FSTC-HT-23-658-68 U.S. ARMY FOREIGN SCIENCE AND TECHNOLOGY CENTER 00 EXPERIMENTAL VARIATION OF NEWCASTLE DISEASE VIRUS COUNTRY: USSR may 2 3 1969 B **TECHNICAL TRANSLATION** Distribution of this document is unlimited. It may be released to the Clearinghouse, Department of Commerce, for sale to the general public. 1 Reproduces by the CLEARINGHOUSE for Frideran Scientiald & Technica Information Springfield Val. 20181

7

TECHNICAL TRANSLATTON

FSTC_HT_23-658-68

EXPERIMENTAL VARIATION OF NEWCASTLE DISEASE VIRUS

bу

V. M. Kolosov, and V. N. Fyurin

SOURCE: VOPROSY VIRUSOLOGII No. 4, pp. 443-451, 1964 USSR

.....

Translated for FSTC by Techtran Corporation

This translation is a rendition of the original foreign text without any analytical or editorial comment. Statements or theories advocated or implied are those of the source and do not necessarily reflect the position or opinion of the US Army Foreign Science and Technology Center. This translation is published with a minimum of copy editing and graphics preparation in order to expedite the dissemination of information. Requests for additional copies of this document should be addressed to the Defense Documentation Jenter, Cameron Station, Alexandria, Virginia, ATTN: OSR-2

EXPERIMENTAL VARIATION OF NEWCASTLE DISEASE VIRUS

In the domestic and foreign literature are a series of reports on modification of viruses during their adaptation to the central nervous system of susceptible or unsusceptible animals.

In 1947 Newcastle disease virus was adapted to a sirenian hampster brain [17], and as a result of this, its virulence for birds decreased and the "brain" virus was recommended as a live vaccine. However, during culturing of this virus in chick embryos, it comparatively rapidly re-established its virulence for birds.

Attempts of several investigators to attenuate this virus by means of adaptation to the organism of white mice and guinea pigs [9, 12, 21], did not give the expected results. In 1949 to 1956, V. N. Fyurin studied modification of Newcastle disease virus (virulent strain T) in the process of its long adaptation to guinea pigs and cther mammals. As a result, a neurotropic variant was obtained, which regularly causes the destruction of guinea pigs upon intra-cerebral infection. However, this straim in the form of a guinea pig brain suspension, was avirulent and completely non-immunogenic for birds. Thus, the "brain" virus proved to be unsuitable as immunizing material. During a series of passages in chick embryos, this virus showed marked immunogenic properties, and some virulence for chicks, which usually did not exceed the residual virulence of the widely known vaccine strain H. However, the genetic similarity of the adapted Newcastle disease virus (strain GNKI) until recent times remained unclear, which to a certain extent prevented its wide use as a live vaccine.

The aim of our search was to fill this gap in the study of the properties of the adapted GNKI strain, and, in particular, to study its genetic similarity by the method of experimental selection.

Haterials and Methods

In the experiments, the GNKI strain of Newcastle's disease virus was used by way of guinea pig brains of the 302nd intra cerebral passage. After three subsequent passages in guinea pig brains, the virus was transferred to 10 to 11 day-old chick embryos, and serially passaged 20 times. The hemagglutination reaction was carried out by the generally accepted method in a volume of 1 ml with a 1% suspension of chick erythrocytes. The hemadsorbtion reaction was carried out by the method of Vogel and Shelokov [22] in infected tissue cultures of chick embryo fibroblasts. Negative colonies were obtained by the method of Hsiung and Melnick [11] in growing culture of chick embryo fibroblasts. Adsorbtion of the neurotropic variants was carried out by the method of Piraino and Hanson [15]. The method of threshold infecting doses of virus was used for separation of virus clones. The passaging of virus in embryos was carried out in two variations: embryos of serial passage were infected in the allantoic cavity by concentrated viruscontaining allantoic culture $(10^{-1} \times 0.1 \text{ ml})$ -variant A; in the other case, for successive infection of embryos, virus was used in dilutions above the LD_{50} (variant B). The enzymatic activity of the virus was

established on the basis of its ability to be adsorbed and to elute from formalinized chick embryos, and also in the hemolysis reaction of the latter according to the method of Khou Yun'-de [5].

