

AD-A034 645

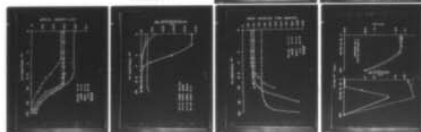
ARMY MEDICAL RESEARCH INST OF INFECTIOUS DISEASES FR--ETC F/G 6/13
EXTREME SENSITIVITY OF STAPHYLOCOCCAL ENTEROTOXIN B AND C PRODU--ETC(U)
JAN 77 R A ALTENBERN

UNCLASSIFIED

NL

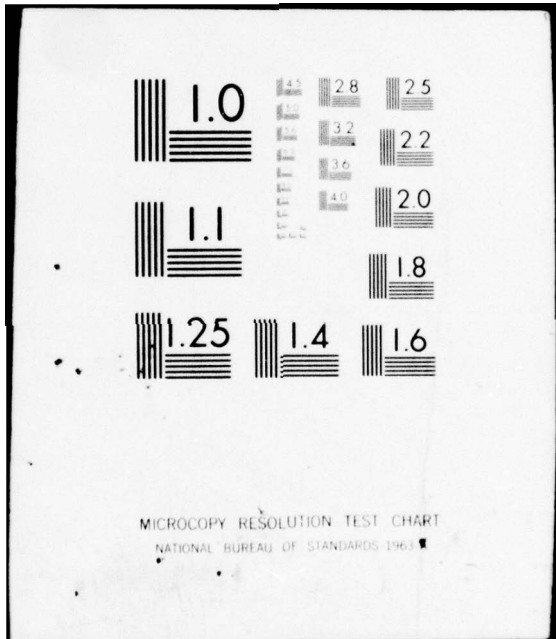
| OF |

AD
A034645



END

DATE
FILMED
2-77



MICROCOPY RESOLUTION TEST CHART
NATIONAL BUREAU OF STANDARDS 1963-A

ADA034645

2
B.S.

Extreme Sensitivity of Staphylococcal Enterotoxin B and C
Production to Inhibition by Cerulenin

ROBERT A. ALTENBERN

U.S. Army Medical Research Institute of
Infectious Diseases, Frederick, Maryland 21701

JAN 1977

D D C
D D C
JAN 19 1977
REGULATED
C

MEMORANDUM for	Wife Section <input checked="" type="checkbox"/>
ATIS	Self Section <input type="checkbox"/>
DDC	
UNANNOUNCED	
JUSTIFICATION	
BY	INSTRUCTIONS/AVAILABILITY CODES
SIG.	ATIS/REG. OF SPECIAL
A	

1473

UNCLASSIFIED

SECURITY CLASSIFICATION OF THIS PAGE (When Data Entered)

2

REPORT DOCUMENTATION PAGE

READ INSTRUCTIONS BEFORE COMPLETING FORM

1. REPORT NUMBER	2. GOVT ACCESSION NO.	3. RECIPIENT'S CATALOG NUMBER
4. TITLE (and Subtitle) 6 Extreme Sensitivity of Staphylococcal Enterotoxin B and C Production to Inhibition by Cerulenin		5. TYPE OF REPORT & PERIOD COVERED 9 Interim rept.
7. AUTHOR(s) 10 Robert A. Altenbern		6. PERFORMING ORG. REPORT NUMBER
9. PERFORMING ORGANIZATION NAME AND ADDRESS U.S. Army Medical Research Institute of Infectious Diseases SGRD-UIP-A Fort Detrick, Frederick, MD 21701		8. CONTRACT OR GRANT NUMBER(s)
11. CONTROLLING OFFICE NAME AND ADDRESS U.S. Army Medical Research and Development Command, Office of The Surgeon General Department of the Army, Washington, DC 20314		10. PROGRAM ELEMENT, PROJECT, TASK AREA & WORK UNIT NUMBERS 16 61102B 3M161102BS03 17 00 018
14. MONITORING AGENCY NAME & ADDRESS (if different from Controlling Office)		13. REPORT DATE 11 3 Jan 77
12 15 P.		13. NUMBER OF PAGES 14 pages
		15. SECURITY CLASS. (of this report) UNCLASSIFIED
		15a. DECLASSIFICATION/DOWNGRADING SCHEDULE
16. DISTRIBUTION STATEMENT (of this Report) Approved for public release; distribution unlimited.		
17. DISTRIBUTION STATEMENT (of the abstract entered in Block 20, if different from Report)		
18. SUPPLEMENTARY NOTES Reprints bearing assigned AD number will be forwarded upon receipt. To be submitted for publication in <u>Antimicrobial Agents and Chemotherapy</u> .		
19. KEY WORDS (Continue on reverse side if necessary and identify by block number) Enterotoxin, <u>Staphylococcus</u> , exoprotein		
20. ABSTRACT (Continue on reverse side if necessary and identify by block number) Production of staphylococcal enterotoxin B and C was completely inhibited by concentrations of cerulenin far too low to affect either growth rate or final growth density. Type A toxin formation was not similarly inhibited.		

