

Award Number: W81XWH-14-1-0141

TITLE:

Testing the Role of p21-Activated Kinases in Schwannoma Formation Using a Novel Genetically Engineered Murine Model that Closely Phenocopies Human NF2 Disease

PRINCIPAL INVESTIGATOR: Jonathan Chernoff, M.D., Ph.D.

CONTRACTING ORGANIZATION:

Institute for Cancer Research
Philadelphia, PA 19111

REPORT DATE: June 2016

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PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

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7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Institute for Cancer Research 333 Cottman Avenue Philadelphia, Pennsylvania 19111 E-Mail: osr@fccc.edu						8. PERFORMING ORGANIZATION REPORT NUMBER			
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14. ABSTRACT <p>The major goal of this research project was to genetically and pharmacologically test the requirement of Group A PAK signaling in Nf2 deficient schwannoma genesis. We planned to accomplish this goal by (1) using our genetically intercrossed Pak1, Pak2 and a dominant negative Group A PAK transgene with our <i>PostnCre;Nf2^{fllox/fllox}</i> mouse schwannoma model in order to generate functional (ABR testing) and histological readouts. In turn these readouts will allow us to utilize a genetic approach to determine if indeed PAK signaling is essential to the development of <i>Nf2</i>-deficient schwannomas; (2) using histological and ABR readouts, we will treat <i>PostnCre;Nf2^{fllox/fllox}</i> mice with three different pharmacologic PAK inhibitors to determine if targeted PAK inhibition in a preclinical model of schwannoma genesis rescues tumor development.</p> <p>We are on track to accomplish the goals we set. In the first year of the funded work, we completed crosses of <i>Pak1^{-/-}</i> and <i>Pak2^{fllox/fllox}</i> mice with <i>PostnCre;Nf2^{fllox/fllox}</i> mice and analyzed the effects of Pak loss on hearing and tumor growth. As we had speculated, loss of Pak1 function partly prevented hearing loss and tumor growth. Studies on Pak2 are ongoing. We also began Aim 2, treating <i>PostnCre;Nf2^{fllox/fllox}</i> mice with Pak small molecule inhibitors.</p>									
15. SUBJECT TERMS NF2, Pak1, Pak2, PID, ABR, Pak inhibitors, mouse models, signal transduction, preclinical studies									
16. SECURITY CLASSIFICATION OF:						17. LIMITATION OF ABSTRACT UU	18. NUMBER OF PAGES 18	19a. NAME OF RESPONSIBLE PERSON USAMRMC	
a. REPORT U		b. ABSTRACT U		c. THIS PAGE U				19b. TELEPHONE NUMBER <i>Include area code</i>	

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INTRODUCTION

Neurofibromatosis type 2 (NF2) is an autosomal dominant genetic disease characterized by benign schwannomas that grow on the cranial and spinal nerves. While technically benign, the tumors are nonetheless progressive and relentless, usually resulting in death before age fifty from inoperable intracranial masses. To date, surgery remains the only effective therapy for these lesions, though this therapy is frequently associated with major morbidities, including loss of hearing. In this award we proposed to evaluate Group A Paks (Pak1 and Pak2) as a therapeutic target in NF2 by intercrossing Group A PAK deficient mice with our NF2 mouse model (*PostnCre;Nf2^{flox/flox}*). Briefly, we hypothesized that the knockdown of Group A Paks in *Nf2* deficient mice will rescue or reduce tumorigenesis. In **Specific Aim 1**, we proposed to characterize Pak's signaling influence on NF2 *in vivo* by assessing hearing thresholds, survival rate, tumor formation and kinome analysis. In **Specific Aim 2**, we proposed to test Pak inhibitors in our NF2 tumor model in order to evaluate whether the inhibition of Pak rescues merlin function to wild-type levels by analyzing readouts generated from tumor growth and auditory brainstem response (ABR).

KEYWORDS: NF2, Pak1, Pak2, PID, ABR, Pak inhibitors

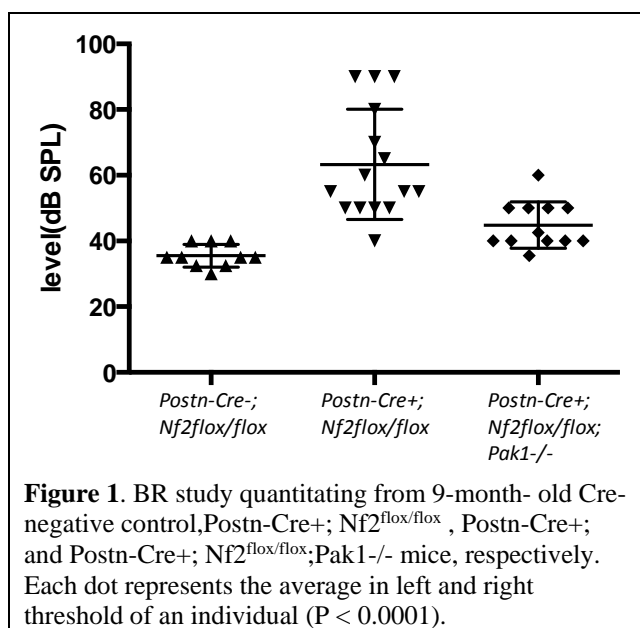
ACCOMPLISHMENTS

(A) Major Goals of the Project

The major goal of this research project was to genetically and pharmacologically test the requirement of Group A PAK signaling in *Nf2* deficient schwannoma genesis. We would accomplish this goal by (1) using our genetically intercrossed Pak1, Pak2 and a dominant negative Group A PAK transgene with our *PostnCre;Nf2^{flox/flox}* mouse schwannoma model in order to generate functional (ABR testing) and histological readouts. In turn these readouts will allow us to utilize a genetic approach to determine if indeed PAK signaling is essential to the development of *Nf2*-deficient schwannomas; (2) using histological and ABR readouts, we will treat *PostnCre;Nf2^{flox/flox}* mice with three different pharmacologic PAK inhibitors to determine if targeted PAK inhibition in a preclinical model of schwannoma genesis rescues tumor development.