Results

The summarized data of the experiment in the study of the genetic homogeneity of the GNKI strain are presented in Table 1, from which it is clear that the "brain" virus in the first passage in chick embryos cause destruction of them after 144 hours, in further passages this period (for variant A) was shortened to 48 to 56 hours. The virulence of the virus increased from 2.75 to 8.26 and 8.74 log ELD_{50} (for var-

iants A and B). Variant B, which was obtained using the method of maximum dilution, possessed the same infectious titre for the embryos as variant A; however, in the progress of the serial passages, the period of destruction of the latter was not shortened, but stabilized in the range 96 to 144 hours. In comparative experiments in chicks, a clear difference in pathogenicity of the indicated variants was observed (Table 2).

-2-

| <u> </u> | 8 | Variant A | | | | | Treatment by the Pir- ainc and Hanson | | | Ê Variant B | | | |
|---|---|-----------|--|----------------------|---------------|--------------------------------------|--|---|--|--|-----------------------|---------------|--|
| Passage | Period of De- struction of Embryos (in h | LOG ELDSO | Hemagglutin- stion Titre | Cytopathic Effect | Hemadsorption | Meth G G 1 J J J J | Period of po | Period of De- struction of Embryos (in h | LOS ELDSO | Hemagguth. ation Titre | Cytopathic. Effect | Hemadsorption | |
| 1st 2nd 3rd 4th 5th 10th 12th 13th 13th 13th 15th | 144 96 72 56 72 60 48 48 56 56 56 | 2.75 | 0 0 128 1 024 512 1 024 1 024 512 512 512 | +++++++000 | | 5,23 6,50 5,23 6,66 6,00 | <u> </u> | 144 120 120 144 120 120 56 120 120 144 120 120 | 2,75 2,6 4,5 6,48 6,5 7,28 7,00 7,5 7,76 7,76 7,60 8,74 | 0 0 0 128 256 128 256 128 256 512 | 000++++++++++ | ***** | |

Table 1 Biological Properties of 2 Variants of Newcastle Disease Virus, Isolated from GNKI Strain

Minur after

Note. Hemagglutination titres are presented in the form of inverted fractions.

As a result of the research carried out, it was established that the "brain" virus did not possess cytopathogenicity and did not form negative colonies in the tissue culture. During serial passage in chick embryos, the virus increased its pathogenicity for the embryos, tissue culture, and also acquired a hemagglutinating property. Virus of the fourth passage in chick embryos caused cytopathic changes and the formation of patches. Negative colonies were not distinguished by form and quantity. Virus isolated from certain colonies was analogous to the control. By applying the method of Piraino and Hanson, we succeeded in experimentally obtaining the aneurotropic variant.

Using the method of serial dilutions during passage of the "brain" virus in chick embryos, we selected the avirulent variant B of the GNKI vaccine strain. Probably, the long adaptation of the virulent, epizootic T strain to a guinea pig brain is accompanied by the appearance of heterogeneous elements (virions) in the virus population, which can be revealed by two methods of culturing of the prototypical virus in the developing chick embryos.

-3-

| ~ | | 1 | _ | | 1 1 | Growth of Chicks (in Days) | | | |
|------|--------|--------------|--------------|----------------------|--|----------------------------|---------------|-------------|--|
| Vari | ant | Passage | (-log LD50 | Method of | Dose (in ELD 50 |) 2 | 15 | 22 | |
| | A B | 20th 20th | 8,26 8,74 | Intramuscularly > | 10 ⁵ ×02 10 ⁵ ×02 | 4/4 0/25 | 14/17 0/35 | 3/4 0/23 | |

Table 2Comparative Virulence for Chicks of Two Variants(A and B) of the GNKI Vaccine Strain of Newcastle Disease Virus

Note.

· 14. E 1

्. च उ∏्≴ Denominator--number of chicks taken in experiment; Numerator--number of stricken chicks.





