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Report No. IITRI C6093-13
(Final Report)

DETECTION AND IDENTIFICATION OF
CHEMICAL SIGNATURES

Ballistic Research Laboratories
Aberdeen Proving Ground, Md.

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IIT RESEARCH INSTITUTE

Report No. IITRI C6093-13
(Final Report)

DETECTION AND IDENTIFICATION OF CHEMICAL SIGNATURES

January 8, 1966 through April 7, 1969
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FOREWORD

This report discusses the experimental procedures used and results obtained during work under Contract DA-18-001-AMC-954(X) entitled, "The Detection and Identification of Chemical Signatures".

Personnel who contributed to this program include, B.K. Krotoszynski, A. Dravnieks, Wanda Bahmet, H.W. Neumann, N.S. Snaw, T.A. Stanley, and Joyce Burton.

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DETECTION AND IDENTIFICATION OF CHEMICAL SIGNATURES

ABSTRACT

In the work on the detection and identification of human chemical signatures, experimental and computerized statistical techniques were first developed, tested, and standardized. The experimental techniques solved the problems of sample collection, recovery, injection into the gas chromatograph, calibration and characterization of signature components. The sample collections were based on the principle of equilibration and were performed by means of Apiezon L-coated Teflon powder in the form of a fluidized bed. The collected samples were transferred into a special injector needle by means of an IITRI-designed transfer system. The injection of samples into the gas chromatograph was performed in a timed, automatic and reproducible manner with a specially constructed injection system. The calibration method permitted correlation of the concentrations of known signature components in air with their respective peak areas. The sensitivity of this novel process is of the order of one part per billion (by volume) of an organic component in air.

The developed techniques were used to collect and analyze vapor samples from humans temporarily isolated from the environment by being placed in a special glass cell containing high purity air. The glass cell was fitted with low-adsorptive Teflon gaskets and a Teflon coated resting platform. The gas chromatographic blanks of air obtained from the system contained a few small size peaks in the high-volatility range of the chromatograph when analyzed by the developed process.

In 65 human chemical signatures collected and analyzed in the packed 10 ft Carbowax 20M column at 125°C, 32 to 48 peaks per signature were observed. Statistical pattern analysis using stepwise discriminant analysis on these human chemical signatures which included blocks of Caucasian males, Indian (from India) males, Caucasian males, and on 10 chemical signatures of assorted

environments showed that valid discriminant functions can be obtained utilizing 5 to 15 of the gas chromatographic ranges. These findings indicated that source-type-characteristic features exist in the moderately resolved composition of vapors from humans obtained gas chromatographically. These features can be used as a basis for detection and identification processes.

Higher resolution gas chromatography at several temperature levels in a 200-ft support-coated open tubular Carbowax 20M column exhibited typically 100 to 130 peaks. Twenty-five of these, located in the gas chromatographic ranges where peaks were most frequently observed in the lower-resolution human chemical signatures, were re-analyzed in a specially designed two-column gas chromatograph in a 50-ft Apiezon L column. Each one of the 25 peaks consisted of 3 to 10 chemical components, or more than 150 components (total) when examined in the two column apparatus. The polarity of the components indicated the presence of considerable amounts of hydrocarbons, carbonyl compounds, and others, such as alcohols. Other materials, exceeding the alcohols in their gas chromatographic polarity, are probably bifunctional compounds or those with a functional group and more than one double bond in the molecule. Analysis of collected samples in a specially designed three-column gas chromatograph utilizing computerized data interpretation similarly indicated a great complexity of composition.

Mass spectrographic identification of the components, with a few exceptions (moth-proofing compounds were found) was not successful. The analysis with the two-column gas chromatograph demonstrated that the principal reason for this difficulty was the lack of resolution, even by analysis using a single 200-ft tubular column. Satisfactory future identification should be possible using the resolved output from the combination, 200-ft Carbowax 20M/50-ft Apiezon L columns in series.

The raw data were tabulated in terms of Kovats retention index and are listed in the Appendixes.

1.0 INTRODUCTION

In the previous project, "Detection and Identification of Chemical Signatures," Contract DA-18-001-AMC-954 (X), the following work was conducted: (1) a glass cell apparatus was designed and constructed for temporary isolation of human subjects in a stream of highly purified air. (2) a vapor collection, sample transfer, and gas chromatographic procedures were developed for obtaining a gas chromatographic representation of organic components in the effluents from the temporary isolated subjects, and (3) several representative gas chromatograms of several subjects of different races and both sexes were obtained.

The objective of the present project was (1) to further improve the vapor collection and evaluation techniques, (2) obtain a sufficient number of chemical signatures (gas chromatographic representations) of vapors from a variety of subjects so that some generalizations would become possible, (3) to characterize, as much as possible, those human vaporous effluent components that may be most suitable for identification of human vapors in various environments, and (4) to prognosticate on the most promising approaches to the effluent composition characterization processes that might be suitable for identification of human-related vaporous components.

As the work progressed, the variability of the chemical signatures among the individuals and the complexity of the vapor composition emerged as the dominant features complicating the tasks (3) and (4) of the objectives. Consequently, much effort had to be spent on techniques of data obtaining and interpretation, to facilitate the tasks (3) and (4). The work consisted of considerable improvement in the techniques in statistical pattern analysis to direct attention to those gas chromatographic features which are most human-related and high-resolution gas chromatography and some mass spectrography to better characterize the chemical nature of the organic fractions of the effluents from humans.

2.0 SELECTION OF EXPERIMENTAL APPROACHES

The previous work indicated that human chemical signatures vary widely among the individuals and to a considerable extent in the same individual and contain many organic components. The volatility of the components encountered covered a broad range, beginning with volatile substances apparently belonging to the lowest members of the homologous series and expected to occur in most atmospheres and extending to substances with much lower volatility such as expected from organic substances with ten or more carbon atoms in molecule. It became obvious that the human-character components may easily occur in the heavier (lower volatility) sections of gas chromatographic spectrum and that the chromatographic peaks observed most likely consist of more than one substance. At that juncture, two courses were possible: (1) work toward improvement of gas chromatographic resolution with detailed study of components encountered in a few individuals, or (2) survey of chemical signatures of many individuals and some environments with intention to locate those gas chromatographic features which would be most suitable for discrimination of humans vs environments so that the later higher-resolution studies could be concentrated on the significantly discriminating ranges.

The second of these routes was selected, to avoid the possibility of wasting initial efforts on identification of those components which later might be found not to be emitted by many humans or be not particularly suitable for detection of vapors from humans in environments. Since the chemical signatures extended into the range of heavier substances, the air sampling technique had to be modified before data accumulation could begin to permit sufficient collection of substances with lower vapor pressures.

The above considerations guided the experimental approach through the following stages:

1. Fluidized bed collector method with suitable vapor transfer devices and calibration apparatus were developed to permit systematic and reproducible chemical signature collection and transfer to the gas chromatograph.
2. After preliminary experimentation with the fluidized bed collectors for human chemical signature collection, standardized samples of organic vapors were obtained in 65 samplings from individuals of both sexes and several races (with some individuals repeated) and in 10 samplings from several environments.
3. The collected samples were analyzed using 10-ft packed bed Carbowax 20M column at 125°C, obtaining 75 gas chromatographic patterns.
4. The patterns were submitted to computerized statistical analysis, to detect those features that are most suitable for discrimination of different vapor source classes e.g., humans vs environment, different groups of humans (males vs females, Indian males vs Caucasian males, etc.). Several statistical approaches and rationales were used, and resulted in pointing out those gas chromatographic ranges which contained components potentially useful for source class discrimination.
5. Several methods of high-resolution gas chromatography were developed to increase the component resolution to a degree where mass spectrographic analysis of individual components would be possible. These methods progressed through: 50-ft and 200-ft support-coated open tubular single-column gas chromatograph; a two-column chromatograph where peaks from the first high-resolution polar column could be individually injected into a second, non-polar column; and a three-column gas chromatograph where the entire pattern could be obtained in polar and non-polar columns and correlated using specifically written computerized peak-matching program. Because of the high complexity of the sample, the three-column patterns lost considerable detail and appeared to present principally the gross composition of the chemical signature. The two-column device was judged to yield most information, and the chemical signature of one individual was studied in detail by repeated analyses with this device, inspecting the ranges indicated significant by the statistical analysis.
6. The initial attempts to mass spectrographically identify the components after resolution in a 50-ft support-coated open tubular Carbowax 20M column did not yield, with one exception, clearly identifiable components. Later work with similar single 200-ft Carbowax 20M column and with the two-column device indicated that

even the peaks emerging from the 200-ft column were further separable into 2 to as many as 10 components. This explained the difficulty experienced in the earlier mass spectrographic attempts of the component identification and pointed the way, in the form of the two-column device, for obtaining gas chromatographic resolution sufficient for more successful mass spectrographic identification.

7. An initial listing system for classification of the components of the chemical signatures in terms of retention (Kovats) indexes was adapted, and some considerations for chemical signature detector devices were evolved.

3.0 EXPERIMENTAL

The various subsections of this chapter deal with the chemical signature collection and vapor transfer techniques, calibration, subject selection, statistical pattern analysis experiments, and several techniques for high-resolution analysis of the collected samples.

3.1 Glass Cell Apparatus

The 18-in I.D. cylindrical glass cell, with Teflon ring seals, Teflon-coated end plate, and a Teflon-coated steel frame stretcher for the support of human subjects in the supine position in the middle of the cell has been described in detail in the previous report.*

For the chemical signature collection, the subjects were enclosed in the cell for 45-min in a stream of highly purified air. The air was taken from hospital grade cylinders. Experience indicated that the commercially available hospital grade air contained organic impurities in varying concentrations sufficient to make the analysis of the vapors emitted by humans uncertain and cumbersome.

Experiments were conducted to design purification procedure that would reduce the contaminant level in the air fed to the cell to a practical minimum. Air first passes through a particle filter (Vetricel, Type VF-6, 47mm Dia, Pore size .45U. Gelman Instrument Company, Ann Arbor, Michigan) then in downflow through an activated charcoal bed (Figure 1) cooled in its lowest one third section by a dry-ice-propanol mixture. The gradual cooling of air in the charcoal bed prevents aerosol formation, since vapors adsorb at approach to their dew point and before an overcooling that can lead to aerosol formation occurs. The diameter of the carbon bed was 4-in and the rate of air flow 0.75 l/sec. The

*Report IITRI No. U6012-4, Contract DA-11-022-AMC-1775(X)

charcoal bed adsorber was activated and stored at 200°C in nitrogen stream between uses, so that each chemical signature collection experiment began using a freshly reactivated adsorber.

The purified air entered the glass cell at very low humidity level, but sufficient humidity was picked up from human subject to result in 40-50 percent relative humidity at the exit of the cell. The exit air temperature typically was 23° to 25°C. Higher humidities for which provisions were made in the initial design of the air preparation train were avoided, since the subjects tended to begin perspiring profusely, and the excess humidity resulted in clouding the glass and in difficulties with water condensation in the vapor collectors kept at room temperature.

The content of oxygen in the air decreased by less than 2 percent absolute in passage through the cell when it was occupied by a subject.

3.2 Fluidized Bed Collectors

The organic vapor collection was conducted using the principle of absorption in highly purified grease films. This principle, used in gas chromatography in the form of "pre-column" collectors, permits collection of organic materials from vapor phase without interfering accumulation of water vapor, if non-polar greases are used. However, in its usual form of a packed bed, consisting of grease or oil-coated inert powders, it has certain disadvantages: the vapors of substances that are more volatile and less soluble in the grease can saturate the collector upon passage of certain limited air volume and the excess begins to pass through the collector, while the less volatile or more oil-soluble vapor components may still continue to be fully removed by the collector from the air passing through the collector. Therefore, at any given volume of air passed, some vapor components will reach the equilibration concentration and their amount in the collector will not further increase, regardless how much more air is passed. Other components will continue to accumulate until their specific

equilibration conditions will be met.

The maximum amount of a substance that can be collected from its vapors in air is determined by several physicochemical variables combined as follows:

The number of molecules of certain species collected is

$$n = 6 \cdot 10^{23} \frac{d}{M} \cdot \frac{1}{\gamma} \cdot \frac{n_a}{n_0} \cdot V \quad (I)$$

Here d and M are the density and the molecular weight of the film substance, respectively; γ is the activity coefficient of the species in the film; n_a is the concentration, molecules/cm³, of the species in the film; n_0 is the concentration in gas at saturation at the same temperature; and V is the volume, cm³, of the film phase in the sampler.

The activity coefficient is large for cases when the vapor is polar while the collecting phase is non-polar. For the same vapor and collecting phase, the amount collected is proportional to the concentration of this vapor in air (n_a) if the collector temperature and consequently n_0 are maintained constant.

For convenience, it can be pointed out that n/n_a represents the volume of air, in cm³, from which the particular vapor can be fully extracted, and by rearranging Eq. 1,

$$\frac{n}{n_a} = 6 \cdot 10^{23} \frac{d}{M} \cdot \frac{1}{\gamma} \cdot \frac{1}{n_0} \cdot V \quad (II)$$

it is seen to be essentially independent of the concentration of the vapor in air, since all terms on the right side are either concentration-independent, or occasionally only partially dependent as is the case with the activity coefficient. The ratio n/n_a is equivalent to the gas chromatographic retention volume of the specific component in the collecting phase at collection temperature--usually room temperature.

In the later gas chromatographic analysis, unless it is conducted using the same stationary phase as in the collector

(Apiezon L), it is impossible to determine for which components the air volume passed through the collector has exceeded the n/n_a volume. The most convenient solution for this indeterminacy was found in developing a fluidized bed collector. Here the collecting phase, in the form of Apiezon L films dispersed on Teflon particles, is placed in a capsule through which air to be sampled is flown at a high rate. The vapors in air reach each particle easily, since the powder is in a violent motion, and an equilibration is reached on all particles essentially simultaneously. For fluidized bed samplers coated with 10-wt percent of Apiezon L, equilibration was reached within 20-30-min. To assure equilibration, the collection was continued for 45-min.

The fluidized bed material is contained in a cylindrical glass column, 5-in long and 1-1/4-in wide (Figure 2). The column is provided with stainless steel screen discs at each of its ends. The diameter of the screen openings is selected to retain the powder in the column during vapor collection and to exert minimum resistance to the airflow.

The fluidized bed material is introduced into the column through a side arm, which is then sealed off. The bed material remains in the column during its conditioning, vapor collection, and vapor transfer. The bed material consists of 10 g of Fluoropak 80 coated with 10% Apiezon L grease; therefore a typical fluidized bed sampler contains 1 g of Apiezon L in the form of a film a few microns thick. From many conventional gas chromatographic phases tried, only Apiezon L permitted later recovery of dissolved organics without artifacts (extra peaks) from decomposition of the trapping phase itself.

Before each vapor collection, the bed material is conditioned by heating the bed at 80°C and flushing it with pure helium gas until all volatile impurities are removed. The conditioning efficiency is established by collecting residual impurities in an injector needle (cf Section 3.3) and gas chromatographing the collected sample. The bed material is considered clean when the

residual impurities produce peaks of negligible areas. Temperatures higher than 80°C maintained for 1 hr led to incipient thermal decomposition of Apiezon L with formation of volatile compounds.

The bed material is then exposed to air carrying the vapors to be collected and analyzed. Air is either pushed or pulled by a small blower through the sampler. In typical vapor collections, the air flows at the rate of 0.5 to 0.75 liters/sec. At rates of this order of magnitude, the powder is constantly floating and is in excellent and uniform contact with flowing air.

During vapor collection the bed is kept at ambient temperatures, which are measured because the number of molecules collected through equilibration is a function of temperature. Experiments with n-butanol vapor in air indicated that at ambient temperatures the n-butanol content of the bed reaches two-thirds of the equilibrium value within 20 min and for all practical purposes reaches equilibrium value within 45 min. The rate-limiting factor is the diffusion of the organic material, e.g., n-butanol, into the greaselike film. Faster equilibration would require lower-viscosity films, which, however, would be more volatile and could introduce artifacts during analysis. Faster equilibration would also be achieved at a higher bed temperature, but a smaller finally collected amount and a loss of simplicity in collection would result.

Since collection involves equilibration, the actual airflow rate through the sampler does not have to be known and is not critical as long as the powder floats and does not pack at the end screens. Therefore it is unnecessary to measure airflow rates during sampling. However, water condensation in the sampler must absolutely be avoided, since condensed water constitutes a competitive collection phase having an unknown, variable volume. Also, if the concentration of the organic components in air changes rapidly during the sampling, the collected amount will not follow the trends well. Therefore the sampler is best suited for sampling from air in which the concentrations of the organics are reasonably

stable. The time constant of the sampler is of order of 20 min. Concentration fluctuations that are much slower or much faster than this value will be unimportant.

3.3 Vapor Transfer and Injection Apparatus

Vapor components trapped in the thin film in the fluidized bed are transferred, by elution with cryogenically purified helium gas (Figure 3), into a stainless steel thin-wall injector needle (Figure 4). During transfer, the bed material is heated to 80°C and the injector needle is cooled to liquid nitrogen temperature in a specially designed copper block immersed at its bottom end in liquid nitrogen. The typical duration of vapor transfer is 1 hr, and 70 to 80% of the organic vapor sample is transferred into the injector needle. Higher efficiency can be achieved by longer transfer at the expense of analytical time. Higher temperatures result in incipient breakdown of Apiezon L and peak artifacts.

The vapor sample in the injector needle is delivered to the chromatographic column by means of an injection system (Figure 5). The essential parts of the injection system include an electrical motor (A) with an eccentric cam, a stainless steel bellows (B), a stainless steel cylindrical bellows enclosure (C), a pressure gauge (D), a stainless steel capillary U-shaped cold trap (E), and a stainless steel injector needle (F).

The motor, acting through the eccentric and a spring-loaded pushing rod, can draw a 1-cm³ volume of carrier gas from the injection port and into the bellows and later employ this volume to inject the sample from the heated injector needle into the septum of the column. The bellows enclosure provides a smooth fit of the bellows, with a minimum of dead space. The pressure gauge indicates the injection port pressure at any column temperature and any temporary pressure change due to injection. The gauge also serves to indicate, by a pressure lower than usual, any leaks in the septum/needle system. The U-shaped trap, which is 1/16-in

in outside diameter and cooled to liquid nitrogen temperature, blocks the passage of impurities from the bellows and the pressure gauge into the injector needle. When not in use, the injection system is kept under the flow of pure helium gas. During this standby interval, helium flows into the system at a measured rate (flowmeter G) and pressure (pressure gauge H) from a separate source through a liquid nitrogen trap (I) and a T-joint (K).

Sample injection involves the following steps. The needle containing the sample and kept in dry ice is attached to the injection system at the outlet of the cold trap by means of a Swagelok connector (L). The helium flow is then discontinued at K by closing the connection with a Swagelok nut provided with a metal rod. The other end of the injector needle is attached to the injection port of the chromatograph (M) by means of a Conax connector equipped with a Teflon sealing plug, and time is allowed for attainment of the injection-port pressure, as indicated by the pressure gauge (D).

A 1-cm³ volume of carrier gas is drawn into the injection system by means of the motor-driven bellows. Again, time is allowed for stabilization of the pressure. To inject the vapor sample, the needle is heated to the selected temperature by passing a 20-amp current along the needle while the vapors are flushed into the septum with the 1-cm³ volume of carrier gas stored previously in the bellows. The needle temperature rises from -80 to 180°C in 5 sec; it then remains at 180°C for the next 5 sec. Injection of helium gas from the bellows starts simultaneously with the heating of the needle and continues for 10 sec. During this interval, the helium from the bellows carries the vaporized sample into the septum while the normal carrier gas flow at 60 cm³/min continues.

The injection system was tested on a large number of various samples, and it was found to produce reproducible and practically quantitative injections, with 95 to 98% of the sample introduced into the septum in one injection.

3.4 Calibration Apparatus

Calibration of the gas chromatographic readout for known compounds is relatively simple. To calibrate for any component of the vapor, the component must first be identified and obtained in a pure state from a commercial or a synthetic supply.* An airflow system (Figure 6) is set up in which breathing-quality air from a commercial pressurized tank is passed through an aerosol filter (A) of a material such as Teflon, a flowmeter (B), and a freshly activated carbon bed (C) cooled in a dry ice-propanol mixture (D) at its exit end. These precautions remove organic impurities from the air. The purified air is warmed to room temperature. It flows through a 2-liter three-neck flask (E), mounted with the necks facing downward, into a clean fluidized bed trap (F). One neck of the flask is connected to an upright small cylinder (G) with a small amount of pure component on its closed bottom. The cylinder is thermostated either at or below the temperature of the flask.

The amount of component that diffuses into the airstream in the flask depends on the vapor pressure of the component and the diffusion coefficient of the component in air. The diffusion rate can be approximately calculated from these values and from the cross section and the length of the gaseous diffusion path formed by the cylinder volume above the component sample. The amount is also directly measurable by experimentally determining the weight loss of component over a longer period of time.

With a controlled and known airflow rate through the flask and the sampler, the molecular concentration of the component in the air flowing through the sampler can be calculated. For instance, during calibration for n-butanol the source intensity

*For unknown components, their approximate concentration in air can be calculated if analysis is conducted using Apiezon L column, and the flame ionization detector's sensitivity is known; the gas chromatographic position gives the retention volume which can be recalculated to ambient temperature using Kovats Index system.

was 6×10^{15} molecules/sec and the airflow was $750 \text{ cm}^3/\text{sec}$. The concentration of n-butanol in the calibration stream was therefore 8×10^{12} molecules/ cm^3 . In a typical calibration this concentration of n-butanol in air resulted in an n-butanol peak with an area of 220 cm^2 at a sensitivity of 0.1×1 (the highest sensitivity setting on the chromatograph). This calibration procedure ties the concentration of a component in air directly to the size of the peak on the chromatogram if a standardized sampler and standardized sample-handling procedures are used. In the above case, 1 cm^2 of peak area at 0.1×1 sensitivity corresponds to 3.6×10^{10} molecules per cubic centimeter of air, and, since smaller peaks are detectable, the detection limit is somewhat lower than parts per billion by volume.

The factor relating the peak area to the concentration in air depends on the partition coefficient, k , for the component i in Apiezon L versus air; $k_i = (n_c)_i / (n_a)_i$ with n_a for concentration in air. It is possible to estimate the value of this coefficient from the amount of Apiezon L in the sampler, the estimated sample transfer efficiency during transfer from the sampler to the gas chromatograph in the standardized 1-hr elution procedure, and the knowledge (obtained by direct calibration) of the absolute number of molecules corresponding to 1 cm^2 of the peak area. For n-butanol in Apiezon L at room temperature, the partition coefficient was 4000. Therefore the described procedure corresponds to a concentration of n-butanol that is 4000-fold greater than that in air, and the analysis is equivalent to a one-shot injection of n-butanol contained in 4 liters of air.

If the molecular weight of Apiezon L is approximately 1000, the activity coefficient for n-butanol in this medium is, very approximately, 3. For larger polar molecules and for smaller but less-polar molecules, lower activity coefficients and correspondingly higher partition coefficients are expected. Table I shows calibration data for some polar and relatively small molecules for which Apiezon L is a poor concentrating medium.

Nevertheless, satisfactory collection is obtained even for these vapors. For the less volatile substances, a higher relative sensitivity is expected since the partition coefficient increases with decrease of the vapor pressure. Similarly, higher enrichment will occur with other, less polar vapors. When a vapor source is enclosed in a stream of air flowing at a known rate through the enclosure and then through the sampler, the total source strength can be calculated. For example, the rate of buta 1 production from a human subject was found in one case to reach 10^{15} molecules/sec.

3.5 Field Sampling Apparatus

For the collection of vapors emitted by humans enclosed in the glass cell (cf. Section 3.1) the fluidized bed sampler was placed at the air exit from the cell. It was possible to place two or three collectors in series and collect several comparable samples simultaneously.

For sampling in open environments, an apparatus shown in Figure 7 was used. It consisted of a 12-volt automobile type vacuum cleaner driven by a portable (airplane type) 12-volt battery. The cleaner pulled the air through the fluidized bed sampler. Provisions were available for water circulation through a glass jacket integral with the sampler, to maintain desired (room temperature) temperature level if the outside air was too cold or too warm. Since fluidized beds act as very efficient heat transfer media, additional pre-thermostating of air before entrance into the sampler would be necessary if ambient temperature extremes were encountered. In the present work, environment temperatures were selected so as to avoid such extremes and simplify the collection and calculations.

3.6 Gas Chromatographic Techniques

3.6.1 Packed Columns

The primary objective of the initial phase of the experimental work was the acquisition of a set of gas chromatographic patterns of human chemical signatures from subjects of various races and sexes as well as from different environments under simple and reproducible analytical conditions. For this purpose, a packed analytical column was selected and the analyses were performed under isothermal conditions at a moderate temperature. The selected column was 10-ft long and 1/8-in O.D. coiled stainless steel tubing packed with 20% Carbowax 20M supported on acid-washed 60/80 mesh Chromosorb P. During each signature analysis, the column was kept at 125°C in the air oven of the dual-channel Aerograph model 204 gas chromatograph. The selection of a packed column was based mainly on its relatively high stability or low rate of secondary aging - a favorable condition for minimizing the effect of column age on its analytical performance. The isothermal analytical conditions were chosen with some sacrifice on component resolution, to maintain maximum retention time reproducibility for each component of a complex mixture analyzed.

In the later stage of this investigation, two packed Apiezon L columns were also used in the three-column apparatus described below. In this apparatus, the packed columns were attached to one branch of the splitter connected to the outlet of a 200-ft open tubular Carbowax 20M column. The packed columns were selected to minimize the change in Apiezon L polarity due to the effluent impurities from the Carbowax column.

3.6.2 High-Resolution Column

In order to gain a deeper insight into the actual complexity of human chemical signatures, modern high separating power and high analytical speed columns were chosen and used in a

single column, a two-column, and a three-column gas chromatographic systems. The selected columns included a 50-ft, 0.020-in I.D. open tubular Carbowax 20M column, a 200-ft, 0.020-in I.D. open tubular Carbowax 20M column, a 50-ft, 0.020-in I.D. open tubular Apiezon L column and another 200-ft, 0.020 I.D. open tubular Carbowax 20M column, all support-coated.

The 50-ft Carbowax 20M column was used in combination with a mass spectrograph to help resolve human signature components in the attempts directed toward their identification.

The 200-ft Carbowax 20M column was used in combination with the 50-ft Apiezon L column in the two-column apparatus. The apparatus was designed to provide a primary resolution of sample components in the polar phase and a secondary resolution of the individual Carbowax 20M peaks in the non-polar phase. The two-column apparatus was used in characterization of chemical signature components obtained from the same Caucasian female. The apparatus is described in the following section of this report.

The second 200-ft Carbowax 20M column was incorporated into the three-column apparatus. In this arrangement, the split outlet of the long open tubular column was connected to two parallel short packed Apiezon L columns. The system was designed to produce simultaneously entire signature patterns in polar and non-polar phases and correlate the patterns by means of a specifically written computerized peak-matching procedure. The system was employed in the investigation of several signatures collected from a Caucasian female who also was the signature donor for the study performed on the two-column apparatus. The description of the system is given below.

3.6.3 Two-Column Apparatus

The modern high-resolution gas chromatographic columns provide a spectacular resolution of complex mixtures; however, many components still remain unresolved and they may be represented by single peaks. The two-column apparatus was designed and

constructed to effect resolution of such multi-component peaks and to provide data useful in characterization of the structure of the resolved components.

The two-column apparatus includes a 200-ft open tubular Carbowax 20M column (0.020-in I.D.), a 50-ft open tubular Apiezon L column (0.020-in I.D.), a 50-ft dummy column, a 3-way solenoid valve, two hydrogen flame ionization detectors and two sniffing ports. A schematic diagram of the arrangement of the apparatus is shown in Figure 8.

In dual column analyses, the Carbowax 20M column provides the primary sample resolution. The effluent from this column is split 1:1, and subsequently all or any part of the split effluent may be directed onto the Apiezon L column for secondary resolution. The peak switching step is effected by means of the solenoid valve. During this step, a switching blip appears on the chromatogram which denotes exactly the portion of the Carbowax 20M chromatogram transferred onto the Apiezon L column. Both analytical columns and the dummy column are provided with 10 cc/min helium flow. The 5 cc/min portion of the Carbowax 20M effluent is supplemented with 35 cc/min of helium and resplit, providing 20 cc/min flow to a flame detector and a sniffing port. The other 5 cc/min flow from the Carbowax column is mixed with 5 cc/min of helium and the total 10 cc/min is directed to either the Apiezon L column or the dummy column. At the exit of the Apiezon L column 30 cc/min of helium is added to the 10 cc/min flow and the total is again split so that 20 cc/min flows to the second detector and 20 cc/min to the second sniffing port. Approximately equal concentrations of sample reach the two sniffing ports; this permits a comparison of relative odor intensities when needed.

A segment of a typical gas chromatogram obtained in the dual-column analysis of a human chemical signature is shown in Figure 9. The upper portion of the figure represents peaks obtained on Carbowax 20M column; the peaks marked with asterisks were injected on Apiezon L column. The lower portion of the figure shows the

components of the injected peaks which were resolved on Apiezon L column.

The proper utilization of the two-column apparatus requires a knowledge of the retention dispersion characteristics for non-polar compounds as well as for selected polar compounds in Apiezon L stationary phase. The n-alkanes are expected to exhibit maximum retention on Apiezon L and the n-alcohols the minimum retardation. Other functional groups such as aldehydes and ketones are expected to fall between the retention times shown by the alkanes and the alcohols. The information on the retention behavior of selected compounds was acquired in a series of calibration experiments. In these experiments, the n-alkane series included C₈, C₁₀, C₁₁, C₁₂, C₁₃, C₁₄, and C₁₅ hydrocarbons, the alcohols were represented by ethanol, 1-propanol, 1-butanol and 1-pentanol; the aldehydes included propionaldehyde, butyraldehyde, hexanal, heptanal, and octanal; the ketone series embraced 2-butanone, 3-pentanone, 2-heptanone, and 2-octanone. The calibration experiments were performed under the same operating conditions and temperature (125°C) as used in the analysis of human samples. The Carbowax column effluent was allowed to flow continuously onto the Apiezon L column. For n-alkanes studied the experimental data were first expressed in terms of the respective elution times from both liquid phases; these data guided the experiments on human samples by providing the maximum time intervals required for a complete elution and display of all components of a given Carbowax 20M peak which was injected onto the Apiezon L column. The n-alkane data were then converted to corrected retention times and these were used in the construction of calibration curves needed in the characterization of components of human chemical signatures in terms of their respective Kovats Indexes on both stationary phases. The calibration data on n-alkanes are given in Table II.

The calibration data on polar compounds were first converted to respective corrected retention times. These retention times

were utilized in determining Kovats Retention Indexes for the selected compounds from previously obtained n-alkane calibration curves. The retention indexes were determined in Carbowax 20M and Apiezon L stationary phases. The respective Kovats Indexes were then employed in the construction of calibration curves for polar compounds which were displayed in the chemical logic field. These curves together with those constructed for other compounds from pertinent literature data served in the description of the structure of components of the human chemical signatures studied. The calibration data on polar compounds are summarized in Table III.

In order to minimize the inherent differences in composition of chemical signatures of different individuals, the collection experiments in this part of the work were limited to one subject and performed in a period of a few days. The selected subject was a young Caucasian female of high hygienic standards. During the experimental period, the subject was on an average diet. A total of four collections was made. In one collection, three fluidized Apiezon L beds were employed in series and two beds in series were used in each of the other collections. In the three-bed experiment, ten Carbowax 20M peaks were injected onto the Apiezon L column; in the remaining collections, eight, four, and three multi-component Carbowax 20M peaks were examined. The examination was performed with the two-column system kept at 125°C. The peaks observed on Apiezon L column and originating from the respective Carbowax peaks were described in terms of Kovats Retention Indexes and the data were plotted in two-dimensional (polar versus non-polar) chemical logic field to produce a tentative description of the structure of the components studied.

3.6.4 Three-Column Apparatus

The three-column gas chromatographic system was constructed to provide a detailed two-coordinate description (polar and non-

polar) of sample components from one single analytical run without the need for peak trapping and reinjection. The system is schematically depicted in Figure 10. The essential components of the system include a 200-ft open tubular Carbowax 20M column, a 6-ft x 1/8-in packed Apiezon L column, a 4-ft x 1/8-in packed Apiezon L column, and three flame ionization detectors. The outlet of the 200-ft column splits into two branches; one branch leads to the flame ionization detector, the other branch to two parallel Apiezon L columns. The outlet of each Apiezon L column connects to a flame ionization detector. Helium gas is delivered to the system from two independent flow trains. One train supplies the Carbowax 20M column with the flow rate of 5cc/min, the other train delivers helium to the Apiezon L columns; the flow rates are 40cc/min and 50cc/min in the long and the short columns respectively. The flame ionization detector residing at the outlet of the Carbowax column is supplemented with auxiliary helium.

The long, open-tubular Carbowax 20M column is used to provide the initial optimum sample separation. The separated sample components split into three fractions; two fractions simultaneously enter both Apiezon L columns; while, also at the same instant, the third fraction is detected by the detector attached to the Carbowax column. The Apiezon L columns are used to obtain the retention behavior for each sample component on a non-polar phase. The parallel arrangement of these columns and the combination of different lengths and carrier gas flow rates provide characteristic ratios of retention times which are useful in the subsequent peak matching procedure.

Each component of the sample is represented by three gas chromatographic peaks which emerge with characteristic sequence and elution times. The first peak appears on the detector receiving the signals from the Carbowax column, the second is eluted from the short Apiezon L column and the third from the long Apiezon L column. An example of a typical gas chromatogram (only a segment is given)

here) of a human chemical signature analyzed on three column apparatus is given in Figure 11. In order to extract useful information, the system is calibrated with various homologous series of compounds

In a typical calibration run a mixture of homologs and selected alkanes is injected into the system and analyzed at constant temperature. For each component of the mixture the characteristic elution times from all three columns are measured and converted to corrected retention data. The conversion requires the knowledge of retention characteristics for each component. This is obtained under similar analytical conditions by incorporating the component into the sample or analyzing methane before injecting the sample into the mixture

The corrected retention data are used in the establishing of the peak shifts which occur on the Apiezon L columns. These shifts are given as the ratios of the corrected retention time for each component on the long Apiezon L column and the short Apiezon L column. The value of these ratios which was found to be constant for all compounds studied is used in the peak matching procedure in the interpretation of the unknown samples. The corrected retention data are also utilized in the computation of the Kovats Retention Indexes for the components of homologous series in the two stationary phases employed. The retention indexes serve to construct calibration curves which are displayed in a chemical logic field for final characterization of components of actual samples

The results of experiments on calibration of the three column apparatus are summarized in Tables IV to X. The data on calibration performed using n-alkanes with the three column system kept at 90°C, 120°C and 150°C are given in Tables IV to VII.

It is noted that the ratios of the corrected retention times observed on Apiezon L column for all compounds studied are practically constant and temperature independent. The average

value of these ratios was used in the computerized indexing of the human chemical signature components.

The results of calibration experiments carried out with polar compounds are summarized in Tables VIII to X. The ratios of the corrected retention times on Apiezon L columns for all compounds studied are also constant and temperature independent. The elution data listed in these tables were used in the three-point match procedure designed to inspect the three gas chromatographic records for the presence of such compounds in human chemical signatures.

3.6.5 Calibration for Conversion to Kovats Indexes

The initial series of gas chromatographic patterns of human chemical signatures was acquired without calibrating the analytical system with a synthetic mixture of n-alkanes. In order to evaluate the daily instrumental variations affecting the positions of the signature components in their respective patterns and to assign Kovats Indexes to those components, a set of two peaks always observed in each pattern was selected and used as pattern "internal standard". One of the selected peaks emerged in the initial section of the patterns with an average retention time of 2.80 ± 0.09 min. (corresponding to n-butanol), the other peak was observed in the final section of the patterns with an average retention time of 27.84 ± 0.47 min. The results of the retention time standardization for the initial series of the chemical signature patterns were then correlated with those obtained in the second series of patterns which were standardized with a synthetic mixture containing hexane, octane, decane, dodecane, tetradecane, hexadecane, and 1-butanol. The correlation facilitated the assignment of the statistical pattern analyses of both series of patterns and provided data needed in the computation of Kovats Indexes for all pattern components.

The Kovats retention indexes were computed for each component of the signatures by using the following equation:

$$I(A) = 100 \cdot (y-x) \frac{\log (t_{ra}/t_{rx})}{\log (t_{ry}/t_{rx})} + (100 \cdot x)$$

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where

$I(A)$ = retention index of compound A
 t_{ra} = retention time of compound A
 t_{rx} = retention time of n-alkane C_xH_{2x+2}
 t_{ry} = retention time of n-alkane C_yH_{2y+2}

For most of the components the calculations were made by interpolating for

$$t_{rx} \sim t_{ra} \sim t_{ry}$$

Only a few values were obtained by extrapolation. To satisfactorily determine the absolute retention values needed in the computation of the Kovats Indexes, it is necessary to know the gas hold-up time of the column under standardized analytical conditions. Since it is impossible to measure this value if the detector is insensible to inert gases, the retention time for methane is usually taken as equal to the gas hold-up time. The value was measured (0.31 min) and used to correct the retention times of the components of the human signature and synthetic mixture.

The patterns of human signatures were then compared with those of various environments and Kovats Index ranges for Carbowax 20M peaks were selected which were most significant for discrimination of humans and environments.

In order to obtain a more precise characterization of the most significant components of signatures, a systematization process was introduced to treat the data obtained on the high-resolution two-column apparatus. The principle underlying the process is illustrated in Figure 12.

The X-axis is the Kovats Index scale for retention in the Carbowax 20M column. The Y-axis is the delay, in minutes, in the Apiezon L (second stage) column, after correction for hold-up time which was obtained by calibration with methane. The Kovats Index scale that corresponds to these delays is shown on the right. The upper diagonal line represents the delay for the n-alkanes and other non-polar saturated hydrocarbons. This is then the maximum delay which a component with a given Kovats Index in Carbowax 20M can

experience in the non-polar Apiezon L column. If the compound is somewhat polar, or possesses double bonds, it will be delayed less and its point in the two-index field of Figure 12 will be lower. The zones where various types of compounds reside, on the basis of literature data, are indicated. Straight chain alcohols are the most polar of the compounds considered here (the glycol zone is also shown) and are located on a straight line. This is drawn using our own experimental points. Points for some other compounds not reported in literature are also shown.

Branched alcohols, or alcohols with OH groups not on the terminal carbon atom lie above the n-alcohol line, but short of the carbonyl compound zone. Double bonds e.g., in the alcohols, bring the points to a zone somewhat below the non-double bond compounds of the same type. Notice that the point for n-butanol lies in the alcohol zone of the continuation of the line formed by higher n-alcohols and at the same horizontal and vertical distance from hexanol. In this particular case, only a rather unusual compound (e.g., some carbonyl compound with more than one double bond) has an outside chance to produce a point that might be confused with n-butanol.

A set of calibration compounds was selected to permit frequent calibration of a plot of Figure 12 for several homologous series. This is desired to increase precision and reliability, since although the plot as it is represented appears to be reliable within ± 10 K.I. units, the actual columns used can differ slightly from the performance of columns on the basis of which Figure 12 was drawn. The causes for difference are slight differences in the average molecular weight of Apiezon L, slight oxidation, contribution of adsorption at column walls, etc.

3.7 Mass Spectrographic Identification Attempts

Various attempts were made to identify components of human chemical signature by mass spectrography. The initial experiments

were performed on signatures collected from a Caucasian male and from an Indian male. In these experiments, the packed analytical Carbowax 20M column was connected to the Hitachi Perkin-Elmer model RMU-6D mass spectrometer with a Watson Bieman sample enrichment device. The column effluent with human signature components was split so that 10cc/min entered the ion chamber and 50cc/min the flame ionization detector. The analysis was performed at 125°C.

Another series of experiments was designed to evaluate the minimum amount of component needed for its positive mass spectrographic identification to collect larger human samples and to improve its resolution. For this purpose, 100-g Apiezon L fluidized bed was constructed and tested in an experiment on signature collection from an Indian male. To improve the component resolution and to facilitate the interpretation of mass spectrographic data, a 50-ft x 0.020-in I.D. support-coated open tubular column with Carbowax 20M as the liquid phase was employed. The column effluent was fed directly into the ion chamber at the rate of 10cc/min. During the analysis, the column temperature was programmed at 2°/min from 60-180°C. The peaks were detected on picnoammeter and timed from the instant of sample injection to the appearance of the maximum current for each peak investigated. In experiments with standard-size beds, each mass spectrographic analysis was preceded by gas chromatographic analysis of human sample collected from the same individual. This was effected by collecting samples on several beds in series. The samples were collected from a young Caucasian female.

