Reprinte; from the AMERICAN JOURNAL OF VETERINARY RESEARCH, Vol.41, No.3, Pages: 399-404 © American Veterinary Medical Association, 1980. All Rights Reserved.

A090912

6



# SELECTED REPORTS AND NOTES

# Modification of Lethality Induced by Staphylococcal Enterotoxin B in Dutch Rabbits,

C. T. Liur Brand R. P. Sanders

## SUMMARY

Intramuscular injection of staphylococcal enterotoxin B (SEB) at a dosage level of 50  $\mu$ g/kg of body weight caused death in Dutch rabbits. Lethality was not modified markedly by morphine pretreatment or by hyperthermia, thyrotoxicosis, propylthiouracil feeding, thyroparathyroidectomy, water deprivation, or fasting. The administration of acetylsalicylic acid to the sEB-inoculated rabbit also failed to protect the rabbits from the effect of SEB. Seemingly, the SEB molecular destruction was not markedly modified by alteration of cellular metabolism, and lethal effects of SEB remained unchanged in the morphineor acetylsalicylic acid-treated rabbits.

When SEB was given to six rabbits 3 days after total-body X-irradiation, fever persisted and three rabbits survived. An identical dose of SEB to nonirradiated rabbits produced fever initially, followed by hypothermia and death of all six rabbits. A single IV injection of a small quantity of purified staphylococcal enterotoxin B (SEB) has been shown to produce death within 24 hours in monkeys,<sup>1-3</sup> dogs,<sup>4,5</sup> and rabbits.<sup>6-7</sup> Although an enteric endotoxin release was suggested as the cause of death in



interest to scientists searching for information to develop means of treating SEB toxemia.

Staphylococcal enterotoxin B is a single polypeptide chain with a molecular weight of 35,300.<sup>12</sup> Warren et al<sup>13</sup> proposed that the closed disulfide loop

TABLE 1—Allocation of 84 Dutch Rabbits into 6 Groups and 12 Subgroups for Studying Modifications of Staphylococcal Enterotoxin B (SEB) Toxemia

Group	(n)	Treatment	Days prior to SEB (100 µg/kg of body weight)	
I-Control*				
la	(4)	Rectal temperature = 38 to 39 C	ND	
Ib	(6)	0.5 ml of 0.02 N NaOH/rabbit/day	6	
Ic	(6)	None	ND	
Id	(6)	None	ND	
Ie	(5)	SEB (50 µg/kg), IM	NA	
lf	(6)	None	ND	
II-Hypermetabolic				
IIa Hyperthermia	(4)	Rectal temperature = 40 to 40.5 C	0.5	
IIb Thyroxine treated	(6)	2 mg/rabbit/day	6	
III-Hypometabolic				
IIIa PTU treated	(5)	25 mg/rabbit/day	70	
IIIb Thyroparathyroidectomized	(6)	Surgically removed	4	
IIIc H <sub>2</sub> O deprived	(6)	Without water	4	
IIId Fasted	(6)	Without food	4	
IV-Morphine treated	(6)	3 mg/rabbit/6 hours	3	
V-Acetylsalicylic acid treated	(6)	217 mg/rabbit/6 hours for 48 hours, be- gun 15 minutes after sgs (50 μg/kg)		
VI-X-irradiated	(6)	500 R total-body exposure	3	

\* Corresponding control vs experimental groups = Ia vs IIa; Ib vs IIb; Ic vs IIIa; Id vs IIIb, IIIc, IIId, and IV; le vs V; If vs VI.

A CALL REAL PROPERTY AND A CALL REAL

PTU = propylthiouracil; NA = not applicable; (n) = number. ND = not done.

rabbits,<sup>7</sup> reduction of body fluid volume, blood pooling in the peripheral capillaries,<sup>3,6</sup> pulmonary edema,<sup>9,10</sup> and activation of the kinin system<sup>11</sup> were considered to be associated with sEB toxemia and death in rhesus macaques. How to modify sEB toxicity in the monkey and rabbit is still of much of SEB is related to stabilization of its native chemical structure. The complete amino acid structure of the SEB molecule was reviewed and summarized by Beisel.<sup>14</sup> Because a major portion of the IV-injected dose of labeled SEB accumulates in the renal proximal tubule.<sup>15,16</sup> the kidney<sup>17</sup> has been sug-

399

Received for publication June 4, 1979. Prom the US Army Medical Research Institute of Infectious Diseases, Fort Detrick, Frederick, MD 21701.