-4-

Enzymatic Activity of the Virus as a Test of the Selection of Avirulent Clones

Clear distinctions in ability to be adsorbed by formalinized chick erythrocytes (Figure 1) were detected in three strains of Newcastle disease virus (A, B, and T strain), which differed in virulence. A difference was also detected in the eluting ability of the indicated variance of the virus. Elution occurs more rapidly and more completely in the virulent T strain. With a reduction in virulence, there is also a decrease in the speed of elution of the virus (Table 3).

Evidently, both adsorption and elution of virus from erythrocytes is in some measure connected with the enzymatic activity of the virus, since loss by the latter of virulent properties is accompanied by diminished enzymatic activity. Having observed this, we use the adsorption method for the experimental selection of variants differing in virulence from the original pathogenic T strain.

| · · · · · · · · · · · · · · · · · · · | Ti | me o | f Elu | tion | (in Hinutes) | | | | |
|---------------------------------------|----------------|---------------|------------------|------------------|------------------|------------------|--------|----|--|
| Viruo | 3 | 0 |] | | | 20 | 180 | | |
| VITUS | GAU | * | GAU | % | GAU | . % | GAU | * | |
| GNKI-B GNKI-Ä T Strain | 0 64 128 | 0 25 50 | 16 255 256 | 12 100 100 | 32 256 256 | 26 100 100 | 64 | 50 | |

Table 3 Dynamics of Elution of Variants of Newcastle Disease Virus

Note: GAU = the number of hemagglutinating units in 1 ml.

With this aim, the virus was precipitated five times by formalinized erythrocytes in order to fully deplete possible virulent virions in the population. After the fifth successive adsorption, the virus found in the supernatant liquid, was titrated and selection of avirulent clones was carried out by the method of serial dilutions in chick embryos. During titration of the non-adsorbed virus particles in the chick embryos, a significant difference was established in the period of their death (Table 4). Embryos infected by the original virus, were destroyed after 48 hours, whereas the virus after adsorption caused their destruction in 96 to 120 hours. After 120 hours, all the living embryos were killed by cold, and each embryo was separately studied for the presence of virus with the help of a drop of RGA on the glass. As a result, two surviving embryos (No. 4 and 10) were detected, which were infected by a virus at dilution 10^{-4} , the allantoic fluid of which contained hemagglutinin. The organs of these embryos did not have visible pathological changes. The allantoic virus culture (TA₁) of embryo No. 10 was passaged five times in chick embryos, and towards the fifth passage, the variant reestablished vir-

ulence (up to the original extent) and enzymatic activity (Table 5 and 6).

| | <u> .</u> | Origina | | | | | | | | | | |
|----------|--------------|---------|-----|------------|------|-----|-------------------|--------|---|------|----------|-----------|
| Dilution | 1 | 2 | 3 - | 4 | 5 | 6 | 7 | 8 | 9 | 10 | Vir | <u>us</u> |
| of Virus | Pe | riod | | Dest | ruct | ion | | mbryg | $\frac{1}{1}$ | hrs' | 1 | 2 |
| | | 1 100 | | 1 | | | <u>, <u> </u></u> | ind ye | <u>, , , , , , , , , , , , , , , , , , , </u> | 1 | <u> </u> | · · · |
| 10-* | 96 | 96 | 96 | 92 | 108 | 108 | 108 | 120 | 120 | + | 48 | 48 |
| 10-3 | · 96 | 108 | 120 | 120 | 120 | - | - | - | _ | - | 48 | 48 |
| 10-4 | [] <u> </u> | | _ | + | | — | | - | - | + • | 48 | 48 |
| 10-4 | [| | — . | ! ' | | · | — I | - | | - | 48 | 48 |

| Tab | le 4 | 4 |
|-----|------|---|
|-----|------|---|

Titration of Non-adsorbed Virus in 10-Day Old Chick Embryos

Note: - equals negative reaction of hemagglutination; + equals positive.

| | | | Table | 5 | | |
|---------|----|-----|-------|----|-------|---------|
| Passage | of | TA, | Virus | In | Chick | Embryos |

| Passage | RGA Titre | Log ELD 50 | Period of Destruction of Embryos (in Hours) | Pathomor- phological Changes | Log LD ₅₀ | Ratio of ELD ₅₀ to LD ₅₀ |
|---------------------------------|---------------------------------|------------------|--|------------------------------------|-------------------------|---|
| lst 2nd 3rd 4th 5th | 256 128 512 256 256 | 7,5 8,0 | 72120 56 4872 4872 4872 | 0 +++ +++ ++++ | 5,4 | 2,6 |

Note: +, +++, ++++ are different degrees of pathomorphological change.