The minimum amount of compound needed for positive mass spectrographic identification was estimated with a mixture of 1-butanol and heptanal. For this purpose, various aliquots of the mixture were analyzed under the same conditions as used for human sample. The aliquots were selected to match major and medium size peaks obtained from human sample. In each case, attempts were made to identify the selected compounds by mass spectrography.

3.8 Statistical Pattern Analyses

As the gas chromatographic data on chemical signatures of humans obtained by the fluidized bed collector technique began to accumulate, it became evident that the composition of the organic vapors from humans varied significantly from individual to individual. Visual inspection of the chromatograms from males and females did not indicate obvious common patterning (except e.g., such as a peak in that location where n-butanol should occur) nor obvious group differences. At that stage of work, patterns consisted of 32 to 48 peaks; the sizes of the peaks in the same gas chromatographic location varied within a wide range from individual to individual, and a comparison of any two gas chromatograms indicated that always a fraction of peaks could be approximately matched with respect to their positions on the gas chromatograms, while others would not match.

The factors that could explain these experimental findings were several. Some did not become apparent until high-resolution two-column gas chromatography was employed toward the end of the project work: higher resolution chromatography indicated that 60 to 100 peaks could be observed, and the two-column apparatus (Section 3.6.3) demonstrated that most of these still could be further resolved into 2 to 10 peaks, each corresponding to at least one organic component. Hence, the 32 to 48 peaks observed in analysis in packed column (Section 3.6.1) each corresponded to mixtures of several components, close in their retention times, and the position of the compound peak would be expected to shift as the proportion of the components in the compound peak would change from the individual to individual. Other causes of the peak position shifts may be the usual experimental fluctuations in the retention times, and the presence of some water in the samples. The amount of water in each injected sample was estimated at a few milligrams. The hydrogen flame ionization detector will not respond to water vapor to a significant extent, but in the Carbowax 20M analytical column the water, while

passing through the column, can temporarily increase the polarity of this stationary phase and can for short time intervals, influence the rate of migration of the components through the column. The retention times of the less polar components will slightly decrease while those of the polar components will increase.

Because of the large variety of the gas chromatographic features observed, it became necessary to develop criteria for selection of those features that would be most significant for identification and classification of human chemical signatures. At this juncture, a series of statistical pattern analysis methods were applied. In the present section, the reasons for use of the different methods will be briefly outlined; the procedures and the results are discussed in Section 4.2 and its subsections.

Procedure A

Various similarity indexes have been proposed in literature for comparisons of patterns or stimuli. In general, the indexes attempt to compare the extent of those features which are similar with those which are dissimilar, or with the total input of features. The crucial part of the procedure is the decision making on the correspondence of features. In the case of the gas chromatographic patterns, the positions of the peaks in two gas chromatograms may either match, or mismatch to varying degrees. The decision must be made as to how much mismatch can be allowed while still considering that one deals with the same feature in both gas chromatograms. A computer program was developed, designated GASCHR to compare each peak in each chromatogram with peaks in the other gas chromatograms with prescribed values of tolerance (typically 4 percent in retention time) in mismatch. The peak matching requires certain positive or negative strain if the match is not exact, and the total algebraic sum of the strain over those peaks which were matched was calculated and printed out. Usually, when approximately the same numbers and amounts of the negative were required, the sum would not differ too much from zero. Some

peaks were matchable, within the prescribed tolerances, to two peaks in the other gas chromatogram.

The peak matching generated, for each pair of patterns, a basis for calculating the various similarity indexes:

Tanimoto Index*. This similarity index is +1 when the two patterns match exactly, and zero when the patterns do not match at all (wherever there is a peak in one pattern, there is none in the other).

The Tanimoto index is calculated as follows: After matching the two patterns within some prescribed tolerance limits (e.g., in one variant of our program, peaks whose retention times in the two patterns differed less than by 4 percent were considered matched and forming an "identical pair"; if one peak in one pattern had several candidate matching peaks, within the chosen tolerance limit, in the other pattern, the peak which matched best was chosen as the partner of the matching pair), sets of peak pairs are taken to describe the correspondence between the patterns. Those peaks of the first pattern which had no match in the second pattern, or in reverse, formed pairs in which one partner was zero. The Tanimoto index S_T is the ratio of the sum of the smaller partners of the pairs divided by the sum of the larger partners of the pairs. A derivative measure of the similarity is Tanimoto "distance" between the patterns

$$D_T = -\ln_2 S_T$$

*Rogers, D.J. and Tanimoto, T.T., "A Computer Program for Classifying Plants," Science 132, 1115, 1960.

Tanimoto, T.T., "Nonlinear Model for a Computer-Assisted Medical Diagnostics Procedure," Trans. N.Y. Acad. Sci. Ser. 2, 23, 576, 1961.

Tanimoto, T.T., "A Class of Exponential Distributions and their Associated Minkowski Geometries," Notices Am. Math. Soc. 8, 432, 1961.

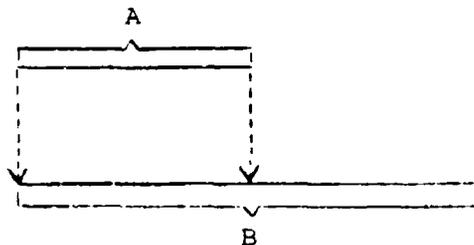
Ornstein, L., "Computer Learning and the Scientific Method: A Proposed Solution to the Informational Theoretical Problem of Meaning," J. Mt. Sinai Hosp. 32, 437, 1965.

It is zero when the patterns are identical, and infinite when the patterns completely mismatch. One of the advantages of this index is its ease of computation and a lack of bias in favor of large differences in one or a few features such as exist in the similarity indexes using quadratic terms. Since the large differences in a few features cannot a priori be assumed significant in pattern classification, it is undesirable to overemphasize their influence in forming the similarity index.

Ekman's Index. Mathematically, it is a simple function of Tanimoto index, but it is closer to the psychophysical concept of similarity presumably operative in the comparison of two stimuli by humans and developed independently of Tanimoto. Ekman and Eisler* consider that the similarity concept follows a process that is mathematically equivalent to taking the sum of those parts of attributes common to both stimuli (here we consider patterns as "stimuli") and dividing this sum by the sum of both full stimuli. Thus, Ekman's similarity index for two lengths A and B is:

$$S_E = \frac{A + A}{A + B}$$

where



This simple concept apparently suffices for reasonably good stimuli recognition by human senses.

Taking then, a bionic approach, we adapted Ekman's concept for the comparison of two patterns through the following procedure:

*G. Ekman, "Some Aspects of Psychophysical Research in Sensory Communications," Rosenlith, ed., MIT Press, 1961, p. 35.

1. Each pattern is first normalized within itself.
2. "Corresponding" peaks within both patterns are indexed by using the 4% rule. This leaves surplus peaks in both patterns. The result is a set of pairs of attributes. For those peaks that have no corresponding peak in the other pattern, the corresponding peak has zero intensity.
3. The normalized intensities of each pair of corresponding peaks are written side by side.
4. The smallest sides of each of the corresponding peaks are added; the sum is multiplied by two.
5. All normalized intensities for both patterns are added.
6. The result of step 4 is divided by that of step 5, and gives S_E (Ekman's similarity index). It will be small (approaching zero) if the similarity is small; it will be +1 for exactly similar patterns.
7. The similarity index, S_E , is converted to a distance value by taking $-\log_2 S_E$. For identical patterns, the "distance" is zero. For highly dissimilar patterns, the distance, D_E , is a positive number exceeding 1, 2, or more.

Correlation Coefficient.

This is the well-known classical measure of correlation between two sets of observations, or, in the case of pattern comparisons, of two patterns at a time, aligned so that the peaks match in their retention times within some prescribed tolerance limits.

The classical correlation coefficient (very related = +1); unrelated = 0; oppositely related = -1) also gives intuitively correct values for distances between points in the correlation space, but is more difficult to compute. Also, it can give a value of +1 for patterns in which additive and multiplicative differences compensate, while patterns are intuitively dissimilar.

In the statistical experiments with the similarity indexes, additional variations were: Tanimoto index averaged over peaks, Ekman index summed over and divided by the number of pairs. To minimize the influence of the quadratics of the correlation coefficient, a correlation coefficient applied to normalized area X transformed logarithmically

$$y = \log (x + 0.02)$$

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was included, in addition to the correlation coefficient applied directly to the normalized areas*.

The rationale behind the study of the similarity indexes was the possibility of developing a taxonomy (classification) of the patterns. If a third pattern has high similarity index to each of the two patterns which also have a high similarity index, these three belong to the same class. Otherwise, a new class can be set up and each next pattern tested for belonging to one or the other class etc. Classes can also branch, cf. Figure 13. Here pattern A has some similarity to B (2 of 5 features match, $S_T = 0.4$), C has more similarity to B (3 of 5 features match, $S_T = 0.6$) but A and C are completely dissimilar (none of the features match)**. Thus interrelation between classes can be gradually developed, and basically must use non-Euclidian spaces for hypergeometric representations (in Euclidian space, if A is close to B and C is close to B, A and B cannot be infinitely far apart).

Although the taxonomy development using similarity indexes indicated that vapors from similar sources in average tended to produce patterns with higher similarity indexes, the indexes per se were low indicating that much extraneous noise (non-essential features) interfered with the classifications. Therefore other methods were applied.

Procedure B

In this procedure, a more mechanical approach was taken to reduce the patterns to a definite number of variables to which

*During this stage of work, the normalized areas were taken as the areas divided by the mean area of peaks in the particular pattern. In later work, second part of Procedure D, the normalization was by dividing the peak areas by the total area of all peaks in the respective pattern.

**Tanimoto distance between Tanimoto units B and C is $-\ln_2 (0.6) = 0.73$.

then stepwise discriminant analysis program was applied. The chromatograms (initially in terms of retention times normalized to n-butanol, later in terms of Kovats Indexes) were divided into ranges of equal width, (or percentage width), disregarding the initial and the last portion of the gas chromatograms. Peaks with the position of the peak in the particular interval were considered to be, with their full area totally in this interval. If several peaks occurred in the same range, their areas were summed and the sum became the intensity coefficient for this range in the respective gas chromatogram. Thus, a "variable" was the total content of organic substances eluted during the particular range, with peaks in this range. By increasing the width of the ranges, the number of range boundaries could be decreased, reducing the number of cases when the same substance eluted close to the boundary could occur in one or the adjoining cell.

After reduction of the chromatograms to a set of number of variables stepwise discriminant analysis was conducted, using BMDO7M Stepwise Discriminant Analysis Computer Program, Version of September 1, 1965, Health Sciences Computing Facility, University of California, Los Angeles.

Procedure C

This procedure also used stepwise discriminant analysis, but instead of selecting the variables by mechanical subdivision into ranges, a histogram study was conducted to find those gas chromatographic locations (in terms of Kovats Indexes) in which peaks most frequently occurred in the chemical signatures of humans. The variables were then formed by putting the center of the range at the maximum frequency point and allowing ± 10 Kovats Index units as an estimated tolerance for the two boundaries of the same range. Thus, in this procedure, the variables were histogram-guided ranges, not necessarily with common boundaries scattered throughout the gas chromatograms. Again, areas of all peaks with the apex in the range formed the intensity value for

the particular variable.

Procedure D

This procedure was a further development of the Procedure C, and differed simply by introduction, into the stepwise discriminant analysis, of the "non-peak" ranges, between the ranges where peaks were frequently observed in humans.

The above various procedures were applied to all peaks, or, in some trials, only to peaks within certain peak size classes, (e.g., only large peaks, above certain size, etc.)

The stepwise discriminant analysis looks for those features which are most significant in discriminating two or more groups of patterns, and forms discriminant functions which are linear equations of the form:

$$X = a_1 V_1 + a_k V_k + \dots$$

Here a_1, a_k etc. are coefficients derived by the computer program, while V_1, V_k etc. are the gas chromatographic variables. The V 's were not taken as the direct sums of peak areas within the particular gas chromatographic range but were logarithmic transforms for the corresponding areas, so as to give more equitable weights to large and small peaks. In the Procedure D, the transform was, e.g.,

$$V_{34} = \sum_{1112}^{1132} \left[1 + \log (A_i + 0.1) \right]$$

where 34 is the "name" of the range; 1112 and 1132 are its boundaries in terms of Kovats Indexes; A_i is the peak area of a peak within this range, taken in terms of its area as percent of the sum of areas of all peaks in the same pattern. If i was the only peak in the range, the expression in the square brackets became V_{34} . If there were several peaks in the range, V_{34} was the sum, as indicated, of the logarithmically transformed peak areas.

Discrimination means that a function X, or several functions (X,Y) are found such that their values for all patterns of one class, (e.g., humans) are distinctly different from the values for another class (environments).

These functions can then be used to discriminate one group of patterns from another. The same principle applies to discrimination of several groups of patterns.

In the stepwise discriminant analysis the success of the procedure can be judged by two criteria. If the number of variables used is small compared with the number of patterns used (much less than by a factor of 2), and function that separates the patterns in two (or more) groups is found, the probability that this is a valid discriminant function is high. The probability increases as the number of patterns is increased and the number of variables needed to classify decreases.

The other criterion is more rigorous. If the discriminant function is obtained by using only a part, e.g., one half of all of the patterns (each pattern used fully), its validity can be tested using the values of variables from the other half of the patterns. If the other half of the patterns is satisfactorily classified into its proper group, the probability is high that the variables selected by the discriminant analysis constitute truly differentiating pattern class features.

3.9 Subject Selection and Preparation

The signature collection experiments were performed on human subjects of both sexes and various races. Except for two middle-age Caucasian males, all subjects were young, healthy specimens. Before the participation in the experiments, the subjects did not use any cosmetic preparations except for neutral soap, and lived on normal mixed diets. To expose the maximum area of skin to air flow the subjects were closed in the cleaned glass system partially naked.

The selected human races included American Caucasian males

and females, Indian males, American Negro subjects, and Mexican and Malayan subjects.

The majority of the Caucasian males were students at the Illinois Institute of Technology in Chicago. The student group included two pairs of brothers. The rest of the Caucasian males comprised two chemists, one free-lance writer and one chemical laboratory technician.

The group of Caucasian females was composed of young office workers employed at IIT Research Institute, house-wives and a few high-school students. This group included a mother and her two daughters.

The Indian group consisted of young males born in India with temporary residence in the United States. All members of this group were students at the Illinois Institute of Technology in Chicago.

The American Negro group included one young male chemist and two young females. The male subject was a husband of one of the females.

The last group included a young Mexican house-wife born in the United States and a young Malayan male. The male subject was also a student at the Illinois Institute of Technology in Chicago.

Other sources of chemical signatures consisted of kitchen and laboratory air and tropical vegetation. The collection of these signatures were performed by means of the portable air sampling apparatus. The kitchen air samples were obtained from the IIT Research Institute Cafeteria kitchen; also the laboratory samples and machine shop samples were taken at IITRI.

The chemical signatures from tropical vegetation were collected in the Palm House of the Lincoln Park Conservatory in Chicago. The collection environment included various types of palm trees, such as sentry palms (Howea forsteriana); fishtail palms, a native of India and the Malay region, (Caryota mitis); and scattered through the underplanting of Fittonia, a

number of ornamental palms, such as Fiji Island fan palms (Eupritchardia pacifica), silver palms (Coccothrinax argentea), and roebelen palms (Phoenix roebelen). There is also a 50-ft fiddleleaf rubber tree (Ficus lyrata) from Africa, a tapioca plant (Mauihot exculenta), a fig tree (Ficus carica), and the common banana (Musa sapientum). The Conservatory also houses flowerless plants such as ferns and mosses. The temperature and the humidity of the Palm House environment range from 60 to 85°F and from 50 to 75%, respectively.

4.0 RESULTS AND DISCUSSION

4.1 Packed Column Data

The experimental data on human chemical signatures of both sexes and various races as well as of different environments are tabulated in the form of copies of computer printout in Appendix I.

In the tabulation, each pattern is identified by the proper number, the signature source ("subject": initials of humans), the date of signature collection, the total intensity and the mean peak intensity.

The arrangement of patterns follows approximately the chronological order of signature collection experiments and is numbered accordingly.

The total intensity is defined as the total area, in sq. mm., of all gas chromatographic peaks of the particular pattern and the mean peak intensity as the total intensity divided by the number of peaks in the same pattern.

The data on each pattern tabulated in Appendix I includes also consecutive numbers of peaks in the pattern, the retention times of peaks in terms of Kovats indices, the peak areas in sq. mm. (calculated to the electrometer sensitivity setting of 0.1×1) and the peak normalized areas (peak area divided by the total intensity).

The gas chromatographic patterns of the signatures studied reflect complex and only partially resolved mixtures. The number of detectable components observed in human signature patterns ranged from 32 to 48 with many indication for the presence of a large number of additional unresolved peaks. The magnitude of the collection samples ranged from 2.6×10^{-7} g for an Indian male to 2.1×10^{-5} g for a Caucasian male.

The total number of detectable components in the environment chemical signatures ranged from 34 to 46 with a strong evidence for the presence of many unresolved peaks. The magnitudes of the collected samples fell within the range of 1.9×10^{-6} g and 1.1×10^{-5} g.

It must be understood that each peak in the tabulation can represent more than one substance and is resolvable, as later work with the two-column device showed, into several peaks when examined in a non-polar Apiezon L column.

The peak areas are approximately proportional to the sum of weight of the components represented by the peak. Table XI lists the relative amount of area contributed per atom of the respective element*.

Since the number of heteroatoms compared with the number of carbon atoms in common compounds with reasonable molecular size is small, the area of multicomponent peaks can be assumed to be approximately proportional to the weight of material it represents.

The peak areas in the Appendix are quoted in mm^2 , taken at the electrometer attenuation of 0.1×1 . For the particular instrument, the calibration factor was approximately:

$$1 \text{ mm}^2 = 0.2 \times 10^{-12} \text{g}$$

The peak areas represent the amount of the organic material within the peak range injected into the chromatograph. The amount present in the collected sample is estimated, from the efficiencies of sample transfer and injection to be 30 percent higher.

The concentration of the respective materials in air from humans could be calculated only if the calibration factor (partition coefficient) was available for the particular molecular species: Table XII represents data on the emission rates of n-butanol from humans. This substance formed a recognizable peak used for internal calibration.

*Johns & Sternberg, Gas Chromatographic Detectors, in Instrumentation in Gas Chromatography, p. 179, Krugers, ed., Centrex Publishing Co., Eindhoven, Holland, 1968.

For unknown substances, very approximate calibration is possible provided that Kovats Index of the unknowns in Apiezon L is known and taken to be, at room collection temperature, the same as at elevated temperatures. The amount G of a substance X collected is

$$G = k \cdot m \cdot v$$

Here k is the partition coefficient of the substance in Apiezon L/Air system at the collection temperature (ambient); m is the concentration, in grams/cm³, of the substance in air, and v is the volume of Apiezon L in the collector. A plot of log k versus the Kovats Indexes of the substances in Apiezon L is a straight line and can be used for approximate estimates of k; v is known, and m can be calculated when G is known from the area of the gas chromatographic peak. The value of k increases by a factor of approximately 2.3 per each 100 units of Kovats Index increase. The emission from the human subject is (750 . . .)/sec when 750 cm³/sec is the rate of airflow through the glass cell from which the human subject vapors are collected.

These remarks are to indicate that in the equilibration sampling (fluidized bed, Section 3.2) approximate calibrations can be obtained even for unknown materials, from their Kovats Index positions in Apiezon L. Since the Apiezon L Kovats Indexes were obtained in the two-column device (Sections 3.6.3 and 4.4) where the first column was a 200-ft open tubular Carbowax 20M column, the areas under peak in the packed Carbowax 20M column cannot be directly related to the high-resolution runs which resulted from the use of two-column device. The packed column analyses were to assist in finding those gas chromatographic regions where the significant features reside. It was more important to concentrate adequately all organic vapors including those with a low volatility, so that they would be detectable at feasible sensitivities and to analyze these in Carbowax 20M column where better peak shapes are obtained for all polar components, including alcohols which chromatograph

poorly on Apiezon L. The use of Carbowax 20M in the fluidized bed collectors was not practical since the sample collected too much water which then interfered with the analysis.

4.2. Pattern Classification

The various approaches to this task were discussed in Section 3.8. The objective of the pattern classification was twofold: (1) to obtain evidence that the complex chemical signatures, in the form of gas chromatograms of vapors from humans and environments, contain features useful for their identification as to the type of their sources, and (2) to indicate those regions in the gas chromatograms that are most significant for the source discrimination and therefore merit first attention in further chemical identification of the components.

4.2.1. Peak Matching Studies

Some pattern comparisons were conducted using simple rank order statistics on gross indexes such as the number of peaks per chemical signature. Here three more or less homogeneous groups of humans on which a sufficient number of chemical signatures were available were used: 12 males from India, 25 Caucasian American males and 23 Caucasian American females.

The average numbers of peaks and the standard deviations in the numbers of peaks (in parentheses) in the chromatograms submitted to the peak matching procedures were as follows: Indian males 41.2 (1.8); Caucasian males 37.3 (2.5); Caucasian females 40.0 (4.2); environments 37.0 (6.1). Wilcoxon-White rank order test applied to the number of peaks per gas chromatogram as the ranking criterion reached 99 percent confidence level (less than 0.01 probability to obtain such differences by chance) when Indian males were compared vs Caucasian males, and when Caucasian males were compared vs Caucasian females; the differences in the peak numbers between Caucasian females and Indian males did not reach 95 percent confidence level.

Further comparisons included data inspection after peak matching.

Most of the pattern comparisons for the purpose of obtaining similarity indexes were conducted after matching of peaks between two patterns at a time, allowing for a ± 4 percent tolerance of retention time normalized to n-butanol after addition of 0.02 to the normalized retention time. The 0.02 term was significant only at the initial time range of the gas chromatographic patterns.

The selected matching criterion was apparently rather liberal, since the number of peaks that could be matched within each pair of chemical signatures taken was in excess of 80 percent of peaks observed in each of the patterns of the pair. To assess if at this tolerance level gross differences existed between members of the same group versus the numbers of the same or other groups, seven randomly selected patterns were chosen within each of the three groups and compared with others. The following observations resulted: from the average number of 37 peaks in 7 Caucasian male patterns, 85 percent found matches in the patterns of other Caucasian males; from the average number of 40.7 peaks in 7 Indian male patterns, 82 percent found matches in the patterns of Caucasian males; from the average number of 39.6 peaks in 7 Caucasian females, 84 percent found matches in the patterns of other Caucasian females. These initial observations did not indicate existence of gross differences in the match frequencies within the group and between the groups.

The relatively high density of the coverage of the gas chromatograms with peaks is one of the factors which may account for this initial result. If the allowed match tolerance is too broad, there will always be a high probability of finding a matching peak. If the tolerance is too narrow, experimental variability of the retention times for the identical substance may be large enough to miss the tolerance limit and give "no match" although actually a perfect chemical identity match existed.

Consequently, more elaborate pattern similarity indexes were explored in which consideration was given not only to match versus nonmatch in the peak positions but also to the magnitudes of the chromatographic areas which represent the abundancies of the components eluted in the respective gas chromatographic range.

The merits of two similarity measures--Tanimoto distance and the correlation coefficient (cf. Section 3.8)--applied to patterns matched to 4 percent tolerances were explored on a part of the data available at the time when the computerized GASCHR program became operational.

The Tanimoto distance in a non-parametric hyperspace is zero for two identical patterns, large for dissimilar, and infinite for completely non-matching patterns in which no peaks in one match the other pattern's peaks in retention times within prescribed tolerances.

The test group consisted of the chemical signatures of 12 Indian males, 10 Caucasian females, and 9 environments, (2 plants, 2 laboratories, and 5 kitchens). The results indicated that considerably variations occurred within each group and that very few Tanimoto distances were below 1.0. However, the average distances for pairs within each group and for intergroup pairs were consistent, and patterns for humans were clustered, in comparison to the average Tanimoto distances between chemical signatures of humans and environments.

Indian males versus Indian males	1.561	(66 pairs)
Caucasian females versus Caucasian females	1.817	(45 pairs)
Indian males versus Caucasian females	1.809	(120 pairs)
Indian males versus environments	2.094	(108 pairs)
Caucasian females versus environments	2.024	(90 pairs)

This tabulation shows that (1) the Indian male group was most consistent within itself (2) the variability among Caucasian females was larger and approximately as large as between Indian males and Caucasian females, and (3) the difference between human and the environmental chemical signatures was larger.

Correlation coefficients applied to the logarithmically transformed peak areas after 4 percent tolerance match showed

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similar relations. The largest cohesivity occurred within the Indian male group.

When patterns were assigned to human versus environment groups by comparing the average Tanimoto distances from the pattern in question to 26 human patterns and to 9 environment patterns correct assignment resulted. Pattern assignment to certain groups of humans on the same comparison basis was inconsistent. It was concluded that if the Tanimoto distance or a similar concept is used, the number of comparison samples should be increased, or some pre-classification of peaks into useful and nonuseful should be conducted.

The similarity indexes for patterns within the same group were relatively small - 0.3 to 0.5, rarely reaching above 0.5. Many mismatches in sizes and appearances of spurious peaks corresponding to occasionally occurring components in vapors from humans lower the index with the result that many Tanimoto distances were above unity and were not good classifiers.

Thus, the pattern classification approaches based on similarity indexes did indicate the possibility of classification. However, it was felt that discriminant analyses where the emphasis is not on the measures of similarity but on the features most useful in class separations may be a more powerful tool for the cases when the classes to which the various patterns belonged were already defined in terms of the vapor sources. Similarity indexes are more specifically suited for developing taxonomies when the observations are not easily assignable to classes, but classes and subclasses have to be uncovered in poorly structured data.

Consequently, later work was concentrated primarily on using discriminant analysis as the classification tool.

4.2.2 Stepwise Discriminant Analyses-Arbitrary Variables

For the stepwise discriminant analyses (cf. Section 3.8) when the chemical identity of the components is uncertain it is

necessary to subdivide the gas chromatographic patterns into a fixed number of ranges serving as variables (vectors), so that each pattern can be expressed in terms of the same number of variables. If each range contains not more than one peak, the intensity value for the particular variable can be the normalized area, or some function of it, of the peak. Experience indicated that if the ranges are small enough to contain either none or very infrequently more than one peak, many cross-overs can occur in a certain range, or in the neighboring range while representing the same component, since the position of a peak can shift by several reasons (cf. Section 3.8).

Initially, 65 ranges were set up, with the width of each range corresponding to 6 percent of the retention time corresponding to the ranges lower retention time boundary; e.g., for the range beginning at 10.0 minutes the boundaries would be 10.0 and 10.6 minutes, respectively. This partitioning resulted in 65 ranges. After preliminary runs, with 49 patterns available at that time in which the retention times were normalized with respect to the n-butanol retention times, 21 adjoining (4 with retention times smaller than that of butanol and 16 with retention times larger than that including the butanol) were selected, since (1) the number of patterns presented for analysis must exceed the number of variables, (2) the retention times of peaks around the butanol peak were more reliable when relative to butanol, and (3) preliminary analysis showed that the majority of peaks useful for distinguishing various types of humans seemed to be in this region.

By using a correlation coefficient matrix, the statistical process extracted successive significant variables, beginning with those which produce the most efficient separation between prescribed groups of patterns. It continued to extract the variables (peaks) in order of their decreasing significance. After each variable had been handled for all patterns, the efficiency of classification based on all peaks extracted up to this point

was evaluated. This process was applied to the 49 patterns grouped in 5 groups: 20 Caucasian males, 12 Indian males, 10 Caucasian females, 5 various humans of other types, and 2 tropical plants.

To compare the efficiency of the classification process by these logical groups, a random classification of the same 49 patterns was also subjected to analysis. Figure 14 shows that after classifying 13 variables, only 3 patterns, or less than 9%, were misclassified when proper groups were used and that 15 patterns, or over 30%, were misclassified in the randomized groups. The partial classification structure in the randomized group arises from the statistical probability that a pattern can be assigned to the proper groups by chance and from the presence of possible other classifiable features that can occur in the gas chromatograms.

Subsequently, the range of the gas chromatogram included in patterns for the stepwise discriminant analysis was expanded and the width of each range representing separate variables was doubled. The purpose of this change was to include more information and reduce the probability, for the peaks close to a range's boundary, to cross over the boundary into the adjoining cell. The range of retention times between 0.327 and 5.73 (relative to n-butanol) was subdivided in 21 ranges, with n-butanol being in the 8th range. The rear boundary of each range was equal to the front boundary's retention time plus 0.02 plus 12 percent of the front boundary's retention time. The intensity of the range was equal to the sum of areas of all peaks having apexes within the range; the actual intensity was transformed logarithmically, with conventional linear increments, to form the intensity vector for the discriminant analysis.

At that stage, three statistical experiments were conducted. In one, discriminant functions were formed using 27 human chemical signatures and 5 chemical signatures of air from a cafeteria kitchen area taken at different times. The separation in terms of two

discriminant functions, X and Y, after computer use of 6 out of 21 variables, is shown in the top part of Figure 15. The derived discriminant functions were used to compute X and Y for additional 24 human chemical signatures. The values obtained were plotted over the repeat plot of the top portion of Figure 15, in the bottom part of Figure 15. Reasonably successful classification of additional signatures was observed with 15 out of 24 falling into the "range circle" somewhat arbitrarily drawn, and several more closer to the center of gravity for the human patterns than to the cooking vapor pattern center.

The second experiment is represented in Figure 16. Here 37 chemical signatures were classified in three groups: 20 Caucasian males, 12 Indian males (students from India, mostly vegetarians) and again 5 cooking vapor samples, using up to 14 variables.

The third experiment consisted of classifying chemical signatures in two groups: 16 females and 35 males. The classification after using 7 variables is shown in Figure 17. This experiment is significant since here only the chemical signatures obtained in the glass cell apparatus (cf. Section 3.1) were used, and the possible question on discriminant features introduced by sampling humans in the cell versus sampling environments in air at large does not arise.

At this stage of the work, normalization of the retention times in terms of Kovats Indexes was introduced (cf. Section 3.6.5) and it appeared desirable to subdivide the chromatogram into Kovats Index increments for the purpose of the stepwise discriminant analysis. This index is a logarithmic function of the retention times, hence equal index intervals approximate equal retention time percentage increments.

One set of the statistical tests was conducted using arbitrary division of the chromatogram into ranges 50 Kovats Index units wide. Thus, 22 ranges were set up: one with K.I. below 600, then 20 ranges each 50 units wide, and the last range with K.I. above 1600. This converts each chromatogram into a set of 22 variables; the intensity

of each variable is equal to the sum of logarithmic transforms of all peak areas with the apexes in the given range. Two discriminant analyses were conducted. One used three groups: 39 males, 26 females, and 10 environments. The other used human signatures, obtained in the glass cell only, and into 3 groups: 25 Caucasian males, 12 Indian males, and 26 Caucasian females, 63 humans in total. Tables XIII and XIV illustrate the success of classifications, in terms of percent of patterns properly assigned to the correct group. Discriminant functions based on 13 variables produced more than 80 percent of correct assignments for the test containing 63 human chemical signatures. Most of the classification was achieved at taking 5 variables only. It can be noted that sometimes upon inclusion of further variables (cf. variables Nos. 3 and 4, female column Table XIII) the percentage of correct assignments can decrease. This effect results from compromises when optimization of group separations leads to a larger deviation from the group center point for an occasional case.

All these experiments conducted by mechanical separation of gas chromatograms into range variables indicated that discriminating features can be found and reasonably successful functions formed after using only a fraction of the variables, but that 5 to 15 variables may be needed to sufficiently define the chemical signature source if the identification is based on vapor gas chromatograms consisting of 30 to 50 peaks.

At this juncture it was decided to explore if a more selective choice of variables (gas chromatographic ranges) can result in better and more valid classifications, or help to pinpoint, for the future gas chromatographic and mass spectrographic identification those narrower ranges where significant discriminating features reside.

4.2.3 Stepwise Discriminant Analysis With Selectivity Chosen Variables

In this part of the statistical experiments, attempts were made to use several rationales in selecting gas chromatographic ranges to serve as variables in the input to the stepwise discriminant analysis program, instead of mechanically dividing the entire gas chromatographic range into a set of adjoining ranges, as was done in the work described in Section 4.2.2.

Histogram Procedure. Here data on the positions of gas chromatographic peaks expressed in Kovats Index units were pooled for all 65 patterns of humans together, or for males and females separately and arranged in the order of increasing Kovats Index. Wherever peaks occurred frequently in the human chemical signatures, the number of peaks, e.g., per 10 Kovats Index units was higher than elsewhere. Figure 18 represents the histograms of the frequency of occurrence of peaks in Caucasian males and females along the Kovats Index scale.

These histograms can be considered as generalized gas chromatographic patterns, with the peaks indicating those ranges where peaks of components most commonly occur; the peak areas of the original gas chromatograms are not indicated by this plot, except that only those original peaks were considered (about 2/3 of all peaks) which were larger than 40 percent of the average area of peaks in the corresponding pattern. There are striking similarities between the occurrence of peaks with the Kovats Index values in the two generalized patterns, but also distinct differences; for instance, substance with K.I. = 1030 (asterisk) much more frequently appears at larger relative concentration in effluents from females than from males. The peak areas are not considered in the histograms.

More histograms were prepared using an available computer program in which the pooled and ordered Kovats Index array is scanned, taking each peak in turn and calculating the Kovats

Index interval between two peaks, the first of which is, e.g., 10 peaks before *i* and the second is 10 peaks after *i*. Wherever this interval is smaller, the peaks occur more frequently. An example of a part of such printout, which is in essence reciprocal to Figure 18, is shown in Figure 19. The points of minima represent those locations on the Kovats Index scale where peaks occurred more frequently.

Selection of Variables. By inspection of a histogram for the entire gas chromatographic range of 65 pooled human chemical signatures, 37 locations were found where the peaks occurred most frequently, that is the minima in the histogram such as shown in Figure 19 were most pronounced. The variables were set to center on the minima and to have a range of ± 10 Kovats Index units around the selected minima. All peaks with apexes in the K.I. range were considered to be fully within the range. Thus, the chromatograms were each converted to 37 variables; cf. Section 3.8 for the intensity transformation procedure used to assign the intensities to the variables. If the center points of two adjoining ranges were closer than 20 K.I. units, the boundary between the ranges was set at the middle point between the centers. This selection left 15 to 20 possible high frequency-of-occurrence ranges not included.

The 37 ranges, called P-ranges (peak-containing) were adjacent at some locations, but separated by ranges in which peaks in the chemical signatures of humans were less common. Table XV lists all types of ranges, consequently numbered, with indication of the range widths and the type of range (P or non-P). The statistical discriminant analyses were conducted using various choices of variable sets. Thus, all 37 P ranges, or 10 of P-ranges, or the non-P ranges were tried; finally, 34 variables were selected, shown in the last column of Table XV, which included 18 P-ranges shown to be most discriminating between human and environment chemical signatures and 16 non-P ranges, also shown.

among the non-P ranges, to be most discriminating. The consideration of non-P ranges was made upon the suggestion of personnel of the Ballistics Research Laboratory who pointed out that the pattern discrimination should be aided by considering, in addition to the presence of certain features, also the lack of other features.

Stepwise Discriminant Analyses. Selected typical experiments are summarized here.

Experiment 1. Discrimination of 65 Human from 10 Environment Chemical Signatures Using P-Ranges Only. Table XVI summarizes the efficiency of classification.

In run A, all 37 variables were submitted; in run B, those ten variables ranges were selected from the 37 ranges where the peaks most frequently occurred in human chemical signatures. It is seen that in test B, the computer needed to use only five ranges as variables to separate 91 percent of the patterns from humans into one coherent group. In test A, 94 percent coherence was accomplished after the use of 11 ranges out of 37, with no improvement beyond that. After the extraction of 17 variables out of 37, the computer reported that no information that would be statistically useful for group separation was detectable in the remaining 20 variables. The degree of coherence in classification reached by this technique of the selection of variables was considerably better than that obtained by a division of the patterns into ordered ranges without regard for the inner logic of the patterns. On the basis of data presented in Table XVI, component identification can be centered on the most significant ranges. It must be noted that the important ranges can include several peaks so that a decision must be reached, by the use of higher resolution columns, as to which are the important peaks within the important range.

Experiment 2. Discriminant Analysis of Human Versus Environment Chemical Signatures Using P and non-P Ranges. Here two runs were made. In Run C, 36 human chemical signatures were selected at random from 65 available signatures, and all 10 environment chemical signatures; the discriminant analysis was conducted using all P-ranges only (marked P in the "Range Width" column of Table XV). In Run D, the discriminant analysis was conducted using all other, non-P-ranges only. Figure 20 compares the relative efficiency of both selections.

Classifications based on either set of variables were eventually comparably successful. The classification based on those Kovats Index ranges in which peaks were frequently observed in samples from humans was more efficient when a smaller number of variables was utilized.

Experiment 3. Derivation of Discriminant Function Using Part of Data and Test of the Function Using the Rest of Data. Here a new selection of the variables (34 in total) was made. It included the most statistically significant variables both from the Kovats Index ranges in which peaks frequently were observed in the vapors from humans (P-ranges) and the intervals between these ranges. The selected variables are marked by crosses in Table XV with the P-ranges also marked with P. Two runs were selected for the report.

In Run E, 36 human chemical signatures, randomly selected from a total of 65 available, and all 10 environment chemical signatures were used to derive the discriminant function. Then the same discriminant function was applied to classify the remaining 29 human chemical signatures. Figure 21 shows that only 4 out of 29 or 14 percent were misclassified if the discriminant function based on 15 variables (computer-selected from 34) was used. The variables were, cf. Table XV, Nos. 18, 30, 35, 37, 44, 47, 49, 54, 55, 56, 57, 59, 61, 63, and 65.

If the discriminant function derivation was stopped at a

fewer than 15 variables, class separations were somewhat poorer:

<u>Variables Included</u>	<u>Percent Correctly Classified in Testing The Discriminant Function</u>
2	71
5	74
15	86

In Run F, a three-way classification was tried. The discriminant functions X and Y were derived for chemical signatures of 26 females of several races, 22 males of several races, and 10 environment samples, or 58 patterns in total; 15 additional male chemical signatures were kept if the additional males were correctly classified by the derived function.

The male group used for derivation consisted of 14 Caucasian and 8 Indian males; the male group withheld for testing contained an additional 11 Caucasian, 2 Negro, and 4 Indian males. The progress of classification during the function derivation is shown in Figure 22. After use of 20 variables out of 34 variables available to the computer, the statistical noise level was reached, with 4 females still not correctly classified. Figure 23 shows the classification field in terms of two discriminant functions X and Y. The hollow triangles represent the male chemical signatures used in derivation. The filled triangles represent the male chemical signatures used to test the functions. Although an intrusion into the environment ("Airs") chemical signatures region occurs, the test points all are far away from the region of the field populated by the female chemical signatures. This experiment again indicates that classifiable features exist in the chemical signatures of males and females.

Other Statistical Experiments. A number of other class separations was tried, e.g., Caucasian males versus Caucasian females versus Indian males, and some four- and five-class separations. None of these was 100 percent successful, but varying lower percentages of success were achieved.

The pattern stepwise discriminant analysis led to the conclusion that under conditions where the chemical signatures are expressed in terms of up to 30 variables, reasonable classifications by the classes of sources (sex, race, humans versus environments) becomes possible with use of 2 to 20 variables. The required number depends on the similarity between the sources; in the case of humans versus environments as few as two to five variables suffice to effect 80 percent levels of success. The derived discriminant functions held reasonably well under tests with additional chemical signature patterns.

The results also indicated those Kovats Index ranges which contain components most useful for the source discrimination and therefore most suitable targets for further chemical identification.

4.3. Three Column Data

The three-column gas chromatographic device (Section 3.6.4) produces readout which can be analyzed in a two-fold manner.

Three Point Match. For the compounds for which the three time points of exit (from the 200 ft Carbowax 20M column and subsequently the short and the long Apiezon L columns) have been established, an inspection of the three gas chromatographic recording tracks permits judging if these compounds are present in the sample of chemical signature. As in any judgement based on gas chromatographic retention times in several stationary phases, there are two types of errors possible: (1) the location of the peak of a known compound can be shifted because of other materials present, so that a slight non match does not rule out the presence of the compound and (2) other compounds can produce peaks in closely similar locations so that a slightly non-matching point can be erroneously taken to represent the particular compound. In the case of a three-column device, with a continuous flow of the components through the system, additional erroneous judgements

are possible. "Coincidence ghosts" can arise if a three-point match, or near-match, results from simultaneous presence of several components; one component may leave the Carbowax 20M column at the same time as the known compound, while other components which left this column at different times can leave the Apiezon L columns at the time points expected for the known compound. These errors can be reduced if the known compound can be added to a duplicate sample and the sample reanalyzed; if the peak in all three points shows no indication of broadening or splitting, the evidence that the known is present in the unknown is very considerably strengthened, while if the peak broadens or splits, errors of coincidence or presence of additional gas chromatographically similar components are likely.