UNCLASSIFIED

405 039

REPORT DOCUMENTATION PAGE	
1. REPORT NUMBER	2. GOVT ACCESSION NO.
3. AUTHOR(s)	4. PERFORMING ORG. REPORT NUMBER
5. AUTHORING ORG. NAME AND ADDRESS	6. PERFORMING ORG. NAME AND ADDRESS
7. AUTHORING ORG. REPORT NUMBER	8. PERFORMING ORG. REPORT NUMBER
9. PERFORMING ORG. REPORT NUMBER	10. PERFORMING ORG. REPORT NUMBER
11. PERFORMING ORG. REPORT NUMBER	12. PERFORMING ORG. REPORT NUMBER
13. PERFORMING ORG. REPORT NUMBER	14. PERFORMING ORG. REPORT NUMBER
15. PERFORMING ORG. REPORT NUMBER	16. PERFORMING ORG. REPORT NUMBER
17. PERFORMING ORG. REPORT NUMBER	18. PERFORMING ORG. REPORT NUMBER
19. PERFORMING ORG. REPORT NUMBER	20. PERFORMING ORG. REPORT NUMBER
21. PERFORMING ORG. REPORT NUMBER	22. PERFORMING ORG. REPORT NUMBER
23. PERFORMING ORG. REPORT NUMBER	24. PERFORMING ORG. REPORT NUMBER
25. PERFORMING ORG. REPORT NUMBER	26. PERFORMING ORG. REPORT NUMBER
27. PERFORMING ORG. REPORT NUMBER	28. PERFORMING ORG. REPORT NUMBER
29. PERFORMING ORG. REPORT NUMBER	30. PERFORMING ORG. REPORT NUMBER
31. PERFORMING ORG. REPORT NUMBER	32. PERFORMING ORG. REPORT NUMBER
33. PERFORMING ORG. REPORT NUMBER	34. PERFORMING ORG. REPORT NUMBER
35. PERFORMING ORG. REPORT NUMBER	36. PERFORMING ORG. REPORT NUMBER
37. PERFORMING ORG. REPORT NUMBER	38. PERFORMING ORG. REPORT NUMBER
39. PERFORMING ORG. REPORT NUMBER	40. PERFORMING ORG. REPORT NUMBER
41. PERFORMING ORG. REPORT NUMBER	42. PERFORMING ORG. REPORT NUMBER
43. PERFORMING ORG. REPORT NUMBER	44. PERFORMING ORG. REPORT NUMBER
45. PERFORMING ORG. REPORT NUMBER	46. PERFORMING ORG. REPORT NUMBER
47. PERFORMING ORG. REPORT NUMBER	48. PERFORMING ORG. REPORT NUMBER
49. PERFORMING ORG. REPORT NUMBER	50. PERFORMING ORG. REPORT NUMBER
51. PERFORMING ORG. REPORT NUMBER	52. PERFORMING ORG. REPORT NUMBER
53. PERFORMING ORG. REPORT NUMBER	54. PERFORMING ORG. REPORT NUMBER
55. PERFORMING ORG. REPORT NUMBER	56. PERFORMING ORG. REPORT NUMBER
57. PERFORMING ORG. REPORT NUMBER	58. PERFORMING ORG. REPORT NUMBER
59. PERFORMING ORG. REPORT NUMBER	60. PERFORMING ORG. REPORT NUMBER
61. PERFORMING ORG. REPORT NUMBER	62. PERFORMING ORG. REPORT NUMBER
63. PERFORMING ORG. REPORT NUMBER	64. PERFORMING ORG. REPORT NUMBER
65. PERFORMING ORG. REPORT NUMBER	66. PERFORMING ORG. REPORT NUMBER
67. PERFORMING ORG. REPORT NUMBER	68. PERFORMING ORG. REPORT NUMBER
69. PERFORMING ORG. REPORT NUMBER	70. PERFORMING ORG. REPORT NUMBER
71. PERFORMING ORG. REPORT NUMBER	72. PERFORMING ORG. REPORT NUMBER
73. PERFORMING ORG. REPORT NUMBER	74. PERFORMING ORG. REPORT NUMBER
75. PERFORMING ORG. REPORT NUMBER	76. PERFORMING ORG. REPORT NUMBER
77. PERFORMING ORG. REPORT NUMBER	78. PERFORMING ORG. REPORT NUMBER
79. PERFORMING ORG. REPORT NUMBER	80. PERFORMING ORG. REPORT NUMBER
81. PERFORMING ORG. REPORT NUMBER	82. PERFORMING ORG. REPORT NUMBER
83. PERFORMING ORG. REPORT NUMBER	84. PERFORMING ORG. REPORT NUMBER
85. PERFORMING ORG. REPORT NUMBER	86. PERFORMING ORG. REPORT NUMBER
87. PERFORMING ORG. REPORT NUMBER	88. PERFORMING ORG. REPORT NUMBER
89. PERFORMING ORG. REPORT NUMBER	90. PERFORMING ORG. REPORT NUMBER
91. PERFORMING ORG. REPORT NUMBER	92. PERFORMING ORG. REPORT NUMBER
93. PERFORMING ORG. REPORT NUMBER	94. PERFORMING ORG. REPORT NUMBER
95. PERFORMING ORG. REPORT NUMBER	96. PERFORMING ORG. REPORT NUMBER
97. PERFORMING ORG. REPORT NUMBER	98. PERFORMING ORG. REPORT NUMBER
99. PERFORMING ORG. REPORT NUMBER	100. PERFORMING ORG. REPORT NUMBER

