

REPORT DOCUMENTATION PAGE				Form Approved OMB No. 0704-0188	
<p>The public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.</p>					
1. REPORT DATE (DD-MM-YY) 30-11-17		2. REPORT TYPE Journal Article		3. DATES COVERED (From - To)	
4. TITLE AND SUBTITLE Exhaled isoprene for monitoring recovery from acute hypoxic stress				5a. CONTRACT NUMBER FA8650-14-D-6516	
				5b. GRANT NUMBER	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) Sean W Harshman 1 , Brian A Geier 2, Anthony V Qualley 1, Leslie A Drummond 3, Laura E Flory 1, Maomian Fan 2, Rhonda L Pitsch 4, Claude C Grigsby 2, Jeffrey B Phillips 3 and Jennifer A Martin 2				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER H0KZ	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) 1 UES Inc., Air Force Research Laboratory, 711th Human Performance Wing/RHXBC, 2510 Fifth Street, Area B, Building 840, Wright- Patterson AFB, OH 45433 3 Naval Medical Research Unit-Dayton, Biomedical Sciences, 2624QStreet, Area B, Building 851, Wright-Patterson AFB, OH 45433 4 The Henry M. Jackson Foundation for the Advancement of Military Medicine, Air Force Research Laboratory, 711th HPW/RHXBC, Wright Patterson AFB, OH 45433				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS(ES) 2 Air Force Materiel Command Air Force Research Laboratory 711 th Human Performance Wing Airman Systems Directorate Human Centered ISR Division Human Signatures Branch Wright-Patterson AFB, OH 45433				10. SPONSORING/MONITORING AGENCY ACRONYM(S) 711 HPW/RHXB	
				11. SPONSORING/MONITORING AGENCY REPORT NUMBER(S) AFRL-RH-WP-JA-2017-0006	
12. DISTRIBUTION/AVAILABILITY STATEMENT Distribution A: Approved for public release. 88ABW-2017-3490, 20 July 2017					
13. SUPPLEMENTARY NOTES Journal of Breath Research https://doi.org/10.1088/1752-7163/aa927d					
14. ABSTRACT Hypoxia-like incidents in-flight have increased over the past decade causing severe safety concerns across the aviation community. As a result, the need to monitor flight crews in real-time for the onset of hypoxic conditions is paramount for continued aeronautical safety. Here, hypoxic events were simulated in the laboratory via a reduced oxygen-breathing device to determine the effect of recovery gas oxygen concentration (21% and 100%) on exhaled breath volatile organic compound composition. Data from samples collected both serially (throughout the exposure), prior to, and following exposures yielded 326 statistically significant features, 203 of which were unique. Of those, 72 features were tentatively identified while 51 were verified with authentic standards. A comparison of samples collected serially between recovery and hypoxia time points shows a statistically significant reduction in exhaled breath isoprene (2-methyl-1,3-butadiene, log ₂ FC=0.399, p=0.005, FDR=0.034, q=0.033), however no significant difference in isoprene abundance was observed when comparing recovery gases (21% or 100% O ₂ , p=0.152). Furthermore, examination of pre-/ post-exposure 1 l bag breath samples illustrate an overall increase in exhaled isoprene abundance post-exposure (log ₂ FC 0.393, p=0.005, FDR=0.094, q=0.033) but again no significant difference between recovery gas (21% and 100%, p=0.798) was observed. A statistically significant difference in trend was observed between isoprene abundance and recovery gases O ₂ concentration when plotted against minimum oxygen saturation (p=0.0419 100% O ₂ , p=0.7034 21% O ₂). Collectively, these results suggest exhaled isoprene is dynamic in the laboratory ROBD setup and additional experimentation will be required to fully understand the dynamics of isoprene in response to acute hypoxic stress.					
15. SUBJECT TERMS exhaled breath, hypoxia, gas chromatography-mass spectrometry, biomarkers					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT: SAR	18. NUMBER OF PAGES 9	19a. NAME OF RESPONSIBLE PERSON (Monitor) Jennifer Martin 19b. TELEPHONE NUMBER (Include Area Code)
a. REPORT Unclassified	b. ABSTRACT Unclassified	c. THIS PAGE Unclassified			

PAPER • OPEN ACCESS

Exhaled isoprene for monitoring recovery from acute hypoxic stress

To cite this article: Sean W Harshman *et al* 2017 *J. Breath Res.* **11** 047111

View the [article online](#) for updates and enhancements.

Related content

- [The identification of hypoxia biomarkers from exhaled breath under normobaric conditions](#)
Sean W Harshman, Brian A Geier, Maomian Fan *et al.*
- [Storage stability of exhaled breath on Tenax TA](#)
Sean W Harshman, Nilan Mani, Brian A Geier *et al.*
- [Wash-out of ambient air contaminations for breath measurements](#)
F Maurer, A Wolf, T Fink *et al.*

Recent citations

- [Characterization of standardized breath sampling for off-line field use](#)
Sean W Harshman *et al*



NEW BREATH BIOPSY PRODUCTS

NEW FEATURES | NEW LOOK

SAME WORLD-LEADING
BREATH RESEARCH PLATFORM

VIEW OUR NEW RANGE

owlstonemedical.com





PAPER

Exhaled isoprene for monitoring recovery from acute hypoxic stress

OPEN ACCESS

RECEIVED

5 October 2017

ACCEPTED FOR PUBLICATION

11 October 2017

PUBLISHED

30 November 2017

Original content from this work may be used under the terms of the [Creative Commons Attribution 3.0 licence](#).

