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TITLE: Metastatic Colorectal Cancer Immunotherapy with GUCY2C-Expressing
Listeria monocytogenes

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CONTRACTING ORGANIZATION: Thomas Jefferson University
Philadelphia, PA 19107

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14. ABSTRACT This objective of this project is to define the ability of a new vaccine to treat colorectal cancer in animal models. The vaccine uses attenuated Listeria monocytogenes to deliver GUCY2C, a colorectal cancer antigen to "antigen-presenting cells" (APCs) to induce GUCY2C-specific T cell responses against GUCY2C which may find and eliminate colorectal cancer. The project tests novel recombinant Listeria monocytogenes vaccines produced in our laboratory in mouse models of colorectal cancer to determine their activity, efficacy, and safety. If successful, the results of these studies could be applied to patients with metastatic colorectal cancer, a disease that is typically fatal.					
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1. INTRODUCTION

This objective of this project is to define the ability of a new vaccine to treat colorectal cancer in animal models. The vaccine uses attenuated *Listeria monocytogenes* to deliver GUCY2C, a colorectal cancer antigen to “antigen-presenting cells” (APCs) to induce GUCY2C-specific T cell responses against GUCY2C which may find and eliminate colorectal cancer. The project tests novel recombinant *Listeria monocytogenes* vaccines produced in our laboratory in mouse models of colorectal cancer to determine their activity, efficacy, and safety. If successful, the results of these studies could be applied to patients with metastatic colorectal cancer, a disease that is typically fatal.

2. KEYWORDS

GUCY2C, *Listeria monocytogenes*, colorectal cancer, cancer vaccine, cancer immunotherapy

3. ACCOMPLISHMENTS

- What were the major goals of the project?

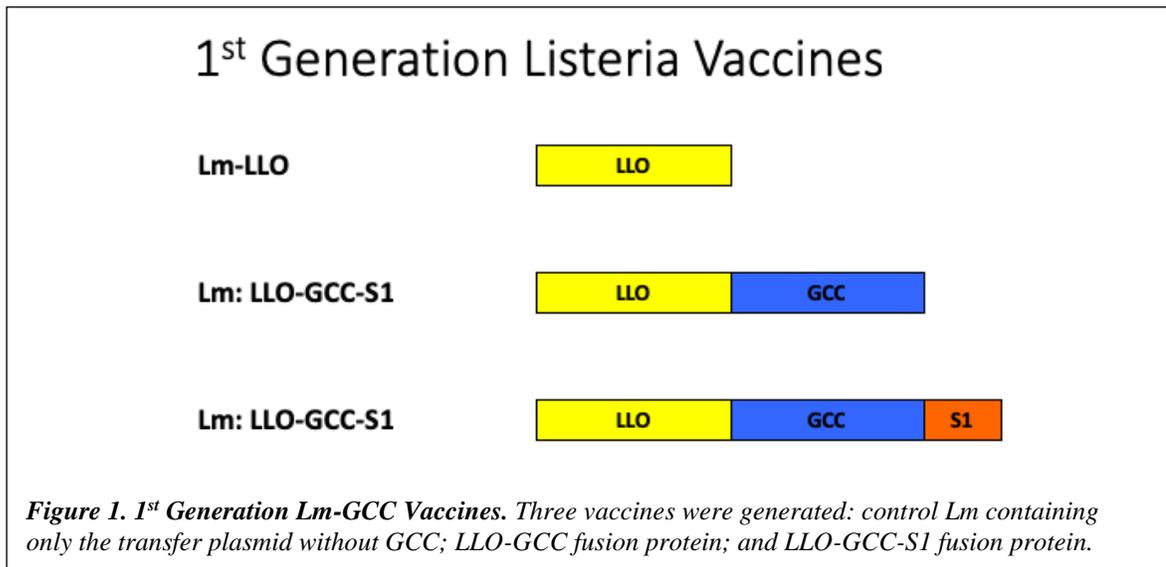
Specific Aim 1. Define the immunogenicity of Lm-LLO-GCC	Anticipated Timeline/ % Complete
Major Task 1: Generate Lm-LLO-GCC and control Lm vaccines	Months 1-4
Milestone Achieved: Produce recombinant Lm vaccines	100%*
Major Task 2: Determine the ability of Lm-LLO-GCC to overcome tolerance in WT mice	Months 1-8
Milestone Achieved: IACUC Approval	100%
Milestone Achieved: Federal (ACURO) Approval	100%
Milestone Achieved: Quantify Lm-LLO-GCC-induced immune responses in <i>WT</i> and <i>KO</i> mice	100%*
Major Task 3: Determine whether GCC-specific responses are CD4⁺ T-cell dependent	Months 7-8
Milestone Achieved: Determine the CD4 ⁺ T-cell-dependence of GCC- specific responses to Lm-LLO-GCC	0%
Specific Aim 2. Quantify the antitumor activity of Lm-LLO-GCC in a metastatic CRC model	
Major Task 4: Compare the antitumor efficacy of Lm-LLO-GCC and Ad5-GCC-S1	Months 9-16
Milestone Achieved: Compare the antitumor efficacy of Lm-LLO-GCC and Ad5-GCC-S1	100%*
Major Task 5: Compare the impact of vector-specific immunity on immune responses to Lm-LLO-GCC and Ad5-GCC-S1	Months 11-14

Milestone Achieved: Quantify immune responses in Lm-LLO-GCC or Ad5-GCC-S1-immunized mice with pre-existing vector-specific immunity	0%
Major Task 6: Compare the impact of vector-specific immunity on Lm-LLO-GCC and Ad5-GCC-S1 antitumor efficacy	Projected Months 11-18 0% Complete
Milestone Achieved: Quantify antitumor efficacy of Lm-LLO-GCC or Ad5-GCC-S1 in mice with pre-existing vector-specific immunity	0%
Specific Aim 3. Demonstrate the safety of Lm-LLO-GCC	
Major Task 7: Examine Lm-LLO-GCC-immunized mice for signs of toxicity	Projected Months 18-23 0% Complete
Milestone Achieved: Establish Lm-LLO-GCC pathology in GCC-expressing and non-expressing tissues	0%

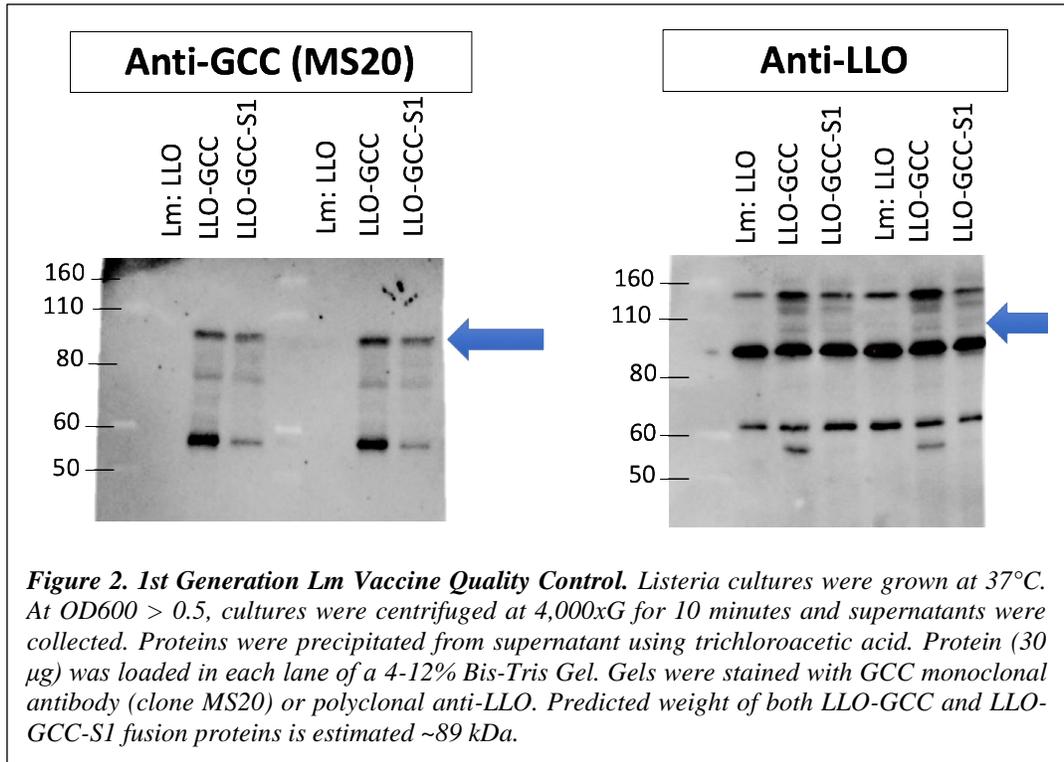
* See also 5. Changes or Problems

What was accomplished under these goals?

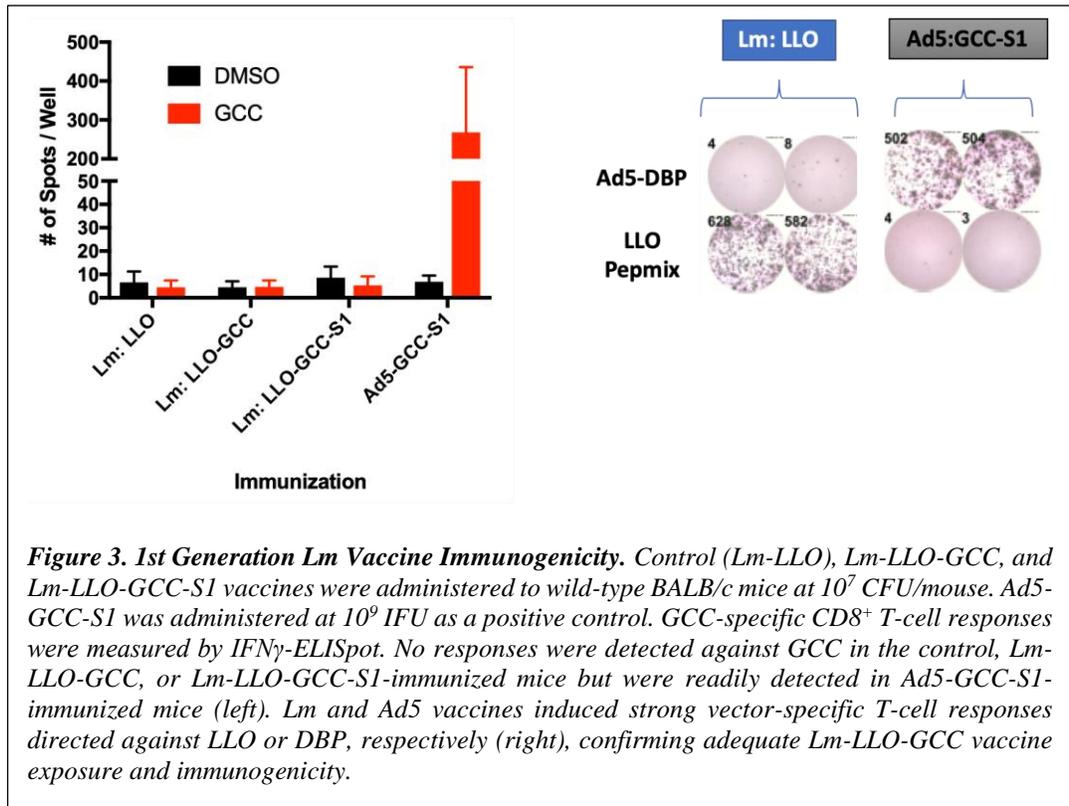
- **1st Generation Recombinant Lm-LLO-GCC Vaccines**
 - The integration plasmid pPL2 was used to induce recombinant GCC or GCC-S1 integration into the genome of attenuated ($\Delta actA/\Delta inLB$) *Listeria monocytogenes* (Lm) downstream of a truncated listeriolysin O (LLO) expression cassette and the LLO promoter, producing an LLO-GCC fusion protein that will be secreted into the cytosol of Lm-infected cells (**Figure 1**). These constructs contain the full extracellular domain of GCC (residues 23-429) that has been used in previous viral vector-based vaccines.