(B) Accomplishments under these Goals

In **Specific Aim 1a.1**, we proposed to characterize the role of individual Pak isoforms (Pak1 and Pak2) by intercrossing these mice with our NF2 mouse model, *Postn-Cre; Nf2^{flox/flox}* to generate the following cohorts of mice; 15 *Postn-Cre-; Nf2^{flox/flox}; Pak1^{+/+}* mice (Control), 15 *Postn-Cre+; Nf2^{flox/flox}; Pak1^{+/+}* mice (NF2-KO), 15 *Postn-Cre-; Nf2^{flox/flox}; Pak1^{-/-}* mice (PAK1-KO), and 15 *Postn-Cre+; Nf2^{flox/flox}; Pak1^{-/-}* (NF2/PAK1-DKO). These cohorts of mice were utilized in order to assess hearing loss via auditory brainstem response (ABR) testing. Preliminary data included in the initial grant proposal indicated that our *Postn-Cre+; Nf2^{flox/flox}* mice showed by eight months of age, complete hearing loss, as compared to control mice, *Postn-Cre-; Nf2^{flox/flox}*. Functionally, the mean hearing threshold of nearly 60 dB in *Postn-Cre+; Nf2^{flox/flox}* mice at the age of 10 months is equivalent, as a human analog, of the inability to hear in a normal conversation, thus indicating severe disruption of an affected individual's life. In the analysis of 9 to 10 month old *Postn-Cre+; Nf2^{flox/flox}; Pak1^{-/-}* mice we observed that these mice showed improved hearing as compared 9 to 10 month old *Nf2* deficient mice (*Postn-Cre+; Nf2^{flox/flox}* mice), as shown in **Figure 1**. Control mice (*Postn-Cre-; Nf2^{flox/flox}; Pak1^{+/+}*) showed no hearing loss at 9 to 10 months of age. In the next year, we will repeat ABR testing on 12 and 14 month old Pak1 deficient cohorts listed above and in parallel begin ABR testing on Pak2 deficient cohorts of mice intercrossed with NF2 mouse model, *Postn-Cre; Nf2^{flox/flox}* to generate the

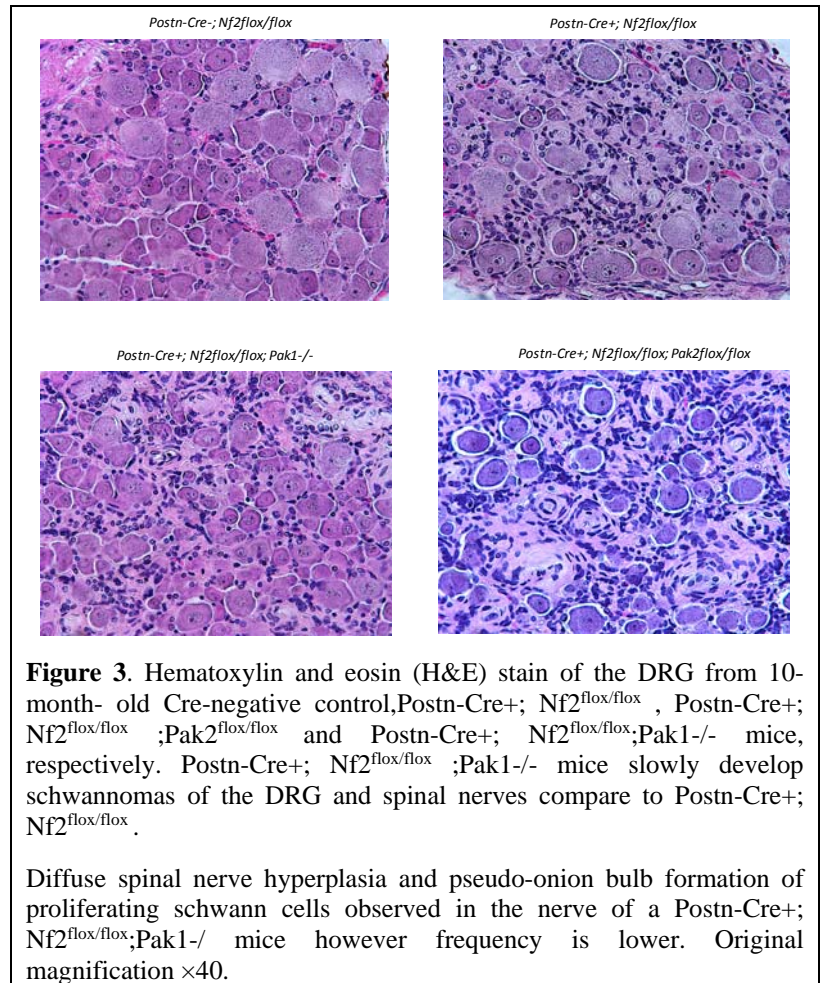
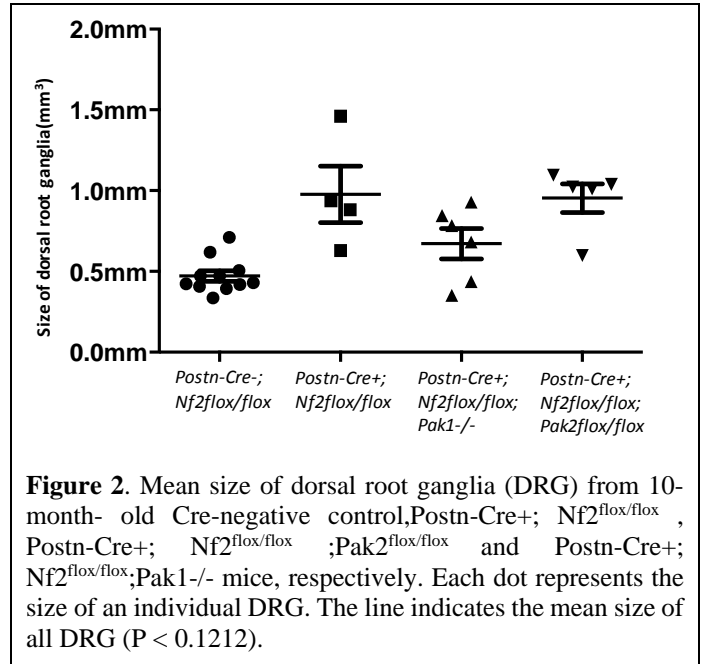


Program Director/Principal Investigator (Last, first, middle): Chernoff, Jonathan following cohorts of mice; 15 *Postn-Cre-; Nf2^{flox/flox}; Pak2^{flox/flox}* mice (Control), 15 *Postn-Cre+; Nf2^{flox/flox}* mice (NF2-KO), 15 *Postn-Cre-; Nf2^{flox/flox}; Pak2^{flox/flox}* mice (PAK2-KO), and 15 *Postn-Cre+; Nf2^{flox/flox}; Pak2^{flox/flox}* (NF2/PAK2-DKO).

