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TITLE: Definition of the T Cell-Mediated Immune Response to Mammaglobin, a Novel Breast Cancer-Associated Protein

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4. TITLE AND SUBTITLE
Definition of the T Cell-Mediated Immune Response to Mammaglobin, a Novel Breast Cancer-Associated Protein

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13. ABSTRACT (Maximum 200 Words)
The elucidation of the immune response to cancer should be of great help in the development of new therapeutic strategies for the treatment of breast cancer. Based on recent advances in our understanding of antigen recognition by T lymphocytes, it has been possible to identify several tumor-associated antigens (TAA) recognized by CTLs. However, the expression for these TAAs has been shown to be relatively low in breast cancer tumor cells. A new protein named mammaglobin has been demonstrated to be exclusively expressed in the mammary epithelium. In addition, 90% of primary breast cancer tumors have high levels of expression of the mammaglobin protein. Given the exclusive mammaglobin expression in breast cancer tumors, this novel protein may prove to be a TAA highly specific for breast cancer that could be utilized in the near future for in vitro breast cancer-specific activation of CTLs. The discovery of mammaglobin-derived antigenic peptides that are highly expressed in breast cancer tumor tissue and are recognized by CTLs offer many exciting future therapeutic options for the treatment of breast cancer.
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Introduction

A novel breast cancer (BC)-specific protein, mammaglobin, has been identified in our laboratories. Analyses of 15 human adult and 3 fetal tissues have demonstrated that mammaglobin is exclusively expressed in the mammary epithelium during proliferation and terminal differentiation. Significantly, high expression of mammaglobin has been found in 50% of human BC cell lines as well as in 62% metastatic primary BC tumors. Therefore, the characterization of the immune response to a specific and highly expressed BC-associated protein, such as mammaglobin should be of great importance toward the development of new immuno-therapeutic strategies for the treatment and prevention of this disease.

Body

Task 1. To determine whether mammaglobin-reactive T cells generated in vitro have the ability to lyse BC tumor cells.

We have obtained 15 different BC cell lines. Seven of these BC cell lines were found to be positive for mammaglobin expression. After HLA typing, three of the mammaglobin-positive BC cell lines were found to be HLA-A*0101, two were found to be HLA-A*0301, and one was HLA-A*0201. We developed lymphoblastoid cell lines (LCL) from peripheral blood mononuclear cells (PBMC) from 3 healthy male individuals that expressed matching HLA-A molecules to at least one of the BC cell lines mentioned above: No. 1: HLA-A*0101, No. 2: HLA-A*0301, and No. 3: HLA-A*0201. Subsequently, The LCL from each individual was transfected with the mammaglobin gene and then used for the development of mammaglobin reactive CD8+ T cell lines in vitro. The cytotoxic activity of these cell lines was tested using mammaglobin-transfected LCLs as well as one of several of the HLA-A matched BC cell lines mentioned above. All the original CD8+ T cell lines generated in vitro by this method showed high levels of cytolytic activity against both parental and mammaglobin-transfected LCLs but not against HLA-A-matched BC cell lines. Cytolytic activity against other HLA-A-matched LCLs but not BC cell lines led us to conclude that the CTL activity is not directed against mammaglobin but most likely against some EBV antigens.

Based on the results presented above, we developed a CD8+ T cell line from individual No. 3 against the mutant T2 cell line (HLA-A*0201) pulsed with a mammaglobin-derived peptide (aa 83-92) that binds the HLA-A*0201 molecule. We also developed CD8+ T cell lines from individual No. 2 against a HLA-A*0301-transfected mutant T2 cell line pulsed with mammaglobin-derived peptides (aa 23-31 or aa 31-39) that bind the HLA-A*0301 molecule. After 3 weekly stimulations, we tested the CTL activity of these CD8+ T cell lines against T2 cells loaded with the corresponding peptide and against an HLA-A-matched BC cell line. No mammaglobin- or BC-specific CTL activity was detected in any of the CD8+ T cell lines tested even in the presence of high levels of IL-2 (100 units/ml for 7 days). The mammaglobin-specificity of one of these cell lines was determined by means of the peptide-specific induction of expression of the CD69 activation marker as shown in Figure 1 (see appendix). Interestingly, mammaglobin-specific T cell lines from 2 healthy female HLA-A*0201-positive individuals using the same protocol could not be maintained in vitro for more than 1 week of culture. Further, we have developed mammaglobin-specific CD4+ T cell lines from three healthy individuals by means of incubating PBMCs in the presence of recombinant mammaglobin protein. As shown in Figure 2 (see appendix), one of the cell lines developed from a healthy male individual displays a mammaglobin-specific proliferative response. The same CD4+ T cell line did not show any cytolitic activity against a mammaglobin-pulsed self LCL. Interestingly, similarly developed CD4+ T cell lines from 2 healthy female individuals did not show any specific proliferative response against self antigen presenting cells pulsed with recombinant mammaglobin protein. These results strongly suggest that females may be tolerant to mammaglobin.

Task 2. To determine whether the BC-specific T cell immune response generated in vivo can recognize mammaglobin-derived antigenic peptides.

We developed CD8+ T cell lines from a HLA-A*0201-positive BC patient against self monocyte-derived dendritic cells (DC) pulsed with the mammaglobin-derived peptides 3-12 (SKM67) or 83-92 (SKM68). After 3 weekly stimulations, we tested the mammaglobin-specificity of both CD8+ T cell lines
by means of peptide-specific proliferative response against SKM67 and SKM68 peptides (Figures 3 and 4, respectively (see appendix)). In addition, the CTL activity of one of these CD8+ T cell lines (anti-SKM68) was tested against self DCs loaded with the corresponding peptide. A moderate mammaglobin-specific CTL activity was detected in this CD8+ T cell line tested as shown in figure 5 (see appendix).

