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Instrumentation Platform for Imaging Cell Membrane Dynamics

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developed, thus combining fluorest spectroscopy. Electrophysiology may vibrational spectroscopy is conduc- light sources that span the whole fir are critical for the custom-designed laser was acquired for identification temporal dynamics at high speeds integrated into the system. The ima with regards to the mid-infrared be morphological and structural detai dynamics, making this an attractive cell membrane dynamics. The imag chemical identification and thermo	tat is compatible for wavelengths rangin cence imaging and differential interfere easurements can be simultaneously perf ted. The imaging platform is custom-des ngerprint region with a tunable quantum a spectroscopic imaging system were put of protein and lipids via their vibrationa and with a strong signal to noise ratio, a ging resolution for the vibrational imagin am, enabling the investigation of subcel is can be simultaneously acquired with a ging platform is versatile and can be ext al diffusion analysis for sensors, various ma limaging platform are ongoing (due to anuscripts to be submitted	nce contrast m formed while flu- igned to suppo a cascade laser urchased and te al absorption sig high-speed loc ig offers sub-diff lular features in chemical inform ed with infrared ended to study aterials charact	icroscopy with infrared vibrational prescence measurements or rt visible, near-infrared and infrared . Different key building blocks that ested: A tunable quantum cascade natures. In order to resolve ck-in amplifier was purchased and fraction limited spatial resolution biological samples. Hence, nation and thermal diffusion nerve stimulation and blocking and tissue sections and to provide rerization and trace and impurity
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DURIP: Instrumentation Platform for Imaging Cell Membrane Dynamics

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Period $\frac{6}{15} - \frac{6}{14} + \frac{2020}{2020}$ (extension of performance from 12 to 24 months)

In this DURIP grant, the acquisition of equipment for microscopy from visible to infrared wavelengths in one imaging platform was proposed, thus combining fluorescence imaging with infrared vibrational spectroscopy. This allows detection of fluorescently labeled probes in biological specimens, identification of label-free bond-specific inherent spectral signatures, characterization of phase transitions and of the underlying structural order of molecules. The development of such a technology platform aims to advance the scientific development and technology progress for external nerve stimulation and inhibition. Specifically, novel insights into the biophysical mechanisms during infrared nerve stimulation and other external neuromodulation schemes will be gained by resolving the structure and dynamics of the cell membrane during different neurostimulation challenges and to analyze cell membrane dynamics.

To build the proposed instrumentation platform, a high speed lock-in amplifier, higher numerical aperture infrared objective and a tunable quantum cascade laser system were purchased. The UHFLI lock-in amplifier (Zurich Instruments) covers a frequency range up to 600 MHz and box car functionality to resolve temporal dynamics with high signal-to-noise. A refractive infrared objective with 0.4 numerical aperture from Pike Technologies was acquired to enhance the overall spatial resolution. A tunable quantum cascade laser from Daylight solutions was purchased since this allows wavelength coverage between 3-12 µm that offers more versatility for spectroscopic characterization in the fingerprint region. The wavelength modules were chosen specifically so that detailed protein, lipid and nucleic acid studies can be performed with the laser source. A customized microscope from Olympus has been tested, which can simultaneously capture fluorescence, differential interference contrast (DIC) and infrared vibrational imaging in one combined platform. The whole setup has been shielded so that low-noise electrical measurements as needed for electrophysiology can be conducted successfully. Overall, the platform enables a large diversity of sample studies from extracted neurons from various animal models to dissociated cells and brain tissue slides where electrophysiology measurements can be pursued simultaneously with imaging. Thus, the proposed instrumentation will provide an important platform to advance research on neuromodulation and the associated cell membrane dynamics during infrared nerve stimulation and blocking. The equipment will be critical to train the next generation of interdisciplinary researchers and students at the interface of engineering, photonics, biophysics and electrophysiology.

The completed microscope showing the full functionality of the electrophysiology setup is shown below in Figure 1. The multi-functionality of the stage and different light pathways allow for easy switching between fluorescence, DIC and photothermal microscope imaging while keeping the sample in position. The electrophysiology measurements can be conducted with extremely low noise level in this configuration. The resolution for the vibrational imaging offers sub-diffraction limited spatial resolution with regards to the mid-infrared beam, enabling the investigation of subcellular features in biological samples. Hence, morphological and structural details can be simultaneously acquired with chemical information and thermal diffusion dynamics, making this an attractive tool to study the mechanisms associated with infrared nerve stimulation and blocking and cell membrane dynamics. The imaging platform is versatile and can be extended to study tissue sections and to provide chemical identification and thermal diffusion analysis for sensors, various materials characterization and trace and impurity detection.

The availability of this novel instrumentation has led to one conference presentation and one submitted manuscript at this point. Several papers are in preparation (on infrared inhibition, photothermal spectroscopy of fibroblast cells, on hydrogel studies) that will highlight the impact of the imaging platform.

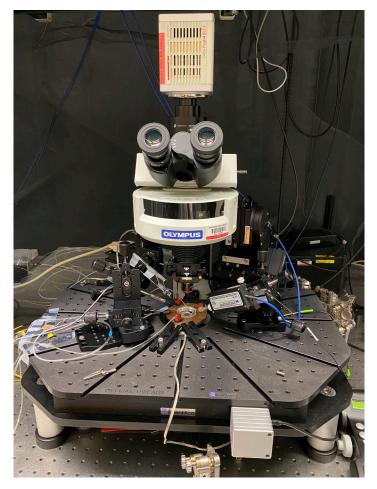


Figure 1: Multi-modular microscope integrating electrophysiology with photothermal chemical imaging. The setup shows the configuration optimized for electrophysiology recordings with various micromanipulators.