A portion of the work was presented at the Federation of American Society for Experimental Biology meeting, Chicago, April 1977 (*Fed Proc* 36:610, 1977). The authors thank Dr. C. L. Hadick for thyroparathyroids-tomy and Measra. R. D. DeLauter and G. Crone for technical amistance.

March, 1980

FILE COPY

- 17 RO 10 27 11



Fig 1—Changes in  $O_2$  consumption, body weight, and rectal temperature of control rabbits (n = 6; group Ic). Time = 22 days.



Fig 2—Effects of water deprivation and fasting on  $O_2$  consumption and body weight of rabbits (n = 6/group; groups life and lild).

gested as the site for SEB catabolism in vivo. Results from in vitro studies<sup>18,19</sup> indicate that lysosomal protease might be responsible for SEB catabolism. However, Canonico et al<sup>20</sup> suggested that SEB was first pinocytized by renal tubular cells and subsequently was sequestered within vesicles having physical and biochemical properties of lysosomes.

Treatment of monkeys<sup>21</sup> or rabbits<sup>22</sup> with a single total-body X-irra-

diation (400 to 500 R) 4 days prior to lethal doses of SEB inoculation prolonged survival. The exact mechanism for the modification of SEB toxicity is not known. Because O<sub>2</sub> consumption decreased markedly after a nonlethal total-body X-ray exposure (500 R) in rabbits,22 the present study was designed to test the hypothesis that experimentally induced hypometabolism (propylthiouracil, thyroparathyroidectomy, water deprivation, and fasting) or hypermetabolism (hyperthermia and thyroxine) would modify SEB toxicity and alter lethality. Other purposes of this study were: (1) to investigate the usefulness of morphine as a sedative<sup>23</sup> for alleviation of pul-monary edema,<sup>24,25</sup> and (2) to see if acetylsalicylic acid would inhibit synthesis of prostaglandins,<sup>26</sup> which might have a major role in causing death<sup>27,28</sup> in the SEB-inoculated rabhits

# Materials and Methods

Male Dutch rabbits (n = 84; Oryctolagus cuniculus), weighing 1.8 to 2.2 kg, were placed in six experimental groups (Table 1). Groups I, II, and III were further allocated among 12 subgroups, of which six subgroups were used as controls. Several different experimental conditions with various durations were induced prior to a single IM inoculation of highly purified SEB<sup>29</sup> (0.1 mg/kg). Acetylsalicylic acid (217 mg/rabbit) was given orally 15 minutes after IM injection of 50 µg of seB/kg and was continued every 6 hours for 2 days (group V). All measurements were made in the same period under the same experimental conditions for control and corresponding treatment groups of rabbits.

Induction of Hypermetabolism-Hyperthermia (group IIa) was induced by placing conscious rabbits into a modified ventilated incubator with an internal temperature of 34 C for 12 hours until the rectal temperature of the rabbits reached 40 to 40.5 C. Rectal temperature was measured with a probe, and values were read on a telethermometer." L-Thyroxine<sup>b</sup> was dissolved in 0.02 N NaOH<sup>30</sup> (4 mg/ml); a daily dose of thyroxine (2 mg/rabbit/day in a 0.5 ml volume) was administered subcutaneously (sc) for 6 days (group IIb). The corresponding controls (group Ib) were given the same volume of 0.02 N NaOH without L-thyroxine, SC.

Induction of Hypometabolism-Induction of various types of hypometabo-

<sup>h</sup> Sigma Chemical Co, St Louis, Mo.

Charles to the State State State State State

Am J Vet Res, Vol 41, No. 3

400

<sup>\*</sup> Yellow Springs Instrument Co, Yellow Springs, Ohio.