Any further distribution of this work must maintain attribution to the author(s) and the title of the work, journal citation and DOI.



Sean W Harshman¹ , Brian A Geier², Anthony V Qualley¹, Leslie A Drummond³, Laura E Flory¹, Maomian Fan², Rhonda L Pitsch⁴, Claude C Grigsby², Jeffrey B Phillips³ and Jennifer A Martin²

¹ UES Inc., Air Force Research Laboratory, 711th Human Performance Wing/RHXBC, 2510 Fifth Street, Area B, Building 840, Wright-Patterson AFB, OH 45433, United States of America

² Air Force Research Laboratory, 711th Human Performance Wing/RHXBC, 2510 Fifth Street, Area B, Building 840, Wright-Patterson AFB, OH 45433, United States of America

³ Naval Medical Research Unit-Dayton, Biomedical Sciences, 2624 Q Street, Area B, Building 851, Wright-Patterson AFB, OH 45433, United States of America

⁴ The Henry M. Jackson Foundation for the Advancement of Military Medicine, Air Force Research Laboratory, 711th Human Performance Wing/RHXBC, Wright-Patterson AFB, OH 45433, United States of America

E-mail: sean.harshman.ctr@us.af.mil

Keywords: exhaled breath, hypoxia, gas chromatography-mass spectrometry, biomarkers

Supplementary material for this article is available [online](#)

Abstract

Hypoxia-like incidents in-flight have increased over the past decade causing severe safety concerns across the aviation community. As a result, the need to monitor flight crews in real-time for the onset of hypoxic conditions is paramount for continued aeronautical safety. Here, hypoxic events were simulated in the laboratory via a reduced oxygen-breathing device to determine the effect of recovery gas oxygen concentration (21% and 100%) on exhaled breath volatile organic compound composition. Data from samples collected both serially (throughout the exposure), prior to, and following exposures yielded 326 statistically significant features, 203 of which were unique. Of those, 72 features were tentatively identified while 51 were verified with authentic standards. A comparison of samples collected serially between recovery and hypoxia time points shows a statistically significant reduction in exhaled breath isoprene (2-methyl-1,3-butadiene, \log_2 FC -0.399 , $p = 0.005$, FDR = 0.034, $q = 0.033$), however no significant difference in isoprene abundance was observed when comparing recovery gases (21% or 100% O₂, $p = 0.152$). Furthermore, examination of pre-/post-exposure 1 l bag breath samples illustrate an overall increase in exhaled isoprene abundance post-exposure (\log_2 FC 0.393, $p = 0.005$, FDR = 0.094, $q = 0.033$) but again no significant difference between recovery gas (21% and 100%, $p = 0.798$) was observed. A statistically significant difference in trend was observed between isoprene abundance and recovery gases O₂ concentration when plotted against minimum oxygen saturation ($p = 0.0419$ 100% O₂, $p = 0.7034$ 21% O₂). Collectively, these results suggest exhaled isoprene is dynamic in the laboratory ROBD setup and additional experimentation will be required to fully understand the dynamics of isoprene in response to acute hypoxic stress.

Introduction

In-flight hypoxia-like events have been a recurrent problem since the invention of high performance aviation. Over the past decade these events have increased, leading to casualties both in the air and on the ground. As a result, hypoxia was identified by the US Naval Safety Center as the number one threat for naval aircrew [1]. Therefore, research into early detection of hypoxia onset is necessary to preserve the

safety of warfighters and civilians while ensuring the continued success of tactical aviation.

Although exhaled breath has been used for prediction of many physiological and disease states, monitoring changes in the volatile organic compound (VOC) profile of exhaled breath has been recently proposed as a potential method for detecting the onset of hypoxia [2–7]. Previous data suggests exhaled breath VOCs change in abundance during periods of reduced blood oxygen saturation [2]. For example, isoprene

(2-methyl-1,3-butadiene) was found to change in response to an overall laboratory simulated hypoxic event [2]. These results established preliminary data in support for the use of exhaled breath VOC abundances to recognize hypoxic conditions.

Isoprene is an abundant compound that is found in exhaled breath and displays a dynamic release profile [8–15]. For instance, isoprene has been previously shown to decrease in certain disease conditions such as lung cancer and acute respiratory distress syndrome (ARDS), while increasing in response to changes in respiratory patterns such as breath holding [5, 9–11]. Additionally, changes in isoprene abundance have been linked to increased immune response and modifications to cholesterol metabolism [9, 16]. Therefore, due to the dynamic nature of isoprene in exhaled breath additional evidence is required to further characterize and understand the release profile during and after acute hypoxic exposures.