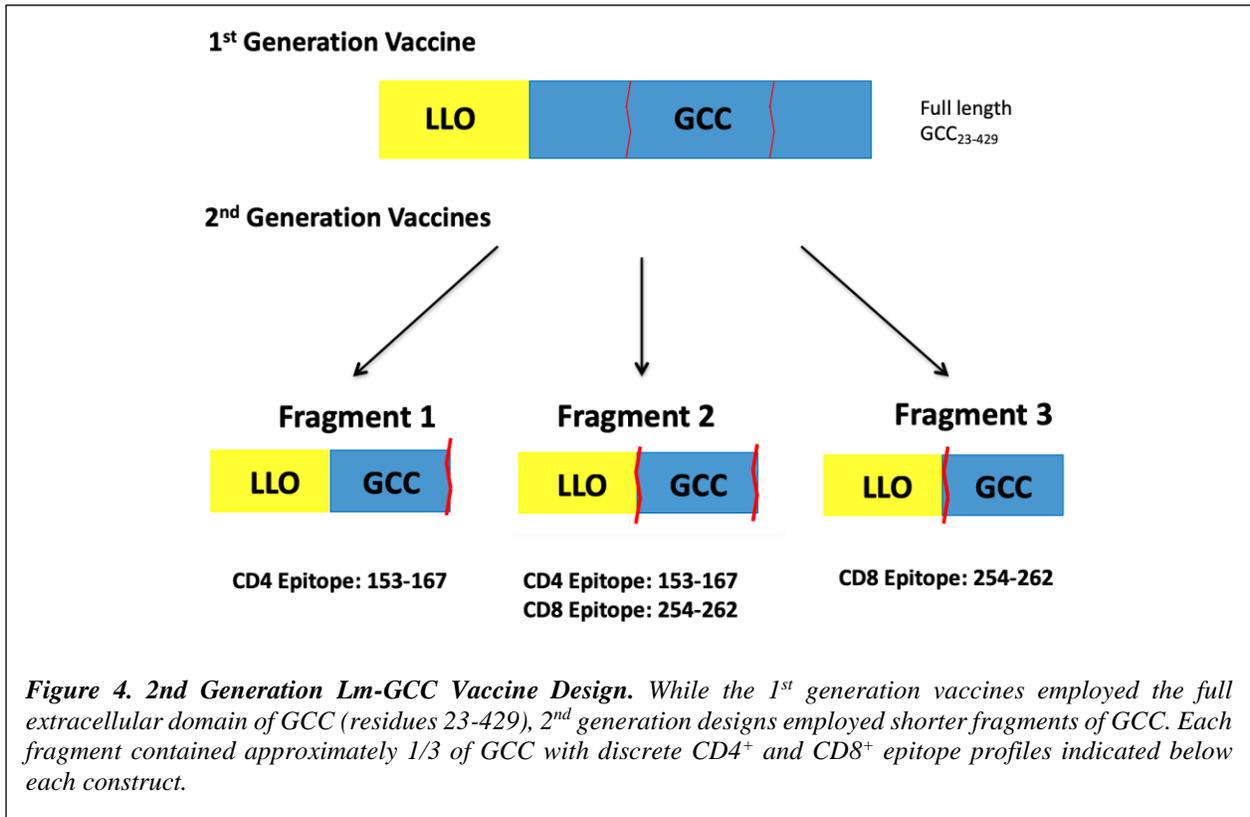
- 1st generation Lm-GCC and control vaccines were quality controlled by confirming GCC expression in supernatants of Lm cultures (**Figure 2**). The LLO-GCC or LLO-GCC-S1 fusion proteins were detected in supernatants using anti-GCC or anti-LLO antibodies.



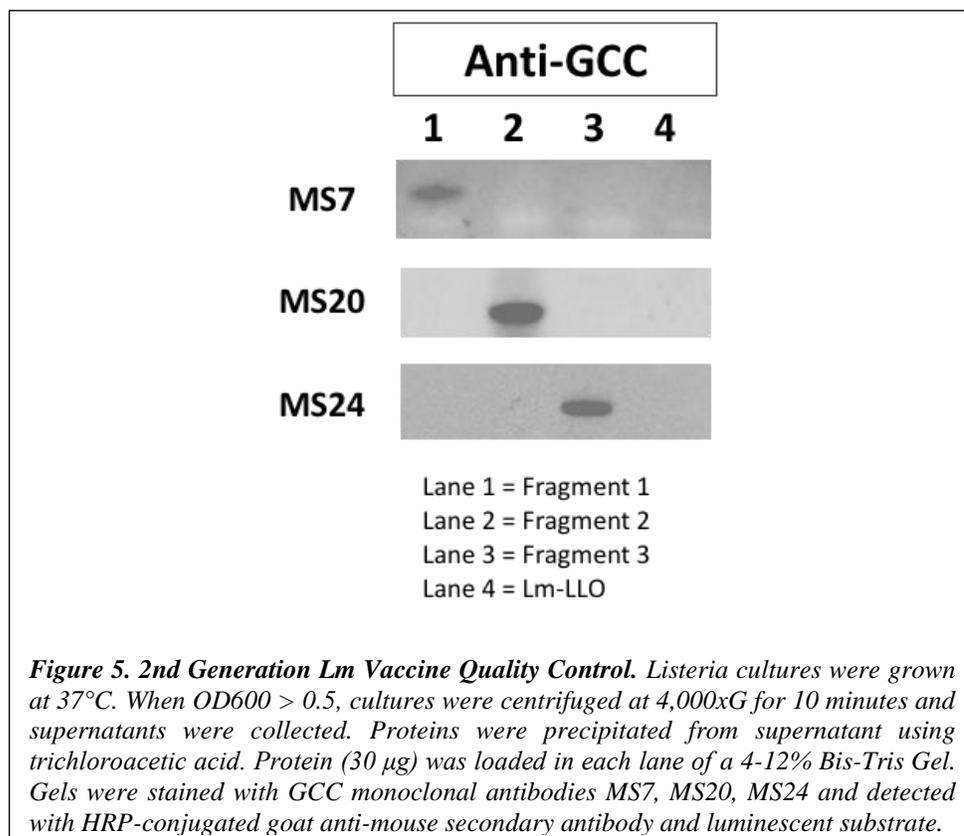
- The immunogenicity of 1st-generation Lm-GCC vaccines was tested in BALB/c mice (**Figure 3**). The well-established adenovirus-based vaccine (Ad5-GCC-S1) was used as a positive control. Following immunization, CD8⁺ T-cell responses to GCC were quantified by IFN γ -ELISpot. As expected control Lm-LLO induced no GCC-specific responses; however, Lm-LLO-GCC and Lm-LLO-GCC-S1 also failed to induce responses. Responses were induced by the positive control Ad5-GCC-S1 vaccine and responses were generated against the antigens LLO and DBP in the Lm and Ad5 vectors, respectively. Together, these data suggested that Lm-LLO-GCC vaccines were poor inducers of GCC-specific responses.



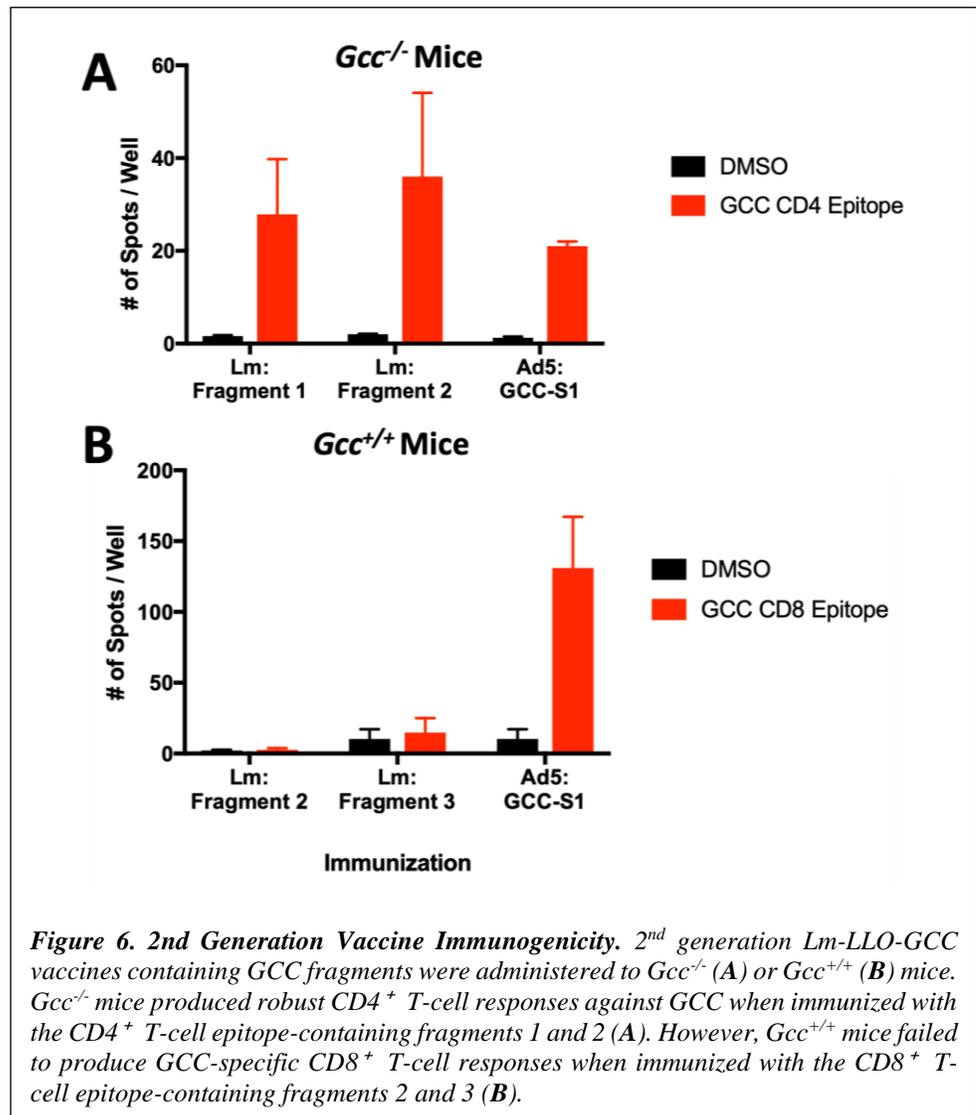
- **2nd Generation Recombinant Lm-LLO-GCC Vaccines**
 - In the context of low LLO-GCC fusion protein expression (**Figure 2**), we hypothesized that the large size of the LLO-GCC fusion protein may be interfering with its expression and induction of immunity. Indeed, similar studies with large tumor antigens fused to LLO show similar difficulty in generating the fusion protein. Therefore, we generated 3 different 2nd generation vaccines, each of which contained about 1/3 of GCC (**Figure 4**). The designs contain different GCC CD4⁺ T-cell and CD8⁺ T-cell epitopes allowing us to test responses to each epitope with two different fragments.



- Expression of the LLO-GCC fusion protein was confirmed in 2nd generation Lm-LLO-GCC vaccines by western blot. Because each 2nd generation vaccine contains a different GCC fragment, multiple GCC-specific monoclonal antibodies were used to detect GCC-LLO fusion proteins in Lm supernatants (**Figure 5**). These data confirm LLO-GCC fusion protein expression by each recombinant Lm vaccine. As expected, the shorter fusion proteins were expressed at higher levels than the full-length LLO-GCC product produced in the 1st generation vaccine (comparison not shown).



- 2nd generation Lm-LLO-GCC vaccines containing GCC fragments were administered to $Gcc^{-/-}$ (**Figure 6A**) or $Gcc^{+/+}$ (**Figure 6B**) mice. Because $Gcc^{-/-}$ mice produce only $CD4^{+}$ T-cell responses and $Gcc^{+/+}$ produce only $CD8^{+}$ T-cell responses, mice received only the vaccine fragments containing those epitopes. $Gcc^{-/-}$ mice produced robust $CD4^{+}$ T-cell responses against GCC when immunized with the $CD4^{+}$ T-cell epitope-containing fragments 1 and 2 (**Figure 6A**). However, $Gcc^{+/+}$ mice failed to produce GCC-specific $CD8^{+}$ T-cell responses when immunized with the $CD8^{+}$ T-cell epitope-containing fragments 2 and 3 (**Figure 6B**). To confirm the lack of $CD8^{+}$ T-cell responses to GCC in $Gcc^{+/+}$ mice, we performed an anti-tumor immunity experiment (**Figure 7**). In agreement with $IFN\gamma$ -ELISpot data (**Figure 6**), Lm-LLO-GCC fragment immunization in $Gcc^{+/+}$ mice showed no antitumor efficacy compared to Ad5-GCC-S1 which was 100% effective (**Figure 7**).