ABSTRACT

Production of staphylococcal enterotoxins B and C was completely inhibited by concentrations of cerulenin far too low to affect either growth rate or final growth density. Type A toxin formation was not similarly inhibited.

The secretion of bacterial exoproteins has been most extensively investigated by Lampen and collaborators (5,7) in the case of penicillinase production by Bacillus licheniformis. From these studies, it appears that lipid of the cytoplasmic membrane participates in the synthesis and transport of penicillinase to the exterior medium. Kimura and Izui (4) have recently reported that membrane fluidity, which is controlled by the fatty acids in the membrane, plays an important role in induction of alkaline phosphatase in Escherichia coli. In this laboratory, there is an ongoing investigation of the role of membrane fatty acids in production of enterotoxin B by Staphylococcus aureus. Fatty acid nutrition in S. aureus can be directed by exposing cells to the minimal inhibitory concentration of cerulenin, an antibiotic which inhibits fatty acid biosynthesis (6), and restoring growth by supplementation with appropriate saturated and unsaturated fatty acids. An unexpected result of these studies was the observation that production of staphylococcal enterotoxins B and C is inhibited by concentrations of cerulenin far too low to affect either the growth rate or final growth density of the strains employed. The present report presents details of this phenomenon.

The strains of S. aureus employed were as follows: ATCC 14458, S-6, 137-H2, and 2909. Both strains 14458 and S-6 are wild type strains which produce enterotoxin B (SEB). Strain 137-H2 is a wild type that elaborates enterotoxin C₁ (SEC). Strain 2909 produces relatively large amounts of enterotoxin A (SEA) and was derived from strain 100 by a multistep mutagenesis procedure (3). Cells were grown in 18 x 150 mm tubes containing 5 ml of NAK medium (2) prepared in 0.067 M phosphate

buffer, pH 7.0, and containing graded concentrations of cerulenin. The tubes were incubated at an angle on a shaker at 37 C. Growth rate was monitored by measuring optical density (absorbance) at 600 nm in a Coleman Jr. spectrophotometer. Final growth density (after 21 h incubation) was assessed by measurement of absorbance at 600 nm of a 1:5 dilution (into water) of the culture. A portion of the final culture was centrifuged to remove cells and the supernatant was assayed for enterotoxin by the Oudin tube method. Assays for α -toxin and coagulase were conducted by microtitration and by conventional serial dilution, respectively.

The minimal inhibitory concentration (MIC) of cerulenin for strains 14458, S-6, and 2909 was approximately 100 μ g of antibiotic per ml; strain 137-H2 was somewhat more sensitive, with an MIC of about 50 μ g/ml (Fig. 1). Concentrations of cerulenin up to at least 10 μ g/ml had no demonstrable effect on the final growth density for strains 14458, S-6, or 2909. The final growth density of strain 137-H2 declined gradually above 3 μ g cerulenin/ml.