Fig 3—Effect of thyroparathyroidectomy (postthyroid) on  $O_2$  consumption, body weight, and rectal temperature of rabbits (n = 6; group IIIb).

lism was attempted. Hypometabolism was by thyroparathyroidectomy induced (group IIIb), water deprivation (group IIIc), or fasting (group IIId) for 4 days prior to SEB inoculation (0.1 mg/kg, 1M). Also, a group of rabbits was fed propylthiouracil (PTU; group IIIa) for 70 days and then were inoculated IM with SEB (0.1 mg/kg). The dose of PTU was 25 mg/rabbit/day; the suspension was given orally through a nasogastric tube, or the tubing was inserted directly into the stomach via the mouth. In the latter case, a metal mouth opener was used to prevent biting.

Morphinization of Rabbits—Six rabbits were given morphine sc (3 mg/rabbit) every 6 hours for 3 days (group IV), and then a single IM dose of 100  $\mu$ g of SEB/kg was given.

Total-body O<sub>2</sub> consumption of the rabbit was measured with a 1-L spirometer.<sup>6</sup> The rabbit was restrained in a plastic box during the measurement of  $O_2$  consumption for at least 15 minutes' conditioning and 9 minutes' actual recording. The measured volume of  $O_2$  consumption was corrected with standard atmospheric pressure, temperature, and vapor pressure according to the gas law.

Total-Body X-Irradiation—The radiation exposure (500 R) was delivered from a 1-meV, 3-mA X-ray-generating unit<sup>4</sup> operating at an effective energy of 475 kiloelectron volt without added filtration. Before irradiation, each rabbit was anesthetized with droperidol and fentanyl<sup>\*</sup> (0.15 ml/kg, IM) and was placed in a plastic acrylic resin cylinder (length, 49.5 cm; diameter, 13.6 cm), which was rotated at 1 rpm in the X-ray beam. The midline ex-

\* Innovar-Vet, Pitman-Moore Inc, Washington Crossing, NJ.

posure rate was 30 R/minute in air at a distance of 118 cm.

A lethal dose of SEB (0.1 mg/kg) was injected IM into nonirradiated (group If) and irradiated (group VI) rabbits.

Recorded Data—The lethality or survival time was recorded for all groups of rabbits studied. Body weight,  $O_2$  consumption, and rectal temperature were measured only in the L-thyroxine-treated, PTU-treated, thyroparathyroidectomized, water-deprived and fasted rabbits, and in their corresponding control groups.

Rectal temperatures were measured with a probe and telethermometer before and after SEB inoculation at various time intervals for a period of 20 hours; deaths were recorded.

# **Results**

Effects of Water Deprivation, Fasting, Thyroparathyroidectomy, or PTU—Typical changes in  $O_2$  consumption, body weight, and rectal temperature for 22 days in normal control Dutch rabbits are illustrated (Fig 1). A steady increase in body weights, with few changes in O<sub>2</sub> consumption and rectal temperature, was shown. However, O<sub>2</sub> consumption and body weights were significantly (P <0.05) decreased 4 days after water deprivation or fasting, as compared with base-line values for the controls (Fig 2). After thyroparathyroidectomy, body weight increased markedly, and O<sub>2</sub> consumption was decreased without change in rectal temperature (Fig 3). Although body weight increased markedly between 8 and 10 weeks after an initial inhibition of growth in the PTU-treated rabbits, O<sub>2</sub> consumption and rectal temperature did not show marked changes during a 10-week PTU-feeding period (Fig 4).

Effects of *L*-Thyroxine—Effects of *L*-thyroxine on body weight and rectal temperature in rabbits are illustrated (Fig 5). Within 6 days, body weights of *L*-thyroxine-injected rabbits sharply decreased, whereas the

TABLE 2—Effect of Hyperthermia on see-Inoculated (0.1 mg/kg) Rabbits

Group	Hyperthermia 12 hours before SEB	Hyperthermia 4 hours after SEB
Control (Ia) (Room tempera- ture)	4/4	3/4
Hyperthermia (lla)	•/•	174
(40.0-40.5 C)	4/4	4/4
Numerator -	No of deaths de	nominatur - N

of rabbits exposed.

