship between the rate sulfate reduction and the rate of sediment accumulation.

Considering factor 1, Sackett et al. (1979) noted that the percentages of organic carbon in Orca sediments are higher than for normal shelf and slope sediments. According to Goldhaber and Kaplan (1974) this would result in higher sulfate reduction rates.

From a comparison of the sulfate reduction rates in the Orca Basin with those in the small basin next to Orca, it can be shown that the rate of sulfate reduction is slower in the Orca Basin, relative to the smaller basin and to other areas of similar sedimentation rate. The sulfate profile in the small basin next to the Orca Basin has been measured by Bernard (1978). A comparison of the initial sulfate gradient,  $(\partial \text{SO}_4 / \partial z)_0$ , of the small basin ( $0.11 \text{ mM SO}_4 \cdot \text{cm}^{-1}$ ) with the Orca Basin ( $0.053 \text{ mM SO}_4 \cdot \text{cm}^{-1}$ ) indicates that the gradient is steeper and thus the rate of sulfate reduction in the other basin is greater than the Orca Basin. Yet the sedimentation rate in the Orca Basin is at least  $20 \text{ cm} \cdot \text{ka}^{-1}$  higher. The sulfate reduction rate should be higher, not



lower in the Orca Basin. If the hypothesis of Berner (1978) were applicable to this area, the sedimentation rate in the small basin next to Orca should have a sedimentation rate twice as fast as the Orca Basin. The two basins are separated by only 25 km. Thus it is reasonable to assume that the same material being deposited due to sedimentation is available to each basin.

The major factor causing the differences in the initial sulfate profiles and sulfate reduction rates of the two basins must be the high salinity brine in the Orca Basin. The hypersaline water in the Orca Basin does not completely inhibit sulfate reduction per se, but the rate of sulfate reduction seems to be decreased as a result of the high salt content, creating a smaller sulfate gradient.

The idea that the rate of microbial activity is slower in the Orca Basin than in other anoxic environments is consistent with the effects of salt stress on microbes (Kushner 1978). A reduced microbial activity rate might also explain some seemingly anomalous carbon isotope data that has been reported for the basin sediments (Sackett et al. 1979). It should be remembered that the production of methane from organic material is a two-step process. One suite of bacteria breaks down organic matter into small organic molecules and carbon dioxide. Methanogenic bacteria then reduce these compounds to methane. The sediment methane data taken from Sackett et al. (1979) are shown in Fig. 37. They



explained the data by suggesting that the source material for the carbon in the Orca Basin must be isotopically lighter than could be accounted for by the isotopic ratios of the products ( $\text{CO}_2$  and  $\text{CH}_4$ ) they observed. They further suggested that the ultimate source for the isotopically light organic carbon in the basin might be methane migrating upward from a deeper reservoir. This methane would be oxidized to  $\text{CO}_2$  and subsequently reduced back to methane, yielding the isotopic balances observed. Several mass balance calculations from mixture models suggested that the source carbon for the source organic material in the basin needed to be  $-42\text{‰}$  to account for values observed.

An alternate argument can be made, if one considers that the rate of sulfate reduction and methane production are lower in this unusual environment than in other areas where methane has been measured and classified isotopically (Bernard 1978). Most of the organic material found in the sediments of the Orca Basin had an isotopic composition essentially the same as other sediments in the region ( $\delta^{13}\text{C} \cong -22\text{‰}$ ). Some depths, however, had isotopically light organic carbon, which may represent slumped sediments, originally deposited in estuarine environments during the Pleistocene (Sackett et al. 1979). For this discussion, I assume that the normal slope organic carbon ( $\delta^{13}\text{C} = -22\text{‰}$ ) is the source carbon for methane and carbon dioxide found in the basin water and sediments. The normal isotopic fractionation for methane produced during this process varies as a function of several factors,

including isotopic content of the source material (Bernard 1978) and temperature of reaction (Gams et al. 1978). A decrease in temperature increases fractionation, perhaps by decreasing the reaction rate, thus allowing increased bacterial selectivity of the source carbon. The idea of enhanced isotopic fractionation at reduced reaction rates has been suggested in several studies. Kaplan and Rittenberg (1964) showed that isotopic fractionation ( $^{32}\text{S}$  vs.  $^{34}\text{S}$ ) during sulfate reduction was directly influenced by the rate of sulfate reduction. The magnitude of the separation between sulfur isotopes was inversely proportional to the rate of sulfate reduction. The same effect should be true for methane carbon fractionations.

In reality, factors other than reaction rate are responsible for changes in fractionation. Fractionation in microbial systems occurs along enzymatic pathways, due to the preferences of transport enzymes for carbon-12 atoms. Under slower reaction rates, or salt stress, the enzymatic pathways may change or the effect of various back reactions (with different fractionations) may become important. These complex systems are difficult to investigate. Thus the simplistic idea of greater fractionation at reduced rates has been generalized to describe the overall effects of these processes.

Also important for this discussion is a consideration of the organic material used during sulfate reduction. It is now well

established that methane can be consumed during sulfate reduction in natural environments (Barnes and Goldberg 1976; Reeburgh 1976; Martens and Berner 1977; Murry et al. 1978). However, methane is not the preferred substrate for sulfate reducing bacteria (Rosenfeld 1947; McKenna and Kalio 1965). Sulfate reducers derive more energy from consuming organic material with a carbon chain length greater than one. In one study where the sulfate reduction rate was low, no methane was consumed (Sorokin 1957). Methanogenesis in the Orca Basin sediments would produce isotopically light methane which would remain unaltered during slow sulfate reduction.

If the rate of methanogenesis is very slow in the Orca Basin sediments, one would expect a greater fractionation than has been observed in other environments. Laboratory fractionation experiments (e.g., Games et al. 1978) are performed under conditions where reaction rates are relatively high, so results can be obtained rapidly. Games et al. (1978) found fractionation factors of 1.044 to 1.061 for methane produced in laboratory cultures. If they had used the carbon dioxide in the Orca Basin as the source material, these fractionations would produce methane in the range  $-71$  ‰ to  $-88$  ‰. If their methane production rates had been slower, the fractionation might be even greater. Greater fractionation would produce methane lighter than  $-88$  ‰, and could produce the  $-105$  ‰ that was measured by Sackett et al. (1979).

The idea of the salt in the basin being the factor which reduces the rates of methanogenesis and sulfate reduction is paramount to this discussion. In the Red Sea brines, Watson and Waterbury (1969) attempted to culture sulfate reducing bacteria in brine at various sodium chloride concentrations. They found that there was a reduction in the rate of sulfate reduction with the increasing salinity and no sulfate reducers could grow at salinities greater than 125 ‰. Some sulfate reduction is taking place in the Orca Basin sediments, as evidenced from the sulfate profiles shown in Fig. 37. However, the rate of sulfate reduction may not be constant throughout the entire sediment column. Microbial activity may be more rapid in certain areas. One of these areas would be the sediment-water interface zone, where the most labile organic material reside. It has often been noted that very rapid sulfate reduction rates occur in a very thin zone near the sediment-water interface (Berner 1972; Sweeney 1972; Goldhaber and Kaplan 1974). Rapid sulfate reduction at the sediment-brine interface could account for both the decrease in methane and the increase in  $^{13}\text{C}$  of the methane in the near surface sediments (Fig. 37). This is the same type of response that is found in coastal sediments in the Gulf of Mexico (Bernard 1978) with methane concentrations decreasing and becoming isotopically heavier near the sediment water interface. Significantly, in the Orca Basin, the heaviest methane is found at the sediment-brine interface.

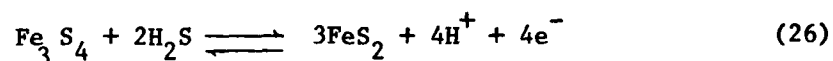
Methane consumption during more rapid sulfate reduction at the sediment-brine interface would also account for the fact that methane is isotopically heavier in the water column ( $\delta^{13}\text{C} = -73.5$  ‰) than in the sediments ( $\delta^{13}\text{C} = -85$  to  $-105$  ‰). The lighter methane would be consumed at the interface leaving the isotopically heavier methane to diffuse out into the water column. Conversely, away from the sediment-brine interface, where microbial populations would be lower (Zobell 1942), rates of sulfate reduction and methanogenesis may increase with depth. This would happen since there is a decreasing chloride gradient with depth. The data in Fig. 37 are consistent with this explanation.  $\delta^{13}\text{C}$  of the methane would increase with depth in the sediment as the sulfate reduction rates increase or as heavier organic carbon is encountered for use in methane production.

In the water column, there is too much scatter in the methane and  $\delta^{13}\text{CH}_4$  data (Fig. 30) to determine the processes affecting the methane distribution there. Methane is obviously consumed at the brine-sea water interface by aerobic oxidation, but there are no other noticeable trends in the deeper data. The high salinity brine in the Orca Basin is fairly well mixed, as was discussed in Chapter II. This mixing is obvious in the methane distribution. In summary, the observed methane profile in the high salinity brine results from production and consumption in the sulfate containing sediment and brine, with only limited loss of

methane from the brine, probably by diffusion across the brine-sea water interface.

#### Mechanisms of H<sub>2</sub>S Removal

Having concluded that sulfate reduction is an active process in the Orca Basin, one must ask where the produced sulfide goes. Free sulfide that is produced is rapidly removed to the sediments as iron sulfide. The iron sulfides that are formed, however, are not pyrite (FeS<sub>2</sub>) but metastable iron sulfides such as greigite (Fe<sub>3</sub>S<sub>4</sub>) and machinawite (FeS) among others (Addy and Behrens 1980). Berner et al. (1979) have noted that the formation of pyrite is directly influenced by the presence of available sulfur. The conversion from metastable iron sulfides to pyrite occurs by the addition of sulfur to FeS (Rickard 1975), and not by the removal of iron. A generalized reaction is (Berner et al. 1979):



Metastable iron sulfides such as greigite and mackinawite are thermodynamically unstable relative to pyrite, in the presence of H<sub>2</sub>S (Berner 1972). Thus the absence of pyrite in the Orca Basin sediments is indicative of the influence of iron in rapidly removing the free sulfide from solution.

The 30-33 μmol·liter<sup>-1</sup> concentrations of iron that were found in the brine (Fig. 36) are two orders of magnitude higher

than the iron values reported for the Black Sea (Brewer and Spencer 1974). In the Black Sea, the hydrogen sulfide concentration is 300-500  $\mu\text{mol}\cdot\text{liter}^{-1}$  and iron sulfides are known to precipitate, as evidenced by their concentrations in the sediment (Berner 1974). The high iron concentration in the Orca Basin has caused almost complete sulfide removal from the Orca Basin waters. With a slow input of microbial sulfide to the brine, the iron that is entering the brine from sediment reaction or from decomposing plankton precipitates all available sulfide. In the Orca Basin, sulfide is the limiting species in FeS precipitation.

#### Other Light Hydrocarbons in the Brine

The production of methane by fermentation shows (Thayer 1931; Stadtman and Barker 1951) that fatty acids are not decarboxylated during methanogenesis and regardless of the length of the carbon chain no hydrocarbons higher than methane should be produced. In spite of this conclusion,  $\text{C}_2$  and  $\text{C}_3$  hydrocarbons are found in many anoxic environments (Table 17) and are probably produced by bacteria (Bernard 1978). In the Orca Basin, ethane concentrations as high as 1300  $\text{nmol}\cdot\text{liter}^{-1}$  were observed, over 50,000 times the concentration found in the deep Gulf water. Propane was also observed just above the interface at a level of 0.4  $\text{nmol}\cdot\text{liter}^{-1}$ ,

almost 20 times the deep Gulf concentration. This type of increase in non-methane gaseous hydrocarbons has also been observed in the Cariaco Trench (Linnenbom and Swinnerton 1969), the Black Sea (Hunt 1974), the Red Sea brine and Lake Nitinat (Swinnerton and Linnenbom 1969) and other anoxic waters (Table 17). Bernard (1978) has shown that these hydrocarbons are also found in anoxic sediments. He attributed these gases to bacterial production. Biological production of  $C_2$  and  $C_3$  hydrocarbons has yet to be satisfactorily explained. The generally accepted mechanisms of anaerobic decomposition (Berner 1971) would preclude microbial production of ethane and propane. Gaseous hydrocarbons higher in molecular weight than methane are normally the result of petroleum production. However, the depth distribution of ethane in the Orca Basin (Fig. 30) indicates an origin similar to methane: biogenic.

#### Methane at the Interface

The maximum in methane, ethane, and propane above the brine-sea water interface was not entirely unexpected. The microbial biomass distribution (LaRock et al. 1979) exhibits a pattern above the interface similar to the observed methane distribution. The two may be associated. The increased density of the brine, relative to the Gulf water, would cause particles falling to the sea floor to be temporarily interrupted in their journey. They would rest at the interface until they reach isotonic balance with the



higher salinity brine. Only after particle densities reach the level of the next brine layer would they settle below the interface. An increased microbial biomass would be expected to correlate with a great number of particles. Degradation of particles at density interfaces has been proposed to account for the near surface methane maxima that have been observed in many areas of the world oceans (Brooks et al. 1980). The main difference here is the depth of maxima above the Orca Basin. This methane maximum is deeper than any previously reported.

The methane depth distribution above the brine interface is also consistent with the concept of methane consumption in oxygenated waters. Rudd et al. (1974) have shown that, in oxygenated waters, methane consumption rates are the highest where the oxygen values are minimal. They made this observation from determinations of methane consumption rates in oxygen minimum layers of several Canadian Shield lakes. As can be seen in Fig. 32, the methane maximum above the interface is in water that has a decreasing oxygen gradient, but where oxygen is still present. This would rule out possible methane consumption during sulfate reduction, since sulfate is not reduced while oxygen is available. Oxygen, from molecular oxygen, must be the oxidizing agent in any microbial methane consumption above the interface.

It must be assumed that interface methane, and presumably ethane and propane, being produced are produced in anaerobic

microenvironments. These microenvironments in the aerobic water column need not be entirely sulfate depleted as Oremland (1975) has reported small amounts of methane production ( $1.81$  to  $1.86 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{hr}^{-1}$ ) in sulfate-rich marine sediments in the Florida Keys. Methane may be produced throughout the entire interface region in these microenvironments, with a constant production rate. This idea, plus surface venting, has been proposed to account for the methane maxima in some surface waters (Scranton and Brewer 1977). If this were the case, the decrease in methane below the maximum would result from an increased rate of methane consumption in the zone where the oxygen concentration is the lowest. The increased consumption at low oxygen concentrations observed by Rudd et al. (1974) may not have real implications for consumption vs. oxygen level. Not coincidentally, the largest population of methane consuming bacteria would also be found in the low oxygen region, since it is closest to the methane source.

#### $\text{N}_2\text{O}$ Consumption During Denitrification

In the sequence of organic decomposition which proceeds from oxygen consumption to carbon dioxide reduction, oxidized nitrogen, especially  $\text{NO}_3^-$  and  $\text{NO}_2^-$ , is consumed by various denitrifying bacteria and converted to gaseous nitrogen compounds, mainly nitrogen gas and ammonia. The geochemistry of molecular nitrogen gas was discussed in Chapter III. The presence of the  $500 \mu\text{mol}\cdot$

liter<sup>-1</sup> of ammonia can be accounted for based on organic decomposition, using the Orca brine excess carbon to nitrogen ratio as a starting point. In measurements of nutrient regeneration in the anoxic sediments, Martens and Berner (1977) show a large NH<sub>3</sub> production, but note very emphatically that there was no production of molecular nitrogen in the sediments. The same is true for the high salinity, anoxic brine in the Orca Basin. Yet, Weiss (1969) has suggested that there is nitrogen production in the Red Sea brines, presumably from decomposition of organic nitrogen. This process causes both the high N<sub>2</sub> concentrations and the high N<sub>2</sub>/Ar ratios that he observed there.

Denitrification is nearly complete in the Orca Basin waters. All NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> have been reduced and the main repository for reduced organic nitrogen is ammonia. Nitrous oxide (Fig. 34) decreases from over 15 nmol·liter<sup>-1</sup> above the brine to only a trace amount in the brine. Thus, almost all the N<sub>2</sub>O has been consumed during denitrification in the Orca Basin waters. Cohen and Gordon (1978) have demonstrated that N<sub>2</sub>O can be consumed not only in completely anoxic waters such as Saanich Inlet, but they noted biochemical N<sub>2</sub>O was consumed by marine denitrifying bacteria. Barbaree and Payne (1967) and Payne et al. (1970) found that cultures of Pseudomonas perfectomarinus can use N<sub>2</sub>O as a terminal oxidant and reduce it to N<sub>2</sub>. Matsubara and Mori (1968) also show that anaerobic cultures of P. denitrificans can reduce N<sub>2</sub>O to

$N_2$  with lactate as the terminal hydrogen donor. Although  $N_2O$  has been depleted in the Orca brine and increased  $N_2$  concentrations have been measured there (Fig. 26), it seems unlikely that  $N_2$  production from  $N_2O$  is significant there. For the organic nitrogen (which is eventually converted to ammonia) to participate in the nitrogen production, it must first be oxidized to  $NO_3^-$ . In anaerobic waters, however, there is little reason to believe that this type of reaction would occur, and if so that the  $NO_3^-$  would then be converted to  $N_2$ . The possibilities of a two step production of  $N_2$  from ammonia in the oxygen step region have been discussed earlier.

In summary, the decomposition processes that produce the reduced gases in the Orca Basin are no different than in other anoxic environments. The rates are just slower. A combination of the slower reaction rates and the high stability of the brine in the Orca Basin makes this environment an ideal laboratory for studying the mechanisms involved in anaerobic decomposition. The isotopic fractionations found there are the greatest reported to date, far greater than could be produced in laboratory experiments over the lifetime of a graduate student. An examination of the degree of sulfur fractionation in this basin would prove valuable for better understanding reduced gas biogeochemistry under hypersaline conditions.

CHAPTER V  
CARBON DIOXIDE AND THE CARBONATE SYSTEM IN  
THE HYPERSALINE BRINE

The Orca Basin brine is basically a sea water solution that has been saturated with sodium chloride (Table 3). Besides sodium chloride, however, the most important compound in higher concentration in the brine than in normal sea water is carbon dioxide. Some carbon dioxide probably entered the basin with the inflowing brine, but most excess  $\text{CO}_2$  was produced in the basin itself. The isotopically light carbon dioxide ( $\delta^{13}\text{C} = -16.4\text{‰}$ ) in the brine (Sackett et al. 1979), compared to the source Gulf water ( $\delta^{13}\text{C} = 0.3\text{‰}$ ), indicates that the excess carbon dioxide has a biogenic source. Biogenic reactions which produce carbon dioxide have been discussed in the previous chapter (see Table 12). The effects of hypersalinity in the Orca Basin on the interactions controlling carbon dioxide system in the Orca Basin are considered here. The previous chapters have been concerned with dissolved gases which generally do not react with water. Carbon dioxide, on the other hand, engages in a series of equilibrium reactions when it comes in contact with water or sea water. These carbonate system reactions will determine the disposition of the biogenic  $\text{CO}_2$  added to the brine from the anaerobic decomposition of organic matter.

Ionic interactions are important in defining the carbon dioxide system. The total ionic strength of the Orca Basin brine (4.10) is quite large relative to normal sea water (~0.7). The ionic strength and ionic balance percentages of the ions in the Orca Basin, sea water and several other brines are given in Table 18. Since the ionic strength of brines is so high, one would expect their physical-chemical properties to be strongly influenced by ionic interactions. Any physical-chemical effects would be due both to the total ion concentration, as well as to the nature of the ions which are important in the interactions in question. Different ions participate to varying degrees in ion pairing (Garrels and Thompson 1962). In sea water, there are interactions between  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Ca}^{++}$ , and  $\text{Mg}^{++}$  with  $\text{HCO}_3^-$ ,  $\text{CO}_3^{=}$ , and  $\text{SO}_4^{=}$ . Johnson and Pytkowicz (1979) have recently calculated that  $\text{Cl}^-$  can also form ion pairs in sea water to produce an  $\text{NaCl}^\circ$  complex. This possibility has interesting potential for the Orca Basin since the ionic balance percentages of  $\text{Na}^+$  and  $\text{Cl}^-$  are significantly higher than sea water (Table 18). Pytkowicz and Atlas (1975) have shown that knowledge of variations in solution composition are sufficient to predict changes in buffer intensity which reflect variations in apparent dissociation constants of the carbonate system. Thus the relative percentages of cations and anions in the Orca Basin, as well as the total ion concentration, play an important role in deciphering carbon dioxide interactions in the brine.

Table 18. Ionic balance percentages and total ionic strength for standard sea water (SSW = 35‰ salinity), Dead Sea brine (DSB = 299‰), Orca Basin brine (OBB = 258‰), Red Sea brine (RSB = 256‰), and East Flower Garden brine (EFG = 218‰).