In **Specific Aim 1a.2**, we proposed to characterize the role of individual Pak isoforms (Pak1 and Pak2) in the NF2 mouse model by assessing tumor formation. Our preliminary data indicate that genetic disruption of Pak 1 or Pak 2 is not completely sufficient to inhibit tumor initiation (**Figures 2 and 3**). However, there is a strong but not statistically significant suggestion that genetic disruption of Pak1 is resulting in a reduction in dorsal root ganglia size as illustrated in **Figure 2**. Though we need to evaluate additional specimens, the histology of the dorsal root ganglia (DRG) of mice containing a disruption of the Pak1 allele appears more benign than the *Postn-Cre+; Nf2^{flox/flox}; Pak2^{flox/flox}* and the *Postn-Cre+; Nf2^{flox/flox}*. However, more studies will be required to verify this observation. We will continue to analyze DRG volume post-ABR testing in 12 and 14 month old *Postn-Cre+; Nf2^{flox/flox}; Pak1^{-/-}*, *Postn-Cre-; Nf2^{flox/flox}; Pak1^{-/-}*, *Postn-Cre+; Nf2^{flox/flox}; Pak2^{flox/flox}*, *Postn-Cre-; Nf2^{flox/flox}; Pak2^{flox/flox}* and *Postn-Cre+; Nf2^{flox/flox}* in the next funding period.

In **Specific Aim 1a.3**, we proposed to characterize the role of individual Pak isoforms (Pak1 and Pak2) in our NF2 mouse model, *Postn-Cre; Nf2^{flox/flox}*, by assessing survival rate. Preliminary data included in the initial grant proposal indicated that our *Postn-Cre+; Nf2^{flox/flox}* mice showed by eight months of age, a decrease in survival as compared to control mice, *Postn-Cre-; Nf2^{flox/flox}*. In the analysis 15 mice per cohort (cohorts listed in Aim1a.1). Though the study is ongoing, the preliminary data may suggest a modest improvement in survival when Pak1 (**Table 1**). However, these studies are ongoing and we are currently still following these cohorts of mice. Therefore in year 3 funding we will continue to observe the, *Postn-Cre; Nf2^{flox/flox} Pak1^{-/-}*, and *Postn-Cre; Nf2^{flox/flox}; Pak2^{flox/flox}* mouse models.

In **Specific Aim 1B**, we planned to cross *Postn-Cre; Nf2^{flox/flox}* mice with ROSA26-LSL-PID mice and analyze tumor growth and hearing loss (Months 6-30). These experiments are ongoing.



	0	1	2	3	4	5	6	7	8	9	10	11	12 (month)		
WT N=15	100	100	100	100	100	100	100	100	100	100	100	100	100	Working on	
Nf2-Cre+ N=15	100	100	93	86	86	80	80	80	80	80	80	80	80	"	
Pak1^{-/-} N=15	100	100	100	100	100	100	100	100	100	100	100	100	100	"	
Nf2;Pak1 N=16	100	100	100	100	100	100	100	100	100	93	93	93	93	"	
Pak2cre+ N=	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
Nf2;Pak2 N=15	100	100	100	100	100	100	93	93	93	93	93	93	93	"	

Table 1. Kaplan-Meier survival data

In **Specific Aim 1C** we proposed to assess kinome activity in tumors from the following cohorts of mice: *Postn-Cre*; *Nf2^{flox/flox} Pak1^{-/-}*, and *Postn-Cre*; *Nf2^{flox/flox} Pak2^{flox/flox}*, and ROSA26-LSL-PID mouse models. Currently, we have preliminary data from the first of these crosses, *i.e.*, *Postn-Cre*; *Nf2^{flox/flox}* vs *Postn-Cre*; *Nf2^{flox/flox}*; *Pak1^{-/-}* mice. Tumors were collected post ABR testing (performed in Aim1a.1) starting at 10 months of age, and at 12 months and at 14 months.

We pooled protein lysates derived from three schwannomas from *Postn-Cre*; *Nf2^{flox/flox}*; *Pak1^{+/+}* and three *Postn-Cre*; *Nf2^{flox/flox}*; *Pak1^{-/-}* mice, respectively, and subjected these lysates to MIBs analysis. The results of this preliminary screen are shown in **Figure 4**. We found elevated kinase activities for MERTK (a receptor tyrosine kinase of the AXL family), GRK5 (a G Protein-Coupled Receptor Kinase), PLK2 (polo-like kinase), etc. We are currently validating these findings using phospho-specific antibodies.

In **Specific Aim 2** we proposed to test small molecule Pak inhibitors on our *Postn-Cre*; *Nf2^{flox/flox}* mouse model. During this reporting period, we completed these studies using FRAX-1036, a specific inhibitor of group A Paks. The compound was well tolerated in our murine Schwannoma model at the 30 mg/kg/d dose and demonstrated robust inhibition of Pak phosphorylation *in vivo* (**Figure 5**).

Despite robust inhibition of Pak phosphorylation, tumor size was not significantly decreased in FRAX-1036 treated mice. ABR analysis revealed a modest decrease in hearing threshold after 12 weeks treatment versus vehicle control treated mice, but the decrease was not statistically significant (**Figure 6**).