-6-

)

| j. | Adso | orption | | | I Elu | Elution | | | | |
|-------------------|-----------------|----------------|--------------|--------------|---------------|-----------------|-------------------|-------------------|--|--|
| Passage | ***** | Reacti | on Tim | e (in M | inutes) | | | | | |
| | 15 | 30 | 45 | 60 | 30 | 60 | 120 | 150 | | |
| lst 2nd 5th | 128 128 8 | 128 64 0 | 64 4 0 | 16 0 0 | 4 16 64 | 32 64 128 | 128 256 256 | 128 256 256 | | |

Table 6 Changes in the Enzymatic Activity of the Virus (Variant TA₁) During Passage in Chick Embryos

Note. 256 GAU/ml were taken in the experiment; the numbers represent the number of GAU in the supernatant fluid.

We assumed that the reversion of variant TA₁, apparently, occurred because of incomplete precipitation of the virulent virions, and therefore the experiment was repeated with subsequent adsorption of the TA virus on the first passage by the same method. The results of the repeated experiment are presented in Table 7.

| Table 7 | |
|--------------------------------------|-------|
| Results of Titration of Non-adsorbed | Virus |
| in 10-Day Old Chick Embryos | |

| DII | ution | | Number of Embryos | | | | | | | | | | | |
|-----|--------------------------------------|----------|-------------------|----------|----------|----------|----------|----------|---------|---------|---------|--|--|--|
| of | Virus | 1 | 2 | a | 4 | 8 | • | 7 | • | • | | | | |
| | 10 ⁻¹ 10 ⁻² | 72 56 | 72 58 | 63 63 | 72 56 | 82 82 | 56 50 | 88 82 | 96 + | 98 + | 87 + | | | |
| | 10-4 | + | + | | - | - | - | - | = | = | - | | | |

Note: Symbols are the same as in Table 4.

-7-

Upon dissection of the destroyed embryos, it was noted that in the majority of them hemorrhage in the body and oxypital hematoma were absent. Allantoic culture of the wirus was taken from embryo No. 1 (infected with virus at a dilution of 10^{-3}), and was passaged three times in embryos by the serial dilutic method. As a result, a variant of the virus (TA₂) was obtained, differing from the original (T) in the period of destruction of the embryos (Figure 3). Not one of the

embryos destroyed by the TA₂ virus variant, has blood flow and oxypital hematoma; moreover, this was characteristic for the original virulent T strain. Sharp differences were detected between the TA₂ variant and the original T strain in pathogenicity for 1- and 10-day old chicks

upon intracerebral and intramuscular methods of infectior (Table S).





The comparative antigenic activity of the TA, variant and the

original T strain were studied with the help of RZGA with rabbit antiserum of the T strain. The serum inhibited hemagglutination of the TA₂ virus (lst, 3rd, 5th and 8th passages) and the T strain at dilutions 1:256--1:512.

-8-

| Table 8 | | | | | | | | | |
|-----------------------------|---|--|--|--|--|--|--|--|--|
| Comparative Characteristics | of Yirulence of TA ₂ Variant and | | | | | | | | |
| T Strain Upon Titration | in Chicks and Chick Embryos | | | | | | | | |

| | La | Leg LD | c at inducts | |
|--------------------------|---------------------------|--------------------|----------------------|--------------------------|
| | 2-2-36 | hadronate | autor j | hadenesseeherst |
| Virus | ĺ | Day eld Calicia | 10-Day and Chicks | Day Old Chicks |
| ΓΑ, ΓΑ, Γ origina: | \$.0 9,0 8.5 3,0 | 1.6 0 7,0 | 1.0 0 0 7.5 | 5.5 6.0 4.5 7.5 |

