Published in Radiation Res. 44. 224-236, 1970

AFRRI SR69-25 DECEMBER 1969



DAMOD



CORRELATION OF RADIATION-INDUCED ULTRASTRUCTURAL CHANGES IN MOUSE HEPATOCYTES WITH ALTERATIONS IN PLASMA CONCENTRATION OF PROTEIN-BOUND NEUTRAL HEXOSES

> ARMED FORCES RADIOBIOLOGY RESEARCH INSTITUTE Defense Atomic Support Agency Bethesda, Maryland

> > Distribution of this document is unlimited.

This report has been approved for open publication by the Department of Defense

All aspects of investigative programs involving the use of laboratory animals sponsored by DOD components are conducted according to the principles enunciated in the "Guide for Laboratory Animal Facilities and Care", prepared by the National Academy of Sciences - National Research Council.

AFRRI SR69-25 December 1969

CORRELATION OF RADIATION-INDUCED ULTRASTRUCTURAL CHANGES IN MOUSE HEPATOCYTES WITH ALTERATIONS IN PLASMA CONCENTRATION OF PROTEIN-BOUND NEUTRAL HEXOSES

A. A. RENE A. S. EVANS

S. (J. BAUM Chairman Experimental Pathology Department

GEORØE

Commander, MSC, USN Chairman Radiation Biology Department

HUGH B. MITCHELL Colonel, USAF, MC Director

ARMED FORCES RADIOBIOLOGY RESEARCH INSTITUTE Defense Atomic Support Agency Bethesda, Maryland

Distribution of this document is unlimited

TABLE OF CONTENTS

																						Page
For	eword (Nonte	chr	nica	l s	um	ma	ry)	•	•	•	•	•	•	٠	•	•	•	•	•	•	•	iii
Abs	tract	•	٠	•	•	•	•	•	•	•	•	•		•	•	•	•	٠	•	•	•	v
Ι.	Introduction	•	٠	٠	•	•	•	•	•	•	•	•	•	•	•	•	•	•	•	•	•	1
ш.	Materials a:	nd i	Met	hoc	ls	•	•	•	•	•	•	•	•	•	•	•	•	•	•	•	•	1
III.	Results .	•	•	•	•	•	•	•	•	•	•	•	•	•	٠	•	•	•	•	•	•	2
IV.	Discussion	•	•	•	•	•	•	•	•	•	•	•	•	•	•		•	•	٠	•	•	8
Refe	erences	•	•	•	•	•	•	•	•	•	•	•	•	•		•		•	•	•	•	11

i

LIST OF FIGURES

Page

Figure 1.	Liver cell of a fed unirradiated control	•	•	•	•	•	•	3
Figure 2.	Liver cell of a starved unirradiated control	•	•	•	•	•	•	3
Figure 3.	Liver cell of an irradiated animal in extremis .		•	•	•	•	•	5
Figure 4.	Liver cell of an irradiated animal in extremis .	•	•	•	•	•	•	5
Figure 5.	Liver cell of an irradiated animal in extremis .	•	•	•	•	•	•	6
Figure 6.	Liver cell of an irradiated animal in extremis .	•	•	•	•	•	•	7
Figure 7.	Liver cell of an irradiated animal in extremis .	•	•	•	•	•	•	7

ii

FOREWORD (Nontechnical summary)

One of the most important of the many functions of the liver is that of combining amino acids, carbohydrates (sugars), and lipids (fats) into complex proteins (biosynthesis). Each stage of these biosynthetic processes is carried out by specific structures within the cell (intracellular organelles). Other structures transfer (secrete) the completed proteins from the intracellular spaces to the blood plasma in which they are transported throughout the body.

In a previous study (AFRRI SR68-4) it was reported that marked differences occurred in the plasma concentrations of protein-bound carbohydrates (PBC) in mice which died after irradiation as compared with animals which survived identical doses.

The present report is a first approach to the explanation of the mechanism of this radiation injury. Thus, the changes demonstrated in the plasma PBC concentration by chemical methods should be referable to abnormalities in the organelles concerned with the biosynthesis and secretion of these complex proteins. Of primary interest, therefore, were the tubules in which the protein portion of the molecules is known to be formed (the rough endoplasmic reticulum), structures which mediate some of the body's utilization of sugars (mitochondria), and the Golgi apparatus, a complex arrangement of tubules which are known to be instrumental in secretory processes.

Mice were subjected to a whole-body dose of 530 rads of mixed gamma-neutron radiation. Irradiated animals, at the point of death, were killed and their livers

iii

rapidly removed and prepared for viewing by the electron microscope. Sections of livers from irradiated survivors and unirradiated mice were similarly prepared and the architecture of the intracellular structures of the groups was compared.

The most dramatic differences in the liver-cell organelles of the irradiated mice as compared with that of unirradiated animals were in the rough endoplasmic reticulum and the Golgi apparatus. In the former, the tubules were greatly distended, suggesting that excessive amounts of material were collected in them. In the same cells, the Golgi apparatus were more numerous and their passages also expanded indicating that the cell was attempting to rid itself of the abnormal volume of accumulated fluid.