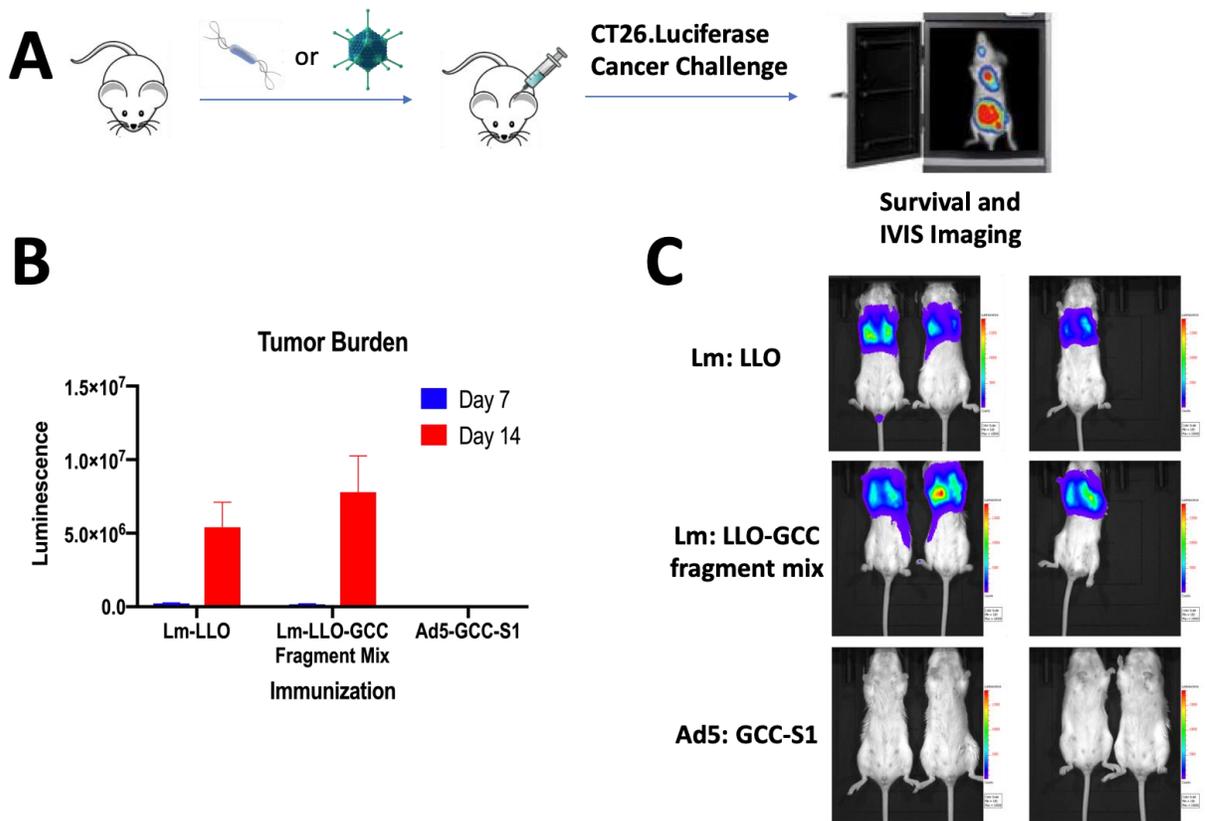
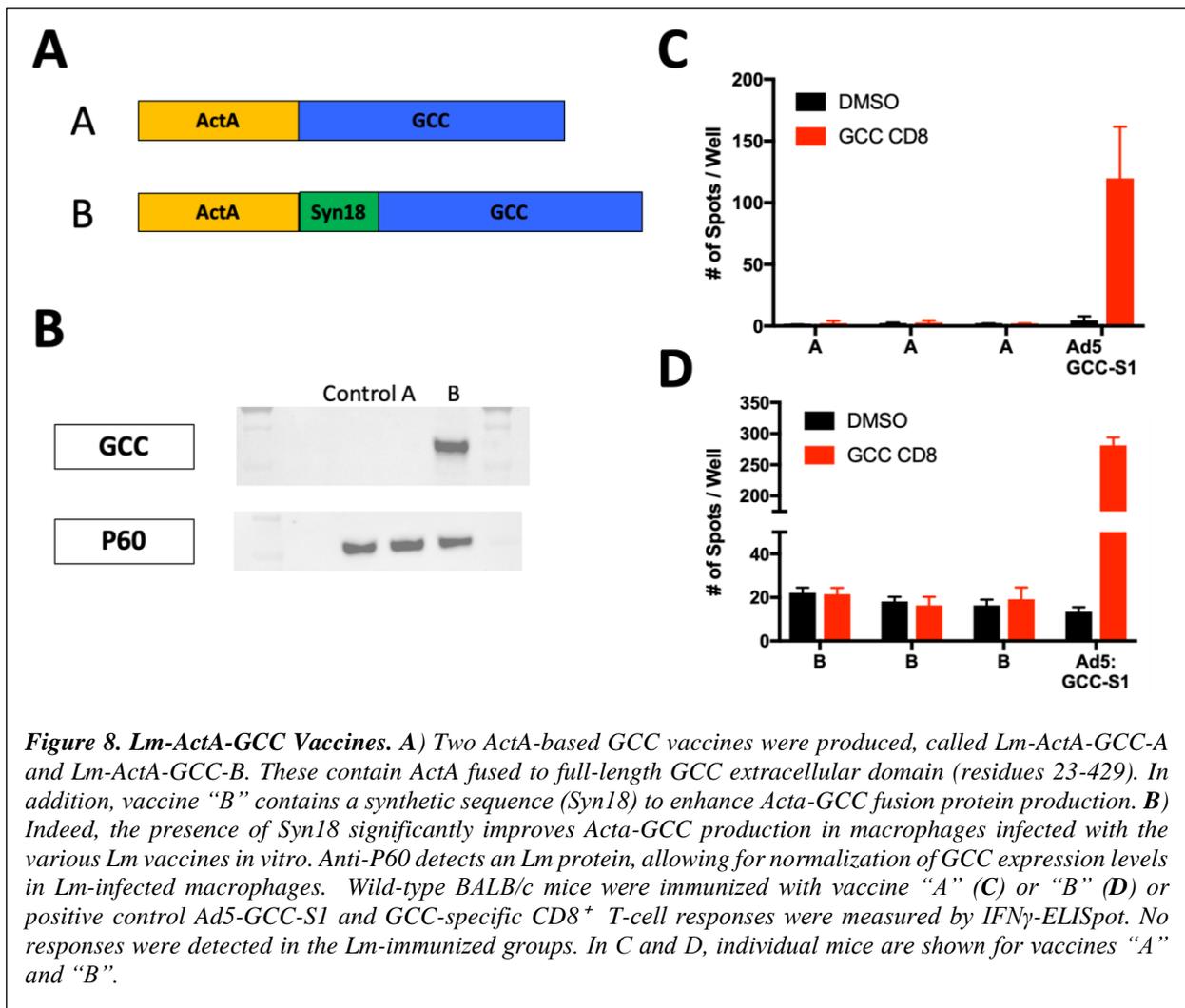
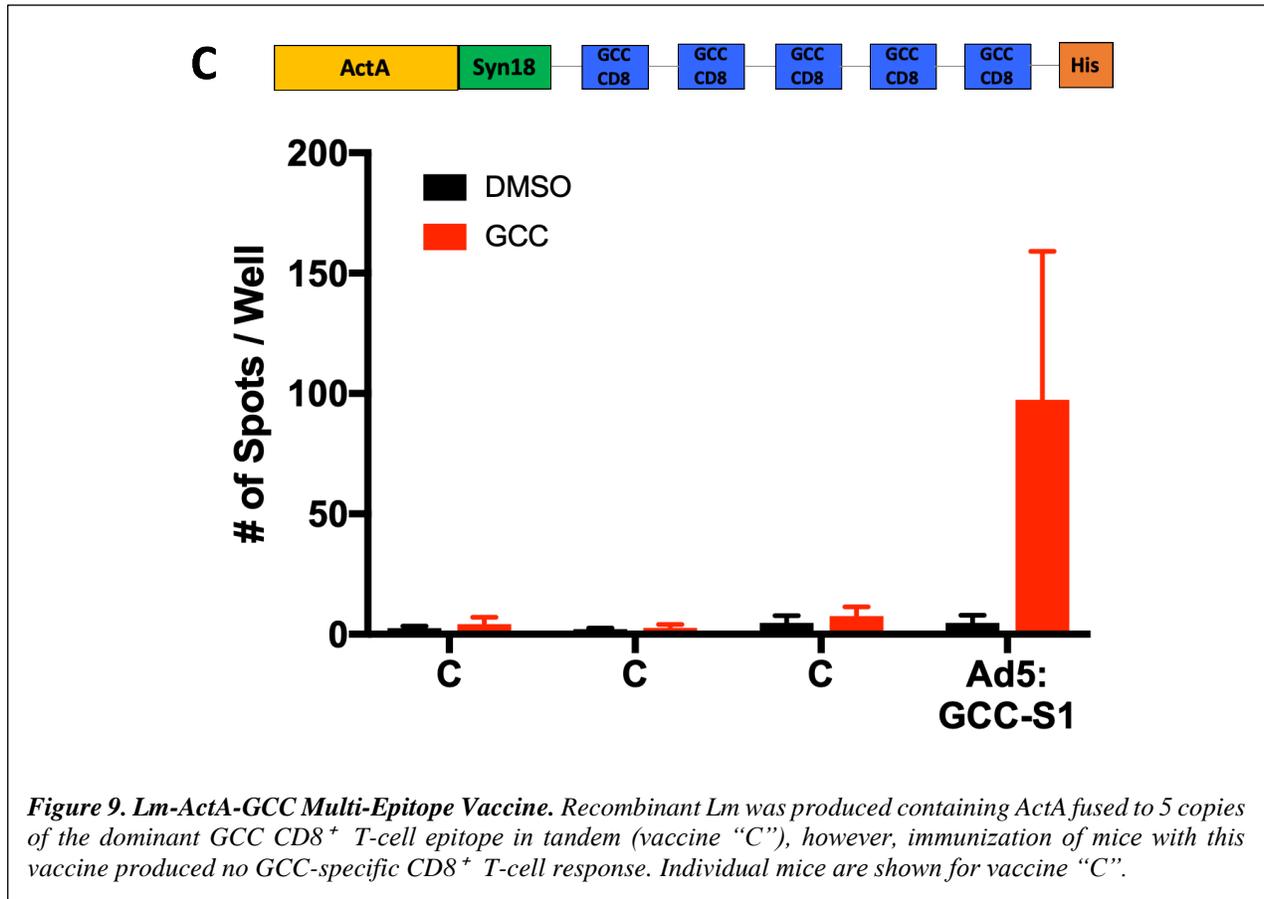


Figure 7. Lm-LLO-GCC Antitumor Immunity. A-C) Wild-type BALB/c mice were immunized with a mixture of Lm-LLO-GCC fragments 1, 2, and 3 (Figures 4-6). Mice received Lm-LLO lacking GCC as a negative control or Ad5-GCC-S1 as a positive control. Mice (n=8 per group) were then challenged with CT26 colorectal cancer cells expressing GCC and luciferase. Tumor burden was imaged and quantified by whole-body bioluminescence imaging (BLI). B-C) Lm-LLO-GCC immunization showed no antitumor activity, while Ad5-GCC-S1 showed complete protection. C) Representative images from day 14.