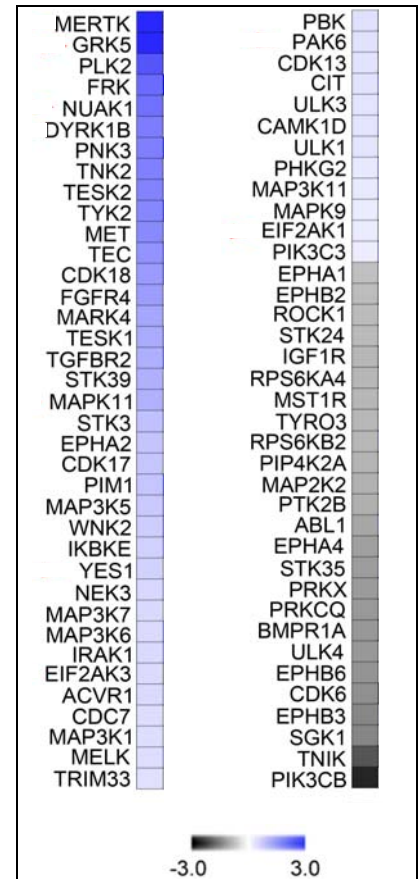


Figure 4. Preliminary MIBs analysis of tumors from *Postn-Cre*+; *Nf2^{flox/flox}* vs. *Postn-Cre*+; *Nf2^{flox/flox}*; *Pak1^{-/-}* mice. Ratios of kinase activity (*Pak1^{-/-}*/WT) are shown, with blue representing elevated activity, black representing diminished activity.

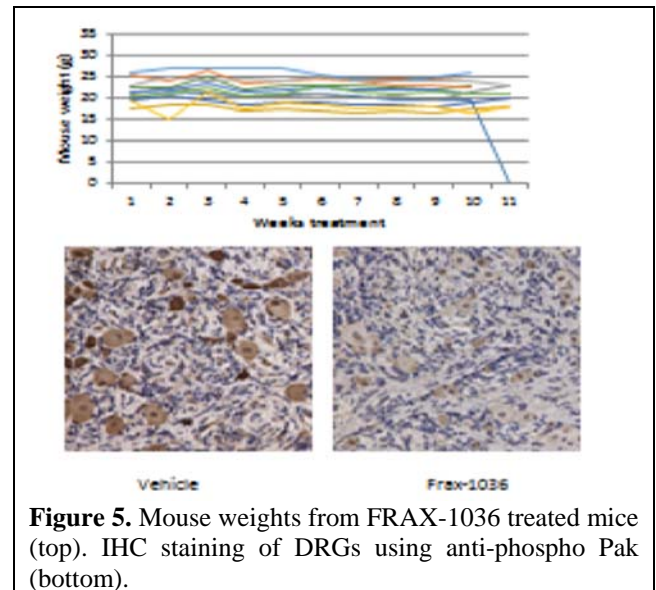


Figure 5. Mouse weights from FRAX-1036 treated mice (top). IHC staining of DRGs using anti-phospho Pak (bottom).

(C) Training and Professional Development Opportunities

Nothing to Report

(D) Disseminated Results to Communities of Interest

Nothing to Report

(E) Plan for Next Reporting period

In order to accomplish the goals and objectives stated above, in the next funding cycle we will complete ABR testing on PID cohorts, and complete kinome readouts and confirmatory data using phospho-specific antibodies.

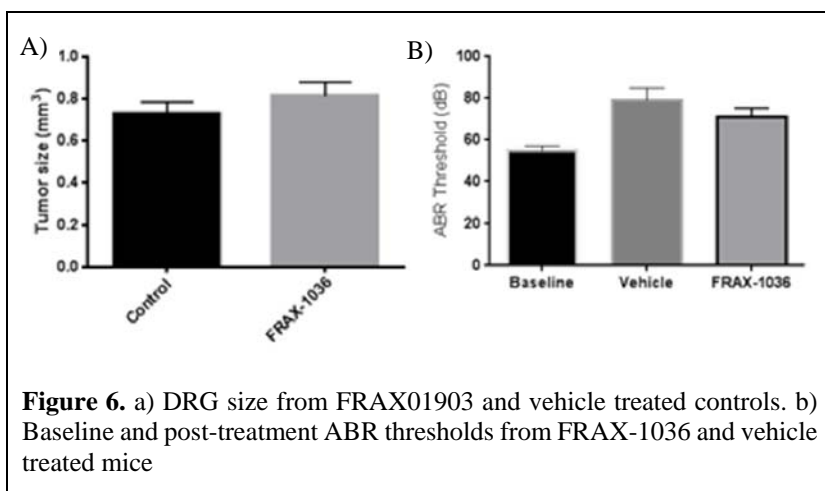


Figure 6. a) DRG size from FRAX01903 and vehicle treated controls. b) Baseline and post-treatment ABR thresholds from FRAX-1036 and vehicle treated mice

IMPACT

Nothing to Report

CHANGES/PROBLEMS

Nothing to Report

PRODUCTS

Nothing to Report

PARTICIPANTS & OTHER COLLABORATING ORGANIZATIONS:

What individuals have worked on the project?

Please see the attached table.

Has there been a change in the active other support of the PD/PI(s) or senior/key personnel since the last reporting period?

Please see Other Support attached. Changes from the last reporting period are marked with a line in the right hand margin.

What other organizations were involved as partners?

Organization Name: Indiana University

Location of Organization: Indianapolis, IN

Partner's contribution to the project: Subaward

Indiana University has a subaward under this grant. Resources include: Facilities; Collaboration; Personnel exchanges

SPECIAL REPORT REQUIREMENTS:

Not Applicable

APPENDICES:

Not Applicable

Fox Chase Cancer Center

Name:	<i>J. Chernoff, MD, PhD</i>
Project Role:	Principal Investigator
eBRAP ID:	<i>chernoff</i>
Nearest person month worked:	<i>1</i>
Contribution to Project:	<i>Overall administration and guidance of research; Management and training of personnel</i>
Funding Support:	<i>N/A</i>
Name:	<i>H.-Y. Chow, PhD</i>
Project Role:	Research Scientist
eBRAP ID:	
Nearest person month worked:	<i>6</i>
Contribution to Project:	<i>Kinome screening using MIBs; Examining tumor tissues by IHC and immunoblot</i>
Funding Support:	<i>N/A</i>

Indiana University

Name:	<i>Wade Clapp, M.D.</i>
Project Role:	Subcontract PI
eBRAP ID:	<i>dclapp</i>
Nearest person month worked:	<i>1</i>
Contribution to Project:	<i>Regulatory activity involved in the genetic intercrosses, laboratory infrastructure, and strategic planning of murine models to validate the Group PAKs</i>
Funding Support:	<i>N/A</i>
Name:	<i>Su-Jung Park, Ph.D</i>
Project Role:	Associate Professor
eBRAP ID:	
Nearest person month worked:	<i>1</i>
Contribution to Project:	<i>Generation and Maintenance of murine models; Genotyping.</i>
Funding Support:	<i>N/A</i>
Name:	<i>Li Jiang, BME</i>
Project Role:	Research Technician
eBRAP ID:	
Nearest person month worked:	<i>3</i>
Contribution to Project:	<i>Maintains the genetically modified murine models; Conducts PCR; Assists in mice dissection and ABR testing.</i>
Funding Support:	<i>N/A</i>

OTHER SUPPORT

Chernoff, Jonathan

Remaining Salary Support from Institutional resources.