The mitochondria, which are normally elongated, were, in the irradiated animals, uniformly spherical. This condition suggested that injury to their limiting membrane had caused them to assume the shape of greatest stability. That they had lost many of their granules indicated that enzyme packets instrumental in the conversion of sugars to energy had been damaged or lost.

These morphological data from electron microscopical observations are in good agreement with the biochemical studies. A cause and effect relationship remains to be established, but it would appear that ionizing radiation may produce damage to intracellular sites of synthesis of complex proteins resulting in abnormal function of the organelles and, in turn, abnormal appearance.

Further, more definitive studies are being designed to resolve the order of occurrence.

iv

ABSTRACT

C3H mice were subjected to a whole-body dose of 530 rads of mixed gammaneutron radiation delivered at a rate of approximately 20 rads/min. The blood plasma concentration of protein-bound carbohydrates, as neutral hexoses, was estimated daily after irradiation. Ultrastructural architecture of liver tissue taken from irradiated animals <u>in extremis</u> was compared with that of survivors and fed and starved unirradiated controls. Among the radiation-induced differences observed in the hepatocytes were moderate to marked dilatation of the rough endoplasmic reticulum, increased Golgi activity, and rounding of the mitochondria with a decrease in numbers of mitochondrial granules. These alterations, together with other differences noted, were correlated with the increased plasma concentration of proteinbound neutral hexoses uniformly found in animals which succumb to radiation injury.

I. INTRODUCTION

Radiation-induced alterations in the blood plasma concentration of proteinbound carbohydrates (PBC) as neutral hexoses have been studied in C3H mice.² In mice which died, the PBC concentration showed a marked increase, while that of the survivors of identical doses changed only slightly or not at all.

Although the detailed mechanism of biosynthesis of the carbohydrate containing plasma proteins has not been defined, it is well established that the liver is the principal site of their formation. The objective of this study was to observe radiationinduced abnormalities in cytoplasmic organelles hypothesized to be instrumental in biosynthesis of complex proteins and to correlate those abnormalities with changes in plasma concentration of glycoproteins.

II. MATERIALS AND METHODS

Young adult male C3H mice,^{*} 6 to 8 weeks of age and weighing between 20 and 24 g were used as experimental animals. They were subjected to a whole-body dose of 530 rads of mixed gamma-neutron radiation from the AFRRI-TRIGA reactor. The dose was delivered at a rate of approximately 20 rads/min. The characteristics of the exposure field and the array for exposure of the mice have been previously described.²

Blood samples (60 to 65 μ l) were taken prior to irradiation and at daily intervals for 10 days postirradiation by snipping about 1 mm from the tip of the tail. The blood flow was collected with a minimum of expression in capillary tubes which had

^{*} Microbiological Associates, Inc., Bethesda, Maryland

previously been coated with EDTA to prevent clotting. The plasma was recovered by centrifugation and stored in an ultralow temperature freezer (-85°C) until analyzed.

To quantify the protein-bound neutral hexoses, the sulfuric acid-orcinol technique of Weimer and Moshin¹⁰ was scaled down to permit duplicate determinations using only 10 μ l of plasma per test. Protein-bound carbohydrates (PBC) were calculated from calibration curves obtained from a series of known equimolar concentrations of galactose and mannose.

Irradiated animals <u>in extremis</u>, 30-day survivors, and starved and fed unirradiated controls were killed by cervical dislocation, and the liver was rapidly removed. The tissues were processed for electron microscopy by washing initially at 4°C in 3.5 percent glutaraldehyde and 0.05 M cacodylate buffer pH 7.2.⁵ The tissues were cut in 1 mm cubes and placed in fresh glutaraldehyde fixative overnight. The specimens were then washed in a solution of 0.05 M cacodylate buffer, pH 7.2, and postfixed in 1 percent osmium tetroxide.⁴ Following fixation, the tissues were dehydrated in graded ethanol solutions and embedded in Maraglas.⁷ The blocks were cut with a Porter-Blum MT2 ultramicrotome and sections mounted on uncoated grids. After staining with uranyl acetate⁶ and lead citrate, ⁹ the sections were examined in an electron microscope.*

III. RESULTS

The mice which survived the irradiation and the unirradiated controls maintained stable plasma concentrations of protein-bound neutral hexoses throughout the

^{*} Siemens Elmiskop 1A

observation period. Small day-to-day fluctuations were seen, but none was greater than could be attributed to the limits of experimental error in the analytical technique. By contrast, the PBC concentrations in the plasma of the animals which died increased from 1.8 to 2.2 times their preirradiation levels.

The only notable difference in ultrastructural architecture between liver parenchymal cells from fed (Figure 1) and starved (Figure 2) unirradiated mice was the appearance of numerous lipid droplets (L) in the latter. Despite inanition, hepatocytes from irradiated animals did not exhibit this lipid mobilization. No notable



Figure 1. Liver cell of a fed unirradiated control.