In Kovats Index system, approximate prediction of the gas chromatographic positions of compounds is possible from rules operative in this system, (cf. Section 3.6.5).

Table XVII lists some three-point matches obtained by comparisons of chemical signatures of humans with the three-point locations of known compounds run through the three-column device independently.

The three-point process, if it is to be utilized either for the components pre-identification or as a part of pattern analysis, must be studied to establish: (1) the experimental reproducibility of the positions of the same compounds in the context of various mixtures, (2) the probability of occurrence of other compounds that can produce, each completely by itself, similar sets of three time points, and (3) the probability of coincidental production of a similar three-point set, by two or three different components.

Computerized Indexing. Here, peaks are matched without reference to known compound points, except that an n-alkane calibration scale is used to convert the readings to Kovats Indexes. A program GA3COL, was written for IBM 360 computer to process the three-column gas chromatographic data and print out the most

probable sets of Kovats Indexes in Carbowax 20M and Apiezon L for the components indicated by peaks.

Each peak emerging from Carbowax 20M column enters both Apiezon L columns simultaneously. The ratio of the corrected retention times on the long: short Apiezon L columns is approximately constant. After calculating this ratio for each Carbowax 20M peak as the possible origin and for the various combinations of Apiezon L peaks within the gas chromatographically feasible retention time range and applying the ratio \pm tolerance criterion, the computer prints out those peak sets that meet the criterion, with the Kovats Index data calculated on the basis of calibration with n-alkanes.

The ratio was explored using known compounds. Its average was 2.05. The critical point is the tolerance allowed. If the tolerance is too small, some components actually present in the sample can exhibit ratio which because of normal experimental variations would be outside the limits and would not appear in the tabulation. If the tolerance is too broad the frequency of mathematical coincidences meeting the limits will begin to increase. On the basis of preliminary experiments tolerance of ± 0.10 was judged reasonable. The computer program can be run with any selected tolerance.

Figure 24 illustrates the data obtained by the computerized analysis of the three-column gas chromatograms of vapors collected from the same subject simultaneously. The figure combines two analyses, at 90° and 150°C. It indicates the utter complexity of the composition and the possible presence of groups of compounds indicated in the upper right corner.

Later work with the two-column system (Sections 3.6.3 and 4.3) confirmed the complexity of peaks eluted from the 200-ft Carbowax 20M column. Since the two-column data were easier to interpret, and a more detailed study was desired in the gas chromatographic ranges that were significant in pattern

classification (cf. Section 4.2.2 and 4.2.3), the work with the three-column system was set aside in favor of that with the two-column system. The limited experience with the three-column system operated isothermally led to the following observations: (1) the system, in principle, can be useful for a rapid overview of new patterns (2) since small errors in short retention times strongly influence the ratios of the retention times, the matching of peaks at the beginning of the gas chromatograms, and for polar components with short retention in Apiezon L throughout the chromatogram, is less certain, so that components actually present can be missed if tolerances there are set too narrow, (3) towards the end of gas chromatograms, resolution increases to the extent that several peak combinations can reasonably meet the matching criterion.

It is evident that the ease of interpretation of the three-column data significantly depends on the number of components present in the sample and on the resolution of columns used. Human chemical signatures exhibit complexity that limits a straightforward interpretation of isothermal analyses.

4.4 Two-Column Data

The data on tentative identification of selected components of human chemical signature acquired by means of two-column apparatus are displayed in Table XVIII. The data were derived from the plot in Figure 25. The Carbowax 20M peaks examined by this technique were arranged in order of increasing Kovats Retention Indexes and numbered accordingly. The selection of peaks to be examined was based on Kovats Index Ranges isolated by the computer program. The ranges comprised peaks with maximum occurrence frequency in chemical signatures of all subjects previously studied. The computer-isolated ranges together with the number of peaks observed in the ranges are given in the second column of Table XVIII. Except for Carbowax 20M peaks 1 and 3, all peaks examined fall within the selected ranges:

peak 1 precedes the range by only 7 Kovats Index units and peak 3 by 12 units.

Since the pattern analysis was based on gas chromatographic data obtained with packed partition column while Table XVIII is based on data from the 200-ft open tubular column with much higher resolution, an exact correspondence between peak positions in the two sets of data is not to be expected.

The twenty-five parent peaks selected in elution from the first (200-ft Carbowax 20M) column resolved, as Table XVIII indicates, into over 150 components on re-analysis in the 50-ft Apiezon L column. The number of components per peak ranged from 3 to 10. Contribution of these components to the parent peak varied over a broad range, but usually one to three daughter components could account for the most of the parent peak area. Thus, the 25 parent peaks resolved in the second (Apiezon L) column, into 36 major components and approximately 120 minor components. The major components are marked by asterisks. Some parent peaks, e.g., one with K.I.=1540, resolved into several components none of which relatively large with respect to the others, so that none of the daughter peaks could be asterisked.

The two Kovats index values for each peak partially characterize the type of the compound. The most convenient identification guide line is through calculation of a difference or dispersion parameter:

$$(K.I. \text{ in Carbowax 20M}) - (K.I. \text{ in Apiezon L})$$

This parameter describes the gas chromatographic "polarity" with respect to Carbowax 20M stationary phase. The parameter is zero, by definition, for n-alkanes; can be a small number for branched alkanes; and increases by approximately 70 units by presence of a double bond in the hydrocarbon. The difference parameter is given in the corresponding five columns of Table XVIII, where it is classified into five ranges.

The first range, 0 to 100, contains alkanes and alkenes. The second range, 100 to 250, may contain dialkenes, ethers, acetals.

substances with heteroatoms in saturated or one-double-bond ring. The third range 250 to 380, is populated by alkanals (aldehydes), ketones, esters. It also can contain benzene and its alkyl derivatives; however, benzene has a Kovats index pair (in Carbowax 20M/Aplazon L) 960/680; toluene 1060/796; o-xylene, 1213/926; m-xylene, 1173/901; p-xylene, 1166/899; and ethylbenzene 1152/884; none of these is represented within error limits of order of ± 10 K.I., in the table and hence most likely are not present among the over 150 compounds shown. Amines also can be present in this range. The fourth range, 380-520, contains alkanols, lower alkenals, and alkadienals. The fifth range, above the dispersion parameter of 520, can contain substances with two polar groups on the molecule, or with several double bonds on alkanol molecules. The lowest member of glycol series, ethyleneglycol, has the index pair of 1580/660, and is not present in Table XVIII.

The indicated range boundaries are only approximate, especially at the lower end of K.I. values. This can be illustrated by the following examples. The n-alkanals are located at discrete K.I. pair values separated by approximately 100 unit steps between the successive members of the series: n-pentanal 985/685, n-hexanal 1078/783, n-heptanal 1188/867 (assumed, in Table, 1220/865), with the dispersion parameter of 300, 295, and 321, respectively. If the polar functional group is partially shielded by non-polar side chains, the molecule behaves in less polar manner; thus

n-butanal	= 326
2-methyl butanal	291
n-hexanal	295
2-ethylhexanal	271

Double bonds, on the other hand, increase the gas chromatographic polarity, cf

2-ethyl hexanal	271
2-ethyl-2-hexenal	341
n-butanal	326
n-butenal (crotonaldehyde)	433
2,4-hexadienal	509

The index pairs, of course, also are located at a point different from the corresponding n-alkanal points. Similar rules apply to ketones and esters:

Butyl Butyrate	$\sigma =$ 287
Isobutyl Butyrate (polar locus shielded)	233
Ethyl butyrate	285
Vinyl butyrate	304
Propyl propionate	286
Isopropyl propionate	270
Propyl acrylate	328
Allyl acrylate	392

Similarly, for n-alkanols the dispersion parameter is in the range of 480-510 (except for the lowest members of the series), decreases if side alkyl groups obstruct the hydroxyl group, and increases if double bonds are introduced.

Thus, although the grouping by ranges in Table XVIII is not ideal, it gives insight into the nature of the components. With the knowledge of the Kovats index in Apiezon L and of the approximate gas chromatographic polarity, estimates of the boiling points of the components are also possible and can be considered to be within $\pm 10^\circ\text{C}$ of the actual value. The estimated boiling points of the components are also tabulated in Table XVIII.

An absolutely positive identification of the specific components from the two gas chromatographic parameters only is not an exact procedure; it merely eliminates many unlikely choices, narrowing the selection to a few possible candidates. Pending the mass spectrographic identification, a closer chemical characterization of the over 150 components was not conducted. Some points fitted reasonably well certain compounds, however; these are indicated in parentheses as the most likely choices in the last column.

A survey of Table XVIII invites several interesting generalizations.

First, there is an unexpectedly frequent presence of alkanes and alkenes, with eight to fourteen carbon atoms, and with C_{12} alkane either not represented or present at low concentrations.

This can be either a physiological effect, or an effect that characterizes urban culture with vehicular exhausts containing unburned hydrocarbons present in air and picked up by human subjects. There are 35 components of this type; 9 of these are among the major 36 daughter-components. Some negative dispersion parameters that show up in the table are inherent in the peak-reinjection procedure where the injection into the second column is considered to occur at the middle of the finite duration injection interval, (e.g., 30 sec) although actually some components are present in the leading edge, others in the trailing edge of the parent peak. An alkane contained in the trailing edge of the injected parent peak will result in a small negative dispersion parameter.

The carbonyl compounds (including esters), with amines also possible, occur in the third dispersion index column, with possible additional cross over into higher and lower ranges. Here, 62 components occur, among these 19 are major components of their respective parent peaks.

The second dispersion index column contains 29 components, but only 2 of these appear to be major components of the respective parent peaks.

The fourth dispersion index column (alkanal, alkadienals) contain 22 components; 5 of these are major.

The fifth group contains 10 components, one of these is a major contributor to the corresponding parent peak. These are multifunctional substances or polyenes with polar functional groups, and their presence may deserve further exploration since they begin to appear in the higher boiling point ranges, repeatedly indicated to be important in discriminant functions. Also, compounds with lesser volatility can be recovered from air with higher efficiencies and perhaps more suitable in detection devices.

To summarize the inspection of Table XVIII, relatively high frequency of hydrocarbons, and the dominance of carbonyl compounds (including perhaps esters) among the major components, and

presence of multifunctional compounds, or carbonyl compounds with more than one double bond appear to be the characteristic features of the human chemical signature studied by the high-resolution two-column technique. Approximate group characterization of the components is summarized as follows:

	Major Components of <u>Parent Peaks</u>	Minor Components of <u>Parent Peaks</u>	<u>Total</u>
Hydrocarbons	9	26	35
Acetals, Ethers, Heterocyclics	2	27	29
Carbonyls, Esters, Amines	19	43	62
Alcohols, Diene Carbonyls	5	17	22
Diene Alcohols, Polyfunctionals	1	9	10
Total	36	122	158

*A few duplications, for peaks close in Kovats Index values on Carbowax 20M column, can be possible.

An inspection of Table XVIII also indicates that from the 25 parent peaks studied, the peaks areas reasonably well represent primarily the content of one single daughter component in 12 cases (one major daughter peak found in each), represent primarily the content of two daughter components in 11 cases, of three daughter components in 3 cases, and none in one case. In the latter, the parent peak consisted of several daughter components none of which excelled in magnitude. These observations should assist in considering the merits of single-column devices vs. two-column devices for pattern classification, at the resolution levels obtainable with 200-ft columns. The presence and the changes in the concentrations of the minor daughter components can, however, influence the position of the apex of the parent peak in the primary gas chromatogram and under certain conditions influence the pattern classification data, by shifting the peak assignment across a range boundary into the range of neighboring variables.

Therefore, the significance of the minor peak components in pattern recognition cannot be ignored.

Aside from the above 25 peaks eluted from the first 200-ft Carbowax 20M column and re-analyzed in the 50-ft Apiezon L column, the gas chromatogram from the first column contained many additional peaks. Data obtained from vapors of the same subject, a Caucasian female, and analyzed at 90°, 120°, and 150°C are listed in Appendix II. There is reasonable similarity between two analyses at 150° (A and B) on two subsequent days from the same subject. A summary of the number of peaks observed in vapors is as follows:

<u>Kovats Index</u> <u>Range</u>	<u>Analysis Temperature, °C</u>				<u>Maximum</u> <u>Number in</u> <u>Respective</u> <u>Range</u>
	<u>90</u>	<u>120</u>	<u>150 (A)</u>	<u>150 (B)</u>	
Below 300	10	2	3	3	10
801-1000	11	11	7	4	11
1001-1200	25	16	6	5	25
1201-1400	11	27	12	13	27
1401-1600		9	20	25	25
1601-1800		2	25	21	25
			7	3	7
TOTAL	57	76	80	74	130

Thus, prior to the subsequent split in the Apiezon L column, a 200-ft Carbowax 20M column, used isothermally at three different temperatures, indicating the presence of approximately 130 peaks.

4.5 Mass Spectrographic Identification of Human Signature Components

The initial results showed that the sizes of the peaks of both human signatures studied fell below the sensitivity of the mass spectrograph.

The experiments on the mixture of n-butanol and heptanal showed that both compounds were positively identified each time their respective gas chromatographic peak sizes matched those of the major and medium-size components of human signature.

The standard gas chromatographic analysis of the human signature collected on the large fluidized bed showed a significant increase in the sample magnitude. The size of the sample found was at least ten-fold that of the sample usually collected on the standard bed. Also, the peak distribution pattern was similar to the pattern previously obtained from the same subject under routine collection conditions.

Mass spectrographic analysis of human chemical signature collected on the large bed and fed directly from the open tubular column produced patterns which in most cases were too complex for immediate interpretation. One analysis indicated the presence of a compound (naphthalene) probably transferred to human from moth-producing materials. The peak of this compound was usually well resolved as observed in the respective gas chromatographic analyses of the sample.

The direct mass spectrographic analysis of effluent carrying human samples collected on standard size beds produced patterns which in all cases studied were too low above the instrument background for positive identification.

The difficulties encountered in the mass spectrographic identification of human signature components result mainly from the lack of sample resolution rather than its size. This is supported by the positive identification of the naphthelene (since its peak was well resolved), by the results of experiments on 1-butanol and heptanal and by recent frequent observations made

during the analysis of human signatures on the two-column apparatus. The results of these experiments indicated that most Carbowax 20M peaks were multi-component peaks.

5.0 PROGNOSIS FOR CHEMICAL SIGNATURE DISCRIMINATION DEVICES

The work conducted indicated that the complex chemical signatures of humans and environments contain features which permit classification by source type using relatively straightforward statistical discriminant techniques. The number of features that must be taken in account and their distribution within the gas chromatographic spectrum depends on the type of sources that must be discriminated. Although it is possible that further research may uncover some specific substances or a few substances that characterize e.g., humans versus environments, it is questionable if signature detection and identification devices should be based entirely on such features. A device that can be based on sufficient redundancy of features is less likely to suffer from interferences. From the findings thus far, it appears that 7 to 15 complex features may be necessary to reasonably characterize the source. The two-column data also indicate that these features consist of several chemical components some of which probably represent useful features while others act only as chemical noise.

Two questions can be posed: (1) which processes are best suited for signature identification and (2) can a sufficiently rapid device be envisioned?

Since the useful features are represented by various molecular species, it would be desirable to use vapor separation methods that permit retention of the molecular identities rather than other processes (such as mass spectrometry on non-separated mixtures) which splitter the molecules. Such methods are the basis of gas chromatography. Separation by interaction with a polar medium, as used in the present work, permits locating the discriminating features. It appears that a useful discriminant system can be based on the principle of pre-separation in polar columns and inspection of the discriminating regions in the second column or columns, cf. 4.4. Vertical cuts through e.g., Carbowax 20M versus Apiezon L field plot in the regions of interest should produce

enough information for identification of source types.

The second question, on the order of speed that may be achieved by such devices, must be answered in two steps: can a preconcentration of organic vapors from air be conducted fast enough, and can the subsequent task of identification of the features be conducted fast enough?

Concentration. Although higher sensitivities to all or to some substances can be achieved with some gas chromatographic detectors especially if helped by very careful work and adjustments, nanogram (10^{-9} g) amounts of individual substances of all types can be relatively easily detected by devices widely used in routine gas chromatographic practice. For a typical substance with a molecular weight of 100, this corresponds to 6×10^{12} molecules. Assuming the desired detectability level at 10^9 molecules/cm³ of air and the maximum gas-sample size into the device's processing channels at 1 cm³, a concentration factor of 6000 is needed to bring the lowest concentrations into the detector's performance range. Solubility of vapors of organic materials in gas chromatographic stationary phases decreases by a factor of such magnitude when temperature is changed from ambient to above 200 to 250 °C. Hence, equilibrium absorption from air at ambient temperatures and desorption at above 200 °C, in principle, permits one-step concentration of the dilute chemical signal to the level needed for a routine detection by usual gas chromatographic detectors.

There is no limitation, in principle, why such a process cannot be conducted with speeds of order of 1 sec. A thermodynamic requirement and a kinetic requirement must be met.

$$(k) (n_a) (v) > 6 \times 10^{12} \quad (I)$$

$$\frac{x^2}{2D} > 1 \quad (II)$$

where

k is the dimensionless partition coefficient for the vapor in the stationary-phase/air system, equivalent to the factor by which the concentration of the vapor molecules in the solution in the phase is larger than their concentration in air when equilibrium is reached

n_a is the concentration, in molecules/cm³, of the vapor in air

v is the volume, in cm³ of the stationary phase, in the concentrating device

x is the thickness of the absorbing film of the stationary phase in the device, cm

D is the diffusion constant of the vapor molecules in the stationary phase, cm² sec⁻¹

Partition coefficients at ambient temperature for vapors of substances with ambient saturated vapor pressures of several millimeters or lower in appropriate stationary phases easily reach into the range of 10^3 or higher. Therefore, the first requirement satisfying the sensitivity of the detectors can be met, for $n_a = 10^9$, with a few cubic centimeters of a suitable stationary phase. The second requirement, from the diffusion laws, must be met to perform absorption equilibration in a time span of order of 1 sec. With a typical $D = 10^{-8}$, the permissible film thickness is of order of 10^{-4} cm, well within technologically feasible limits.

Resolution. The complete resolution of the chemical signal captured from air would consist of a full inventory of the molecular species present in air at the time of inspection by the analog. As in any process, compromise must be accepted between resolution and time. Resolution is possible at two stages of processing -- in sending the vapor mixture that constitutes the signal with assistance of a carrier gas through a gas chromatographic partition tube, and in using species-selective detectors on the signals resolved in the first stage. Most detectors and their electronic circuitry have very fast response, and their specificity cannot be improved by simply allowing more time. Therefore, signal processing time and resolution depend

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primarily on partition-tube performance.

The resolution that can be produced per second can be estimated from the following equation*:

$$\frac{R_{23}}{t} = (r_{23} - 1) \frac{k}{(1+k)^2} \frac{\sqrt{N}}{t_m}$$

where

R_{23} is the resolution of the vapors of substances 2 and 3 ($R_{23} = 1$ indicates very distinct resolution with a negligible overlap of signals)

r_{23} is the ratio of partition coefficients of the vapors 2 and 3 in the stationary phase at resolution temperature

t is the time in seconds

k is the retardation ratio in the partitioning tube and depends on the volume of gas space, the stationary phase in the tube, and the partition coefficient of the vapor in the stationary phase

N is the equivalent plate number of the partitioning tube

t_m is the gas holdup time (volumetric flow rate of carrier gas through the tube divided by gas phase volume)

We now use this equation to estimate what ratio, r_{23} , in partition coefficients of 2 and 3 can be ascertained in 1 sec ($R_{23}/t = 1$) in a typical short partition tube. For capillary tubes, k is of order of 0.2 to 20, and a 10-cm-long tube can typically be equivalent to $N=100$. Selecting $k=1$, and $t_m=0.1t$, $r_{23}=1.04$. A difference of 4% in the partition coefficients of 2 and 3 should suffice to tell these components apart within 1 sec. Other estimates** show that a "100-plate" partition tube can permit recovery of substance 2 with less than 5% impurity of the substance 3 and, in reverse, if both are present in equal amounts

*Loyd, R.J., Ayers, B.O., and Karasek, F.W., Anal. Chem., 32, p. 43, 698 (1960).

**Littlewood, A.B., Gas Chromatography, p. 136, Acad. Press, New York, 1962.

in the original mixture and their partition coefficients differ by 40% or more.

In the case of very complex vapor mixtures, many compounds are present, and the above resolution is insufficient for a complete inventory. There is a continuous overlapping of the substances with small differences in the retention times. In usual cases, low-resolution processing then results in a poorly differentiated effluent from the partition tube with peaks of noticeable size concentrations considerably in excess of the total concentration of other poorly differentiated substances. Any portion of the effluent from the above "100-plate" partition column, taken during the time interval of $t \pm 0.4t$, contains over 95% of the component or components with the retention time t , and a lesser fraction of components with retention times within the $t \pm 0.4t$ boundary.

In the notation of Kovats retention index (cf. Section 3.6.5), the retention times are referred to the scale of n-alkanes with a logarithmic transformation to produce intervals of 100 Kovats Index units between each of two consecutive members of the homologous series of n-alkanes. Two compounds with retention times of t and $t + 0.4t$ produce an index difference of 50 K.I. units. The interval between C_2 (200 K.I.) and C_{20} (2000 K.I.) alkanes carries 36 intervals of 50 K.I. units each. Thus, in principle, a parallel multichannel unit with separate non-polar phase channels operated at different processing regimes (temperatures, flow rates, and times) can split the Kovats Index continuum corresponding to compounds with boiling points up to 400°C into 30 to 40 cells. Each cell is populated by molecular species with relatively similar partition coefficients in the gas/non-polar phase system. The detector response at the exit of each low-resolution channel is a complex function of the population distribution within the cell with frequency maxima at those time points at which some more-than-usually abundant molecular species elute.

Various discriminators can then be applied to the population of each of the cells. Two types of discriminators can be derived from gas chromatographic practice. One is a polar or otherwise specific stationary phase in a low-resolution column with its characteristics being the shift of the elution positions with respect to the n-alkane position. The other is the sensitivity of the detector to some atomic species, e.g., a sulfur-sensitive detector indicates the distribution of sulfur compounds among the cells and within each cell. The species-sensitive detector can apply to the initial multichannel signals or to the signals first distorted by the second-stage discriminator columns. Since the dispersion shifts easily exceed 50 K.I. units, a low-resolution discriminator is sufficient to produce a rather informative dispersed signal. The shifts are characteristic of functional groups in the molecule and of the steric clustering of non-polar parts of the molecule around the functional group.

Thus, there is no fundamental difficulty in development of processes that can characterize chemical signatures of humans within a few seconds to a degree sufficient for classification by sources. The actual choice of the processing steps requires however a massive experimental and theoretical effort.

6.0 SUMMARY AND CONCLUSIONS

Within the context of the present work, chemical signatures are defined as compositional characteristics of organic vapors emanated into air by various sources. The objective of the conducted work was to establish if features exist in the chemical signatures of humans that can be used to distinguish humans from environments and to distinguish among humans. A further objective was to characterize those features as far as possible within the limits of the effort and presently available technology. An all-glass Teflon cell was available from the previous project (DA-11-022-AMC-1775(X)). This cell permitted isolating humans temporarily, from the surrounding air and maintain them in a flowing, highly purified air stream so that better samples, not confused by a continuous arrival of contaminants from air, could be collected.

It was soon discovered that many of the usual sample collection procedures, such as: collection in vessels, cryogenic cooling of air, or adsorption on carbon, were unsuitable for the chemical signature acquisition when the chemical signatures consisted of a large variety of organic materials available in air at concentrations as low as parts per billion.

Consequently, considerable effort was directed toward the development of improved techniques for the acquisition and processing of samples of chemical signatures from humans and various environments. The essential features of this development involved new systems for sample collection, transfer, and injection into the gas chromatograph. The sample acquisition was based on the concept of collection through equilibration. The collectors were special fluidized beds of Teflon powder coated with a thin film of Apiezon L stationary phase. The fluidized bed configuration of the collector produced the best possible contact of samples with air. The Apiezon L permitted later recovery of the dissolved sample with minimum artifacts from decomposition of the trapping phase itself. In a typical

collection of a sample from a human, the bed material was exposed to air carrying the vapors at the flow rate of 0.75 liters per sec. The environment chemical signatures were collected by pulling the air through the sampler by a small blower. Since the collection involved equilibration, the actual airflow rate through the sampler did not have to be known and was not critical as long as the powder was kept in the fluidized state.

A special calibration procedure was devised to correlate the concentration of a known sample component in air directly to the size of the peak on the gas chromatogram, to check the collection efficiency and to establish the time required for attainment of equilibrium concentrations for various types of compounds.

The recovery of the collected sample from the trapping film and its injection into the gas chromatograph for analysis was effected by a special sample transfer and injection system. The sample transfer system was designed to allow the transfer of the vapors from the bed maintained at 80°C into a special stainless steel thin wall needle which was kept at liquid nitrogen temperature. The transfer was effected with cryogenically purified gas. The sample injection system was constructed to deliver the sample from the injector needle to the gas chromatographic column in a timed, automatic and reproducible manner.

The sample acquisition step was found to require 45 min to produce reproducible and equilibrated samples. The typical duration of the sample transfer step was 6 min. During this period 70 to 80 percent of the vapor sample was found in the injector needle. Higher efficiency could be achieved with a longer transfer time but at the expense of increased analytical time. The sample injection step resulted in reproducible and practically quantitative injections with 95 to 98 percent of the sample transferred from the needle and introduced into the injection port of the gas chromatograph.

The standardized techniques developed in the initial phase

of this work were used to obtain 65 gas chromatographic patterns of human signatures and 10 patterns of various environments. The human signatures were collected from healthy subjects of both sexes and several races. The races included American Caucasian males and females, Indian males (born in India), a few American Negroes, and a Mexican and a Malayan subject.

The first series of chemical signatures were analyzed at 125°C using a 10-ft, 1/8-in O.D. packed bed columns with a Carbowax 20M stationary phase. Later work, conducted on a smaller variety of samples from humans, utilized 50 and 200-ft long support-coated open tubular columns with a Carbowax 20M coating. These provided much higher resolution and were used isothermally at several temperature levels (90°, 125°, and 150°C). In still later work, a specially modified two-column gas chromatograph was used in which the sample was first analyzed in a 200-ft support-coated open tubular Carbowax 20M column and the gas chromatographic peaks found previously to occur most frequently in the human chemical signatures were further analyzed in a 50-ft support-coated open tubular Apiezon L column. This procedure allowed the resolution of peaks into further components and a subsequent estimation of their gas chromatographic polarities. A three-column gas chromatograph was also used, combined with a computer-analysis of data, to characterize the components of the chemical signatures in similar terms.

The great complexity of the chemical signatures was revealed by the experimental survey of the samples from 65 humans, and 10 environments in analysis in the 10-ft packed columns. At this stage, the data were studied by several computerized statistical analysis techniques, to discover if: (1) valid discriminant functions can be extracted permitting classification of the signatures by the types of their sources, (2) how many gas chromatographic features need to be used to obtain classifications, and (3) indicate those features which are most significant in forming discriminant functions and therefore deserve priority in

their chemical identification.

Attempts at mass spectrographic or other identification of components at various degrees of gas chromatographic resolution were also made. These indicated that the highest degrees of resolution in a two-column system will be needed to resolve the chemical species present in the chemical signatures of humans to a degree where meaningful identifications of the individual components becomes possible. This work remains an objective for the future, and in many cases will require further enrichment of the sample to reach concentrations sufficient for identification.

The data were organized in a Kovats Index system and are tabulated in the Appendix.

The following conclusions were reached:

- (1) At the resolution available isothermally with 10-ft packed gas chromatographic columns and using collection and sample processing methods to ppb of organic species in air, the human chemical signatures exhibited from 32 to 48 features per signature.
- (2) Statistical pattern analysis conducted on the chemical signatures at the above resolution levels indicated that from 2 to 15 gas chromatographic variables are needed to reach meaningful classification of the signatures by their sources. The number of variables (features) that must be utilized increases as the similarity of sources (e.g. Caucasian males vs. females) or variability within classes of sources (e.g. males of several races vs. females of several races) increases.
- (3) Some classifications (e.g. humans vs. environments) based on classical stepwise discriminant analyses of patterns resulted in discriminant function which reasonably successfully classified additional test cases.
- (4) There were indications that the decision boundaries for the chemical signature classifications in statistical hyperspaces may be more complex than accessible to the classical stepwise discriminant procedures.
- (5) Detection and source discrimination devices based on the principle of chemical signatures must most probably be based on detection and correlation of 5 to 15 features at resolution levels outlined in the conclusion of paragraph #1.

- (6) Chromatographic analysis with resolution obtainable isothermally at several temperature levels from 200-ft support-coated Carbowax 20M column exhibited typically 100 to 130 peaks. Twenty-five of these, reanalyzed isothermally on 50-ft support-coated Apiezon L column separated further into 3 to 10 chemical components, thus producing over 150 observable components. The total number of components observable at this level of two-stage resolution in the chemical signature of a selected human, can be estimated to be in the range between 500 to 1000.
- (7) Classification of the observed components, by their gas chromatographic polarity, with attention to those components most frequently observed in humans, indicated the presence of considerable amounts of hydrocarbons, carbonyl compounds and a few alcohols. Compounds considerably more polar than alcohols were also observed and probably belong to species with more than two polar groups per molecule, or contain more than one double bond, in addition to a polar group.
- (8) Mass spectrographic identification succeeded only on two compounds which originate most probably from moth-proofing materials and therefore represent features characteristic of advanced material culture. The surprising frequency-of-occurrence of hydrocarbons may be related to a similar environment permeated by vehicular exhaust. Future mass spectrographic identification should be attempted only on components after their resolution in two-column system.

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Table I
 EXAMPLES OF CALIBRATION FOR COLLECTION OF SELECTED COMPOUNDS
 IN APIEZON L FLUIDIZED BED

Vapor	Detectability Limit With Present Technique, (molecules/cm ³)/cm ² a
n-Xylene	1.1 x 10 ¹⁰
n-Butanol	3.6 x 10 ¹⁰
n-Hexanol	5.2 x 10 ⁹
Octanone	7.2 x 10 ¹⁰
n-Heptaldehyde	6.1 x 10 ⁹
Acetic Acid	2.6 x 10 ¹¹
n-Propionic Acid	4.1 x 10 ¹⁰
n-Butyric Acid	5.7 x 10 ⁹
Ethyl Propionate	1.2 x 10 ¹¹
Ethyl Butyrate	6.1 x 10 ¹⁰
Dibutylamine	1.7 x 10 ¹¹

^aThe number of molecules per cubic centimeter of air that results in a 1-cm² peak at the highest sensitivity of the gas chromatograph

Table II
 CALIBRATION OF TWO-COLUMN SYSTEM WITH n-ALKANES¹

n-Alkane ²	Elution Time, Min.		Corrected Retention Time, Min.	
	Carbowax 20M	Apiezon L	Carbowax 20M	Apiezon L
C ₈	4.74	5.83	0.71	0.73
C ₁₀	6.18	9.39	2.15	2.85
C ₁₁	7.77	13.68	3.74	5.55
C ₁₂	10.53	21.65	6.50	10.76
C ₁₃	15.15	36.42	11.12	20.91
C ₁₄	22.92	63.80	18.89	40.52
C ₁₅	37.24	118.36	33.21	80.76

¹The system at 125°C

²Elution data for methane: Carbowax 20M, 4.03 min; Apiezon L, 4.39 min

Table III
 CALIBRATION OF TWO-COLUMN APPARATUS WITH POLAR COMPOUNDS¹

Compound ²	Corrected Retention Time, Min. ³		Kovats Retention Index	
	Carbowax 20M	Apiezon L	Carbowax 20M	Apiezon L
Ethanol	1.12	0.06	880	420
1-Propanol	2.18	0.12	1005	523
1-Butanol	4.15	0.30	1118	636
1-Pentanol	7.65	1.03	1235	726
Propionaldehyde	0.53	0.11	743	510
Butyraldehyde	1.00	0.20	859	600
Hexanal	3.53	0.67	1092	783
Heptanal	6.39	1.20	1198	870
Octanal	11.27	2.46	1304	976
2-Butanone	1.12	0.17	881	585
3-Pentanone	1.80	0.31	967	667
2-Heptanone	6.36	1.11	1197	859
2-Octanone	10.68	2.17	1290	958

¹The apparatus at 125°C

²Elution data for methane: Carbowax 20M, $M_C=4.03$ min; Apiezon L, $M_A=4.39$ min; $M_A-M_C=0.36$ min

³ $C_C = E_C - M_C$; $C_A = (E_A - E_C) - (M_A - M_C)$ where

C_C and C_A - Corrected retention times on Carbowax 20M and Apiezon L, in minutes, respectively

E_C and E_A - Elution times on Carbowax 20M and Apiezon L, in minutes, respectively

Table IV
 CALIBRATION OF THREE-COLUMN APPARATUS WITH n-ALKANES¹

n-Alkane ²	Elution Time, Min.		Corrected Retention Time, Min.		Ratio ³ Apiezon L Long/Short
	Carbowax 20M	Apiezon L Short Long	Carbowax 20M	Apiezon L Short Long	
C ₇	5.16	6.24 7.39	0.84	0.92 1.90	2.07
C ₉	8.13	12.56 17.12	4.02	4.27 8.83	2.07
C ₁₀	12.06	21.48 31.54	7.95	9.26 19.15	2.07
C ₁₁	19.20	39.33 59.51	15.09	19.99 39.98	2.00
C ₁₂	33.21	76.21 ⁴ 121.21 ⁴	29.10	43.00 88.00	2.05

¹The Apparatus at 90°C

²Methane elution data: Carbowax 20M, 4.11 min; Apiezon L-short, 4.27 min; Apiezon L-long, 4.44 min

³Average value, 2.05 ± 0.02

⁴From graphical extrapolation

Table V
 CALIBRATION OF THREE-COLUMN APPARATUS WITH n-ALKANES¹

n-Alkane ²	Elution Time, Min.		Corrected Retention Time, Min.			Ratio ³ Apiezon L Long/Short
	Carbowax 20M	Apiezon L Short Long	Carbowax 20M	Apiezon L Short Long	Apiezon L Long/Short	
C ₈	4.77	5.73 6.75	0.80	0.80 1.68	2.10	
C ₉	5.40	7.10 8.89	1.43	1.54 3.19	2.07	
C ₁₀	6.60	9.80 13.04	2.63	3.04 6.14	2.02	
C ₁₁	8.61	14.54 20.66	4.64	5.77 11.75	2.02	
C ₁₂	12.03	23.27 34.77	8.06	11.08 22.44	2.03	
C ₁₃	17.76	39.93 60.71	13.79	22.00 42.95	1.95	

¹The apparatus at 120°C

²Methane elution data: Carbowax 20M, 3.97 min; Apiezon L short, 4.13 min; Apiezon L-long, 4.27 min

³Average value, 2.03 ± 0.04

Table VI
 CALIBRATION OF THREE-COLUMN APPARATUS WITH n-ALKANES¹

Compound ²	Elution Time, Min.		Corrected Retention Time, Min.		Ratio ³ Apiezon L Long/Short
	Carbowax 20M	Apiezon L Short	Carbowax 20M	Apiezon L Long	
C ₈	4.24	4.79	0.40	0.82	2.05
C ₉	4.50	5.35	0.66	1.43	2.04
C ₁₀	4.92	6.30	1.08	2.54	2.07
C ₁₁	5.61	8.03	1.77	4.57	2.01
C ₁₂	6.81	10.86	2.97	7.91	2.03
C ₁₃	8.64	15.63	4.80	14.00	2.05
C ₁₄	11.69	23.25	7.85	23.61	2.07

¹The apparatus at 150°C

²Methane elution data: Carbowax 20M, 3.84 min; Apiezon L-short, 3.99 min; Apiezon L-long, 4.14 min

³Average value, 2.04 ± 0.02

Table VII
 CALIBRATION OF THREE-COLUMN APPARATUS WITH n-ALKANES¹

Compound ²	Elution Time, Min.		Corrected Retention Time, Min.		Ratio ³ Apiezon L Long/Short
	Carbowax 20M	Apiezon L Short Long	Carbowax 20M	Apiezon L Short Long	
C ₈	4.22	4.81 5.40	0.38	0.44 0.88	2.00
C ₉	4.48	5.38 6.28	0.64	0.75 1.50	2.00
C ₁₀	4.86	6.36 7.86	1.02	1.35 2.70	2.00
C ₁₁	5.64	8.09 10.59	1.80	2.30 4.65	2.02
C ₁₂	6.84	11.01 15.27	3.00	4.02 8.13	2.02
C ₁₃	8.67	15.83 23.12	4.83	7.01 14.15	2.02
C ₁₄	11.52	23.83 36.39	7.68	12.16 24.57	2.02

¹The apparatus at 150°C

²Methane elution data: Carbowax 20M, 3.84 min; Apiezon L-short, 3.99 min; Apiezon L-long, 4.14 min

³Average value, 2.01 ± 0.01

Table VIII
 CALIBRATION OF THREE-COLUMN APPARATUS WITH ALDEHYDES¹

Compound ²	Elution Time, Min.		Corrected Retention Time, Min.		Ratio ³ Apiezon L Long/Short		
	Apiezon L		Apiezon L				
	Short	Long	Short	Long			
Propionaldehyde	5.07	5.61	0.99	0.10	0.20	2.00	
Butyraldehyde	5.67	6.01	1.59	0.18	0.38	2.11	
Hexanal	8.94	9.79	10.74	4.86	0.69	1.46	2.11
Heptanal	12.36	13.88	15.55	8.28	1.36	2.85	2.09
Octanal	18.09	20.85	23.93	14.01	2.60	5.50	2.12

¹The apparatus at 120°C

²Elution data for methane: Carbowax 20M, 4.08 min; Apiezon L-short, 4.24 min; Apiezon L-long, 4.42 min

³Average value: 2.08 ± 0.04

Table IX
 CALIBRATION OF THREE-COLUMN APPARATUS WITH n-ALCOHOLS¹

Compound ²	Elution Time, Min.		Corrected Retention Time, Min.			Ratio ³ Apiezon L Long/Short		
	Carbowax 20M	Apiezon L Short Long	Carbowax 20M	Apiezon L Short Long	Apiezon L Long/Short			
Methanol	4.23	4.40	4.60	4.87	0.36	0.03	0.07	2.33
Ethanol	4.44	4.64	4.87	5.15	0.57	0.06	0.13	2.17
1-Propanol	4.89	5.15	5.44	6.11	1.02	0.12	0.25	2.08
1-Butanol	5.76	6.11	6.49	7.65	1.89	0.21	0.43	2.05
1-Pentanol	7.12	7.65	8.21	10.06	3.25	0.39	0.79	2.03
1-Hexanol	9.21	10.06	10.95		5.34	0.71	1.44	2.03

¹ System at 150°C

² Elution data for methane: Carbowax 20M, 3.87 min; Apiezon L-short, 4.01 min; Apiezon L-long, 4.17 min

³ Average value: 2.11 ± 0.09

Table X
 CALIBRATION OF THREE-COLUMN APPARATUS WITH ALDEHYDES¹

Compound ²	Elution Time, Min.		Corrected Retention Time, Min.		Ratio ³ Apiezon L Long/Short
	Carbowax 20M	Apiezon L Short Long	Carbowax 20M	Apiezon L Short Long	
Propionaldehyde	4.26	4.46 4.68	0.40	0.06 0.12	2.18
Butyraldehyde	4.55	4.79 5.06	0.70	0.10 0.21	2.10
Hexanal	5.97	6.45 7.01	2.10	0.34 0.74	2.17
Heptanal	7.35	8.13 9.01	3.48	0.64 1.36	2.12
Octanal	9.48	10.86 12.28	5.61	1.24 2.50	2.02

¹System at 150°C

²Elution data for methane: Carbowax 20M, 3.87 min; Apiezon L-short, 4.01 min; Apiezon L-long, 4.17 min

³Average value: 2.12 ± 0.06

Table XI
 CONTRIBUTIONS TO EFFECTIVE CARBON NUMBER
 IN HYDROGEN FLAME IONIZATION DETECTOR

Aliphatic C	1.0
Aromatic C	1.0
Olefinic C	0.95
Acetylenic C	1.30
Ketone, Aldehyde C	0
Nitrile C	0.3
Ether O	-1.0
Primary Alcohol O	-0.6
Secondary Alcohol O	-0.75
Tertiary Alcohol O	-0.25
Ester O	-0.25
Two or more Cl on Single	
Aliphatic C	-0.12 each Cl
Cl on Olefinic C	+0.05
Amine N	as O in corresponding alcohols

Table XII
n-BUTANOL EMISSION FROM VARIOUS HUMANS

Subject	Peak Area, cm ²	Rate Molecules/Sec
CAUCASIAN MALE		
A	2.00	0.62 x 10 ¹⁴
	1.60	0.49
	3.20	0.99
B ^a	7.78	2.42
	5.52	1.71
	3.28	1.02
	0.53	0.16
	1.76	0.55
C ^b	48.00	14.90
CAUCASIAN FEMALE		
D	11.64	3.61
E	6.08	1.89
F	46.30	14.40
INDIAN MALE		
G	3.60	1.12
H	4.13	1.28
I	3.84	1.19
J ^c	14.30	4.43
	4.00	1.24
	19.88	6.16
K	4.64	1.44
L	5.04	1.56
M	9.92	3.07
N	1.28	0.40
O	3.50	1.08
P	2.08	0.65
NEGRO MALE		
Q	5.28	1.64
NEGRO FEMALE		
R	60.48	18.75
S	56.40	

^aFirst 3 measurements taken when subject was on average mixed diet; 4th and 5th measurements taken 7 and 8 days later, respectively, after subject had been on special diet of fish, cheese, milk and tea.