401

<sup>&#</sup>x27; Warren E. Collins Inc. Braintree, Mass.

<sup>&</sup>lt;sup>d</sup> General Electric Co, Milwaukee, Wis.



Fig 4—Effect of oral propylthiouracil (25 mg/day) on  $O_2$  consumption, rectal temperature, and body weight in rabbits (n = 5; group Illa) for 10 weeks.



Fig 5—Changes in body weight and rectal temperature of control and L-thyroxine-treated rabbits (group IIb).

control group showed a slight increase in body weight. The rectal temperatures among control and L-thyroxinetreated rabbits were not significantly different (P > 0.05). Effects of SEB in Hypermetabolic Rabbits—The effect of hyperthermia and L-thyroxine administration on SEB-exposed rabbits is summarized (Tables 2 and 3). Whether hyperthermia was applied before or after SEB, the number of deaths was essentially the same as compared with the number of deaths of controls (Table 2). Although all rabbits in the thyroxinetreated group died, control rabbits, which were given 0.5 ml of 0.02 NNaOH/day for 6 days, survived. The failure of these control animals to respond in the same way as did the other controls is not explained and is presumed to have occurred fortuitously.

Effects of SEB in Hypometabolic, Morphinized, and PTU-Treated Rabbits—Effects of SEB (0.1 mg/kg) on survival in hypometabolic (induced by thyroparathyroidectomy, water deprivation, and fasting) and morphinized rabbits are summarized (Tables 4 and 5). Significant difference was not observed in survival between control groups and each of the corresponding experimental groups.

Effects of SEB in X-Irradiated Rabbits-Changes in rectal temperature of irradiated and nonirradiated rabbits given SEB (100  $\mu$ g/kg) 3 days after irradiation are illustrated (Fig 6). Fever was observed within 2 hours after SEB inoculation in both groups and persisted in the irradiated rabbits: decreased rectal temperatures below base-line values were found in nonirradiated rabbits. Although there was a trend for the rectal temperature to reach preinoculation values between 8 and 10 hours after SEB, all six nonirradiated rabbits died within 10 hours. However, three of six irradiated rabbits survived 20 hours after the **SEB** injection.

## Discussion

The cause of SEB-induced death in rabbits has been attributed to endotoxin release from the intestine.<sup>7</sup> Pulmonary edema has been shown to be an important factor leading to death during SEB toxemia in rhesus macaques.<sup>10</sup> Further, Pettit et al<sup>11</sup> reported that the kinin system was activated in monkeys after a lethal IV dose of SEB. In the present study, acetylsalicylic acid failed to modify survival time after IM inoculation of SEB in rabbits, suggesting that prostaglandins may not be directly involved in the pathogenesis of shock and death.

Propylthiouracil is an antithyroid drug which decreases the formation of thyroid hormones by preventing iodi-

Am J Vet Res, Vol 41, No. 3

TABLE 3—Effect of Hypermetabolism (Induced by L-Thyroxine) on SEB Challenge-Exposed (0.1 mg/kg) Rabbits (Groups Ib and IIb)

Variable	Ib (contr	ols; n = 6)	IIb (thyroxine treated; n =		
	Day 0	Day 6	Day 0	Day 6	
Rectal temperature (C)	38.8 ± 0.1	39.2 ± 0.2	39.3 ± 0.1	$39.3 \pm 0.2$	
O <sub>2</sub> consumption (L/hours/m <sup>2</sup> )	7.0 ± 0.5	$9.3 \pm 1.0$	$7.8 \pm 0.2$	$14.2 \pm 0.3^{\circ}$	
Body weight (kg)	1.59 ± 1.10	$1.62 \pm 0.09$	1.67 ± 0.06	1.25 ± 0.05*	
Survival after SEB on day 6 (%)		100		0*	

• P < 0.05 vs controls.