- Gr = mitochondrial granule
- M = mitochondria
- RER = rough endoplasmic reticulum
- N = nucleus

- Figure 2. Liver cell of a starved unirradiated control.
 - L = lipid droplet
 - M = mitochondria
 - Bc = bile canaliculus
 - Mv = microvilli

differences were seen between the hepatocytes of the 30-day survivors and the fed, unirradiated controls. Assessments of radiation-induced alterations in fine structure were therefore made by comparison with specimens from normal, fed unirradiated mice.

The most dramatic changes observed in the cytoplasmic organelles in hepatocytes of irradiated animals were in the rough endoplasmic reticulum (RER) and Golgi apparatus (Go) (Figures 3-7).

In contrast to the uniformity of the cisternae and the regular distribution of ribosomes on the thin single unit membrane of the RER of the normal mice (Figure 1), the irradiated mice exhibited moderate to marked dilatation of this structure. The cisternae of some of the RER of the irradiated animals were greatly distended, forming lakes (La) of sparse filamentous material of low electron density suggesting accumulation of cell products (Figure 3). In some areas the dilated cisternae were separated by narrow cytoplasmic septa. In other areas, cisternal fragmentation occurred forming vesicular images with and without granules very much like the spherical fragments of microsomal fractions seen after cell fractionation. In some instances, the dilation was so extensive that "enwrapping" of the mitochondria by the cisternae occurred (Enr). In many cells the dilation of the RER was associated with a loss of ribosomes (degranulation). These detached granules were seen as clusters of free ribosomes (Fr) in the cytoplasm ground substance. Alterations in the agranular reticulum were not apparent except in the vicinity of the Golgi where a large number of vesicles were present, presumably originating from the smooth endoplasmic reticulum.

There was a marked increase in the Golgi activity in the hepatocytes of the irradiated animals (Figures 3 and 4). The lamellar system was frequently dispersed over a large area. In some cells the Golgi apparatus showed marked enlargement exhibiting a large number of dilated cisternae. The cisternae were allied with vesicles containing concentrated cell products.



Figure 3. Liver cell of an irradiated animal in extremis.

- Go = Golgi apparatus
- La = lakes
- Cs = cytoplasmic septa
- Enr = enwrapping
- M = mitochondria
- N = nucleus
- \rightarrow = vesicles



- Figure 4. Liver cell of an irradiated animal in extremis.
 - Go = Golgi apparatus
 - RER = rough endoplasmic reticulum
 - M = mitochondria

Vacuoles containing condensation products were present in greater numbers after irradiation and there was indication of polarity in the packets of cisternae which often contained a visible flocculent precipitate (Figure 5, Sp).



Figure 5. Liver cell of an irradiated animal <u>in extremis.</u> <u>M</u> = mitochondria RER = rough endoplasmic reticulum Go = Golgi apparatus N = nucleus Sp = secretory product

The hepatocyte of the unirradiated mice (Figure 1) contained numerous mitochondria (M) which varied in shape according to the angle of cut relative to the longitudinal axis, from near circular to elongated cylinders. Mitochondrial granules (Gr), 20 to 30 nm in diameter, were distributed throughout their matrices. By way of contrast (Figure 6), the hepatic mitochondria in irradiated mice were uniformly circular or near circular in cross section indicating that they had lost their elongated conformation and had rounded up into spheres. The mitochondrial matrix granules of the irradiated animals were fewer in number than those of the starved and fed unirradiated controls. Indeed, in most of the electron photomicrographs of the irradiated animals there was a decrease in the number of matrix granules. In some instances (Figure 7), there appeared to be either an increase in number or fragmentation of the cristae in the mitochondria. Whether there was an increase or decrease in the number of mitochondria per cell was not ascertained.



Go Go Go Go Go RER

Figure 6. Liver cell of an irradiated animal in extremis. Go = Golgi apparatus M = mitochondria Fr = free ribosome Bc = bile canaliculus Figure 7. Liver cell of an irradiated animal in extremis.

- Go = Golgi apparatus
- Mf = myelin-like figure
- M = mitochondria
- RER = rough endoplasmic reticulum

A sporadic finding (Figure 7) was the occasional formation of myelin-like structures (Mf) in the cytoplasm of irradiated hepatocytes.

The nuclei of irradiated cells did not show regular morphological changes. Slight distension of the nuclear envelope in some cells was seen (Figure 5).

IV. DISCUSSION

Regulation of the concentration of many of the protein constituents of the blood is an important function of normal liver. Radiation interferes with this function by damaging organelles intimately concerned with such metabolic regulation. Whether one assumes that the augmentation of circulating glycoproteins in moribund animals is a result of increased synthesis, decreased catabolism, or both, is not relevant to this discussion. The correlation of ultrastructural changes with alterations in plasma concentrations of protein-bound neutral hexoses as pursued here is operational and does not permit differentiation of specific cause and effect mechanisms. These conditions do not, however, preclude speculation on the overall relationship of radiationinduced damage of certain organelles to abnormal plasma glycoprotein concentrations.