^bSubject is a chemist working with various organic solvents.

^cSubject is a strict vegetarian; other Indian subjects mostly vegetarians.

Table XIII
 DISCRIMINANT ANALYSIS OF CHEMICAL SIGNATURES
 OF MALES, FEMALES, AND AIR FROM VARIOUS ENVIRONMENTS

Consecutive Number*	Variables Considered Corresponding Kovats Index Range	Number of Cases Considered		
		Males, 39	Females, 26	Air, 10
		Cases Correctly Classified, %		
1	1250-1300	77	34	70
2	1350-1400	77	38	60
3	1450-1500	82	51	90
4	1050-1100	84	48	90
5	650- 700	84	62	90
6	> 1600	84	62	90
.				
.				
.				
.				
15	850- 900	87	79	90

* In order selected by the computer

At 15 variables considered, the following Kovats Index ranges have not been selected by the computer as useful discriminants: 700-750; 800-850; 950-1000; 1000-1050; 1150-1200; 1400-1450; 1550-1600.

Table XIV
 DISCRIMINANT ANALYSIS OF CHEMICAL SIGNATURES
 OF CAUCASIAN MALES, INDIAN MALES, AND CAUCASIAN FEMALES

Consecutive Number*	Variables Considered Corresponding Kovats Index Range	Number of Cases Considered		
		Caucasian Males, 25	Indian Males, 12	Caucasian Females, 23
		Cases Correctly Classified, %		
1	1250-1300	60	41	61
2	700- 750	56	75	73
3	1350-1400	64	75	69
4	650- 700	64	75	69
5	1550-1600	72	75	69
.				
.				
.				
13	> 1600	83	83	80

* In order selected by the computer

At 13 variables considered, the following Kovats Index ranges have not been selected by the computer as useful discriminants:
 < 600; 600-650; 850-900; 900-950; 1150-1200; 1200-1250;
 1300-1350; 1400-1450; 1500-1550.

Table XV
 LIST OF VARIABLES (K.I. RANGES)
 (Those ranges in which peaks most frequently
 occurred in vapor from humans are marked P)

Consecutive Number of Range, (Variable)	K.I. Range From-To	Range Width, K.I. Units	Variables Selected for Latest Class Separations
1	0- 465	465	
2	465- 485	20 P	
3	485- 516	31	
4	516- 536	20 P	x P
5	536- 590	54	x
6	590- 610	20 P	x P
7	610- 645	35	
8	645- 665	20 P	x P
9	665- 666	1	
10	666- 686	20 P	x P
11	686- 706	20 P	x P
12	706- 722	16	
13	722- 740	18 P	
14	740- 758	18 P	
15	758- 799	41	x
16	799- 819	20 P	
17	819- 824	5	x
18	824- 842	18 P	x P
19	842- 860	18 P	
20	860- 882	22	
21	882- 902	20 P	
22	902- 904	2	x
23	904- 924	20 P	x P
24	924- 968	44	
25	968- 987	19 P	
26	987-1005	18 P	x P
27	1005-1025	20	x
28	1025-1045	20 P	
29	1045-1061	16	
30	1061-1081	20 P	x P
31	1081-1084	3	
32	1084-1104	20 P	
33	1104-1112	8	x
34	1112-1132	20 P	
35	1132-1140	8	x
36	1140-1160	20 P	
37	1160-1171	11	x
38	1171-1191	20 P	
39	1191-1195	4	
40	1195-1215	20 P	
41	1215-1232	17 P	
42	1232-1250	18 P	
43	1250-1265	15	x
44	1265-1285	20 P	
45	1285-1297	12	
46	1297-1315	18 P	x P
47	1315-1334	19 P	x P
48	1334-1340	6	
49	1340-1360	20 P	x P
50	1360-1372	12	x
51	1372-1392	20 P	x P
52	1392-1401	9	x
53	1401-1419	18 P	
54	1419-1437	17 P	x P
55	1437-1454	17	x
56	1454-1472	18 P	x P
57	1472-1490	18 P	x P
58	1490-1500	10	x
59	1500-1517	17 P	x
60	1517-1535	18 P	x P
61	1535-1590	55	x
62	1590-1610	20 P	
63	1610-1615	5	x
64	1615-1635	20 P	
65	1635-1800	165	x

Table XVI
 SUCCESS IN STATISTICAL PATTERN CLASSIFICATION
 AS A FUNCTION OF THE NUMBER OF VARIABLES
 (Variables Correspond to Range Numbers in Table XV)

Consecutive Numbering	<u>All 37 P-Ranges (Run A)</u>		<u>10 P-Ranges Only (Run B)</u>	
	<u>Identification Number of Variable of Table XV</u>	<u>% of Human Signatures Classified in "Human Class"</u>	<u>Identification Number of Variable of Table XV</u>	<u>% of Human Signatures Classified in "Human Class"</u>
1	64	71	64	71
2	56	78	56	78
3	57	83	57	83
4	47	83	26	87
5	46	83	44	91
6	26	86		
7	18	86		
8	10	no readout		
9	30	88		
10	8	no readout		
11	16	94		
Additional		92 to 94		

Table XVII

SEVERAL IDENTIFICATIONS OF COMPONENTS IN CHEMICAL SIGNATURE
FROM HUMAN SUBJECT BY THREE-POINT MATCH
IN THREE-COLUMN APPARATUS¹

Compound	E l u t i o n T i m e , M i n .					
	Carbowax 20M		Aplezon L-Short		Aplezon L-Long	
	Calibration	Signature ²	Calibration	Signature ²	Calibration	Signature ²
Methanol	4.23	4.29	4.40	4.48	4.60	4.68
Ethanol	4.44	4.38	4.64	4.63	4.87	4.86
Butyraldehyde	4.55	4.53	4.79	4.77	5.06	5.05
Heptanal	7.35	7.53	8.13	8.27	9.01	9.03
Octanal	9.48	9.48	10.86	10.68	12.28	12.45

¹The apparatus at 150°C

²Human subject: Caucasian female L.B.

Table XVIII
 CHARACTERIZATION OF HUMAN CHEMICAL SIGNATURE COMPONENTS ON TWO-COLUMN GAS CHROMATOGRAPHIC APPARATUS¹

Peak Examined Consecutive Number	Highest Occurrence Frequency Peaks in Human Chemical Signatures	Component Characterization										Approximate Boiling Point, °C	Closest Tentative Identification
		Kovats Index Carbowax 20M	Kovats Index Apiezon L2	0 - 100	100 - 250	250 - 380	380 - 520	520 - 1,200	1,200 - 1,400	1,400 - 1,600	1,600 - 1,800		
1	722-740, 29P	715	463*					252*				50	(methylal)
			567		142							70	
			615		100							80	
2	739-819, 45P	810	545*					370				60	(acetone)
			440					265*				50	
			656		154							100	
			688		122							90	
			795			15						110	(octane)
			822			-12						120	
3	882-902, 45P	879	545*					334*				75	(butyral nonane)
			902*			-23*						140	(methanol)
			316							563		70	
			657		222							90	
			718		161							105	
			865		14							130	
4	882-902, 45P	877	545*					332*				80	(butyral nonane)
			699*			-22*				484		140	(ethanol)
			393									80	
			640		237							90	
			656		221							100	
5	968-987, 48P	985	633*					352*				90	(pentanal)
			990*			-5*						160	
			645					300				110	
			775									120	
			795		210							120	
			953		200							150	
			965			32						150	
			985			20						150	
						0						160	
6	987-1005, 67P	1000	1002*			-2*						160	(propanol)
			525*							475*		100	
			697									120	
			850					303				140	
			930		150							145	
			945			70						150	
			965			55						155	
			975			25						160	
7	1025-1045, 34P	1030	545							480		110	
			697									115	
			1005			25		333				165	

Table XVIII (continued)
 CHARACTERIZATION OF HUMAN CHEMICAL SIGNATURE COMPONENTS ON TWO-COLUMN GAS CHROMATOGRAPHIC APPARATUS¹

Peak Examined - Consecutive Number	Highest Occurrence Frequency Peaks in Human Chemical Signature - Kovats Index Range Carbomax 20M/125°C	Component Characterization						Approximate Boiling Point, °C	Closest Tentative Identification	
		L = I _{20M} /Apl								
		Kovats Index Carbomax 20M	Abiezon 12	0 - 100	100 - 250	250 - 380	380 - 520			> 520
8	1025-1045, 34P	620*						120		
		715*						125		
9	1061-1081, 48P	1060*		-10*		335*		180		
		775				275		130		
		822			228				140	
		850			200				145	
		859			192				150	
		866		84					155	
		888		62					160	
		883*							130	(hexanal)
		1100*	1078	-22*		295*			185	(undecane)
		828				250			140	
		1054		24					170	
		1065		13					175	
1084		-5					175			
1093		-15					180			
10	1084-1104, 45P	747*						125		
		1100*		-5*		348*		120		
		577						110		
		718					518	120		
		813				377		130		
		1050		45		282		175		
		1062		33				180		
		1083		12				180		
		1090		5				180		
		630*							120	
11	1084-1104, 45P	1103*		0*				180	(undecane)	
		936				267		145		
		865			238			145		
		1050		53				175		
		1062		41				180		
		1085		18				180		
		987*							175	(heptanal)
		865	1220		233*			355	150	
		903						317	160	
		947						273	170	
1055			165				185			

Table XVII (continued)
 CHARACTERIZATION OF HUMAN CHEMICAL SIGNATURE COMPONENTS ON TWO-COLUMN GAS CHROMATOGRAPHIC APPARATUS¹

Peak Examined Consecutive Number	Highest Occurrence Frequency Peak in Human Chemical Signature	Retention Time		Kovats Index		Approximate Boiling Point, °C		Closest Tentative Identification	
		Carbowax 20M, 125°C	Carbowax 20M, 125°C	Carbowax 20M	Apiezon Lz	250 - 380	380 - 520		
13	1265-1285, 60P	1275	1014*	261*	392	180			
			883			150			
			914			321	170		
			977			298	175		
			1037	238			180		
14	1297-1315, 54P		1018	315*		170		(octanal)	
			907	254*	385	180			
			1010	282		175			
15	1297-1315, 54P		974*	318*		170		(octanal)	
			760			> 140			
			980			180			
			992			180			
			1044	248		186			
			1140	152		205			
			1180	112		210			
16	1297-1315, 54P		972*	323*		175			
			893			170			
			920			402			
			950			375			
			1003			345			
			1045			292			
			1070			260			
17	1140-1160, 64P		1140			190			
				225		205			
				155					
			950*			170			
			925			175			
18	1372-1392, 65P		1345			180			
				395*		240			
				420		180			
						185			
						190			
						195			
						180			
19	1372-1392, 65P		965*	418*		180			
			1390*			240			
			965			180			
			1035			180			
			1052			190			
				348					
				331					
				269					

Table XVIII (continued)
CHARACTERIZATION OF HUMAN CHEMICAL SIGNATURE COMPONENTS ON TWO-COLUMN GAS CHROMATOGRAPHIC APPARATUS¹

Peak Examined Consecutive Number	Highest Occurrence Frequency Peaks in Human Chemical Signatures Kovats Index Range Carbowax 20M, 125°C	Component Characterization										Approximate Boiling Point, °C	Closest Tentative Identification	
		C20M ¹ Apl												
		Kovats Index Carbowax 20M		Apiezon L2		100 - 250		250 - 380		380 - 520				520 - 720
19	1372-1392, 65P	1390	1075*	100	100	100	250	250	380	380	520	520	190	
			1235*			155*							225	
			753										> 150	
20	1372-1392, 65P	1392	1077*	100	100	100	250	250	380	380	520	520	190	
			635										> 130	
			1054										185	
21	1419-1437, 50P	1443	1162*	100	100	100	250	250	380	380	520	520	210	(tetradecane)
			946										180	
			1068										195	
22	1454-1472, 69P	1468	1153*	100	100	100	250	250	380	380	520	520	220	(heptanol)
			734										> 150	
			923										> 180	
23	1472-1490, 66P	1490	1192*	100	100	100	250	250	380	380	520	520	220	
			735										195	
			1003										210	
24	1472-1490, 66P	1493	1177*	100	100	100	250	250	380	380	520	520	220	
			897										190	
			980										205	
25	1472-1490, 66P	1493	1012	100	100	100	250	250	380	380	520	520	220	
			1075										225	
			1150										225	
26	1472-1490, 66P	1493	1187	100	100	100	250	250	380	380	520	520	220	
			1195										225	
			1213										230	

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24	Three-Column Data Interpretation Assisted by GA3COL Computer Program
25	Display of Two-Column Data on Human Chemical Signatures

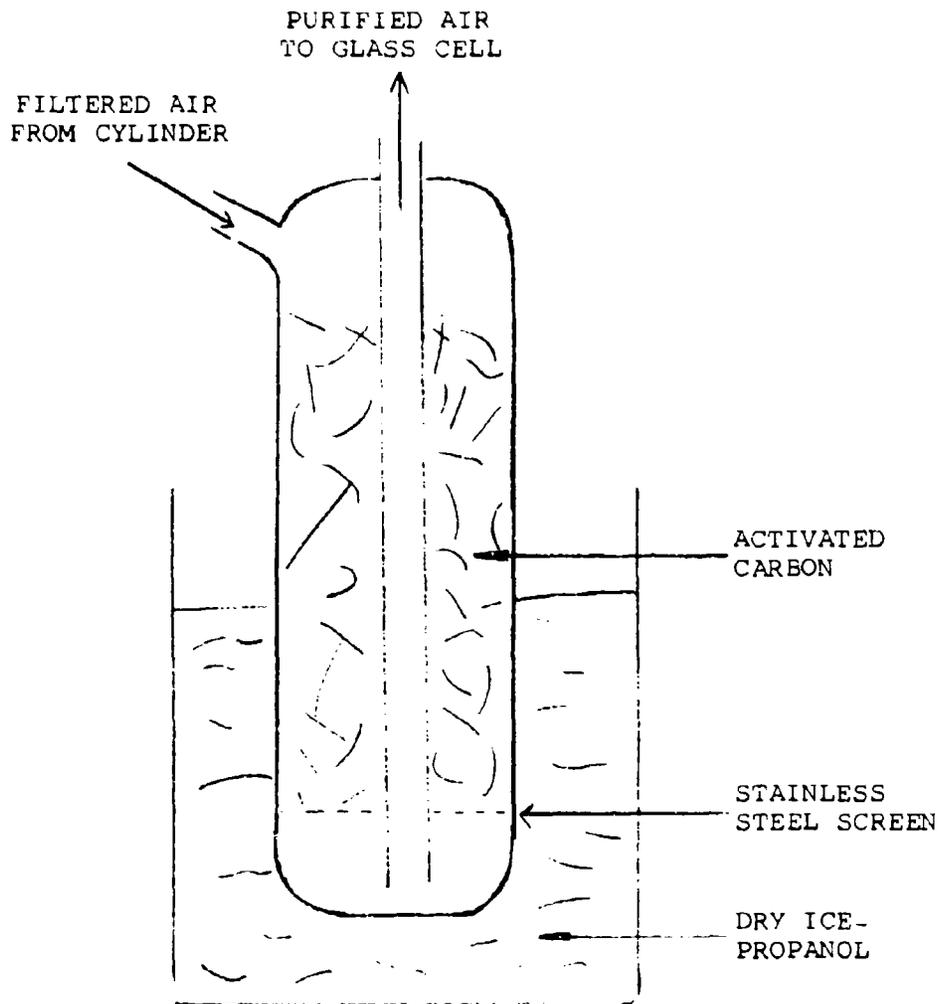


Figure I

CARBON BED FOR REMOVAL OF RESIDUAL ORGANIC VAPOR
FROM HOSPITAL GRADE CYLINDER AIR

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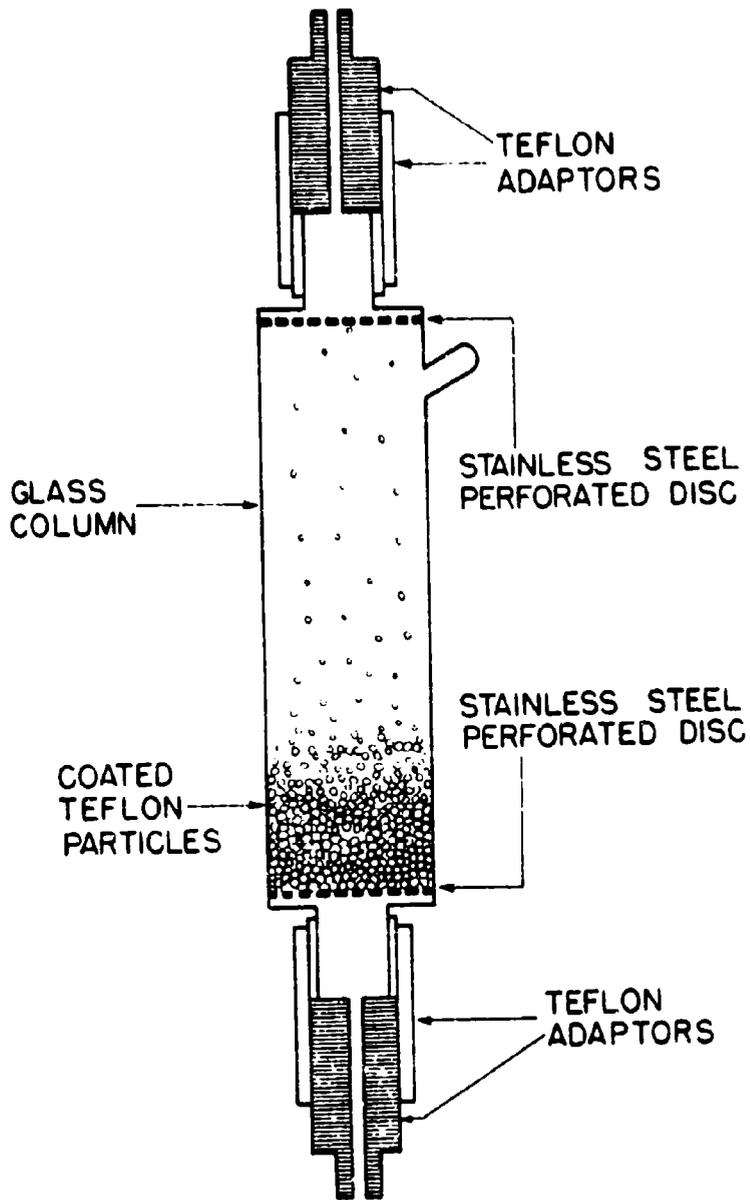


Figure 2
FLUIDIZED BED SAMPLER

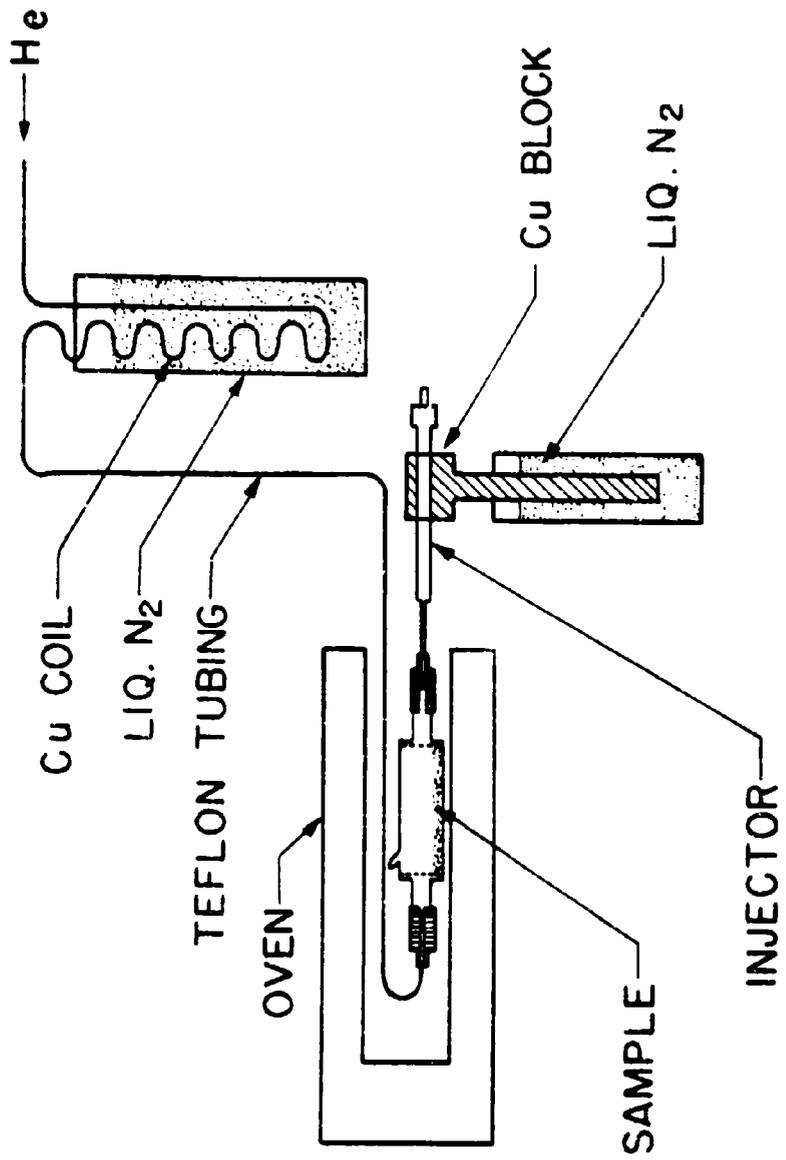
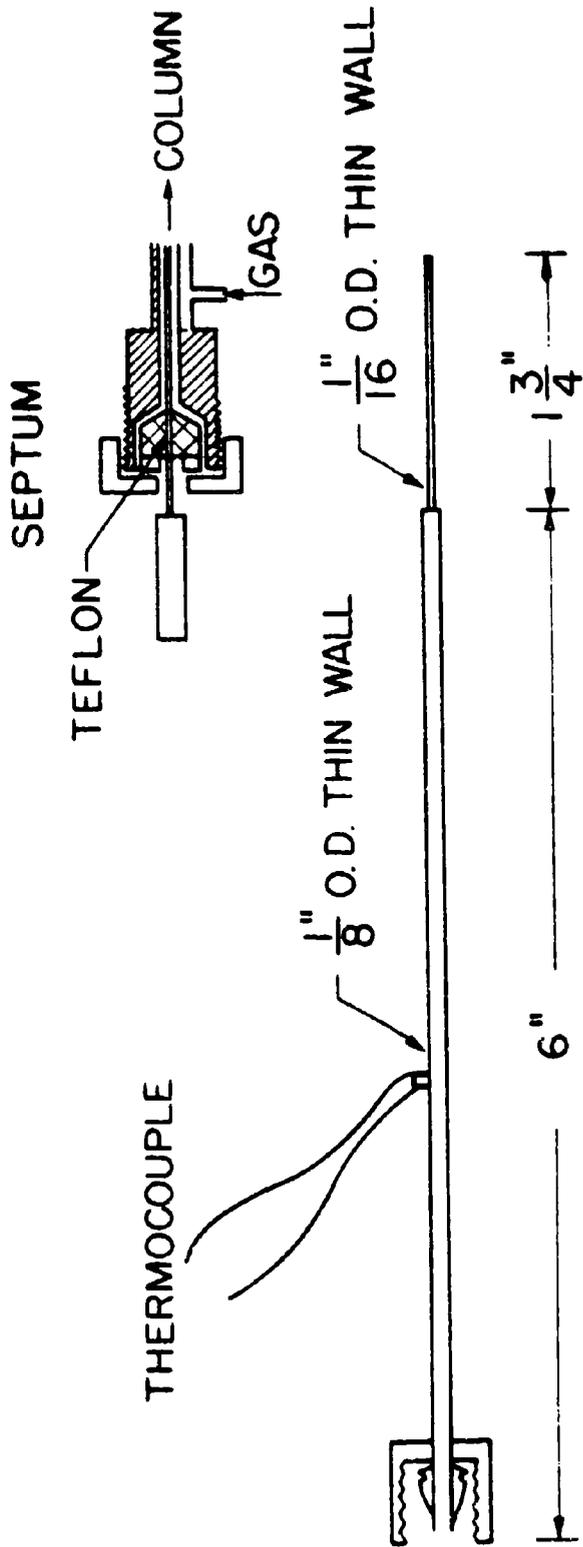


Figure 3
TRANSFER OF SAMPLE FROM BED SAMPLER TO INJECTOR



(Material : Stainless Steel)