Brine	Na <sup>+</sup>	K <sup>+</sup>	Mg <sup>++</sup>	Ca <sup>++</sup>	Cl <sup>-</sup>	SO <sub>4</sub> <sup>=</sup>	HCO <sub>3</sub> <sup>-</sup>	I
SSW	77.7	1.66	17.3	3.3	90.3	9.34	0.36	0.52
DSB	28.2	2.88	55.0	13.9	99.7	0.25	0.06	
OBB	96.2	0.39	2.01	1.32	98.1	1.77	0.12	4.10
RSB	91.9	1.26	1.51	5.36	99.6	0.36	0.05	6.36
EFG	95.1	0.26	2.42	2.22	97.5	2.43	0.06	

### Alkalinity Considerations

In describing the geochemistry of carbon dioxide in aquatic systems, three parameters must be considered: pH, total CO<sub>2</sub>, and alkalinity. While pH and total CO<sub>2</sub> can be directly measured, alkalinity is a more difficult to estimate. In sea water, the total or titration alkalinity is the result of anions of various weak acids which are not charge balanced with hydrogen ions. The major acids to be considered are carbonic and boric, since their concentrations are normally much larger than the secondary group: phosphoric, silicic, sulfuric, and hydrofluoric acids. Only weak acids with dissociation constants smaller than the first dissociation constant of carbonic acid are included in the titration alkalinity (Edmond 1972). (Throughout this chapter, the dissociation constant will be understood to mean the apparent dissociation, K', which has the activity coefficients included.) For normal sea water, the titration alkalinity is described as:

$$\begin{aligned} \text{TA} = & [\text{HCO}_3^-] + 2[\text{CO}_3^{2-}] + [\text{B}(\text{OH})_4^-] + [\text{Si}(\text{OH})_3\text{O}^-] \\ & + 2[\text{HPO}_4^{2-}] + 3[\text{PO}_4^{3-}] + [\text{OH}^-] - [\text{H}^+] \end{aligned} \quad (27)$$

In anoxic waters, terms representing the nitrogen and sulfur products of anaerobic decomposition ([NH<sub>3</sub>], [HS<sup>-</sup>]) can also be added to the other weak acids which contribute to the total alkalinity (Knull and Richards 1969; Gaines and Pilson 1972). Since the concentrations of weak acids other than carbonic and



boric are relatively low in sea water, they are generally excluded from the total alkalinity. Boric acid contributes only a few percent to the titration alkalinity of normal sea water. However, it needs to be considered for precise alkalinity determinations of normal oceanic samples.

In the Orca Basin, carbonic acid is by far the major contributor to the titration alkalinity. The dissociation constants for acids other than carbonic acid are poorly known even for normal sea water, and are non-existent for hypersaline solutions. Thus, for my discussions of carbonate systems in the Orca Basin, I have ignored all alkalinity components other than carbonic acid and its dissociation products. This omission causes no concern for borate, silicate, or phosphate. It may, however, be an inappropriate assumption with regard to  $\text{NH}_3$ . The ammonia level in the Orca Basin is about  $500 \mu\text{mol}\cdot\text{liter}^{-1}$ , or about 10% of the total carbonate concentration. However, it is not the total species concentration which determines the contribution to alkalinity, but rather the weak acid contribution determined from both the dissociation constants and the total concentration. For ammonia, the contribution to alkalinity would be described as:

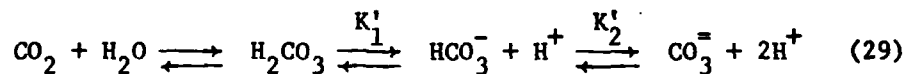
$$A_{\text{NH}_3} = [\text{NH}_3] \frac{K'_{\text{NH}_3}}{a_{\text{H}} + K'_{\text{NH}_3}} \quad (28)$$

where  $A_{\text{NH}_3}$  is the ammonia alkalinity,  $[\text{NH}_3]$  is the total ammonia concentration and  $K'_{\text{NH}_3}$  is the apparent dissociation constant for

ammonia. One can assume  $pK'_{\text{NH}_3} = 9.2$  (Sillen and Martell 1964), which is the dissociation constant for infinite dilution, since no equilibrium values are available even for sea water. Using the pH of the Orca Basin (6.83), it can be calculated that the  $500 \mu\text{mol}\cdot\text{liter}^{-1}$  of ammonia in the Orca Basin contributes only  $2.13 \mu\text{eq}\cdot\text{liter}^{-1}$  to the alkalinity. Millero et al. (1979) calculated  $A_T$  to be  $4750 \mu\text{eq}\cdot\text{liter}^{-1}$ , thus making the ammonia contribution insignificant - if  $K'_{\text{NH}_3}$  is correctly assumed. For the anoxic water of the Pettaquamscutt River, Gaines and Pilson (1972) assumed that the total dissolved ammonia was equal to its alkalinity contribution. This assumption would require  $pK'_{\text{NH}_3}$  to be around 6.9 and may be inappropriate. Yet, the dissociation constant for ammonia is unknown. There is even some confusion as to whether the ammonia contribution should be considered a weak acid (Gieskes 1974) or a weak base (Knull and Richards 1969). Considering the uncertainty as to how to treat the ammonia contribution to the alkalinity, I have chosen to disregard it.

#### Carbonate System Interactions

Carbon dioxide which enters oceanic waters from the atmosphere, or from decomposition processes, participates in equilibria which may be summarized by a generalized equation:



Associated with each step in this series is an equilibrium constant which can be expressed in terms of thermodynamic activities of the participating species. The basic equations necessary for quantifying the interactions of carbon dioxide with its parent solution are (Skirrow 1965):

$$K'_1 = \frac{a_{H^+} [HCO_3^-]}{[H_2CO_3]} \quad \text{First apparent dissociation constant} \quad (30)$$

$$K'_2 = \frac{a_{H^+} [CO_3^{2-}]}{[HCO_3^-]} \quad \text{Second apparent dissociation constant} \quad (31)$$

$$A_c = [HCO_3^-] + 2[CO_3^{2-}] \quad \text{Carbonate alkalinity} \quad (32)$$

$$C = [H_2CO_3] + [HCO_3^-] + [CO_3^{2-}] \quad \text{Total dissolved carbon (refers to inorganic carbon)} \quad (33)$$

$$A_B = TB \frac{K'_B}{a_{H^+} K'_B} \quad \text{Borate alkalinity} \quad (34)$$

where primes indicate that the constants are apparent dissociation constants, brackets designate molar concentrations and  $a_{H^+}$  is the activity of the hydrogen ion (see Table 19 for complete notations).

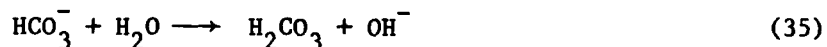
A detailed knowledge of the carbonic acid equilibrium system is of considerable practical importance. In principle, it allows the concentrations of various species (e.g.  $HCO_3^-$ ,  $CO_3^{2-}$ ,  $H_2CO_3$ , and total  $CO_2$ ) to be calculated by measuring any two of the parameters

Table 19. Notation for carbonate calculation.

$a_H$	activity of hydrogen ions, defined by $\text{pH} = -\log a_H$
$K_1$	first apparent dissociation constant of carbonic acid and $\text{p}K_1 = -\log K_1$
$K_2$	second apparent dissociation constant of carbonic acid and $\text{p}K_2 = -\log K_2$
$K_B$	first apparent dissociation constant of boric acid and $\text{p}K_B = -\log K_B$
$C$	total dissolved inorganic carbon
$A_0$	titration alkalinity, initial
$A_c$	carbonate alkalinity
$T_B$	Total dissolved boron concentration
$A_B$	borate alkalinity
$\gamma_{H^+}$	activity coefficient of the hydrogen ion where $[\text{H}^+] = \frac{a_H}{\gamma}$
$N_a$	normality of HCl used in titrations
$A_t^n$	titration alkalinity at the nth step of an HCl titration

pH,  $p\text{CO}_2$ , total  $\text{CO}_2$  or alkalinity (Park 1969). In a brine system, however, the situation is more complicated, since such calculations require the knowledge of the apparent dissociation constants for each reaction. While these dissociation constants have been well established for normal sea water at various temperatures and salinities (Mehrbach et al. 1973), brine values vary depending on both the ionic composition and total ionic strength.

Few brine carbonate systems parameters have been studied in detail. Amit and Bentor (1971) performed a series of fresh water dilutions on the dead Sea brine. They noted that the pH of the brine increased with dilution - a phenomena not found in solutions devoid of  $\text{HCO}_3^-$  salts. The pH-increase with decreasing salinity was attributed to depression of bicarbonate dissociation in strong brines. Upon dilution, the bicarbonate dissociates, hydroxide ions are formed and the pH increases



Their dilution curves (given in Fig. 38) provide a good comparison for the Orca Basin. The Dead Sea brine and the Orca Basin brine differ in several ways. The Dead Sea brine is a chloride brine characterized by relatively high concentrations of calcium and magnesium, low sodium and very low sulfate and inorganic carbon concentrations (Table 3). The Orca Basin brine is a sodium chloride saturated sea water with slightly higher sulfate and

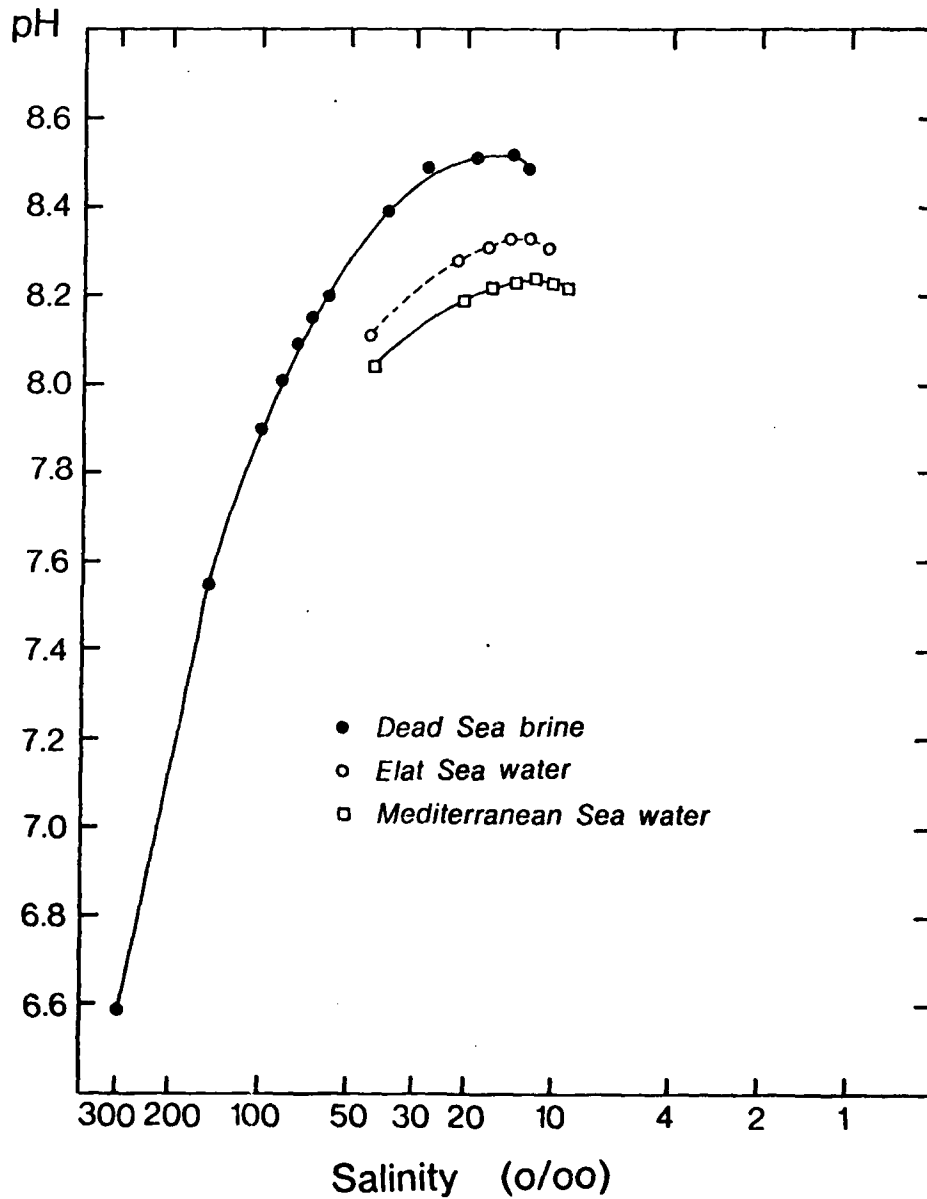


Fig. 38. pH versus log salinity dilution curves for the Dead Sea brine and two other waters (after Amit and Bentor 1971).

calcium, a deficiency in magnesium and about twice the total inorganic carbon as sea water. Thus two differences which will cause varying carbonate interactions between these two brines are the differences in ion interactions and in the total  $\text{CO}_2$  levels.

Sass and Ben-Yaakov (1977) have done a thorough study of the carbonate system in the Dead Sea brines. They developed a generalized method to determine the alkalinity, total carbonate and dissociation constants of any system. Applying this method to the Dead Sea brines, they observed that as the salt content increases, the dissociation constants also increase, resulting in a consequential pH decrease. Their work confirmed the earlier conclusions of Amit and Bentor (1971) in this regard. More importantly, their results showed that the chemistry of carbonate system in hypersaline waters differ only in degree of interactions and not in nature from the carbonate chemistry in less saline waters. Thus, the carbonate system equations that have been developed to describe sea water carbonate equilibria can also be applied to hypersaline waters such as the Orca Basin brine.

The Orca Basin carbon dioxide system was investigated using both the pH-dilution process of Amit and Bentor (1971) and the titration techniques used by Sass and Ben-Yaakov (1977). Field measurements were made of total carbon dioxide and pH. Laboratory titrations of brine with HCl and  $\text{NaHCO}_3$  are also used to unravel the carbonate system interactions. Treatment of the data reflects

a desire of the author to make comparisons between the Orca Basin brine, the Dead Sea brine, and deep Gulf sea water. For this reason, the calculation methodology involved is parallel to that of Sass and Ben-Yaakov (1977).

### Methods

#### Sampling

Sampling requirements for total  $\text{CO}_2$  are less stringent than for other Orca Basin gases. This is true since the  $\text{CO}_2$  in the brine is mainly tied up as bicarbonate and carbonate. Less than one percent of the effervescing gas from the brine was  $\text{CO}_2$ . Brine samples were taken on several cruises for pH, total  $\text{CO}_2$ , and alkalinity analysis. Both Niskin and Nansen bottles were used with no obvious systematic variations between bottle types. pH samples were transferred by gravity flow to 125 ml hard plastic bottles which were allowed to overflow and then sealed with rubber stoppers. pH values were measured (by Tonalee Carlson) as soon as the brine had come to thermal equilibrium at 25°C.

Brine was sampled for total  $\text{CO}_2$  analysis using 12 ml plastic syringes attached to the Nansen or Niskin bottles with tygon tubing. A three-port Luer-tip stopcock was used to flush the syringe so an air free sample could be obtained. Some samples were also taken on cruise EN-32 using the all glass sample containers shown in Fig. 21. All samples were kept refrigerated



and in the dark until analysis aboard ship - usually within 12 hours.

Alkalinity analyses were performed in the laboratory, rather than aboard ship. Samples for alkalinity titration were collected in 300 ml glass bottles with plastic-on-metal crimp shut tops. Each bottle was filled and about 100 ml of brine was allowed to overflow. 20 ml of brine was decanted and one ml of a saturated  $\text{NaN}_3$  solution was added to each bottle to inhibit bacterial removal of  $\text{CO}_2$ . Samples for alkalinity titration were stored at  $4^\circ\text{C}$  and in the dark until analysis in the laboratory at Texas A&M University.

#### Analytical Techniques

pH. The pH of the Orca brine was measured at  $25^\circ\text{C}$  using Beckman glass electrodes in conjunction with a Radiometer Model PHM64 pH-millivolt meter. Millivolt measurements were made and converted to pH by making the same measurements on three different National Bureau of Standards (NBS) buffers of known pH, and calculating the brine pH using linear regression. Pykowicz et al. (1974) have shown that operationally defined hydrogen ion activity measurements ( $\text{pX}$ , where  $X = \text{ka}_\text{H}$  and  $k$  represents the effects of liquid junction and asymmetry potentials) compare well with pH, since  $k$  is almost unity.

The problem of electrode junction potential should be briefly mentioned since previous studies (Ben-Yaakov and Sass 1977) had

questioned the validity of pH measurements in hypersaline waters of the Dead Sea brine (Amit and Bentor 1971). Like this study, Amit and Bentor (1971) used conventional pH electrodes with inherent liquid junction potentials. Ben-Yaakov and Sass (1977) constructed a pH electrode without a liquid junction and made independent measurements of the pH of synthetic Dead Sea brine. Their measurements were strikingly similar to the values reported by Amit and Bentor (1971). Their verification confirmed that the potential problem resulting from the liquid junction in conventional pH electrodes is not a significant factor in making pH measurements in hypersaline solutions.

Total carbon dioxide. A gas chromatographic system which uses the on-line gas stripping techniques of Swinnerton et al. (1962) was used for total CO<sub>2</sub> analysis. A schematic of the system is shown in Fig. 39. One ml of concentrated phosphoric acid was injected via a septum (S) into the glass stripping chamber (GS). After all air had been stripped from the acid, a 6 to 12 ml brine sample was injected into the stripper where the bicarbonate and carbonate are converted to gaseous CO<sub>2</sub>. The CO<sub>2</sub> is swept from the stripper by a helium carrier gas flowing at 20 ml·min<sup>-1</sup>. The gas is dried using either a 20 mesh drierite or 40/60 mesh silica gel drying column (DC) and passed to a 40/60 mesh silica gel separation column (SiGel, 1.0 m x 0.64 cm O.D. stainless steel) at 50°C for separation of air and CO<sub>2</sub>. The gas chromatograph used

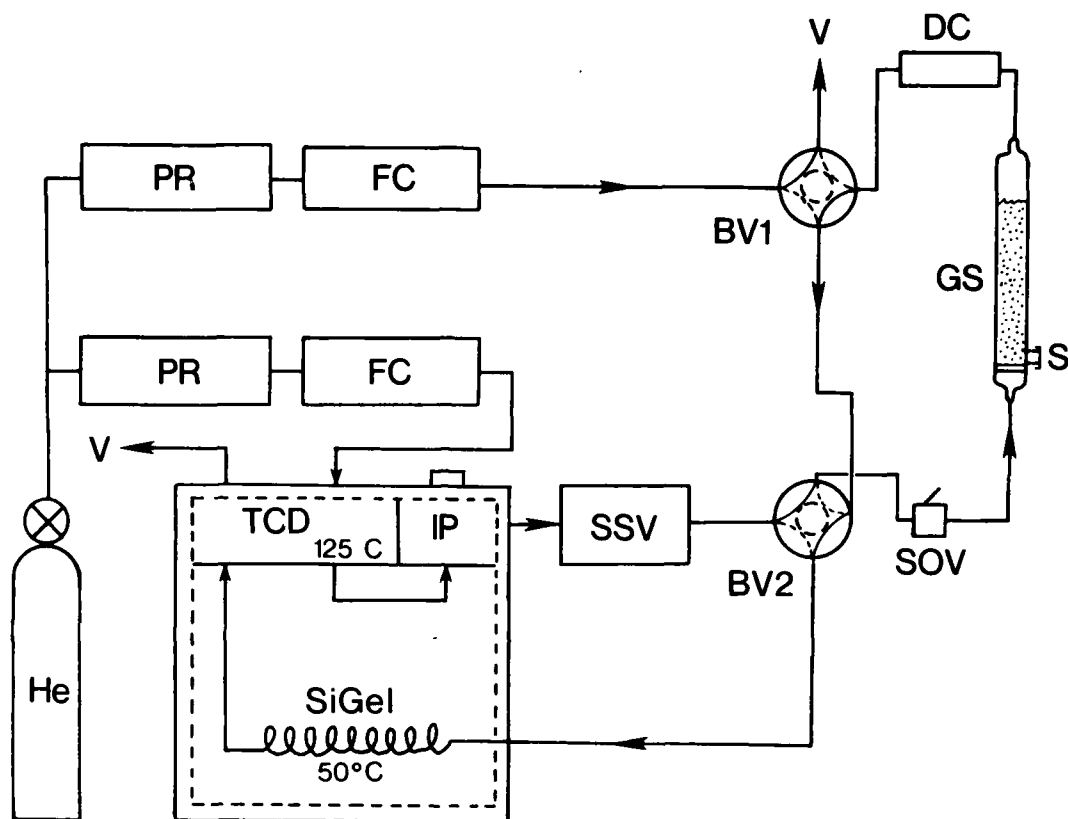


Fig. 39. Schematic presentation of the gas chromatographic system used to measure total carbon dioxide in the Orca Basin brine. Acid is injected into the gas stripping chamber (GS) via a rubber septum (S) and followed by injection of the brine sample. The helium (He) carrier gas flow forces the carbon dioxide from the chamber through a drying column (DC) and into the gas chromatograph. The carbon dioxide is separated from air in the sample by a silica gel column (SiGel) thermostated at 50°C. The carbon dioxide is measured with a thermal conductivity detector (TCD) operating at 125°C. Standard gas (pure carbon dioxide) is introduced into the system via a 6-port standard sample valve (SSV) and analyzed in the same manner as the water samples.

was a Hewlett-Packard 5830 instrument equipped with a thermal conductivity detector operated at 125°C. Baseline stability and peak shape were monitored using the printer-plotter associated with the unit, and peak area integration was performed by the microprocessor which controls the gas chromatograph. Matheson bone-dry (99.7%) carbon dioxide gas was used as a standard. Standard loop volumes of the pure CO<sub>2</sub> were injected through the stripper for calibration of the system. Calculations of total CO<sub>2</sub> were made using Eq. 14 and the appropriate data for carbon dioxide.