Pronounced suppression of SEB production by cerulenin concentrations above 1 μ g/ml was noted for strains 14458 and S-6 (Fig. 2). Formation of SEC by strain 137-H2 was completely inhibited by a cerulenin concentration (3 μ g/ml) which had no effect on final growth density. In contrast, there appeared to be no suppression of SEA production by strain 2909 by concentration of cerulenin that did not inhibit growth.

Addition of cerulenin up to 20 μ g/ml to preformed enterotoxin B in NAK medium did not alter the assay value by the Oudin method, thus ruling out possible inactivation of SEB or inhibition of the antigen-antibody reaction by the antibiotic. Similar tests on SEA and SEC were not performed.

Determinations of mass doubling times from optical density data during exponential growth revealed there was no increase in doubling time for strains 14458, S-6, and 2909 up to at least 10 µg cerulenin/ml (Fig. 3). These data parallel the effects on final growth density depicted in Fig. 1. There was also agreement for strain 137-H2 between the concentration of cerulenin required to induce suppression of final growth density and extension of the mass doubling time (5 µg/ml and above).

Cerulenin is an unstaured fatty acid amide (6) and it might be expected that other saturated or unsaturated fatty acids would antagonize the inhibition of production of enterotoxin B by cerulenin. Accordingly, two sets of tubes of NAK medium were prepared, one set without cerulenin and the other containing 10 µg cerulenin/ml. Graded concentrations of a mixture of saturated fatty acids were added to each series. The saturated fatty acid mixture (SFA) was composed of equal concentrations (w/v) of lauric, myristic, palmitic, stearic, arachidic, behenic, and lignoceric acids. Figure 4 shows that the final growth density of the cultures was affected to the same degree by SFA regardless of the presence or absence of cerulenin in the medium. There was no suppression of SEB formation by the SFA mixture in cerulenin-free medium until final growth density was depressed by the added fatty acids. In contrast, in tubes containing cerulenin at 10 µg/ml, a narrow concentration range of SFA mixture partially reversed the total suppression of SEB formation by cerulenin. A mixture of 10 unsaturated fatty acids from C₁₆ to C₂₄ was unable to reverse cerulenin-induced suppression of SEB production without concomitant severe inhibition of growth (data not shown).

Employing strain 14458, it was found that low concentrations of cerulenin, which were noninhibitory for growth rate or final growth density, also strongly suppressed production of α -toxin and coagulase. It appears that production of SEB, α -toxin and coagulase is severely inhibited by the same concentration of cerulenin (Table 1). It is noteworthy that the inhibition is a general effect, altering production of all three exoproteins in the same manner.

The mechanism of this effect is unknown. The concentration of cerulenin required to suppress production of enterotoxins, α -hemolysin and coagulase is only a small fraction of the minimal inhibitory concentration. Since neither the final growth density nor the growth rate is demonstrably affected by these low antibiotic concentrations, it is suggested that interference with fatty acid biosynthesis does not occur, although subtle modifications of synthesis of fatty acids cannot be excluded by these data. The effects of cerulenin reported here resemble the inhibition of coagulase release by very low, subinhibitory concentrations of chloramphenicol (1). The observation that strain 2909 is resistant to inhibition of SEA production when growth rate and final density are unaffected is not surprising. Strain 2909 was derived from wild type strain 100 by a 13-step mutagenesis procedure and is far removed in many of its growth properties from the original wild type. The very low production of SEA by wild type strains such as strain 100 (less than 3 $\mu\text{g/ml}$) precluded determination by Oudin assay of the effect of cerulenin in subinhibitory concentrations on formation of this toxin type.

ACKNOWLEDGEMENTS

I am grateful to Allen R. Knott for performing Oudin assays for the enterotoxins and to Anna Johnson for conducting α -hemolysin titrations.

LEGEND TO FIGURES

FIG. 1. Effect of graded concentrations of cerulenin on final growth density of S. aureus strains.

FIG. 2. Suppression of enterotoxin formation by graded concentrations of cerulenin. Enterotoxin in $\mu\text{g/ml}$ divided by absorbance of 1:5 dilution yields a measure of toxin production per unit cell mass. Strain number in parentheses. The lower limit of determination of enterotoxin amounts of all types by Oudin technique is 3 $\mu\text{g/ml}$.