Comparative Hepolytic Properties of the Virus Variants

By studying the hemolytic properties of Newcastle disease virus, Kilham [13] established the effect of the pH of the medium, temperature and form of erythrocytes on its activity. The author showed that the hemolytic properties of the virus may be inactivated by specific antisera and formalin. It was later established [7, 8, 12, 14, 19] that the majority of strains of Newcastle disease virus possess hemolytic properties, which increase after dialysis of the virus, its purification by the method of freezing and thawing, replacement of allantoic fluid with physiological solution, buffer, sodium citrate, etc. Hemolytic activity sharply decreased at a temperature of 4°. In our research, a comparative study of the hemolytic properties was carried out with virants A, B, and TA₂ of the Newcastle disease virus

| | | | Table 9 | | | |
|-----------|-----------|-----|------------|------------|------------|----|
| Comparati | ive Chara | cte | ristics of | Henolytic | Propert es | o£ |
| | Variants | of | Newcastle | Disease V | irus | |
| | | | . IV | irus Strai | n | |

| Erythroo | yt: | e\$ | • | | | т | * | В | |
|--------------------------------|-----|-----|---|---|---|-----------------------|----------------------|------------------------|--|
| Horse Guinea Pig Chicken | • | • | • | • | • | 0,18 0,438 0,07 | 0,11 0,19 0,03 | 0.91 0.048 0.011 | |

Note. The numbers represent optical density, of a range of error of determinations of ± 0.03 .

-9-

From Table 9 it is clear that the virulent T strain possesses a higher hemolytic activity, the avirulent B variant possesses almost none, and the A variant occupies an intermediate position between the T strain and the B variant. Evidently, the changes in the virulent properties of the T virus in the process of adaptation to guinea pigbrains, are accompanied by a reduction of hemolytic activity.

Interest has been shown in studying the inner connection of hemagglutinizing and hemolytic activity in variants of Newcastle disease virus by the method proposed by McCollum and Bradly and based on the use of chicken erythrocytes [14]. The interconnection between hemolytic and hemagglutinizing properties of the virus have not been established; evidently, this property is determined by various structures and biochemical components of the virus capsid. In the T strain clearly expressed hemolytic activity was observed up to a dilution of 1:40, and variants of this virus (TA, A, B) did not produce hemolysis, but possessed relatively high hemagglutinating properties.

Discussion

In analyzing the data obtained, one may draw the conclusion that virulence and enzymatic activity are related in Newcastle disease virus. V. D. Solov'yev, T. G. Orlov, L. A. Porubel', and I. N. Vasil'yeva [4], in studying genetic markers of virus strains of the A2 group, obtained contradictory results. In their experiments, the non-pathogenic strains possessed high hemagglutinating and enzymatic activity; the pathogenic strains weakly agglutinated chicken and mammalian erythrocytes, and did not possess enzymatic activity. Similar data was also obtained by A. A. Kolchurina [2] with vaccine and pathogenic strains of group A2 virus. Evidently, the lack of agreement of the results of our study and that of the indicated authors may be explained by a difference in the biological properties of the viruses.

By studying the beginning stages of interaction of the virus with the cell, several authors [1, 6, 20] arrived at the conclusion that the adsorption of the virus by the cell depends slightly on its sensitivity to the virus and, evidently, occurs under the influence of a physico-chemical force, and inherent Brownian motion, which is general for all suspended particles. But in the hemagglutinating virus, the interaction begins between its enzyme and the surface receptors of the cell, as a result of which, the destruction of the neuraminic bonds occurs. In our research, the adsorption process, besides the physical factors, evidently, still served as some specific bonds of the virus with the receptors of the cell, while the extremeties bonds depended on the virulence of the virus. The adsorption of the virus by the cell is so specific, that it does not serve as a framework of a simple physical phenomenon. The rate of elution of Newcastle virus from erythrocytes also depends on its virulence, which may be used in selective work. This hypothesis is confirmed b, the results of experimental selection of the avirulent TA_p variant from the original pop-

ulation of the epizootic T strain. The T_{2}^{λ} variant differs in its

enzymatic and virulent properties from the original. We also carried analogous experiments with the virus of the classical chicken plague (Rostok strain); as a result, the avirulent R_4 and R_5 variants were

obtained [3].

In 1957, Padgett and Nalker [16], using various periods of elution of virus of the group, obtained variants possessing different enzymatic activities.