The sample was injected into the gas stripper in either of two ways. Syringe samples were injected directly into the stripper through a septum using a 20-gauge needle. Brine collected in the glass sample containers (Fig. 21) were flushed into the stripper by diverting the helium stream through the samples using their two 3-port stopcocks (see Fig. 21). Before the stopcocks were turned to divert the helium flow through the sampler, the carrier gas flow was purged through the system for five minutes to remove all contaminating air and carbon dioxide. No significant differences were found between either type of sample (syringe or glass container), thus confirming that little CO<sub>2</sub> is lost during the sample collection or by diffusion through the plastic syringe.

Dilution techniques. The dilution agent in all experiments described in this work was distilled water. The water used was stirred in contact with the atmosphere for 24 hours which was long

enough to reach  $\text{CO}_3^{=}$  -  $\text{HCO}_3^-$  -  $\text{CO}_2$  equilibrium with  $\text{CO}_2$  in air. All dilutions were weight dilutions and experiments were carried out at room temperature ( $-23^\circ\text{C}$ ) with the brine and distilled water at the same temperature. Two sets of brine samples were diluted. One set contained preserved brine which was opened to the air immediately prior to dilution. These samples had a total  $\text{CO}_2$  concentration of  $4.41 \text{ mmol}\cdot\text{liter}^{-1}$ . A second set of samples were opened and equilibrated with the atmosphere with stirring for 24 hours. The total  $\text{CO}_2$  of these samples had been reduced to a constant  $3.44 \text{ mmol}\cdot\text{liter}^{-1}$  after 24 hours of equilibration. Total  $\text{CO}_2$  analyses were run on several of the dilutions from both sets to examine whether the total  $\text{CO}_2$  dilution curve followed the predicted dilution relationship. pH measurements were made on diluted brine samples over the range 262‰ to 7‰. For comparison, Gulf of Mexico surface and deep water samples were diluted from 35‰ down to 4‰ salinity with concurrent pH measurements at each dilution. pH measurements on the diluted samples were made after the brine and dilution water had been mixed together for three minutes. The instrumentation and techniques used for pH determinations were the same as those described in the earlier section on pH measurements.

Titration with HCl. The sample titration with HCl is performed using techniques similar to Edmond (1970). A 187 ml volume of brine was titrated using 1.0 N HCl in a closed titration flask

with constant stirring. Titrations were carried out in a thermostated water bath at 25°C. Acid was delivered through an immersed capillary tip of a microburette (Guilmont Model S4200A), accurate to 0.001 ml. The pH of the titrated solution was monitored using the techniques described previously (p. 197). All inlets into the sample chamber were air tight and the sample filled all cavities - thus preventing CO<sub>2</sub> loss during the titration. In their study of the Dead Sea brines, Sass and Ben-Yaakov (1977) performed their HCl titration on 50 ml samples open to the atmosphere. They experienced some loss of gaseous CO<sub>2</sub> during their titration and could not accurately calculate the total CO<sub>2</sub> concentrations from their results. Such a problem was avoided here by performing the HCl titrations in a closed vessel.

NaHCO<sub>3</sub> titration. Orca Basin brine samples and 50% (w/w) dilutions of brine with distilled water were titrated with 0.5N NaHCO<sub>3</sub>. Titrations were carried out at room temperature on a 50 ml sample continuously stirred in a 100 ml Pyrex beaker open to the atmosphere. Titrant was added to the beaker using the same burette used for the HCl titration. pH was measured as described above. A total of 5 ml of NaHCO<sub>3</sub> was added to the brine in increments of 0.25 ml. pH was measured one minute after the addition of each aliquot, a time sufficient to allow equilibrium to be reached.

## Model Calculations

## HCl Titration: General Equation

In any chemical system, the total (or titration) alkalinity is equal to the sum of the titratable bases available, minus the available hydronium ion. In oceanic waters, the major components of total alkalinity are the carbonate alkalinity ( $A_c$ ), borate alkalinity ( $A_B$ ) and hydroxide ion ( $OH^-$ ). The most general form of the total alkalinity equation is given as:

$$A_T = A_c + A_B + [OH^-] - [H^+] \quad (36)$$

The  $[OH^-]$  concentration of sea water is low (relative to  $A_c$  and  $A_B$ ) and is generally ignored. Thus Eq. 37 reduces to

$$A_T = A_c + A_B - [H^+] \quad (37)$$

During HCl titration of a water sample, the addition of  $H^+$  ions alters the charge balance of the system such that Eq. 32 no longer represents the original carbonate alkalinity. Also, as addition of acid occurs, the original alkalinity ( $A_o$ ) and total inorganic carbon (C) decreases due to dilution. At any nth step in the titration with HCl, the total alkalinity at that step,  $A_T^n$ , can be expressed by an equation which considers both the change in the charge balance as well as the dilution effect (mass balance):

$$A_T^n = \frac{A_o V_o - V_a N_a}{V_o + V_a} \quad (38)$$

where  $V_o$  is the initial volume of sample titrated,  $V_a$  is the volume of acid added by all the steps up to and including the nth step and  $N_a$  is the normality of the HCl. By inspection, when  $V_a$  equals zero (before titration begins),  $A_t^n = A_o$ .

Expanding Eq. 38, we can write a two term equation

$$A_t^n = \frac{A_o V_o}{V_a + V_o} - \frac{V_a N_a}{V_o + V_a} \quad (39)$$

such that the separate effects of dilution and titration on the alkalinity during HCl addition can be observed. The first term on the right hand side of Eq. 39 describes dilution, while the second term represents the lowering of the alkalinity by the titration with HCl. Using Eq. 39 and the carbonate system equations given as Eq. 30-34, one can derive a general equation for an acid titration which can be used to determine the alkalinity and the magnitude of dissociation constants involved. In this process, we are ignoring the contributions to alkalinity from species other than carbonate components. Even in normal oceanic waters, borate and hydroxide play only a minor role in determining the total alkalinity. In the Orca Basin, where total carbonate levels are twice that of sea water, the borate and hydroxide would have even a more diminished effect. With these assumptions, Eq. 36 reduces to

$$A_T = A_c - [H^+] \quad (40)$$



and is considered to be the total alkalinity in the following discussion. Effects on Orca Basin alkalinity by other species (e.g.  $\text{H}_2\text{S}$ ,  $\text{NH}_4^+$ ) are also ignored since the dissociation constants for other than carbonate species are not known in hypersaline systems. Secondly, the titrations performed on laboratory samples were devoid of any  $\text{H}_2\text{S}$  (and probably  $\text{NH}_4$ ) since the samples were stored for several months. During initial degassing, most gases - but not total  $\text{CO}_2$  - were lost from the brine due to stripping by the effervescing nitrogen and methane (see Chapter III).

One final identity is needed, before Eq. 39 is transformed into a relationship from which alkalinity and the dissociation constants can be calculated. That identity is the relationship between carbonate alkalinity ( $A_c$ ) and total carbonate (C). By simple algebraic manipulation of Eqs. 30-33, we arrive at the relationship

$$A_c = C \left( \frac{a_H K'_1 + 2K'_1 K'_2}{a_H^2 + a_H K'_1 + K'_2} \right) = C(Q) \quad (41)$$

If we now equate Eq. 39 and 40,

$$A_c - [\text{H}^+] = \frac{A_o V_o}{V_a + V_o} - \frac{V_a N_a}{V_o + V_a} \quad (42)$$

and multiplying by  $(V_a + V_o)$  we have

$$A_o V_o - (V_a + V_o) A_c + (V_a + V_o) [\text{H}^+] = V_a N_a \quad (43)$$

Now, remembering that at any point  $n$  on a titration curve, the total carbonate is going to be diluted such that

$$C_n = C \frac{V_o}{V_a + V_o} \quad (44)$$

Substituting Eq. 44 for  $C$  in Eq. 41, and the combined term into Eq. 43 gives

$$\begin{aligned} A_o V_o - (V_a + V_o) \frac{V_o}{V_a + V_o} C \cdot Q \\ + (V_a + V_o) [H^+] = N_a V_a \end{aligned} \quad (45)$$

where

$$Q = \frac{a_H K'_1 + 2K'_1 K'_2}{a_H^2 + a_H K'_1 + 2K'_1 K'_2}$$

Substituting  $\frac{a_H^+}{\gamma}$  for  $[H^+]$  and dividing through by  $V_o$  gives a complete generalized titration equation

$$A_o - C \cdot Q + \frac{a_H^+}{\gamma} \left( \frac{V_a + V_o}{V_o} \right) = \frac{N_a V_a}{V_o} \quad (46)$$

which can be used to calculate the alkalinity of a sample. Sass and Ben-Yaakov (1977) arrived independently at the same equation

which they used in a different form:

$$\begin{aligned} \frac{1}{V_o} [A_o V_o + \frac{a_H}{\gamma_{H^+}} (V_o + V_a) - N_a V_a] \\ = C \frac{a_H K'_1 + 2K'_1 K'_2}{a_H^2 + a_H K'_1 + K'_1 K'_2} \end{aligned} \quad (47)$$

They have used a two step process to solve Eq. 47 for  $A_o$ ,  $C$ ,  $K_1'$ ,  $K_2'$ , and  $\gamma_{H^+}$ . The first step involves setting the right hand side equal to zero, and then solving the left hand side for  $A_o$  and  $\gamma_{H^+}$  by linear regression.  $A_o$  and  $\gamma_{H^+}$  are then used as constants to solve all of Eq. 47 using a series of guesses for  $K_2'$  and linear regressions techniques to determine the best fit values of  $K_1'$ ,  $K_2'$ , and  $C$ . Their solution technique was applied to the Orca Basin brine titrations. The mathematics of their solution are outlined for completeness in Table 20.

#### $\text{NaHCO}_3$ Titration

Additional knowledge of the carbonate system can also be gained by addition of one individual component to a solution and examining the resultant perturbations. The parameters that can easily be monitored are carbonate alkalinity, total  $\text{CO}_2$  and pH. The interdependence of carbonate alkalinity, total  $\text{CO}_2$  and pH is described by a Deffeyes (1965) diagram showing the variations of alkalinity with input of carbon dioxide at various pH values. Fig. 40 gives an example of a typical Deffeyes diagram for normal salinity sea water. The pH is uniquely determined for each value of total  $\text{CO}_2$  and alkalinity. Also shown on this figure are the relative changes in carbonate alkalinity and total  $\text{CO}_2$  due to the addition of bases or acids, carbon dioxide, bicarbonate, ions, carbonate ions or distilled water. By adding  $\text{NaHCO}_3$  to a solution, both the alkalinity and the total inorganic carbon are increased

Table 20. Sass and Ben-Yaakov (1977) mathematical solution to the general titration formula. Primes on constants are omitted for simplicity.

General equation, LHS = RHS

$$\frac{1}{V_o} \left[ AV_o + \frac{a_H}{\gamma_{H^+}} (V_o + V_a) - N_a V_a \right] = C \frac{a_H K_1 + 2K_1 K_2}{a_H^2 + a_H K_1 + K_1 K_2} \quad (47)$$

STEP 1: At low pH, RHS is negligibly small and can be neglected, thus

$$a_{H^+} (V_o + V_a) = -AV_o \gamma_{H^+} + N_a \gamma_{H^+} \cdot V_a \quad (48)$$

This equation has the linear form  $y = m + Kx$ , where

$$\begin{aligned} y &= a_{H^+} (V_o + V_a) & m &= AV_o \gamma_{H^+} \\ x &= V_a & K &= N_a \gamma_{H^+} \end{aligned}$$

$\gamma_{H^+}$  and A can be determined by linear regression since  $a_{H^+}$  and  $(V_o + V_a)$  change for each titration step.

STEP 2: Define Z as the LHS of the general equation and rearrange

$$\frac{Z(a_{H^+})^2}{2K_2 + a_{H^+}} = CK_1 - K_1 Z \left( \frac{K_2 + a_{H^+}}{2K_2 + a_{H^+}} \right) \quad (49)$$

Using A and  $\gamma_{H^+}$  from Step 1, Z becomes a known quantity. By guessing  $K_2$ , Eq. 49 has a linear form with

$$\begin{aligned} y &= \frac{Z(a_{H^+})^2}{2K_2 + a_{H^+}} & m &= CK_1 \\ x &= Z \left( \frac{K_2 + a_{H^+}}{2K_2 + a_{H^+}} \right) & K &= -K_1 \end{aligned}$$

Using linear regression techniques, C,  $K_1$  and  $r^2$  (coefficient of determination) are determined for each guess of  $K_2$ . By iteration techniques, successive values of  $K_2$  are used until the  $r^2$  value closest to 1.0 is obtained. The calculated  $K_1$  and C at that value of  $K_2$  thus provide the best fit of the titration points to the model.

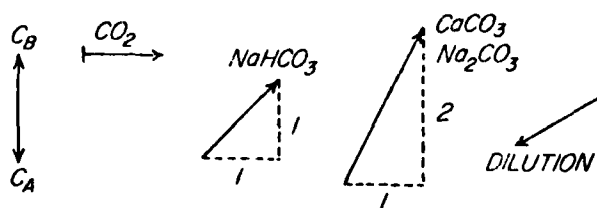
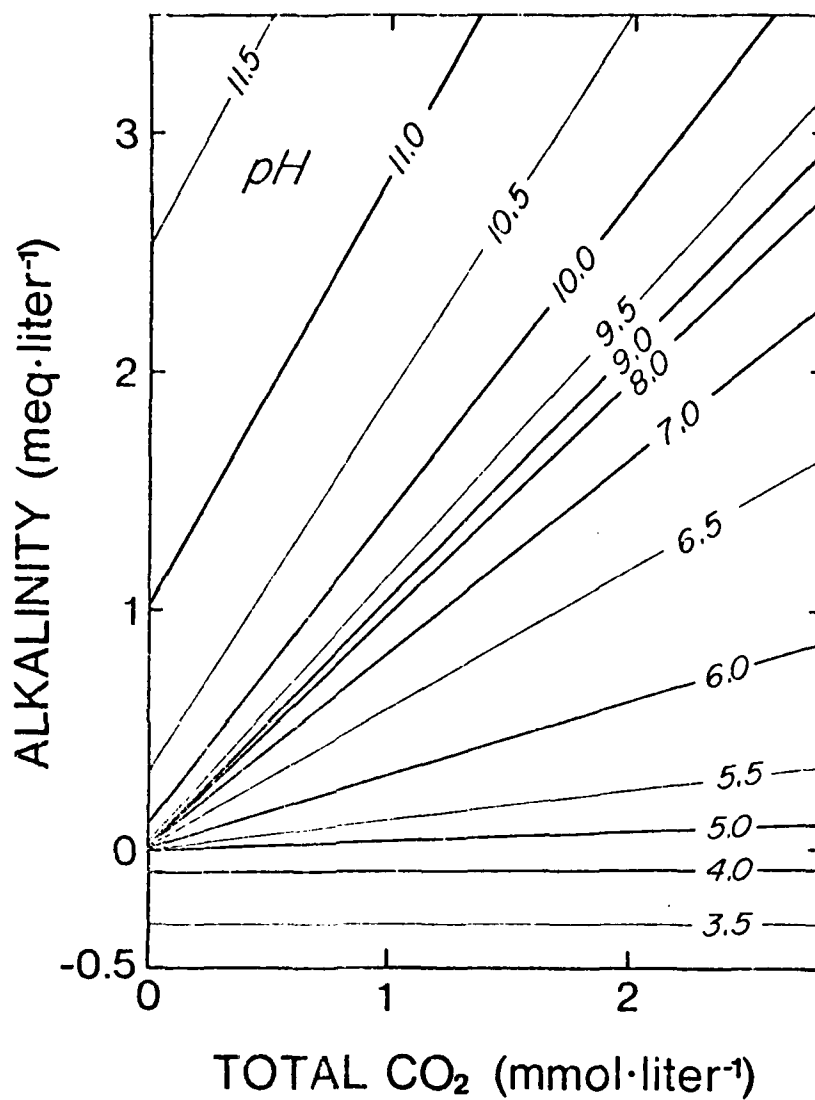


Fig. 40. Typical Deffeyes diagram for sea water (after Deffeyes 1965).

by the same amount. Such a titration thus leaves the ratio of  $A_c/C$  unchanged during the  $\text{NaHCO}_3$  addition. The pH, however, will change during the titration. It will either increase or decrease, depending upon whether the  $A_c/C$  of the sample is greater or less than one. This pH change interpretation can be shown schematically, for any sea water or brine system by a simplified, general  $A_c$  versus C diagram. The diagram (Fig. 41) is derived from Eq. 41 and contains straight pH locus lines for fixed values of  $A_c/C$ . For  $A/C$  equal to one, Eq. 41 reduces to

$$(a_{\text{H}^+})^2 = K_1'K_2' \quad (50)$$

or

$$\text{pH}_{12} = 0.5(\text{pK}_1' + \text{pK}_2') \quad (51)$$

where  $\text{pH}_{12}$  is the mean of  $\text{pK}_1'$  and  $\text{pK}_2'$ . As  $\text{NaHCO}_3$  is added to a brine (or sea water) solution, the pH increase or decrease to a constant saturation pH which approaches the limiting value of  $\text{pH}_{12}$  (Weyl 1961). Thus,  $\text{NaHCO}_3$  titration of a sample will give an independent estimate of  $K_1'K_2'$ . This titration will also allow one to determine whether alkalinity is greater or less than total inorganic carbon. This determination can be visualized by examining dashed lines 1 and 2 in Fig. 41. These lines represent a  $\text{NaHCO}_3$  titration curve of a sample. pH lines (solid) increase from lower right to upper left. As the sample is titrated,  $A_c$  and C increase

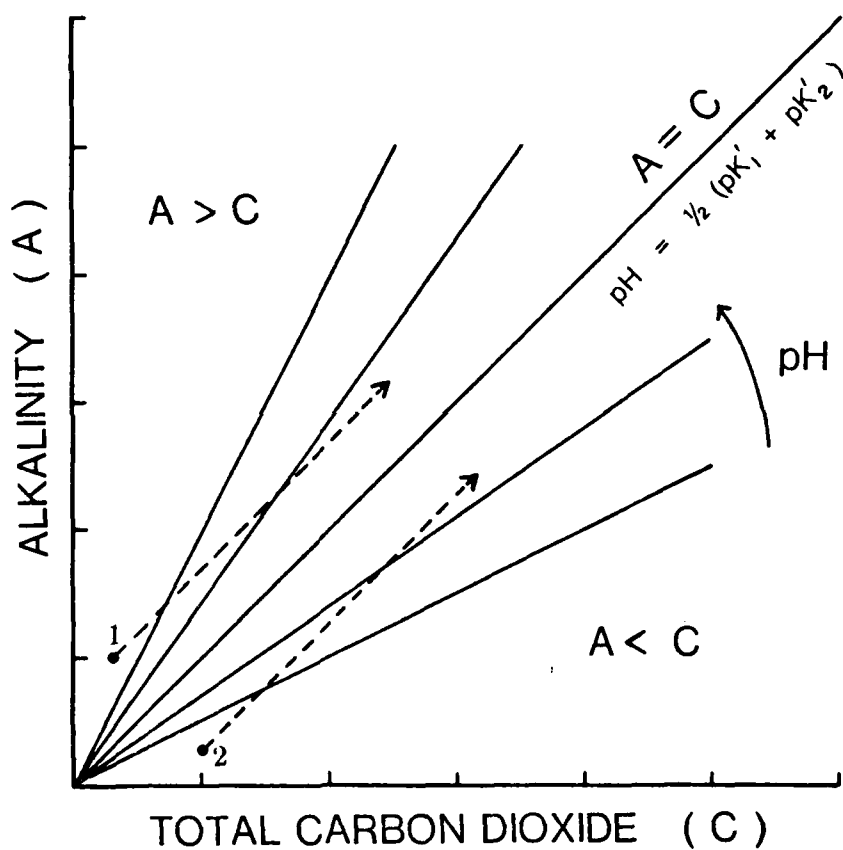


Fig. 41. Theoretical alkalinity (A) versus total  $CO_2$  (C) plot showing lines of constant pH. Lines 1 and 2 represent the results of a  $NaHCO_3$  titration with  $A > C$  (1) or  $A < C$  (2).

at the same rate. If  $A = C$ , no pH change is observed. For  $A < C$  (line 1), pH increases and if  $A > C$  (line 2), pH decreases during the titration, at least up to bicarbonate saturation. Thus, a  $\text{NaHCO}_3$  titration of a sample yields independent information as to whether the alkalinity of a sample is greater than or less than the total  $\text{CO}_2$ .