**Key Research Accomplishments**

✦ Normal females are tolerant to mammaglobin.
✦ Breast cancer patients respond to mammaglobin, tolerance may be broken.

**Reportable Outcomes**

There have been no manuscripts, abstracts, presentations, patents and licenses applies for and/or issued, degrees obtained that are supported by this award, development of cell lines, tissue or serum repositories, informatics such as databases and animal models, etc., funding applied for based on work supported by this award, employment or research opportunities applied for and/or received on experiences/training supported by this award.

**Conclusions**

Overall, the data presented herein indicate that healthy female individuals have developed both CD4+ and CD8+ T cell tolerance to mammaglobin-derived peptides while healthy male individuals have not. Since mammaglobin is significantly over-expressed in BC tumors, the possibility exist that T cell tolerance may have been broken in vivo in these patients. Indeed, our positive results obtaining CD8+ T cell lines from a BC patient using peptide-pulsed DCs indicate this possibility. We have started limiting dilution analyses to determine the precursor frequency of mammaglobin-specific CD4+ as well as CD8+ T cells in both BC patients as well as healthy control female individuals in order to confirmed these results.

**References**

None used.

**Appendices**

Figure 1: Mammaglobin-specific activation of CD8+ T cells
Figure 2: Specific CD4+ T cell proliferative response to mammaglobin
Figure 3: Antigen specific proliferative response of induced CTLs after stimulation with cognate peptide (HLA-A2)
Figure 4: Antigen specific proliferative response of induced CTLs after stimulation with cognate peptide (HLA-A2)
Figure 5: Induction of CTL responses by peptide pulsed DCs
Appendices

Figure 1: Mammaglobin-specific activation of CD8+ T cells

Figure 2: Specific CD4+ T cell proliferative response to mammaglobin

Figure 3: Antigen specific proliferative response of induced CTLs after stimulation with cognate peptide (HLA-A2)

Figure 4: Antigen specific proliferative response of induced CTLs after stimulation with cognate peptide (HLA-A2)

Figure 5: Induction of CTL responses by peptide pulsed DCs
Figure 1: Mammaglobin-specific Activation of CD8+ T cells

**Stimulators**

- T₈ cells plus irrelevant HLA-A2-binding peptide
- T₈ cells plus mammaglobin peptide 83-92
Incorporation, BSA (5 μg/ml) or medium alone (control) were used as negative controls.

Response to recombinant mammaglobin (5 μg/ml) was determined by means of 3H-thymidine incorporation. PBMCs and 30 μg/ml of recombinant human IL-2. On day 4, the proliferative response to PBMCs was recorded. On day 7, the CD4+ T cells were purified and re-stimulated in the presence of PBMCs from a healthy male individual were stimulated with 10 μg/ml of recombinant mammaglobin protein. Figure 2: Specific CD4+ T cell proliferative response to recombinant mammaglobin protein.

![Graph showing proliferative response](image)

Figure 2: Specific CD4+ T cell proliferative response to recombinant mammaglobin protein.
The assay was performed after 6 stimulation. Phased with H3 Thymidine, and harvested after an additional 16 hours. Residual or irrelevant peptide (HLA-A3), 48 hours later, the cultures were residue of irrelevant specific HLA-A2 restricted peptide or similar anchor. Mammoglobin specific HLA-A2 restricted peptide (SKM-67) or SKM-68, another 5x10^6 coated with the co-cultate peptide (SKM-67) or SKM-68, another HLA-A2 (HLA-A2) were cultured in presence of autologous irradiated DC peptide). triplicate wells containing 5x10^4 cells from Ck breast patient peptide. 

Figure 3. Ten days after the last stimulation with SKM-67(HLA-A2

Counts per minute

Coagulate Peptide

of Induced CTLs after Stimulation with Antigen-Specific Peptide Response

FIG 3.
The assay was performed after 6 stimulations, and harvested after an additional 16 hours. Pulsed with H3 Thymidine, and harvested after 48 hours later, the cultures were resided or irrelevant peptide (HLA-A3). Another 5x10^3 cells were coated with the cognate peptide (SKM-68) or SKM-67, another HLA-A2 specific peptide. Similar anchor mammoglobin-specific HLA-A2 restricted peptide of similar anchor mamoglobin specific HLA-A2 were cultured in presence of autologous irradiated DC (HLA-A2), triplicate wells containing 5x10^4 cells from Ca-breast patient.

Figure 4. Ten days after the last stimulation with SKM-68-HLA-A2 peptide, no peptide, IR-67, SKM-68. Counts per minute with cognate peptide response of induced CTLs after stimulation.
Target pulsed for 2h with 50 µg of the conjugate peptide.

Determined in a standard 4h 31Cr release assay using autologous DC as a CTL response in vitro. Cytotoxic activity of induced CTL's was

Days in RPMI-1640 supplemented with GM-CSF and IL-4. DC pulsed

Figure 3. Adherent peripheral blood mononuclear cells were grown for 7

E:T

![Diagram showing E:T ratio with bars for DC alone, IRP-peptide, and SKM-68.]

Peptide pulsed dendritic cells

Induction of CTL response by

Figure 5.