In this manuscript, 203 unique features were assessed for differential abundance between serially applied sea-level, hypoxia and recovery exposures, each of a 5 min duration, applied via a reduced oxygen-breathing device (ROBD-2) [2]. The results illustrate the dynamic nature of isoprene during hypoxic exposures, with significant differences in abundance observed between hypoxia and recovery time points. Additionally, analysis of breath pre-/post-bag samples illustrated that isoprene had significant differences in abundance that are correlated with minimum oxygen saturation and recovery gas. These results provide additional evidence for the use of changes in exhaled breath isoprene to monitor hypoxic respiratory stress.

Experimental

Human subject recruitment

All human volunteer test subjects were male, non-smoking, active duty military in the United States Air Force with ages ranging from 22 to 39 (supplemental data 1 is available online at stacks.iop.org/JBR/11/047111/mmedia). The study was approved for human use by the Institutional Review Board at the Naval Medical Research Unit-Dayton (NAMRUD.2015.003). Written consent was obtained from each test subject prior to initiation of the experiment.

Experimental setup

Each volunteer test subject was placed in a modified 20 P flight mask and HGU 68/P flight helmet as shown in supplemental data 2(A) [2]. Oxygen concentration was controlled to the flight mask via ROBD (Evironics, Tolland CT, supplemental data 2(B)). Blood pressure, pulse and blood oxygen saturation (SpO_2) were recorded as shown and described in supplemental data 2(A). Test subjects were exposed to two separate normobaric simulated flight profiles on different days. Each exposure consisted of three sequential

five-minute periods of sea-level oxygen (21% O_2), 25 000 feet equivalent oxygen (7620 m, 8% O_2) and either sea-level (21% O_2) or 100% O_2 recovery (supplemental data 2(C)). Test subjects were progressed to the recovery gas if their individual SpO_2 fell below 55% or upon their request. A summary of the exposure data and traces of test subjects' SpO_2 are provided in supplemental data 3(A) and (B). A timeline of the experiment illustrating collection periods for bags and time series samples also indicates duration of exposure for each of the O_2 conditions (supplemental data 4).

Breath collection

Exhaled breath samples were collected directly on to preconditioned stainless steel Tenax TA thermal desorption (TD) tubes affixed with brass caps having polytetrafluoroethylene ferrules when not in use (Markes International, South Wales, UK).

Exhaled breath was collected serially from each test subject throughout the exposure, from each test subject ($n = 8$) from the exhalation port side of the modified flight mask, as shown in supplemental data 5(A). Exhaled breath volatiles were captured on to Tenax TA TD tubes in one-minute (550 ml min^{-1}) increments via a Logistically Enabled Sampling System-Portable (LESS-P, SignatureScience LLC, Austin, TX, USA). Flow rate to the LESS-P was continuously monitored throughout the experiment by an inline DryCal Defender 510 l flow meter (Bios International Corp, Butler, NJ, USA). Refer to supplemental data 5(B) and (C) for representative flow rate measurements. All sample TD tubes were capped and stored at 4°C for less than one week until GC-MS analysis [17].

Exhaled breath samples were also collected in 1 l ALTEF polypropylene bags, prior to and following the entire hypoxic exposure, using the previously established breath exhalation protocol (supplemental data 4, Jansen Inert Products, Coral Springs, FL) [2]. Volatiles were immediately transferred from each bag on to Tenax TA TD tubes by a MultiRAE Pro pump (270 ml min^{-1} , 550 ml total volume). Maximal concentrations of carbon dioxide (CO_2 , %) were recorded via the MultiRAE pump to ensure end tidal breath sample quality (supplemental data 5(D)) [18, 19]. All sample TD tubes were capped and stored, less than one week, at 4°C until analysis [17].

Gas chromatography-mass spectrometry (GC-MS)

All samples were analyzed on a Thermo Scientific Trace Ultra-ISQ gas chromatograph affixed with a single quadrupole mass spectrometer following TD on a Markes International TD-100 as described previously (TD, Waltham, MA, USA) [2, 20]. Please refer to supplemental data 6 for a detailed description of the TD-GC-MS methodology.

Table 1. A summary of the feature results from the statistical comparisons across all samples.