Figure 4
SAMPLE INJECTOR

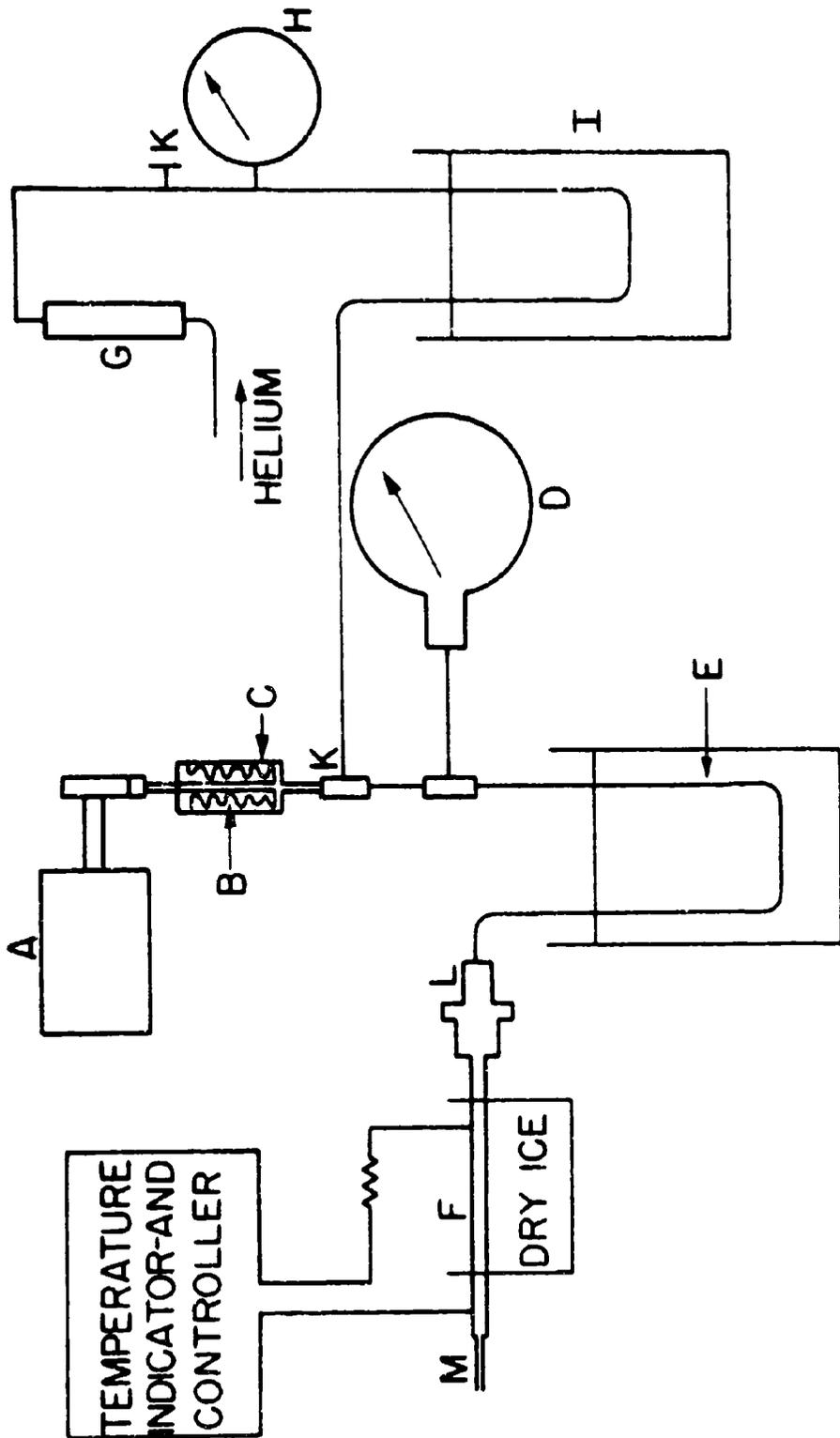


Figure 5
SAMPLE INJECTION SYSTEM

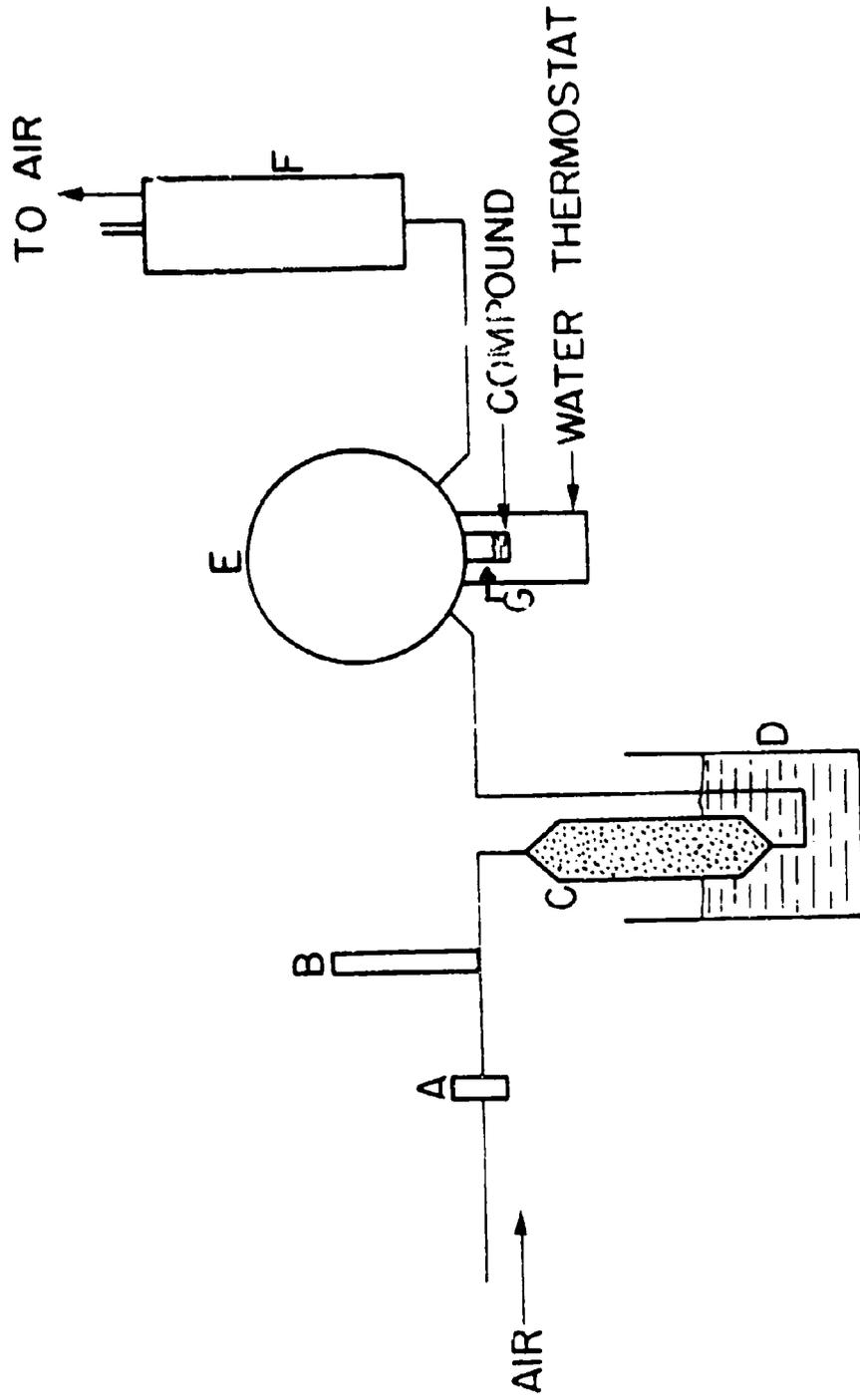


Figure 6
CALIBRATION SYSTEM

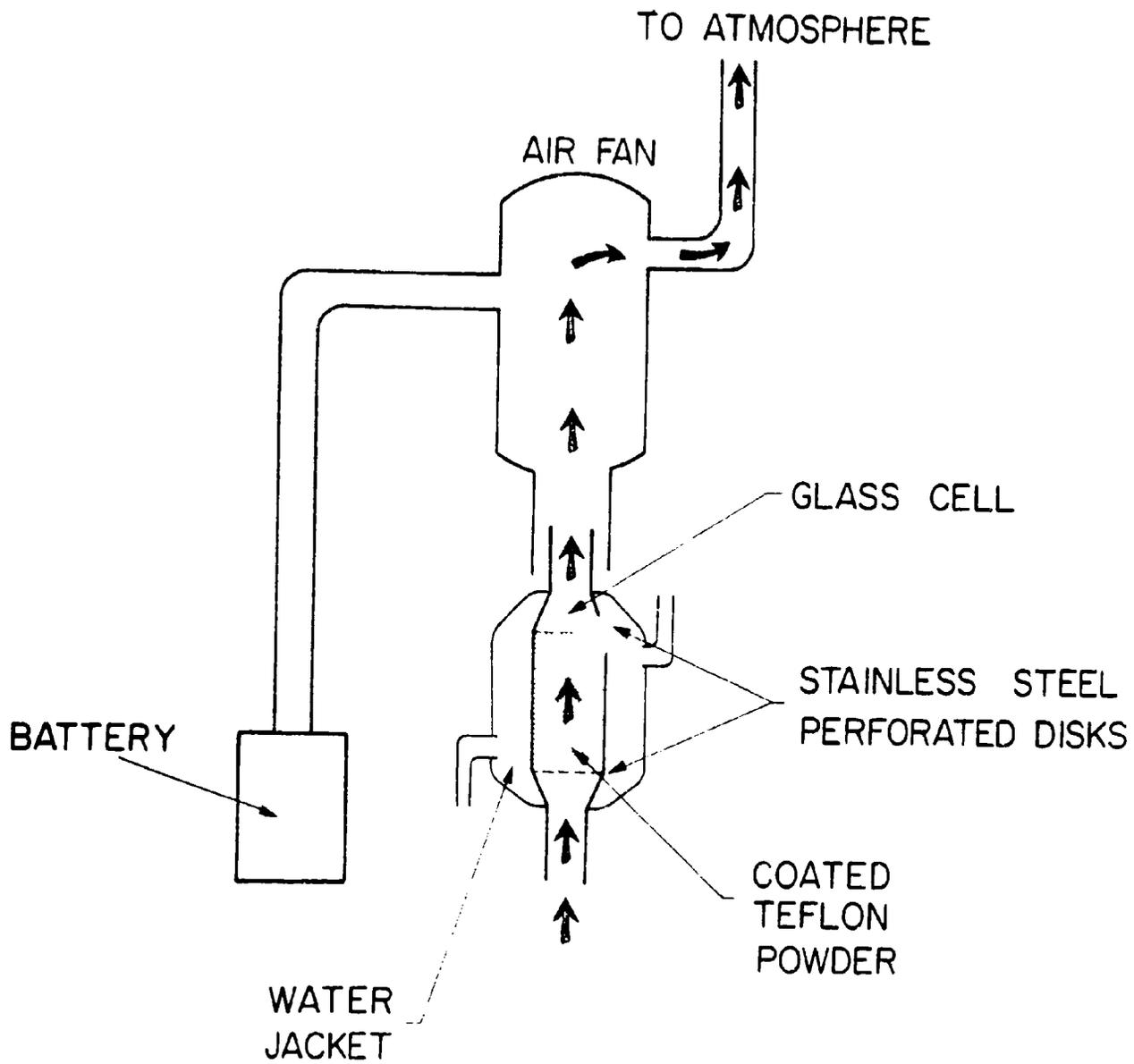


Figure 7
 PORTABLE AIR SAMPLING APPARATUS

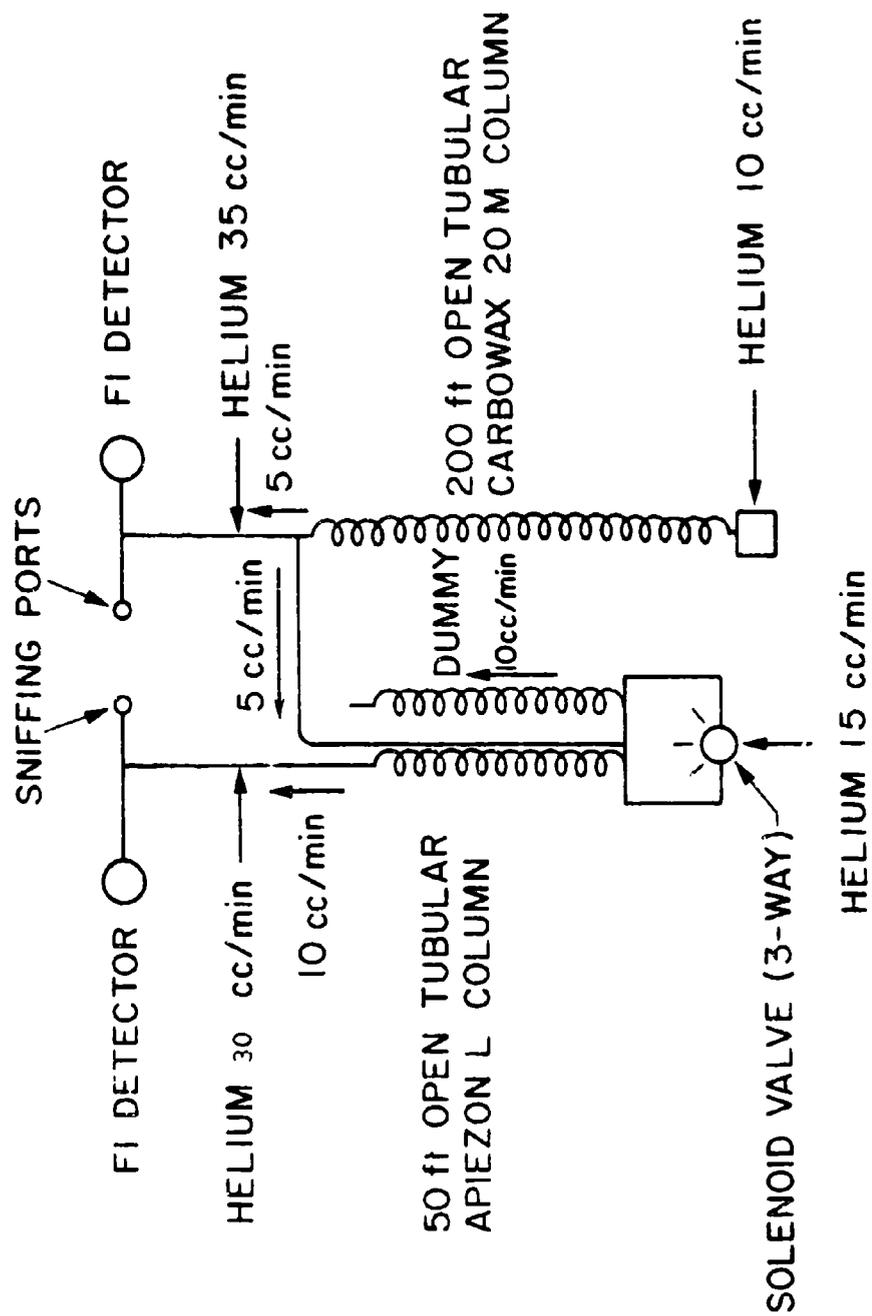


Figure 8
TWO-COLUMN APPARATUS

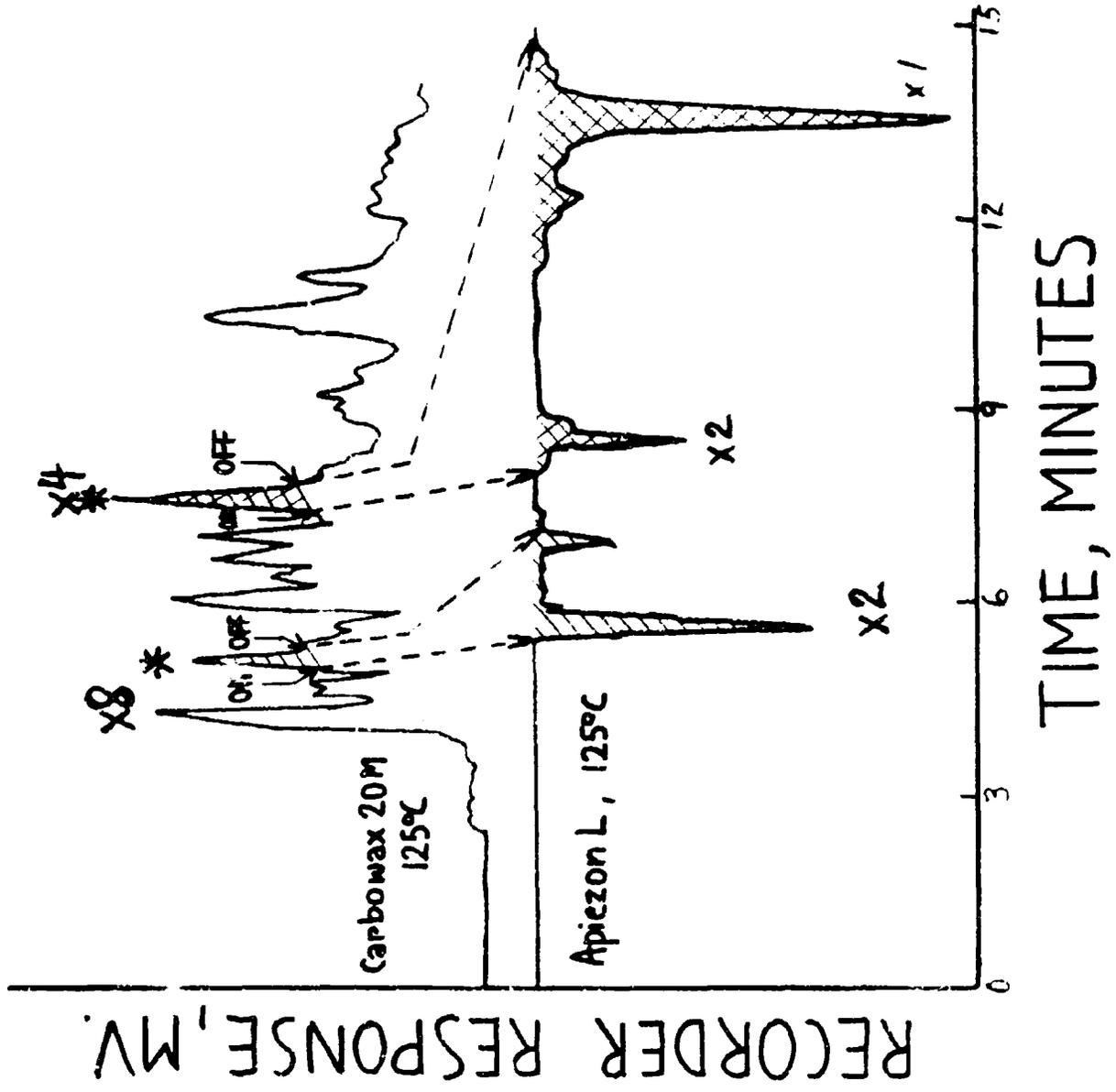


Figure 9
 SECONDARY RESOLUTION OF TWO HUMAN SIGNATURE COMPONENTS
 ON APIEZON L COLUMN OF TWO-COLUMN APPARATUS

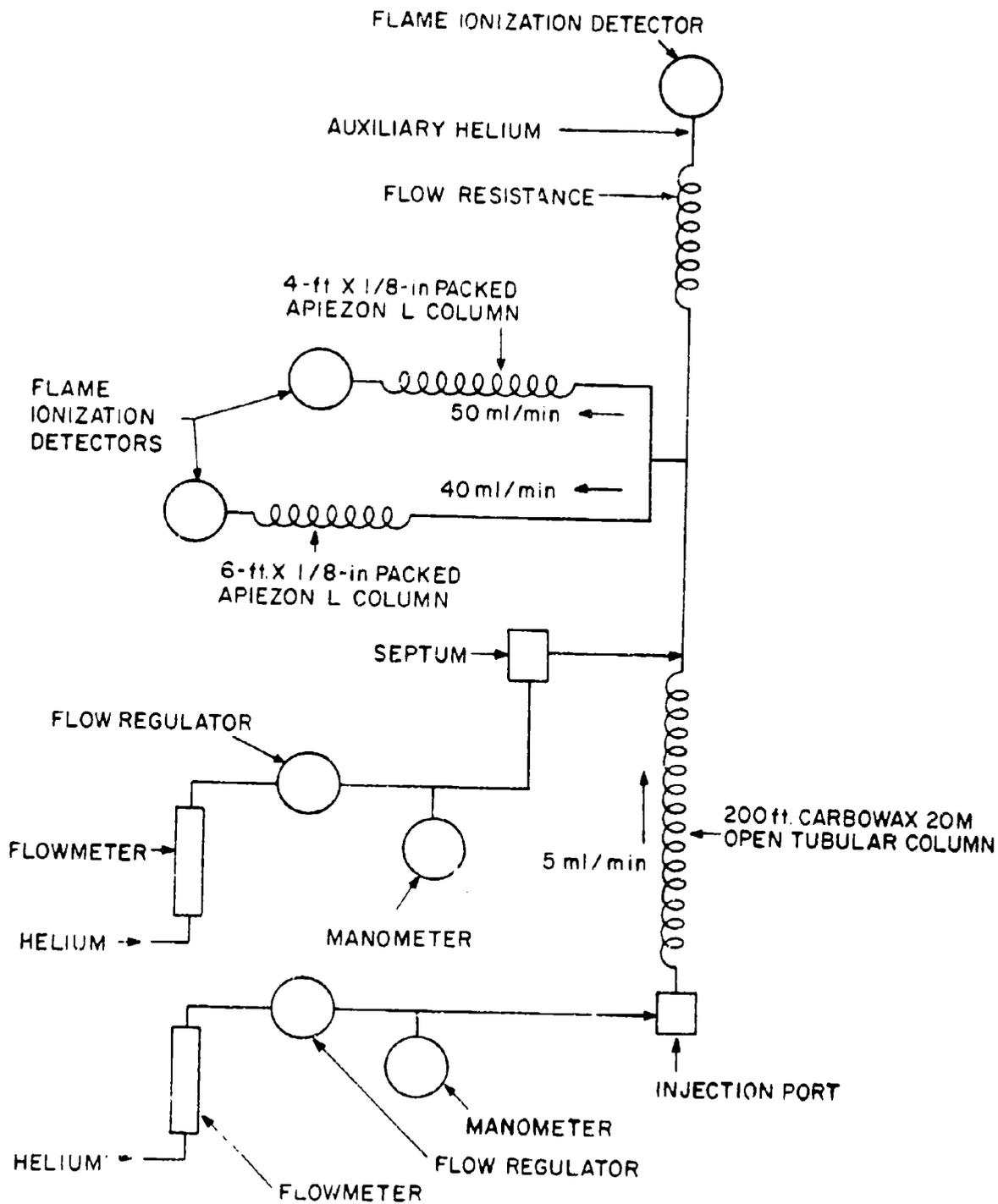


Figure 10
THREE COLUMN APPARATUS

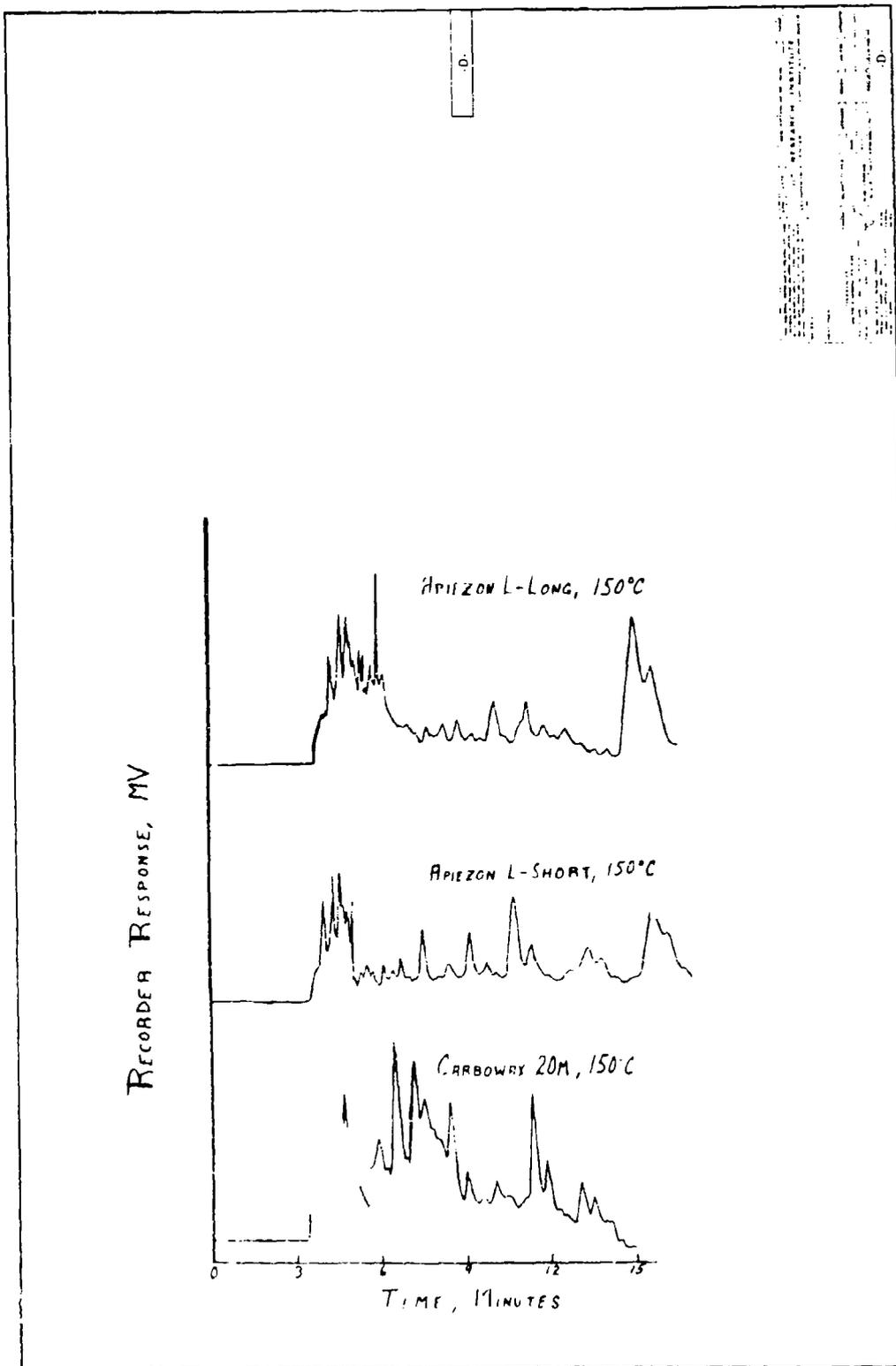


Figure 11
A SEGMENT OF A TYPICAL GAS CHROMATOGRAM OF
HUMAN CHEMICAL SIGNATURE OBTAINED ON THREE-COLUMN APPARATUS

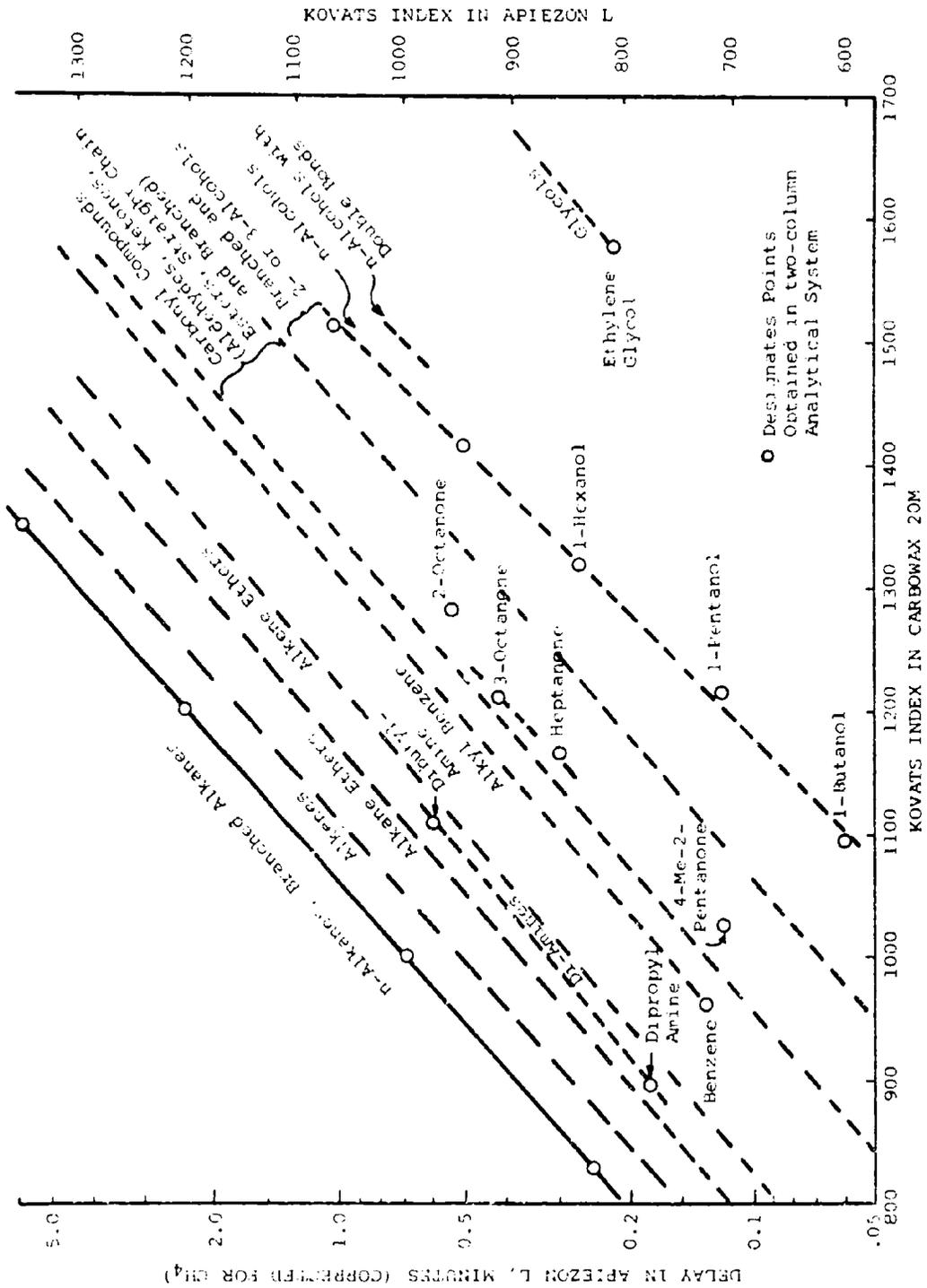


Figure 12
 CHEMICAL LOGIC FIELD FOR CLASSIFICATION OF DATA OBTAINED FROM THE TWO-COLUMN SYSTEM

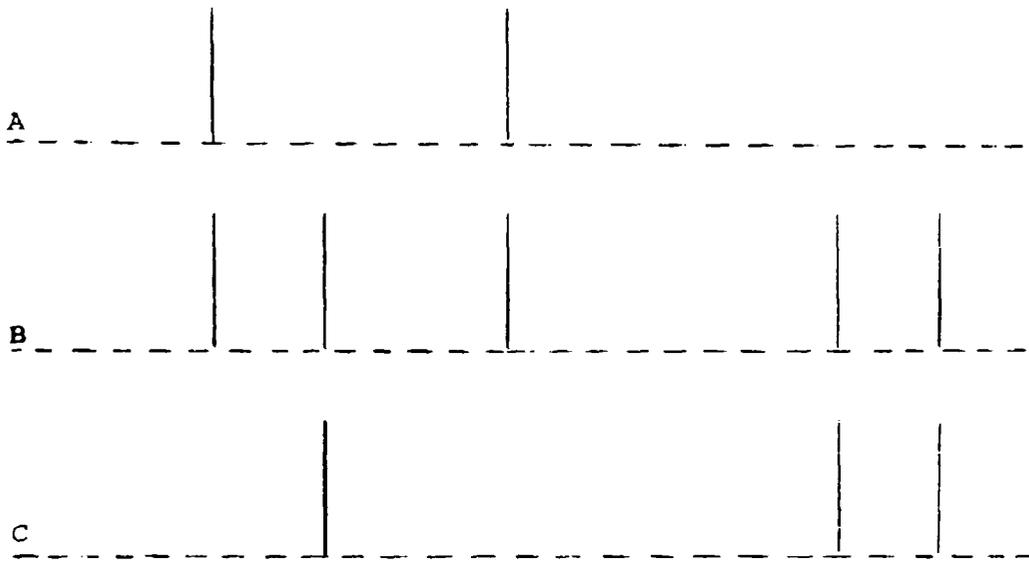


Figure 13

PROBLEMS IN PATTERN SIMILARITY COMPARISONS

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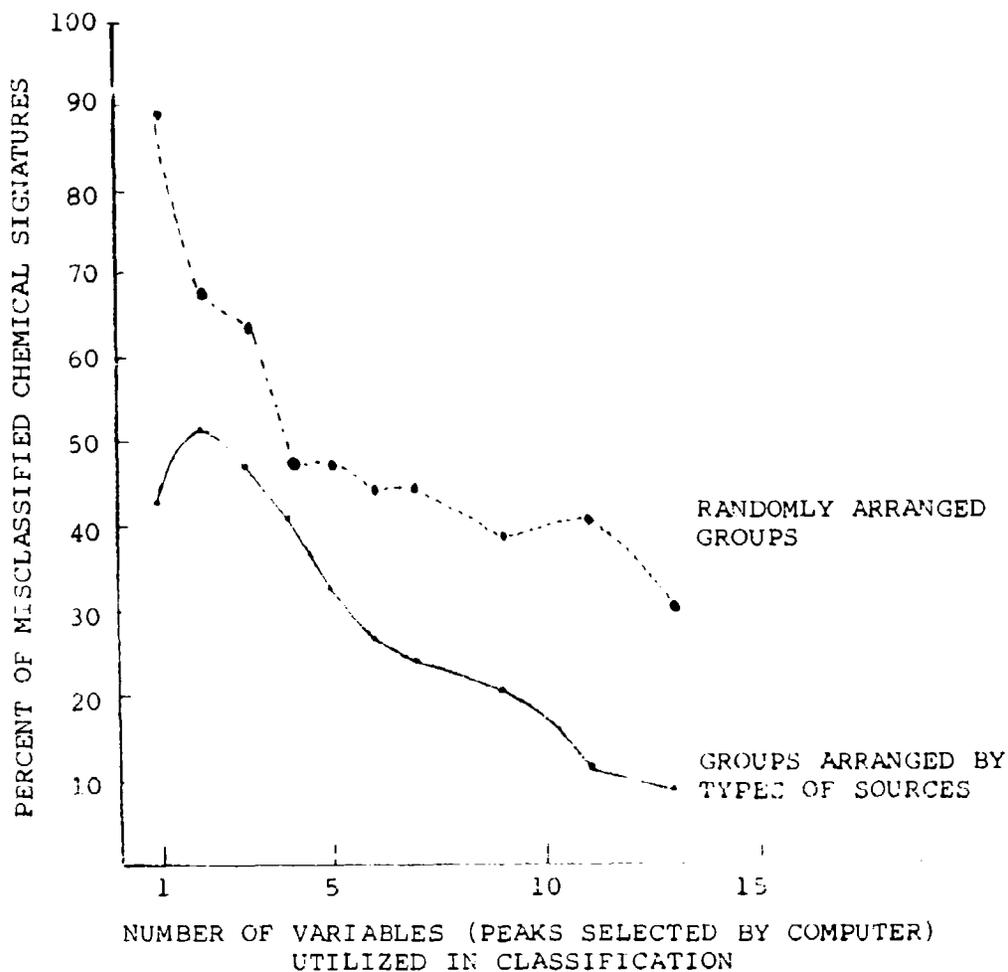


Figure 14

IMPROVEMENT IN CORRECT CLASSIFICATION OF INDIVIDUAL CHEMICAL SIGNATURES WITH INCREASE IN THE NUMBER OF COMPUTER-SELECTED PEAKS EMPLOYED IN THE CLASSIFICATION

Classification based on 49 signatures arranged in 5 groups.

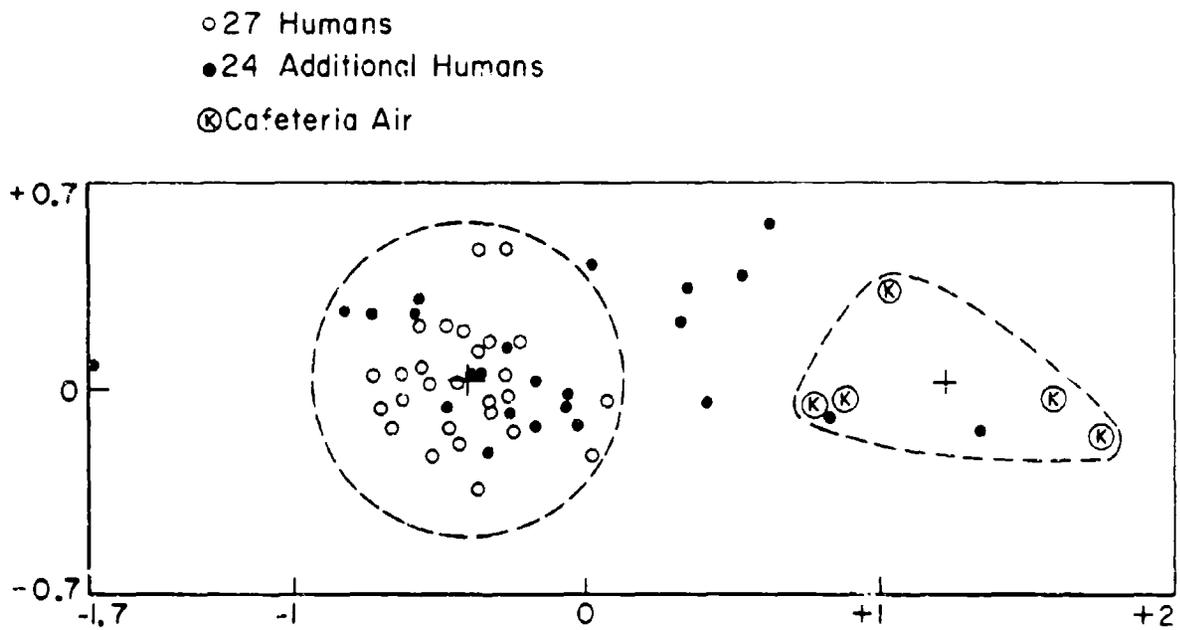
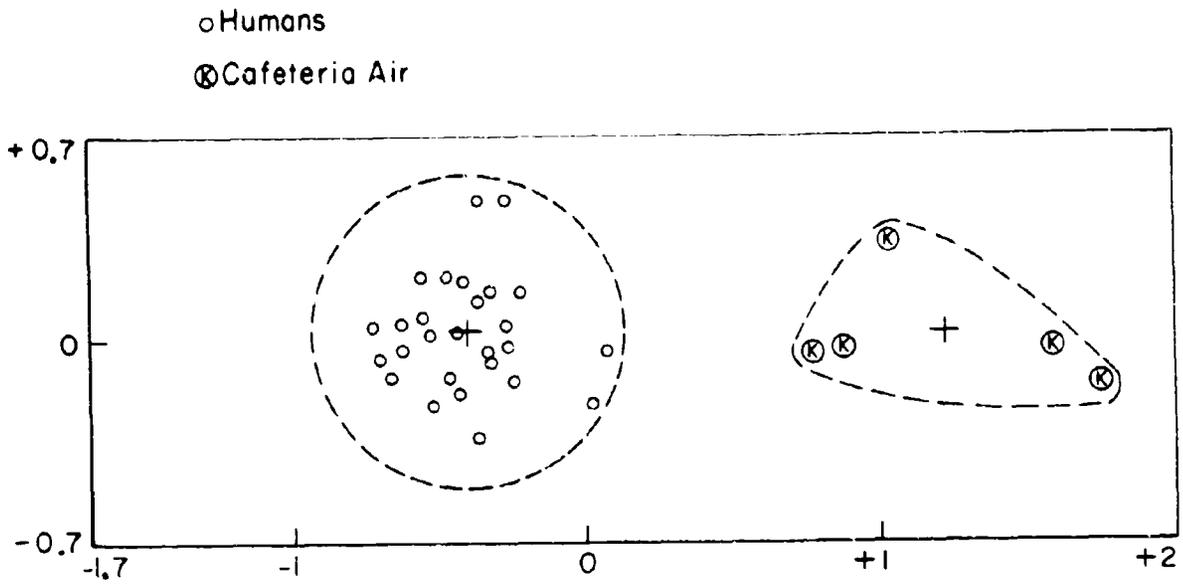


Figure 15

DISCRIMINANT FUNCTIONS FIELD FOR HUMANS VS. COOKING AREA VAPORS
AND CLASSIFICATION OF ADDITIONAL HUMANS USING DERIVED FUNCTIONS

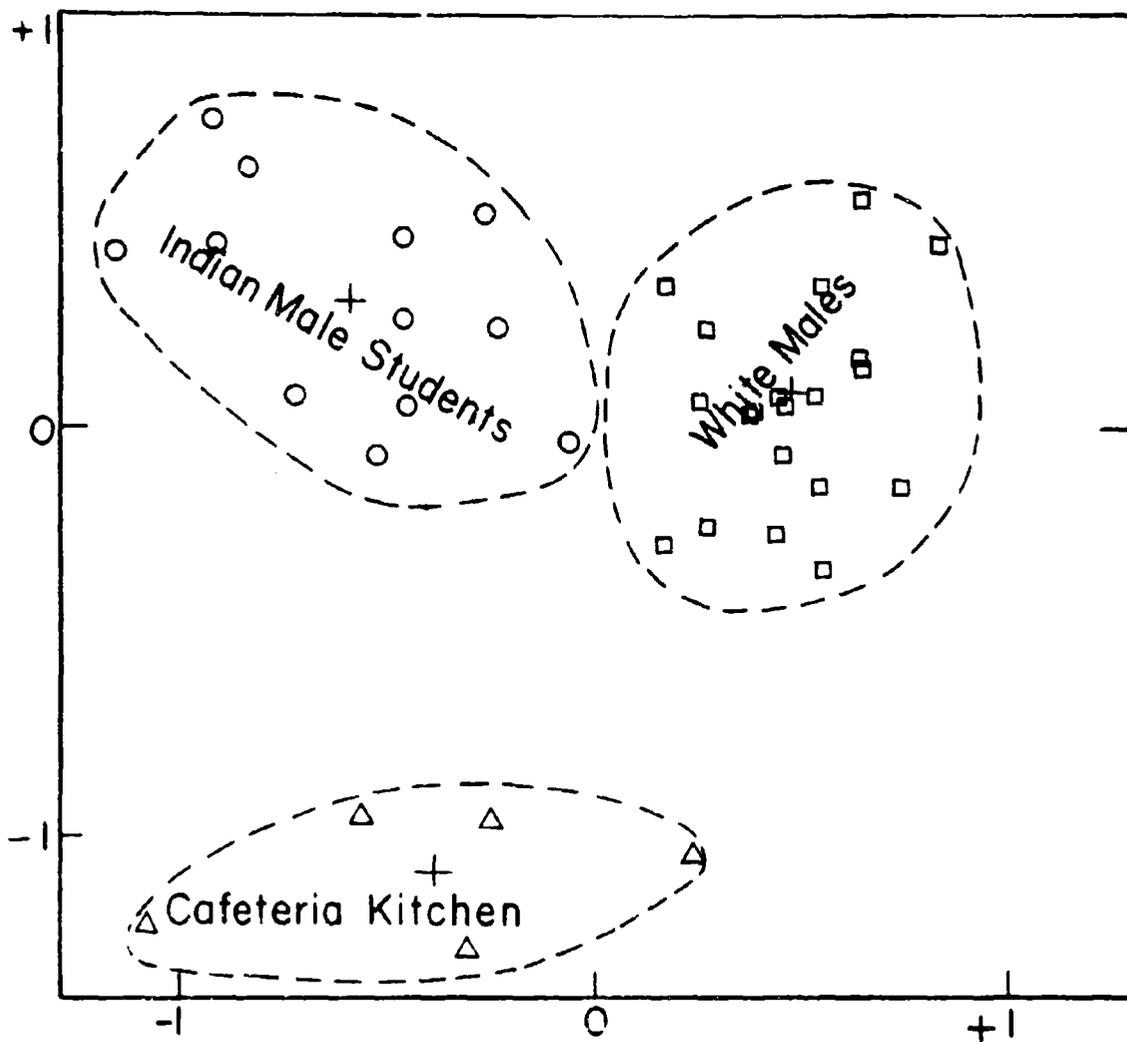


Figure 16
 STATISTICAL CLASSIFICATION PLANE FOR VAPOR
 CHROMATOGRAMS OF TWO TYPES OF MALES AND FOOD PREPARATION AREA

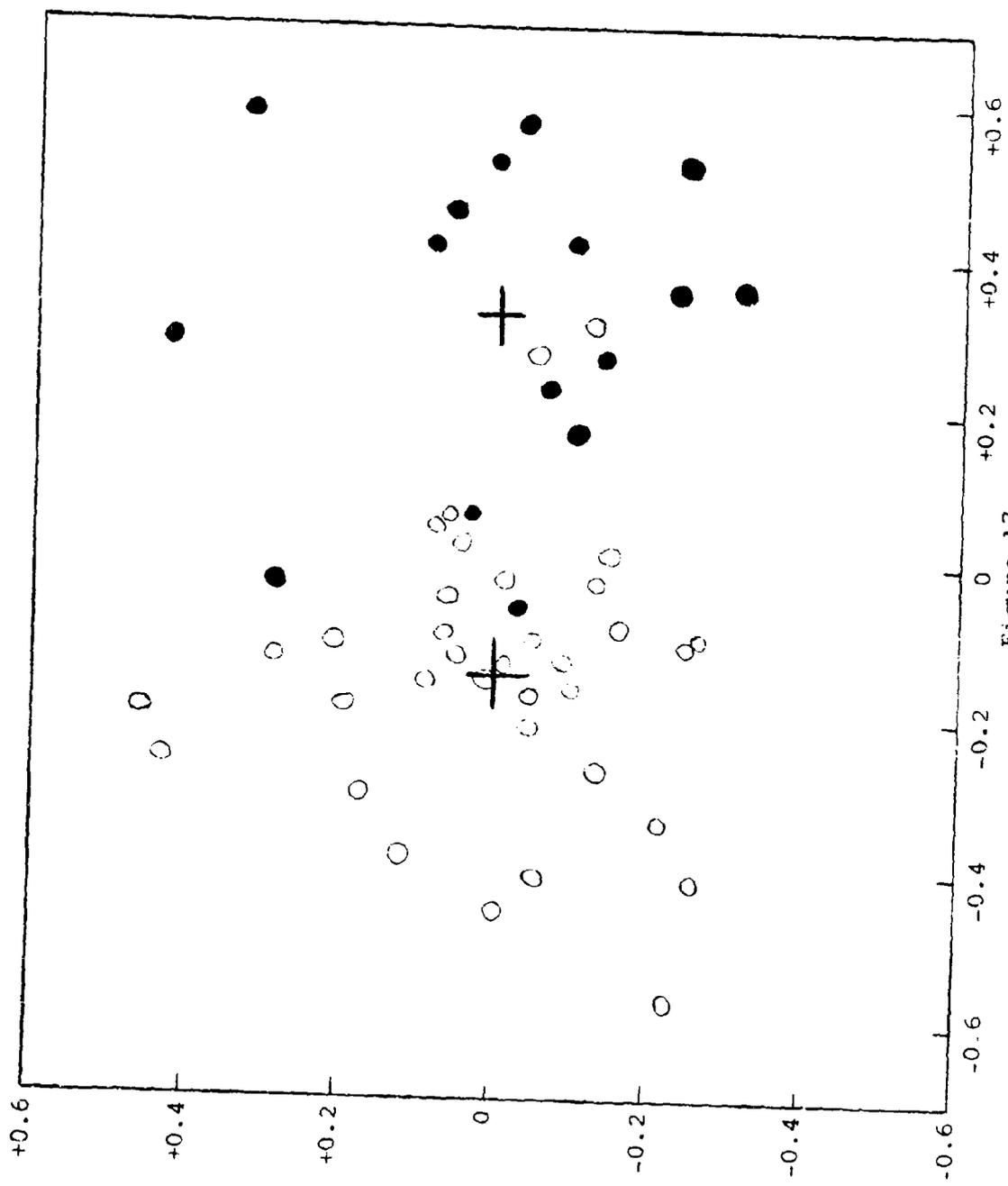


Figure 17

CLASSIFICATION FIELD FOR GLASS CELL EXPERIMENTS ONLY

Subjects: 35 males, 16 females

Variables: Input, 21. Utilized: 7. Crosses indicate the centers for groups

NOTES TO FIGURE 17

Wilcoxon-White ranking test, applied to the positions of points in the negative direction of X discriminant function only yielded 162 as the rank sum for females (rank sum for the smallest group). The critical point for error probability of 0.1 percent is 252. Thus, the validity of the X function in classification of males versus females has considerably less than 1:1000 chance of being in error.