#### Buffer Intensity

The buffer intensity or buffer capacity,  $B$ , of a solution is the amount of acid or base required to change the pH by one unit. Since Eq. 41 represents the carbonate alkalinity-total  $\text{CO}_2$ -pH relationship of a solution, it is the starting point for determining the buffer intensity. By taking the derivative of Eq. 41 with respect to  $a_{\text{H}^+}$ , one arrives at an expression for the buffer intensity of a solution (Pytkowicz and Atlas 1975)

$$B = 2.303 \cdot C \cdot K_1' a_{\text{H}} \frac{a_{\text{H}}^2 + 4K_1' a_{\text{H}} + K_1' K_2'}{(a_{\text{H}}^2 + K_1' a_{\text{H}} + K_1' K_2')^2} \quad (52)$$

The plus signs on the hydrogen activity are omitted here for simplicity. Thus for two systems, the buffer intensity (capacity) of each system can be compared for any pH values, if the total  $\text{CO}_2$  and equilibrium constants are known.



## Results and Discussion

### pH and Total CO<sub>2</sub> Relationships

In the aerobic Gulf of Mexico waters, in the region of the Orca Basin, profiles of pH and total carbon dioxide (Fig. 42) are typical of the oceanic environment. Surface pH is about  $8.2 \pm 0.1$  and decreases with depth to slightly less than 7.8 at 500 meters, before increasing to 7.9 in the deeper water. The total CO<sub>2</sub> depth profile is almost a mirror image of the pH distribution. Total CO<sub>2</sub> increases with depth are a major cause of the decreasing pH. These increases in Gulf CO<sub>2</sub> levels result from the aerobic respiration of marine organisms (see Table 12), which causes an oxygen minimum where total CO<sub>2</sub> is highest and pH is lowest.

Below 2100 m in the Orca Basin, the total CO<sub>2</sub> again increases and pH decreases as oxygen levels begin to decrease. Fig. 43 shows the pH and total CO<sub>2</sub> profiles through the Orca Basin interface into the brine. pH decreases to 6.83 in the high salinity brine, while total CO<sub>2</sub> increases to an average of about 4.45 mmol·liter<sup>-1</sup>. Sackett et al. (1979) have shown that the excess CO<sub>2</sub> in the brine (relative to deep Gulf values) results from introduction of isotopically light, biogenic CO<sub>2</sub> into the Orca Basin water. The pH decrease reflects this increase in CO<sub>2</sub>, but the change in pH may also be affected by the total ion composition of the brine. This possibility will be discussed in the next section.

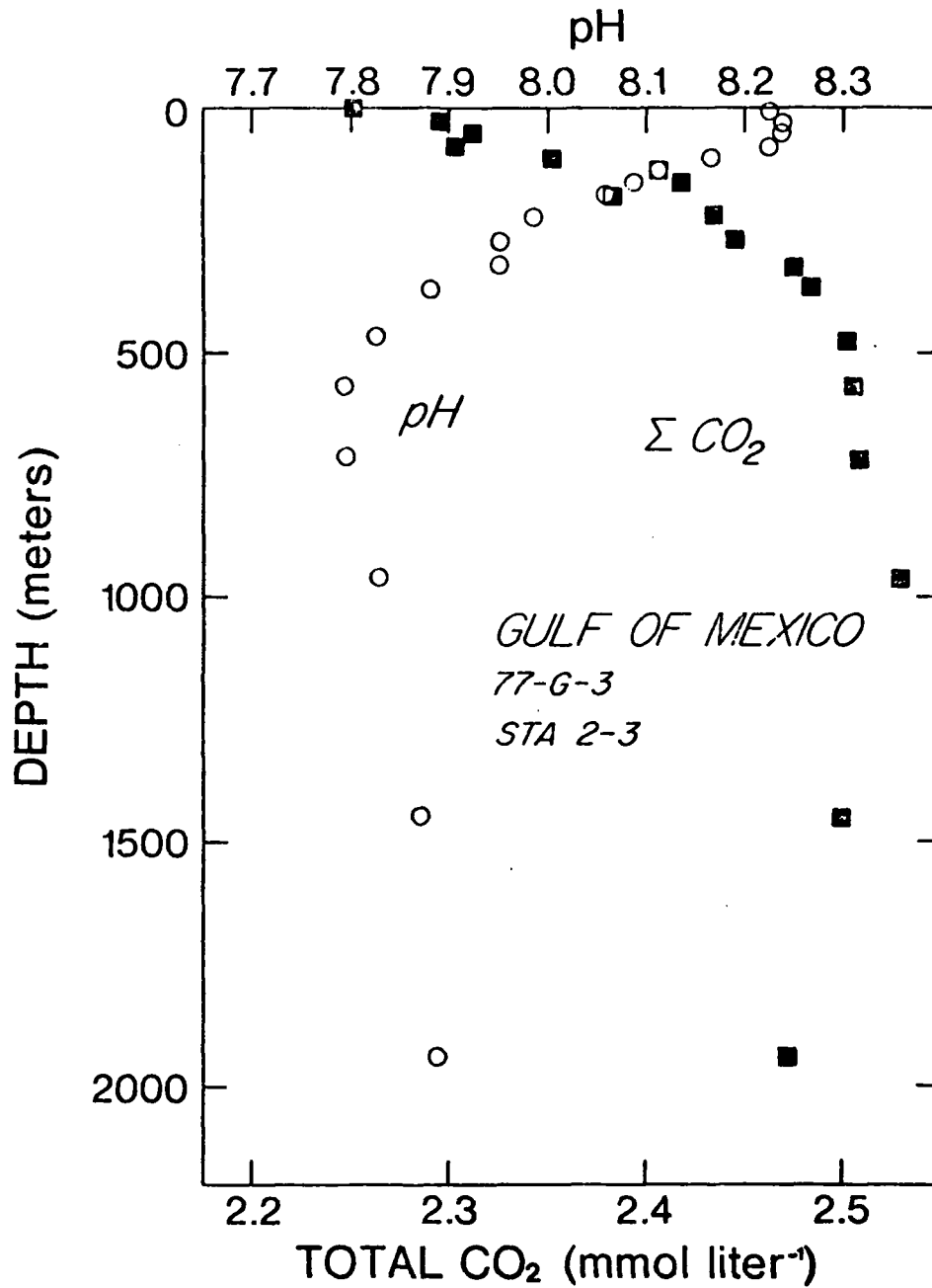


Fig. 42. pH and total carbon dioxide depth distribution in the northwestern Gulf of Mexico.

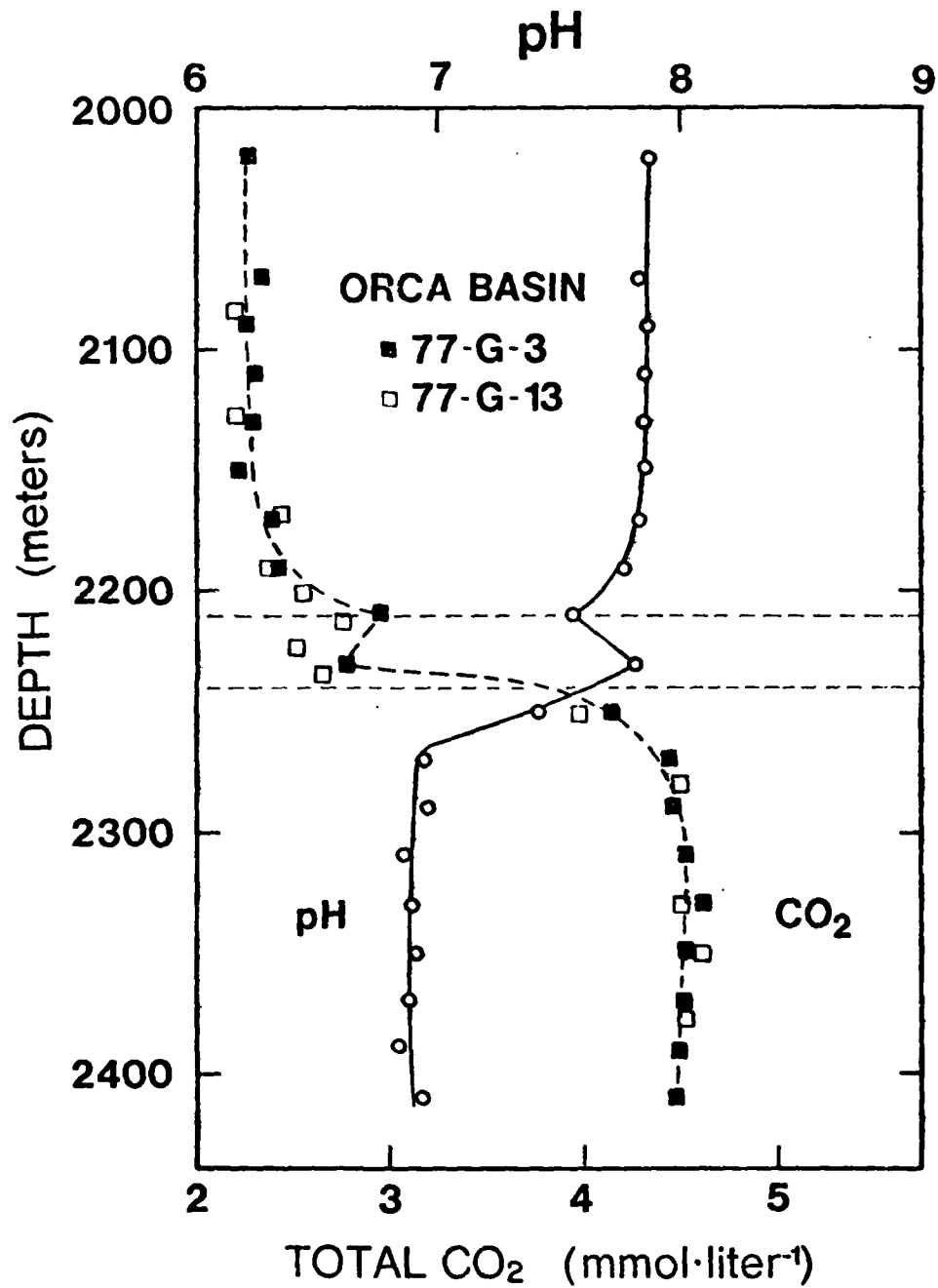


Fig. 43. pH and total carbon dioxide profiles through the Orca Basin brine interface and into the high salinity brine.

In the interface region, there is an obvious discontinuity in both the pH and total CO<sub>2</sub> depth distribution. Both parameters follow a predictable pattern (based on O<sub>2</sub> consumption and CO<sub>2</sub> production) down to about 2210 m, then change sharply, before continuing their expected increase (CO<sub>2</sub>) or decrease (pH) into the high salinity brine (see Fig. 43). The total CO<sub>2</sub> profile shows only a one point maximum on cruise 77-G-3, but the inverse relationship with pH contributes to the reliability of this data point. Sackett et al. (1979) also observed the same discontinuity in their total CO<sub>2</sub> data at this level. However, they chose to ignore the single point maximum and they drew a smooth curve through the other data points in this region. Data from cruise 77-G-13 is more detailed in this zone (open squares in Fig. 43). These data confirm the CO<sub>2</sub> discontinuity that was observed along with pH and oxygen anomalies in this zone.

A detailed plot of pH and oxygen with depth through the interface region is shown in Fig. 44. Data from two different years are plotted for pH. It is apparent that the discontinuity in the pH-depth profile occurs in transition zone T3, the zone where salinity increases from 50-70‰ and oxygen concentration is low, but present. Thus, the unusual oxygen step-region has a characteristic pH profile change associated with it, along with the large temperature gradient shown in Fig. 10. The decreasing pH down to 2210 m reflects increasing production of CO<sub>2</sub> through the

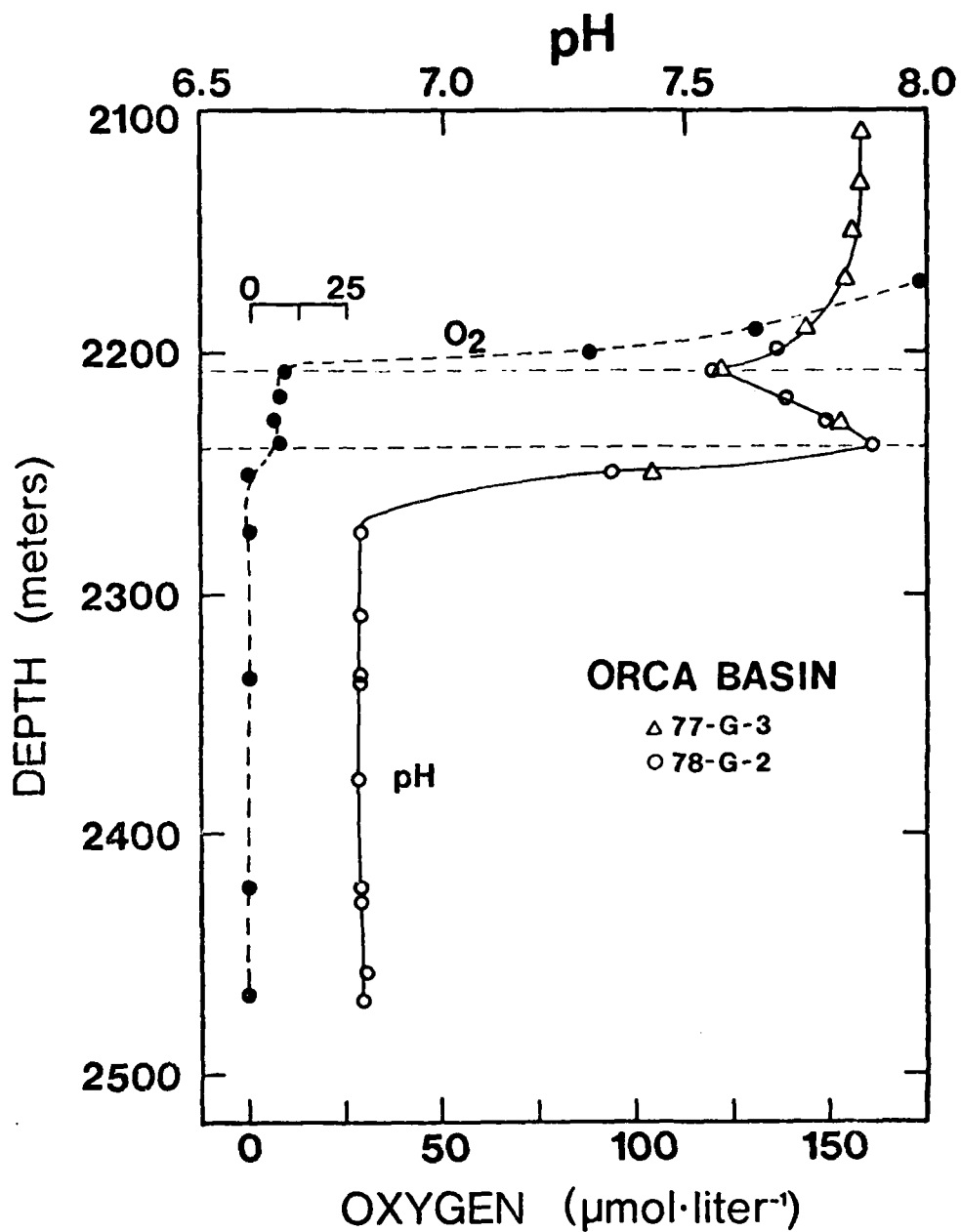


Fig. 44. Detailed plot of pH and dissolved oxygen through the interface region of the Orca Basin.

increasing density region. The  $\delta^{13}\text{CO}_2$  data of Sackett et al. (1979) support this concept, with increasingly lighter (isotopically)  $\text{CO}_2$  found as the interface is traversed.

The increase in pH and decrease in total  $\text{CO}_2$  through the oxygen step region (transition zone T3) are not completely understood. The factors acting to cause this discontinuity must be a complex of mixing and biological processes. Both temperature and salinity change rapidly in this region. Physical mixing processes may be important, but significant transport of carbon dioxide from the brine is questionable, since transport through the interface probably occurs only by molecular diffusion. Another, probably more significant process would be degradation of organic material at the top of this low oxygen region. Particles settling on to the interface would undergo decay, mediated by microbial populations which are as high as this interface (LaRock et al. 1979) as at some sediment-sea water interfaces (Zobell 1942). Degradation of organic matter would produce more  $\text{CO}_2$  where the bacteria biomass was highest, at the top of this zone. Decreasing pH would result from this  $\text{CO}_2$  increase. The pH increase might also be affected by ammonia production there, or by some slight diffusion of ammonia into this region from the brine below. Addition of ammonia to the water would cause the pH to increase. However, the magnitude of this effect is unknown. A detailed survey of physical and biological parameters in this low oxygen zone is needed to explain the pH and  $\text{CO}_2$  distributions.

In the high salinity brine, there are no obvious trends in either pH or total  $\text{CO}_2$ . Sackett et al. (1979) also found

relatively constant values for all other carbon budget components in the high salinity brine. They reported a  $\delta^{13}\text{C}$  of the inorganic carbon of  $-6.4 \pm 1.0\text{‰}$ . This negative  $\delta^{13}\text{C}$  value is indicative of biogenic input of carbon dioxide into the brine. They estimated that the biogenic  $\text{CO}_2$  input must have a  $\delta^{13}\text{C}$  of  $-33\text{‰}$  to produce the net  $\delta^{13}\text{C}$  value observed. Average total  $\text{CO}_2$ , pH and  $\delta^{13}\text{CO}_2$  values for several brines are compared in Table 21. With no change in the dissociation constants, Eq. 41 predicts that pH will decrease as total  $\text{CO}_2$  increases. This is indeed the case for the Orca Basin. There is a two-fold increase in  $\text{CO}_2$  levels due to  $\text{CO}_2$  production in the brine. However, with the highest total  $\text{CO}_2$  of all the brines listed in Table 21, the Orca Basin brine also has the highest pH. All three brines have evidence of biogenic  $\text{CO}_2$  input, based on the  $\delta^{13}\text{C}$  data. The higher Orca Basin pH must result from the Orca Basin brine having a higher buffer capacity than either the Dead Sea or Red Sea brines.

If one assumes that the origin of the excess  $\text{CO}_2$  is biogenic, then the major concern becomes the species distribution and effect of this excess  $\text{CO}_2$  in the brine. Kennett and Penrose (1978) have noted that the calcite microfossils in the Orca Basin sediments are well preserved. This effect must result from a change in the carbonate system distribution in the brine. Other (non-brine) Gulf sediments with high  $\text{CO}_2$  levels show no exceptional calcium carbonate preservation. The following sections describe the

Table 21. Total carbon dioxide,  $\delta^{13}\text{C}-\text{CO}_2$  and pH from several brine areas.

Brine	Total $\text{CO}_2$ ( $\text{mmol}\cdot\text{liter}^{-1}$ )	$\delta^{13}\text{C}$ (‰)*	pH
Orca Basin	4.57	-16.4	6.83
Red Sea *			
Discovery Deep	0.60	-16.8	6.5
Atlantis II Deep	2.80	-5.6	5.61
Dead Sea †	2.55	-	6.3

\* Data from Weiss (1969).

† Data from Sass and Ben-Yaakov (1977).



experimental data used explain why the pteropod shells are well preserved in the Orca Basin.