The endoplasmic reticulum and the Golgi apparatus have been implicated in biosynthesis and secretion of complex proteins in cells and appear to be related to each other structurally and functionally. In the present study, these two membranous systems were the most severely altered structures in the hepatocytes of moribund irradiated animals.

The lakes formed by the extreme dilation of the RER has been termed "hydropic degeneration"⁸ on the assumption that it reflected an increase in cellular water. That there appears to be an accumulation of fluid in the RER of hepatic cells after

irradiation need not, however, be in itself "degenerative". This accumulation could represent stored secretory product. In addition, the large number of OH⁻ groups found in all glycoproteins containing a considerable number of sugar residues interact with water molecules of the solvent, probably by H-bond formation, which results not only in increased solubility and a larger effective molecular volume than might be indicated by the molecular weight, but actual entrainment of water.

The marked increase in Golgi activity in the moribund animals' hepatocytes undoubtedly represented a compensatory attempt of the cells to eliminate the excessive amounts of products accumulated in the cisternae of the RER. The fact that the number and complexity of the Golgi apparatus vary directly with the secretory activity of various types and different functional states of cells has established their role in secretion.

It has been suggested that in cells elaborating products rich in complex carbohydrates, such as the hepatocytes in the present study, the synthesis of the oligosaccharide component may take place in the Golgi complex itself and may be combined there with protein synthesized elsewhere in the cell.³ If this be the case, the condition of the RER and Golgi provides support for the view that damage to regulatory mechanisms leads to excessive production of materials which must be carried to extracellular spaces to prevent further deterioration and death of the cell. In an occasional cell, evidence of necrosis was observed.

The most obvious changes in the mitochondria of the hepatocytes of irradiated mice were the loss of their normal elongated conformation and loss of matrix granules. That they had apparently rounded up into spheres, together with what appeared

to be mild swelling, suggested that damage to their limiting membranes had forced them to take a more stable shape. The loss of matrix granules was probably associated with the cells' change in functional state. The disappearance or decrease in numbers of these granules has been observed in cells after stimulation or secretory activity. ^{1,8} This condition has also been observed in cells exposed to an anoxic environment and is believed to be associated with a depressed rate of phosphorylation.⁸

The data presented here permit the following conclusions to be drawn. The condition of the cytoplasmic membranes and organelles of the hepatocytes of irradiated mice correlates well with the increased plasma concentration of protein-bound neutral hexoses seen in these animals. The distension of the cisternae of the RER appears to be the result either of an accumulation of excess products synthesized in the RER or excess products synthesized and incorporated in the Golgi which had overflowed and backed up into the channels communicating with the cisternae of the RER. The high amount of Golgi activity is believed to be a reflection of a compensatory response of the cell to eliminate excessive products via their secretory processes.

REFERENCES

- 1. Bulger, R. E. Fine structure of the rectal (salt-secreting) gland of the spiny dogfish, <u>Squalus acanthias</u>. Anat. Record 147:95-127, 1963.
- Evans, A. S., Quinn, F. A., Brown, J. A. and Strike, T. A. Effect of ionizing radiation on total protein-bound neutral hexoses in the plasma of mice. Radiation Res. 36:128-137, 1968.
- 3. Fawcett, D. W. An Atlas of Fine Structure: The Cell, Its Organelles and Inclusions. Philadelphia, Pennsylvania, W. B. Saunders Company, 1966.
- 4. Palade, G. E. A study of fixation for electron microscopy. J. Exptl. Med. 95:285-298, 1952.
- 5. Sabatini, D. D., Bensch, K. and Barrnett, R. Cytochemistry and electron microscopy. J. Cell Biol. 17:19-58, 1963.
- Sjöstrand, F. S. A new ultrastructural element of membranes in mitochondria and of some cytoplasmic membranes. J. Ultrastructure Res. 9:340-361, 1963.
- 7. Spurlock, B. O., Kattine, V. C. and Freeman, J. A. Technical modifications in Maraglas embedding. J. Cell Biol. 17:203-207, 1963.
- 8. Trump, B. F. and Ericsson, J. L. E. In: The Inflammatory Process, Zweifach, B. W., Grant, L. and McCluskey, R. T., editors. New York and London, Academic Press, 1965.
- 9. Venable, J. H. and Coggeshall, R. A simplified lead citrate stain for use in electron microscopy. J. Cell Biol. 25:407-408, 1965.
- Weimer, H. E. and Moshin, J. R. Serum glycoprotein concentrations in experimental tuberculosis of guinea pigs. Am. Rev. Tuberculosis 68:594-602, 1953.



