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SUBJECT OF INVESTIGATION

STUDIES ON THE MODE OF ACTION OF ANTIBACTERIAL DRUGS.

RESPONSIBLE INVESTIGATOR

Katsuhiko Tago.

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ANTIBACTERIAL DRUGS

RESPONSIBLE INVESTIGATOR

Dr. Katsuhiko Tago
Assistant, Chief, Tuberculosis Section
Kitasato Institute
Tokyo, Japan
1. Preparation of Cl₄-leucomycin by fermentation using radio-active precursors.

a. Materials and Methods.

(1) Strain used: Streptomyces kitasatoensis was used in these studies. The strain was transferred to slants of either leucomycin medium or starch synthetic medium to which 2% agar was added. The slant cultures were incubated 3 to 7 days. The spores were gently scraped from the culture surface to form a spore inoculum.

(2) Medium used: The following two media were used throughout the studies.

(a) Leucomycin medium.

<table>
<thead>
<tr>
<th>Component</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glucose</td>
<td>4.0g</td>
</tr>
<tr>
<td>NaCl</td>
<td>0.5g</td>
</tr>
<tr>
<td>K₂HPO₄</td>
<td>0.1g</td>
</tr>
<tr>
<td>(NH₄)₂SO₄</td>
<td>0.3g</td>
</tr>
<tr>
<td>C₆H₇NO₇</td>
<td>0.5g</td>
</tr>
<tr>
<td>Urea</td>
<td>0.05g</td>
</tr>
<tr>
<td>Soi-bean powder</td>
<td>2.0g</td>
</tr>
<tr>
<td>Yeast extract</td>
<td>0.5g</td>
</tr>
<tr>
<td>CaCO₃</td>
<td>0.3g</td>
</tr>
<tr>
<td>Ag. dest.</td>
<td>100 ml</td>
</tr>
</tbody>
</table>

Adjust pH 7.4. Agar is added 2.0% in solid medium.

(b) Starch-synthetic medium.

<table>
<thead>
<tr>
<th>Component</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>Soluble starch</td>
<td>0.2g</td>
</tr>
<tr>
<td>K₂HPO₄</td>
<td>0.05g</td>
</tr>
<tr>
<td>MgSO₄</td>
<td>0.04g</td>
</tr>
<tr>
<td>CaCl₂</td>
<td>0.02g</td>
</tr>
<tr>
<td>NaNO₃</td>
<td>0.005g</td>
</tr>
<tr>
<td>Asparagin</td>
<td>0.005g</td>
</tr>
<tr>
<td>Fe₂(SO₄)₃</td>
<td>0.001g</td>
</tr>
<tr>
<td>Ag. dest.</td>
<td>100 ml</td>
</tr>
</tbody>
</table>

Adjust pH 7.4. Agar is added 2.0% in solid medium.
(3) **Method of fermentation:** 100 ml. of starch medium was distributed to 250 ml. Erlenmeyer flasks and inoculated 1.0 ml. of 48 hrs. culture of *S. kitasatoensis* in leucomycin media. Labeled precursors were added to the medium either as solid prior to sterilization or as aqueous solution which had been passed through an ultrafine bacteriological filter and added after sterilization. The fermentation was carried out at 30°C on a shaker.

(4) **Isolation of leucomycin:** After filtration of mycelium, the culture filtrate was extracted twice with 50 ml. of ether which was evaporated in vacuo. Dried leucomycin was solved with 1.0 ml. of ether and diluted with adequate amount of aq. dest.

(5) **Estimation of radio-activity:** 1.0 ml. of solution of leucomycin was distributed to aluminum cup and vacuum-dried. The 14-C content of dried preparation was estimated by use of 'Riken' radiation counter model RSC-3B.
b. Result

(1) Fig. 1 shows the relationship between pH value of culture filtrates and incubation period. All media showed the reversal of pH value on the fourth day of the incubation. Among the media tested, leucomycin medium showed the most significant reversal and starch media inoculated spores of *S. kitasatoensis* slightest reversal. Production of leucomycin in media was paralleled to the reversal of media. Contents of leucomycin in leucomycin-medium was 40 mcg. per ml, and the lowest starch medium inoculated spores was 5 mcg. per ml. The optimal concentration of soluble starch in synthetic medium was 0.2 g per liter.