The hemolytic activity of Newcastle disease virus was studied in experiments with erythrocytes of chicken, horse, and guinea pig. The guinea pig erythrocytes proved to be most sensitive. We did not succeed in establishing an interaction between hemagglutinating and hemolytic activity. Analogous results were presented in the work of V. M. Zhdanov and A. C. Bukrinskaya [1] and Khou Yun'-de [5], having studied the different variants of Senday virus. A direct dependence was established between virulence and hemolytic activity.

Results

1. The process of adaptation of Newcastle virus to a heterologous animal (guinea pig) was accompanied by the appearance of genetically heterogeneous virions, which were successfully isolated by the method of serial dificien.

2. The attenuation of the virus was accompanied by reduction of its enzymatic activity (adsorption, elution, hemolysis), while a direct dependence was established between virulence and enzymatic activity. 5. The possibility of obtaining weakly virulent variants of the virus from pathogenic strains according to their enzymatic activities was demonstrated. Published principles of experimental selection, probably, can be used in investigations on the direction of alteration of these viruses.

BIBLIOGRAPHY

 Zhdanov, V. M. and A. G. Bukrinshaya, Acta virol., Vol. 6, p. 105, 1962.

2. Kolchurina, A. A., Vopr. virusol., No. 5, p. 559, 1963.

- 3. Syurin, V. N., V. M. Kolosov, and G. A. Ivanova, et al., Vorp. veterinarn. virusoî., No. 1, p. 327, 1964.
- 4. Solov'yev, V. D., T. G. Orlova, and L. A. Porubel' et al., *Vc: v. virusol.*, No. 6, p. 684, 1961.
- 5. Khou, Yun'-de, Acta virol., Vol. 6, p. 114, 1962. *
- Allison, A. C. and R. C. Valentine, *Ciochim. biophys. Acta*, Vol. 40, p. 393, 1960.
- Atanasiu, P., T. Edipides and J. Basset, Ann. Inst. Pasteur, Vol. 89, p. 523, 1955.
- 8. Burnet, F. M., Nature, Vol. 164, p. 100, 1949.
- 9. Bradly, C. A. et al., Am. J. vet. Res., Vol. 7, p. 307, 1946.
- 10. Granoff, A. and W. Henle, J. Immunol., Vol. 72, p. 322, 1954.
- 11. Hsiung, G. D. and J. L. Melnick, Ibid., Vol. 78, p. 128, 1957.
- Kilham, L., L. N. Loomis and J. H. Peers, Am. J. vet. Res., Vol. 13, p. 95, 1952.
- 13. Kilham, L., Proc. Soc. exp. Bible., Vol. 71, p. 63, New York, 1949.

-12-

- McCollum, W. H. and C. A. Bradly, Am. J. vet. Res., Vol. 16, p. 584, 1955.
- 15. Piraino, F. and R. P. Hanson, Virology, Vol. 8, p. 383, 1959.
- Padgett, B. L. and D. L. Walker, J. exp. Med., Vol. 106, p. 53, 1957.
- 17. Reagan, R. L. and M. G. Lillie et al., Am. J. vet. Res., Vol. 8, p. 136, 1947.
- 18. Sagik, B. et al., J. exp. Med., Vol. 99, p. 251, 1954.
- 19. Sagik, B. P. and S. Levine, Virology, Vol. 3, p. 401, 1957.
- 20. Valentine, R. C. and A. C. Allison, Biochim. biophys. Acta, Vol. 34, p. 10, 1959.
- 21. Verge, J and L. Placidi, C. R. Acad. Sci., Vol. 242, p. 422, Paris, 1956.