Controls were given 0.5 ml of 0.02 N NaOH/day for 6 days. Thyroxine dosage = 2 mg/rabbit/day sc, in 0.5 ml of 0.02 N NaOH. n = No. of rabbits.

TABLE 4—Effects of Hypometabolism on SEB	Challenge-Exposed (0.1 mg	/kg) Rabbits (6/group)
--	---------------------------	------------------------

	O <sub>2</sub> consumption (L/hour/m <sup>2</sup> )				
Treatment	Prior to treat- ment	Prior to ses	SEB given after procedures (days)	Survival time* (hours)	Survival (%)
Control	6.08 ± 0.26	6.53 ± 0.16	ND	15 ± 3.4	17
Thyroparathyroidectomized	6.64 ± 0.56	$4.24 \pm 0.31$	4	$20.6 \pm 3.4$	33
H <sub>2</sub> O deprived	$7.84 \pm 0.42$	6.07 ± 0.49	4	$12.3 \pm 2.0$	0
Fasting	$7.68 \pm 0.69$	$5.83 \pm 0.16$	4	62 ± 47.1	50

\* Refers only to rabbits that died.

ND = not done.

ł

TABLE 5-Effects of SEB (0.1 mg/kg) on Survival of Rabbits Given Morphine or Propylthiouracil

_	O <sub>2</sub> consumption	(L/hour/m²)	Days after medica- tion	Time to death* (hours)	Survival (%)
Treatment (n)	Prior to treatment	Prior to ses			
Control (6)	6.08 ± 0.26	6.53 ± 0.16	ND	19.4 ± 3.1	17
Morphine (5)	ND	ND	3	11.3 ± 1.25	20
PTU (5)	6.32 ± 0.43	6.21 ± 0.59	70	$15.5 \pm 2.5$	0

\* Only for rabbits that died.

ND = not done; PTU = propylthiouracil.



Fig 6—Changes in rectal temperature of total-body X-irradiated and nonirradiated rabbits given staphylococcal enterotoxin B (100  $\mu$ g/kg) 3 days after irradiation (n = 6/group; group If and VI).

nation of tyrosine.<sup>31</sup> In the present study, daily oral feeding of PTU for a period of 70 days did not alter  $O_2$  consumption and rectal temperature in Dutch rabbits. Only the body

growth rate was inhibited during the first 3 weeks after initiation of PTU feeding. Because few changes occurred in the PTU-treated rabbits, it was not surprising to observe that PTU (25 mg/day) failed to modify SEB toxemia in rabbits with the dose schedule used.

If pulmonary edema is the main cause of death in SEB-intoxicated rabbits, pretreatment with morphine for 4 days might exert some beneficial effects by the sedative action and decrease in cardiac load.<sup>23,24</sup> According to our recent findings,<sup>f</sup> pulmonary edema did not occur at death of SEBinoculated rabbits. Furthermore, Pettit et al<sup>7</sup> showed that endotoxemia was induced after SEB injection IV. This may explain why morphine did not have any protective action against lethal SEB toxemia in Dutch rabbi\*s.

Experimentally induced hypermetabolism, including hyperthermia and pretreatment with L-thyroxine, did not alter SEB lethality in rabbits. These results indicate that SEB molecules are not easily catabolized or detoxified in the body, even with increased cellular metabolism. The findings that hypometabolism failed to prolong survival during SEB toxemia suggested that a decrease in O<sub>2</sub> consumption alone after 500 R of totalbody X-irradiation<sup>22</sup> was not the key factor for survival in Dutch rabbits after an IM inoculation of SEB.

Rapoport et al<sup>32</sup> demonstrated that the administration of antiserum to SEB in rhesus macaques prevented a rapid disappearance of *iv-injected* SEB from the circulation. However, Beisel<sup>8</sup> pointed out that if macaques were hyperimmunized by repeated IV injections of an SEB antigen, typical anaphylactic reactions developed. Liu et al<sup>21,g</sup> reported that pretreatment with 400 R of total-body X-irradiation and continuous positive pressure were beneficial to rhesus macaques after an IV inoculation of SEB. Recently, a system of hemoperfusion through an activated charcoal column has been developed by Liu and Sanders.<sup>h</sup> The adsorptive capacity of activated charcoal to SEB in vitro and in rhesus macaques has also been shown.