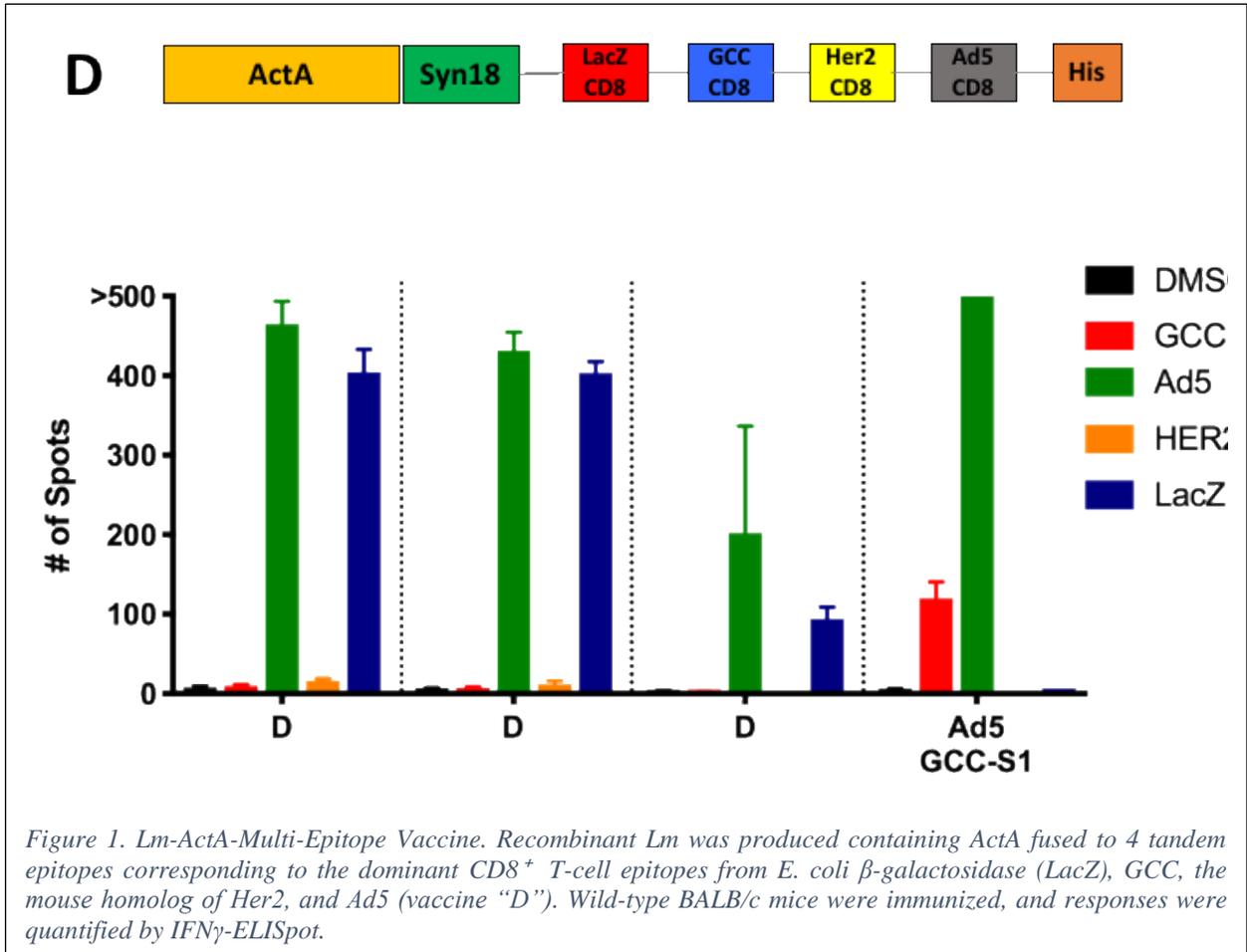
- **2nd Generation Recombinant Lm-LLO-GCC Vaccines**
 - In the context of poor Lm-LLO-GCC vaccine immunogenicity in mice receiving vaccines containing full-length GCC or fragments of GCC (**Figures 1-7**), we hypothesized that GCC may not be properly processed and presented in the context of LLO fusion proteins. Therefore, we produced recombinant Lm vaccines containing GCC fused to ActA, another Lm protein that is highly expressed in the cytosol of Lm-infected cells (**Figure 8**). These vaccines induced GCC fusion protein production in Lm-infected macrophages and expression was significantly enhanced by the inclusion of the Syn18 enhancer sequence (**Figure 8B**). However, the Lm-ActA-GCC vaccines failed to induce GCC-specific CD8⁺ T-cell responses in *Gcc*^{+/+} mice (**Figure 8C-D**).



- Next, we tried to enhance the GCC-specific CD8⁺ T-cell responses by creating a novel construct containing 5 tandem repeats of the GCC CD8⁺ T-cell epitope but lacking other domains of GCC (**Figure 9**). These epitopes were fused to ActA and Syn18 to produce what we expected to be a highly immunogenic construct capable of inducing CD8⁺ T-cell responses. However, immunization of wildtype BALB/c mice with this vaccine failed to generate any responses (**Figure 9**).



- Finally, we hypothesized that there may be something fundamentally flawed with our construct design and it cannot generate CD8⁺ T-cell responses against *any* antigen. To test that hypothesis, we produced a novel construct containing ActA-Syn18 fused to the dominant CD8⁺ T-cell epitope of GCC, as well as CD8⁺ T-cell epitopes from *E. Coli* β-galactosidase (LacZ), mouse Her2, and Ad5 DNA-binding protein (DBP) (Figure 10). While that vaccine produced robust responses against LacZ and Ad5, it failed to generate meaningful responses against GCC and Her2 (Figure 10). Currently, we are generating new Lm constructs designed to enhance antigen processing and presentation of GCC CD8⁺ T-cell



epitopes.

- What opportunities for training and professional development has the project provided?
 - Nothing to report.
- How were the results disseminated to communities of interest?

- Results were disseminated to the local scientific community during poster presentations detailed in Section 6. Products, Inventions, Patent Applications, and or Licenses
- **What do you plan to do during the next reporting period to accomplish the goals?**
 - We have designed and are currently producing several novel Lm constructs that we expect to result in increased GCC epitope presentation and generation of GCC-specific CD8⁺ T-cell responses. Over the remaining project period, we will complete production of these vaccines and test their immunogenicity. Together, those studies and the completed work will establish the potential utility of *Listeria*-based vaccines targeting GCC for colorectal cancer immunotherapy.

4. IMPACT

- **What was the impact on the development of the principal discipline(s) of the project?**
 - Nothing to report
- **What was the impact on other disciplines?**
 - Nothing to report
- **What was the impact on technology transfer?**
 - Nothing to Report
- **What was the impact on society beyond science and technology?**
 - Nothing to Report

5. CHANGES/PROBLEMS

- **Changes in approach and reasons for change**
 - The overall approach, objectives, and scope have not changed. Rather, our initial design for Lm-GCC vaccines proved to have low immunogenicity. We are currently exploring alternative vaccine designs that will allow us to complete our planned objectives. These are detailed in 3. Accomplishments.
- **Actual or anticipated problems or delays and actions or plans to resolve them**
 - We have encountered a significant delay in exploring Lm-GCC vaccines reflecting the poor immunogenicity of the vaccines we have generated on this project. In the context of previous literature and human clinical trials employing *Listeria*-based vaccines for cancer, this was unanticipated. We have generated new Lm-GCC vaccine designs to improve its immunogenicity and complete the objectives of the proposal. However, we may find that no Lm-GCC vaccine is sufficiently immunogenic to induce GCC-specific immune responses and effective antitumor immunity. In that context, our studies may reveal previously unexplored Lm vaccine biology which we will pursue. Thus, while Lm-GCC vaccines may “fail” during the course of this project, we may identify novel mechanisms regulating Lm vaccine efficacy which may be applied to Lm vaccines targeting GCC or other cancer antigens in future studies.
- **Changes that had a significant impact on expenditures**
 - Animal expenditures have been significantly less than expected, reflecting the poor immunogenicity of Lm-GCC vaccines. Because of the low immunogenicity, we have spent most of our time iterating and testing new Lm vaccine designs to enhance immunogenicity, rather than explore the safety and efficacy of Lm-GCC vaccines in models of metastatic colorectal cancer. Because this project was highly novel and explores a previously untested approach to colorectal cancer vaccines, the poor immunogenicity of Lm-GCC vaccines could not be anticipated.
- **Significant changes in use or care of human subjects, vertebrate animals, biohazards, and/or select agents**
 - Nothing to Report
- **Significant changes in use or care of human subjects**
 - Nothing to Report
- **Significant changes in use or care of vertebrate animals.**
 - Nothing to Report
- **Significant changes in use of biohazards and/or select agents**
 - Nothing to Report

6. PRODUCTS, INVENTIONS, PATENT APPLICATIONS, AND/OR LICENSES

- **Publications, conference papers, and presentations**
 - **Journal publications.**
 1. Flickinger JC, Rodeck U, Snook AE. *Listeria monocytogenes as a Vector for Cancer Immunotherapy: Current Understanding and Progress*. Vaccines (Basel). 2018 Jul 25;6(3).
 - Published
 - Acknowledgement of federal support: yes
 - **Books or other non-periodical, one-time publications.**
 1. Nothing to report
 - **Other publications, conference papers, and presentations.**
 1. Presentations
 - Flickinger JC, McCarthy G, Caudill J, Waldman SA, Snook AE. *Development of A Listeria-Based Vaccine Targeting Guanylyl Cyclase C for Metastatic Colorectal Cancer*. Sigma Xi Student Presentation Day, April 10, 2018, Philadelphia, PA
 - Flickinger JC, McCarthy G, Caudill J, Waldman SA, Snook AE. *Development of A Listeria-Based Vaccine Targeting Guanylyl Cyclase C for Metastatic Colorectal Cancer*. Sidney Kimmel Cancer Center Education Retreat, September 24, 2018, Philadelphia, PA
 - Flickinger JC, McCarthy G, Caudill J, Waldman SA, Snook AE. *Development of A Listeria-Based Vaccine Targeting Guanylyl Cyclase C for Metastatic Colorectal Cancer*. Thomas Jefferson University 10th Annual MD/PhD Retreat, September 29, 2018, Philadelphia, PA
- **Website(s) or other Internet site(s)**

Nothing to report
- **Technologies or techniques**

Nothing to report
- **Inventions, patent applications, and/or licenses**

Nothing to report
- **Other Products**

Nothing to report

7. PARTICIPANTS & OTHER COLLABORATING ORGANIZATIONS

- What individuals have worked on the project?

Name:	Adam Snook
Project Role:	Principal Investigator
Researcher Identifier (e.g. ORCID ID):	https://orcid.org/0000-0001-9216-4560
Nearest person month worked:	4
Contribution to Project:	Dr. Snook is the Principal Investigator of this project
Funding Support:	NA

Name:	Tara Abraham
Project Role:	Doctoral student
Researcher Identifier (e.g. ORCID ID):	NA
Nearest person month worked:	12
Contribution to Project:	Tara Abraham has performed work in the area of ELISpot analyses.
Funding Support:	NA

Name:	Trevor Baybutt
Project Role:	Doctoral Student
Researcher Identifier (e.g. ORCID ID):	NA
Nearest person month worked:	12
Contribution to Project:	Trevor Baybutt worked in the areas of recombinant <i>Listeria monocytogenes</i> generation and antitumor efficacy studies.
Funding Support:	NA

Name:	John Flickenger
Project Role:	Doctoral Student
Researcher Identifier (e.g. ORCID ID):	NA
Nearest person month worked:	10
Contribution to Project:	John Flickinger has performed work in the area of recombinant <i>Listeria monocytogenes</i> generation, immunoassays, and antitumor efficacy studies.
Funding Support:	NA

- **Has there been a change in the active other support of the PD/PI(s) or senior/key personnel since the last reporting period?**

Snook, Adam
Other support:

The below Active Support/award ended 12/31/17:

Snook (PI)

Title: Elimination of Adoptive Cell Therapy (ACT) Toxicity by Hypoxic Regulation of Antigen Receptors

Funding Agency: W.W. Smith Charitable Trust

Goals of Project: These studies will define the utility of hypoxia-regulated tumor-targeted receptor expression in ACT. Selective expression of CARs/TCRs in hypoxic microenvironments employing promoters with hypoxia-response elements should eliminate on-target and off-target toxicity, while preserving antitumor efficacy, broadening the repertoire of targetable antigens and permitting safe and effective ACT for all malignancies.

Specific aims/tasks:

Aim 1. Quantify hypoxia-induced, CD19-specific CAR-T cell antitumor efficacy.

Aim 2. Quantify hypoxia-induced, CD19-specific CAR-T cell autoimmunity.

Estimated start and end date: 1/1/16-**12/31/17**

Level (%) of effort in the project: **0 calendar/0% effort**

Point of Contact at the funding agency: Louise Havens

OVERLAP

None

The below Active Support/award ended 11/29/17:

Rodeck (PI)

Title: Combination Immunoradiotherapy Preclinical Evaluation of Efficacy, Sequencing and Biomarkers in Prostate Cancer

Funding Agency: Advaxis

Goals of Project: This sponsored research agreement examines efficacy and biomarkers of combination immunotherapies including a listeria based cancer vaccine in prostate cancer.

Specific aims/tasks: see above goal

Estimated start and end date: 11//30/215-11/29/17

Level (%) of effort in the project: **0 calendar/0% effort**

Point of Contact at the funding agency: Robert Petit, EVP CSO

OVERLAP

None

The below Active Support is new support.

Waldman (PI)

Title: Novel gene delivery systems for colorectal cancer

Funding Agency: Targeted Diagnostics and Therapeutics, Inc.

Total Costs: \$1,500,000

Goals of Project: This project will cover various clinical trials and is a supplement to the above award listed.

Specific aims/tasks: The goal of this project is to further define the utility of guanylyl cyclase C as a novel vaccine target to prevent disease recurrence in GI cancers in a Phase I/II clinical trials.