DISTRIBUTION LIST

AIR FORCE

The Surgeon General, U. S. Department of the Air Force. Washington, D. C. 20333 (1)

- Executive Officer, Director of Professional Services, Office of the Surgeon Ceneral, Hq. USAF (AFMSPA) T-8. Washington, D. C. 20333 (1)
- Hcadquarters, U. S. Air Force (AFMSPAB), Washington, D. C. 20333 (1)
- USAFSAM (SMBR), ATTN: Chief, Radiobiology Branch, Brooks AFB, Texas 78235 (1)
- Air Force Weapons Laboratory, ATTN: WLIL (1), ATTN: WLRB-2 (1), Kirtland AFB, New Mexico 87117 (2)
- Chief, Nuclear Medicine Department, P. O. Box 5088, USAF Hospital, Wright-Patterson AFB, Ohio 45433 (1)
- Office of the Command Surgeon (ADCSC), Hq. ADC, USAF, Ent AFB, Colorado 80912 (1)

Commander, 6571st Aeromedical Research Laboratory, Holloman AFB, New Mexico 88330 (2)

ARMY

The Surgeon Ceneral, U. S. Department of the Army, Washington, D. C. 20315 (1)

Surgeon Ceneral, ATTN: MEDDH-N, U. S. Department of the Army, Washington, D. C. 20315 (1)

USACDC CSSG, Doctrine Division, Fort Lee, Virginia 23801 (1)

CG, USCONARC, ATTN: ATUTR-TNG (NBC), Fort Monroe, Virginia 23351 (1)

Commanding Officer, U. S. Army Medical Research Laboratory, Fort Knox, Kentucky 40121 (1)

Commanding Officer, USA Nuclear Medical Research Detachment, Europe, APO New York, New York 09180 (2)

Army Research Office, ATTN: Chief, Scientific Analysis Branch, Life Sciences Division, 3045 Columbia Pike, Arlington, Virginia 22204 (1)

Division of Nuclear Medicine, Walter Reed Army Institute of Research, Walter Reed Army Medical Center, Washington, D. C. 20012 (5)

- Commanding Officer, U. S. Army Environmental Hygiene Agency, ATTN: USAEHA-RP, Edgewood Arsenal, Maryland 21010 (1)
- Commandant, U. S. Army Medical Field Service School, ATTN: MEDEW ZNW, Fort Sam Houston. Texas 78234 (1)

NAVY

Chief, Bureau of Medicine and Surgery, U.S. Navy Department, Washington, D.C. 20390 (1)

Chief, Bureau of Medicine and Surgery, ATTN: Code 71, U.S. Navy Department, Washington, D.C. 20390 (1)

Director, Biological Sciences Division, Office of Naval Research, Washington, D. C. 20360 (1)

Commanding Officer, Naval Aerospace Medical Institute, NAMC, ATTN: Research Director, Pensacola, Fla. 32512 (3) Head, Animal Behavioral Sciences Branch, Naval Aerospace Medical Institute, Naval Aerospace Medical Center, Pensacola, Florida 32512, ATTN: Dr. John S. Thach, Jr. (1)

Commanding Officer, U. S. Naval Hospital, ATTN: Director, REEL, NNMC, Bethesda, Maryland 20014 (1) Commanding Officer, Nuclear Weapons Training Center, Atlantic, Nuclear Warfare Department, Norfolk, Virginia 23511 (1)

<u>D.O.D</u>.

Director, Defense Atomic Support Agency, Washington, D. C. 20305 (1)

Director, Defense Atomic Support Agency, ATTN: DDST, Washington, D. C. 20305 (1)

Director, Defense Atomic Support Agency, ATTN: Chief, Medical Directorate, Washington, D. C. 20305 (4)

Director, Defense Atomic Support Agency, ATTN: Technical Library (APTL), Washington, D. C. 20305 (2)

Commander, Field Command, Defense Atomic Support Agency, ATTN: FC Technical Library, Sandia Base, Albuquerque, New Mexico 87115 (1)

Director, Armed Forces Institute of Pathology, Washington, D. C. 20305 (1)

Administrator, Defense Documentation Center, Cameron Station, Bldg. 5, Alexandria, Virginia 22314 (20)

OTHER COVERNMENT

- U. S. Atomic Energy Commission, Headquarters Library, Reports Section, Mail Station G-17, Washington, D. C. 20545 (1)
- U. S. Atomic Energy Commission, Division of Biology and Medicine, Washington, D. C. 20545 (1)

OTHER GOVERNMENT (continued)

 U. S. Atomic Energy Commission, Bethesda Technical Library, 7920 Norfolk Avenue, Bethesda, Maryland 20014 (1)
 National Aeronautics and Space Administration, ATTN: Lt. Col. Charles M. Barnes, USAF, DB-3, MSC, Houston, Texas 77058 (1)

- National Bureau of Standards, ATTN: Chief, Radiation Physics Division, Washington, D. C. 20234 (1)
- U.S. Public Health Service, Deputy Chief, Division of Radiological Health, Washington, D. C. 20201 (1)
- U.S. Public Health Service, Radiological Health Laboratory, ATTN: Library, 1901 Chapman Avenue, Rockville, Maryland 20852 (1)
- U.S. Public Health Service, Northeastern Radiological Health Laboratory, 109 Holton Street, Winchester, Massachusetts 01890 (1)
- U. S. Public Health Service, Southwestern Radiological Health Laboratory, P. O. Box 684, Las Vegas, Nevada 89101 (1)
- U. S. Public Health Service, National Center for Radiological Health, Information Office, Room 3, Twinbrook Laboratory, RBE Program, 1901 Chapman Avenue, Rockville, Maryland 20852 (1)