(2) Twenty hours after inoculation of *S. kitasatoensis* in 100 ml. of starch-synthetic medium, 0.1 mc. of C\(^{14}\)-sodium acetate or C\(^{14}\)-Starch-U were added as the precursor. Three days after addition of them, radioactive leucomycin was extracted from the culture filtrate. C\(^{14}\)-acetate-leucomycin showed very high specific activity: 208,000 cpm per \(\mu\) mol, whereas C\(^{14}\)-starch leucomycin was only 263 cpm per \(\mu\) mol.
2. Fractionation of Staphylococcal cells treated with radioactive leucomycin.

a. Materials and Methods.

(1) Cold TCA fraction: All low-molecular weight compounds soluble in 5% (w/v) trichloroacetic acid are contained. The organism was suspended in 2 ml. cold water, added 0.5 ml. cold 25% (w/v) TCA; after 10 min., centrifuged at 4,000 g for 5 min., and decanted extract.

(2) Aqueous ethanol-soluble fraction: Ethanol-soluble "proteins" and lipid are contained. The residue was suspended in 2.5 ml. 75% (w/v) ethanol in water; after 10 min. at room temperature centrifuged (4,000 g 10 min.) and decanted extract.

(3) Hot TCA fraction: Breakdown products of nucleic acid and teichoic acid. The residue was suspended in 2.5 ml. 5% (w/v) TCA, heated 6 min. at 90°C, cooled, centrifuged (4,000 g 10 min.) and decanted extract.

(4) Trypsin-solubilized: Trypsin-degraded proteins. The residue was suspended in 0.95 ml. 0.05 N-NH₄HCO₃ containing 0.005 N-NH₄OH:0.05 ml. of solution containing 1 mg. crystalline trypsin per ml. was added. Incubated 2 hrs. at 37°C or until digestion was complete. Centrifuged (4,000 g, 10 min.) and decanted extract.

(5) Residue: Mucopeptide of wall. The residue was suspended in 1.0 ml. of water.
b. Results.

Staphylococcal cells treated with $^{14}$-leucomycin were fractionated by a modified method of Park et al., and radio activity of each fraction was estimated. As shown in Table 2, the high radio activity were revealed in the trypsin digested protein fraction and cell wall mucopeptide fraction, whereas low radio activity were found from ethanol soluble lipid fraction and nucleic acid fraction.

Table 1. Fractionation of Staphylococcus aureus treated with $^{14}$-leucomycin.

<table>
<thead>
<tr>
<th>Fraction</th>
<th>Contents of fraction</th>
<th>Opm</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Cold TCA</td>
<td>All low-molecular weight compounds soluble in 5% (w/v) TCA</td>
<td>35.0</td>
</tr>
<tr>
<td>2. Aqueous ethanol soluble</td>
<td>Ethanol-soluble protein and lipid</td>
<td>11.5</td>
</tr>
<tr>
<td>3. Hot TCA</td>
<td>Breakdown products of nucleic acid and teichoic acid.</td>
<td>0</td>
</tr>
<tr>
<td>4. Trypsin-solubilized</td>
<td>Trypsin-degraded protein</td>
<td>52.5</td>
</tr>
<tr>
<td>5. Residue</td>
<td>Mucopeptide of wall</td>
<td>70.0</td>
</tr>
</tbody>
</table>
3. Preparation of ribosomes from staphylococcal cells treated with radioactive leucomycin.

a. Materials and Methods

Bacterial extracts were prepared by grinding washed whole cells with alumina. The alumina and cells (3:1) were homogeneously mixed in an ice cold mortar. To the white paste-like material 6 volume of extracting solvent was added equal to the wet weight of cells. The solvent contained: Tris buffer (0.1M pH 7.8); MgCl₂ (7 x 10⁻²M); and KCl (6 x 10⁻²M). The mixture of solvent, cells and alumina was vigorously ground for 5 min. Precipitable material was removed by centrifugation (10 min, 10,000g). The precipitate was re-extracted two more times as before except 2 volumes of extracting solvent were used each times; the supernatants of all centrifugations were then combined and recentrifuged (120 min, 10⁵,000g). The radiation count of the precipitate was performed as mentioned before.