Comparison	# Signif. features	# Unique features	# Compound tent. ID	# Verified compound IDs
Hypoxia v. Placebo: Increase	61	54	23	17
Hypoxia v. Placebo: Decrease	37	29	18	13
Recovery v. Hypoxia: Increase	42	38	19	18
Recovery v. Hypoxia: Decrease	26	21	15	13
Recovery v. Placebo: Increase	98	82	36	28
Recovery v. Placebo: Decrease	36	30	15	12
Bag: Post v. Pre: Increase	9	8	5	5
Bag: Post v. Pre: Decrease	4	3	1	1

Feature registration, statistical analysis, compound identification and confirmation

XCMS R library distribution (v 3.5) was leveraged to process raw GC-MS data files [21–23]. Exhaled breath peaks were detected within 1 m/z, registered and aligned using parameters consistent for GC-MS data. Refer to supplemental data 7 for the R script. A table of 3147 features was returned for downstream statistical analysis. Initially, the features were excluded if null values were reported in more than 10% of samples and/or retention time standard deviation post registration was greater than 1.5 s. The remaining matrix of 1742 features was quantile normalized and log₂ transformed. Redundant peaks, i.e. different m/z at a common apex, are an expected occurrence when generating peak tables from raw GC-MS samples. In general, the presence of redundant peaks leads to high measurement co-linearity, which may mislead cluster formation or statistical significance tests. To limit this effect, time binning was performed to identify the most abundant ion for each XCMS feature retention time when rounded to the nearest second. Four hundred and twenty-nine features were available for statistical analysis. Four comparisons of peak abundances were performed, three within serial sampling (hypoxia versus sea-level, recovery versus sea-level, recovery versus hypoxia) and a pre-/post-bag sampling comparison, representing a total of 1716 individual comparisons. For serial sampling, comparisons were constructed by computing differences in average abundances taken within sea-level, hypoxia or recovery time periods. The average was based on at most five samples within either sea-level or hypoxia time periods.

All statistical analyses were performed using the Matlab and Matlab Statistics Toolbox (R2013a, MathWorks, Natick, MA). Wilcoxon signed rank test (Matlab function `signrank.m`) was applied to every feature for each paired comparison (i.e. hypoxia/sea-level paired observations) to determine if the group had a non-zero median distribution. Storey q-values (Matlab function `mafdr.m`) were calculated to control for false discovery rate (FDR). An FDR of 10% was applied to identify feature comparisons that deviated from the null. All statistically significant features were manually inspected. The feature list was manually reduced by removing features corresponding to

redundant ions, i.e. multiple ions of the same compound at a specific retention time, siloxanes (column bleed), and noise. The remaining features were tentatively identified by spectral comparison to the NIST 11 Mass Spectral Library (v.2.0, National Institute of Standards and Technology, Gaithersburg, MD). Tentatively identified compounds were verified by mass spectral and retention time comparison to neat standards. Please refer to supplemental data 8 for neat standard chemical information, supplemental data 9 for tentative identification verification methodology and supplemental data 10 for ID verification results.

Results

Test subject blood oxygen saturation

Individual variation of minimum blood oxygen saturation in response to ROBD reduced oxygen exposures has been observed previously [2, 24]. To monitor this effect, finger blood oxygen saturation (%SpO₂) was measured and plotted (supplemental data 3(A)). The data show a high amount of variability (47%–76%) in the minimum %SpO₂ achieved across test individuals (supplemental data 3(B)). These results confirm previous observations, of variable minimum %SpO₂ by test subjects, related to ROBD hypoxic exposures [2]. The results suggest that physical aspects of an individual may be impactful to the blood's ability to store and transport O₂.

Overview of exhaled breath results

The statistical comparisons of exhaled breath VOC abundances yielded 326 significant features amongst all comparisons (serial and bag). Of those 326 features, 203 correspond to unique features, 72 to tentatively identifiable compounds and 51 of those compound identifications subsequently verified by retention time and spectral matching to neat standards. Refer to table 1 for summary of the individual comparison results and supplemental data 10 for tentative ID verification data summary. Due to the large number of features and verified compounds identified, focus will be placed on isoprene and those compounds that were previously observed to change in response to hypoxic respiratory stress [2].

LESS-P time series analysis

The goal of this study was to determine the effects of recovery gas O₂ concentration on exhaled breath VOC composition. A comparison of the mean abundances of the recovery gas time period to the mean hypoxia time period yielded 68 significant features (p -value ≤ 0.05 , FDR ≤ 0.1 , q -value ≤ 0.1) with 31 verified compound identifications (table 1 and supplemental data 12). Among these features, isoprene (2-methyl-1,3-butadiene) was found to significantly decrease (\log_2 FC -0.399 , $p = 0.005$, FDR = 0.034, $q = 0.033$) in abundance with under recovery conditions with no observed difference between recovery gas O₂ composition ($p = 0.152$, figure 1(A)).

While the goal of this study was to evaluate recovery oxygen's effects on exhaled breath from acute hypoxia exposures, additional samples were acquired across the exposure time course. Of those remaining time series results, 98 features and 30 verified compound identifications were a result of abundance comparisons between hypoxia and sea-level time periods (table 1 and supplemental data 11). A survey of the results highlighted two compounds, pentanal and 2-pentanone, that were previously found to change in abundance in response to reduced oxygen content when compared to sea-level [2]. To further explore the relationship of these compounds to hypoxia exposures, \log_2 abundance ratios (mean hypoxia abundance/mean sea-level abundance) were calculated and plotted (figures 1(B) and (C)). The results show a statistically significant increase in 2-pentanone abundance (\log_2 FC 0.244, $p = 0.012$, FDR = 0.063, $q = 0.062$) and a significant reduction in pentanal abundance (\log_2 FC -0.283 , $p = 0.022$, FDR = 0.09, $q = 0.088$) under hypoxic conditions. These data support previous observations of reduced pentanal abundance under hypoxic conditions when compared to sea-level time points [2]. Interestingly, the results show an inverse response in the 2-pentanone abundance when compared with previous results [2]. While inconsistencies in the data are present, these results support further studies of pentanal and 2-pentanone changes in response to hypoxic stress.