#### Brine Dilution Experiment

The results of the pH-dilution experiment for the Orca brine and Gulf of Mexico sea water are shown in Fig. 45. Four dilution experiments were undertaken, two with brine and two with Gulf of Mexico sea water. In all of the dilutions, the pH increased initially and then decreased after going through a maximum. Amit and Bentor (1971) described this type of pH response with dilution as being due to the "hidden alkalinity" effect. Sass and Ben-Yaakov (1977) noted however, that this response is actually due to the change in the ionic strength of the solution, which in turn causes an increase in the dissociation constants of the carbonate system, thus removing hydrogen ions from solution.

Upon dilution with distilled water, the pH of both Orca Basin brine samples increased progressively. The two brines were prepared differently before the experiment. One sample was stirred for 24 hours before the dilution was begun, while the other was fresh, preserved brine right out of the bottle. The effect of the stirring was to reduce the total carbon dioxide of the brine down from  $4.41 \text{ mmol}\cdot\text{liter}^{-1}$  to  $3.44 \text{ mmol}\cdot\text{liter}^{-1}$ . The brine with the higher total carbonate (closed diamonds in Fig. 45) has an initial pH of about 7.3 and increases only to a pH of about 7.8 at a

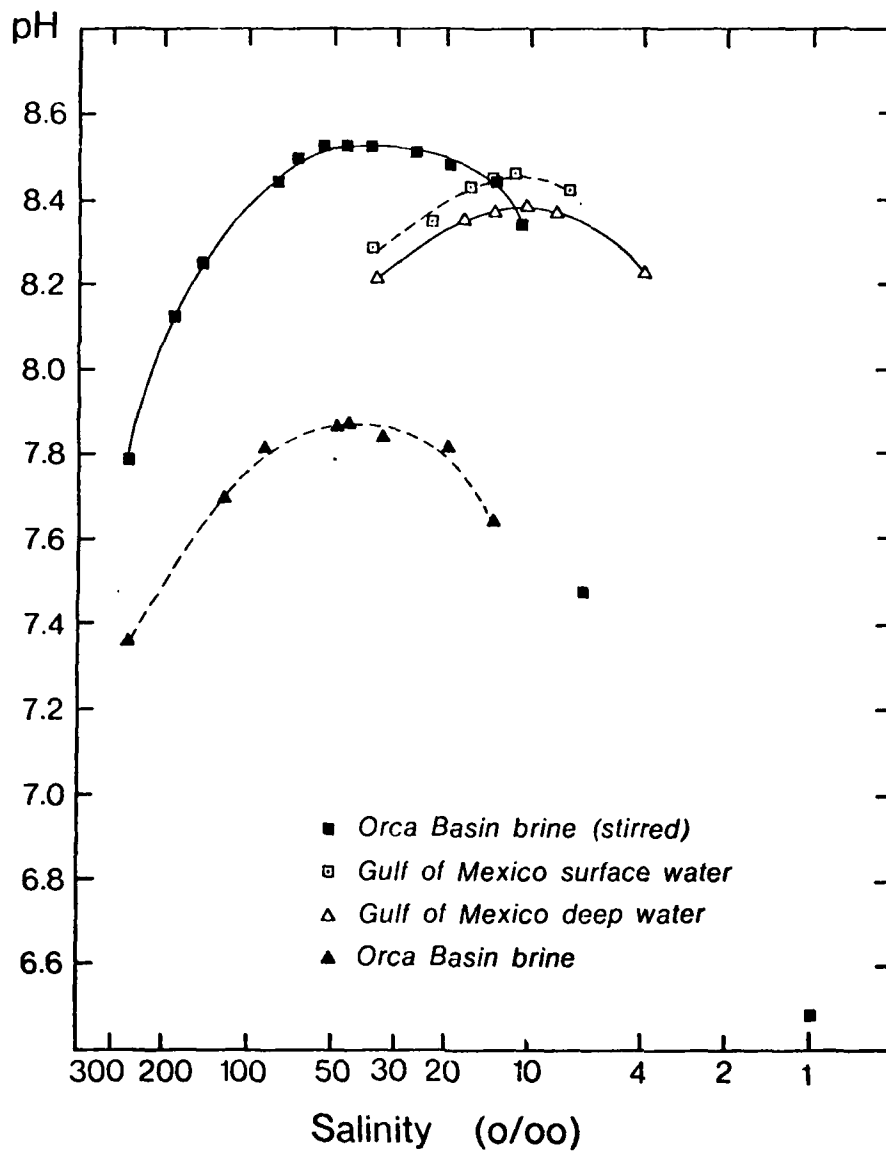


Fig. 45. pH versus log salinity dilution curves for two samples of Orca Basin brine and for surface and deep water from the Gulf of Mexico.

salinity of 35 ‰. The carbonate depleted brine (initial pH = 7.8) reached a pH of 8.5 at 35 ‰ salinity. The differences between the two brine dilution curves are a direct result of one sample having more carbon dioxide.

Fig. 46 shows that relatively little, if any, carbon dioxide was lost during the dilution experiment. The carbon dioxide in the two experiments both show a regular dilution curve, with only the slopes different, due to different initial amounts of carbon dioxide in the brines. Yet the initial carbon dioxide difference makes all the difference in the pH-dilution curves. The brine with the higher total  $\text{CO}_2$  never approximates the pH of the Gulf sea water. The brine with the lower  $\text{CO}_2$  constant has the upper part of its dilution curve in the sea water pH range. The Dead Sea dilution curves of Amit and Bentor (1971), presented in Fig. 38, show a relationship similar to the low  $\text{CO}_2$  curve for the Orca Basin.

The importance of biogenic carbon dioxide to the carbonate system in the Orca Basin is obvious from the pH-dilution results. In the Dead Sea, the surrounding area is fairly desolate and little organic carbon enters the Dead Sea brine. With the absence of biogenic carbon dioxide in the Dead Sea, the total carbonate measures only  $2.5 \text{ mmol} \cdot \text{kg}^{-1}$ , a value close to the levels found in the nearby Elat Sea water (Amit and Bentor 1971). Thus when the Dead Sea brine is diluted down to sea water salinities, the carbonate content is much reduced, and the pH of the diluted brine

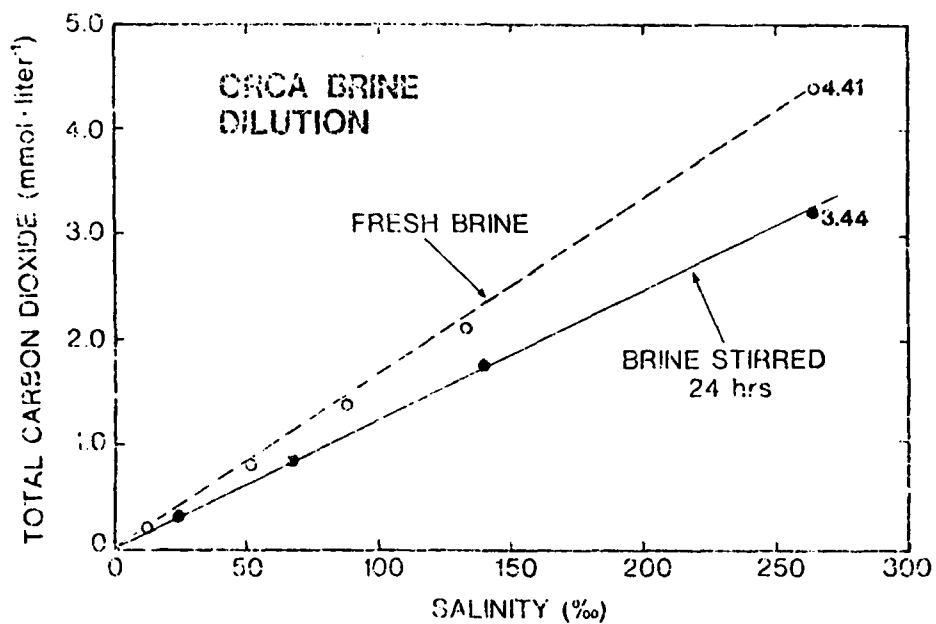


Fig. 46. Change in total carbon dioxide with dilution of Orca Basin brine. These  $\text{CO}_2$  measurements were made on the same samples analyzed for pH and shown in Fig. 45. Total  $\text{CO}_2$  values for each brine sample are shown in units of  $\text{mmol}\cdot\text{liter}^{-1}$ .

exceeds the pH of the Elat Sea water. In the Orca Basin with the high total carbonate, the dilution pH values are lower than the sea water pH values. This effect must result from the higher total  $\text{CO}_2$  of the Orca brine, as well as the fact that the salt did not originate from present Gulf of Mexico sea water.

The pH-dilution curve of the Orca brine is also shifted to the left, compared to the dilution curve for the Gulf of Mexico water. This effect was not observed by Amit and Bentor (1971) who compared the Dead Sea brines to the Elat and Mediterranean Sea waters. This shift means that a higher pH was reached at a higher salinity in the Orca brine, as compared to the Dead Sea brine. This response may be due to the difference in ionic nature of the two brines (see Table 3). The Dead Sea brine is higher in calcium and magnesium and relatively depleted in sulfate. In contrast to the Orca Basin, and in contrast to almost all other known surface water bodies (Nissebaum 1975), magnesium is the dominant cation in the Dead Sea brine.

The pH-dilution curve is controlled predominately by a relaxation of the dissociation constants as more water is added to the brine. The two brines would be expected to respond differently if changes in the dissociation constants were due to the different ionic compositions of the brines. Ben-Yaakov and Goldhaber (1973) noted that sea water dissociation constants of the carbonate system vary in sensitivity to changes in different ions.

They defined a sensitivity parameter for the change in each constant with respect to the major ions. They showed that the apparent constants are especially sensitive to a variation in the concentration of magnesium in solution. Ionic composition, as well as the total ionic concentration, are important in determining the carbonate system dissociation constants. The differences in the ionic balances between the Dead Sea and Orca Basin can be dramatically seen when referenced to the chloride content of each brine. Relative to chloride, and compared with average Gulf of Mexico sea water, Mg and Ca are enriched by a factor of about three in the Dead Sea, but they are depleted by a factor of one-third and one-tenth, respectively, in the Orca Basin (Table 22). Ion complexations due to the higher magnesium and calcium levels in the Dead Sea brine would thus allow the pH levels to increase more slowly with dilution, than in the Orca Basin brine where the concentrations of those important ions is lower.

#### Alkalinity Versus Total $\text{CO}_2$

As was discussed earlier, titration of a bicarbonate solution with one of the carbonate system components can yield useful information as to the species distribution, when the data is interpreted on a Deffeyes diagram. The results of a titration of the Orca brine with 1.0 N sodium bicarbonate solution is shown in Fig. 47. With reference to Fig. 41, the initial increase in brine

Table 22. Ion enrichment factors relative to sea water for calcium and magnesium (referenced to chloride).

Brine	Ca Enrichment	Mg Enrichment
Orca Basin *	0.346	0.105
Red Sea †	1.436	0.078
Dead Sea ‡	3.716	2.828

\* Shokes et al. (1977).

† Brewer et al. (1969).

‡ Amit and Bentor (1971).

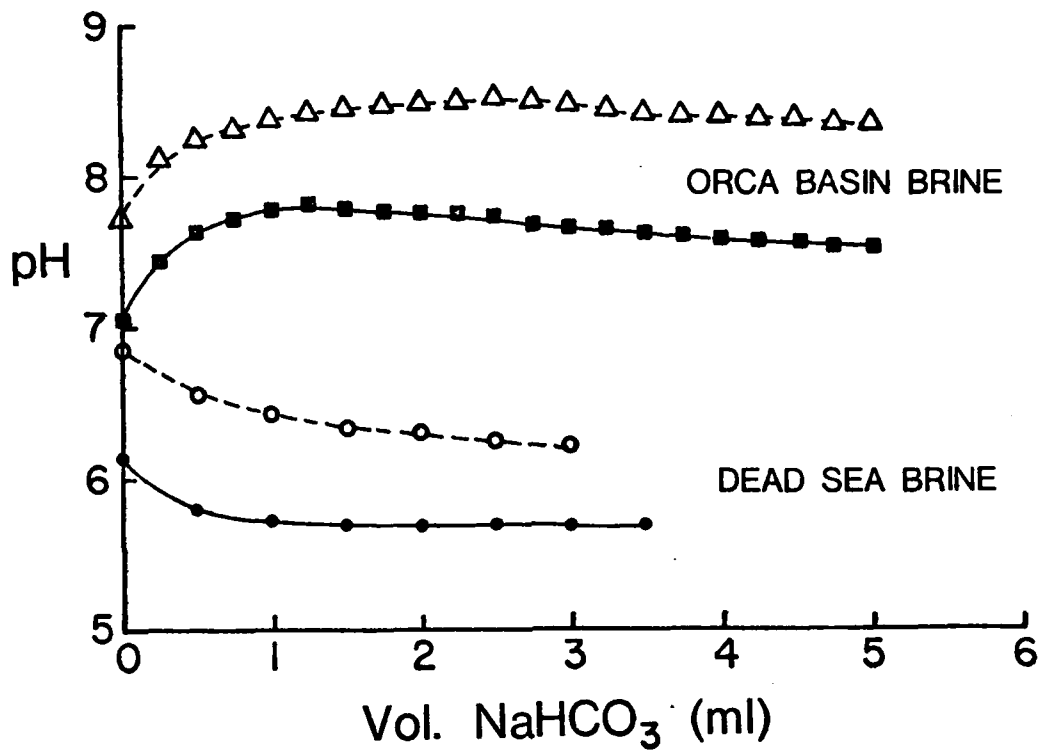


Fig. 47. Change in pH of the Orca Basin brine and Dead Sea brines with addition of 0.5 N NaHCO<sub>3</sub>. Solid symbols are pure brine and open symbols are 50% dilutions (Dead Sea brine data from Sass and Ben-Yaakov 1977).



pH with the addition of the  $\text{NaHCO}_3$  indicates that the total carbonate level in the Orca Basin is higher than the alkalinity. The Dead Sea data of Sass and Ben-Yaakov (1977) are shown for comparison. In the Dead Sea brine, the alkalinity was greater than the total carbon dioxide, as seen by the reduction of pH with the addition of bicarbonate (see Fig. 47).

It should be pointed out here, that the brine that was used for titration with sodium bicarbonate was stored (with sodium azide poisoning) for several months before titration. It is possible that there was some change in the carbonate composition before titration. I make this comment since Millero et al. (1979) have reported that the total alkalinity of the brine is  $4.015 \pm 0.008$   $\text{meq}\cdot\text{kg}^{-1}$  for Orca Basin brine. This value translates to  $4.75$   $\text{meq}\cdot\text{liter}^{-1}$  using a value of 1.185 for the density of the brine at room temperature. This alkalinity value is greater than the  $4.57$   $\text{mmol}\cdot\text{liter}^{-1}$  total inorganic carbon value reported by Sackett et al. (1979) and also higher than the  $4.50$   $\text{mmol}\cdot\text{liter}^{-1}$  determined in this study. It is possible that the ammonia in the water is causing the higher alkalinity that was measured by Millero et al. (1979). Thus the influence of other factors may affect the alkalinity in the Orca Basin, as they do for other anoxic hypersaline waters (Gaines and Pilson 1972).

To confirm the conclusion that A/C was less than unity, several brine samples were left open to the air and stirred for 24

hr to remove  $\text{CO}_2$ . Titration of these samples with 0.5 N  $\text{NaHCO}_3$  produced a titration curve in which the pH decreased with addition of titrant. The lower  $\text{CO}_2$  sample had an alkalinity greater than the total  $\text{CO}_2$  concentration. This titration also showed that the observed pH changes were not due to the brine dilution, since dilution alone, at either high or low total  $\text{CO}_2$  levels, would increase the pH of the brine.

#### Alkalinity in the High Salinity Brine

As discussed in the carbonate model section, using the calculation methods of Sass and Ben-Yaakov (1977) with an acid titration of a solution allows calculation of the dissociation constants as well as alkalinity and total carbonate. A typical titration with HCl of Orca Basin brine is depicted in Fig. 48. Also shown is an HCl titration of deep Pacific Ocean sea water (salinity=34.8‰). The following observations can be made from this titration:

- (1) The pH of the inflection point of the HCl titration is a function of the salinity of the solution.
- (2) The shape of the HCl titration curve for the Orca brine is similar to the sea water curve, but shifted downward (to lower pH) due to the high carbonate content of the brine and the difference in dissociation constants resulting from the high salinity.

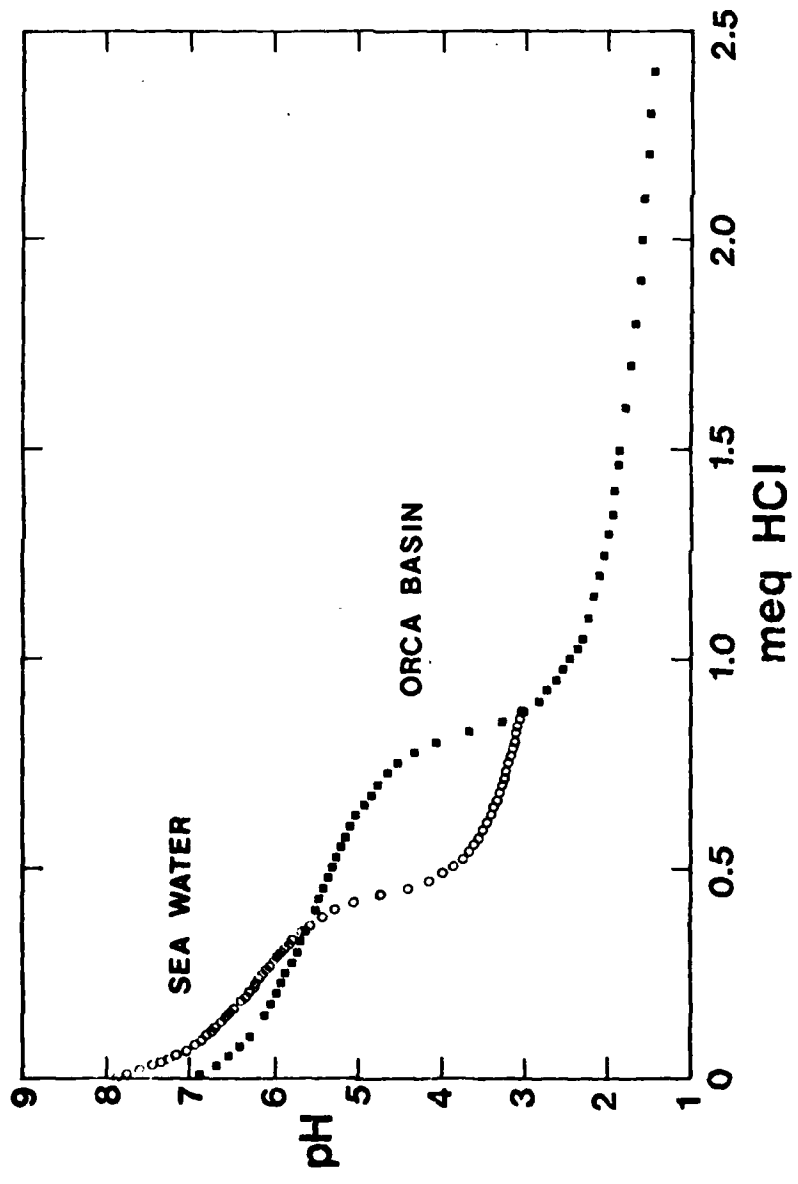


Fig. 48. Representative HCl titration curves of sea water and Orca Basin brine. Volume of sample was 180 ml for each.

For interpretation of the data in Fig. 48, the buffer capacity (the incremental change in pH for a given volume of HCl added) was calculated and plotted for each series of data points. The results of this calculation are presented in Fig. 49. The buffer capacity versus pH curve is also plotted for the sea water HCl titration as a comparison. Maxima and minima in these curves correspond to the  $pK'$  values of the dissociation constants of the carbonate system in each solution. For example, the  $pK'_2$  value for sea water would correspond to the minimum at a buffer capacity of about 0.25. The  $pK'_1$  value for sea water corresponds to the buffer capacity maximum at a pH of about 6.1. Since the pH of the Orca Basin is so low, there is no upper minimum to use in graphically determining the  $pK'_2$ . The  $pK'_1$  corresponds to the maximum in buffer capacity which occurs at a pH of 5.5. A  $pK'_1$  of 5.5 for the brine represents a factor of three increase in the  $K'_1$  dissociation constant, compared to sea water.