Estimated start and end date: 01/01/17-12/31/18

Level (%) of effort in the project: 1.2 calendar/10% effort

Point of Contact at the funding agency: Harry A. Arena, CEO

OVERLAP

None

- **What other organizations were involved as partners?**

Nothing to Report

8. SPECIAL REPORTING REQUIREMENTS

- **COLLABORATIVE AWARDS:**
 - Not applicable
- **QUAD CHARTS:**
 - Not applicable

9. APPENDICES

See pages to follow:

Listeria monocytogenes as a Vector for Cancer Immunotherapy: Current Understanding
Immunotherapy: Current Understanding and Progress

Review

Listeria monocytogenes as a Vector for Cancer Immunotherapy: Current Understanding and Progress

John C. Flickinger Jr. ¹, Ulrich Rodeck ² and Adam E. Snook ^{1,*} 

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² Department of Dermatology, Thomas Jefferson University, 1020 Locust Street, Philadelphia, PA 19107, USA; Ulrich.Rodeck@jefferson.edu

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Received: 22 June 2018; Accepted: 23 July 2018; Published: 25 July 2018



Abstract: *Listeria monocytogenes*, a Gram-positive facultative anaerobic bacterium, is becoming a popular vector for cancer immunotherapy. Indeed, multiple vaccines have been developed utilizing modified *Listeria* as a tool for generating immune responses against a variety of cancers. Moreover, over a dozen clinical trials testing *Listeria* cancer vaccines are currently underway, which will help to understand the utility of *Listeria* vaccines in cancer immunotherapy. This review aims to summarize current views on how *Listeria*-based vaccines induce potent antitumor immunity and the current state of *Listeria*-based cancer vaccines in clinical trials.

Keywords: *Listeria*; cancer; vaccine; immunotherapy; bacteria

1. Introduction

The proficiency of the immune system to recognize and eliminate cancer cells, a process summarily called immunosurveillance, is well documented [1]. Failure of immunosurveillance enables the clinical development of cancer and has motivated the search for strategies to rearm and restore effective immune responses to malignant cells. One such strategy includes vaccine development against cancer. In general, cancer vaccines are composed of antigens found in tumor cells, often referred to as tumor-associated antigens (TAA), paired with adjuvants designed to induce an immune response. Antigen-specific T-cell responses induced by cancer vaccines have the potential to produce more targeted elimination of cancer cells than conventional chemotherapy, as well as lead to durable memory responses capable of challenging cancer recurrence. This review aims to summarize one particular approach to cancer vaccination, employing *Listeria monocytogenes* vectors, and its current progress in terms of preclinical development and clinical trials.

Listeria monocytogenes (Lm) is a Gram-positive bacteria most widely known for its ability to infect humans and produce a variety of symptoms, including gastroenteritis, meningitis, and encephalitis [2]. In general, the human immune system mounts potent innate and adaptive immune responses capable of controlling Lm infections. As a result, serious infection by Lm is rare and typically limited to elderly, pregnant, or immunocompromised patients [2]. Decades worth of research on the properties that make Lm immunogenic and how these properties can be exploited have led to its advancement as a vaccine platform for cancer immunotherapy in clinical trials.

Lm has numerous features that make it an attractive vector for cancer immunotherapy. While other vectors may be inhibited through neutralizing antibodies, Lm infection triggers only modest humoral responses which fail to block reinfection [3,4]. This permits repeated administration of Lm-based

vectors to boost T-cell responses in patients as needed. Additionally, and in contrast to DNA or peptide vaccines, Lm strongly induces both innate and adaptive immune responses. Indeed, the ability of bacteria to activate innate immune mechanisms and elicit antitumor responses was first appreciated over 100 years ago when William Coley observed instances of spontaneous cancer regression in sarcoma patients who acquired bacterial skin infections [5]. Beyond innate immune mechanisms, and in contrast to other bacterial vectors, such as *Salmonella*, Lm is a potent stimulator of cytotoxic lymphocytes and cell-mediated immunity [4]. Over 50 years ago, George Mackaness further demonstrated that mice exposed to sublethal doses of Lm developed long-lived, antibody-independent immune responses which protected against a future Lm challenge administered at lethal doses [6]. These and other observations eventually led to the exploration of Lm as a vaccine vector with the aim of inducing similar cell-mediated immune responses towards foreign antigens. This concept was reduced to practice by Paterson and colleagues using Lm-expressing β -galactosidase antigen to induce a cytotoxic lymphocyte response that killed cancer cells expressing β -galactosidase [7]. Since the publication of this seminal paper over 25 years ago, numerous Lm vaccines have been developed for the treatment of a variety of cancers. Here, we will provide an overview of the mechanisms underlying Lm-based cancer vaccines and summarize the current state of Lm vaccines in cancer clinical trials.

2. Pathogenesis of *Listeria* Infection

In nature, Lm typically enters through the gastrointestinal system after ingestion of contaminated food, where it crosses the intestinal epithelium and spreads into the bloodstream [2]. However, in the context of immunotherapeutic applications, Lm is largely administered intravenously, bypassing the intestinal epithelium. Once in the bloodstream, Lm can enter a variety of organs, including the liver, brain, and placenta [2,8]. While it can survive in the extracellular environment, its preferred niche is the cell cytoplasm where it undergoes replication [9]. As Lm transitions into intracellular compartments, numerous immune mechanisms are activated. These mechanisms are essential for understanding Lm vaccine therapy and are illustrated in Figure 1.

Lm entry into mammalian cells is primarily mediated through a family of proteins called internalins. Of particular interest are the internalin A (InlA) and internalin B (InlB) proteins, which guide Lm invasion through binding to receptors on host cells and triggering receptor-mediated endocytosis [8]. Importantly, InlA and InlB bind to different receptors and promote infection of different cell types. InlA binds E-cadherin which is found on epithelial cells, while InlB interacts with the hepatocyte growth factor, Met [10,11]. In addition to entry via cell surface receptors, Lm is actively taken up via phagocytosis by antigen-presenting cells (APCs) [2]. Importantly, as Lm interacts with the outside surfaces of mammalian cells, innate immune responses against Lm begin to mount. On the surface of the cell, as well as inside phagosomes, pathogen-associated molecular patterns (PAMPs) expressed by Lm are recognized by mammalian Toll-like receptors (TLRs), leading to the activation of NF- κ B signaling and the expression of pro-inflammatory genes [2,12].

Upon internalization into phagosomes, it appears that Lm can undergo one of two fates. It is estimated that the majority of bacteria are killed upon phagosome–lysosome fusion [13], providing a source of antigens for MHC class II-dependent presentation and priming of Lm-specific CD4⁺ T-cell responses [4]. However, Lm has evolved mechanisms to escape lysosomal degradation and enter the cytosols of infected cells by a well-defined mechanism. As Lm transitions from the extracellular environment into the host cell, it begins to express the transcription factor PrfA [9]. PrfA activates numerous virulence genes, including those responsible for phagosome escape, such as the pore-forming toxin listeriolysin O (LLO) and two different phospholipases, PlcA and PlcB. Secreted LLO molecules oligomerize at the phagosomal membrane to form a β -barrel pore-like structure through which Lm can pass [14], while phospholipases mediate exit through direct hydrolysis of membrane lipids [15]. Once outside the phagosome, Lm-secreted peptides enter the host cell cytosol where they can be degraded by proteasomes and loaded onto MHC class I molecules for presentation to cytotoxic T

cells [4]. The direct secretion of antigens into the cytosol, as well as prior degradation in phagosomes, results in the induction of potent CD4⁺ and CD8⁺ T-cell responses to Lm antigens [4,16,17].

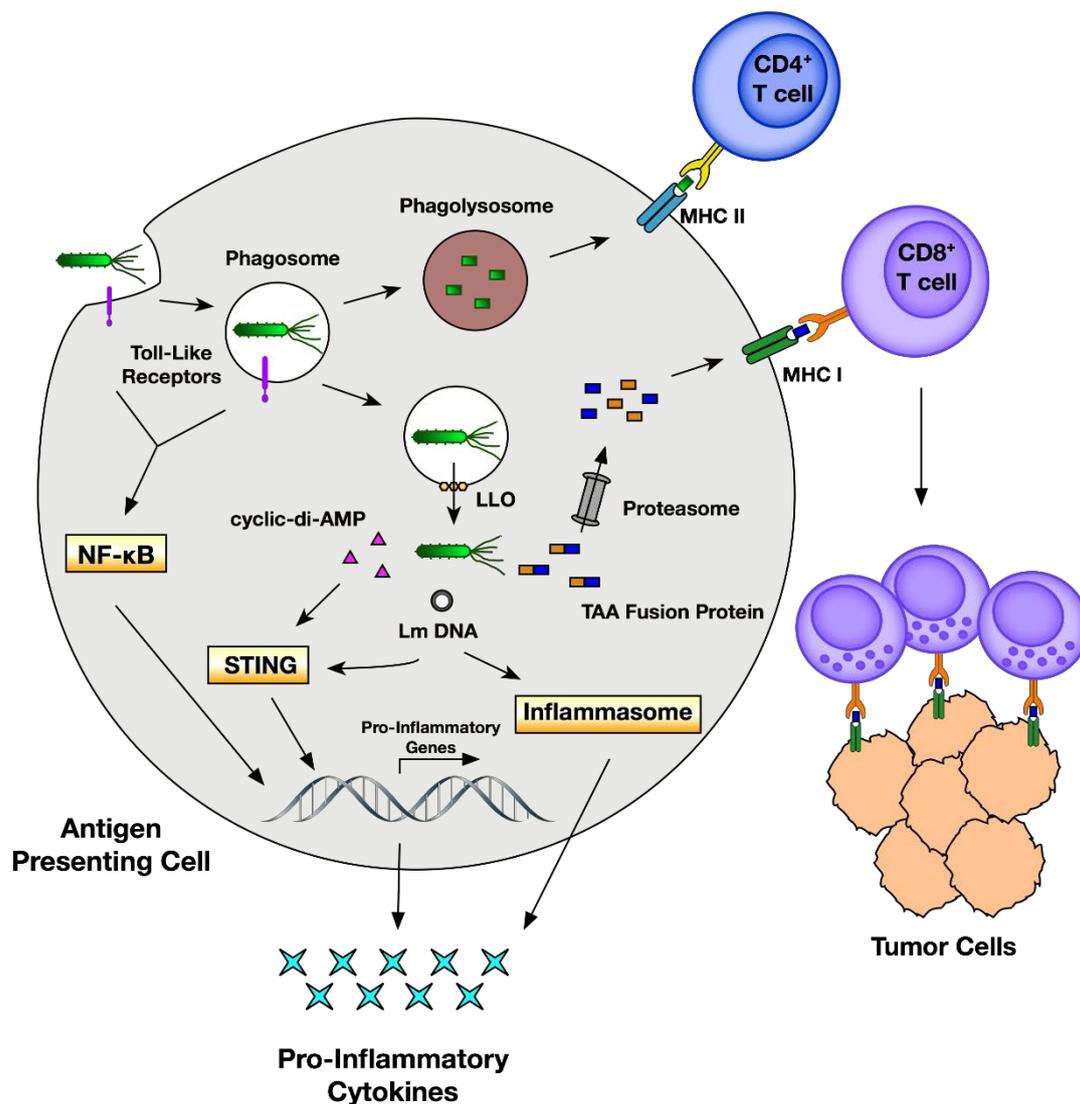


Figure 1. Innate and adaptive immune responses to recombinant *Listeria*. *Listeria monocytogenes* (Lm) is internalized by antigen-presenting cells into phagosomes. During entry, Lm is sensed by toll-like receptors leading to the activation of NF-κB and synthesis of pro-inflammatory genes. Phagosomes may then fuse with lysosomes to form phagolysosomes where Lm can be killed, leading to loading of its antigens onto MHC class II for activation of CD4⁺ helper T cells. Alternatively, Lm can express the pore-forming toxin listeriolysin O (LLO) to perforate phagosomes and gain entry into the cytosol. Once in the cytosol, recombinant Lm secretes tumor-associated antigens (TAA) as fusion proteins with Lm antigens into the cytosol where they can be degraded by proteasomes and loaded onto MHC class I for activation of TAA-specific CD8⁺ cytotoxic T lymphocytes. Additionally, cytosolic Lm triggers further induction of pro-inflammatory pathways through secretion of the cyclic dinucleotide cyclic-di-AMP and detection of Lm DNA. Secreted cyclic-di-AMP directly stimulates the STING pathway and negatively regulates the NF-κB inhibitor RECON (not shown) [18], while the presence of genomic Lm DNA can lead to the activation of STING and inflammasome pathways, both of which contribute to the transcription of pro-inflammatory genes and cytokines.