OTHER

Argonne National Laboratory, Library Services Department, Report Section Bldg. 203, RM-CE-125, 9700 South Cass Avenue, Argonne, Illinois 60440 (1)

Dr. Donald G. Baker, Radiobiology Department, Zellerbach Saroni Tumor Institute, 1600 Divisadero Street, San Francisco, California 94115 (1)

Brookhaven National Laboratory, Information Division, ATTN: Research Library, Upton, Long Island, New York 11973 (2)

Dr. J. S. Burkle, Director of Nuclear Medicine, York Hospital, York, Pennsylvania 17403 (1)

Director, Radiobiology Laboratory, University of California, Davis, California 95616 (1)

University of California, Lawrence Radiation Laboratory, Library, Bldg. 50, Room 134, Berkeley, Calif. 94720 (1) University of California, Lawrence Radiation Laboratory, Technical Information Division Library L-3, P. O. Box 808, Livermore, California 94551 (2)

University of California, Laboratory of Nuclear Medicine and Radiation Biology, Library, 900 Veteran Avenuc, Los Angeles, California 90024 (1)

Director, Collaborative Radiological Health Laboratory, Colorado State University, Fort Collins, Colorado 80521 (1) Dr. L. W. Davis, Radiology Department, University of Pennsylvania, 3400 Spruce Street, Philadelphia, Pa. 19104 (1) Professor Merril Eisenbud, New York University, Tuxedo, New York 10987 (1)

 Dr. T. C. Evans, Radiation Research Laboratory, College of Medicine, University of Iowa, Iowa City, Iowa 52240 (1)
 Dr. Arnold Feldman, Institute of Radiology, School of Medicine, Washington University, 510 South Kingshighway, St. Louis, Missouri 63110 (1)

Mr. Orin Gelderloos, Department of Biological Sciences, Northwestern University, Evanston, Illinois 60201 (1) General Dynamics/Fort Worth, ATTN: Librarian, P. O. Box 748, Fort Worth, Texas 76101 (1)

Gulf General Atomic Incorporated, ATTN: Library, P. O. Box 608, San Diego, California 92112 (1)

Hazleton Nuclear Science Corporation, ATTN: Library, 4062 Fabian Way, Palo Alto, California 94303 (1)

IIT Research Institute, ATTN: Document Library, 10 West 35th Street, Chicago, Illinois 60616 (1)

Dr. R. F. Kallman, Department of Radiology, Stanford University, Palo Alto, California 94305 (1)

Dr. L. S. Kelly, Donner Laboratory, University of California at Berkeley, Berkeley, California 94720 (1) Los Alamos Scientific Laboratory, ATTN: Report Librarian, P. O. Box 1663, Los Alamos, New Mexico 87544 (1) Director, Nuclear Science Conton, Loring and California (1997)

Director, Nuclear Science Center, Louisiana State University, Baton Rouge, Louisiana 70803 (2)

Lovelace Foundation for Medical Education & Research, Document Library, 5200 Gibson Boulevard, S. E. Albuquerque, New Mexico 87108 (1)

- Dr. Ross A. McFarland, Guggenheim Professor of Aerospace Health & Safety, Harvard School of Public Health, 665 Huntington Avenue, Boston, Massachusetts 02115 (1)
- Dr. J. I. Marcum, Rand Corporation, 1700 Main Street, Santa Monica, California 90401 (1)

Massachusetts Institute of Technology, M.I.T. Libraries, Technical Reports, Room 14 E-210, Cambridge, Massachusetts 02139 (1)

Dr. Charles W. Mays, Physics Group Leader, Radiobiology Division, University of Utah, Salt Lake City, Utah 84112 (1)

Dr. B. D. Newsom, Colony Oaks, Apt. 32, 18100 Nassau Bay Drive, Nassau Bay, Texas 77058 (1)

Ohio State University, Nuclear Reactor Laboratory, 1298 Kinnear Road, Columbus, Ohio 43212 (1)

Dr. Harvey M. Patt, Laboratory of Radiobiology, University of California, San Francisco Medical Center, San Francisco, California 94122 (1)

Purdue University, Nuclear Engineering Library, Lafayette, Indiana 47907 (1)

Dr. S. M. Reichard, Director, Division of Radiobiology, Medical College of Georgia, Augusta, Georgia 30902 (1) University of Rochester, Atomic Energy Project Library, P. O. Box 287, Station 3, Rochester, New York 14620 (1)

OTHER (continued)

Dr. H. H. Rossi, 630 West 168th Street, New York, New York 10032 (1)

Dr. Eugene L. Saenger, Director, Radioisotope Laboratory, Cincinnati General Hospital, Cincinnati, Ohio 45229 (1) Sandia Corporation Library, P. O. Box 5800, Albuquerque, New Mexico 87115 (1)

Scientific Committee on the Effects of Atomic Radiation, ATTN: Library, United Nations Room 3267, United Nations Plaza, New York, New York 10017 (1)