Application of the iterative solution technique of Sass and Ben-Yaakov to this titration allows more accurate calculation of the dissociation constant. Using the titration data plotted in Fig. 48 and the mathematical sequence given in Table 20, the alkalinity, the total carbon dioxide, and the carbonate dissociation constants were calculated for the Orca Basin and for sea water, for comparison. The results of this calculation are presented in Table 23. Analysis of other samples gave lower carbon dioxide

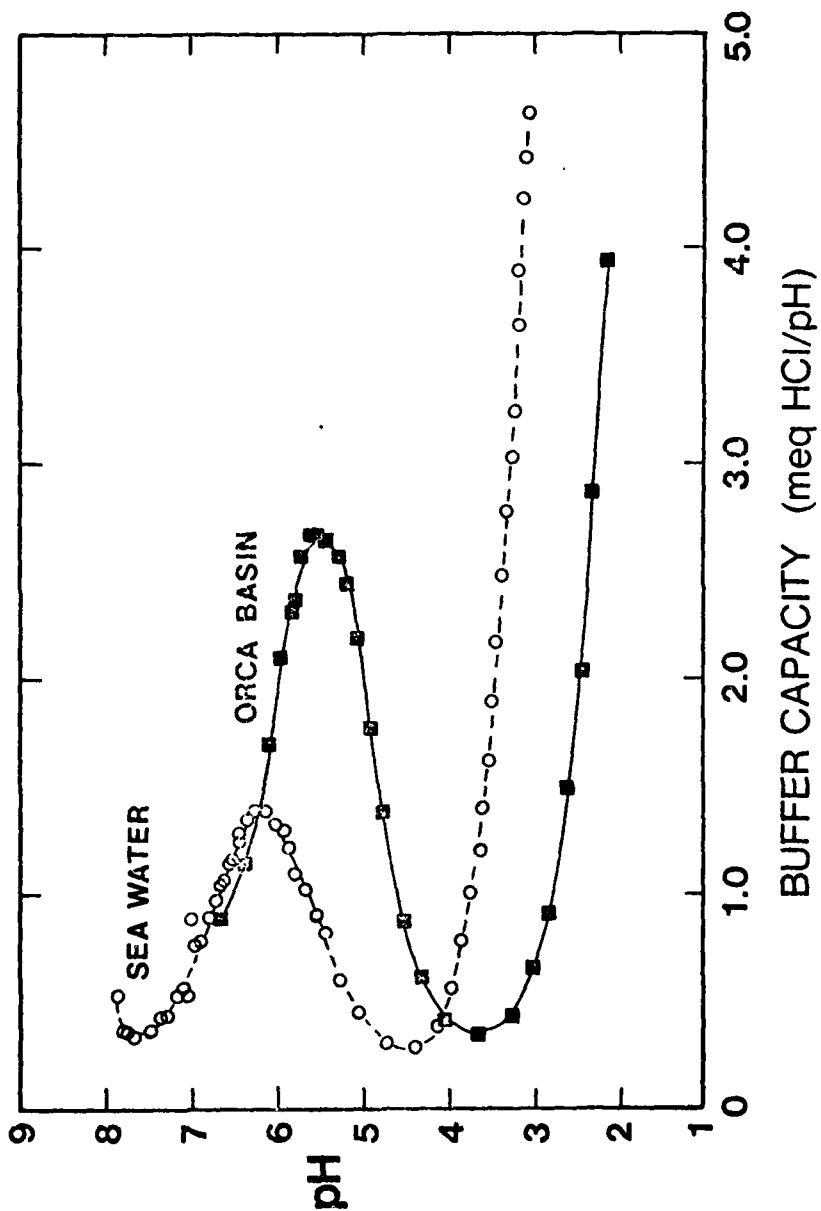


Fig. 49. Buffer capacity (the incremental change in pH for a given addition of HCl) plotted as a function of pH. This type of plot can be used to estimate the dissociation constants, as described in the text. Titration data and samples are the same as for Fig. 48.

Table 23. Calculated alkalinity and carbonate dissociation constants for seawater and Orca Basin brine.

Brine	Alkalinity ( $\text{meq}\cdot\text{liter}^{-1}$ )	Total $\text{CO}_2$ ( $\text{mmol}\cdot\text{liter}^{-1}$ )	$\text{pK}_1$	$\text{pK}_2$
Sea water	2.20	2.13	5.80	9.14
Orca Basin brine	4.701	4.750	5.49	8.34
Dead Sea brine *	3.86	4.41	5.09	6.23

\* From Sass and Ben-Yaakov (1977).

concentrations, but also lower alkalinity concentrations, indicative of some  $\text{CO}_2$  loss.

The dissociation constants increase with increasing salinity in the Orca Basin. In the Dead Sea brine, Sass and Ben-Yaakov (1977) also noted this increase in the dissociation constants. Their values are given in Table 23 for comparison. It is immediately obvious that the dissociation constants of the Dead Sea brine are altered more (relatively to sea water) due to the increasing salt content, than are the constants for the Orca Basin. This magnitude of difference can be seen more dramatically when the distribution of carbonate system species is calculated for the Orca Basin and the Dead Sea brines. The results of the species distributions as a function of pH are given in Fig. 50. The results are presented as relative percentages of each species as a function of pH. Presented in this manner, the relative distributions in each solution can be directly compared without considering the total carbonate concentration.

The distribution of the carbonate species in the Dead Sea brine is very different from either sea water or Orca Basin brine. At the natural pH of the Dead Sea, the concentrations of carbonate and bicarbonate ions are approximately equal. This is atypical for the other systems, where bicarbonate is by far the dominant inorganic carbon ion in solution at in situ pH values.

The shape of the distribution in the Orca Basin is very much like that of sea water, but all distribution curves are shifted

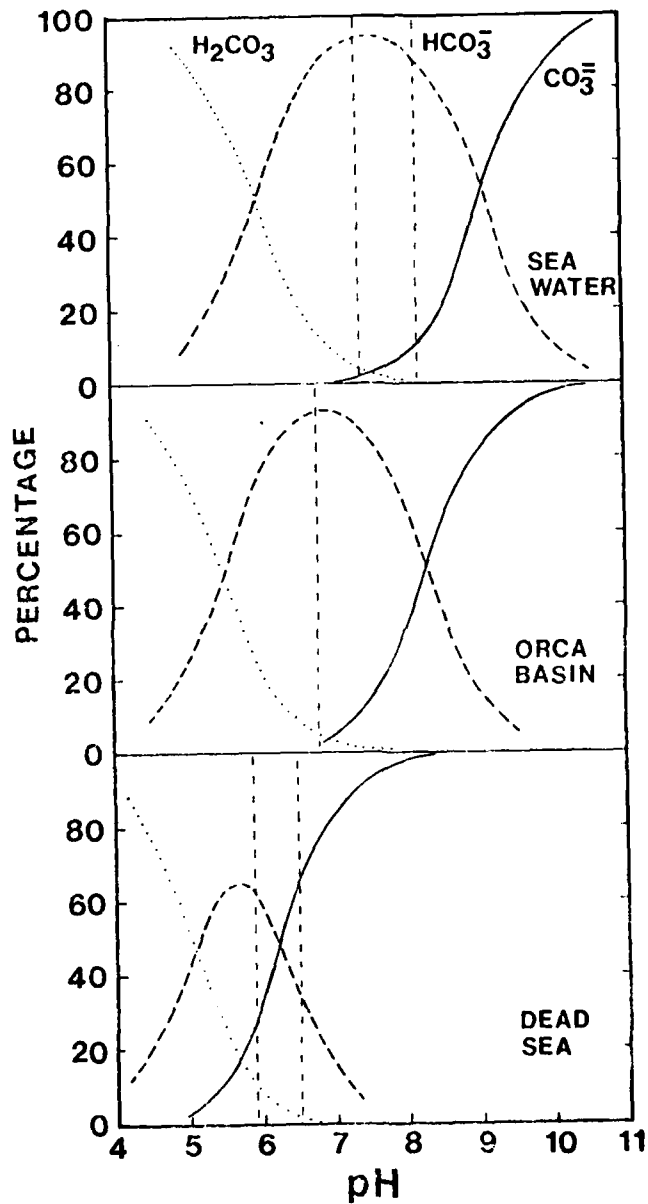


Fig. 50. Carbonate system species distributions for sea water, the Orca Basin brine and the Dead Sea brines. The vertical dashed lines indicate the pH ranges of the natural systems.



toward lower pH values. Orca Basin brine has a pH of 6.83. This small difference in the species profiles underscores the chemical nature of the Orca Basin brine. The Orca Basin brine is like sea water that has been saturated with sodium chloride. On the other hand, the major ionic differences between the Dead Sea brine and sea water play havoc with the carbonate species distribution there. Sass and Ben-Yaakov (1977) have attributed most of the disparity in the Dead Sea carbonate system to the concentration of magnesium in the Dead Sea brine. The bicarbonate to carbonate dissociation constant is about threefold more sensitive to variations in magnesium concentration than is the  $K'_1$  (Ben-Yaakov and Goldhaber 1977). This effect is attributed to the relatively high stability of  $MgCO_3^0$  complex as compared to that of  $MgHCO_3^-$  (Garrels and Thompson 1962). The high  $K'_2$  dissociation constant for the Dead Sea brine causes the observed distributions. Since both  $K'_1$  and  $K'_2$  are greater, but only two or three times sea water values, the Orca Basin brine distributions are similar to sea water.

#### Pteropod Preservation in the Sediments

The consequences of salinity causing an increase in the dissociation constants, combined with the biogenic total  $CO_2$  in the brine has resulted in such a high concentration of carbonate ion in the sediments, that there is rather striking preservation

of carbonaceous material in the Orca Basin sediments. Even pteropod tests, which are made of the more soluble aragonite, are preserved in the sediment. Fig. 51 shows several electron microscope photographs (taken by L. A. Barnard) of some of the pteropods found in the Orca Basin sediments. As noted by Kennett and Penrose (1978), the preservation is striking. There is no evidence of even the tiny etch marks often observed on calcium carbonate tests undergoing slow dissolution in typical marine sediments (Alexandersson 1975). Fig. 52 shows one of the pteropods found in the basin, and two additional magnifications of one portion of the pteropod. These micrographs show that there is absolutely no dissolution of this soluble aragonitic species (Cavolinia longirostris).

The observation of the well-preserved microfossils in the sediment, with no evidence of  $\text{CaCO}_3$  precipitation, implies the Orca Basin sediments are in equilibrium with respect to calcium carbonate. Since the concentration of carbonate ion in the brine is no higher than in normal sea water, one must consider the solubility coefficient of calcium carbonate in the brine sediments to be different than in normal marine environments. Also there is an excess of  $17 \text{ mmol} \cdot \text{Ca} \cdot \text{kg}^{-1}$  brine in the Orca Basin. This excess calcium would contribute to the preservation of the calcite and aragonite tests in the brine. But, where did this excess calcium come from? Could it be from past dissolution of



Fig. 51. Scanning electron photographs of three pteropod species preserved in the Orca Basin sediments. The number above the length scale on each micrograph gives the scale length in micrometers.

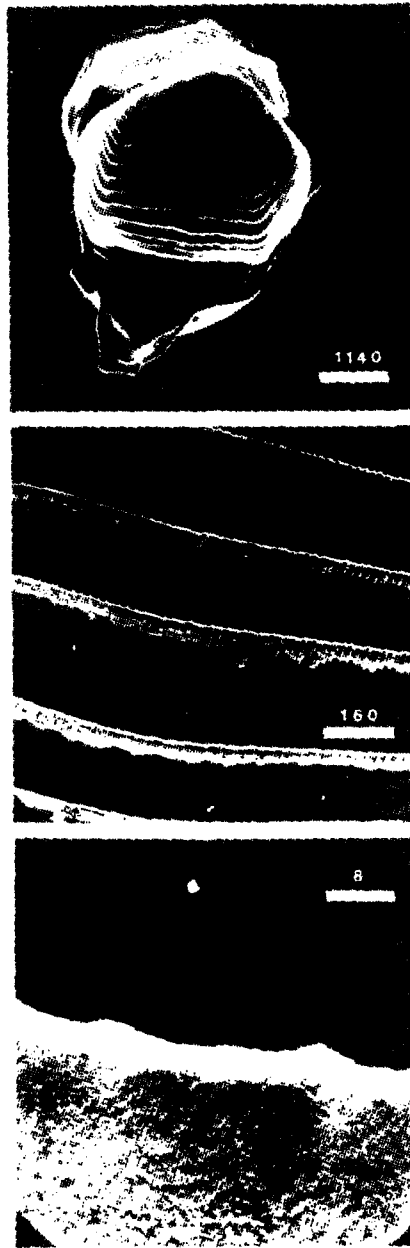


Fig. 52. Three scanning electron photographs of a *Cavolinia longirostris* pteropod found in the Orca Basin sediments. The number above each length scale on each micrograph gives the scale length in micrometers. Even at high magnification, there is no evidence of aragonite dissolution in the Orca Basin sediments.

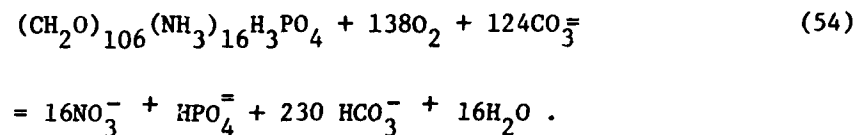
calcium carbonate in the brine? This seems improbable since the pteropods in the brine are preserved throughout the sediment column (Addy and Behrens 1980). In one core section at a depth of 689 cm, Addy and Behrens (1980) found that the percentage of sand increased significantly, to 21%. The increase in sand was entirely skeletal, calcium carbonate tests. In fact, they noted that the preservation of calcium carbonate in the sediments increased the sedimentation rate by as much as  $21 \text{ cm} \cdot \text{ka}^{-1}$ . Thus, the excess calcium must have an alternate source.

#### Excess Calcium and $K_{sp}$ of Brine

It will be shown here that this excess calcium originated with the inflowing brine and not from calcium carbonate dissolution in the basin.  $\text{CaCO}_3$  can decompose to form



With this dissolution,  $\text{Ca}^{++}$  increases (1 mole) along with alkalinity (2 equivalents) and total  $\text{CO}_2$  (1 mole). At the same time, organic material is decomposing in the brine producing  $\text{CO}_2$ ,  $\text{HNO}_3$  (or  $\text{NH}_3$ ) and  $\text{H}_3\text{PO}_4$  (Table 12). In sea water  $\text{CO}_2$ ,  $\text{HNO}_3$  and  $\text{H}_3\text{PO}_4$  react with  $\text{CO}_3^{-}$  (Brewer et al. 1975). Thus, the combined decomposition equation (aerobic) which includes  $\text{CO}_3^{-}$  reaction can be written as



This combined reaction scheme (Eq. 54) results in an increase of 106 moles in  $\Sigma\text{CO}_2$  and a decrease in 17 equivalents in alkalinity. Chen (1978) has discussed this combined effect of  $\text{CaCO}_3$  dissolution and decomposition of organic matter in the ocean. He represents the decomposing of  $x \mu\text{mol}$  of  $\text{CaCO}_3$  and  $y \mu\text{mol}$  of organic matter in sea water by the expressions:

$$\Delta\text{TA} = 2x - 17y \quad (55)$$

$$\Delta\Sigma\text{CO}_2 = x + 106y \quad (56)$$

$$\Delta\text{NO}_3 = 16y \quad (57)$$

where TA is in microequivalents and  $\Sigma\text{CO}_2$  and  $\text{NO}_3$  are in micromoles. The same equations are applicable to anaerobic decomposition since the same ratios are involved, only  $\text{NH}_3$  is produced as the terminal nitrogen product. In examining the Orca Basin, Eq. 57 becomes

$$\Delta\text{NO}_3 + \Delta\text{NH}_3 = 16y \quad (58)$$

since both aerobic (initially) and anaerobic processes have been active in bringing the brine to its present state. Eqs. (55) and (56) and Eqs. (56) and (57) were solved by Chen (1978) for  $x$ , the  $\text{CO}_3^{\equiv}$  concentration which is equivalent to  $\Delta\text{Ca}$ , the change in

calcium concentration (see Eq. 54). The results are

$$\Delta\text{Ca} = x = 0.46288 \Delta\text{TA} + 0.074236 \Delta\Sigma\text{CO}_3 \quad (59)$$

$$\Delta\text{Ca} = x = 0.5 \Delta\text{TA} + 0.53125 \Delta\text{NO}_3 \quad (60)$$

Since alkalinity, total  $\text{CO}_2$ , nitrate and ammonia have all been measured for the Orca Basin, the predicted  $\Delta\text{Ca}^{++}$  can be calculated showing the increase in  $\text{Ca}^{++}$  from possible dissolution of  $\text{CaCO}_3$  and decomposition of organic matter. These calculations are shown in Table 24. Eq. 59 predicts a  $\Delta\text{Ca}^{++}$  of a  $0.96 \text{ mmol}\cdot\text{kg}^{-1}$  while Eq. 60 predicts  $1.12 \text{ mmol}\cdot\text{kg}^{-1}$  for an average of  $1.04 \text{ mmol}\cdot\text{kg}^{-1}$ . This increase is far below the measured  $\Delta\text{Ca}^{++}$  (compared to deep Gulf water) of  $17.0 \text{ mmol}\cdot\text{kg}^{-1}$  (Table 3). It can also be calculated (Table 24) that if enough  $\text{Ca}^{++}$  were put into Gulf sea water to reach the Orca Basin levels, the resultant alkalinity would be approximately  $35 \text{ meq}\cdot\text{kg}^{-1}$ . Thus the excess  $\text{Ca}^{++}$  in the Orca Basin must have entered with the original brine, probably from the dissolution of calcium sulfate along with NaCl.

The complete preservation of the aragonitic pteropod shells in the Orca Basin is unusual. While some other anoxic environments have partial preservation of pteropod tests (see, for example, Berger and Soutar 1970), preservation is complete in the Orca Basin sediments. Since no  $\text{CaCO}_3$  precipitation occurs in the Orca Basin, one can assume that Orca brine is in equilibrium with respect

Table 24. Calculations showing the possible changes in total calcium content resulting from the increases in alkalinity, total  $\text{CO}_2$ , and ammonia in the Orca Basin.

---

Date for Calculations

$$\Delta\text{Ca}^{++} = 17.0 \text{ mmol}\cdot\text{kg}^{-1} \text{ (Shokes et al. 1977)}$$

$$\Delta\text{TA} = 4.015 \text{ meq}\cdot\text{kg}^{-1} \text{ (Millero et al. 1979 and this study)}$$

$$\Delta\text{ECO}_2 = 1.913 \text{ mmol}\cdot\text{kg}^{-1} \text{ (Sackett et al. 1979 and this study)}$$

$$\Delta\text{NO}_3^- + \text{NH}_3 = 441 \text{ }\mu\text{mol}\cdot\text{kg}^{-1} \text{ (this study)}$$

Basic Equations

$$\text{a. } \Delta\text{Ca}^{++} = 0.46288 \Delta\text{TA} + 0.074236 \Delta\text{ECO}_2$$

$$\text{b. } \Delta\text{Ca}^{++} = 0.5 \Delta\text{TA} + 0.53125 (\Delta\text{NO}_3^- + \text{NH}_3)$$

$\text{Ca}^{++}$  Predicted Changes

$$\text{a. } \Delta\text{Ca}^{++} = 0.46288 (1765 \text{ }\mu\text{eq}\cdot\text{kg}^{-1}) + 0.074236 (1.913 \text{ }\mu\text{mol}\cdot\text{kg})$$

$$\Delta\text{Ca}^{++} = 958 \text{ }\mu\text{mol}\cdot\text{kg}$$

$$\text{b. } \Delta\text{Ca}^{++} = 0.5(1765 \text{ }\mu\text{eq}\cdot\text{kg}^{-1}) + 0.53125(441 \text{ }\mu\text{mol}\cdot\text{kg}^{-1})$$

$$\Delta\text{Ca}^{++} = 1117 \text{ }\mu\text{mol}\cdot\text{kg}^{-1}$$

$$\text{Average} = 1037 \text{ }\mu\text{mol}\cdot\text{kg}^{-1}$$

Alkalinity Required to Produce Orca  $\Delta\text{TA}$

$$\text{a. } \Delta\text{TA} = \frac{\text{Ca}^{++} - 0.074236 \text{ CO}_2}{0.46288}$$

$$\Delta\text{TA} = 36.4 \text{ }\mu\text{eq}\cdot\text{kg}^{-1}$$

$$\text{b. } \Delta\text{TA} = \frac{\text{Ca}^{++} - 0.53125 (\text{NO}_3^- + \text{NH}_3)}{0.5}$$

$$\Delta\text{TA} = 34.5 \text{ }\mu\text{eq}\cdot\text{kg}^{-1}$$

$$\text{Average TA} = 35.5 \text{ }\mu\text{eq}\cdot\text{kg}^{-1}$$


---



to aragonite, and the apparent solubility product ( $K'_{sp}$ ) for aragonite can be calculated. This constant is defined in terms of total concentration, i.e. the product of the total calcium ( $Ca^{++}$ ) and total carbonate ( $CO_3^{=}$ ) ions. Total calcium is given in Table 3. Carbonate ion concentration is obtained from the relationship:

$$[CO_3^{=}] = \frac{A_c K_2'}{(a_H^+ + 2K_2')} \quad (61)$$

Using the pH of the brine as 6.83 and a  $pK_2'$  of 8.34, one obtains a value of  $3.2 \times 10^{-6} \text{ mol}^2 \cdot \text{kg}^{-2}$  for  $K'_{sp}$  at 25°C. This value is about six times larger than the corresponding value for sea water (Ben-Yaakov and Goldhaber 1973). If the Orca brine had normal sea water calcium concentration (see Table 3), the ion product for  $[Ca^{++}][CO_3^{=}]$  would be  $1.2 \times 10^{-6} \text{ mol}^2 \cdot \text{kg}^{-2}$ . This would make the brine undersaturated with respect to aragonite by a factor of three. Thus, it is the high calcium content of the brine which causes the complete preservation of the aragonitic pteropod shells in the Orca Basin sediments.