Having escaped the phagosome, Lm is free to move throughout the host cell cytosol via expression of the virulence factor actin assembly-inducing protein (ActA). Anchored to the surface of Lm,

ActA protein interacts with the Arp2/3 complex to promote nucleation and polymerization of actin monomers into filaments [19]. Forces generated via actin polymerization are capable of projecting Lm throughout the cytosol as well as into the plasma membrane of the infected cell, forming a protrusion that can be internalized by a neighboring cell and leading to dissemination of the infection. [19].

Similar to the sensing of Lm on the cell surface and inside phagosomes by toll-like receptors, additional pattern recognition receptors recognize Lm in the cytosol and further stimulate pro-inflammatory pathways. One route is through the activation of retinoic acid-inducible gene I (RIG-I)-like receptors by Lm RNA, leading to the activation of the mitochondrial antiviral signaling protein (MAVS) and transcription of interferon (IFN) genes [20,21]. Further transcription of IFN genes is mediated through activation of the STING pathway through at least two mechanisms. First, Lm can secrete cyclic diadenosine monophosphate (cyclic-di-AMP), directly activating STING [22]. Second, Lm lysis can expose genomic DNA leading to activation of the cyclic GMP-AMP synthase, production of cGAMP, and activation of STING [23]. Regardless of the route, activation of the STING pathway promotes the transcription of type I IFN genes, including IFN- β [22,24]. Importantly, IFN- β and activation of the STING pathway improve the priming of CD8⁺ T cells in the tumor microenvironment, leading to potent antitumor responses [25,26]. Paradoxically, Lm mutants, secreting higher levels of cyclic-di-AMP, elicit reduced T-cell responses, suggesting that the role of STING activation in Lm infection is not fully understood [24]. Finally, Lm infection can activate NLRP3 and AIM2 receptors, leading to activation of the inflammasome and further increasing pro-inflammatory cytokines [2,27].

3. Development of Attenuated *Listeria* Strains for Vaccination

Because wild-type *Listeria* is pathogenic and not suitable for clinical use, considerable effort has been expended to improve Lm safety. The ideal Lm strain would minimize pathogenicity while maximizing immunogenicity towards the target antigen. Multiple strategies to attenuate Lm have been developed towards that end.

3.1. Deletion of Virulence Genes

A widely employed strategy to attenuate Lm involves deleting genes responsible for Lm tropism and cell-to-cell transmission. As mentioned previously, Lm tropism into non-phagocytic cells, such as hepatocytes, is mediated by the virulence factor internalin B. Therefore, deletion of *inlB* is expected to limit liver toxicity and render phagocytic antigen-presenting cells the primary target of Lm infection. Indeed, $\Delta inlB$ mutants display reduced hepatocyte entry while infecting monocytes as effectively as wild-type strains [28]. Further attenuation can be achieved by deleting the virulence factor *actA* [28]. In wild-type Lm, ActA expression allows Lm to polymerize host actin filaments to maneuver through the host cytosol and spread from one cell into another [2,9]. Deletion of *actA* is thus expected to confine Lm to the cytoplasm of the cell it initially infects, limiting the spreading of infection and reducing toxicity. Evidence for this is supported by experiments demonstrating that $\Delta actA$ strains have a much higher LD₅₀ (>1000-fold) than wild-type strains [29]. By deleting both *inlB* and *actA*, it is possible to selectively infect antigen-presenting cells and reduce off-target toxicity. Vaccines using $\Delta actA/\Delta inlB$ strains also exhibit rapid clearance of infection from the liver and spleen compared to single $\Delta actA$ or $\Delta inlB$ mutants [28]. Importantly, despite a rapid clearance of Lm, $\Delta actA/\Delta inlB$ strains demonstrate comparable antitumor responses [28]. The Lm $\Delta actA/\Delta inlB$ strain, also known as Live Attenuated Double-Deleted (LADD) (Aduro BioTech Inc., Berkeley, CA, USA), forms the basis for many vaccines in clinical trials.

3.2. Episomal Replacement of Virulence or Metabolic Genes

While attenuation can be achieved through gene deletion, the deletion of a gene followed by reintroduction of that gene via plasmid can also result in attenuation. It is likely that episomal gene expression leads to different quantities or spatial patterns of gene product than expression from the host genome [17]. As described above, PrfA is a master transcription factor that regulates the transcription

of multiple Lm virulence factors, including LLO, ActA, and phospholipases [9]. Indeed, Lm strains deficient in *prfA*, such as XFL-7, are severely attenuated [30,31]. Moreover, due to the lack of LLO and phospholipase expression, *prfA* mutants are incapable of phagolysosome escape and invasion into the host cytosol [31]. Partial restoration of virulence is achieved by the addition of plasmid-encoded PrfA into which the tumor-associated antigens (TAA) are cloned as well [30,32,33]. The result is an immunogenic strain that is significantly attenuated compared to wild-type Lm [30,32,33].

A similar attenuation strategy has been employed utilizing the $\Delta dal/\Delta dat$ strain (Lmdd). Lm requires the *dal* and *dat* genes to synthesize D-alanine to build peptidoglycan and lipoteichoic acid [34]. Strains that are deficient in both of these genes fail to replicate in vitro unless supplemented with exogenous alanine [35,36]. Furthermore, immunogenicity of the Lmdd strain in vivo depends on co-administration of alanine [37]. However, by transforming the Lmdd strain with a plasmid encoding the *dal* gene under a constitutively active promoter, it is possible to partially restore virulence and achieve immunogenicity without alanine supplementation [38]. Often, the Lmdd strain is further attenuated by the deletion of *actA* creating what is often called the LmddA strain [38,39]. Both the LmddA and the XFL-7 strain are currently being evaluated in clinical trials by Advaxis Inc., Princeton, NJ, USA.

3.3. Killed but Metabolically Active

Heat-killed Lm and strains deficient in phagolysosomal escape have long been noted to be poor inducers of CD8⁺ T-cell responses [40,41], suggesting that Lm entry into the host cytosol is required for the effective activation of CD8⁺ T cells. As a result, Lm vaccine development has relied on attenuated, rather than killed, Lm formulations. However, live attenuated Lm strains are capable of inducing listeriosis in some cases and may not be safe to use in immunocompromised patients [4,42]. In an effort to further improve the safety profile of Lm vaccines, killed but metabolically active (KBMA), Lm vaccines have been developed. Lm strains deficient in nucleotide excision repair can be photochemically inactivated by the DNA crosslinking agent psoralen coupled with UV irradiation [43]. This process induces irreversible DNA damage and renders Lm incapable of replication. However, for a period of hours after photochemical treatment, Lm is capable of invading host cells, escaping into the cytosol, and secreting TAA products to elicit antitumor CD8⁺ T-cell responses [43,44]. Initially, KBMA vaccines were found to invoke inferior protective responses compared to live, attenuated vaccines [45]. However, enhanced CD8⁺ T-cell responses have been achieved using KBMA vaccines with a constitutively activating *prfA* mutation [44]. While promising, KBMA Lm vaccines have yet to enter clinical trials.

4. Fusion with *Listeria* Antigens Enhances Antitumor Responses

While it is possible to generate therapeutic immune responses by engineering Lm to express a TAA alone, nearly all Lm vaccines express TAAs as chimeric proteins fused to a native Lm antigen. Initial studies piloting Lm vaccines fused TAAs to highly-secreted Lm proteins as a method to increase the delivery of TAAs into the cytosols of host cells [46]. However, it is now well appreciated that certain Lm proteins, when fused to TAA, can also act as adjuvants, boosting therapeutic efficacy. The two most common fusion partners with TAAs include modified versions of LLO and ActA.

4.1. Listeriolysin O

Listeriolysin O (LLO) is a virulence factor in the Lm life cycle and perforates the phagosome to aid Lm entry into the cytosol. Because LLO could also perforate the plasma membrane, vaccines incorporating LLO fusion proteins are typically engineered to express it as a truncated version (tLLO) which is deficient in the hemolytic domains necessary for pore formation [30,33]. Effective antitumor immunity using tLLO-TAA constructs was first demonstrated by Gunn et al. [30]. In these studies, mice were injected with a cancer cell line expressing the human papillomavirus (HPV) antigen E7 and then vaccinated with Lm expressing E7 alone or E7 as a fusion protein with tLLO (tLLO-E7). While E7

expressing Lm had almost no effect on tumor growth, immunization with tLLO-E7-expressing Lm induced complete regression of 75% of the tumors [30]. Since these initial observations, numerous studies examining the role of tLLO in antitumor immunity have been conducted, and it is believed that tLLO enhances vaccine responses through a combination of different mechanisms.

It has been suggested that tLLO enhances the delivery of TAA to the proteasome, facilitating antigen processing and MHC class I presentation [47]. Indeed, LLO contains PEST-like sequences and destabilizing N-terminal residues that enhance protein degradation and affect cytosolic LLO levels [48–50]. This hypothesis has been supported by direct comparison of Lm vaccines expressing either E7 alone, E7 fused to the isolated PEST sequence (PEST-E7), E7 fused to a version of tLLO lacking the PEST sequence (Δ PEST-E7), or the full tLLO sequence fused to E7 (tLLO-E7). Immunization with PEST-E7 elicited increased CD8⁺ T-cell responses and antitumor immunity compared to vaccination with E7 alone [47], suggesting that PEST sequences can improve vaccine responses towards TAAs. Consistent with this view, vaccination with Δ PEST-E7 generated reduced CD8⁺ T-cell responses and antitumor immunity compared to tLLO-E7 [47]. However, immunization with Δ PEST-E7 also elicited superior antitumor responses compared to the E7 vaccine, suggesting that tLLO may enhance responses through additional mechanisms independent of PEST sequences [32,47]. One possibility is that tLLO may signal through pattern recognition receptors. Indeed, LLO and other pore-forming toxins can activate TLR4 [51]. Initially, these immunogenic properties were thought to depend on LLO cholesterol-binding domains which are involved in LLO-induced pore formation [51]. However, later experiments indicated that even non-hemolytic forms of LLO are capable of inducing dendritic cell maturation and the synthesis of proinflammatory cytokines, suggesting that LLO might be signaling through pattern recognition receptors independent of its cytotoxic attributes [52]. However, this study was unable to identify a specific pattern recognition receptor responsible for these effects.

Since many TAAs targeted by vaccines are also expressed by normal tissues, effective vaccines must overcome tolerance [4]. Importantly, multiple lines of evidence suggest that tLLO-TAA fusion constructs can remodel the immunosuppressive tumor microenvironment and overcome tolerance mechanisms. While Lm expressing E7 alone elicits poor antitumor responses, its effectiveness is substantially increased by regulatory T-cell depletion [30]. Moreover, experiments examining tumor-infiltrating CD4⁺CD25⁺ regulatory T cells (Tregs) revealed that vaccination with Lm expressing E7 alone increased the abundance of Tregs, while the tLLO-E7 vaccination decreased Tregs [53,54], supporting the hypothesis that tLLO may enhance antitumor responses through changes in Tregs. These results may be partly explained by a preferential expansion of CD4⁺Foxp3⁻ cells upon vaccination with tLLO-E7 [54]. Interestingly, vaccination with Lm strains expressing tLLO alone appears to affect the CD4⁺ effector and regulatory T-cell numbers similarly to those obtained with the tLLO-E7 fusion protein [54]. In addition, tLLO-TAA constructs reportedly reduce the number and functional activity of myeloid-derived suppressor cells (MDSCs) in the tumor microenvironment, potentially contributing to the ability of Lm to break peripheral tolerance against TAAs [55]. Interestingly, immunity induced by tLLO-TAA fusion constructs does not appear to be limited by central tolerance mechanisms. Experiments using transgenic mice expressing the E7 antigen in the thymus, resulting in central tolerance to E7, demonstrated that tLLO-E7 vaccines could expand low-avidity E7-specific T cells and produce antitumor responses [56]. Additionally, tLLO fusions widen the breadth of T-cell receptors (TCRs) recognizing self-antigen. In support of this notion, DNA vaccines containing a fusion of the self/tumor antigen HER2 with tLLO induced T-cell responses towards subdominant epitopes [32]. In this study, DNA vaccines encoding HER2 elicited responses to only two HER2 epitopes, while a tLLO-HER2 vaccine elicited responses against four HER2 epitopes [32]. However, Lm-based, rather than DNA-based, vaccines expressing HER2 fragments fused with tLLO induced response against nine HER2 epitopes, suggesting that even broader responses are generated in the context of Lm [32].