Scope Publications, Franklin Station, P. O. Box 7407, Washington, D. C. 20004 (1)

Dr. Arthur R. Tamplin, Biophysicist, Information Integration Group, University of California, Lawrence Radiation Laboratory, L-612, Livermore, California 94550 (1)

Radiation Biology Laboratory, Texas Engineering Experiment Station, Texas A. & M. University, College Station, Texas 77840 (2)

Texas Nuclear Corporation, ATTN: Director of Research, Box 9267 Allandale Station, Austin, Texas 78756 (1) Western Reserve University, Department of Radiology, Division of Radiation Biology, Cleveland, Ohio 44106 (1) Mr. Lionel Zamore, 601 Brightwater Court, Brooklyn, New York 11235 (1)

FOREIGN

International Atomic Energy Agency, Kaerntnerring 11, Vienna I. 1010, Austria (1)

European Atomic Energy Community, C.E.E.A., Library, 51 rue Belliard, Brussels 4, Belgium (1)

Dr. L. G. Lajtha, Paterson Laboratories, Christie Hospital and Holt Radium Institute, Manchester, England (1)

Dr. L. F. Lamerton, Biophysics Department, Institute of Cancer Research, Surrey Branch, Belmont, Sutton, Surrey, England (1)

National Lending Library for Science and Technology, Boston Spa, Yorkshire, England (1) Directorate of Medical and Health Services, FAF (Federal Armed Forces), Bonn, Ermekeilstr. 27, West Germany (1) Abteilung fur Strahlenbiologie im Institut fur Biophysik der Universitat Bonn, 53 Bonn-Venusberg, Annaberger Weg 15, Federal Republic of Germany (2)

Prof. Dr. H. Langendorff, Direktor des Radiologischen Instituts der Universitat, 78 Freiburg im Breisgau, Albertstrasse 23, Germany (1)

Dr. Helmut Mitschrich, Academie des Sanitaets-und Gesundheits, Weseus BW, Spezialstab ATV, 8 Muenchen Schwere-Reiterstr. 4, Germany (2)

Prof. Dr. F. Wachsmann, Gesellschaft fur Strahlenforschung m.b.H., 8042 Neuherberg bei Muenchen, Institut fur Strahlenschutz, Ingolstadter Landstrasse 1, Muenchen, Germany (1)

Joachim Emde, Col. Director ATV/Stab, ABC- und Selbstschutzschule, SpezStATV/R, 8972 Sonthofen 2/Allgaeu, Berghoferstrasse 17, West Germany (1)

Dr. M. Feldman, Section of Cell Biology, The Weizmann Institute of Science, Rehovoth, Israel (1)

Dr. G. W. Barendsen, Radiobiological Institute TNO, Rijswijk, Netherlands (1) Puerto Rico Nuclear Center, ATTN: Reading Room, College Station, Mayaguez, Puerto Rico 00708 (2)

Dr. H. Cottier, Pathological Institut der Universitat, Bern, Switzerland (1)

UNC LASSIFIED

Security Classification							
DOCUMENT CON	TROL DATA - R&D	d whan t	he overall report is classified)				
(Security classification of the, body of abstract and indexin	annotation must be after	REPOR	T SECURITY CLASSIFICATION				
Armed Forces Radiobiology Research Inst	itute	UNCLASSIFIED					
Defense Atomic Support Agency	2 8	2b GROUP					
Bethesda, Maryland 20014		N/A					
3. REPORT TITLE		TO A T	CHANGES IN MOUSE				
CORRELATION OF RADIATION-INDUCED) ULTRASTRUCTU		TON OF PROTEIN-				
HEPATOCYTES WITH ALTERATIONS IN .	PLASMA CONCEN	ILAI	ION OF PROTEIN-				
BOUND NEUTRAL HEXOSES							
5 AUTHOR(S) (Last nama, first name, initial)	· · · · · · · · · · · · · · · · · · ·						
René, A. A. and Evans, A. S.							
6. REPORT DATE	78 TOTAL NO. OF PAG	E.\$	75 NO. OF REFS				
December 1969	19		10				
B.A. CONTRACT OR GRANT NO.	94. ORIGINATOR'S REPO	DRT NUM	BER(3)				
	AFREI SR 69-	-25					
B. PROJECT NO.							
с. MC 3 90203	9b. OTHER REPORT NO(S) (Any other numbers that may be assis						
	this report)						
d.							
10. A VAIL ABILITY/LIMITATION NOTICES							
Distribution of this document is unlimited							
11. SUPPL EMENTARY NOTES	12. SPONSORING MILITA	RY ACTI	IVITY				
	Defense Atomio	Suppo	ort Agency				
Washington D C 20305							
13. ABSTRACT		_					
C3H mice were subjected to a whole	e-body dose of 530	rads	of mixed gamma-neutron				
radiation delivered at a rate of approxim	nately 20 rads/mir	ı. The	e blood plasma concen-				
tration of protein-bound carbohydrates, a	s neutral hexoses	, was	estimated daily after				
irradiation. Ultrastructural architecture	of liver tissue tal	ken fro	om irradiated animals <u>in</u>				
extremis was compared with that of survi	vors and fed and s	starved	d unirradiated controls.				
Among the radiation-induced differences	observed in the he	patocy	tes were moderate to				
marked dilatation of the rough endoplasm	ic reticulum, incr	eased	Golgi activity, and round				
ing of the mitochondria with a decrease in	n numbers of mito	chondr	ial granules. These				
alterations together with other difference	es noted, were co	rrelat	ed with the increased				
plasma concentration of protein-bound ne	utral hexoses unif	ormly	found in animals which				
succumb to radiation injury.		v					
Subduits to future injury.							