CHAPTER VI  
SUMMARY AND CONCLUSIONS

The advective, diffusive, and reactive processes acting upon the dissolved gases in the hypersaline waters of the Orca Basin are no different than in other marine, anoxic environments. Only the magnitude of the constants and the rates of reaction are affected by the increased concentrations of dissolved salts in the Orca Basin water. The processes differ from other less saline areas only in degree of interaction. This study has shown the potential of dissolved gas studies in understanding those processes.

This investigation confirmed that the Orca Basin brine is the result of the low temperature dissolution of a sea floor salt deposit by Gulf of Mexico sea water, with the resultant brine flowing into the depths of the basin. The similarity of the nitrogen to argon ratio in the Orca Basin water (exclusive of salt), to that of deep Gulf of Mexico water (also corrected for the salt content) supports the hypothesis of low temperature solution of a sill salt. This first postulate was based on the ion content of the brine, relative to evaporite and solution deposit systems. The dissolved nitrogen and argon data provided an independent confirmation of that mechanism of brine formation, and furthermore, gave a temperature maximum for the dilution water of less than 5°C. The bottom water temperature in the Gulf of Mexico is 4.2°C.

The step-like feature in the oxygen concentration was perhaps the most interesting of the observations involving dissolved gases. A simple consideration of the possible mixing processes which occur at the brine-sea water interface revealed that it would be possible to account for this step-like feature from mixing processes alone. More work is needed to prove this conclusively. Detailed, continuous, simultaneous measurements of temperature, salinity, dissolved oxygen, and pH across the brine-sea water transition regions should be made with great precision in order to indicate accurately the nature of the mixing processes at the Orca Basin interface.

Measurements of other parameters indicated that the brine-sea water interface in the Orca Basin is physically active, chemically reactive, and behaves much like the sediment-sea water interface in other anoxic regions. This fact was most apparent from the dissolved molecular nitrogen and ammonia distributions in the interface and transition zones. The observed distributions of dissolved molecular nitrogen in the low oxygen region were strikingly similar to the near surface sediment distributions that were reported by Barnes et al. (1975) for the southern California borderland basin sediments. This similarity illustrates the important laboratory potential of the Orca Basin system. The interface there is like a sediment-sea water interface, except that it can be sampled over a length scale of meters, rather than the

centimeters and millimeters required for sediment work. In reviewing the other constituents, such as nitrous oxide and ammonia, these parameters also change, going from the Gulf water into the brine, in the same manner that sea water-sediment interstitial water transitions are made in many environments. Thus, the Orca Basin interface region could be useful in examining the transition processes of consumption and production by which concentration gradients change rapidly across interface boundaries.

The dissolved reduced gases in the high salinity brine are produced by the same decomposition processes that are operative in other anoxic basins. The high concentrations of methane and ethane in the brine result from the great stability of the brine in the basin, which has a residence time of over 6000 years. Both methane and ethane are produced in the basin. However, in spite of the high concentrations observed, empirical evidence from isotopic composition of the carbon compounds (methane and carbon dioxide), along with the unusual depth distributions of iron and hydrogen sulfide (when compared with other anoxic areas), indicated that the rates of microbial decomposition are slower in the Orca Basin. The presence of preserved Sargassum seaweed deep in the sediments is further evidence for reduced decomposition rates in the Orca Basin. Reduced microbial activity produces little free sulfide to react with the iron in the brine. High concentrations of iron have accumulated in the water column.

The carbonate system in the high salinity brine is similar to sea water which has simply been saturated with sodium chloride. The relative distributions of the carbonate species are the same as those found in normal salinity sea water. The low pH in the brine results from the input of  $2.2 \text{ mmol} \cdot \text{liter}^{-1}$  of biogenic carbon dioxide. The carbonate dissociation constants differ only because of the different degree of interaction caused by the higher salt content. The apparent solubility product constant of aragonite in the brine is about six times larger than the corresponding value for sea water at  $25^\circ\text{C}$ . Thus it must be the high concentrations of calcium in the brine which has caused the preservation of pteropod shells in the basin sediments.

This investigation has shown the utility of dissolved gas in examining the active physical and biological processes in unusual environments. Such an examination should also answer questions about the Orca Basin that had been posed during previous studies. An examination of both reactive and unreactive gases has yielded significant information about the formation of the brine, as well as about the rates of reaction in the brine. The objectives of this work have been met with solutions that are internally consistent. Dissolved gases can be powerful geochemical tracers for oceanic processes.

## REFERENCES

- Adams, D.D. 1973. A laboratory model for plankton decomposition in anaerobic and aerobic sea water. Ph.D. Thesis, Dalhousie University, Halifax, Nova Scotia, Canada. 228 p.
- Addy, S.K. and E.W. Behrens. 1980. Time accumulation of hypersaline anoxic brine in Orca basin (Gulf of Mexico). *Mar. Geol.* 37: 241-252.
- Alexandersson, E.J. 1975. Etch patterns on calcareous sediment grains: petrographic evidence of marine dissolution of carbonate minerals. *Science* 189: 47-48.
- Amit, O. and Y.K. Bendor. 1971. pH-dilution curves of saline waters. *Chem. Geol.* 7: 307-313.
- Antoine, J.W. and W.R. Bryant. 1969. Distribution of salt and salt structures in the Gulf of Mexico. *Amer. Assoc. Pet. Geol. Bull.* 53: 2543-2550.
- Armstrong, F.A.J., C.R. Stearns, and J.D.H. Strickland. 1967. The measurement of upwelling and subsequent biological processes by means of the Technicon AutoAnalyzer and associated equipment. *Deep-Sea Res.* 14: 381-389.
- Atkinson, L.P. and F.A. Richards. 1967. The occurrence and distribution of methane in the marine environment. *Deep-Sea Res.* 14: 673-684.
- Atlas, E.L., S.W. Hager, L.I. Gordon, and P.K. Park. 1971. A practical manual for use of the Technicon AutoAnalyzer in seawater nutrient analyses; revised. Oregon State University, Department of Oceanography, Technical Report 215. 49 p.
- Barabee, J.M. and W.J. Payne. 1967. Products of denitrification by a marine bacterium as revealed by gas chromatography. *Mar. Biol.* 1: 136-139.
- Barnes, R.O., K.K. Bertine, and E.D. Goldberg. 1975.  $N_2:Ar$ , nitrification and denitrification in southern California borderland basin sediments. *Limnol. Oceanogr.* 20: 962-970.
- Barnes, R.O. and E.D. Goldberg. 1976. Methane production and consumption in anoxic marine sediments. *Geology* 4: 297-300.

- Benard, H. 1901. Les tourbillons cellulaires dans un nappes liquide transportant de la chaleur par convection en regime permanent. *Ann. Chim. Phys.* 23: 62-144.
- Ben-Yaakov, S. 1970. A method for calculating the in situ pH of seawater. *Limnol. Oceanogr.* 15: 326-328.
- Ben-Yaakov, S. and M.B. Goldhaber. 1973. The influence of sea water composition on the apparent constants of the carbonate system. *Deep-Sea Res.* 20: 87-99.
- Ben-Yaakov, S. and E. Sass. 1977. Independent estimate of pH of Dead Sea brine. *Limnol. Oceanogr.* 22: 374-376.
- Berger, W.H. and A. Soutar. 1970. Preservation of plankton shells in an anaerobic basin off California. *Geol. Soc. Amer. Bull.* 81: 275-282.
- Bernard, B.B. 1978. Light hydrocarbons in marine sediments. Ph.D. Thesis, Texas A&M University, College Station, Tx. 144 p.
- Bernard, B.B. 1979. Methane in marine sediments. *Deep-Sea Res.* 26A: 429-443.
- Bernard, B.B., J.M. Brooks, and W.M. Sackett. 1976. Natural gas seepage in the Gulf of Mexico. *Earth Planet. Sci. Lett.* 31: 48-54.
- Berner, R.A. 1971. Principles of chemical sedimentology. McGraw-Hill Book Co., 240 p.
- Berner, R.A. 1972. Sulfate reduction, pyrite formation and the oceanic sulfur budget, p. 347-361. In D. Dryssen and D. Jagner [eds.], *The changing chemistry of the oceans*. Almquist & Wiksell.
- Berner, R.A. 1974. Iron sulfides in Pleistocene deep Black Sea sediments and their paleo-oceanographic significance, p. 524-531. In E.T. Degens and D.A. Ross [eds.], *The Black Sea - geology, chemistry and biology*, Mem. 20. Amer. Assoc. Pet. Geol. George Banta Company.
- Berner, R.A. 1978. Sulfate reduction and the rate of deposition of marine sediment. *Earth Planet. Sci. Lett.* 37: 492-498.
- Berner, R.A., T. Baldwin, and G.R. Holdren, Jr. 1979. Authigenic iron sulfide as paleosalinity indicators. *J. Sed. Petrol.* 49: 1345-1350.
- Borchert, H. and R.O. Muir. 1964. Salt deposits; the origin, metamorphism and deformation of evaporites. Van Nostrand. 338 p.

- Bouma, A.H., L.B. Smith, B.R. Sidner, and T.R. McKee. 1975. Submarine geomorphology and sedimentation patterns of the Gyre intraslope basin, northwest Gulf of Mexico. Technical Report 75-9-T, Department of Oceanography, Texas A&M University, College Station, Tx. 163 p.
- Braitsch, O. and A.G. Herrmann. 1963. Zur Geochemie des Broms in Salinaren Sedimenten, Teil I. Experimentelle Bestimmung der Br-Verteilung in verschiedenen natürlichen Salzsystemen. *Geochim. Cosmochim. Acta* 27: 361-391.
- Breed, R.S., E.G.D. Murray, and N.R. Smith. 1957. *Bergley's manual of determinative microbiology*, 7th ed. The Williams and Wilkins Co. 1094 p.
- Brewer, P.G., C.D. Densmore, R. Munns, and R.J. Stanley. 1969. Hydrography of the Red Sea brines, p. 138-147. In E.T. Degens and D.A. Ross [eds.], *Hot brines and recent heavy metal deposits in the Red Sea*. Springer-Verlag.
- Brewer, P.G. and D.W. Spencer. 1974. Distribution of some trace elements in Black Sea and their flux between dissolved and particulate phases, p. 137-143. In E.T. Degens and D.A. Ross [eds.], *The Black Sea - geology, chemistry, and biology*, Mem. 20. Amer. Assoc. Pet. Geol. George Banta Company.
- Brewer, P.G., G.T.F. Wong, M.P. Bacon, and D.W. Spencer. 1975. An oceanic calcium problem. *Earth Planet. Sci. Lett.* 26-81-87.
- Brisou, J. and N. Vargues. 1961. Proteolysis and nitrate reduction in sea water, p. 410-414. In C.H. Oppenheimer [ed.], *Symposium on marine microbiology*, Charles C. Thomas.
- Broenkow, W.W. 1969. The distribution of nonconservative solutes related to the decomposition of organic matter in anoxic basins. Ph.D. Thesis, University of Washington, Seattle, Wash. 207 p.
- Broenkow, W.W. and J.D. Cline. 1969. Colorimetric determination of dissolved oxygen at low concentrations. *Limnol. Oceanogr.* 14: 450-454.
- Brooks, J.M., T.J. Bright, B.B. Bernard, and C.R. Schwab. 1979. Chemical aspects of a brine pool at the East Flower Garden Bank, northwestern Gulf of Mexico. *Limnol. Oceanogr.* 24: 735-745.
- Brooks, J.M., D.F. Reid, and B.B. Bernard. 1980. Methane in the upper water column of the Gulf of Mexico. *Deep-Sea Res.* In press.



- Burke, Jr., R.A., J.M. Brooks, and W.M. Sackett. 1980. Light hydrocarbons in Red Sea brines and sediments. *Geochim. Cosmochim. Acta* (submitted).
- Carlucci, A.F. and P.M. McNally. 1969. Nitrification by marine bacteria in low concentrations of substrate and oxygen. *Limnol. Oceanogr.* 14: 736-739.
- Carpenter, J.H. 1965. The accuracy of the Winkler method for dissolved oxygen analysis. *Limnol. Oceanogr.* 10: 135-140.
- Chen, C.A. 1978. Decomposition of calcium carbonate and organic carbon in the deep oceans. *Science* 201: 735-736.
- Claypool, G.E. and I.R. Kaplan. 1974. The origin and distribution of methane in marine sediments, p. 99-139. *In* I.R. Kaplan [ed.], *Natural gases in marine sediments*. Plenum Press.
- Cline, J.D. 1969. Spectrophotometric determination of hydrogen sulfide in natural waters. *Limnol. Oceanogr.* 14: 454-458.
- Cline, J.D. and F.A. Richards. 1972. Oxygen deficient conditions and nitrogen reduction in the eastern tropical North Pacific Ocean. *Limnol. Oceanogr.* 17: 885-900.
- Cohen, Y. 1978. Consumption of dissolved nitrous oxide in an anoxic basin, Saanich Inlet, British Columbia. *Nature* 272: 235-237.
- Cohen, Y. and L.I. Gordon. 1978. Nitrous oxide in the oxygen minimum of the eastern tropical North Pacific: evidence for its consumption during denitrification and possible mechanisms for its production. *Deep-Sea Res.* 25: 509-524.
- Collins, A.G. 1975. *Geochemistry of oilfield waters*. Elsevier Scientific Publishing Company. 496 p.
- Cooke, R.C. 1973. The use of activated charcoal for the removal of oxygen from gas systems. *Limnol. Oceanogr.* 18: 150-152.
- Craig, H.C. 1969. Geochemistry and origin of the Red Sea brines, p. 208-242. *In* E.T. Degens and D.A. Ross [eds.], *Hot brines and recent heavy metal deposits in the Red Sea*. Springer-Verlag.
- Crank, J. 1975. *The mathematics of diffusion*. 2nd edition. Clarendon Press. 414 p.
- Crozier, T.E. and S. Yamamoto. 1974. Solubility of hydrogen in water, sea water, and sodium chloride solutions. *J. Chem. Eng. Data* 19: 242-244.

- Davis, J.B. and H.F. Yarbrough. 1966. Anaerobic oxidation of hydrocarbons by Desulfovibrio desulfuricans. Chem. Geol. 1: 137-144.
- Deffeyes, K.S. 1965. Carbonate equilibria: a graphic and algebraic approach. Limnol. Oceanogr. 10: 412-426.
- Degens, F.T. and D.A. Ross [eds.]. 1974. Hot brines and recent heavy metal deposits in the Red Sea. Springer-Verlag. 633 p.
- Deuser, W.G., E.T. Degens, and G.R. Harvey. 1973. Methane in Lake Kivu: New data bearing on its origin. Science 181: 51-54.
- Douglas, E. 1964. Solubilities of oxygen, argon, and nitrogen in distilled water. J. Phys. Chem. 68: 169-174.
- Edmond, J.M. 1970. High precision determination of titration alkalinity and total carbon dioxide content of sea water by potentiometric titration. Deep-Sea Res. 17: 737-750.
- Edmond, J.M. 1972. The thermodynamic description of the CO<sub>2</sub> system in seawater: development and current status. Proc. Roy. Soc. Edinburgh 72: 371-387.
- Emery, K.O. and D. Hogan. 1958. Gases in marine sediments. Bull. Amer. Assoc. Petrol. Geol. 42: 2174-2188.
- Fanning, K.A. and M.E.Q. Pilson. 1972. A model for the anoxic zone of the Cariaco Trench. Deep-Sea Res. 19: 847-863.
- Fleming, R.H. 1940. Composition of plankton and units for reporting population and production. Proc. Sixth Pac. Sci. Assoc. 3: 535-540.
- Fredericks, A.D. 1972. Distribution of organic carbon, p. 6-7. In Vivian C. Bushnell [ed.], Serial atlas of the marine environment. Folio 22, Chemistry, primary productivity, and benthic algae of the Gulf of Mexico. Amer. Geograph. Soc.
- Froelich, P.N., G.P. Klinkhammer, M.L. Bender, N.A. Luedtke, G.R. Heath, D. Cullen, and P. Dauphin. 1979. Early oxidation of organic matter in pelagic sediments of the eastern equatorial Atlantic: suboxic diagenesis. Geochim. Cosmochim. Acta. 43: 1075-1090.
- Fuhs, G.S. 1961. Der mikrobielle Abbau von Kohlenwasserstoffen. Arch. Mikrobiol. 39: 374-422.
- Gaines, A.G. and M.E.Q. Pilson. 1972. Anoxic water in the Pettaquamscutt River. Limnol. Oceanogr. 17: 42-49.

- Games, L.M., J.M. Hayes, and R.P. Gunsalus. 1978. Methane-producing bacteria: natural fractionation of the stable carbon isotopes. *Geochim. Cosmochim. Acta.* 42: 1295-1297.
- Garrels, R.M. and M.E. Thompson. 1962. A chemical model for sea water at 25°C and one atmosphere total pressure. *Amer. J. Sci.* 260: 57-66.
- Garrison, L.E. and R.G. Martin, Jr. 1973. Geologic structures in the Gulf of Mexico basin. U.S. Geological Survey, Prof. Paper 773. 85 p.
- Gieskes, J.M. 1974. The alkalinity-total carbon dioxide system in sea water, p. 123-151. In E.D. Goldberg [ed.], *The Sea*, v. 5: Marine Chemistry. Wiley Interscience.
- Gille, J. 1967. Interferometric measurement of temperature gradient reversal in a layer of convecting air. *J. Fluid Mech.* 30: 371-384.
- Glueckauf, E. 1951. The composition of atmospheric air, p. 3-10. In T.F. Malone [ed.], *Compendium of meteorology*, *Amer. Meteor. Sci.*
- Goering, J.J. 1968. Denitrification in the oxygen minimum layer of the eastern tropical Pacific Ocean. *Deep-Sea Res.* 15: 157-164.
- Goldhaber, M.B. and I.R. Kaplan. 1974. Controls and consequences of sulfate reduction rates in recent marine sediments. *Soil Sci.* 119: 42-55.
- Gordon, K.I., Y. Cohen, and D.R. Standley. 1977. The solubility of molecular hydrogen in seawater. *Deep-Sea Res.* 24: 937-941.
- Guinasso, N.L., Jr. 1977. Continuous temperature profile in the Orca Basin. *EOS, Trans., Amer. Geophys. Union.* 58: 1175.
- Hammond, D.E. 1974. Dissolved gases in Cariaco Trench sediments: Anaerobic diagenesis, p. 71-89. In I.R. Kaplan [ed.], *Natural gases in marine sediments*. Plenum Press.
- Harned, H.S. and B.B. Owen. 1950. *Physical chemistry of electrolytic solutions*. Reinhold Publishing Co. 645 p.
- Holser, W.T. 1966. Bromide geochemistry of salt rocks, p. 248-275. In J.L. Rau [ed.], *Second Symposium on Salt*, v. 1, Northern Ohio Geological Soc.
- Hunt, J.M. 1974. Hydrocarbon geochemistry of the Black Sea, p. 599-604. In E.T. Degens and D.A. Ross [eds.], *The Black Sea - geology, chemistry, and biology*, Mem. 20, Amer. Assoc. Pet. Geol., George Banta Company.