ACTIVE

R01 NS066927 (PI: Li, Vanderbilt Univ.)	3/1/2015 - 2/29/2020	10.0%
NIH	\$29,750	1.20 calendar

Pathophysiology of Conduction Block in HNPP

This project is a subcontract to Vanderbilt University.

Procuring Contracting/Grants Officer: Roddy Smith, Neurology Dept, 1161 21st Ave. South, Nashville, TN 37203, 615-936-8950

R01 CA148805 (PI: Chernoff / Testa)	4/22/2016 - 3/31/2021	10.0%
NIH	\$237,500	1.20 calendar

Role of STE20 Protein Kinases in Malignant Mesothelioma

The major goals of this project are to: 1) Define the response of MM mice to Pak small molecule inhibitors and identify how tumors adapt to such inhibitors in vivo; and 2) Delineate the role of the Hippo pathway in Merlin-related signaling and MM pathology in vivo.

Procuring Contracting/Grants Officer: Candace Cofie, 9609 Medical Center Dr., Bethesda, MD 20892, 240-276-6317

T32 CA009035 (PI: Chernoff)	9/16/2011 - 8/31/2016	NA
NIH	\$523,729	calendar

Training Program in Cancer Research

The overall goal of this program is to prepare postdoctoral scientists for independent careers in basic and/or translational research focused on cancer.

Procuring Contracting/Grants Officer: Bryan Benton, BG 9609 RM2W514, 9609 Medical Center Drive, Rockville MD 20852, (240-276-5863

IRG-15-175-21 (PI: Chernoff)	1/1/2016 - 12/31/2018	NA
ACS	\$90,000	calendar

Institutional Research Grant No Salary

This is an Institutional Grant from the American Cancer Society that provides start-up funds for promising junior investigators to initiate innovative laboratory, clinical and translational research.

Procuring Contracting/Grants Officer: Virginia Krawiec, Extramural Grants Dept., 350 Williams St., Atlanta, GA 30303, 404-329-5734

P30 CA006927 (PI: Fisher)	7/21/2011 - 6/30/2016	15.0%
NIH	Partial Salary	1.80 calendar

Comprehensive Cancer Center Program at Fox Chase

The major goal of this Cancer Center Support Grant is to provide partial salary support for professional personnel, including senior and program leadership, administration, planning and evaluation, and developmental funds, as well as support for 4 established peer-reviewed Research Programs, 12 Shared Research Resources and 1 Support Element.

Procuring Contracting/Grants Officer: Emily Tran, OGA 9609 Medical Center Dr., West Tower, 2nd Flr., Rockville, MD 20850, 240-276-6324

Program Director/Principal Investigator (Last, first, middle): Chernoff, Jonathan

W81XWH-14-1-0141 (PI: Chernoff) 5/15/2014 - 5/14/2017 10.0%
DOD \$190,195 1.20 calendar
Testing the Role of p21 Activated Kinases in Schwannoma Formation Using a Novel Genetically Engineered Murine Model that Closely Phenocopies Human NF2 Disease
The major goals of this project are: 1) Does Pak signaling influence NF2-related pathology in vivo?; and 2) Are small molecule inhibitors of Pak effective in preclinical models of NF2?
Procuring Contracting/Grants Officer: Jodi Cardoza, US AMRAA, 820 Chandler St., Fort Detrick, MD 21702 (301) 619-2693

R01 CA142928 (PI: Chernoff) 1/1/2015 - 12/31/2019 20.0%
NIH \$237,500 2.40 calendar
Targeting the Kinome in Neurofibromatosis Type 1
The major goals of this project are to determine: 1) What is the status of the kinome in NF1-/- Schwann cells and mast cells, and how does this change upon Mek, Akt/mTOR, or Pak inhibition?; 2) What is the cellular basis for Pak's function in NF1-related tumors? and 3) Are small molecule Pak inhibitors effective in preclinical models of NF1? Can whole kinome analysis predict pathways for drug resistance in treated mice?
Procuring Contracting/Grants Officer: Candace Cofie, 9609 Medical Center Dr., Bethesda, MD 20892, 240-276-6317

(PI: Chernoff) 2/1/2016 - 1/31/2017 5.0%
BCA \$92,593 0.60 calendar
Dissecting a Breast Cancer Amplicon using CRISPR/Cas
The major goal of this project is to use a single pool of CRISPR-Cas constructs targeting all 198 genes in the 11q13.5-14.1 cluster between (and including) CCND1 and GAB2, and determine the relative contribution of each gene to the proliferation and survival of an IntClust-2 breast cancer cell line versus a non Intclust-2 breast cancer cell line.
Procuring Contracting/Grants Officer: Sharon Phillips, 48 Maple Ave., Greenwich, CT 06830, 203-861-0014

W81XWH-15-1-0192 (PI: Bumber, UNMCC) 9/15/2015 - 9/14/2017 2.0%
DOD No Salary 0.24 calendar
Msi2 Regulates the Aggressiveness of Non-Small Cell Lung Cancer (NSCLC)
This project is a subcontract to University of New Mexico Cancer Center (UNMCC). The major goals of this project are: 1) To evaluate the functional role of Msi2 in human lung cancer cell lines; and 2) To establish how Msi2 expression predicts tumor phenotypes, patient outcomes, and invasion related signaling in primary human NSCLC tumors. Dr. Chernoff serves as a Co-Mentor to the prime PI at UNMCC.
Procuring Contracting/Grants Officer: Rena Vinyardk, SPO, MSC09 5220 1 University of New Mexico, Albuquerque, NM 87131, 505-272-0159

OVERLAP

None

COMPLETED

None

Clapp, D. Wade

ACTIVE

K12 HD068371 (PI: Clapp)	03/01/13-11/30/17	5%
NIH/NICHD	\$400,000	0.60 Calendar Months

Indiana Pediatric Scientist Award (IPSA)

Goals and Aim: Leveraging \$400,000 n existing strong foundation of training, mentorship and research, the Indiana University School of Medicine (IUSM) Indiana Pediatric Scientist Award (IPSA) program will accelerate the success of junior faculty in bridging the gap between training and independence. The program will provide our junior faculty with a blueprint for success, structured mentorship and guidance and progressive goals in order to achieve independent funding within a five year period.