Breath bag analysis

To identify the overall changes in exhaled breath following a hypoxic exposure with different recovery gas O₂ compositions, comparisons of VOC abundances from bags collected prior to and following the overall exposure event were performed. Collectively, the data yielded a small number (13) of statistically different features (p -value ≤ 0.05 , FDR ≤ 0.1 , q -value ≤ 0.1) with only 6 features being verified (table 1 and supplemental data 14). Among the six significant features, isoprene was found to have a significant increase in abundance (\log_2 FC 0.393, $p = 0.005$, FDR = 0.094, $q = 0.033$), post-exposure and recovery, with no observed effect between recovery gas

compositions (figure 2(A)). These results suggest increase in isoprene abundance, compared to the breath prior to exposure, may be indicative of an overall hypoxic and recovery event.

Our previous study identified a correlation between increase in isoprene abundance under 100% O₂ recovery to the minimal SpO₂ achieved by test individuals [2]. To determine if this effect was observed, the \log_2 abundance ratios (post/pre) for isoprene were plotted against the individual minimum blood oxygen saturation (figure 2(B)). Isoprene abundance significantly increases ($p = 0.0419$) in the exhaled breath of those individuals recovered under 100% O₂, shown by a positive slope of the fit, similar to those results previously observed (figure 2(B)) [2]. However, the effect is not observed ($p = 0.7034$) in those recovered on 21% O₂ gas (figure 2(B)). These results suggest exhaled isoprene is affected by the oxygen concentration or gas delivery provided to the test subject following a hypoxic exposure.

Discussion

As hypoxic events continue to plague the aviation community the need to monitor flight crew is necessary for continued safety. Here, further evidence is provided in support for the use of exhaled breath VOC content to monitor hypoxic stress. The data illustrate the dynamic nature of isoprene in the ROBD setup. In the serial samples, a significant decrease in exhaled isoprene is observed. The significant dynamics of isoprene are not surprising as isoprene has been repeatedly shown to change in response to a large group of stimuli including age, posture, and disease state [8–15]. For example, decreased exhaled isoprene has been shown in other respiratory diseases such as ARDS and lung cancer [5, 9, 10]. Additionally, exhaled isoprene has been shown to be derived from the cholesterol metabolic pathway where decreased cholesterol is linked to decreased breath isoprene [16]. As hypoxia has been shown to negatively influence cholesterol metabolism, the observation of decreased breath isoprene in hypoxic individuals is a plausible hypothesis for the results obtained [25].

The observation that 2-pentanone and pentanal significantly change in response to hypoxic stress is not novel [2]. However, previous results suggest that 2-pentanone significantly decreased under hypoxic conditions whereas the current study shows an increase in compound abundance. While it is currently unknown why the inversion of 2-pentanone abundance was observed, these results support changes of these compounds have been found among multiple studies suggesting development of targeted analytical assays to better quantitate specific exhaled breath compounds of interest.

Pre-/post-bag analysis shows an increase in exhaled isoprene with a correlation to minimum SpO₂ levels