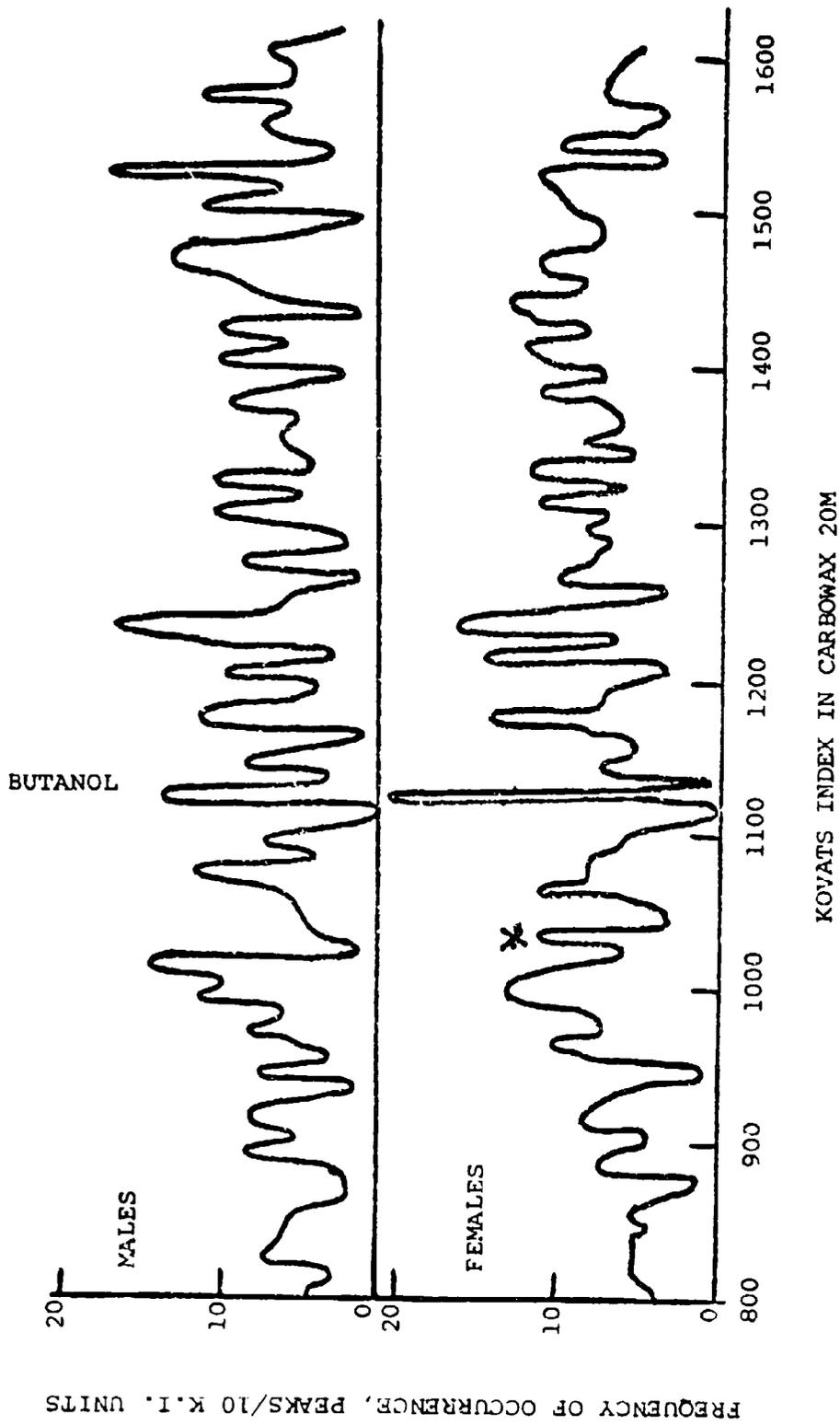


Figure 18
 DENSITY OF OCCURRENCE OF PEAKS IN CHEMICAL SIGNATURES
 OF CAUCASIAN MALES AND FEMALES ON A KOVATS INDEX PLOT

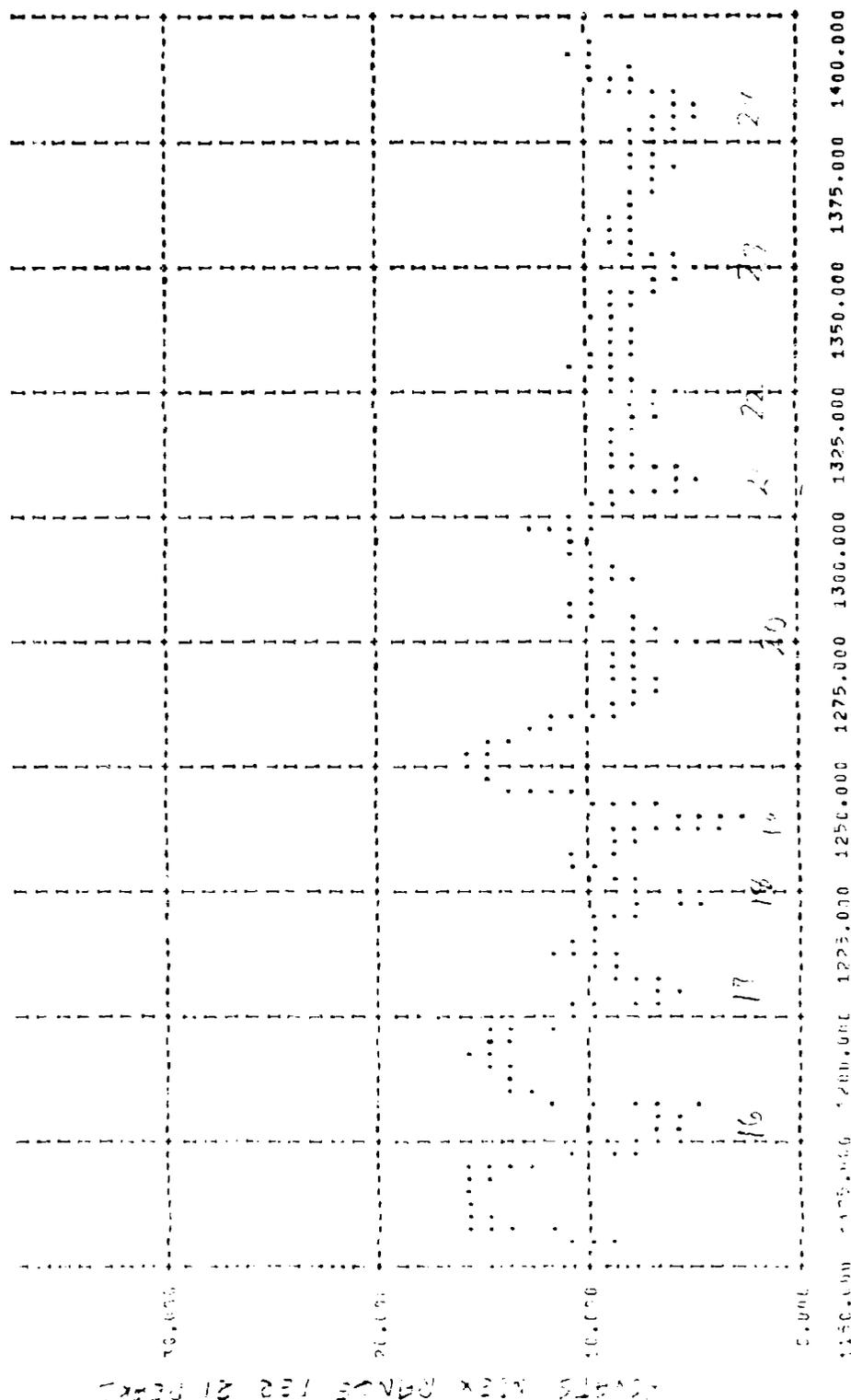


Figure 19
 COMPUTER PRINTOUT OF HISTOGRAM OF PEAK OCCURRENCE
 FREQUENCY IN KOVATS INDEX RANGE BETWEEN 1150 AND 1400 UNITS
 IN FOOLED CHROMATOGRAMS FROM ALL HUMANS STUDIED

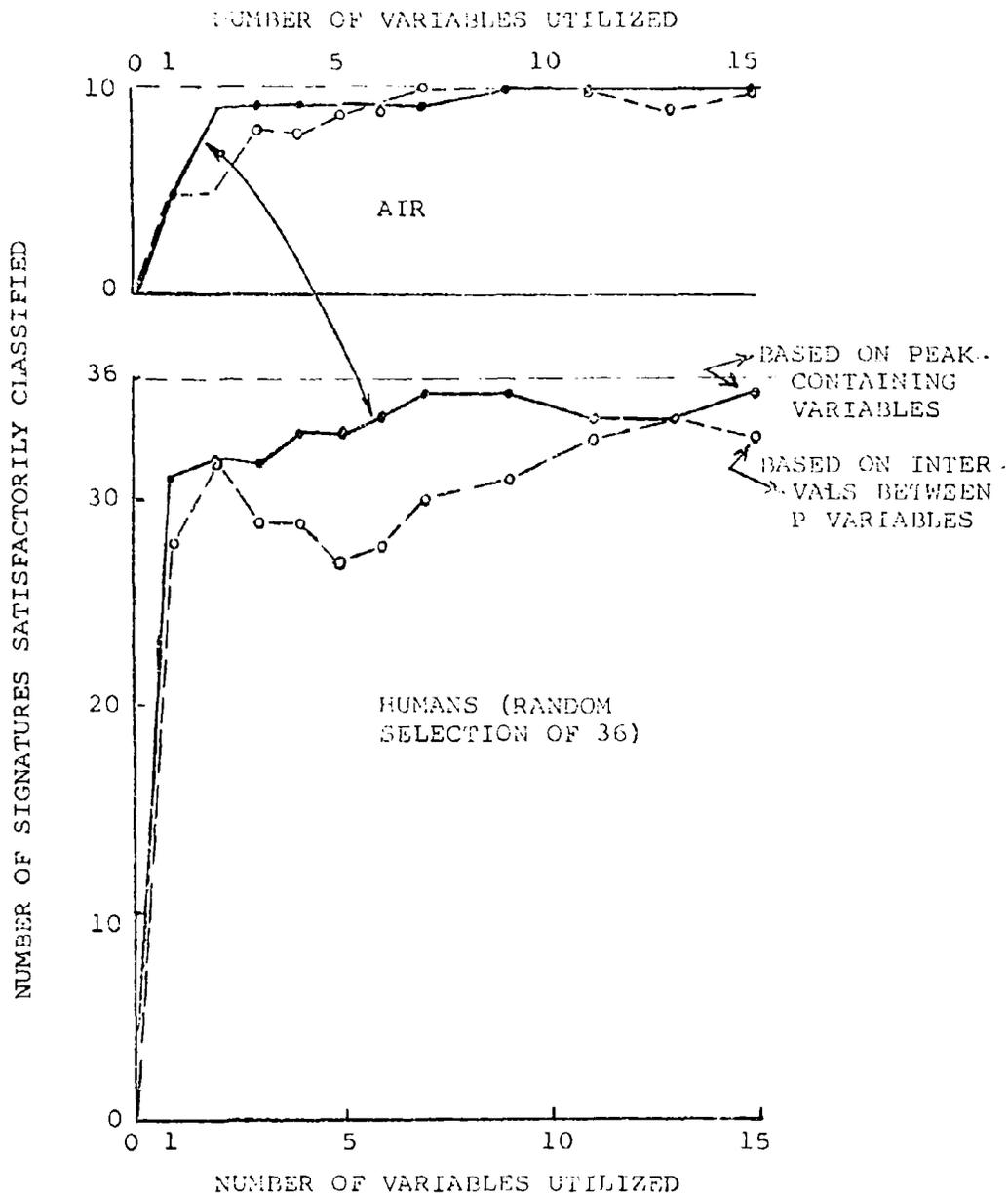


Figure 20
 COMPARISON IN THE IMPROVEMENT OF TWO-WAY CLASSIFICATION
 OF CHEMICAL SIGNATURES: USING VARIABLES (KOVATS INDEX RANGES)
 THAT FREQUENTLY CONTAIN PEAKS IN HUMAN SIGNATURES
 (P-VARIABLES, FULL LINE), OR VARIABLES THAT CORRESPOND TO
 RANGES IN BETWEEN (DASHED LINE)

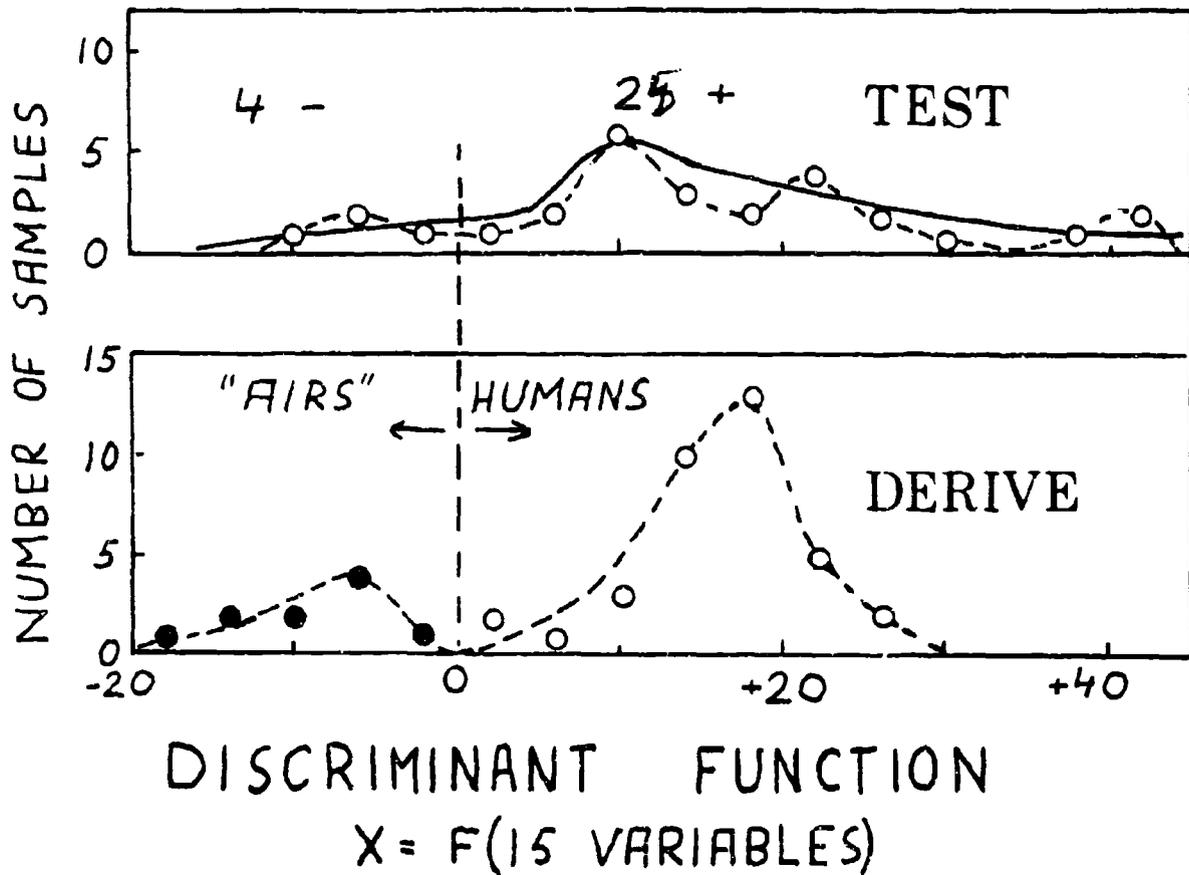


Figure 21

DERIVATION OF DISCRIMINANT FUNCTION FOR CLASSIFICATION
 OF HUMAN AND ENVIRONMENT CHEMICAL SIGNATURES
 AND TEST OF THE FUNCTION ON 29 ADDITIONAL HUMAN CHEMICAL SIGNATURES

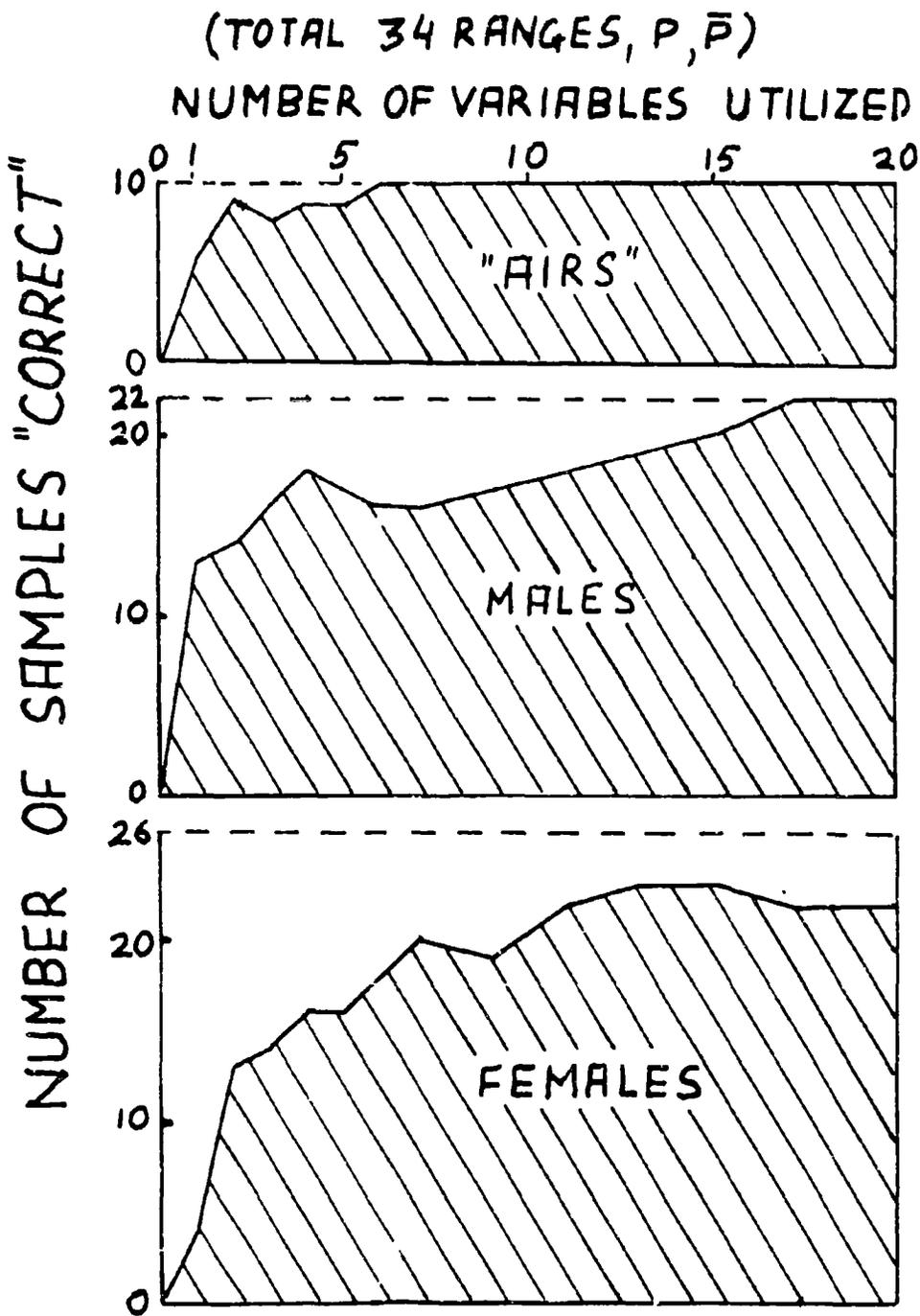


Figure 22
 IMPROVEMENT IN CORRECT PATTERN ASSIGNMENTS WITH
 INCREASE IN THE NUMBER OF VARIABLES UTILIZED BY THE
 STEPWISE DISCRIMINANT ANALYSIS PROGRAM

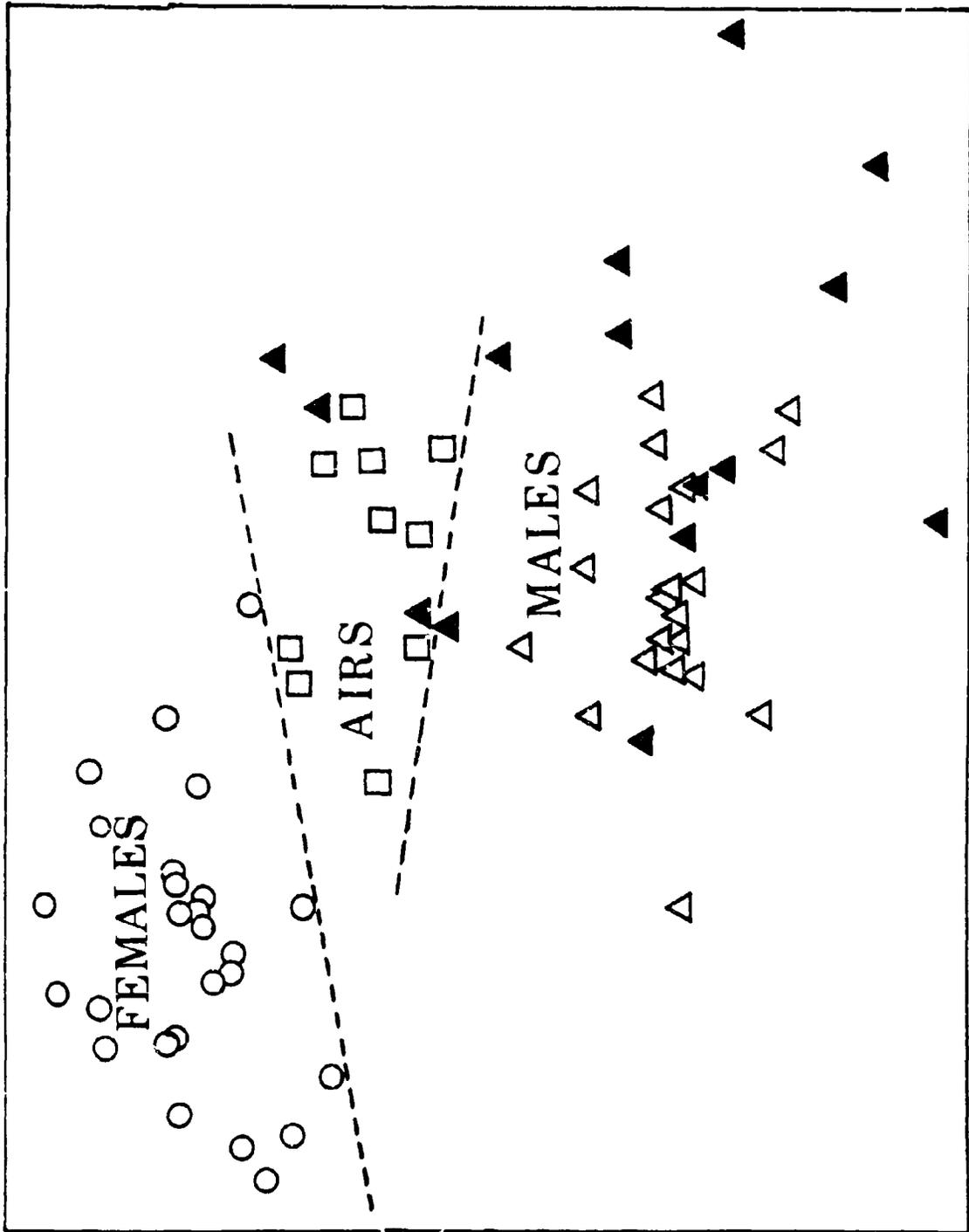


Figure 23

DERIVATION OF THREE-CLASS DISCRIMINANT FUNCTIONS FIELD (HOLLOW SYMBOLS) AND TEST OF THE DERIVED FUNCTIONS WITH ADDITIONAL MALE CHEMICAL SIGNATURE PATTERNS (FILLED TRIANGLES)

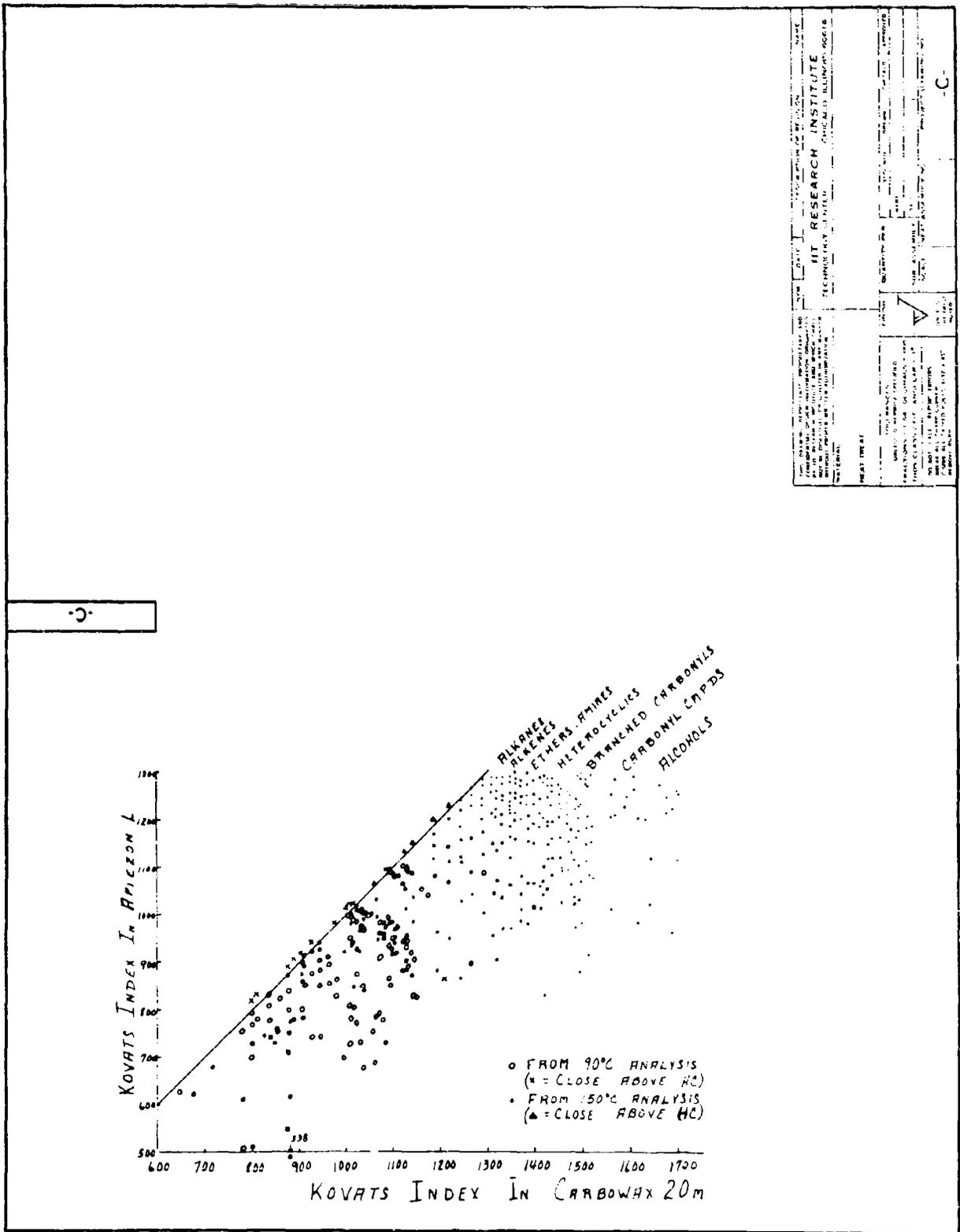


Figure 24
 THREE-COLUMN DATA INTERPRETATION ASSISTED BY GA3COL
 COMPUTER PROGRAM
 SUBJECT: CAUCASIAN FEMALE L.B.

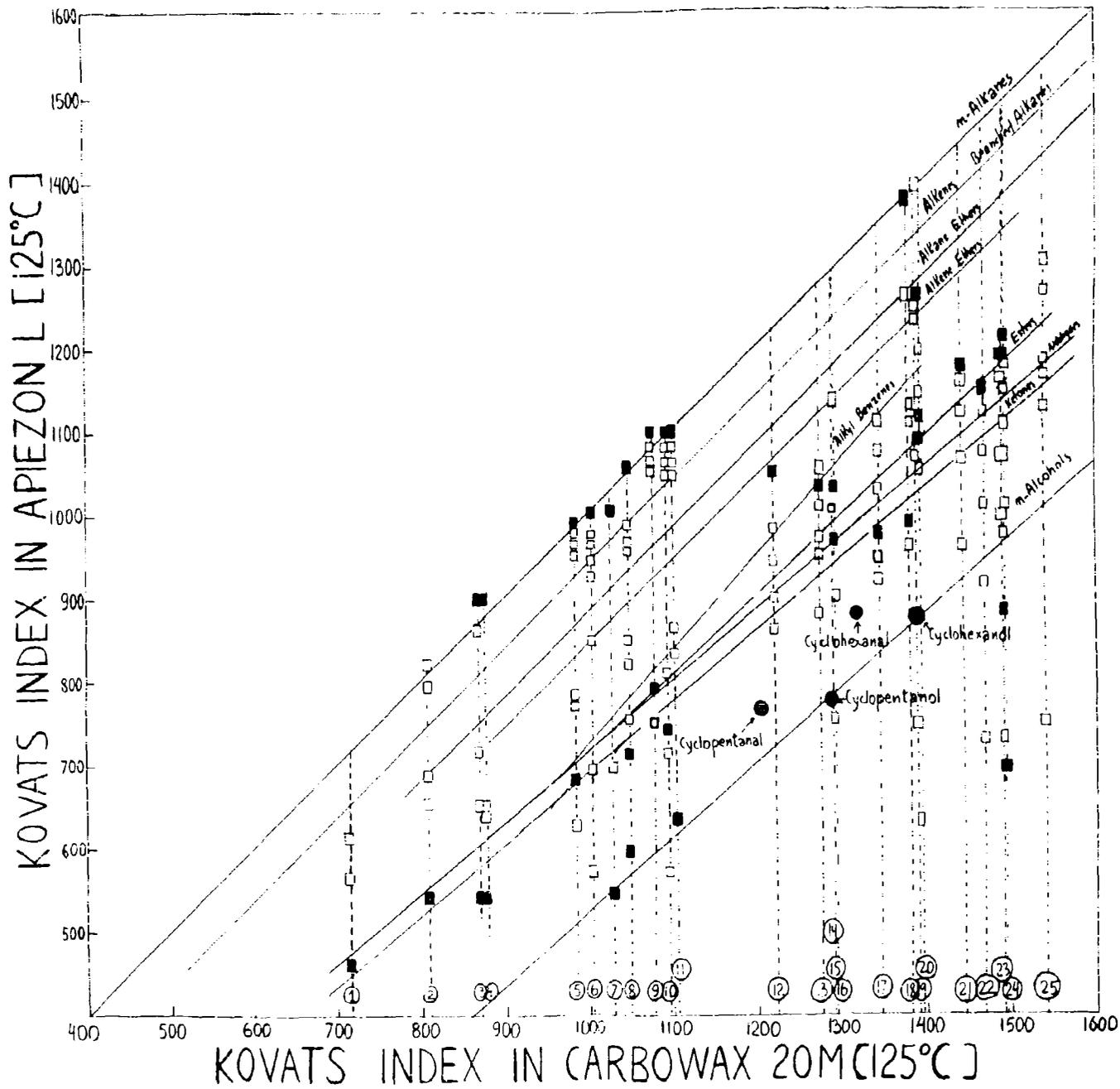


Figure 25
 DISPLAY OF TWO-COLUMN DATA ON HUMAN CHEMICAL SIGNATURES

APPENDIX I

Tabulation of Chemical Signature Data

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EXPLANATION OF ARRANGEMENT

Human chemical signatures are arranged in numbered order, approximating the chronological order in which the data were calculated. The environment chemical signatures begin after the human chemical signature No. 69.

The tabulation is a copy of the computer printout.

The upper line in each pattern describes the source, ("SUBJECT"), gives the consecutive pattern number (one per pattern), and initials if the source was a human subject.

The second line is the data of the chemical signature collection.

The third line (Total Intensity) gives the total area, in sq. mm. at 0.1 x 1 electrometer setting, of all gas chromatographic peaks of the particular pattern, and the mean peak intensity (total area of the pattern's peaks divided by the number of peaks in the same pattern).

The columns in the tabulation are as follows:

- (1) First column- consecutive number of the peak in the particular pattern.
- (2) Second column - retention time in terms of Kovats retention index-cf. text, section 3.6.5.
- (3) Third column- the area of the peak, in sq. mm. calculated to the electrometer sensitivity setting of 0.1 x 1.
- (4) Fourth column- normalized area, equal to the area of the particular peak divided by the "Total Intensity"-- sum of areas of all peaks in the same pattern.

NOT REPRODUCIBLE

SUBJECT: ... JAD
 DATE: ...
 TOTAL INTENSITY: ...

AREA: ... 1961.

YEAR	RETENTION TIME	AREA	PERCENTAGE	PER AREA	NORMALIZED AREA
1	574.04	5.74	22	12.00	2.000
2	585.21	1.97	24	31.00	6.836
3	596.38	1.03	24	20.00	9.854
4	607.55	0.37	25	52.00	1.140
5	618.72	1.04	26	48.00	1.092
6	629.89	7.85	27	24.00	9.526
7	641.06	7.55	28	12.00	7.639
8	652.23	3.44	29	18.00	6.281
9	663.40	1.74	30	25.00	6.954
10	674.57	2.37	31	17.00	6.253
11	685.74	0.97	32	13.00	3.016
12	696.91	0.97	33	5.75	1.752
13	708.08	1.03	34	24.00	1.150
14	719.25	1.21	35	43.25	0.947
15	730.42	0.32	36	11.00	3.224
16	741.59	0.32	37	47.50	9.223
17	752.76	7.01	38	29.70	6.524
18	763.93	1.03	39	1.80	6.255
19	775.10	5.32	40	25.00	6.570
20	786.27	2.17	41	30.75	6.656
21	797.44	0.03	42	30.00	6.789
22	808.61	0.03	43	24.00	6.526

SUBJECT: ... BD
 DATE: ...
 TOTAL INTENSITY: ...

AREA: ...

YEAR	RETENTION TIME	AREA	PERCENTAGE	PER AREA	NORMALIZED AREA
1	574.04	1.04	20	24.00	6.710
2	585.21	1.97	21	11.00	4.315
3	596.38	0.37	22	32.00	9.272
4	607.55	0.37	23	26.00	6.758
5	618.72	1.04	24	20.00	6.092
6	629.89	7.85	25	9.00	8.642
7	641.06	7.55	26	7.00	2.131
8	652.23	3.44	27	10.00	6.064
9	663.40	1.74	28	12.00	6.414
10	674.57	2.37	29	12.00	6.379
11	685.74	0.97	30	24.00	2.310
12	696.91	0.97	31	13.00	2.250
13	708.08	1.03	32	13.00	2.250
14	719.25	1.21	33	5.75	1.404
15	730.42	0.32	34	54.00	1.598
16	741.59	0.32	35	14.00	5.426
17	752.76	7.01	36	5.75	1.770
18	763.93	1.03	37	9.00	2.664
19	775.10	5.32	38	36.00	1.125
20	786.27	2.17	39	7.00	2.974

KS

SUBJECT
DATE

10-3-65

10-3-65

MALE

ES

35316

947

NOT REPRODUCIBLE

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	524.96	532.	1.441	23	1217.76	244.	0.268
2	587.21	104.	0.282	24	1304.44	80.	1.074
3	635.20	303.	1.517	25	1319.02	390.	4.995
4	727.65	305.	8.274	26	1335.48	51.	0.643
5	815.62	815.	2.210	27	1354.19	50.	0.633
6	856.61	603.	2.167	28	1367.69	100.	2.265
7	862.54	285.	0.740	29	1381.70	177.	2.629
8	905.87	250.	6.435	30	1394.39	50.	0.653
9	967.32	480.	1.390	31	1406.07	163.	2.048
10	967.83	512.	1.387	32	1422.43	32.	4.076
11	967.23	724.	6.065	33	1434.18	116.	1.450
12	1007.88	440.	1.303	34	1452.39	170.	2.209
13	1072.53	888.	2.495	35	1461.26	44.	0.262
14	1121.91	720.	1.955	36	1488.24	44.	0.262
15	1164.53	852.	2.275	37	1507.45	56.	0.851
16	1181.49	544.	1.847	38	1521.94	228.	2.851
17	1202.04	32.	0.087	39	1541.48	100.	1.265
18	1223.49	2015.	5.461	40	1547.60	112.	0.141
19	1244.21	1843.	4.984	41	1575.44	96.	1.175
				42	1597.44	260.	3.264
				43	1625.57	345.	4.357
				44	1642.14	960.	1.205

SUBJECT 10-3-65 MALE ES
DATE 10-3-65
TOTAL INTENSITY: 35316

947

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	524.96	532.	1.441	20	1409.64	1240.	3.301
2	587.21	104.	0.282	21	1506.96	204.	0.715
3	635.20	303.	1.517	22	1522.23	250.	0.935
4	727.65	305.	8.274	23	1547.75	236.	0.693
5	815.62	815.	2.210	24	1567.58	400.	1.084
6	856.61	603.	2.167	25	1579.81	1150.	3.240
7	862.54	285.	0.740	26	1610.77	576.	1.590
8	905.87	250.	6.435	27	1631.57	259.	0.721
9	967.32	480.	1.390	28	1652.20	144.	0.350
10	967.83	512.	1.387	29	1666.75	150.	0.257
11	967.23	724.	6.065	30	1679.04	360.	0.975
12	1007.88	440.	1.303	31	1686.76	128.	0.347
13	1072.53	888.	2.495	32	1511.79	472.	1.279
14	1121.91	720.	1.955	33	1525.67	2280.	6.198
15	1164.53	852.	2.275	34	1539.82	780.	2.113
16	1181.49	544.	1.847	35	1587.30	184.	0.453
17	1202.04	32.	0.087	36	1581.75	300.	0.813
18	1223.49	2015.	5.461	37	1602.10	182.	0.385
19	1244.21	1843.	4.984	38	1638.90	360.	0.975
				39	1625.57	1792.	4.854

NOT REPRODUCIBLE

SUBJECT 10014
 DATE 10/25/58
 TOTAL INTENSIVE 14925

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	1264.07	457	0.210
2	1275.78	670	2.777
3	1279.13	1870	7.041
4	1280.19	147	0.624
5	1274.55	144	0.624
6	1305.20	384	1.678
7	1325.32	816	3.266
8	1343.05	192	0.839
9	1363.22	352	1.520
10	1352.74	384	1.678
11	1409.41	792	3.266
12	1426.49	960	4.195
13	1463.25	1152	5.035
14	1507.65	1176	5.113
15	1523.52	936	4.091
16	1538.11	680	2.972
17	1579.52	360	1.573
18	1601.69	144	0.624
19	1625.57	972	4.335

NOT REPRODUCIBLE

SUBJECT 10014
 DATE 10/25/58
 TOTAL INTENSIVE 14925

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	1264.07	457	0.210
2	1275.78	670	2.777
3	1279.13	1870	7.041
4	1280.19	147	0.624
5	1274.55	144	0.624
6	1305.20	384	1.678
7	1325.32	816	3.266
8	1343.05	192	0.839
9	1363.22	352	1.520
10	1352.74	384	1.678
11	1409.41	792	3.266
12	1426.49	960	4.195
13	1463.25	1152	5.035
14	1507.65	1176	5.113
15	1523.52	936	4.091
16	1538.11	680	2.972
17	1579.52	360	1.573
18	1601.69	144	0.624
19	1625.57	972	4.335

NOT REPRODUCIBLE

Subject	INDICA	MALE	MEH	DATE	11-12-56	14145.	479.
TOTAL INTENSIVE	RETENTION TIME	PEAK AREA	NORMALIZED AREA	RETENTION TIME	PEAK AREA	NORMALIZED AREA	
1	473.59	95.	0.479	1656.50	40.	0.397	
2	574.58	64.	0.334	1624.84	40.	0.158	
3	589.15	75.	0.397	1639.62	100.	1.588	
4	651.48	72.	0.375	1756.14	60.	0.555	
5	711.21	364.	1.901	1669.34	120.	1.191	
6	751.80	589.	3.071	1309.92	72.	0.715	
7	797.95	200.	1.045	1326.47	40.	0.382	
8	826.78	800.	4.597	1550.14	70.	0.695	
9	832.43	615.	3.218	1371.05	24.	0.238	
10	911.30	319.	1.665	1383.14	140.	1.171	
11	931.70	280.	1.503	1411.56	44.	0.407	
12	964.04	420.	2.194	1466.18	270.	2.660	
13	1002.22	392.	2.048	1477.65	25.	2.322	
14	1005.35	624.	3.259	1494.23	40.	0.397	
15	1032.80	476.	2.486	1509.25	104.	1.032	
16	1063.45	620.	3.233	1523.85	164.	1.632	
17	1093.16	368.	1.922	1540.90	486.	4.863	
18	1121.91	700.	3.655	1562.29	260.	2.580	
19	1163.66	480.	2.507	1581.28	440.	4.407	
20	1208.25	144.	0.757	1598.93	324.	3.216	
21	1123.37	140.	1.384	1625.57	1590.	15.900	

SUBJECT	INDICA	MALE	MEH	DATE	11-12-56	14145.	479.
TOTAL INTENSIVE	RETENTION TIME	PEAK AREA	NORMALIZED AREA	RETENTION TIME	PEAK AREA	NORMALIZED AREA	
1	473.59	95.	0.479	1225.68	21	7.104	
2	574.58	64.	0.334	1236.94	27	2.716	
3	589.15	75.	0.397	1277.82	520.	2.716	
4	651.48	72.	0.375	1308.14	120.	0.627	
5	711.21	364.	1.901	1315.74	880.	4.597	
6	751.80	589.	3.071	1350.03	168.	0.878	
7	797.95	200.	1.045	1367.20	96.	0.501	
8	826.78	800.	4.597	1382.21	520.	2.716	
9	832.43	615.	3.218	1394.97	40.	0.209	
10	911.30	319.	1.665	1411.56	224.	1.170	
11	931.70	280.	1.503	1424.97	1296.	6.789	
12	964.04	420.	2.194	1466.03	1040.	5.432	
13	1002.22	392.	2.048	1480.23	360.	1.860	
14	1005.35	624.	3.259	1493.50	84.	0.334	
15	1032.80	476.	2.486	1510.88	484.	2.528	
16	1063.45	620.	3.233	1524.59	1840.	9.611	
17	1093.16	368.	1.922	1562.50	204.	1.066	
18	1121.91	700.	3.655	1578.50	96.	0.501	
19	1163.66	480.	2.507	1598.98	100.	0.542	
20	1208.25	144.	0.757	1625.57	1440.	7.522	

SUBJECT: WHITE MALE RB
 DATE: 3-27-73
 TOTAL INTENSITY: 10676.00
 PEAK INTENSITIES: 2000

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	34.929	316.0	3.355	17	1140.00	2450.0	2.282
2	90.000	497.4	5.074	18	1176.00	2730.0	2.767
3	114.93	498.5	5.072	19	1213.21	3208.0	3.851
4	140.94	1095.0	1.095	20	1240.28	1320.0	1.325
5	172.13	1097.4	1.097	21	1251.77	2170.0	2.201
6	195.41	1120.0	1.120	22	1345.98	1530.0	1.543
7	222.07	180.0	1.800	23	1369.82	4200.0	6.254
8	232.84	2305.0	2.305	24	1400.00	4130.0	4.163
9	242.48	2725.0	2.725	25	1437.34	1800.0	1.820
10	262.46	3765.0	3.765	26	1465.74	4300.0	4.418
11	270.03	2094.0	2.094	27	1503.02	800.0	0.816
12	281.11	2511.0	2.511	28	1542.10	2610.0	2.639
13	291.25	1913.5	1.914	29	1550.59	4110.0	4.130
14	301.44	2360.0	2.360	30	1566.85	1080.0	1.092
15	308.54	2240.0	2.240	31	1585.47	450.0	0.457
16	313.01	1100.0	1.100	32	1606.19	7100.0	7.185
				33	1625.57	1720.0	1.748

SUBJECT: WHITE MALE RB
 DATE: 3-27-73
 TOTAL INTENSITY: 10676.00
 PEAK INTENSITIES: 3574

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	475.90	2080.0	1.914	19	1664.54	1008.0	0.783
2	512.27	832.0	0.832	20	1288.50	500.0	0.435
3	740.02	4800.0	4.800	21	1305.63	3670.0	2.954
4	755.81	1504.0	1.504	22	1343.09	640.0	0.497
5	808.57	1248.0	0.970	23	1359.63	2200.0	1.710
6	935.76	5280.0	4.103	24	1376.46	330.0	0.261
7	855.47	2455.0	17.607	25	1412.00	5580.0	4.336
8	913.09	1410.0	9.491	26	1446.00	1320.0	1.025
9	950.12	2622.0	2.622	27	1474.80	2000.0	1.554
10	1030.95	5280.0	4.103	28	1485.27	600.0	0.466
11	1057.32	2664.0	4.525	29	1512.89	4440.0	3.451
12	1075.24	1175.0	1.691	30	1520.87	1870.0	1.455
13	1097.58	1728.0	1.343	31	1537.04	694.0	0.550
14	1121.91	2400.0	1.875	32	1551.04	1190.0	0.929
15	1132.55	4800.0	2.732	33	1559.97	3900.0	3.031
16	1156.34	1840.0	1.430	34	1573.54	742.0	0.581
17	1201.91	3450.0	2.685	35	1613.52	8160.0	6.345
18	1230.82	3040.0	2.353	36	1625.57	2608.0	2.182

SUBJECT WHITE BIRCH SA
 DATE 3-22-67
 TOTAL INTENSIVE 100000

17
 100 INTENSIVE 3799

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	4740	9700	1000	23	12810	12900	6.757
2	5000	3000	3000	24	13080	10700	6.100
3	5000	3000	3000	25	13210	18057	10.57
4	7000	14000	14000	26	13370	16200	9.925
5	8100	6000	6000	27	13500	7070	4.019
6	8400	2000	2000	28	13700	2080	1.187
7	8700	12000	12000	29	13900	7000	3.860
8	9100	4000	4000	30	14070	1980	1.088
9	9700	2000	2000	31	14310	3070	1.670
10	9800	10000	10000	32	14450	1900	1.000
11	9900	9000	9000	33	14570	2000	1.050
12	10000	4000	4000	34	14700	3000	1.580
13	10100	5000	5000	35	14700	3000	1.580
14	10100	5000	5000	36	14700	3000	1.580
15	10200	5000	5000	37	14700	3000	1.580
16	10400	5000	5000	38	14700	3000	1.580
17	10400	5000	5000	39	14700	3000	1.580
18	10500	5000	5000	40	14700	3000	1.580
19	10600	5000	5000	41	14700	3000	1.580
20	10700	5000	5000	42	14700	3000	1.580
21	10800	5000	5000	43	14700	3000	1.580
22	10900	5000	5000	44	14700	3000	1.580