22. Vogel, J., Shelokov A. Science, Vol. 126, p. 358, 1957.

-13-

| (Security classification of title, body of about | ENT CONTROL DATA - R & D | | | | |
|--|--|---|--|--|--|
| | : and country excatation must be entered when the urstall | report in classified) | | | |
| PRISHATING ACTIVITY (Converse author) | ZA. REPORT SECURI | Y CLASSIFICATION | | | |
| IS Army science and reemotogy ex- | 26. GROUP | ISSIFIED | | | |
| epartment of the Army | | | | | |
| REPORT TITLE | | | | | |
| Experimental Variation of No. | wcastle Disease Virus | | | | |
| | | | | | |
| RECHIPTINE NOTES (Type of report and inclusive d | Lipa) | | | | |
| Translation | | | | | |
| au Thor(3) (Firef name, middle Millai, Jael name) | | | | | |
| V. M. Kolosov, and V. N. Fyr | arin | | | | |
| | | | | | |
| NEPORT DATE | 74. TOTAL NO. OF PAGES 76. N 13 | D. OF REFS | | | |
| CONTRACT OR GRANT NO | Se. ORIGINATOR'S REPORT NUMBERIS | <u>N/A</u> | | | |
| · · · · · · · · · · · · | | | | | |
| PROJECT NO. 0703006 | | | | | |
| | FSTC-HT-23-658-68 | | | | |
| 922 3 020 2 3 01 | the report) | werd max may be estimat | | | |
| | ACSI Control Number (N | one) | | | |
| DISTRIBUTION STATEMENT | | | | | |
| Distribution of this document | (S. 344) - 31 HT | | | | |
| Distribution of this document. | | | | | |
| SUPPLEMENTARY NOTES | 112 SPONSORING MILITARY ACTIVITY | | | | |
| | S Army Foreign Science | and Technology | | | |
| | Center | | | | |
| | and a second | | | | |
| | | | | | |
| . Adoptation of Nowcast] | a disease virus to guines pigs was | accom- | | | |
| | - UISCASC VIIVA LU EULIUU VALV WWW | | | | |
| Adaptation of newcast | genetically dissimilar virions. W | nich | | | |
| nained by the appearance of were successfully isolated | genetically dissimilar virions, w by the method of serial dilution. | nich Atten- | | | |
| were successfully isolated vation of the virus was acc | genetically dissimilar virions, w by the method of serial dilution. companied by a reduction in its enz | nich Atten- ymatic | | | |
| were successfully isolated vation of the virus was acc activity (adsorption, eluti | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel | hich Atten- ymatic ationship | | | |
| natived by the appearance of were successfully isolated uation of the virus was acc activity (adsorption, eluti was established between vir | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En | hich Atten- ymatic ationship zymatically | | | |
| was established between vir active variants were precip | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy | Atten- ymatic ationship zymatically tes, and | | | |
| nained by the appearance of were successfully isolated uation of the virus was acc activity (adsorption, eluti was established between vir active variants were precip an avirulent TA2 variant, i | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was | hich Atten- ymatic ationship zymatically tes, and shown to activity | | | |
| native variants were precip an avirulent TA2 variant, i be less active to select wea | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was y. It is suggested that enzymatic kly virulent variants from pathoge | hich Atten- ymatic ationship zymatically tes, and shown to activity hic strains | | | |
| naimed by the appearance of were successfully isolated vation of the virus was acc activity (adsorption, eluti was established between vir active variants were precip an avirulent TA2 variant, i be less active enzymaticall could be used to select wea of the virus. | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was y. It is suggested that enzymatic kly virulent variants from pathoge | hich Atten- ymatic stionship zymatically tes, and shown to activity hic strains | | | |
| named by the appearance of were successfully isolated uation of the virus was acc activity (adsorption, eluti was established between vir active variants were precip an avirulent TA2 variant, i be less active enzymatically could be used to select wea of the virus. | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was y. It is suggested that enzymatic kly virulent variants from pathoge | hich Atten- ymatic ationship zymatically tes, and shown to activity hic strains | | | |
| named by the appearance of were successfully isolated uation of the virus was acc activity (adsorption, eluti was established between vir active variants were precip an avirulent TA2 variant, i be less active enzymatically could be used to select wea of the virus. | genetically dissimilar virions, w by the method of serial dilution. companied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was y. It is suggested that enzymatic kly virulent variants from pathoge | hich Atten- ymatic ationship zymatically tes, and shown to activity hic strains | | | |
| named by the appearance of were successfully isolated vation of the virus was acc activity (adsorption, eluti was established between vir active variants were precip an avirulent TA2 variant, i be less active enzymaticall could be used to select wea of the virus. | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was y. It is suggested that enzymatic kly virulent variants from pathoge | hich Atten- ymatic ationship zymatically tes, and shown to activity hic strains | | | |
| named by the appearance of were successfully isolated uation of the virus was acc activity (adsorption, eluti was established between vir active variants were precip an avirulent TA2 variant, i be less active enzymaticall could be used to select wea of the virus. | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was y. It is suggested that enzymatic kly virulent variants from pathoge | hich Atten- ymatic ationship zymatically tes, and shown to activity nic strains | | | |
| named by the appearance of were successfully isolated vation of the virus was acc activity (adsorption, eluti was established between vir active variants were precip an avirulent TA2 variant, i be less active enzymatically could be used to select wea of the virus. | genetically dissimilar virions, w by the method of serial dilution. companied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was y. It is suggested that enzymatic kly virulent variants from pathoge | hich Atten- ymatic ationship zymatically tes, and shown to activity hic strains | | | |
| named by the appearance of were successfully isolated vation of the virus was acc activity (adsorption, eluti was established between vir active variants were precip an avirulent TA2 variant, i be less active enzymaticall could be used to select wea of the virus. | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was y. It is suggested that enzymatic kly virulent variants from pathoge | hich Atten- ymatic ationship zymatically tes, and shown to activity hic strains | | | |
| Adaptation of Newcasti pained by the appearance of were successfully isolated uation of the virus was acc activity (adsorption, eluti was established between vir active variants were precip an avirulent TA2 variant, i be less active enzymaticall could be used to select wea of the virus. | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was y. It is suggested that enzymatic kly virulent variants from pathoge | hich Atten- ymatic ationship zymatically tes, and shown to activity hic strains | | | |
| named by the appearance of were successfully isolated uation of the virus was acc activity (adsorption, eluti was established between vir active variants were precip an avirulent TA2 variant, i be less active enzymatically could be used to select wea of the virus. | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was y. It is suggested that enzymatic kly virulent variants from pathoge | hich Atten- ymatic ationship zymatically tes, and shown to activity hic strains | | | |
| Adaptation of Newcastin paimed by the appearance of were successfully isolated uation of the virus was acc activity (adsorption, eluti was established between vir active variants were precip an avirulent TA2 variant, i be less active enzymaticall could be used to select wea of the virus. | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was y. It is suggested that enzymatic kly virulent variants from pathoge | hich Atten- ymatic ationship zymatically tes, and shown to activity hic strains | | | |
| D . 1473 SEPLACES PROTECT | genetically dissimilar virions, w by the method of serial dilution. ompanied by a reduction in its enz on, hemolysis), while a direct rel ulents and enzymatic activity. En itated with formalinized erythrocy solated from the supernatent, was y. It is suggested that enzymatic kly virulent variants from pathoge With the supernate of the supernate | hich Atten- ymatic ationship zymatically tes, and shown to activity nic strains | | | |