Role: Principle Investigator

Program Officer: Karen Winer, 6100 Executive Blvd., Room 4B11, MSC 7510, Bethesda MD, 20892.

W81XWH-12-1-0155 (PI: Korf, UAB)	05/15/13-05/15/17	1%
DOD, University of Alabama, Birmingham	\$152,024	0.12 Calendar Months

NFRP Neurofibromatosis Clinical Trials Consortium

Goals: Implement the clinical proposal; collect and transmit the data at Riley Hospital for Children at Indiana University Health. Participate in results preparation of the NF Consortium Infrastructure for presentation and publication.

Specific Aims:

Primary Aim: To estimate the objective response rate to Cabozantinib at 12 months in adults with NF1 plexiform neurofibromas by volumetric MRI imaging.

Secondary Aims:

1. To further assess the tolerability and toxicity of Cabozantinib in patients with NF1
2. To estimate the objective response rate of up to 2 non-target plexiform neurofibromas to Cabozantinib by MRI
3. To determine the quality of life (QOL) response to Cabozantinib in patients with NF1 plexiform neurofibromas
4. To access activity of Cabozantinib on mast cell activity by mast cell culture and FACS
5. To describe changes by flow cytometry in peripheral blood monocyte counts, circulating endothelial cells, and plasma angiogenic factors during treatment with Cabozantinib
6. To describe the baseline and change in 17 circulating cytokine factors related to proliferating cells
7. To characterize the pharmacokinetic profile of Cabozantinib in this population

Role: Site PI (Overall PI: Bruce Korf)

Program Officer: Naba Bora, 1077 Patchel St. Fort Detrick, Frederick, MD, 21702-9218

(PI: Clapp)	05/01/14-04/30/17	10%
Children's Tumor Foundation	\$ 338,730	1.2 Calendar Months

Experimental Therapeutics targeting the NF2 kinome: an integrated rational approach

Goals: The goal of this application is to bring a multidisciplinary group of investigators together in a vertically integrated program that allows consistency of screening approaches, utilizes both established and newly developed cellular and murine models of NF2-associated tumors, allows validation of the mechanisms of action of drugs and investigation of the biologic result of target inhibition in both VS and meningiomas as well as exploration of the cellular compensatory events influencing drug response with the ultimate goal of developing clinical studies that have the highest chance of success for patients.

Specific Aims:

Aim 1: Utilize NF2/(Nf2)-deficient schwannoma and meningioma cells to screen:

Program Director/Principal Investigator (Last, first, middle): Chernoff, Jonathan

- a) Established FDA-approved drugs and targeted therapeutics in the clinical pipeline that have rational linkage to Nf2-dependent pathways in collaboration with industry partners
- b) A library of late-stage and FDA-approved drugs in collaboration with the National Center for Advancement of Translational Science (NCATS) to allow an unbiased drug screen

Aim 2: Utilize genetically engineered (VS) and xenograft (meningioma) models to test drugs that meet in vitro go/no go criteria:

- a) Vestibular schwannomas: PeriostinCre; Nf2^{flox/flox} mice will be treated with drugs successful in the in vitro screen against schwannomas. The preclinical endpoints will be restoration of baseline hearing in 50% of mice and 50% mean reduction in paraspinal tumor volume by MRI and at autopsy in 75% of mice compared to mice treated with a vehicle control.
- b) Meningiomas: Ben-Men-1 cells (benign NF2-deficient meningioma cells) will be implanted intracranially and mice will be treated with drugs successful in the in vitro screen against meningiomas. The preclinical endpoint will be 50% mean reduction in tumor volume by MRI and luciferase activity in 75% of mice compared to mice treated with a vehicle control.

Aim 3: Identify at least 1 drug to move forward for a clinical trial.

- a) Drs. Blakeley and Welling will be involved in all stages of go/no-go decision making and will design the appropriate clinical trial for the agent(s) with the best portfolio based on:
 - a) Acceptable safety profile
 - b) Efficacy in vitro and in vivo against cell culture models of schwannoma and meningioma - Preference will be for joint activity against VS and meningioma and for drugs with delineated mechanisms in NF2 tumors
 - c) Drugs that are in active clinical development with an industry partner

Aim 4: Delineate cell and molecular validation of the best performing experimental compounds

- a) Identify cellular mechanisms of action of drugs in tumors in vitro and in vivo
- b) Utilize immunohistochemistry and western blot to validate inhibition of the biochemical target in tumorigenic cells of the in vivo models
- c) Conduct kinome analyses on drugs that pass the established go/no go criteria for at least one in vivo model to evaluate patterns of adaptation to therapy and the differences between meningioma and VS therapeutic responses
- d) Confirm synthetic lethality via genetic confirmation with shRNA

Role: Multi-PI (David W. Clapp, Jaishri Blakeley, Scott Plotkin and James Guesella)

Program officer: Annette Bakker, 95 Pine Street, 16th Floor, New York, NY, 10005

W81XWH-14-1-0141 (PI: Chernoff, FCCC)

05/14/14-05/13/17

10%

DOD/NFRP

\$116,797

1.2 Calendar Months

Testing the Pharmacologic and genetic role of P21 activated kinase inhibition on Schwannoma Formation using a Novel Genetically Engineered Murine Mouse Model that Closely Phenocopies Human NF2 Disease

Goals: The objective of this project is to evaluate Pak as a therapeutic target in NF2.