SUBJECT WHITE BIRCH RD
 DATE 3-22-67
 TOTAL INTENSIVE 100000

17
 100 INTENSIVE 3799

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	5200	10000	10000	19	12000	5000	3.000
2	5700	10000	10000	20	12000	5000	3.000
3	6000	10000	10000	21	12000	5000	3.000
4	7000	10000	10000	22	12000	5000	3.000
5	8000	10000	10000	23	12000	5000	3.000
6	8500	10000	10000	24	12000	5000	3.000
7	8500	10000	10000	25	12000	5000	3.000
8	9000	10000	10000	26	12000	5000	3.000
9	9500	10000	10000	27	12000	5000	3.000
10	9500	10000	10000	28	12000	5000	3.000
11	9500	10000	10000	29	12000	5000	3.000
12	9500	10000	10000	30	12000	5000	3.000
13	9500	10000	10000	31	12000	5000	3.000
14	9500	10000	10000	32	12000	5000	3.000
15	9500	10000	10000	33	12000	5000	3.000
16	9500	10000	10000	34	12000	5000	3.000
17	9500	10000	10000	35	12000	5000	3.000
18	9500	10000	10000	36	12000	5000	3.000

SUBJECT WHITE MALE RR 21
 DATE 3-30-57
 TOTAL INTENSIVE 134900 PEAK INTENSIVE 3749

PEAK	RETENTIVE TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTIVE TIME	PEAK AREA	NORMALIZED AREA
1	50.15	308	4.378	19	1252.99	12000	9.378
2	67.90	1610	3.88	20	1275.96	384	6.285
3	74.53	348	3.18	21	1324.23	4000	3.557
4	79.57	348	3.69	22	1357.74	336	0.249
5	84.34	348	3.507	23	1374.17	2500	1.897
6	85.42	311	2.384	24	1397.17	240	0.176
7	86.71	188	1.371	25	1424.65	7200	5.394
8	91.94	1130	3.507	26	1460.32	2800	2.134
9	95.17	950	2.845	27	1475.38	2400	1.838
10	97.55	627	4.404	28	1493.35	704	0.522
11	1014.08	176	4.268	29	1502.82	336	0.249
12	1015.53	187	1.35	30	1525.41	1344	9.953
13	1017.59	439	1.724	31	1534.63	268	1.947
14	1111.01	448	2.11	32	1564.31	2576	1.909
15	1122.21	473	0.615	33	1576.01	3360	2.450
16	1175.35	448	3.41	34	1592.42	1584	1.174
17	1181.79	240	2.90	35	1625.57	14500	10.780
18	1211.51	575	2.943	36	1641.01	512	3.951

SUBJECT WHITE MALE KW 22
 DATE 3-30-57
 TOTAL INTENSIVE 87550 PEAK INTENSIVE 2403

PEAK	RETENTIVE TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTIVE TIME	PEAK AREA	NORMALIZED AREA
1	477.24	2490	3.509	17	1276.57	3740	4.673
2	585.51	530	3.06	18	1305.71	5250	6.505
3	747.41	334	6.914	19	1325.25	2350	2.914
4	800.11	292	3.31	20	1336.93	2170	2.695
5	836.33	450	3.317	21	1368.61	3640	4.507
6	851.40	374	5.464	22	1422.61	3500	4.339
7	854.67	483	3.357	23	1440.70	740	0.920
8	971.33	128	3.11	24	1458.77	1270	1.575
9	992.59	240	2.741	25	1465.34	1548	1.856
10	1015.89	2300	2.531	26	1505.50	896	0.111
11	1111.91	410	1.129	27	1525.94	1160	1.367
12	1145.03	480	5.944	28	1631.43	520	1.141
13	1175.49	3780	4.681	29	1661.82	3360	0.411
14	1205.34	3690	4.577	30	1574.07	348	0.428
15	1241.07	5040	6.24	31	1592.51	1470	0.180
16	1251.51	440	5.746	32	1625.97	360	1.233
				33	1642.88	2370	0.294

SUBJECT: WHITE FEMALE MO
 DATE: 5-2-67
 TOTAL INTENSIVE: 120594

PEAK	RETENTION TIME	AREA	AREA%	AREA	AREA%	AREA	AREA%
1	47.661	715	0.59	17.00	2.00	7.00	0.58
2	51.447	87	0.07	50.00	6.00	7.00	0.58
3	51.620	100	0.08	320	3.80	4.00	0.33
4	53.110	500	0.41	790	9.50	1.10	0.09
5	54.000	500	0.41	4000	48.00	5.00	0.41
6	56.040	300	0.25	2700	32.40	3.00	0.25
7	56.890	300	0.25	3800	45.60	4.00	0.33
8	57.000	200	0.16	1600	19.20	2.00	0.16
9	58.110	500	0.41	1600	19.20	2.00	0.16
10	61.170	1700	1.41	2300	27.60	3.00	0.25
11	61.593	400	0.33	100	1.20	0.10	0.08
12	62.535	200	0.16	7100	85.20	10.00	0.83
13	64.191	500	0.41	400	4.80	0.50	0.41
14	64.603	1000	0.83	1000	12.00	1.50	0.12
15	67.324	400	0.33	400	4.80	0.50	0.41
16	67.671	1000	0.83	2000	24.00	3.00	0.25

SUBJECT: WHITE FEMALE MO
 DATE: 5-2-67
 TOTAL INTENSIVE: 120594

PEAK	RETENTION TIME	AREA	AREA%	AREA	AREA%	AREA	AREA%
1	50.000	500	0.41	2000	24.00	3.00	0.25
2	67.300	400	0.33	100	1.20	0.10	0.08
3	70.000	1000	0.83	1000	12.00	1.50	0.12
4	70.000	500	0.41	500	6.00	0.70	0.58
5	84.000	500	0.41	500	6.00	0.70	0.58
6	90.000	1000	0.83	1000	12.00	1.50	0.12
7	87.000	1000	0.83	1000	12.00	1.50	0.12
8	90.000	1000	0.83	1000	12.00	1.50	0.12
9	92.000	2000	1.66	2000	24.00	3.00	0.25
10	95.000	1000	0.83	1000	12.00	1.50	0.12
11	95.000	300	0.25	300	3.60	0.40	0.33
12	100.000	500	0.41	500	6.00	0.70	0.58
13	100.000	500	0.41	500	6.00	0.70	0.58
14	105.000	700	0.58	700	8.40	1.00	0.83
15	105.000	700	0.58	700	8.40	1.00	0.83
16	105.000	700	0.58	700	8.40	1.00	0.83
17	120.000	1000	0.83	1000	12.00	1.50	0.12

SUBJECT MALAYAN
 DATE 4-5-67
 TOTAL INTENSIVE 75,000

45
 4119

PEAK	RETENTION TIME	AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	475.00	430	0.001	14	1175.85	2352	3.124
2	587.00	300	0.000	17	1209.00	3440	4.507
3	595.00	300	0.000	20	1239.31	2300	3.000
4	730.00	700	0.001	21	1255.52	1150	1.500
5	760.00	800	0.001	22	1315.71	2630	3.434
6	800.00	800	0.001	23	1348.63	500	0.653
7	840.00	800	0.001	24	1365.73	1970	2.574
8	850.00	800	0.001	25	1385.17	600	0.777
9	900.00	1000	0.001	26	1420.05	3120	4.144
10	920.00	1000	0.001	27	1455.25	1020	1.335
11	950.00	1000	0.001	28	1470.06	1180	1.578
12	980.00	1000	0.001	29	1523.81	5100	6.655
13	1000.00	1000	0.001	30	1561.02	500	0.653
14	1050.00	1000	0.001	31	1571.02	1530	2.000
15	1100.00	1000	0.001	32	1592.15	500	0.653
16	1150.00	1000	0.001	33	1625.57	6420	8.427
17	1170.00	1000	0.001	34	1644.91	1740	2.280

SUBJECT WHITE
 DATE 4-5-67
 TOTAL INTENSIVE 113250

46
 2004

PEAK	RETENTION TIME	AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	527.75	2320	2.091	20	1174.25	3450	3.052
2	600.00	430	0.381	21	1206.74	4400	3.890
3	620.00	1000	0.890	22	1235.37	4000	3.537
4	630.00	800	0.712	23	1262.54	400	0.353
5	730.00	800	0.712	24	1313.50	2010	1.795
6	760.00	700	0.618	25	1345.42	200	0.179
7	800.00	700	0.618	26	1365.61	700	0.618
8	840.00	1000	0.890	27	1385.11	6100	5.374
9	850.00	1200	1.062	28	1410.01	3120	2.748
10	900.00	1320	1.175	29	1454.45	1110	0.977
11	950.00	770	0.678	30	1480.00	1750	1.535
12	980.00	600	0.526	31	1499.09	400	0.353
13	1000.00	700	0.618	32	1523.89	5000	4.435
14	1050.00	1100	0.970	33	1552.75	1100	0.970
15	1070.00	1000	0.890	34	1571.05	500	0.443
16	1070.00	1000	0.890	35	1574.29	700	0.618
17	1070.00	1000	0.890	36	1570.49	1700	1.500
18	1100.00	1000	0.890	37	1585.73	500	0.443
19	1150.00	1000	0.890	38	1625.57	7110	6.251
20	1170.00	1000	0.890	39	1645.91	2000	1.795

SUBJECT: MITHRAMINE
 DATE: 3-15-67
 TOTAL INTENS: 85730

ION: 17
 PEAK AREA: 8176

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	473.96	33.88	0.41	20	1170.53	3360	2.82
2	508.17	100	1.21	21	1203.54	4324	3.65
3	651.61	151	1.81	22	1259.09	4410	3.73
4	700.27	108	1.29	23	1275.19	2010	1.71
5	772.08	1176	14.14	24	1306.91	6810	5.78
6	805.57	1440	17.15	25	1317.57	2660	2.26
7	849.19	494	5.88	26	1340.29	1344	1.13
8	850.82	1372	16.24	27	1361.01	4800	4.05
9	904.62	1490	17.62	28	1379.17	1900	1.59
10	971.93	3435	40.06	29	1415.56	8900	7.51
11	983.73	4083	47.74	30	1452.41	3674	3.08
12	1051.19	472	5.62	31	1470.05	2470	2.07
13	1078.59	1407	16.65	32	1479.00	3144	2.64
14	1117.57	2254	26.3	33	1498.07	1344	1.13
15	1202.59	6710	79.33	34	1523.50	12004	10.0
16	1251.54	3600	42.1	35	1561.55	2510	2.1
17	1349.57	3720	43.6	36	1574.13	2800	2.35
18	1421.91	7000	82.7	37	1589.66	960	0.8
19	1451.21	5512	64.7	38	1625.57	10440	8.78
				39	1643.53	1630	1.36

SUBJECT: MITHRAMINE
 DATE: 3-15-67
 TOTAL INTENS: 85730

ION: 17
 PEAK AREA: 8176

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	525.70	9815	114.7	18	1230.40	6600	5.61
2	675.30	1120	13.1	19	1241.98	4700	3.98
3	747.51	4824	56.4	20	1262.11	1824	1.54
4	811.53	1524	17.8	21	1312.03	900	0.75
5	844.33	4784	55.9	22	1318.98	974	0.81
6	894.57	1664	19.5	23	1339.45	274	0.23
7	919.37	4220	49.2	24	1356.98	390	0.32
8	958.06	1472	17.2	25	1376.75	324	0.27
9	1012.18	10124	119.4	26	1394.55	864	0.72
10	1052.08	2680	31.1	27	1475.60	1520	1.27
11	1056.62	1100	12.8	28	1477.95	264	0.22
12	1084.79	5064	59.1	29	1472.91	2300	1.93
13	1105.82	800	9.3	30	1519.22	720	0.6
14	1121.91	2520	29.3	31	1530.34	1070	0.9
15	1122.58	1624	18.9	32	1548.13	1260	1.06
16	1152.13	5184	60.6	33	1564.00	470	0.39
17	1211.75	1220	14.2	34	1625.57	880	0.73
				35	1641.59	510	0.42

SUBJECT WHITE FEMALE SR
 DATE 11-15-67
 TOTAL INTENSITY 2584

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PLAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	52.37	1834	0.711	19	1240.24	2740	2.845
2	60.88	1800	0.696	20	1260.20	760	0.795
3	65.31	2689	0.809	21	1272.54	480	0.341
4	67.17	370	0.143	22	1294.80	954	0.870
5	68.75	4344	0.452	23	1304.26	3754	3.925
6	70.82	1859	0.362	24	1358.58	780	0.793
7	72.77	1463	0.561	25	1379.08	2080	2.170
8	73.70	684	0.803	26	1394.73	314	0.320
9	73.70	5425	0.425	27	1428.45	5854	6.145
10	74.48	3416	0.916	28	1438.47	2104	6.205
11	75.13	3274	0.163	29	1480.87	3454	3.648
12	75.91	1820	0.603	30	1503.16	144	1.146
13	76.40	3360	0.514	31	1526.95	858	9.253
14	76.40	4580	0.773	32	1561.44	2116	2.213
15	78.43	3304	0.514	33	1574.70	3474	3.832
16	78.43	3644	0.444	34	1593.09	1380	1.443
17	78.43	11848	0.864	35	1614.03	120	0.120
18	78.43	7654	0.803	36	1614.57	9440	10.343
				37	1642.07	4214	4.400

SUBJECT WHITE FEMALE RC
 DATE 11-15-67
 TOTAL INTENSITY 11344

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PLAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	52.37	4095	0.111	17	1290.42	1080	0.820
2	54.21	864	0.66	18	1310.82	680	0.517
3	72.48	810	0.829	19	1330.92	208	0.154
4	72.48	4494	0.449	20	1363.74	4000	3.500
5	80.87	1112	1.112	21	1384.75	1404	1.000
6	80.87	2454	0.543	22	1417.44	4800	3.640
7	82.42	2115	0.348	23	1438.77	4404	4.005
8	70.40	5348	0.348	24	1474.45	684	0.520
9	74.13	1341	1.341	25	1475.74	1440	1.094
10	74.13	4174	0.417	26	1500.31	740	0.553
11	75.97	4297	0.429	27	1544.74	1644	4.424
12	76.47	3044	0.304	28	1574.14	1160	0.924
13	76.47	3743	0.374	29	1581.16	2080	1.580
14	76.47	1474	0.147	30	1594.94	700	6.501
15	76.47	1942	1.942	31	1624.57	4544	3.430
16	76.47	5474	0.547	32	1642.33	1170	0.893

SUBJECT WHITE MALE
 DATE 1-2-50
 TOTAL INTENSIVE 50750

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	874.22	3027	0.005	20	1149.25	3027	0.005
2	884.20	1484	0.003	21	1193.59	1600	0.003
3	874.17	994	0.002	22	1227.24	4320	0.008
4	713.52	2784	0.005	23	1258.94	4400	0.008
5	801.03	1135	0.002	24	1274.37	2170	0.004
6	874.20	1054	0.002	25	1328.51	6400	0.012
7	874.20	2372	0.004	26	1375.05	3200	0.006
8	991.26	4171	0.008	27	1396.92	500	0.001
9	821.02	1249	0.002	28	1429.16	10810	0.020
10	745.79	1334	0.002	29	1433.62	3312	0.006
11	954.32	1344	0.002	30	1480.28	2500	0.004
12	1003.16	4000	0.007	31	1493.99	1872	0.003
13	1015.22	2015	0.003	32	1505.23	4000	0.007
14	1025.69	3720	0.007	33	1527.50	11870	0.022
15	1027.56	1370	0.002	34	1536.75	2880	0.005
16	1061.25	4320	0.008	35	1563.77	3160	0.006
17	1105.44	4520	0.008	36	1575.04	4600	0.008
18	1121.91	3372	0.006	37	1591.15	1470	0.002
19	1145.62	4100	0.007	38	1625.57	15312	0.028
				39	1641.88	6032	0.011

SUBJECT WHITE FEMALE
 DATE 1-2-50
 TOTAL INTENSIVE 50750

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	474.22	1420	0.003	21	1226.73	304	0.000
2	555.20	752	0.001	22	1234.93	2112	0.004
3	722.04	2624	0.005	23	1279.75	360	0.000
4	805.03	700	0.001	24	1291.04	500	0.000
5	874.20	1824	0.003	25	1311.03	1560	0.002
6	894.55	672	0.001	26	1338.99	1710	0.003
7	891.66	868	0.001	27	1353.27	560	0.000
8	914.09	570	0.001	28	1380.56	280	0.000
9	935.35	704	0.001	29	1392.57	670	0.001
10	964.32	640	0.001	30	1413.22	352	0.000
11	975.29	480	0.000	31	1427.77	880	0.001
12	995.54	768	0.001	32	1441.35	2640	0.004
13	1003.18	2752	0.005	33	1457.59	2520	0.004
14	1064.03	140	0.000	34	1482.96	2880	0.005
15	1092.62	228	0.000	35	1493.60	792	0.001
16	1105.44	492	0.000	36	1514.27	3600	0.006
17	1141.91	704	0.001	37	1536.09	4752	0.008
18	1154.70	72	0.000	38	1557.31	3320	0.005
19	1186.64	136	0.000	39	1566.57	2800	0.004
20	1210.96	288	0.000	40	1596.02	3344	0.005
				41	1625.57	3332	0.005

SUBJECT: Neosulfonamide
 DATE: 9-2-66
 TOTAL INTENSIVITY: 10720

33

Hud

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	553.96	421	1.23632	21	1236.32	300	1.115
2	575.51	72	1.05578	7	1055.78	200	0.8723
3	701.71	54	1.27401	23	1274.01	384	1.3180
4	775.95	704	1304.92	24	1304.92	200	0.765
5	802.12	1160	4.194	25	1321.02	1440	5.2504
6	851.30	1898	3.298	26	1345.67	874	0.4905
7	863.95	744	2.533	27	1363.53	112	0.4005
8	904.95	312	1.127	28	1378.34	720	2.602
9	934.55	724	1.127	29	1402.98	404	1.677
10	992.59	1120	4.887	30	1422.17	1482	5.2350
11	1012.34	344	1.260	31	1452.24	300	1.3301
12	1023.49	575	2.982	32	1464.81	680	3.180
13	1035.80	1224	4.924	33	1474.21	530	1.8214
14	1061.87	1400	5.054	34	1507.94	1154	4.1123
15	1127.21	304	1.821	35	1523.52	1240	4.2510
16	1141.91	1056	3.31	36	1534.71	300	1.084
17	1147.57	1312	4.741	37	1570.22	192	0.694
18	1183.28	672	2.824	38	1605.69	244	1.062
19	1205.14	715	3.741	39	1609.63	170	0.614
20	1211.33	1954	5.814	40	1613.54	400	0.173
				41	1625.57	700	2.746

SUBJECT: Neosulfonamide
 DATE: 9-2-66
 TOTAL INTENSIVITY: 74176

34

Hud

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	426.24	108	0.178	10	1243.00	5376	5.718
2	503.00	144	0.742	11	1274.62	2800	3.058
3	675.51	272	2.287	22	1310.25	1200	1.359
4	725.73	475	3.829	23	1324.81	3328	3.534
5	845.74	4288	4.553	24	1340.22	200	0.237
6	905.93	2645	3.312	25	1370.23	800	0.844
7	946.44	1200	7.041	26	1374.01	704	0.748
8	942.44	640	0.683	27	1395.37	98	0.102
9	942.44	548	0.680	28	1410.25	760	0.807
10	1002.72	2615	4.970	29	1424.77	1924	4.043
11	1010.36	1376	1.941	30	1446.34	84	0.089
12	1035.01	925	0.981	31	1465.31	1020	1.083
13	1072.63	5072	7.633	32	1485.80	200	0.276
14	1104.92	5942	8.363	33	1510.47	1800	1.911
15	1121.81	11262	11.975	34	1525.70	2024	4.149
16	1145.53	5720	7.255	35	1540.43	432	0.454
17	1181.93	4484	4.889	36	1556.01	200	0.211
18	1213.01	3940	4.377	37	1582.11	252	0.268
19	1215.13	6228	6.922	38	1607.98	480	0.510
				39	1625.57	974	1.034

SUBJECT: 4500100000 BI
 DATE: 7/20/67
 TOTAL INTENSIVITY: 1705.

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	5.00	1876	1.071	24	16.00	2800	1.625
2	5.50	1876	1.071	25	17.00	2800	1.625
3	6.00	1876	1.071	26	18.00	2800	1.625
4	6.50	1876	1.071	27	19.00	2800	1.625
5	7.00	1876	1.071	28	20.00	2800	1.625
6	7.50	1876	1.071	29	21.00	2800	1.625
7	8.00	1876	1.071	30	22.00	2800	1.625
8	8.50	1876	1.071	31	23.00	2800	1.625
9	9.00	1876	1.071	32	24.00	2800	1.625
10	9.50	1876	1.071	33	25.00	2800	1.625
11	10.00	1876	1.071	34	26.00	2800	1.625
12	10.50	1876	1.071	35	27.00	2800	1.625
13	11.00	1876	1.071	36	28.00	2800	1.625
14	11.50	1876	1.071	37	29.00	2800	1.625
15	12.00	1876	1.071	38	30.00	2800	1.625
16	12.50	1876	1.071	39	31.00	2800	1.625
17	13.00	1876	1.071	40	32.00	2800	1.625
18	13.50	1876	1.071	41	33.00	2800	1.625
19	14.00	1876	1.071	42	34.00	2800	1.625
20	14.50	1876	1.071	43	35.00	2800	1.625
21	15.00	1876	1.071	44	36.00	2800	1.625
22	15.50	1876	1.071	45	37.00	2800	1.625
23	16.00	1876	1.071	46	38.00	2800	1.625

SUBJECT: 4500100000 BI
 DATE: 7/20/67
 TOTAL INTENSIVITY: 591.

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	5.00	1876	1.071	19	19.00	2800	1.625
2	5.50	1876	1.071	20	20.00	2800	1.625
3	6.00	1876	1.071	21	21.00	2800	1.625
4	6.50	1876	1.071	22	22.00	2800	1.625
5	7.00	1876	1.071	23	23.00	2800	1.625
6	7.50	1876	1.071	24	24.00	2800	1.625
7	8.00	1876	1.071	25	25.00	2800	1.625
8	8.50	1876	1.071	26	26.00	2800	1.625
9	9.00	1876	1.071	27	27.00	2800	1.625
10	9.50	1876	1.071	28	28.00	2800	1.625
11	10.00	1876	1.071	29	29.00	2800	1.625
12	10.50	1876	1.071	30	30.00	2800	1.625
13	11.00	1876	1.071	31	31.00	2800	1.625
14	11.50	1876	1.071	32	32.00	2800	1.625
15	12.00	1876	1.071	33	33.00	2800	1.625
16	12.50	1876	1.071	34	34.00	2800	1.625
17	13.00	1876	1.071	35	35.00	2800	1.625
18	13.50	1876	1.071	36	36.00	2800	1.625
				37	37.00	2800	1.625

47

SUBJECT WHITE MALE HN

DATE 7-7-50 1950. TOTAL INTENSITY 572.

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PKAR	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	527.85	65	0.445	19	1152.33	70	0.248
2	567.62	110	0.569	20	1169.37	200	1.035
3	625.85	72	0.373	21	1199.88	90	0.666
4	675.16	270	1.397	22	1219.97	120	0.621
5	686.65	312	1.616	23	1236.43	490	2.535
6	745.20	560	2.894	24	1268.50	104	0.538
7	800.91	1760	8.107	25	1279.78	64	0.331
8	841.50	240	1.159	26	1307.60	490	2.566
9	872.81	480	2.484	27	1334.02	120	0.621
10	895.04	706	3.643	28	1351.04	480	2.515
11	927.55	1920	9.935	29	1372.52	150	0.770
12	942.98	672	3.477	30	1387.33	42	0.217
13	984.75	416	2.153	31	1416.30	930	4.812
14	993.30	95	0.497	32	1470.42	1140	5.859
15	1008.91	848	4.385	33	1486.06	91	0.471
16	1179.19	480	2.484	34	1523.38	2700	13.971
17	1181.38	180	0.931	35	1555.73	180	0.931
18	1127.91	490	2.473	36	1595.52	144	0.745
				37	1625.57	125	0.647

48

SUBJECT WHITE MALE HN

DATE 2-16-50 1950. TOTAL INTENSITY 646.

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PKAR	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	569.47	168	0.722	19	1203.05	210	0.928
2	610.02	220	0.945	20	1222.97	504	2.106
3	637.44	420	1.805	21	1235.10	1440	6.188
4	720.70	280	1.203	22	1257.41	160	0.668
5	745.01	220	0.945	23	1276.39	320	1.375
6	845.17	800	3.436	24	1292.36	960	4.125
7	860.66	1730	7.460	25	1331.32	1100	4.727
8	894.69	688	2.957	26	1343.39	476	2.046
9	917.79	732	3.144	27	1365.82	80	0.344
10	936.55	302	1.295	28	1405.60	1350	5.601
11	977.95	240	1.031	29	1432.32	250	1.000
12	1001.05	384	1.650	30	1459.63	1560	6.704
13	1015.12	912	3.919	31	1473.37	280	1.203
14	1050.05	120	0.516	32	1507.60	1920	8.251
15	1072.85	672	2.888	33	1534.69	160	0.668
16	1097.93	300	1.289	34	1578.95	180	0.774
17	1121.91	640	2.750	35	1601.37	150	0.645
18	1157.03	352	1.513	36	1625.57	250	1.074

SUBJECT: 44111
 DATE: 10-10-66
 TOTAL INTENSITY: 123456

AD: 34
 660
 961

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PKAS	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	475.57	133	0.717	20	1260.77	100	0.274
2	574.24	135	0.772	21	1277.56	38	1.051
3	722.58	140	3.833	22	1309.22	128	0.450
4	853.56	171	1.029	23	1325.85	2560	7.009
5	851.91	150	3.944	24	1347.81	320	0.876
6	913.47	272	7.447	25	1364.91	432	1.183
7	921.55	384	1.051	26	1382.50	3760	10.295
8	974.35	244	0.785	27	1406.76	768	2.103
9	975.65	426	3.505	28	1425.27	5888	16.121
10	1033.19	544	1.449	29	1447.76	288	0.789
11	1032.75	720	1.971	30	1464.13	762	2.141
12	1039.26	204	1.427	31	1472.81	600	1.645
13	1042.93	201	0.569	32	1507.26	700	1.917
14	1121.91	1536	5.705	33	1525.55	1632	4.468
15	1147.74	150	0.611	34	1535.23	240	0.657
16	1176.51	52	1.511	35	1557.82	400	1.091
17	1204.41	288	0.789	36	1578.69	1020	2.793
18	1243.75	800	2.245	37	1601.20	680	1.862
19	1255.97	432	1.183	38	1625.57	1320	3.614

SUBJECT: 44111
 DATE: 12-20-66
 TOTAL INTENSITY: 36348

AD: 40
 976

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PKAS	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	545.70	1848	4.857	20	1233.99	920	2.418
2	554.01	774	20.353	21	1260.85	288	0.757
3	704.24	485	4.876	22	1271.76	140	0.368
4	853.00	704	1.850	23	1305.84	432	1.151
5	845.34	960	2.523	24	1317.60	288	0.757
6	845.52	1472	3.867	25	1326.00	64	1.758
7	850.81	1276	3.354	26	1345.17	140	0.368
8	944.57	1024	2.691	27	1369.44	96	0.258
9	971.42	800	2.103	28	1385.75	672	1.766
10	975.72	96	0.252	29	1407.07	640	1.703
11	1014.05	528	1.388	30	1426.93	1258	3.306
12	1033.43	320	0.841	31	1451.53	198	0.520
13	1070.05	816	2.145	32	1466.73	1560	4.111
14	1097.59	264	0.694	33	1507.63	1440	3.785
15	1121.91	352	0.925	34	1524.50	1296	3.406
16	1151.97	344	1.304	35	1531.81	1240	3.254
17	1178.25	360	0.946	36	1571.37	936	2.460
18	1203.02	108	0.284	37	1580.57	544	1.430
19	1222.15	680	1.787	38	1599.94	720	1.892
				39	1625.57	2448	7.738

SUBJECT MITE MALL AD 700 CAPLX BEFORE FLIT 41
 DATE 12-13-66 8541- MEAN INTENSITY= 2375

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	47.85	156	0.193	19	1155.24	100	0.117
2	60.00	278	0.325	20	1170.13	240	0.281
3	595.39	132	0.155	21	1205.76	70	0.082
4	717.15	1152	1.349	22	1273.95	526	0.618
5	775.86	304	0.355	23	1239.20	288	1.8124
6	791.81	382	0.417	24	1280.41	208	0.244
7	815.53	1455	1.705	25	1296.05	1560	1.826
8	882.20	175	0.209	26	1316.83	216	0.253
9	905.34	175	0.209	27	1343.42	1824	2.135
10	958.86	112	0.131	28	1377.84	840	0.983
11	973.32	288	0.337	29	1406.06	612	9.497
12	985.54	275	0.323	30	1423.15	5336	6.247
13	1002.81	400	0.468	31	1461.87	3108	3.639
14	1028.56	576	0.674	32	1505.46	13524	15.866
15	1055.25	448	0.524	33	1525.70	22400	26.225
16	1074.84	280	0.326	34	1567.92	8832	10.340
17	1097.15	168	0.197	35	1602.23	9856	11.534
18	1121.91	655	0.768	36	1625.57		

SUBJECT MITE MALE AD 1ST LAPX BEFORE FLIT 42
 DATE 12-12-66 64036 MEAN INTENSITY= 1817

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	408.19	44	0.064	20	1205.86	72	0.104
2	525.34	112	0.162	21	1227.45	800	1.246
3	602.02	72	0.104	22	1237.83	2248	3.662
4	657.92	76	0.110	23	1268.85	240	0.348
5	734.36	1106	1.599	24	1295.09	500	0.724
6	791.81	515	0.747	25	1309.79	804	1.165
7	815.53	1475	2.136	26	1320.03	1450	2.115
8	897.20	272	0.394	27	1345.28	400	0.574
9	912.51	555	0.861	28	1362.36	720	1.043
10	932.68	72	0.104	29	1379.56	5016	7.266
11	967.98	128	0.185	30	1406.32	3296	4.774
12	985.54	128	0.185	31	1427.64	1248	1.785
13	995.17	200	0.290	32	1463.19	11840	17.150
14	1007.34	276	0.400	33	1507.82	3600	5.226
15	1025.24	455	0.661	34	1528.65	10676	15.459
16	1058.79	312	0.452	35	1554.84	4596	7.231
17	1105.05	240	0.348	36	1558.27	9200	13.326
18	1121.91	1106	1.599	37	1601.90	2304	3.337
19	1178.19	288	0.417	38	1625.57	1968	2.834

SUBJECT WHITE MALE AD 151 011 2 10-1-66 42

DATE 12-19-66 1-11-7. PEAK INTENSITIES 478.

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	474.85	922.	3.534	20	1217.16	378.	2.026
2	500.69	168.	0.6503	21	1248.00	142.	1.057
3	717.15	605.	3.347	22	1266.29	143.	0.767
4	765.05	512.	2.816	23	1276.76	96.	0.528
5	804.01	440.	1.921	24	1294.15	288.	1.585
6	835.84	72.	0.395	25	1318.70	924.	5.086
7	890.23	112.	0.617	26	1338.88	144.	0.793
8	905.24	395.	2.185	27	1360.40	72.	0.396
9	945.17	168.	0.625	28	1376.79	646.	3.556
10	952.55	152.	0.637	29	1401.53	396.	2.180
11	965.12	256.	1.407	30	1416.38	1834.	8.994
12	1011.78	400.	2.202	31	1454.90	506.	2.765
13	1040.46	348.	1.915	32	1465.50	1154.	6.341
14	1065.67	88.	0.484	33	1502.32	1480.	8.147
15	1085.12	464.	2.554	34	1525.27	2436.	13.409
16	1121.87	108.	0.594	35	1578.31	876.	4.932
17	1155.70	175.	0.969	36	1587.55	300.	1.982
18	1182.64	50.	0.275	37	1599.01	274.	1.387
19	1202.35	580.	3.193	38	1625.57	646.	3.556

SUBJECT WHITE MALE MCD 44

DATE 10-19-66 84078. MEAN INTENSITIES 2249.

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	474.22	480.	0.567	19	1179.80	3460.	4.110
2	610.20	260.	0.307	20	1223.95	3072.	3.628
3	678.11	260.	0.307	21	1241.72	1260.	1.514
4	723.65	1160.	1.370	22	1272.62	1232.	1.455
5	768.52	160.	0.189	23	1282.65	288.	0.340
6	808.97	325.	0.385	24	1324.79	6016.	7.105
7	827.80	2412.	2.848	25	1356.47	364.	0.433
8	865.55	128.	0.151	26	1388.30	4864.	5.744
9	898.27	580.	0.587	27	1427.53	8400.	9.920
10	918.75	960.	1.134	28	1470.42	5440.	6.424
11	964.58	528.	0.624	29	1483.71	1440.	1.701
12	978.71	304.	0.359	30	1495.77	575.	0.680
13	992.41	2160.	2.551	31	1512.90	528.	0.624
14	1035.17	1824.	2.154	32	1528.48	9360.	11.054
15	1089.15	2968.	3.486	33	1548.25	1152.	1.360
16	1093.69	1920.	2.267	34	1570.52	1344.	1.587
17	1121.91	9600.	11.337	35	1591.20	960.	1.143
18	1144.10	544.	0.642	36	1604.69	1800.	0.213
				37	1625.57	7200.	8.503

SUBJECT INDIAN MALE RS 45
 DATE 11-11-58
 TOTAL INTENSITY 50000
 WAVELENGTH 13500
 RELATIVE INTENSITY

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PLAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	975.08	478	0.956	23	1177.08	1080	1.815
2	915.25	180	0.360	44	1266.74	480	0.480
3	625.01	60	0.120	25	1221.52	1100	1.855
4	641.69	440	0.770	26	1239.17	163	2.745
5	745.03	1320	2.640	27	1268.40	1080	1.825
6	765.32	240	0.480	28	1289.50	280	0.471
7	797.89	152	0.304	29	1300.85	280	0.471
8	833.82	732	1.464	30	1320.03	1232	2.070
9	865.68	440	0.880	31	1349.75	440	0.739
10	893.28	240	0.480	32	1367.61	260	0.444
11	917.69	672	1.344	33	1377.45	1360	2.292
12	935.36	300	0.600	34	1408.75	580	0.980
13	965.96	320	0.640	35	1421.10	2112	3.545
14	982.57	300	0.600	36	1445.65	610	0.639
15	995.02	420	0.840	37	1463.04	3090	5.205
16	1001.94	530	1.060	38	1476.43	1460	2.360
17	1005.52	550	1.100	39	1487.01	950	1.600
18	1034.71	640	1.280	40	1509.10	640	1.076
19	1061.88	560	1.120	41	1522.09	3200	5.445
20	1094.34	720	1.440	42	1542.11	13920	23.420
21	1114.91	800	1.600	43	1571.80	7420	12.472
22	1145.33	340	0.680	44	1625.57	6530	10.984

SUBJECT INDIAN MALE RS 46
 DATE 12-5-58
 TOTAL INTENSITY 67240
 WAVELENGTH 13500
 RELATIVE INTENSITY

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PLAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	977.23	104	0.155	27	1268.00	940	1.398
2	974.22	324	0.482	23	1227.13	1070	1.570
3	625.42	140	0.208	44	1240.38	3800	5.747
4	655.20	240	0.354	25	1269.74	1080	1.582
5	717.74	940	1.404	26	1280.90	1360	2.023
6	735.04	940	1.404	27	1310.74	1770	2.651
7	805.03	1820	2.713	28	1337.18	2120	3.105
8	815.62	4800	7.139	29	1351.70	672	0.999
9	857.48	1340	1.997	30	1391.27	200	0.291
10	883.62	2160	3.212	31	1400.55	160	0.238
11	905.87	4800	7.139	32	1414.34	720	1.071
12	927.81	1848	2.746	33	1439.91	1530	2.284
13	969.32	3760	5.592	34	1454.84	440	0.654
14	995.41	1250	1.868	35	1467.35	672	0.999
15	1003.18	1144	1.701	36	1479.58	392	0.583
16	1034.64	2500	3.714	37	1515.50	1260	1.874
17	1071.41	4410	6.566	38	1541.91	5860	8.745
18	1102.56	1404	2.088	39	1568.55	240	0.357
19	1111.91	3476	5.193	40	1580.20	580	0.853
20	1152.64	1360	2.023	41	1590.35	200	0.292
21	1165.03	1764	2.623	42	1625.57	648	0.954

SUBJECT INDIAN 4401

DATE 7-1-67

TOTAL INTENSITY 173.

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	467.88	66.	0.981	41	1121.61	7100.	5.879
2	476.02	96.	0.791	42	1122.61	70.	0.989
3	503.85	92.	0.735	43	1205.80	500.	0.707
4	515.44	68.	0.961	44	1226.98	130.	1.857
5	525.17	95.	0.791	45	1239.80	140.	1.979
6	725.83	369.	0.427	46	1265.44	76.	1.018
7	759.79	140.	1.979	47	1274.02	500.	0.791
8	803.34	132.	1.865	48	1308.95	70.	0.989
9	825.91	372.	1.242	49	1330.65	120.	1.696
10	825.53	235.	0.279	50	1347.85	104.	1.470
11	805.75	198.	4.795	51	1370.75	300.	0.424
12	916.25	394.	4.256	52	1381.43	90.	1.272
13	931.29	130.	6.221	53	1408.72	100.	1.413
14	975.75	175.	2.487	54	1428.31	170.	2.402
15	1152.31	682.	0.954	55	1458.47	24.	0.359
16	1162.01	168.	2.374	56	1465.89	144.	2.035
17	1162.34	68.	0.904	57	1480.39	180.	2.544
18	1315.09	575.	6.142	58	1507.82	140.	1.979
19	1363.11	325.	4.522	59	1520.24	540.	7.631
20	1162.35	84.	1.187	60	1578.89	130.	1.922
				61	1625.57	600.	8.679

SUBJECT WHITE 9396R.

DATE 7-1-67

TOTAL INTENSITY 2185.