•

•

.

•

:

•

۱

UNCLASSIFIED

| ◀. | KEY WORDS | | | LIN | K 8 | LIN | кс |
|-----|---------------------------------------|------|----------|------------------|-------|------|----|
| | | ROLE | WT | ROLE | wr | ROLE | WT |
| | | | | | | |] |
| | Newcastle disease virus | ł | ł | | | | l |
| | adsorption . | | | | | | |
| | hemolysis | | | ł | ł | | |
| | virulence | i | 1 | 1 | 1 | | |
| | enzymatic activity | | | 1 | | | |
| _ | | 1 | l | 1 | ļ | 1 | |
| - | | | | | | 1 | |
| | | | | | | | |
| | | | | ļ | 1 | Ì | |
| | | | | ł | | } | |
| | | | | 1 | | 1 | |
| | | | | 1 | | [| |
| . ' | | | | | | • | |
| | | [| | 1 | | | |
| | | | | | | | |
| | | | ь. | | | | |
| | · · · · · · · · · · · · · · · · · · · | | | | | | |
| | | | | · · | | | |
| | | | | | - | | |
| | |] | | а ^н , | | | |
| | · | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | [] | •. |
| | | | | | | | |
| | | | | | | | |
| | | | | | | · · | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | • | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |
| | | | UNCL | ASSIFI | ED | · | |
| | | | Security | Classifi | ation | | |
| | | | | | | | |