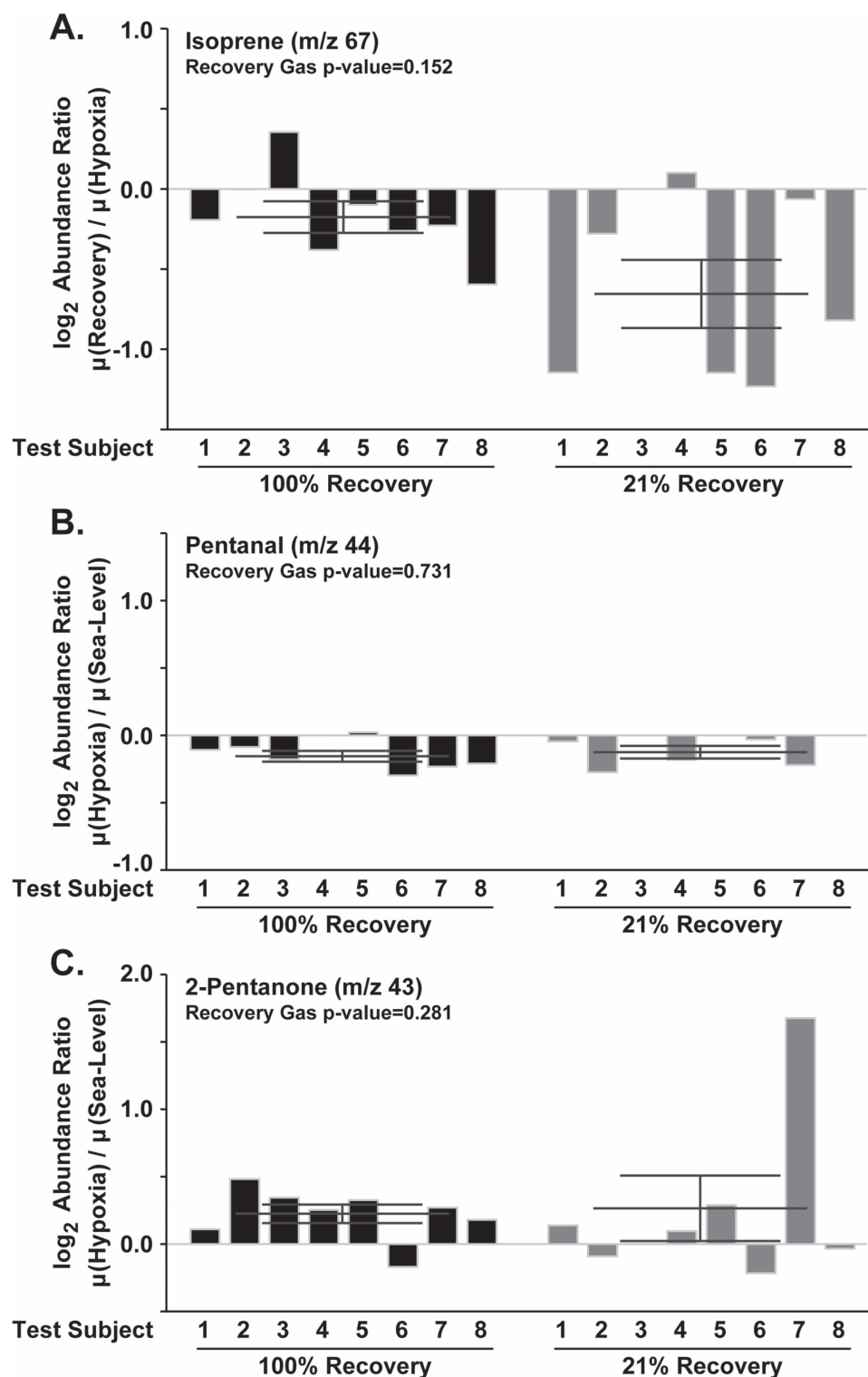
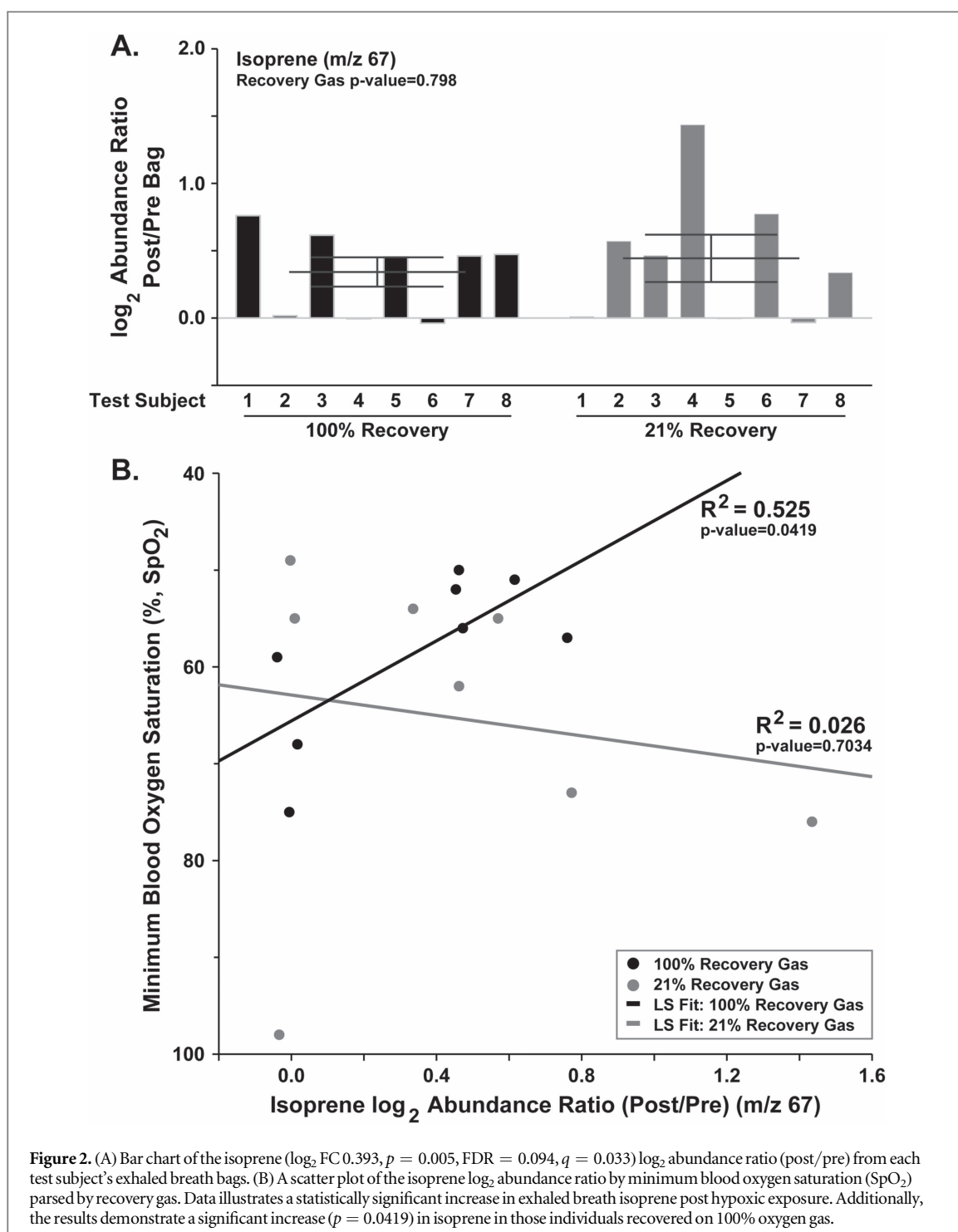