4.2. ActA

A different widely used strategy for Lm vaccination involves fusing TAAs to ActA. Similar to fusion with tLLO, these approaches tend to rely on truncated ActA sequences of varying lengths. Some investigators have used the first 100 amino acids of ActA [44,57], while others used the first 420 amino acids of the protein [58,59]. Similar to tLLO, fusion of E7 with ActA produces superior antitumor immunity compared to E7 alone [60]. However, explanations for superior therapeutic responses are incomplete. Like tLLO, ActA contains N-terminal PEST domains which may facilitate proteasomal degradation and enhance TAA processing [4,61]. However, experiments have also suggested that ActA may exert immunostimulatory effects independent of ActA–TAA fusion. Using protein vaccines containing either ActA, E7, or ActA covalently conjugated to E7 (ActA-E7), immunization with a mixture of ActA and E7 produced similar CD8⁺ T-cell responses and antitumor immunity compared to vaccination with ActA-E7 fusion protein [60]. It has been speculated that ActA may possess PAMP-like attributes, leading to the production of pro-inflammatory cytokines and maturation of antigen-presenting cells [60]. Much like tLLO, the precise mechanisms for its adjuvant-like properties are not entirely understood.

At least two studies have directly compared Lm vaccines expressing ActA-E7 to those expressing tLLO-E7 [56,59]. Vaccination with either ActA-E7 or tLLO-E7 resulted in similar levels of circulating E7-specific CD8⁺ T cells and yielded similar antitumor effects [59]. Interestingly, vaccination with tLLO-E7 produced higher percentages of tumor-infiltrating E7-specific CD8⁺ T cells [58,59]. Both vaccines generated similar levels of E7-specific responses in transgenic mice expressing E7, indicative of a comparable potency in breaking tolerance [56]. While it is unclear which strategy, if any, is superior, it is well-established that fusion of TAA to either ActA or tLLO can boost immune responses towards TAAs and enhance antitumor immunity [4,30].

5. *Listeria* Vaccines in Clinical Trials

To date, over 30 clinical trials testing 10 different Lm cancer vaccines have been initiated (Table 1). Two companies have been at the forefront of advancing Lm-based vaccines through clinical trials, with each using unique approaches. In general, Lm-based vaccines developed by Aduro BioTech Inc. are derived from the LADD[®] strain. TAA genes are cloned into integration vectors allowing for stable incorporation into the Lm genome [62]. Expression of TAAs is regulated by the ActA promoter and occurs as a fusion protein with a modified form of ActA [17]. In contrast, Lm vaccines manufactured by Advaxis Inc. are based on either the XFL-7 or the LmddA strains described above. Expression of TAAs occurs episomally as a fusion protein with tLLO under control of the hly (LLO) promoter [38,39].

5.1. ADXS11-001 (*Axalimogene Filolisbac* [AXAL])

ADXS11-001, also known as Axalimogene Filolisbac or AXAL, is a vaccine against cancers caused by the human papillomavirus (HPV). HPV expresses the E6 and E7 oncoproteins which directly promote cell division, genomic instability, and tumorigenesis by interfering with the functions of the tumor suppressor proteins p53 and RB [63]. Approximately 5% of all cancers world-wide are caused by HPV infection, with most cases being attributed to HPV subtypes 16 and 18 [64]. AXAL is based on the XFL-7 strain, engineered to secrete the E7 protein from HPV 16 fused to tLLO [30,58,59]. Numerous preclinical studies have demonstrated its ability to induce regression of HPV-transformed tumors, leading to its advancement to multiple clinical trials [30,58,59].

Table 1. *Listeria* cancer vaccines in clinical trials.

Vaccine	Antigen	Cancer Indication	Drug Combination	Phase	Completion	Studies	Identifier	Company
ADXS11-001	HPV 16 E7	Cervical	Vaccine alone	I	2009	[65]	N/A	Advaxis
				I/II	12/2018 ¹		NCT02164461	
				II	04/2016		NCT01116245	
				II	10/2018 ¹		NCT01266460	
		III	06/2021	NCT02853604				
		Cervical and Oropharyngeal	Vaccine + chemotherapy	II	2017	[66]	CTRI/2010/091/001232	
				I/II	12/2019		NCT02291055	
		Oropharyngeal	Vaccine alone	I	11/2014		NCT01598792	
II	08/2019			NCT02002182				
Anal	Vaccine + chemoradiation Vaccine alone	I/II	02/2018	[67]	NCT01671488			
		II	03/2022		NCT02399813			
Lung	Vaccine + chemotherapy	II	03/2019 ¹		NCT02531854			
ADXS31-142	PSA	Prostate	Vaccine + αPD-1	I/II	12/2019	[68]	NCT02325557	
ADXS31-164	HER2	HER2 + Solid Tumors	Vaccine alone	I/II	12/2018		NCT02386501	
ADXS-NEO	Personal Neo-antigens	Colon, Lung, Head and Neck	Vaccine alone	I	09/2020		NCT03265080	
CRS-100 (ANZ-100)	None	Hepatic metastases	Vaccine alone	I	02/2008	[69]	NCT00327652	
CRS-207	Mesothelin	Pancreatic, Lung, Ovarian and Mesothelioma	Vaccine alone	I	02/2009	[69]	NCT00585845	Aduro
				II	08/2016	[70]	NCT02004262	
		II	02/2017	[71]	NCT01417000			
		II	01/2019		NCT02243371			
		II	10/2019		NCT03190265			
		II	06/2023		NCT03006302			
		Ovarian, Fallopian and Peritoneal	Vaccine + αPD-1 + IDO1 inhibitor	I/II	12/2018		NCT02575807	
		Gastroesophageal	Vaccine + αPD-1	II	05/2019		NCT03122548	
Mesothelioma	Vaccine + chemotherapy Vaccine + αPD-1	I	12/2018	[72]	NCT01675765			
		II	03/2019 ¹		NCT03175172			
ADU-623	EGFRvIII and NY-ESO-1	Brain	Vaccine alone	I	12/2018	[73]	NCT01967758	
pLADD	Personal Neo-antigens	Colorectal	Vaccine alone	I	12/2020		NCT03189030	
JNJ-64041757(ADU-214)	EGFRvIII and mesothelin	Lung	Vaccine alone Vaccine + αPD-1	I	03/2020	[74]	NCT02592967	Janssen ²
				I/II	03/2022		NCT03371381	
JNJ-64041809(ADU-741)	Multiple prostate antigens	Prostate	Vaccine alone Vaccine + anti-androgen	I	06/2018		NCT02625857	
				II	09/2018		NCT02906605	

¹ Primary completion; ² Licensed from Aduro.

Most clinical trials studying AXAL have been in patients with cervical cancer. After a Phase I clinical trial demonstrated safety [65], multiple Phase II trials were initiated. Results from a Phase II trial in India comparing AXAL and AXAL plus cisplatin to a historical control were recently published [66]. In this trial, patients with refractory cervical cancer received either three doses of AXAL or four doses of AXAL with five infusions of cisplatin. While median overall survival (OS) was equivalent, patients receiving AXAL or AXAL plus cisplatin experienced 1.5 to 2-fold increases in overall survival at 12 months (34.9%) and 18 months (24.8%) [66]. In addition to combination with chemotherapy, AXAL is also being tested in combination with immune checkpoint inhibitors. Preclinical models using AXAL with α PD-1 inhibitors have demonstrated a reduction in Treg and MDSC abundance and an increased antitumor response compared to AXAL alone [75]. A Phase I/II trial combining AXAL with the α PD-1 inhibitor durvalumab was initiated but briefly put on hold after a patient died of respiratory failure (ClinicalTrials.gov identifier NCT02291055). After an investigation, the FDA reinstated this trial and it is currently ongoing. Of great interest, AXAL was recently advanced into the double-blind, randomized Phase III AIM2CERV clinical trial, comparing AXAL to a placebo in patients with high-risk, locally-advanced cervical cancer (ClinicalTrials.gov identifier NCT02853604). This is currently the only Phase III trial testing an Lm-based vaccine. Patients in the AIM2CERV trial will receive chemoradiation with curative intent followed by multiple infusions of AXAL or placebo. Dosing of AXAL or the placebo is to occur every 3 weeks for the first 3 months, followed by an infusion every 8 weeks for a total of five doses or until disease recurrence. The primary endpoint will compare disease-free survival (DFS) in patients receiving AXAL or placebo and is expected to be met in June 2020.

While most trials with AXAL have been in cervical cancer, AXAL is also being pursued in patients with HPV-associated oropharyngeal, anal, and lung cancers. In oropharyngeal cancer, a Phase I trial was terminated early after a patient suffered a dose-limiting toxicity (ClinicalTrials.gov identifier NCT01598792). Despite this setback, a Phase II trial testing AXAL as a monotherapy in anal cancer continues to recruit patients (ClinicalTrials.gov identifier NCT02002182). In anal cancer, a Phase I trial studying AXAL in combination with radiation and the chemotherapies mitomycin and 5-FU was recently completed (ClinicalTrials.gov identifier NCT01671488). Of the nine patients who completed treatments, eight were progression-free at a median follow-up time of 42 months [67]. Based on those results, a subsequent Phase II trial assessing AXAL as a monotherapy in anal cancer was initiated (ClinicalTrials.gov identifier NCT2399813). This trial is expected to be completed in March 2022.

In addition to AXAL, Advaxis is also pursuing a second-generation version called ADXS-DUAL in collaboration with Bristol-Myers Squibb. ADXS-DUAL possess E7 from both HPV 16 and HPV 18 serotypes. Although the majority of clinical trials by Advaxis have focused on AXAL, Advaxis is also advancing Lm vaccines against the tumor antigens PSA (prostate-specific antigen) and HER2 (human epidermal growth factor 2).

5.2. ADXS31-142

PSA is a serine protease whose expression is normally confined to the prostate. While its utility as a serum diagnostic marker in prostate cancer remains controversial, it is well-accepted that PSA can act as an immunotherapeutic target in prostate cancer [76]. Early studies piloting tLLO-PSA vaccines were based on the XFL-7 strain of Lm; however, clinical studies use ADXS31-142 based on the LmddA strain. Both versions have been shown to modestly reduce tumor burden and the number of tumor-infiltrating Tregs in mice inoculated with the prostate cancer cell line TPSA23 [33,38]. Based on the hypothesis that radiation therapy may induce immunogenic tumor cell death and promote favorable inflammatory environments for T-cell activation, follow-up studies assessed ADXS31-142 in combination with radiation therapy [77,78]. While ADXS31-142 or radiation alone slowed TPSA23 tumor growth, combination therapy led to multiple instances of complete tumor regression [77]. Synergistic antitumor responses were associated with significant increases in PSA-specific T cells in the spleen and tumor [77] as well as enhanced intratumoral Th1 responses [78]. Recent reports have

demonstrated an additional therapeutic benefit by combining ADXS31-142 and radiation therapy with α PD-1 or α PD-L1 inhibitors [78].

Currently, ADXS31-142 is being tested in a Phase I/II trial in patients with metastatic castration-resistant prostate cancer. The trial (ClinicalTrials.gov identifier NCT02325557), also known as the KEYNOTE-046 trial, is assessing progression-free survival following ADXS31-142 alone or in combination with the α PD-1 antibody pembrolizumab. While the trial is not yet completed, an early report found that patients vaccinated with ADXS31-142 generated T-cell responses against multiple prostate antigens beyond PSA through a process known as epitope spreading [68]. In this context, it is believed that PSA-specific T cells inducing prostate cancer cell death and the production of inflammatory cytokines result in the release of additional tumor antigens and induction of immune responses against additional epitopes, ultimately broadening the therapeutic scope of the vaccine.

5.3. ADXS31-164

HER2 is a receptor tyrosine kinase that is frequently overexpressed by cancers, including breast, salivary gland, and bone [79,80]. HER2 expression is often associated with poor prognosis, as HER2 signaling has been shown to directly activate pathways responsible for tumor growth and survival [79]. While antibody-based therapies targeting HER2 have been developed, many patients still experience recurrence after treatment. In addition to antigen escape, antibody-based therapies against HER2 primarily induce antibody-dependent cytotoxicity in the absence of adaptive immune responses. Indeed, poor Th1 responses after anti-HER2 antibody therapy strongly correlated with recurrence, leading to speculation that vaccines boosting Th1 responses could improve the standard of care [81,82].