- Johnson, K.S. and R.M. Pytkowicz. 1979. Ion association of chloride and sulphate with sodium, potassium, magnesium, and calcium in seawater at 25°C. *Mar. Chem.* 8: 87-94.
- Kaplan, I.R. and S.C. Rittenberg. 1964. Microbiological fractionation of sulfur isotopes. *J. Gen. Microbiol.* 34: 195-212.
- Karl, D.M., P.A. LaRock, and D.J. Schultz. 1977. Adenosine triphosphate and organic carbon in the Cariaco Trench. *Deep-Sea Res.* 24: 105-113.
- Kennett, J.P. and N.L. Penrose. 1978. The occurrence of fossil holocene seaweed and attached calcareous polychaetes in an anoxic basin, Gulf of Mexico. *Nature* 276: 172-173.
- Kester, D.R. 1975. Dissolved gases other than CO<sub>2</sub>, p. 498-556. In J.P. Riley and G. Skirrow [eds.], *Chemical Oceanography*, v. 2, 2nd ed., Academic Press.
- Knoll, J.R. and F.A. Richards. 1969. A note on the sources of excess alkalinity in anoxic waters. *Deep-Sea Res.* 16: 205-212.
- Kushner, D.J. 1968. Halophilic bacteria, v. 10, p. 73-99. In W.W. Umbreit and D. Perlman [eds.], *Advances in applied microbiology*. Academic Press.
- Kushner, D.J. 1978. *Microbial life in extreme environments*. Academic Press. 465 p.
- Laking, P.N. 1974. *The Black Sea, its geology, chemistry, biology: a bibliography*. Woods Hole Oceanographic Institution. 368 p.
- Lamb, H. 1945. *Hydrodynamics*. Dover Publications, 371 p.
- LaRock, P.A., R.D. Lauer, J.R. Schwartz, K.K. Watanabe, and D.A. Wiesenburg. 1979. Microbial biomass and activity distribution in an anoxic, hypersaline basin. *Appl. Environ. Microbiol.* 37: 466-470.
- Larsen, H. 1967. Biochemical aspects of extreme halophilism, p. 97-132. In A.H. Rose and J.F. Wilkinson [eds.], *Advances in microbial physiology*. Academic Press.
- Lingane, J.J. and R.L. Pecsok. 1948. Preparation of standard chromous sulfate or chromous chloride solution of determinate concentration. *Anal. Chem.* 20: 425-428.
- Linnenbom, V.J. and J.W. Swinnerton. 1969. Distribution of low molecular weight hydrocarbons and excess molecular nitrogen in the Cariaco Trench, p. 125-130. In *Symposium on investigation and resources of the Caribbean Sea and adjacent regions*. Rep. 71.1, FAO Fisheries, Willemstad, Curacao.

- McAuliffe, C. 1971. GC determination of solutes by multiple phase equilibration. *Chem. Tech.* 1: 46-51.
- McKee, T.R., L.M. Jeffrey, B.J. Presley, and U.G. Whitehouse. 1978. Holocene sediment geochemistry of continental slope and intra-slope basin areas, northwest Gulf of Mexico, p. 313-326. In A.H. Bouma, G.T. Moore, and J.M. Coleman [eds.], *Framework, facies, and oil-trapping characteristics of the upper continental margin.* Amer. Assoc. Pet. Geol.
- McKee, T.R. and B.R. Sidner. 1976. An anoxic high salinity intra-slope basin in the northwest Gulf of Mexico, p. 125-131. In A.H. Bouma [ed.], *Beyond the Shelf Break.* AAPG Marine Geology Communications Short Course, Amer. Assoc. Pet. Geol.
- McKenna, E.J. and R.E. Kallio. 1965. The biology of hydrocarbons. *Annu. Rev. Microbiol.* 19: 183-208.
- McKinney, R.E. and R.A. Conway. 1957. Chemical oxygen in biological waste treatment. *Sewage Ind. Waste* 29: 1097-1106.
- Manheim, F.T. and J.L. Bischoff. 1969. Geochemistry of pore waters from Shell Oil Company drill holes on the continental slope of the northern Gulf of Mexico. *Chem. Geol.* 4: 63-82.
- Martens, C.S. and R.A. Berner. 1977. Interstitial water chemistry of anoxic Long Island Sound sediments. I. Dissolved gases. *Limnol. Oceanogr.* 22: 10-25.
- Martens, C.S., R.A. Berner, and J.K. Rosenfeld. 1978. Interstitial water chemistry of anoxic Long Island Sound sediments. 2. Nutrient regeneration and phosphate removal. *Limnol. Oceanogr.* 23: 605-617.
- Matsubara, T. and T. Mori. 1968. Studies on denitrification: IX. Nitrous oxide, its production and reduction to nitrogen. *J. Biochem.* 64: 863-871.
- Mehrbach, C., C.H. Culberson, J.E. Hawley, and R.M. Pytkowicz. 1973. Measurement of the apparent dissociation constants of carbonic acid in seawater at atmospheric pressure. *Limnol. Oceanogr.* 18: 897-907.
- Merrell, W.J., Jr. 1971. Wind-induced inertial oscillations in a linearly stratified sea. Ph.D. Dissertation, Texas A&M University, College Station, Texas. 97 p.
- Millero, F.J., A.L. Surdo, P. Chetirkin, and N.L. Guinasso. 1979. The density and speed of sound of Orca basin waters. *Limnol. Oceanogr.* 24(2): 218-225.

- Munk, W.H. 1966. Abyssal recipes. *Deep-Sea Res.* 13: 707-730.
- Murphy, J. and J.P. Riley. 1962. A modified single solution method for determination of phosphate in natural waters. *Anal. Chim. Acta.* 27: 31-36.
- Murray, J.W., V. Grundmanis, and W.M. Smithie, Jr. 1978. Interstitial water chemistry in the sediments of Saanich Inlet. *Geochim. Cosmochim. Acta.* 42: 1011-1026.
- Murray, R.G.E. and S.W. Watson. 1963. An organelle confined within the cell wall of Nitrosocystis oceanus (Watson). *Nature* 197: 211-212.
- Neumann, G. and W.J. Pierson, Jr. 1966. Principles of physical oceanography. Prentice-Hall, Inc. 545 p.
- Newman, F.C. 1976. Temperature steps in Lake Kivu: a bottom heated saline lake. *J. Phys. Oceanogr.* 6: 157-163.
- Nissenbaum, A. 1975. The microbiology and biogeochemistry of the Dead Sea. *Microb. Ecol.* 2: 139-161.
- Novelli, G.D. and C.E. Zobell. 1947. Assimilation of petroleum hydrocarbons by sulfate-reducing bacteria. *J. Bacteriol.* 47: 447-448 (abstract).
- Nowlin, W.D. Jr. 1972. Winter circulation patterns and property distributions, p. 3-51. In L.R.A. Capurro and J.L. Reid [eds.], Contributions on the physical oceanography of the Gulf of Mexico, v. 2, Gulf Publishing Co.
- Oremland, R.S. 1975. Methane production in shallow water, tropical marine sediments. *Appl. Microbiol.* 30: 602-608.
- Ostland, H.G. 1974. Expedition "Odysseus 65": radiocarbon age of Black Sea deep water, p. 127-132. In E.T. Degens and D.A. Ross [eds.], The Black Sea - geology, chemistry, and biology. Mem. 20, Amer. Assoc. Pet. Geol. George Banta Company.
- Park, K. 1969. Oceanographic CO<sub>2</sub> system: An evaluation of ten methods of investigation. *Limnol. Oceanog.* 14: 179-186.
- Payne, W.P., P.S. Riley, and C.P. Cox, Jr. 1970. Separate nitrite, nitric oxide, and nitrous oxide reducing fractions from Pseudomonas perfectomarinus. *J. Bacteriol.* 106: 356-36.
- Prabhakara, C., G. Dalu, and V.G. Kunde. 1974. A search for global and seasonal variation of methane from Nimbus 4 Iris measurements. *J. Geophys. Res.* 79: 1744-1749.

- Pytkowicz, R.M. and E. Atlas. 1975. Buffer intensity of seawater. *Limnol. Oceanogr.* 20: 222-229.
- Pytkowicz, R.M., S.E. Ingle, and C. Merbach. 1974. Invariance of apparent equilibrium constants with pH. *Limnol. Oceanogr.* 19: 665-669.
- Quayle, J.R. 1972. The metabolism of one-carbon compounds by microorganisms, p. 119-203. In A.H. Rose and D.W. Tempest [eds.], *Advances in microbial physiology*, v. 7, Academic Press.
- Ramm, A.E. and D.A. Bella. 1974. Sulfide production in anaerobic microcosms. *Limnol. Oceanogr.* 19: 110-118.
- Redfield, A.C. 1934. On the proportions of organic derivatives in sea water and their relation to the composition of plankton, p. 176-192. In James Johnstone Memorial Volume. University of Liverpool Press.
- Reeburgh, W.S. 1969. Observations of gases in Chesapeake Bay sediments. *Limnol. Oceanogr.* 14: 368-375.
- Reeburgh, W.S. 1976. Methane consumption in Cariaco Trench waters and sediments. *Earth Planet. Sci. Lett.* 28: 337-344.
- Richards, F.A. 1955. The Cariaco Trench. *Oceanus* 3: 9-10.
- Richards, F.A. 1957. Oxygen in the ocean, p. 185-238. In J.W. Hedgpeth [ed.], *Treatise on marine ecology and paleoecology*, v. 1, Mem. 67, Geol. Soc. Amer.
- Richards, F.A. 1965. Anoxic basins and fjords, p. 611-645. In J.P. Riley and G. Skirrow [eds.], *Chemical oceanography*, v. 1. Academic Press.
- Richards, F.A. 1975. The Cariaco Basin (Trench). *Oceanogr. Mar. Biol. Annu. Rev.* 13: 11-67.
- Richards, F.A. and B.B. Benson. 1961. Nitrogen/argon and nitrogen isotope ratios in two anaerobic environments, the Cariaco Trench in the Caribbean Sea and Dramsfjord, Norway. *Deep-Sea Res.* 7: 254-264.
- Richards, F.A., J.D. Cline, W.W. Broenkow, and L.P. Atkinson. 1965. Some consequences of the decomposition of organic matter in Lake Nitinat, an anoxic fjord. *Limnol. Oceanogr.* 10 (Suppl.): R185-R201.
- Rosenfeld, W.D. and S.R. Silverman. 1959. Carbon isotope fractionation in bacterial production of methane. *Science* 130: 1658-1659.

- Rosenfeld, W.S. 1947. Anaerobic oxidation of hydrocarbons by sulfate-reducing bacteria. *J. Bacteriol.* 54: 664-665.
- Rudd, J.W.M., R.D. Hamilton, and N.E.R. Campbell. 1974. Measurement of microbial oxidation of methane in lake water. *Limnol. Oceanogr.* 19: 519-524.
- Sackett, W.M., J.M. Brooks, B.B. Bernard, C.R. Schwab, H. Chung, and R.A. Parker. 1979. A carbon inventory for Orca Basin brines and sediments. *Earth Planet. Sci. Lett.* 44: 73-81.
- Sackett, W.M., S. Nakaparshin, and D. Dalrymple. 1970. Carbon isotope effects in methane production by thermal cracking, p. 37-53. In G.D. Hobson and G.C. Spears [eds.], *Advances in organic geochemistry*. 1966. Pergamon Press.
- Sass, E. and S. Ben-Yaakov. 1977. The carbonate system in hypersaline solutions: Dead Sea brines. *Mar. Chem.* 5: 183-199.
- Schmidt, U. 1978. The latitudinal and vertical distribution of molecular hydrogen in the troposphere. *J. Geophys. Res.* 83: 941-946.
- Schmidt, U. 1979. The solubility of carbon monoxide and hydrogen in water and sea-water at partial pressures of about  $10^{-5}$  atmospheres. *Tellus* 31: 68-74.
- Schoener, A. and G.T. Rowe. 1970. Pelagic *Sargassum* and its presence among the deep-sea benthos. *Deep-Sea Res.* 17: 923-925.
- Scranton, M.I. and P.G. Brewer. 1977. Occurrence of methane in the near-surface waters of the western subtropical North Atlantic. *Deep-Sea Res.* 24: 127-138.
- Shokes, R.F., P.K. Trabant, B.J. Presley, and D.F. Reid. 1977. Anoxic, hypersaline basin in the Northern Gulf of Mexico. *Science* 196: 1443-1446.
- Sillen, L.G. and A.E. Martell. 1964. Stability constants of metal-ion complexes. The Chemical Society (London). 754 p.
- Skirrow, G. 1965. The dissolved gases — carbon dioxide, p. 227-322. In J.P. Riley and G. Skirrow [eds.], *Chemical Oceanography*, v. 1, Academic Press.
- Solorzano, L. 1969. Determination of ammonia in natural waters by the phenylhypochlorite method. *Limnol. Oceanogr.* 14: 799-801.
- Sorokin, Y.I. 1957. On the ability of sulfate-reducing bacteria to utilize methane for the reduction of sulfates to hydrogen sulfide. *Dokl. Akad. Nauk. SSSR* 115: 816-818.



- Stadtman, T.C. and H.A. Barker. 1951. Studies on methane fermentation. IX. The origin of methane in acetate and methanol fermentation by Methanosarcina. J. Bacteriol. 61: 81-86.
- Stookey, L.L. 1970. Ferrozine - a new spectrophotometric reagent for iron. Anal. Chem. 42: 779-781.
- Sverdrup, H.U., M.W. Johnson, and R.H. Fleming. 1942. The oceans — their physics, chemistry, and general biology. Prentice-Hall. 1087 p.
- Sweeney, R.E. 1972. Pyritization during diagenesis of marine sediments. Ph.D. Thesis, University of California, Los Angeles. 194 p.
- Swinerton, J.W. and V.J. Linnenbom. 1967. Determination of C<sub>1</sub> to C<sub>4</sub> hydrocarbons in sea water by gas chromatography. J. Gas Chrom. 5: 570-573.
- Swinerton, J.W. and V.J. Linnenbom. 1969. Low molecular weight hydrocarbon analyses of Atlantis II waters, p. 251-253. In E.T. Degens and D.A. Ross [eds.], Hot brines and recent heavy metal deposits in the Red Sea. Springer-Verlag.
- Swinerton, J.W., V.J. Linnenbom, and C.H. Cheek. 1962. Determination of dissolved gases in aqueous solutions by gas chromatography. Anal. Chem. 34: 483-485.
- Technicon. 1976. Ammonia in water and sea water. Technicon Industrial Systems, Industrial Methods No. 154-71W, 2 p.
- Thayer, L.A. 1931. Bacterial genesis of hydrocarbons from fatty acids. Bull. Amer. Assoc. Pet. Geol. 15: 441-453.
- Thorstenson, D.C. and F.T. MacKenzie. 1974. Time variability of pore water chemistry in recent carbonate sediments, Devil's Hole, Harrington Sound, Bermuda. Geochim. Cosmochim. Acta. 38: 1-19.
- Tompkins, R.E. and L.E. Shephard. 1979. Orca Basin: Depositional processes, geotechnical properties, and clay mineralogy of holocene sediments within an anoxic hypersaline basin, northwest Gulf of Mexico. Mar. Geol. 33: 221-238.
- Trabant, P.K. and B.J. Presley. 1978. Orca Basin, an anoxic pool on the continental slope, Northwest Gulf of Mexico, p. 303-311. In A.H. Bouma, G.T. Moore, and J.M. Coleman [eds.], Framework, facies, and oil-trapping characteristics of the upper continental margin. AAPG Studies in Geology No. 7.

- Trüper, H.G. 1969. Bacterial sulfate reduction in the Red Sea hot brines, p. 263-271. In E.T. Degens and D.A. Ross [eds.], Hot brines and recent heavy metal deposits in the Red Sea. Springer-Verlag.
- Turner, J.S. 1965. The coupled turbulent transport of salt and heat across a sharp density interface. *Inter. J. Heat Mass Trans.* 8: 759-767.
- Turner, J.S. 1968. The behaviour of a stable salinity gradient heated from below. *J. Fluid Mech.* 33: 183-200.
- Turner, J.S. 1973. Buoyancy effects in fluids. University Press, Cambridge. 367 p.
- Turner, J.S. and H. Stommel. 1964. A new case of convection in the presence of combined vertical salinity and temperature gradients. *Proc. U.S. Nat. Acad. Sci.* 52: 49-53.
- Vaccaro, R.F. 1965. Inorganic nitrogen in sea water, p. 365-408. In J.P. Riley and G. Skirrow [eds.], *Chemical Oceanography*, v. 1, Academic Press.
- Valley, S.L. 1965. Handbook of geophysics and space environments. McGraw-Hill. 683 p.
- Valyashko, M.G. 1956. Geochemistry of bromine in processes of halogenesis and use of bromine content as a genetic and prospecting criterion. *Geochemistry* 6: 33-49.
- Vary, P.S. and M.J. Johnson. 1967. Cell yields of bacteria grown on methane. *Appl. Microbiol.* 15: 1473-1478.
- Vogel, A.I. 1968. Quantitative inorganic analysis. John Wiley & Sons. 446 p.
- Watson, S.W. and J.B. Waterbury. 1969. The sterile hot brines of the Red Sea, p. 272-281. In E.T. Degens and D.A. Ross [eds.], Hot brines and recent heavy metal deposits in the Red Sea. Springer-Verlag.
- Weast, R.C. 1969. Handbook of chemistry and physics, 50th ed., Chemical Rubber Co., Cleveland, Ohio. 2356 p.
- Weatherburn, M.W. 1967. Phenol-hypochlorite reaction for determination of ammonia. *Anal. Chem.* 39: 971-974.
- Weiss, R.F. 1968. Piggyback samples for dissolved gas studies on sealed water samples. *Deep-Sea Res.* 15: 695-699.

- Weiss, R.F. 1969. Dissolved nitrogen, argon, and total carbonate in the Red Sea brines, p. 254-262. In E.T. Degens and D.A. Ross [eds.], Hot brines and recent heavy metal deposits in the Red Sea. Springer-Verlag.
- Weiss, R.F. 1970. The solubility of nitrogen, oxygen, and argon in water and seawater. *Deep-Sea Res.* 17: 721-735.
- Weiss, R.F. 1971. Solubility of helium and neon in water and seawater. *J. Chem. Eng. Data* 16: 235-241.
- Weiss, R.F. 1974. Carbon dioxide in water and seawater: the solubility of a non-ideal gas. *Mar. Chem.* 2: 203-215.
- Weiss, M.F. and H. Craig. 1973. Precise shipboard determination of dissolved nitrogen, oxygen, argon, and total inorganic carbon by gas chromatography. *Deep-Sea Res.* 20: 291-303.
- Weiss, R.F. and T.K. Kyser. 1978. Solubility of krypton in water and seawater. *J. Chem. Eng. Data* 23: 69-72.
- Weiss, R.F. and B.A. Price. 1980. Nitrous oxide solubility in water and seawater. *Mar. Chem.* 8: 347-359.
- Wertlieb, D. and W. Vishniac. 1967. Methane utilization by a strain of Rhodopseudomonas gelatinosa. *J. Bacteriol.* 93: 1772-1774.
- Weyl, P.K. 1961. The carbonate saturometer. *J. Geol.* 69: 32-44.
- Wiesenburg, D.A. 1975. Processes controlling the distribution of methane in the Cariaco Trench, Venezuela. M.S. Thesis, Old Dominion University, Norfolk, Va. 106 p.
- Wiesenburg, D.A., L.A. Barnard, and D.R. Schink. 1977. Nitrogen in the Orca Basin: effects of denitrification at the brine-sea water interface. *EOS, Transactions, Amer. Geophys. Union.* 58: 1175.
- Wiesenburg, D.A. and N.L. Guinasso, Jr. 1979. Equilibrium solubilities of methane, carbon monoxide, and hydrogen in water and sea water. *J. Chem. Eng. Data* 24: 356-360.
- Wilson, T.R.S. 1978. Evidence for denitrification in aerobic pelagic sediments. *Nature* 274: 354-356.
- Winkler, L.W. 1888. Die Bestimmung des im Wasser gelosten Sauerstoffes. *Chem. Ber.* 21: 2843-2855.
- Yamamoto, S., J.B. Alcauskas, and T.E. Crozier. 1976. Solubility of methane in distilled water and seawater. *J. Chem. Eng. Data* 21: 78-80.

- Zobell, C.E. 1942. Changes produced by microorganisms in sediments after deposition. *J. Sed. Petrol.* 12: 127-136.
- Zobell, C.E. 1946. Marine microbiology. Chronica Botanica Co. 240 p.

## VITA

Denis Alan Wiesenbure was born to Otto Karl Wiesenbure and Denise Higginbotham in Pascagoula, Mississippi on November 4, 1948. He attended parochial and public schools in Pascagoula, Mississippi, and graduated from Pascagoula High School in May, 1966.

He attended Duke University in Durham, North Carolina and received a Bachelor of Arts degree in Chemistry in June, 1970. He attended Old Dominion University in Norfolk, Virginia, from 1970 to 1974 and received a Masters of Science degree in Oceanography in August, 1975. From September, 1974, to June, 1975, he was enrolled at Dalhousie University in Halifax, Nova Scotia, Canada, where he studied chemical oceanography.

In 1975, he came to Texas A&M University as a graduate student in the Oceanography Department. He currently resides with his wife, Jean Herbert, and his son, Heath Karlton, at 1005 Milner Drive, College Station, Texas.

The typist for this dissertation was Bee Gee Hart.