FIG. 3. Alteration of mass doubling times of S. aureus strains by cerulenin.

FIG. 4. Saturated fatty acid-induced reversal of cerulenin suppression of enterotoxin B formation by 14458.

TABLE 1. Suppression of production of α -hemolysin and coagulase by strain 14458 by graded concentrations of cerulenin.

μg Cerulenin/ml	Titers	
	α -toxin ^a	Coagulase ^b
0	256	4
1	128	2
2	64	2
3	32	0
4	8	0
5	4	0
8	0	0
10	0	0
15	0	0
20	0	0

^aReciprocal of highest dilution showing 50% lysis.

^bReciprocal of highest dilution displaying any clot.

LITERATURE CITED

1. Altenbern, R. A. 1966. On the nature of albumin-promoted coagulase release by Staphylococcus aureus. *J. Infect. Dis.* 116:593-600.
2. Altenbern, R. A. 1976. Enterotoxin B formation by fermentation mutants of Staphylococcus aureus. *Can. J. Microbiol.* 22:182-188.
3. Friedman, M. F., and M. B. Howard. 1971. Induction of mutants of Staphylococcus aureus 100 with increased ability to produce enterotoxin A. *J. Bacteriol.* 106:289-291.
4. Kimura, K., and K. Izui. 1976. Importance of membrane fluidity in the induction of alkaline phosphatase, a periplasmic enzyme, in Escherichia coli. *Biochem. Biophys. Res. Commun.* 70:900-906.
5. Lampen, J. O. 1974. Movement of extracellular enzymes across cell membranes, pp. 351-374. In M. A. Sleight and D. H. Jennings (ed.), *Transport at the cellular level*, Cambridge University Press, Cambridge, England.
6. Omura, S. 1976. The antibiotic cerulenin, a novel tool for biochemistry as an inhibitor of fatty acid synthesis. *Bacteriol. Rev.* 40:681-697.
7. Yamamoto, S., and J. O. Lampen. 1976. Membrane penicillinase of Bacillus licheniformis 749/C: sequence and possible repeated tetrapeptide structure of the phospholipopeptide region. *Proc. Natl. Acad. Sci. U.S.A.* 73:1457-1461.