b. Results

Radioactivity of ³¹⁴-sodium acetate treated leucomycin was 25 cpm per mg N and that of ³¹⁴-staphyl-leucomycin treated was 15 cpm per mg N. For the estimation of such a low radiactivity, it will be necessary to use a gas-flow windowless radiation-counter.
4. Preparation of \(^{14}\text{C}\)-leucomycin by biological method.

a. Materials and methods.

A trained female mongrel dog was anesthetized with an intravenous injection of sodium pental, the cystic duct was tied off with braided silk through a medial abdominal incision. The inner part of a lucite Thomas duodenal fistula was placed opposite the duodenal papilla at which the common bile duct opens; The outer part will be inserted in a lateral abdominal incision about one inch to the right of the middle line and one inch from the last rib. Leucomycin tartrate which assayed 1,010 mcg. of leucomycin activity per mg., was administrated intravenously in a dose of 150 mg. per kg. body weight. Bile collected was made for the subsequent six hours by cannulation of the common bile duct with the dog lying at rest. Bile was extracted with five equal volume of chloroform and the green chloroform solution was washed with water, dried over anhydrous sodium sulfate and evaporated to dryness under vacuum.

b. Results.

About 80 mg. of light yellow powder, which is assumed de-\(\text{N}\)-methyl-leucomycin was extracted with chloroform from the dog bile. The antibacterial activity of this powder was 200 mcg. per mg. Seventy six mg. of this powder was methylated with CH\(_3\)J in methanol and 28 mg solid material was obtained. But this material was assayed the equivalent of only 250 mcg. leucomycin per mg. As the amount of the sample was too little, the completion of methylation was not clear; but from its low antibacterial activity, it might not be true leucomycin. This fact will be clarified when \(^{14}\text{CH}_{3}\)-methyl-jodid is used in our future experiment.
5. Radiography

E. coli strain K-12 and Staphylococcus aureus FDA 209-P were employed for this experiment. The organisms were cultured in synthetic media containing all the essential amino acids, purines, pyrimidines, vitamins, glucose as energy source and other minerals. Before the addition of labelled compound, logarithmic growth had been maintained by shaking at 37°C for several hours. The organisms were harvested by centrifugation, washed once with 0.1 M phosphate buffer pH 7.2 and resuspended in fresh media containing 5-14C adenine sulfate instead of unlabelled adenine at the concentration of 0.21 micromol/l. Then, the incubation was continued in a water bath at 37°C with shaking. 1 ml of samples were taken at different times and added to the same volume of cold 0.5 M perchloric acid. After the 30 minutes preservation in the cold, the materials were washed and resuspended in 0.1 M phosphate buffer pH 7.2. This suspension was spotted on a slide glass treated with egg albumin solution. Preparation of autoradiogram was carried out with colloidion-calcinium bromide method developed by Czibor et al. The advantage of this procedure is at good resolution obtained.

In the technique of the autoradiography, the time of exposure must be determined empirically. Therefore, the standardization of the procedure is now in progress to obtain a constant result.
FIG. 1 REVERSAL OF PH VALUE

- LEUCOMYCIN MEDIUM
- STARCH MEDIUM
- SOLUBLE-STARCH MEDIUM
- STARCH MEDIUM SPORE INOCULUM
- SOLUBLE-STARCH MEDIUM SPORE INOCULUM

INCUBATION PERIOD IN DAYS

PH

8.0
7.0
6.0
5.0
1 2 3 4 5 6
FIG. 2 PRODUCTION OF LEUCOMYCIN

- LEUCOMYCIN MEDIUM
- STARCH MEDIUM
- SOLUBLE-STARCH MEDIUM
- STARCH MEDIUM SPORE INOCULUM
- SOLUBLE-STARCH MEDIUM SPORE INOCULUM

MCC/ML LEUCOMYCIN

INCUBATION PERIOD IN DAYS