Figure 1. Bar charts of test subjects \log_2 abundance ratios measured during time course for (A) isoprene (\log_2 FC -0.399 , $p = 0.005$, FDR = 0.034, $q = 0.033$), (B) pentanal (\log_2 FC -0.283 , $p = 0.022$, FDR = 0.09, $q = 0.088$), (C) 2-pentanone (\log_2 FC 0.244 , $p = 0.012$, FDR = 0.063, $q = 0.062$). Error bars signify the standard error of the mean. Data shows examples of statistically significant changes in exhaled breath volatile organic compound abundance in response to hypoxic exposure and recovery independent of the recovery gas O_2 composition.

achieved and the oxygen concentration used in the recovery from the hypoxic exposure. In evaluating these results, it was observed that 100% O_2 recovery gas was provided at a positive pressure ($\sim 2''$ H_2O) while the 21% O_2 was provided at a much lower pressure (similar to on demand). Therefore it is plausible that the effect

observed maybe due to changes in respiration induced from recovery gas pressure rather than the O_2 concentration itself [13, 26, 27]. This hypothesis is further supported by the increased levels of isoprene observed in breath holding and exercise experiments [11, 12]. However further experimentation utilizing alternative



methods for exposure, such as a reduced oxygen breathing environment, will be required to fully investigate this hypothesis.

Conclusion

In conclusion, the results illustrate the dynamic nature of isoprene in the exhaled breath of test subjects during and following acute hypoxic exposures. These data provide support for additional research into the use of exhaled breath for non-invasively monitoring personnel with acute hypoxic exposures. However, due to the

dynamic nature of exhaled VOCs, alternative exposure methodologies, such as a mask-off nitrogen tent, and targeted analytical assays must be utilized to fully characterize the exhaled breath dynamics in response to these exposures.

Acknowledgments

Support for this work was provided by UES Inc. under subcontract from the United States Air Force (FA8650-14-D-6516). Opinions, interpretations, conclusions and recommendations are those of the

authors and not necessarily endorsed by the United States Government. The authors would like to thank Mr Gregory Sudberry and Ms Tenika Dearmond for their help and scholarly input for this manuscript.