Fig 1

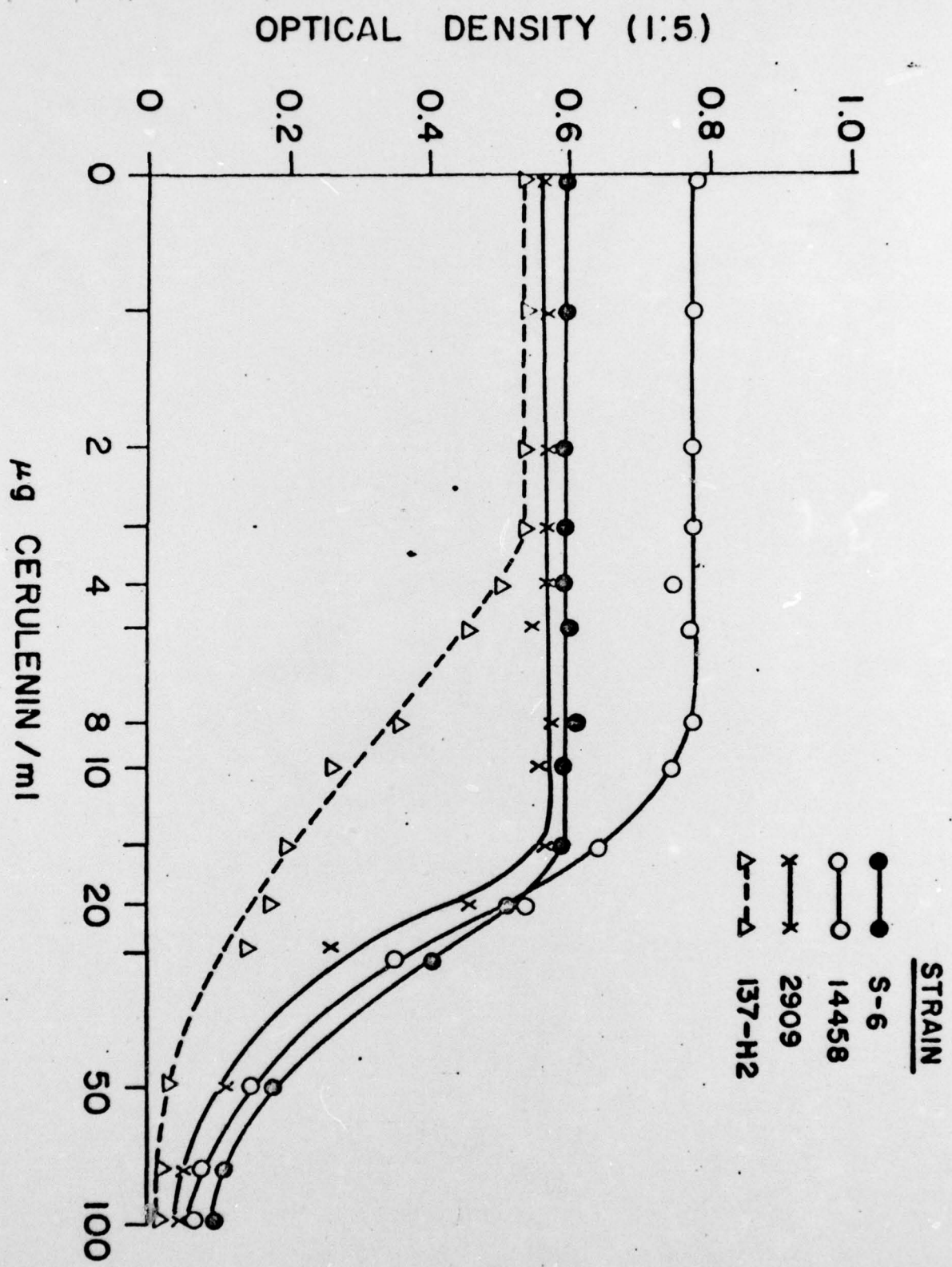