DD FORM. 1473

Security Classification								
		LINK A			КВ	LINKC		
NET WORDS		ROLE	₩T	ROLE	wT	ROLE	ю т	
INST	RUCTIONS							
 ORIGINATING ACTIVITY: Enter the name and address of the contractor, subcontractor, grantee, Department of De- fense activity or other organization (corporate author) issuing the report. REPORT SECURITY CLASSIFICATION: Enter the over- all security classification of the report. Indicate whether "Restricted Dats" ia included. Marking is to be in accord- ance with appropriate security regulationa. GROUP: Automatic downgrading is specified in DoD Di- rective 5200.10 and Armed Forces Industrial Manual. Enter the group number. Also, when applicable, show that options! markinga have been used for Group 3 and Group 4 as author- ized. REPORT TITLE: Enter the complete report title in all capital letters. Titles in all cases should be unclassifica- tion, show title classification in all capitals in parenthesis immediately following the title. DESCRIPTIVE NOTES: If appropriate, enter the type of report, e.g., interim, progress, aummary. annual. or final. 	10. AVA itations c imposed t such as: (1) (2) (3) (3) (4) (4)	LABILIT on further oy securit "Qualifie" eport fror "Foreign report by "U. S. Go this repor users sha "U. S. mi report dire shall requ "All dista ified DDC	Y/LIMIT dissemin y classif d request n DDC." snnounce DDC is n vernment t directly ll request litary age ectly from test throu ribution oc c uaers sh	ATION N ation of (ication, t ers may of ment and ot author sgencie: from DD t through c through mccies ma h DDC of gh f this repu	OTICES: the report ising star obtain cop i dissemin ized." s may obt C. Other ay obtain Other qual	Enter an other the odard stat bies of th astion of qualified copies of lifted use ntrolled.	ny lim- an those ements is this s of I DDC 	
 Give the incluaive dates when a specific reporting period is covered. 5. AUTHOR(S): Enter the name(s) of author(s) sa shown on or in the report. Enter last name, first name, middle initisl. If military, show rank and branch of service. The name of the principal author is an absolute minimum requirement. 6. REPORT DATE: Enter the date of the report as day, month wear or morth was a first principal author is an absolute service. 	If the Services, cste this 11. SUP tory note 12. SPO	report ha Departme fact and PLEMEN' S.	as been fu ent of Co enter the TARY NC	mmerce, price, if DTES: U	to the Off for ssle t known se for sdo VITY: F	ice of Te o the pub litional e	chnical lic, indi- xplans-	
 7a. TOTAL NUMBER OF PAGES: The total page count should follow normal pagination procedures, i.e., enter the number of pagea containing information. 	the depar ing for) t 13. ABST summsry it may sl	rtmental p he resear RACT: E of the do so sppear	roject of ch and de nter an a cument in elsewhe	fice or la evelopment bstract g dicstive re in the	boratory nt. Inclusiving a br of the rep body of t	sponsorin de addres ief and fa port, even he techni	g (pay- s. actual though cal re-	

7b. NUMBER OF REFERENCES Enter the total number of referencea cited in the report.

8a. CONTRACT OR GRANT NUMBER: If appropriate, enter the applicable number of the contract or grant under which the report was written.

8b, 8c, & 8d. PROJECT NUMBER: Enter the appropriate military department identification, such as project number, subproject number, system numbers, task number, etc.

9a. ORIGINATOR'S REPORT NUMBER(S): Enter the officisl report number by which the document will be identified and controlled by the originating activity. This number must be unique to this report.

9b. OTHER REPORT NUMBER(S): If the report has been assigned any other report numbers (either by the originator or by the sponsor), siso enter this number(s).

port. If additional space is required, a continuation sheet shall be sttsched.

It is highly desirable that the abstract of classified reports be unclassified. Each paragraph of the abstract shall end with an indication of the military security classification of the information in the paragraph, represented as (TS), (S). (C), or (U).

There is no limitation on the length of the abstract. How-ever, the suggested length is from 150 to 225 words.

14. KEY WORDS: Key words are technically meaningful terms or short phrases that characterize a report and may he used as index entries for cataloging the report. Key words must be selected so that no security classification is required. Iden-fiers, such as equipment model designation, trade name, miltary project code name, geographic location, may be used as key words but will be followed hy an indication of technical context. The assignment of links, rules, and weights is optional.

GP0 885-448

UNCLASSIFIED Security Classification