Due to its large molecular mass, early Lm-based HER2 vaccine designs consisted of five different Lm vaccines, with each expressing tLLO fused to a unique region of the rat HER2 extracellular or intracellular domain [32]. Individually, each Lm strain antagonized the growth of the HER2 expressing NT-2 tumor cell line to a similar extent. Based on this proof of concept, a single Lm vaccine against human HER2, known as ADXS31-164, was subsequently developed [39,83]. ADXS31-164 is derived from the attenuated LmddA strain and is engineered to secrete a chimeric tLLO fusion protein containing immunogenic regions from two extracellular domain sequences and one intracellular domain sequence of HER2 [83]. The chimeric HER2 protein is designed to encompass most of the known human MHC class I epitopes of the HER2 protein. Preclinical studies with this vaccine demonstrated the ability of ADXS31-164 to reduce the number of regulatory T cells in tumors and increase the CD8⁺ T-cell/Treg ratio [39]. Impressive results with ADXS31-164 in preclinical studies has led to its advancement to clinical trials in canines and humans.

Recently, results from a Phase I trial of ADXS31-164 in canine osteosarcoma were published [84]. In this trial, canines received three doses of ADXS31-64 following amputation and chemotherapy. Out of 18 immunized dogs, 15 developed T-cell responses against HER2. Impressively, this led to a significant reduction in metastatic disease and increase in survival. Canines vaccinated with ADXS31-164 had a median survival time of 956 days, and 56% were still alive three years after chemotherapy, compared to a historical control of 423 days and 22%, respectively [84]. Canines with immune responses to the intracellular HER2 domain had the most robust effects on survival, potentially related to the fact that the intracellular HER2 domain is indispensable for oncogenic cell signaling [85]. In contrast to HER2/neu antibodies, which are encumbered by adverse effects, including cardiotoxicity [86], ADXS31-164 was not associated with cardiotoxicity in canines [84]. In 2014, Advaxis licensed the use of ADXS31-164 in canines to Aratana Therapeutics Inc. where it was renamed AT-014. The USDA, in early 2018, granted conditional clinical approval for the use of AT-014 in canine osteocarcinoma. Currently, AT-014 is the only Lm vaccine to receive any clinical approval. While data from mice and dogs indicate promising potential, results from human trials have yet to be released. Currently, an ongoing Phase I/II trial is examining the use of ADXS31-164 in HER2 expressing solid tumors. This trial is expected to be completed in December of 2018.

5.4. CRS-100 (ANZ-100)

While most antitumor effects of Lm vaccines are attributed to Lm secretion of TAAs leading to TAA-specific T-cell responses, studies utilizing CRS-100 highlight the powerful effects that Lm vaccines can have on the innate immune system. CRS-100 consists of the live, attenuated $\Delta actA/\Delta inlB$ strain of Lm and does not express exogenous antigens. Despite this, the CRS-100 vaccination has been shown to mediate antitumor effects, primarily through inducing antigen-independent NK cell activity. In preclinical modes of mice with hepatic colorectal cancer metastases, the CRS-100 vaccination induced recruitment of NK cells to the liver, leading to antitumor responses [87]. The therapeutic effects of the vaccine were eliminated by depletion of NK cells. Interestingly, while initial antitumor immunity was mediated by NK cells, challenge experiments in surviving vaccinated mice found that tumors were rejected through memory CD8⁺ T-cell responses, suggesting that primary destruction of the tumor by NK cells can trigger adaptive immune responses [87]. A Phase I dose escalation study of CRS-100 in patients with hepatic metastases showed similar effects on NK cells. A single injection of CRS-100 upregulated expression of the NK cell maturation marker CD38 and stimulated the production of Th1 cytokines in a dose-dependent manner [69]. Moreover, this trial was the first to demonstrate the safety and tolerability of the LADD[®] strain in a clinical setting. It appears that the development of CRS-100 has been halted in favor of constructs that incorporate TAAs which may directly stimulate antigen-specific CD8⁺ T-cell responses [17].

5.5. CRS-207

CRS-207 is an Lm vaccine expressing the tumor-associated antigen mesothelin as a fusion protein with a modified form of ActA [69]. Mesothelin is a glycoprotein that is normally confined to serosal cells lining the pleura, peritoneum, and pericardium [88]. Overexpression of mesothelin has been observed in pancreatic, ovarian, lung, and multiple other cancers, making it an attractive target for immunotherapy [88]. Clinical trials testing CRS-207 have primarily been performed in patients with metastatic pancreatic ductal adenocarcinoma (PDAC). After Phase I testing demonstrated the safety of CRS-207 in patients with pancreatic, mesothelioma, lung, and ovarian cancers (ClinicalTrials.gov identifier NCT00585845), multiple Phase II trials in PDAC were initiated using a prime-boost strategy. In these trials, CRS-207 is given as a boost after initial priming with GVAX and cyclophosphamide (Cy/GVAX) therapy. GVAX is a cancer vaccine composed of irradiated pancreatic cancer cells engineered to secrete GM-CSF [89]. Cyclophosphamide is typically given prior to GVAX based on studies demonstrating inhibitory effects on regulatory T cells and an increased survival benefit compared to GVAX alone [89]. The use of CRS-207 in combination with Cy/GVAX has been supported by at least two observations. First, early Phase I testing of CRS-207 found an increased survival benefit in a small cohort of PDAC patients who received prior GVAX therapy compared to those who did not [69]. Moreover, preclinical studies in mouse PDAC models utilizing GVAX prime with CRS-207 boost demonstrated synergistic induction of mesothelin-specific T cells and inhibition of tumor growth [71]. Early Phase II testing in patients with metastatic PDAC demonstrated increased median OS in patients receiving GVAX with CRS-207 compared to Cy/GVAX alone (6.1 vs. 3.9 months) [71]. The induction of mesothelin-specific T cells was found to correlate with increased overall survival [71]. Interestingly, there was no observed difference in the number of circulating mesothelin-specific CD8⁺ T cells between Cy/GVAX and Cy/GVAX + CRS-207. Based on the results from this study, a Phase IIb study (ECLIPSE) was initiated which compared CRS-207 vs. Cy/GVAX with CRS-207 vs. conventional chemotherapy. Data presented at a conference hosted by the American Society for Clinical Oncology indicated no survival benefit from either CRS-207 alone (5.4 months), or the combination of CRS-207 with Cy/GVAX (3.8 months), compared to standard-of-care chemotherapy (4.6 months) [70]. Additional trials exploring CRS-207 and Cy/GVAX in combination with α PD-1 and α CTLA-4 immune checkpoint inhibitors as well as with indoleamine dioxygenase (IDO) inhibitors are underway.

These are complemented by other trials using CRS-207 in patients with mesothelioma, gynecologic, and gastroesophageal cancers, and data in mesothelioma patients have been published. These data showed that, after two infusions of CRS-207 in combination with six cycles of chemotherapy, 59% of patients exhibited a partial response, and an additional 35% experienced stable disease [72]. Further testing demonstrated an increase in tumor-infiltrating lymphocytes post-vaccination [72]. While numerous trials with CRS-207 are currently ongoing, Aduro has discontinued further clinical advancement of CRS-207.

5.6. ADU-623

ADU-623 is currently the only Lm vaccine being developed for the treatment of brain cancers. ADU-623 is a bivalent vaccine that expresses EGFRvIII and NY-ESO-1 antigens [73]. EGFRvIII is the most commonly occurring EGFR (epidermal growth factor receptor) mutation, producing a neoantigen formed from the spontaneous deletion of exons 2–7 which encode sequences in the extracellular domain [90]. Oncogenic EGFRvIII signaling may directly promote tumor growth and is frequently present in glioblastoma, lung, and other cancers [90]. NY-ESO is a cancer-testis antigen, the expression of which is restricted to germ cells in healthy adults. However, aberrant expression of NY-ESO-1 has been observed in neuroblastoma, esophageal, and other cancers [91]. Currently, a Phase I trial assessing ADU-623 in patients with high-grade astrocytomas is ongoing (ClinicalTrials.gov identifier NCT01967758). The trial is expected to be completed in December 2018.

5.7. JNJ-64041757 and JNJ-64041809

In 2014, Aduro licensed the Lm vaccines JNJ-64041757 and JNJ-64041809 to Janssen Biotech. JNJ-64041757 (previously known as ADU-214) is a bivalent Lm vaccine expressing EGFRvIII and mesothelin antigens [4]. Early reports from a Phase I trial in non-small cell lung cancer patients suggest that JNJ-64041757 exhibits a safety profile similar to CRS-207 and is capable of generating mesothelin-specific T-cell responses [74]. Based on these results, a subsequent Phase I/II trial assessing JNJ-64041757 alone or in combination with the α PD-1 inhibitor nivolumab was launched (ClinicalTrials.gov identifier NCT03371381). The trial is expected to be completed in March 2020. JNJ-64041809 (previously known as ADU-741) contains Lm expressing multiple prostate cancer-associated antigens [4]. Currently, it is undergoing Phase I clinical testing in patients with metastatic castration-resistant prostate cancer. A Phase II trial testing JNJ-64041809 in combination with the nonsteroidal antiandrogen apalutamide was initiated, but it was withdrawn prior to enrollment (ClinicalTrials.gov identifier NCT02906605).

5.8. pLADD and ADXS-NEO

During tumorigenesis, cancer cells undergo genetic alterations which can lead to the cumulative production of mutated protein sequences not found in healthy tissues [92]. These mutated proteins, termed neoantigens, may undergo proteasomal degradation and MHC loading, forming unique epitopes presented only by the tumor cells harboring the underlying mutations. In theory, vaccines targeting neoantigens may generate stronger T-cell responses due to the absence of tolerance mechanisms as well as fewer off-target effects [92]. Based on this rationale, two Lm vaccines targeting personalized neoantigens, ADXS-NEO and pLADD, were recently approved for Phase I testing. Proof of concept for ADXS-NEO was demonstrated using CT26 and MC38 colorectal cancer cell lines, and this was presented at the American Association for Cancer Research annual meeting [93,94]. The development of personalized vaccines involved whole-exome sequencing to identify tumor-specific mutations followed by analysis with predictive algorithms to identify potential neoepitopes. After screening to verify immunogenicity, the neoantigens were cloned into Lm and found to induce tumor regression upon immunization [93,94]. A Phase I trial testing ADXS-NEO in patients with metastatic head and neck, colon, and lung cancer is underway (ClinicalTrials.gov identifier NCT03265080). The personalized Lm vaccine pLADD is similarly undergoing Phase I testing in patients with microsatellite stable metastatic colorectal cancer

(ClinicalTrials.gov identifier NCT03189030). In addition to personalized vaccines, Advaxis is currently developing ADXS-HOT. Because sequencing for neoantigens is a time-consuming and expensive process, ADXS-HOT is designed to be an “off-the-shelf” therapy containing a collection of frequently-mutated somatic, cancer testis, and oncofetal antigens [95]. However, the clinical details concerning this vaccine have not been released.

5.9. Adverse Effects of *Listeria* Vaccine Therapy

In general, the most common side effects of Lm vaccines are grade 1 or 2 adverse events, including transient fever, chills, nausea, vomiting, and hypotension [66,67,69,71]. Increases in circulating liver enzymes, as well as lymphopenia, have also been reported upon administration of vaccines [65,69,71]. In at least two cases, systemic listeriosis was reported after administration but was adequately treated with antibiotics [42,96]. Overall, multiple clinical trials have demonstrated an excellent safety profile with attenuated Lm vaccines in humans.

6. Conclusions and Future Directions

While preclinical models have shown impressive therapeutic benefits of Lm vaccines, currently, no Lm vaccines are FDA-approved. It is important to note that Lm vaccines are still relatively nascent, with most clinical trials only being initiated in recent years. As these trials advance, further improvements to Lm vaccine technology will likely be made in the coming years. Indeed, after initial clinical trials used Lm expressing a single tumor antigen, Lm vaccine technology has evolved to include vaccines expressing multiple tumor antigens. By targeting multiple antigens, it may be possible to induce therapeutic responses that are less susceptible to antigen escape. Yet, even vaccination against multiple tumor antigens may still not be sufficient to induce optimal therapeutic responses. Tumors consist of many cell types and the tumor microenvironment has been hypothesized to be a protective niche that may limit antitumor responses. Thus, many strategies moving forward are likely to combine Lm vaccines with approaches that remodel the tumor microenvironment. One such strategy has been to directly target angiogenesis using Lm vaccines. Indeed, Lm vaccines targeting tumor-associated angiogenic proteins, including CD105 and VEGFR2, have demonstrated inhibited tumor growth [97–100] as well as the ability to induce secondary antitumor immune responses via epitope spreading [98,99]. While exciting, these vaccines have not yet been tested clinically. Other studies are likely to explore combination approaches using Lm vaccines with radiation, chemotherapy, immune checkpoint inhibitors, and other drugs. These studies are expected to firmly establish the utility of Lm vaccines in the emerging immunotherapeutic landscape for the benefit of patients with a variety of malignancies.

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