Prevention of SEB-induced death by pretreatment with total-body Xirradiation (500 R) has been shown in the present study and in previous studies.<sup>22</sup> Mechanisms of survival appeared to be correlated with the abil-

<sup>r</sup> Unpublished data, 1977.

March, 1980

403

<sup>&</sup>lt;sup>4</sup> Liu CT, DeLauter RD, Griffin MJ: Pulmonary dysfunction in rhesus monkeys during staphylococcal B enterotoxemia (abstr). *Physiologist* 19:273, 1976.

<sup>&</sup>lt;sup>h</sup> Liu CT, Sanders RP: Development of a hemoperfusion system with activated charcoal for rhesus monkeys (abstr). Fed Proc 38:538, 1979.

ity to maintain fever after IM injection of SEB. Sustained fever after SEB challenge exposure may imply that SEBinduced fever in the irradiated host may originate centrally due to modification of hypothalamus functions. One of the possible reasons for survival with pretreatment of X-irradiation may be a result of sustained fever. because fever has been demonstrated to increase survival in the lizard and other animal models.<sup>33-35</sup> However, experimentally induced hyperthermia, by increasing rectal temperature 1 C. had no effect on survival during SEB toxemia in Dutch rabbits.

## References

1. Liu CT, Griffin MJ, Faulkner RT: Effect of staphylococcal enterotoxin B on body fluid compartments in conscious rhesus monkeys. J Med Primatol 5:336-344, 1976.

2. Liu CT, DeLauter RD, Faulkner RT: Cardiovascular and hepatic responses of rhesus macaques to staphylococcal enterotoxin B. Am J Vet Res 38:1849-1854, 1977.

3. Elsberry DD, Rhoda DA, Beisel WR: Hemodynamics of staphylococcal B enterotoxemia and other types of shock in monkeys. J Appl Physiol 27:164-169, 1969.

4. Skinner DG, Hayes MA: Effect of staphylococcal toxin on renal function: Irreversible shock. Ann Surg 162:161-180, 1965.

5. Prohaska JV, Kocandrle V, Houttuin E: Hemodynamic changes in response to staphylococcal endotoxin and enterotoxin. Bull Soc Int Chir 25:727-736, 1966.

6. Israel J, Oldstone M, Levenson S, et al: Mechanism of action of staphylococcal toxin in rabbits. *Proc Soc Exp Biol Med* 108:709-711, 1961.

7. Pettit GW, Elwell MR, Jahrling PB: Possible endotoxemia in rabbits after intravenous injection of *Staphylococcus aureus* enterotoxin B. *J Infect Dis* 135:646-648, 1977.

8. Beisel WR: Pathophysiology of staph-

after intravenous administration to monkeys. Toxicon 10:433-440, 1972.

9. Finegold MJ: Interstitial pulmonary edema. An electron microscopic study of the pathology of staphylococcal enterotoxemia in rhesus monkeys. *Lab Invest* 16:912-924, 1967.

10. Liu CT, DeLauter RD, Griffin MJ, et al: Effect of staphylococcal enterotoxin B on functional and biochemical changes of the lung in rhesus monkeys. *Toxicon* 16:543-550, 1978. 11. Pettit GW, Yamada T, Wing DA, et al:

 Pettit GW, Yamada T, Wing DA, et al: Enterotoxic shock in rhesus monkeys. The role of selected bloodborne factors. *Circ Res* 43:398– 405, 1978.

12. Spero L, Stefanye D, Brecher PI, et al: Amino acid composition and terminal amino acids of staphylococcal enterotoxin B. *Biochemistry* 4:1024-1030, 1965.

13. Warren JR, Spero L, Metzger JF: Stabilization of native structure by the closed disulfide loop of staphylococcal enterotoxin B. Buochim Biophys Acta 359:351-363, 1974.