Specific Aims:

Aim 1: Does Pak signaling influence NF2-related pathology *in vivo*? If group A Paks are important for growth and motility signaling in cells lacking NF2, then loss of Pak function should slow or prevent pathologies associated with loss of the *Nf2* gene in mice. We have already constructed *Pak1*^{-/-} and *Pak2*^{flox/flox} mice, as well a targeted transgenic knock-in mouse that conditionally expresses the PID, a peptide inhibitor of all group A Paks, in any tissue expressing Cre recombinase. We will cross these transgenic mice, as well as our existing *Pak1* knock-out mice, with Periostin (*Postn*)-Cre; *Nf2*^{flox/flox} mice to answer the following questions

- a) Does loss of the *Pak1* or *Pak2* gene alone ameliorate loss of the *Nf2* gene in schwannoma formation? What signaling pathways mediate these effects?

Program Director/Principal Investigator (Last, first, middle): Chernoff, Jonathan

b) Does suppression of group A Pak kinase function by a genetically encoded peptide inhibitor ameliorate loss of the *Nf2* gene *in vivo*? What signaling pathways mediate these effects?

c) What is the basal state of the kinome in NF2 schwannomas and how is the kinome in Schwann cells or tumors reprogrammed upon loss of Pak function?

Aim 2: Are small molecule inhibitors of Pak effective in preclinical models of NF2? Selective small molecule Pak inhibitors have recently entered clinical trials. In NF2 cell culture and in xenograft settings, such small molecule inhibitors have proven effective in preventing tumor growth and in inhibiting Merlin signaling. We will test three of the most advanced Pak inhibitors to determine if they are beneficial in improving auditory function in *Postn-Cre; Nf2^{flox/flox}* mice and in reducing tumor growth:

a) Do inhibitors of group A Paks restore normal hearing and/or promote tumor regression in *Postn-Cre; Nf2^{flox/flox}* mice?

b) Do pan-Pak (group A plus group B) inhibitors restore normal hearing and/or promote tumor regression in *Postn-Cre; Nf2^{flox/flox}* mice?

Role: Co-Investigator (PI: Jonathan Chernoff)

Program officer: Scott Linton, U.S. Army Medical Research and Material Command, Fort Detrick, Maryland, 21702-5012

(PI: Clapp)	07/15/11-07/14/16	5%
Children's Tumor Foundation	\$243,996	0.60 Calendar Months
Children's Tumor Foundation /Neurofibromatosis Therapeutics Acceleration Program (NTAP)		

Drug Discovery Initiative Neurofibromatosis Preclinical Consortium Center

Goals: The goal is to identify improved ways to monitor response of plexiform neurofibromas to therapy.

Specific Aims:

Aim 1: The first study will be to test MEK inhibitor (AZD 5362) as a single agent and then in combination with the AKT inhibitor (AZD 6244) in order to determine the effects these agents have on the plexiform neurofibromas.

Aim 2: In continuation with our previous funded studies with the AKT inhibitor know as, AZD 6244, and in collaboration with Dr. Brigette Widemann, we will examine alternate dosing schedules in order to determine the proper dosing for long term treatments.

Aim 3: These studies will utilize the transforming growth factor-beta inhibitor, LY2157299, in order to identify TGF- β as novel molecular target to inhibit neurofibroma formation. Due to our success translating preclinical results to clinical trials, we foresee this as a potential drug that could be accelerated to in human testing if successful in our preclinical work

Aim 4: We hypothesize that treatment with PI3K/dual mTORC1/2 inhibitor, LY3023414, will provide significant tumor reduction of plexiform neurofibromas *in vivo* and reduce contributions to tumor progression from NF1^{+/-} mast cells.

Role: Principle Investigator

Program Officer: Annette Bakker, 95 Pine Street, 16th Floor, New York, NY, 10005

P50 CA096519 (PI: Clapp/Shannon)	09/01/15-08/31/20	20%
NIH/NCI	\$2,300,000	2.40 Calendar Months

Developmental HyperActive Ras Tumor SPORE

Goals: The overall goal of this SPORE is to implement effective targeted molecular therapies for neoplasms and cancers characterized by *NF1* mutations by conducting integrated, mechanistically based translational research. This highly-qualified, collaborative group will address the following overarching objectives:

1. To evaluate novel therapeutics in validated preclinical models and in the treatment of patients with NF1.
2. To Identify risk factors of individuals with NF1 to acquire spontaneous and treatment-associated second malignancies
3. To decrease tumor associated morbidity and mortality of patients with NF1

Role: Multi-PI (David W. Clapp and Kevin Shannon)

Program Director/Principal Investigator (Last, first, middle): Chernoff, Jonathan

Program Officer: Igor Kuzmin, 9609 Medical Center Drive, Room 3W112, MSC 9726, Bethesda, MD, 20892

(PI: Ferrer)	09/01/15-08/31/18	10%
DOD	\$236,642	1.2 Calendar Months

Discovery of drug combinations for the treatment of Neurofibromatosis malignancies

Goals: To identify compounds that can inhibit signaling pathways regulated by Ras in NF1

Specific Aims:

Aim 1: High Throughput (HTS) screening of a library of chemotherapeutic agents to identify drugs that induce cells death of plexiform NF1-/- Schwann Cells.

Aim 2: Dose Response Matrix screening to identify synergistic drug combinations in plexiform NF1-/- Schwann Cells.

Aim 3: Determination of therapeutic effects of synergistic combinations in inhibiting tumor growth in mouse models of pNF1.

Role: Co-Investigator (PI: Marc Ferrer)

Program Officer: Akua Roach, U.S. Army Medical Research and Material Command, Fort Detrick, Maryland, 21702-5012

T32 DK007519	07/01/85-06/30/16	NA
NIH/NIDDK	\$499,659 No Salary Support	NA

Regulation of Hematopoietic Cell Production

Goals: The goal of this application is to continue training the next generation of scientists in the clinically-relevant medical area of the regulation of hematopoietic cell production.

Program Officer: Terry R. Bishop, The National Institute of Diabetes and Digestive and Kidney Diseases, Bethesda, MD, 20892-2560.