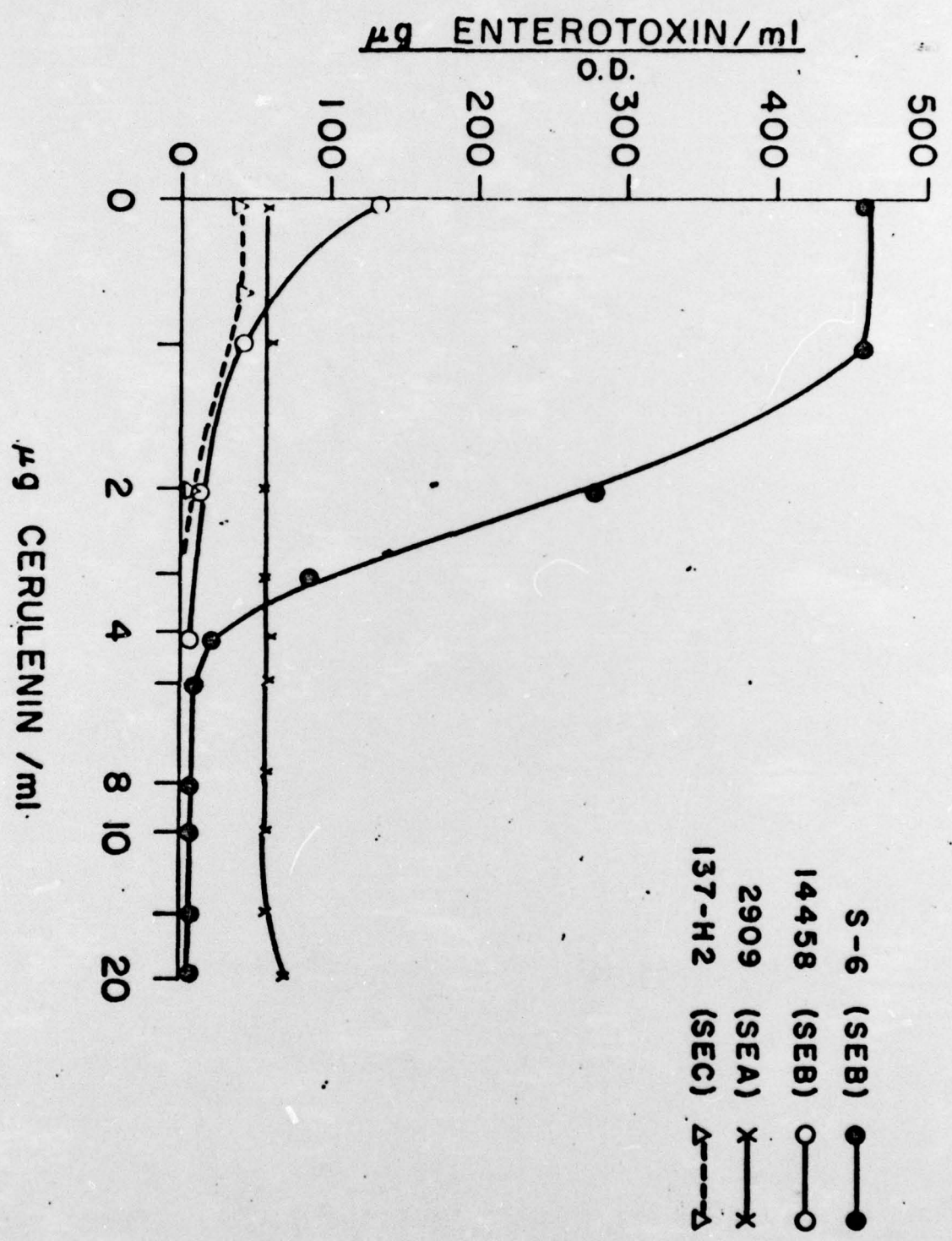