14. Beisel WR: Enterotoxin-mediated diseases, in Newberne PM (ed): *Trace Substances* and *Health*. New York, Marcel Dekker Inc, 1976, pp 1-66.

15. Morris EL, Hodoval LF, Beisel WR: The unusual role of the kidney during intoxication of monkeys by intravenous staphylococcal enterotoxin B. *J Infect Dis* 117:273-284, 1967.

16. Normann SJ, Jaeger RF, Johnsey RT: Pathology of experimental enterotoxemia. The *in vivo* localization of staphylococcal enterotoxin B. *Lab Invest* 20:17-25, 1969.

17. Normann SJ: Renal fate of staphylococcal enterotoxin B. Lab Invest 25:126-132, 1971.

18. Canonico PG: Lysosomal catabolism of a protein toxin: Staphylococcal enterotoxin B. Biochim Biophys Acta 322:251-257, 1973.

19. Normann SJ, Stone CM: Renal lysosomal catabolism of staphylococcal enterotoxin B. Lab Invest 27:236-241, 1972.

 Canonico PG, Henriksen EL, Ayala E, et al: Subcellular localization of staphylococcal enterotoxin B in rabbit kidney. Am J Physiol 226;J333-1337, 1974.
Liu CT, Griffin MJ, Hilmas DE: Effect

21. Liu CT, Griffin MJ, Hilmas DE: Effect of staphylococcal enterotoxin B on cardiorenal functions and survival in x-irradiated rhesus macaques. Am J Vet Res 39:1213-1217, 1978. 22. Liu CT, Hilmas DE: Effect of x-irradiation on survivai of rabbits with staphylococcal B enterotoxemia. *Radiat Res* 76:402-409, 1978.

23. Wikler A: Sites and mechanisms of action of morphine and related drugs in the central nervous system. *Pharmacol Rev* 2:435-506, 1950.

24. Rodbard S: Bronchomotor tone. A neglected factor in the regulation of the pulmonary circulation. Am J Med 15:356-367, 1953.

25. Murphree HB: Narcotic analgesics 1: Opium alkaloids, in DiPalma JR (ed): Drill's Pharmacology in Medicine, ed 4. New York, McGraw-Hill Book Co, 1971, pp 342-343.

26. Hammond AL: Aspirin: New perspective on everyman's medicine. Science 174:48, 1971.

27. Greenberg S, Englebrecht JA, Wilson WR: Cardiovascular pharmacology of prostaglandin  $B_1$  and  $B_2$  in the intact dog. *Proc Soc Exp Biol Med* 143:1008-1013, 1973.

28. Flynn JT, Appert HE, Howard JM: Arterial prostaglandin  $A_1$ ,  $E_1$ , and  $F_{2a}$  concentrations during hemorrhagic shock in the dog. *Circ Shock* 2:155-163, 1975.

29. Schantz EJ, Roessler WG, Wagman J, et al: Purification of staphylococcal enterotoxin B. Biochemistry 4:1011-1016, 1965.

30. Liu CT, Overman RR: Effect of toxic doses of L-thyroxine on tissue water, electrolytes and plasma protein in rats. *Proc Soc Exp Biol Med* 117:232-236, 1964.

31. Astwood EB: Thyroid and antithyroid drugs, in Goodman LS, Gilman A (ed): The Pharmacological Basis of Therapeutics, ed 3. New York, The Macmillan Co, 1966, pp 1484-1491.

32. Rapoport MI, Hodoval LF, Grogan EW, et al: The influence of specific antibody on the disappearance of staphylococcal enterotoxin B from blood. *J Clin Invest* 45:1365-1372, 1966.

33. Kluger MJ, Ringler DH, Anver MR: Fever and survival. Science 188:166-168, 1975.

34. Bernheim HA, Kluger MJ: Fever: Effect of drug-induced antipyresis on survival. Science 193:237-239, 1976.

35. Haahr S, Mogenson S: Function of fever in infectious disease. *Biomedicine* 28:305-307, 1978.



Am J Vet Res, Vol 41, No. 3

- ALASSA

ACC STREET