R42 HL099150 (PI: Vincent / Goebel)	12/01/15-11/30/18	2%
NIH/NHLBI	\$241,728	0.24 Calendar Months

HLS13-04, Lentiviral Vectors for the Treatment of Fanconi Anemia

Goals: The ultimate goal of this work is to commercialize an approved gene therapy based treatment for FANCA. The commercialization of a FA treatment will serve as a platform therapy for other orphan diseases, as the vector design allows replacing the FA gene with other therapeutic genes suitable for the treatment of immunodeficiencies, sickle cell anemia, thalassemia, hemophilia and other genetic disease of the hematopoietic system.

Specific Aims:

Aim 1: Complete Method Validation, Generate Clinical Vector Product, and Perform An Additional In Vitro Immortalization Assay. Rimedion submitted an Investigational New Drug Application (IND 15855) to the FDA in November of 2013 and comments were received in February of 2014. The FDA is in agreement with the method validation for transducing patient samples, the design of the clinical trial, and the production method and testing of the final vector product. Funds are requested to perform the final full-scale transduction validation and to generate the clinical vector. In terms of safety, no additional animal safety work was required but the FDA did request an additional replicate of the In Vitro Immortalization Assay be performed.

Aim 2: Conduct a Phase I trial in subjects with Fanconi anemia A by introducing a functioning FANCA gene into autologous CD34+ stem and progenitor cells. Cells will be treated ex vivo and reinfused into the circulation through the intravenous route. The study will initially introduce gene corrected cells without chemotherapy given the existing evidence suggesting a growth advantage for FA corrected cells. If insufficient vector transduced cells are noted after the initial three patients, escalating doses of cyclophosphamide will be given as the preparative regimen prior cell infusion (3 patients per dose, with two dose levels). The Phase II STTR request funds to treat the initial six patients on the trial.

Role: Co-Investigator (PI: Bill Vincent and Scott W. Goebel)

Program Officer: Pankaj Qasba, 6701 Rockledge Dr., Rockledge II, Room 10042, Bethesda, MD, 20892.

NF150083 (PI: Shekhar) 05/01/16-04/31/19 10%
DOD \$274,944 1.20 Calendar Months

Molecular Mechanisms and Therapeutics Development for Social and Communication Learning Deficits in NF1

Goals: The goal of this application is to further elucidate the molecular mechanisms underlying the behavioral and communication disruptions exhibited by *Nf1*^{+/-} mice and test novel therapeutic approaches to treating these debilitating symptoms seen in subsets of NF1 patients.

Specific Aim I: To determine the neural networks and molecular mechanisms involved in the social and communication deficits seen with NF1 mutation

Aim 1A: Map the brain networks and molecular mechanisms involved in the social learning and ultrasonic vocalization communications in *Wt* versus *Nf1*^{+/-} mice utilizing pERK immunocytochemistry (ICC) and NARG gene expression panels

Aim 1B: Demonstrate the functional organization of the putative brain networks for behavioral deficits seen in *Nf1*^{+/-} mice by optogenetically modulating their function through stimulating or inhibiting identified neurons in the relevant pathways.

Specific Aim II: To test if blocking *Ras-MAPK over* activity by targeting two different downstream molecular targets with novel pharmacological inhibitors will rescue the deficits in *Nf1*^{+/-} mice.

Role: Investigator (PI: Anantha Shekhar)

Program Officer: Danielle Reckley, 820 Chandler Street, Bldg. 843, Fort Detrick, MD 21702-5014.

OVERLAP

None

COMPLETED

R01 CA0155294

P50 NS052606

Kolton Pharmaceuticals – “Testing the pharmacokinetic effect anti-C-Kit antibody has on the Genesis of Plexiform Neurofibromas”

Kolton Pharmaceuticals – “Testing the pharmacokinetic effect KTN3379 has on the Genesis of Plexiform Neurofibromas”

Chow, Hoi Yee

ACTIVE

W81XWH-14-1-0141 (PI: Chernoff) 5/15/2014 - 5/14/2017 50.0%
DOD Salary only 6.00 calendar
Testing the Role of p21 Activated Kinases in Schwannoma Formation Using a Novel Genetically Engineered Murine Model that Closely Phenocopies Human NF2 Disease
The major goals of this project are: 1) Does Pak signaling influence NF2-related pathology in vivo?; and 2) Are small molecule inhibitors of Pak effective in preclinical models of NF2?
Procuring Contracting/Grants Officer: Jodi Cardoza, US AMRAA, 820 Chandler St., Fort Detrick, MD 21702 (301) 619-2693

R01 CA142928 (PI: Chernoff) 1/1/2015 - 12/31/2019 50.0%
NIH Salary only 6.00 calendar
Targeting the Kinome in Neurofibromatosis Type 1
The major goals of this project are to determine: 1) What is the status of the kinome in NF1-/- Schwann cells and mast cells, and how does this change upon Mek, Akt/mTOR, or Pak inhibition?; 2) What is the cellular basis for Pak's function in NF1-related tumors? and 3) Are small molecule Pak inhibitors effective in preclinical models of NF1? Can whole kinome analysis predict pathways for drug resistance in treated mice?
Procuring Contracting/Grants Officer: Candace Cofie, 9609 Medical Center Dr., Bethesda, MD 20892, 240-276-6317

OVERLAP

None

COMPLETED

None

Park, Su-Jung

ACTIVE

W81XWH-14-1-0141 (PI: Chernoff, FCCC) 05/14/14-05/13/17 10%
DOD/NFRP \$116,797 1.2 Calendar Months

Testing the Pharmacologic and genetic role of P21 activated kinase inhibition on Schwannoma Formation using a Novel Genetically Engineered Murine Mouse Model that Closely Phenocopies Human NF2 Disease

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Role: Investigator (PI: Jonathan Chernoff)

Program officer: Scott Linton, U.S. Army Medical Research and Material Command, Fort Detrick, Maryland, 21702-5012

NF150083 (PI: Shekhar) 05/01/16-04/31/19 10%
DOD \$274,944 1.20 Calendar Months

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Program Director/Principal Investigator (Last, first, middle): Chernoff, Jonathan

the relevant pathways.

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Role: Investigator (PI: Anantha Shekhar)

Program Officer: Danielle Reckley, 820 Chandler Street, Bldg. 843, Fort Detrick, MD 21702-5014.

OVERLAP

None

COMPLETED

None