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	449.78	80.	8.085	22	1273.50	7100.	7.668
2	480.40	84.	7.089	23	1310.48	1232.	1.311
3	522.58	144.	6.152	24	1325.46	4100.	4.427
4	593.54	52.	8.055	25	1341.15	1440.	1.532
5	728.35	1152.	1.226	26	1375.23	4800.	5.108
6	805.48	624.	0.864	27	1381.58	4032.	4.291
7	826.24	1342.	1.481	28	1409.54	2040.	2.171
8	891.25	1470.	2.043	29	1425.96	800.	0.851
9	1055.15	722.	0.765	30	1440.14	1140.	1.217
10	929.67	720.	0.765	31	1453.94	704.	0.749
11	951.59	1536.	1.635	32	1468.75	2880.	3.065
12	969.35	1800.	1.915	33	1485.61	11400.	12.132
13	1001.55	1008.	1.073	34	1510.53	1008.	1.073
14	1041.67	7040.	7.492	35	1523.05	1848.	1.957
15	1172.48	9802.	5.118	36	1541.55	616.	0.656
16	1121.91	2912.	3.099	37	1555.82	936.	0.996
17	1144.20	1280.	1.362	38	1574.04	3224.	3.431
18	1171.95	6360.	2.511	39	1586.04	3680.	3.878
19	1207.89	4604.	4.904	40	1598.74	280.	0.298
20	1230.14	1440.	1.532	41	1602.72	320.	0.341
21	1251.54	1480.	1.593	42	1611.40	404.	0.419
				43	1625.57	2720.	2.895

SUBJECT WHITE FEMALE DC

DATE	7-15-57	375000	9000
TOTAL INTENSIVE	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	135.55	212	0.054
2	414.37	170	0.042
3	527.75	435	0.112
4	675.27	420	0.111
5	730.02	1400	0.381
6	777.42	2720	0.719
7	827.13	1650	0.438
8	887.90	1460	0.387
9	881.22	4515	0.765
10	928.34	1550	0.760
11	945.33	2680	0.711
12	951.09	6970	0.787
13	965.31	6950	2.070
14	975.19	35800	7.475
15	1025.24	20600	7.102
16	1076.43	5370	0.548
17	1141.71	13520	3.247
18	1151.61	25500	6.754
19	1171.55	30600	8.137
20	1187.74	25220	0.754
21	1215.70	50700	10.755
22	1242.45	24	0.006
23	1267.49	73	0.019
24	1278.40	24	0.006
25	1296.53	25	0.007
26	1331.55	26	0.007
27	1350.95	27	0.007
28	1383.01	28	0.008
29	1392.26	29	0.008
30	1417.04	30	0.008
31	1433.53	31	0.008
32	1449.35	32	0.009
33	1484.23	33	0.009
34	1496.14	34	0.009
35	1524.62	35	0.009
36	1534.89	36	0.010
37	1550.10	37	0.010
38	1568.63	38	0.010
39	1586.54	39	0.011
40	1598.56	40	0.011
41	1635.57	41	0.011
42	1636.33	42	0.011

SUBJECT WHITE FEMALE MS

DATE	7-16-57	245000	66700
TOTAL INTENSIVE	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	524.96	300	0.171
2	600.00	6912	2.174
3	675.17	765	0.311
4	735.04	1792	0.724
5	805.03	4560	1.837
6	835.03	4712	1.503
7	891.98	3680	1.450
8	914.04	6980	1.398
9	922.83	7020	2.852
10	970.58	7650	3.110
11	1004.83	6950	3.343
12	1074.74	4720	1.724
13	1141.91	26200	8.740
14	1174.88	16138	6.530
15	1215.91	11770	4.768
16	1236.21	10410	4.217
17	1257.91	6150	2.484
18	1283.05	15740	7.554
19	1318.57	15	0.004
20	1335.63	20	0.005
21	1353.06	21	0.005
22	1367.30	22	0.005
23	1391.76	23	0.006
24	1417.13	24	0.006
25	1437.62	25	0.006
26	1446.11	26	0.006
27	1479.87	27	0.007
28	1491.24	28	0.007
29	1528.21	29	0.008
30	1545.60	30	0.008
31	1560.73	31	0.008
32	1577.35	32	0.009
33	1587.75	33	0.009
34	1597.61	34	0.009
35	1613.95	35	0.010
36	1625.57	36	0.010
37	1638.66	37	0.010

SUBJECT: 44-38861-1000 SN
 DATE: 7-2-67
 TOTAL INTENSITY: 1000000

PEAK	RETENTION TIME	AREA	NORMALIZED AREA	AREA	NORMALIZED AREA	RETENTION TIME	AREA	NORMALIZED AREA
1	44.00	17400	1.7400	17400	1.7400	1275.07	17400	1.7400
2	50.00	12750	1.2750	12750	1.2750	1310.93	12750	1.2750
3	55.00	4900	0.4900	4900	0.4900	1345.47	4900	0.4900
4	60.00	1800	0.1800	1800	0.1800	1376.03	1800	0.1800
5	65.00	1800	0.1800	1800	0.1800	1385.36	1800	0.1800
6	70.00	8350	0.8350	8350	0.8350	1412.04	8350	0.8350
7	75.00	17400	1.7400	17400	1.7400	1428.34	17400	1.7400
8	80.00	17400	1.7400	17400	1.7400	1440.80	17400	1.7400
9	85.00	17400	1.7400	17400	1.7400	1475.56	17400	1.7400
10	90.00	17400	1.7400	17400	1.7400	1483.23	17400	1.7400
11	95.00	17400	1.7400	17400	1.7400	1523.11	17400	1.7400
12	100.00	17400	1.7400	17400	1.7400	1541.21	17400	1.7400
13	105.00	17400	1.7400	17400	1.7400	1575.67	17400	1.7400
14	110.00	17400	1.7400	17400	1.7400	1606.04	17400	1.7400
15	115.00	17400	1.7400	17400	1.7400	1625.57	17400	1.7400
16	120.00	17400	1.7400	17400	1.7400	1647.74	17400	1.7400
17	125.00	17400	1.7400	17400	1.7400			
18	130.00	17400	1.7400	17400	1.7400			

NOT REPRODUCIBLE

SUBJECT	DATE	TIME	MADE	EN	EXPL.	4000	1000	2000
1	525072	16800	16800	16800	16800	16800	16800	16800
2	525073	16800	16800	16800	16800	16800	16800	16800
3	525074	16800	16800	16800	16800	16800	16800	16800
4	525075	16800	16800	16800	16800	16800	16800	16800
5	525076	16800	16800	16800	16800	16800	16800	16800
6	525077	16800	16800	16800	16800	16800	16800	16800
7	525078	16800	16800	16800	16800	16800	16800	16800
8	525079	16800	16800	16800	16800	16800	16800	16800
9	525080	16800	16800	16800	16800	16800	16800	16800
10	525081	16800	16800	16800	16800	16800	16800	16800
11	525082	16800	16800	16800	16800	16800	16800	16800
12	525083	16800	16800	16800	16800	16800	16800	16800
13	525084	16800	16800	16800	16800	16800	16800	16800
14	525085	16800	16800	16800	16800	16800	16800	16800
15	525086	16800	16800	16800	16800	16800	16800	16800
16	525087	16800	16800	16800	16800	16800	16800	16800
17	525088	16800	16800	16800	16800	16800	16800	16800
18	525089	16800	16800	16800	16800	16800	16800	16800
19	525090	16800	16800	16800	16800	16800	16800	16800
20	525091	16800	16800	16800	16800	16800	16800	16800
21	525092	16800	16800	16800	16800	16800	16800	16800

PEAK	RETENTION TIME	PLAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PLAK AREA	NORMALIZED AREA
1	1276.72	8064	0.4331	17	1631.10	4890	5.278
2	1314.89	1064	0.4835	18	1673.08	1024	1.100
3	1330.16	3756	2.556	19	1699.90	3600	4.148
4	1347.01	672	0.8268	20	1706.34	3400	4.218
5	1357.78	336	0.4204	21	1725.31	1100	1.210
6	1361.78	512	0.4402	22	1737.02	3312	3.577
7	1377.80	2724	2.181	23	1744.72	4200	4.623
8	1387.13	3700	2.812	24	1749.24	1900	2.074
9	1397.48	624	0.4490	25	1753.24	1176	1.270
10	1407.86	2400	1.734	26	1758.92	1344	1.452
11	1422.57	2400	1.800	27	1772.78	1344	1.452
12	1437.02	2400	1.884	28	1786.23	336	0.353
13	1448.14	7400	5.872	29	1802.80	2156	2.329
14	1454.92	1656	1.300	30	1814.98	8192	8.898
15	1463.70	1648	1.291	31	1824.21	224	0.242
16	1475.07	4592	3.605	32	1837.23	344	0.368
17	1487.81	1300	1.021	33	1852.87	352	0.380
18	1493.64	1756	1.384	34	1871.49	480	0.502
19	1517.26	2800	2.196	35	1884.84	1456	1.566
20	1521.85	2800	2.196	36	1888.27	1792	1.930
21	1527.24	1456	1.143	37	1897.27	1792	1.930

SUBJECT: A-111-70 56

DATE: 10-12-67 20-3900

TOTAL INTENSIVE: 10-12-67 20-3900

PEAK	RETENTION TIME	MEAN AREA	NORMALIZED AREA	AREA	RETENTION TIME	MEAN AREA	NORMALIZED AREA
1	11.70	11130	4.764	11130	11.70	4100	1.967
2	12.50	11880	4.864	11880	12.50	6272	2.995
3	13.00	11770	4.805	11770	13.00	7600	3.606
4	13.50	8192	3.324	8192	13.50	2450	1.130
5	14.00	8704	3.524	8704	14.00	5368	2.504
6	14.50	5850	2.405	5850	14.50	1604	0.755
7	15.00	6780	2.800	6780	15.00	896	0.428
8	15.50	1940	0.861	1940	15.50	4800	2.294
9	16.00	4880	1.980	4880	16.00	2688	1.264
10	16.50	4100	1.780	4100	16.50	480	0.229
11	17.00	5700	2.405	5700	17.00	2352	1.123
12	17.50	1677	0.677	1677	17.50	2440	1.165
13	18.00	5008	2.085	5008	18.00	800	0.382
14	18.50	1920	0.821	1920	18.50	2400	1.148
15	19.00	4100	1.780	4100	19.00	6392	3.053
16	19.50	5700	2.405	5700	19.50	1008	0.481
17	20.00	2736	1.171	2736	20.00	1624	0.776
18	20.50	2912	1.246	2912	20.50	8400	4.012
19	21.00	5824	2.495	5824	21.00	300	0.143
20	21.50	4570	1.958	4570	21.50	10600	5.158
21	22.00	4484	1.910	4484	22.00	8704	4.157
22	22.50	2376	1.017	2376	22.50	1820	0.869
23	23.00	3688	1.539	3688	23.00	7192	3.435
24	23.50	4100	1.780	4100			

SUBJECT: A-111-70 56

DATE: 10-12-67 20-3900

TOTAL INTENSIVE: 10-12-67 20-3900

PEAK	RETENTION TIME	MEAN AREA	NORMALIZED AREA	AREA	RETENTION TIME	MEAN AREA	NORMALIZED AREA
1	11.70	11130	4.764	11130	11.70	4100	1.967
2	12.50	11880	4.864	11880	12.50	6272	2.995
3	13.00	11770	4.805	11770	13.00	7600	3.606
4	13.50	8192	3.324	8192	13.50	2450	1.130
5	14.00	8704	3.524	8704	14.00	5368	2.504
6	14.50	5850	2.405	5850	14.50	1604	0.755
7	15.00	6780	2.800	6780	15.00	896	0.428
8	15.50	1940	0.861	1940	15.50	4800	2.294
9	16.00	4880	1.980	4880	16.00	2688	1.264
10	16.50	4100	1.780	4100	16.50	480	0.229
11	17.00	5700	2.405	5700	17.00	2352	1.123
12	17.50	1677	0.677	1677	17.50	2440	1.165
13	18.00	5008	2.085	5008	18.00	800	0.382
14	18.50	1920	0.821	1920	18.50	2400	1.148
15	19.00	4100	1.780	4100	19.00	6392	3.053
16	19.50	5700	2.405	5700	19.50	1008	0.481
17	20.00	2736	1.171	2736	20.00	1624	0.776
18	20.50	2912	1.246	2912	20.50	8400	4.012
19	21.00	5824	2.495	5824	21.00	300	0.143
20	21.50	4570	1.958	4570	21.50	10600	5.158
21	22.00	4484	1.910	4484	22.00	8704	4.157
22	22.50	2376	1.017	2376	22.50	1820	0.869
23	23.00	3688	1.539	3688	23.00	7192	3.435
24	23.50	4100	1.780	4100			

I-31

SUBJECT WHITE FEMALE PP

DATE 10-23-57 PEAK INTENSIVE 14973. TOTAL INTENSIVE 14973.0

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	51.420	1415.	0.074	19	1195.20	11200.	7.863
2	54.400	415.	0.092	20	1244.17	11424.	8.021
3	54.600	545.	0.291	21	1405.57	2088.	1.487
4	73.172	504.	0.254	22	1498.08	13440.	9.436
5	74.534	432.	0.203	23	1339.32	4048.	2.842
6	80.700	544.	0.282	24	1353.11	7176.	5.038
7	83.075	2304.	1.018	25	1369.70	2976.	2.089
8	87.140	1632.	1.114	26	1401.86	12096.	8.492
9	92.014	864.	0.407	27	1438.24	4536.	3.185
10	950.74	1728.	1.213	28	1465.64	5632.	3.954
11	975.27	1968.	1.362	29	1478.82	792.	0.556
12	945.54	4320.	3.033	30	1503.04	4624.	3.294
13	1025.47	1008.	0.708	31	1511.78	2632.	1.848
14	1095.12	480.	0.370	32	1535.73	840.	0.590
15	1062.22	3024.	2.123	33	1553.37	2256.	1.580
16	1044.14	1344.	0.917	34	1568.25	850.	0.597
17	1147.92	6512.	5.076	35	1584.44	240.	0.169
18	1167.45	960.	0.556	36	1602.26	2376.	1.668
				37	1619.13	1028.	0.722

SUBJECT WHITE FEMALE FS

DATE 10-23-57 PEAK INTENSIVE 14007. TOTAL INTENSIVE 14007.0

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	51.420	476.	0.374	20	1242.70	13312.	2.322
2	50.000	344.	0.260	21	1285.02	5760.	1.005
3	67.222	324.	0.257	22	1300.23	14720.	2.568
4	73.172	1920.	1.435	23	1313.43	6528.	1.139
5	80.700	990.	0.713	24	1335.13	3572.	0.623
6	83.160	1440.	1.051	25	1353.58	11440.	1.995
7	850.22	712.	0.514	26	1360.19	5888.	1.027
8	890.34	2472.	1.742	27	1385.87	9568.	1.669
9	925.83	4080.	2.874	28	1398.04	5824.	1.016
10	954.56	4752.	3.389	29	1411.48	2692.	0.469
11	991.36	2616.	1.855	30	1430.25	7616.	1.328
12	1006.00	2568.	1.826	31	1453.54	5544.	0.967
13	1045.87	4920.	3.470	32	1465.40	15776.	2.752
14	1050.59	1920.	1.365	33	1479.79	4032.	0.703
15	1084.02	2724.	1.925	34	1514.86	35228.	6.155
16	1144.14	2748.	1.923	35	1544.58	4320.	0.754
17	1157.39	2376.	1.683	36	1583.10	7680.	1.340
18	1194.40	4080.	2.874	37	1605.07	5104.	0.890
19	1214.70	1632.	1.184	38	1618.40	3472.	0.600

SUBJECT WHITE MALE JK

65

DATE 11-1-57

TOTAL INTENSIVE 10000

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	345.89	809	0.410	20	1274.15	8.630
2	423.79	256	0.448	21	1245.78	8.424
3	527.54	403	0.410	22	1498.69	12.480
4	633.00	1240	0.887	23	1348.48	2856
5	649.37	1440	0.983	24	1363.77	2112
6	720.79	1064	0.722	25	1398.69	2880
7	755.41	800	0.545	26	1405.15	3420
8	892.00	1680	1.147	27	1433.07	1360
9	1122.22	1916	0.689	28	1449.11	1512
10	1092.75	928	0.634	29	1461.59	1632
11	895.67	4895	2.343	30	1497.54	1476
12	913.55	4560	1.746	31	1508.65	4940
13	981.26	4800	3.277	32	1537.46	1320
14	983.56	13056	8.914	33	1546.67	1470
15	1051.27	15680	10.705	34	1564.54	360
16	1077.55	4800	3.277	35	1583.16	132
17	1103.61	3480	2.376	36	1599.50	1604
18	1141.85	10080	6.862	37	1614.65	2.602
19	1194.74	4792	4.665	38	1636.64	1.721

SUBJECT WHITE FEMALE CK

66

DATE 10-2-57

TOTAL INTENSIVE 186260

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	362.65	1328	0.713	20	1246.03	3.170
2	558.59	2352	1.264	21	1266.95	1.057
3	622.56	1032	0.558	22	1297.00	5.212
4	712.85	1512	0.812	23	1312.54	1.890
5	774.53	7424	3.986	24	1332.51	1.701
6	622.75	2176	1.168	25	1349.89	3.307
7	867.97	3504	1.965	26	1364.38	1.598
8	938.39	1056	0.567	27	1400.00	5.927
9	983.92	4192	2.251	28	1444.21	2.432
10	1013.10	1792	0.962	29	1435.21	4.104
11	1065.14	2592	1.392	30	1447.63	1.744
12	1083.96	3504	1.879	31	1461.93	3.608
13	1098.43	1820	0.977	32	1498.49	7.422
14	1124.63	6480	3.479	33	1506.80	4.467
15	1133.62	2368	1.271	34	1521.03	1.134
16	1170.72	4032	2.165	35	1546.84	11.339
17	1194.82	8736	4.690	36	1598.27	4.991
18	1214.91	1792	0.962	37	1610.56	0.691
19	1239.31	2800	1.503	38	1615.67	1.392
				39	1634.86	3.608

SUBJECT WHITE MALE FO
 DATE 10-18-67
 TOTAL INTENSIVE 5822

67

PEAK INTENSIVE 1991

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	30.65	974	1.000	22	1229.76	1700	3.012
2	45.21	174	1.190	23	1249.27	1154	1.972
3	508.29	74	1.273	24	1268.29	476	0.815
4	603.00	400	5.087	25	1290.66	1872	3.204
5	674.22	450	5.411	26	1330.59	140	0.240
6	732.85	1524	1.753	27	1358.51	1122	0.192
7	767.63	552	5.945	28	1349.43	612	1.048
8	811.39	1305	1.725	29	1365.94	468	0.801
9	855.89	552	5.945	30	1400.00	1450	2.492
10	901.12	482	5.411	31	1403.84	1872	3.204
11	955.12	720	1.231	32	1436.44	852	1.424
12	969.49	4548	4.601	33	1442.23	792	1.350
13	995.27	495	5.272	34	1463.93	2080	3.550
14	1012.89	1148	2.130	35	1475.96	150	0.257
15	1022.52	3100	5.977	36	1479.10	308	0.527
16	1064.78	2495	4.271	37	1506.09	1950	3.338
17	1091.45	2580	9.037	38	1508.31	1624	2.789
18	1120.82	3120	5.342	39	1536.08	1232	2.109
19	1145.94	3024	5.170	40	1549.44	1550	2.629
20	1167.13	1308	1.725	41	1590.50	2760	4.724
21	1182.62	2856	3.220	42	1617.72	1368	2.392

SUBJECT WHITE MALE BK
 DATE 10-11-67
 TOTAL INTENSIVE 34451

68

PEAK INTENSIVE 9568

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	394.69	4900	5.769	19	1220.97	10472	3.040
2	500.00	7144	0.612	20	1234.33	22176	6.438
3	523.71	482	0.251	21	1254.87	9850	2.861
4	734.95	1280	0.372	22	1260.50	5600	10.875
5	800.00	2080	0.604	23	1305.23	9784	2.899
6	867.51	895	0.260	24	1341.13	29440	6.547
7	893.64	1248	0.362	25	1358.47	9850	2.891
8	911.73	1520	0.557	26	1390.03	28800	6.261
9	984.25	2520	0.715	27	1428.44	12076	3.212
10	989.44	3360	1.497	28	1444.10	624	1.812
11	1012.89	12000	3.484	29	1454.92	52	1.510
12	1043.84	10416	3.024	30	1491.18	16	4.808
13	1070.88	5836	0.740	31	1527.77	32	1.015
14	1091.65	1745	5.168	32	1548.41	124	0.418
15	1135.15	2400	2.392	33	1571.99	2160	0.825
16	116.14	1176	3.270	34	1543.02	6368	1.813
17	1185.04	2112	6.131	35	1607.94	10400	3.19
18	1207.37	11076	3.252	36	1630.77	4096	1.189

69

SH

SUBJECT WHITE, FEMALE

DATE 7-20-67

1705

MEAN INTENSITY

195596

TOTAL INTENSITY

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	127.46	1792.	1.152	22	1257.92	4800.	3.085
2	525.80	3216.	1.067	23	1281.79	1732.	0.792
3	633.84	1200.	0.771	24	1291.06	960.	0.617
4	751.72	2086.	1.342	25	1308.50	4592.	2.951
5	747.32	1344.	0.864	26	1319.68	2976.	1.913
6	811.87	1232.	0.792	27	1344.82	800.	0.514
7	833.82	2424.	1.558	28	1360.04	1662.	1.068
8	853.74	960.	0.617	29	1378.75	640.	0.411
9	918.91	1792.	1.152	30	1411.15	3200.	2.057
10	951.48	784.	0.504	31	1419.90	2240.	1.440
11	985.01	2256.	1.450	32	1446.33	2580.	1.658
12	1004.53	1080.	0.692	33	1462.92	2632.	1.692
13	1037.57	640.	0.411	34	1473.13	2632.	1.692
14	1088.02	3960.	2.545	35	1491.29	364.	0.234
15	1120.99	4972.	3.213	36	1511.98	2924.	1.886
16	1158.03	784.	0.504	37	1520.85	3008.	1.933
17	1174.30	5760.	3.762	38	1550.37	1200.	0.771
18	1205.21	2720.	1.748	39	1559.97	504.	0.324
19	1238.41	3952.	2.540	40	1573.61	720.	0.463
20		6496.	4.175	41	1589.95	120.	0.077
21		6312.	4.152	42	1614.57	45244.	28.628

SUBJECT AIR SAMPLE MACHINE SHOP (PHYSICS) 70
 DATE 10-24-67
 TOTAL INTENSITY 32731.0 HEAD INTENSITY 9352.0

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	197.61	432.	0.132	18	1230.96	20008.	6.113
2	448.41	6720.	2.053	19	1258.05	920.	0.281
3	525.97	3850.	1.173	20	1289.40	34450.	10.537
4	728.40	1920.	0.587	21	1300.00	5600.	1.711
5	773.82	3520.	1.075	22	1318.06	5760.	1.760
6	810.95	980.	0.293	23	1337.39	8928.	2.728
7	831.34	1732.	0.527	24	1351.54	1920.	0.587
8	875.34	5080.	1.552	25	1355.08	21504.	6.570
9	927.50	6122.	1.873	26	1421.57	2496.	0.753
10	965.98	20195.	6.173	27	1448.76	8600.	2.648
11	1000.00	4800.	1.465	28	1480.78	8352.	2.552
12	1012.31	26780.	8.243	29	1500.80	3120.	0.953
13	1073.79	24304.	7.425	30	1527.18	3404.	1.040
14	1122.48	26880.	8.212	31	1541.52	3640.	1.113
15	1157.23	11500.	3.524	32	1557.65	600.	0.183
16	1183.38	39680.	12.123	33	1590.82	4896.	1.496
17	1201.55	6400.	1.955	34	1608.02	960.	0.293
				35	1612.00	528.	0.161

SUBJECT: LACROSTIN 1
 DATE: 5-28-67
 TOTAL INTENSITY: 127,496

PEAK	RETENTION TIME	PEAK AREA	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	5.8641	4000	20	9728	3.509
2	7.3498	1177	21	7476	6.900
3	8.3111	1000	22	16348	5.968
4	9.7726	1177	23	7508	2.756
5	10.7339	6874	24	1880	2.487
6	11.1952	1177	25	6708	2.531
7	12.6565	1177	26	8768	3.180
8	13.1178	1177	27	17472	1.781
9	13.5791	2493	28	21440	7.734
10	14.0404	6874	29	1476	1.434
11	14.5017	1177	30	3368	1.222
12	14.9630	1177	31	4840	3.550
13	15.4243	1177	32	7140	2.540
14	15.8856	6874	33	6440	2.373
15	16.3469	1177	34	1680	0.680
16	16.8082	705	35	1680	0.188
17	17.2695	1944	36	2128	0.722
18	17.7308	4840	37	400	0.346
19	18.1921	705	38	600	0.218
20	18.6534	8400	39	600	0.218

SUBJECT: LACROSTIN 2
 DATE: 5-28-67
 TOTAL INTENSITY: 127,496

PEAK	RETENTION TIME	PEAK AREA	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	5.8641	6874	40	41600	5.070
2	7.3498	1177	41	13216	3.104
3	8.3111	1177	42	8768	2.092
4	9.7726	2493	43	4000	2.254
5	10.7339	1177	44	4948	1.171
6	11.1952	6874	45	6768	1.569
7	11.6565	1177	46	7632	1.794
8	12.1178	2493	47	12672	2.475
9	12.5791	6874	48	17880	5.150
10	13.0404	1177	49	8328	1.851
11	13.5017	1177	50	14436	0.351
12	13.9630	1177	51	14960	0.761
13	14.4243	4144	52	2736	2.684
14	14.8856	2493	53	4128	0.077
15	15.3469	15354	54	5760	0.135
16	15.8082	4742	55	368	0.056
17	16.2695	3230	56	1380	0.455
18	16.7308	1654	57	1656	0.357
19	17.1921	3576	58	2760	0.605
20	17.6534	15232	59	1740	0.033

SUBJECT TROPICAL PLANT 2V2 EXPERIMENT 1967
 LAIR 12-15-64 74877
 TOTAL INTENSITIES 74877

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	418.01	4480	5.98	24	1318.01	4480	5.98
2	571.4	3714	4.95	25	1329.10	3714	4.95
3	1035.98	1036	1.38	26	1335.98	1036	1.38
4	1251.80	1252	1.65	27	1363.27	1252	1.65
5	1352.7	1353	1.79	28	1374.91	1353	1.79
6	1406.08	1407	1.86	29	1406.08	1407	1.86
7	1435.03	1435	1.89	30	1428.01	1435	1.89
8	1440.2	1440	1.90	31	1435.03	1440	1.90
9	1448.35	1448	1.92	32	1452.97	1448	1.92
10	1453.17	1453	1.93	33	1452.97	1453	1.93
11	1458.44	1458	1.94	34	1464.35	1458	1.94
12	1464.35	1464	1.95	35	1473.16	1464	1.95
13	1473.16	1473	1.96	36	1488.12	1473	1.96
14	1488.12	1488	1.97	37	1502.54	1488	1.97
15	1502.54	1503	1.98	38	1512.92	1503	1.98
16	1512.92	1513	1.99	39	1524.72	1513	1.99
17	1524.72	1525	2.00	40	1537.81	1525	2.00
18	1537.81	1538	2.01	41	1551.87	1538	2.01
19	1551.87	1552	2.02	42	1565.15	1552	2.02
20	1565.15	1565	2.03	43	1578.56	1565	2.03
21	1578.56	1579	2.04	44	1590.52	1579	2.04
22	1590.52	1591	2.05	45	1594.12	1591	2.05
23	1594.12	1595	2.06	46	1625.57	1595	2.06
24	1625.57	1626	2.07				

SUBJECT TROPICAL PLANT 2V2 EXPERIMENT 1967
 LAIR 12-15-64 74877
 TOTAL INTENSITIES 74877

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	418.01	4480	5.98	19	1318.01	4480	5.98
2	571.4	3714	4.95	20	1329.10	3714	4.95
3	1035.98	1036	1.38	21	1335.98	1036	1.38
4	1251.80	1252	1.65	22	1363.27	1252	1.65
5	1352.7	1353	1.79	23	1374.91	1353	1.79
6	1406.08	1407	1.86	24	1406.08	1407	1.86
7	1435.03	1435	1.89	25	1428.01	1435	1.89
8	1440.2	1440	1.90	26	1435.03	1440	1.90
9	1448.35	1448	1.92	27	1452.97	1448	1.92
10	1453.17	1453	1.93	28	1452.97	1453	1.93
11	1458.44	1458	1.94	29	1464.35	1458	1.94
12	1464.35	1464	1.95	30	1473.16	1464	1.95
13	1473.16	1473	1.96	31	1488.12	1473	1.96
14	1488.12	1488	1.97	32	1502.54	1488	1.97
15	1502.54	1503	1.98	33	1512.92	1503	1.98
16	1512.92	1513	1.99	34	1524.72	1513	1.99
17	1524.72	1525	2.00	35	1537.81	1525	2.00
18	1537.81	1538	2.01	36	1551.87	1538	2.01
19	1551.87	1552	2.02	37	1565.15	1552	2.02
20	1565.15	1565	2.03	38	1578.56	1565	2.03
21	1578.56	1579	2.04	39	1590.52	1579	2.04
22	1590.52	1591	2.05	40	1594.12	1591	2.05
23	1594.12	1595	2.06	41	1625.57	1595	2.06

NOT REPRODUCIBLE

K01

SUBJECT KITCHEN 003701

DATE 12-25-67

TOTAL INTENSITY 183000

PEAK	RETENTION TIME	PEAK AREA	UNNORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	UNNORMALIZED AREA
1	583.83	1584	5.810	30	1219.67	6280	5.751
2	598.10	330	3.007	31	1241.19	6144	3.250
3	615.45	118	0.892	32	1253.72	2810	1.535
4	718.14	117	0.861	33	1276.00	2560	1.376
5	751.80	849	8.456	34	1291.42	2640	1.434
6	817.73	426	3.447	35	1306.82	7200	3.426
7	855.78	2176	14.181	36	1353.11	2912	1.588
8	908.93	400	3.110	37	1370.14	448	0.424
9	924.59	172	0.868	38	1410.95	1124	0.614
10	948.63	135	0.874	39	1441.70	1480	0.785
11	977.84	272	1.611	40	1485.53	2720	1.443
12	1056.91	465	3.154	41	1510.14	676	0.631
13	1058.24	1815	12.254	42	1524.62	644	0.610
14	1092.06	2080	11.341	43	1538.10	342	0.670
15	1102.20	793	4.327	44	1548.15	1270	0.118
16	1121.91	2073	11.306	45	1566.24	210	0.029
17	1154.91	1138	5.172	46	1579.62	36	0.354
18	1184.23	1050	5.754	47	1599.34	158	1.288
19	1205.28	1150	5.281	48	1609.58	2380	

K02

SUBJECT KITCHEN 003702

DATE 12-21-67

TOTAL INTENSITY 114000

PEAK	RETENTION TIME	PEAK AREA	UNNORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	UNNORMALIZED AREA
1	629.40	328	3.077	19	1241.44	21120	4.978
2	555.20	284	0.867	20	1261.57	592	1.403
3	732.71	788	3.181	21	1281.05	768	1.810
4	815.62	792	0.187	22	1287.89	676	1.564
5	934.35	528	1.245	23	1314.17	13520	3.187
6	964.83	733	1.775	24	1336.54	330	0.792
7	985.55	454	1.071	25	1354.26	1800	0.424
8	1013.85	1420	3.470	26	1372.07	824	0.195
9	1025.60	1280	3.017	27	1383.59	100	0.024
10	1042.68	1496	3.981	28	1409.20	25600	6.034
11	1057.24	2376	5.491	29	1440.04	8208	1.935
12	1061.25	5736	13.577	30	1463.05	8320	1.461
13	1109.27	1582	3.741	31	1509.32	34176	8.056
14	1171.91	3540	8.444	32	1535.44	1124	0.284
15	1173.11	4242	9.957	33	1543.55	488	0.114
16	1185.70	2300	5.431	34	1559.44	420	0.099
17	1212.08	1036	2.444	35	1593.46	500	0.114
18	1234.83	1104	2.502	36	1604.82	870	0.205
				37	1625.57	192	0.045

SUBJECT

KITCHEN DOORS 3

DATE

4-27-51

ANALYST

W.P.

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	42.42	1000	0.01	22	119.09	17192	6.052
2	50.00	1888	0.02	23	122.19	17200	6.057
3	57.88	4228	0.09	24	126.57	10240	3.613
4	73.04	3710	0.11	25	129.74	5880	2.076
5	78.98	3936	0.12	26	130.12	24480	8.638
6	80.03	3740	0.11	27	131.97	5920	2.089
7	85.03	4672	0.13	28	134.42	6420	2.283
8	87.45	4420	0.12	29	135.88	9410	3.271
9	89.68	5470	0.15	30	136.12	15080	5.389
10	91.8	5700	0.16	31	142.87	5440	2.090
11	94.35	6120	0.17	32	144.83	4780	1.688
12	94.77	6700	0.19	33	145.93	8520	2.931
13	94.84	1600	0.05	34	150.26	170	0.004
14	101.28	2780	0.09	35	152.05	4820	1.719
15	103.85	2520	0.08	36	154.88	440	0.155
16	105.97	2520	0.08	37	157.97	500	0.178
17	108.03	1920	0.06	38	158.93	480	0.169
18	109.65	1720	0.05	39	167.26	2200	0.800
19	111.91	950	0.03	40	167.40	2920	1.056
20	115.81	360	0.01	41	171.17	1400	0.497
21	118.85	1300	0.04	42	182.57	4700	1.739

SUBJECT

KITCHEN DOORS 4

DATE

4-27-51

ANALYST

W.P.

PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA	PEAK	RETENTION TIME	PEAK AREA	NORMALIZED AREA
1	31.94	340	0.01	16	119.59	6340	2.270
2	52.41	500	0.01	17	118.70	1920	0.197
3	60.00	420	0.01	18	141.88	1310	0.221
4	65.13	180	0.005	19	143.40	750	0.419
5	73.42	3840	0.11	20	146.04	1440	0.632
6	81.42	4070	0.11	21	148.91	1920	0.779
7	83.42	3360	0.09	22	151.75	2560	0.839
8	90.93	4520	0.12	23	153.13	5030	1.812
9	92.15	26120	0.68	24	158.02	5980	1.945
10	97.00	720	0.02	25	160.00	2300	0.412
11	97.75	6420	0.16	26	162.72	2710	1.837
12	101.44	13120	0.31	27	165.78	2800	0.940
13	102.42	9800	0.22	28	168.77	5040	1.861
14	104.52	3370	0.08	29	168.91	1370	0.410
15	105.22	14070	0.34	30	172.80	3520	1.170
16	107.11	1740	0.04	31	174.80	7800	1.130
17	111.41	1110	0.02	32	178.91	1880	0.510

NOT REPRODUCIBLE

SUBJECT	AREA	PERCENTAGE	PERCENTAGE	PERCENTAGE	PERCENTAGE	PERCENTAGE	PERCENTAGE
TOTAL	TOTAL	TOTAL	TOTAL	TOTAL	TOTAL	TOTAL	TOTAL
1	10000	10000	10000	10000	10000	10000	10000
2	10000	10000	10000	10000	10000	10000	10000
3	10000	10000	10000	10000	10000	10000	10000
4	10000	10000	10000	10000	10000	10000	10000
5	10000	10000	10000	10000	10000	10000	10000
6	10000	10000	10000	10000	10000	10000	10000
7	10000	10000	10000	10000	10000	10000	10000
8	10000	10000	10000	10000	10000	10000	10000
9	10000	10000	10000	10000	10000	10000	10000
10	10000	10000	10000	10000	10000	10000	10000
11	10000	10000	10000	10000	10000	10000	10000
12	10000	10000	10000	10000	10000	10000	10000
13	10000	10000	10000	10000	10000	10000	10000
14	10000	10000	10000	10000	10000	10000	10000
15	10000	10000	10000	10000	10000	10000	10000
16	10000	10000	10000	10000	10000	10000	10000
17	10000	10000	10000	10000	10000	10000	10000

SUMMARY OF PATTERNS BY GROUPS

Consecutive Pattern No.	H U M A N S				
	Caucasian		Indian	Other	
	Males	Females	Males	Males	Females
1		X			
2		X			
3		X			
4			X		
5			X		
6			X		
7			X		
8			X		
9			X		
10			X		
11			X		
12			X		
13	X				
14	X				
15	X				
16	X				
17		X			
18	X				
19	X				
20	X				
21	X				
22	X				
23	X				
24		X			
25				X	
26	X				
27		X			
28		X			
29		X			
30		X			
31					X
32		X			
33				X	
34					X
35					X
36	X				
37	X				
38	X				
39	X				
40	X				
41	X				
42	X				
43	X				
44	X				
45			X		
46			X		
47			X		
48	X				
49		X			
50		X			
51		X			
52 NOT USED					
53 NOT USED					
54 NOT USED					
55		X			
56		X			
57		X			
58 NOT USED					
59	X				
60	X				
61		X			
62		X			
63		X			
64		X			
65	X				
66		X			
67		X			
68	X				
69		X			
Total	25	23	12	2	3
Environments	10				

APPENDIX II

Gas Chromatographic Positions of Peaks Obtained in Analysis of Vapors from the Same Caucasian Female

Analysis: Isothermal, 200-ft support-coated
open tubular column with Carbowax 20M.
Data are listed on Kovats Retention
Index Scale.

NOTES

- (1) For those ranges of the chromatograms which were outside of the calibrated range, the peak positions are given in minutes of retention time instead of Kovats Indexes.
- (2) Values below K.I. = 700 correspond to the very initial parts of the gas chromatograms and are much less exact (of order of ± 30 units at extreme early range) than the values in the higher K.I. ranges.
- (3) The last column represents difference in Kovats Indexes between peaks and shoulders observed in Run B and the closest matching peak in Run A. A plus (+) mark indicates that peaks in pattern B did not seem to have a reasonable counterpart in the pattern A within the limits of experimental error. A dash (-) means that a peak was observed in Run A but apparently had no satisfactory match in Run B.

Difference in
Kovats Indexes
Between Runs A & B
(See Note 3.)

Appendix Table Page 7

		Analysis Temperature, °C					
90°		120°		150° (A)		150° (B) Repeat	
Kovats Index	Min.	Kovats Index	Min.	Kovats Index	Min.	Kovats Index	Min.
	4.13				3.84		3.84
	4.26						4.04
435							(0.00 min.)
690				692			-
648							-
675				697			-
715						730	+ (33?)
758		752					
777							
799							
809		803					
837				823			
		842				844	4
				848			
853						867	14
		870					
872				881			
888							
		891					
		902					
903				909			
910						916	7
927		916					
		938					
945				942			
		954					
963						955	13
		967					
977				969			-
		981					
		995					
1005				999			-
1038		1005					
						1010	6
		1013					
				1016			
1017							
1024		1024					
1031				1031			-
1035							
1037							
		1038					
1047							
1057							
		1059					
				1066			
						1070	4
		1071					
1074							
1076							
		1077					
1082							
		1088					
1089							
1093							
1096							
1101							
		1108					
1109							
		1116					
1122							
1127				1130			

Difference In
Kovats Indexes
Between Runs A & B
(See Note 3.
Appendix Title Page)

Analysis Temperature, °C							
90°		120°		150° (A)		150° (B) Repeat	
Kovats Index	Min.	Kovats Index	Min.	Kovats Index	Min.	Kovats Index	Min.
1131						1131	1
1139		1133					
1143				1147			
1151		1147					
		1153					
1166						1160	13
1175		1170					
	11.95	1184		1188		1184	4
		1199				1207	
		1209					
		1215					
		1225		1224			2
		1236				1226	
		1247		1247		1246	1
		1259					
				1266		1262	4
		1274				1276	4
		1280					
		1290				1291	2
				1293			
		1300					
		1308		1311			
		1313				1316	4
				1320			
		1321					
		1326		1332			
		1335				1336	4
		1340					
		1348		1350		1350	0
		1352					
		1353					
		1360		1364		1365	1
		1368					
		1379		1374		1380	6
		1386					
				1388		1389	1
		1390					
		1393				1399	1
		1395					
		1398					
				1400			
		1405				1412	2
				1414			
		1415				1418	3
				1421			
		1432		1428		1428	0
				1439		1435	0
		1442					
				1452		1451	1
		1453					
						1457	1

Difference In
Kovats Indexes
Between Runs A & B
(See Note 3,
Appendix Title Page)

90°		120°		150° (A)		150° (B)		Repeat
Kovats Index	Min.	Kovats Index	Min.	Kovats Index	Min.	Kovats Index	Min.	
		1468		1468				
				1470		1476		-
				1476				1
		1479				1480		1
				1481		1487		1
				1488		1495		0
				1495				
				1500		1502		2
						1508		2
				1510		1516		1
				1517		1524		-
				1524		1532		0
1536								+
						1544		+
						1551		-
						1558		2
				1560		1568		+
						1569		1
		1572		1570				
						1574		+
						1577		0
						1584		+
						1589		1
				1590		1595		2
				1597		1608		3
		1611		1611		1618		2
				1620		1630		- (2)
						1635		+
						1635		1
				1634		1642		3
				1645		1652		2
				1654		1658		2
				1660		1678		3
				1681		1688		1
				1687		1699		-
				1690				2
				1701		1713		-
				1702				+
				1719		1721		2
						1733		3
				1736		1744		3
				1747				
				1754		1759		5
						1762		3
				1765				-
				1767				-
				1769		1780		+
						1781		1
				1782				-
				1783				-
				1784				-
				1786		1790		4
				1810				
						1811		1
						1813		+
						1830		-

UNCLASSIFIED

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13. ABSTRACT In the work on the detection and identification of human chemical signatures, experimental and computerized statistical techniques were first developed, tested, and standardized. The experimental techniques solved the problems of sample collection, recovery, injection into the gas chromatograph, calibration and characterization of signature components. The sample collections were based on the principle of equilibration and were performed by means of Apiezon L-coated Teflon powder in the form of a fluidized bed. The collected samples were transferred into a special injector needle by means of an IITRI-designed transfer system. The injection of samples into the gas chromatograph was performed in a timed, automatic and reproducible manner with a specially constructed injection system. The calibration method permitted correlation of the concentrations of known signature components in air with their respective peak areas. The sensitivity of this novel process is of the order of one part per billion (by volume) of an organic component in air.			

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14. KEY WORDS	LINK A		LINK B		LINK C	
	ROLL	WT	ROLL	WT	ROLL	WT
Chemical Signatures - Detection Chemical Signatures - Identification Gas Chromatography						

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