Fig 2



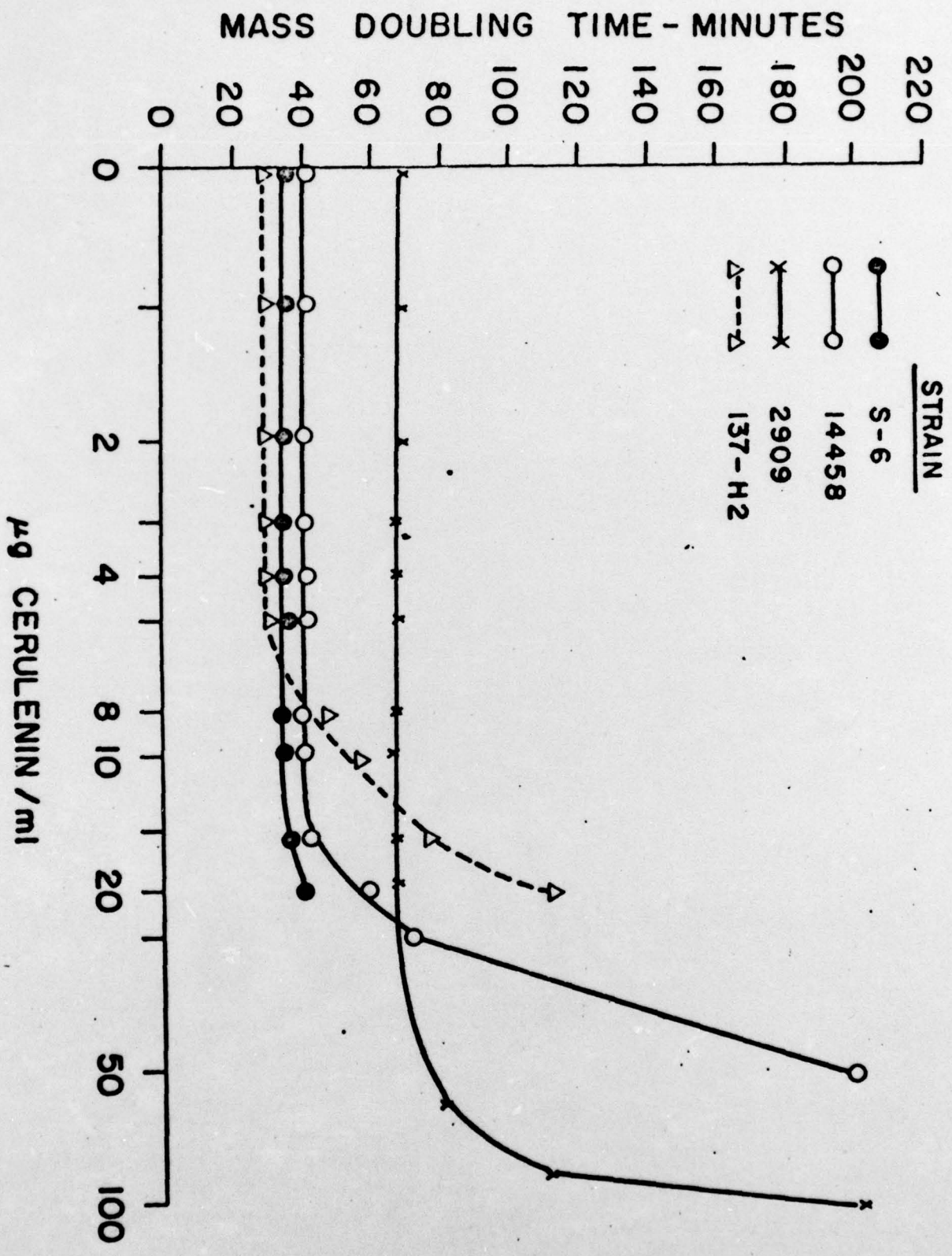
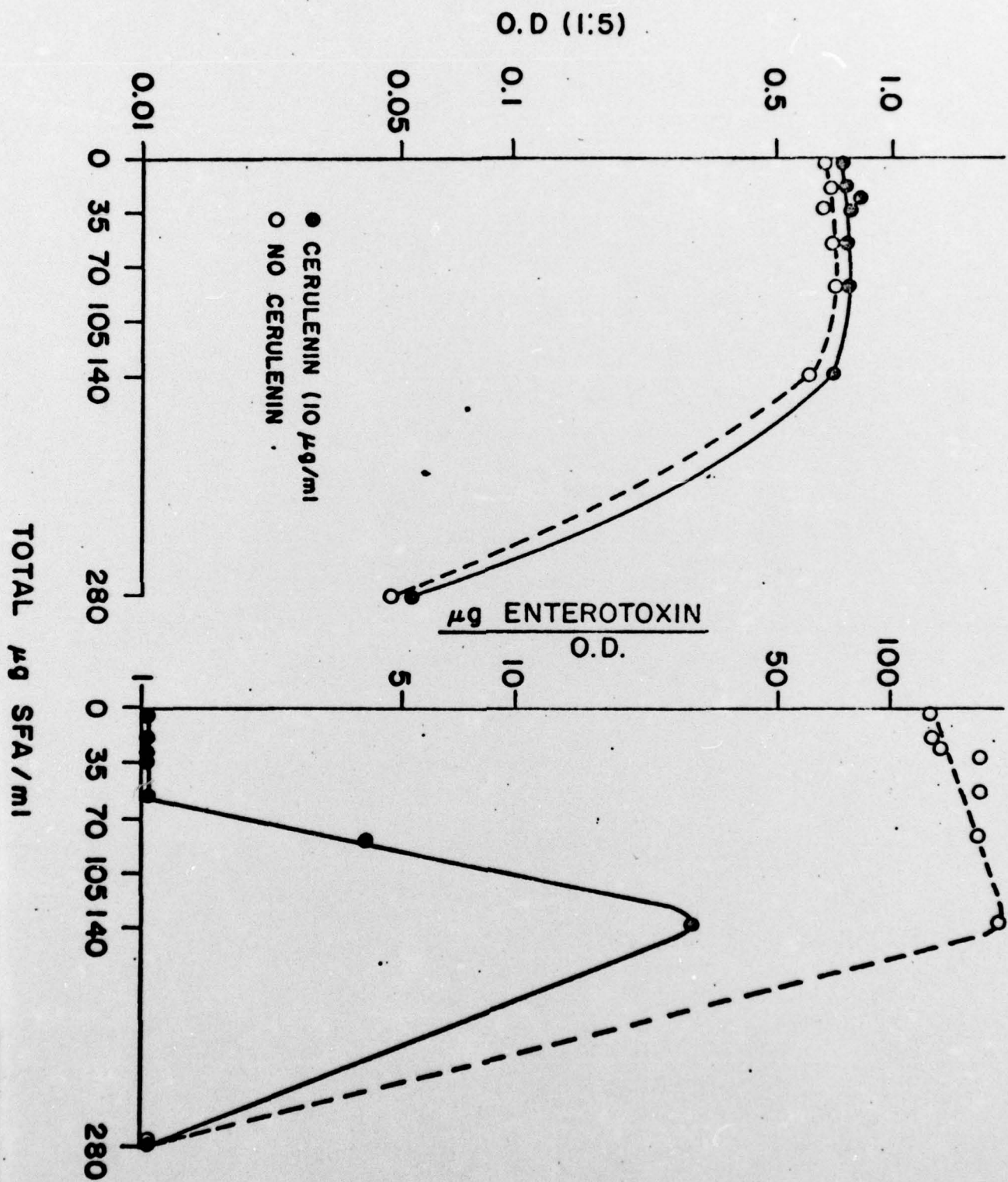


Fig 3



194