ORCID iDs

Sean W Harshman  <https://orcid.org/0000-0001-5517-3847>

References

- [1] Ostrander G 2005 Hypoxia in the Hornet—What we know, and what we're doing *Approach* **50** 10–2
- [2] Harshman S W, Geier B A, Fan M, Rinehardt S, Watts B S, Drummond L A, Preti G, Phillips J B, Ott D K and Grigsby C C 2015 The identification of hypoxia biomarkers from exhaled breath under normobaric conditions *J. Breath Res.* **9** 047103
- [3] Horváth I, Lazar Z, Gyulai N, Kollai M and Losonczy G 2009 Exhaled biomarkers in lung cancer *Eur. Respir. J.* **34** 261–75
- [4] Westhoff M, Litterst P, Freitag L, Urfer W, Bader S and Baumbach J-I 2009 Ion mobility spectrometry for the detection of volatile organic compounds in exhaled breath of patients with lung cancer: results of a pilot study *Thorax* **64** 744–8
- [5] Bajtarevic A et al 2009 Noninvasive detection of lung cancer by analysis of exhaled breath *BMC Cancer* **9** 348
- [6] Poli D, Goldoni M, Corradi M, Acampa O, Carbognani P, Internullo E, Casalini A and Mutti A 2010 Determination of aldehydes in exhaled breath of patients with lung cancer by means of on-fiber-derivatisation SPME-GC/MS *J. Chromatogr. B* **878** 2643–51
- [7] Fens N, Zwinderman A H, van der Schee M P, de Nijs S B, Dijkers E, Roldann A C, Cheung D, Bel E H and Sterk P J 2009 Exhaled breath profiling enables discrimination of chronic obstructive pulmonary disease and asthma *Am. J. Respir. Crit. Care Med.* **180** 1076–82
- [8] Sukul P, Trefz P, Kamysek S, Schubert J K and Miekisch W 2015 Instant effects of changing body positions on compositions of exhaled breath *J. Breath Res.* **9** 047105
- [9] Fuchs D, Jamnig H, Heininger P, Klieber M, Schroecksnadel S, Fiegl M, Hackl M, Denz H and Amann A 2012 Decline of exhaled isoprene in lung cancer patients correlates with immune activation *J. Breath Res.* **6** 027101
- [10] Schubert J K, Miekisch W and Noldge-Schomburg G 2002 *Disease Markers in Exhaled Breath* (Amsterdam: IOS Press)
- [11] Sukul P, Trefz P, Schubert J K and Miekisch W 2014 Immediate effects of breath holding maneuvers onto composition of exhaled breath *J. Breath Res.* **8** 037102
- [12] King J, Kupferthaler A, Unterkofler K, Koc H, Teschl S, Teschl G, Miekisch W, Schubert J, Hinterhuber H and Amann A 2009 Isoprene and acetone concentration profiles during exercise on an ergometer *J. Breath Res.* **3** 027006
- [13] Hornuss C, Zagler A, Dolch M E, Wiepcke D, Praun S, Boulesteix A-L, Weis F, Apfel C C and Schelling G 2012 Breath isoprene concentrations in persons undergoing general anesthesia and in healthy volunteers *J. Breath Res.* **6** 046004–8
- [14] Alkhouri N, Singh T, Alsabbagh E, Guirguis J, Chami T, Hanouneh I, Grove D, Lopez R and Dweik R 2015 Isoprene in the exhaled breath is a novel biomarker for advanced fibrosis in patients with chronic liver disease: a pilot study *Clin. Transl. Gastroenterol.* **6** e112
- [15] Lechner M, Moser B, Niederseer D, Karlseder A, Holzknacht B, Fuchs M, Colvin S, Tilg H and Rieder J 2006 Gender and age specific differences in exhaled isoprene levels *Respir. Physiol. Neurobiol.* **154** 478–83
- [16] Stone B G, Besse T J, Duane W C, Evans C D and DeMaster E G 1993 Effect of regulating cholesterol biosynthesis on breath isoprene excretion in men *Lipids* **28** 705–8
- [17] Harshman S W et al 2016 Storage stability of exhaled breath on Tenax TA *J. Breath Res.* **10** 046008
- [18] Schubert J K, Spittler K-H, Braun G, Geiger K and Guttmann J 2001 CO₂-controlled sampling of alveolar gas in mechanically ventilated patients *J. Appl. Physiol.* **90** 486–92
- [19] Birken T, Schubert J, Miekisch W and Noldge-Schomburg G 2006 A novel visually CO₂ controlled alveolar breath sampling technique *Technol. Health Care* **14** 499–506
- [20] Kwak J, Fan M, Harshman S W, Garrison C E, Dershem V L, Phillips J B, Grigsby C C and Ott D K 2014 Evaluation of Bio-VOC sampler for analysis of volatile organic compounds in exhaled breath *Metabolites* **4** 879–88
- [21] Smith C A, Want E J, O'Maille G, Abagyan R and Siuzdak G 2006 XCMS: processing mass spectrometry data for metabolite profiling using nonlinear peak alignment, matching, and identification *Anal. Chem.* **78** 779–87
- [22] Tautenhahn R, Böttcher C and Neumann S 2008 Highly sensitive feature detection for high resolution LC/MS *BMC Bioinform.* **9** 504
- [23] Benton H P, Want E J and Ebbels T 2010 Correction of mass calibration gaps in liquid chromatography-mass spectrometry metabolomics data *Bioinformatics* **26** 2488–9
- [24] Phillips J B, Horning D and Funke M E 2015 Cognitive and perceptual deficits of normobaric hypoxia and the time course to performance recovery *Aerosp. Med. Hum. Perform.* **86** 357–65
- [25] Matsumoto K, Taniguchi T, Fujioka Y, Shimizu H, Ishikawa Y I and Yokoyama M 2000 Effects of hypoxia on cholesterol metabolism in human monocyte-derived macrophages *Life Sci.* **67** 2083–91
- [26] Dolch M E, Frey L, Hornuss C, Schmoelz M, Praun S, Villinger J and Schelling G 2008 Molecular breath-gas analysis by online mass spectrometry in mechanically ventilated patients: a new software-based method of CO₂-controlled alveolar gas monitoring *J. Breath Res.* **2** 037010
- [27] Brock B, Kamysek S, Silz J, Trefz P, Schubert J K and Miekisch W 2017 Monitoring of breath VOCs and electrical impedance tomography under pulmonary recruitment in mechanically ventilated patients *J. Breath Res.* **11** 1–10